

Appendix 3: Synonyms

Appendix 3a: Synonyms by Name

This appendix contains all of the synonyms found in the literature (excluding new name combinations and lapsi calami). For each of these synonyms the current name is included to be able to link the name to the information in the main text and the other appendices.

Original Name

?*Necromantis fragmentum* Ravel, 2016
 ?*Palaeophyllophora tunisiensis* Ravel, 2016
 ?*Vespertiliavus aenigma* Ravel, 2016
[Epomophorus (Nanonycteris)] Matschie, 1899
[Epomophorus macrocephalus] var. *angolensis* Gray, 1870
[Epomophorus macrocephalus] var. *unicolor* Gray, 1870
[Nyctinomus (Nyctinomus) pumilus] Var. *Major* Trouessart, 1897
[Pipistrellus Kuhl] latastei Laurent, 1937
[Rhinolophus hipposideros] *escaleræ* K. Andersen, 1918
[Triaenops] *furinea* Tate, 1941

[Vespertilio auritus] *β aegyptius* J.B. Fischer, 1829
[Vespertilio pipistrellus] var. *β Aegyptius* J.B. Fischer, 1829
[Vesperugo (Vesperus) serotinus] Var. *Gabonensis* Trouessart, 1897
[Xantharpyia (Myonycteris)] Matschie, 1899
acrotis G.M. Allen, 1914
Adelonycteris H. Allen, 1892
Aegyptonycteridae Simmons, Seiffert and Gunnell, 2016
Aegyptonycteris Simmons, Seiffert and Gunnell, 2016
Aegyptonycteris knightae Simmons, Seiffert and Gunnell, 2016
Aëorestes Fitzinger, 1870
Afrillonura Rosina and Pickford, 2021
Afrillonura namibensis Rosina and Pickford, 2021
Afronycteris Monadjem, Patterson and Demos, 2020
Afropipistrellus Thorn, Kock and Cuisin, 2007
Afropterus Lavocat, 1961
Afropterus gigas Lavocat, 1961
Afropterus gigas Lavocat, 1961
Allomops J.A. Allen, 1917
Alobus Peters, 1868
Amblyotus Kolenati, 1858
Anamygdon Troughton, 1929
ANIMALIVORA Gill, 1872
Aquias Gray, 1847
Aristippe Kolenati, 1863
Asellia Gray, 1838
Asellia patrizii de Beaux, 1931
Asellia tridens diluta K. Andersen, 1918
Asellia tridens italo-somalica de Beaux, 1931
Asellia tridens pallida Laurent, 1937
Asellia vetus Lavocat, 1961
Attalepharca Menu, 1987
Austronomus Troughton, 1941
Austronomus Iredale and Troughton, 1934
Barbastella Gray, 1821
Barbastella barbastellus guanchae Trujillo, Ibáñez and Juste, 2002

Current Name

Necromantis fragmentum (Ravel, 2016)
Palaeophyllophora tunisiensis (Ravel, 2016)
Vespertiliavus aenigma (Ravel, 2016)
Nanonycteris Matschie, 1899

Epomophorus angolensis Gray, 1870

Epomophorus wahlbergi (Sundevall, 1846)

Mops (Chaerephon) major (Trouessart, 1897)

Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Rhinolophus hipposideros (Bechstein, 1800)

Paratriaenops pauliani (Goodman and Ranivo, 2008)
Plecotus christii Gray, 1838

Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)

Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Myonycteris Matschie, 1899
Rhinolophus fumigatus Rüppell, 1842
Eptesicus Rafinesque, 1820
AEGYPTONYCTERIDAE Simmons, Seiffert and Gunnell, 2016
Aegyptonycteris Simmons, Seiffert and Gunnell, 2016
Aegyptonycteris knightae Simmons, Seiffert and Gunnell, 2016
Myotis Kaup, 1829
Afrillonura Rosina and Pickford, 2021
Afrillonura namibensis Rosina and Pickford, 2021
Afronycteris Monadjem, Patterson and Demos, 2020
Nycticeinops Hill & Harrison, 1987
Megaderma E. Geoffroy St.-Hilaire, 1810
Megaderma gigas (Lavocat, 1961)
Megaderma gaillardi (Trouessart, 1898)
Mops (Mops) Lesson, 1842
Pipistrellus Kaup, 1829
Eptesicus Rafinesque, 1820
Myotis Kaup, 1829
CHIROPTERA Blumenbach, 1779
Rhinolophus Lacépède, 1799
Eptesicus Rafinesque, 1820
Asellia Gray, 1838
Asellia patrizii de Beaux, 1931
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Asellia italosomalica de Beaux, 1931
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Hipposideros vetus (Lavocat, 1961)
Pipistrellus Kaup, 1829
Tadarida Rafinesque, 1814
Tadarida Rafinesque, 1814
Barbastella Gray, 1821
Barbastella barbastellus (Schreber, 1774)

Original Name

Barbastellus Kaup, 1829
Barbastellus communis Gray, 1838
Brachipposideros Sigé, 1968
Brachyotus Kolenati, 1856
Brachyura Peters, 1865
Brevipalatus Hand and Archer, 2005
C[oleura] g[allarum] nilosa Thomas, 1915
C[oleura] gallarum Thomas, 1915
C[oleura] silhouettæ Thomas, 1915
Capaccinius Bonaparte, 1841
Cardioderma Peters, 1873
Cardioderma leakeyi Gunnell, Butler, Greenwood and Simmons, 2015
Casinycteris Thomas, 1910
Casinycteris argynnis Thomas, 1910
Casinycteris campomaanensis Hassanin, 2014
Cateorus Kolenati, 1856
Cephalotes teniotis Rafinesque, 1814
Cephalotidæ Gray, 1821
Cercopteropus Burnett, 1829
Cerivoula Blanford, 1891
Chærephon (*Lophomops*) J.A. Allen, 1917
Chærephon (*Lophomops*) *abæ* J.A. Allen, 1917
Chærephon (*Lophomops*) *chapini* J.A. Allen, 1917
Chærephon (*Lophomops*) *cristatus* J.A. Allen, 1917
Chærephon (*Lophomops*) *shortridgei* Thomas, 1926
Chærephon atsinanana Goodman, Buccas, Naidoo, Ratrimomanarivo, Taylor and Lamb, 2010
Chærephon emini Wroughton, 1911
Chærephon frater J.A. Allen, 1917

Chærephon jobimena Goodman and Cardiff, 2004
Chærephon lancasteri Hayman, 1938
Chærephon leucostigma G.M. Allen, 1918
Chærephon nigeriæ Thomas, 1913
Chærephon nigeriæ Thomas, 1913

Chærephon pumilus naivashæ Hollister, 1916

Chærephon russatus J.A. Allen, 1917
Chærephon (*Lophomops*) *nigri* Hatt, 1928

Chærephon pumilus elphicki Roberts, 1926

Chalinolobus Peters, 1867
Chalinolobus argentatus Dobson, 1875
Chalinolobus congicus Noack, 1889
Chambinycteris Ravel, 2016
Chambinycteris pusilli Ravel, 2016
Chamtwaria Butler, 1984
Chamtwaria pickfordi Butler, 1984
Cheiomelinae Legendre, 1984
CHEIROPTERA Gray, 1821
CHEIROPTERA Fleming, 1822
Cheiroptères G. Cuvier, 1816
CHIROPTERA Blumenbach, 1779
Chiropteren Duméril, 1806
Chiropteri de Rochebrune, 1883
Chiropteria Rafinesque, 1815
Chiropteriformes Kinman, 1994
Choerephon (*Lophomops*) *langi* Roberts, 1932

Choerephon websteri Dollman, 1908

Chrysonycteris Gray, 1866
Chrysopteron Jentink, 1910
Cistugo Thomas, 1912

Current Name

Barbastella Gray, 1821
Barbastella barbastellus (Schreber, 1774)
Brachipposideros Sigé, 1968
Myotis Kaup, 1829
EMBALLONURIDAE Gervais, 1855
Brevipalatus Hand and Archer, 2005
Coleura afra (Peters, 1852)
Coleura gallarum Thomas, 1915
Coleura seychellensis Peters, 1868
Myotis Kaup, 1829
Cardioderma Peters, 1873
Cardioderma leakeyi Gunnell, Butler, Greenwood and Simmons, 2015
Casinycteris Thomas, 1910
Casinycteris argynnis Thomas, 1910
Casinycteris campomaanensis Hassanin, 2014

Eptesicus Rafinesque, 1820
Tadarida teniotis (Rafinesque, 1814)
PTEROPODIDAE Gray, 1821
Rousettus Gray, 1821
Kerivoula Gray, 1842
Mops (*Chaerephon*) Dobson, 1874
Mops (*Chaerephon*) *major* (Trouessart, 1897)

Mops (*Chaerephon*) *chapini* (J.A. Allen, 1917)

Mops (*Chaerephon*) *leucogaster* (A. Grandidier, 1869)
Mops (*Chaerephon*) *chapini* (J.A. Allen, 1917)

Mops (*Chaerephon*) *atsinanana* (Goodman, Buccas, Naidoo, Ratrimomanarivo, Taylor and Lamb, 2010)
Mops (*Mops*) *condylurus* (A. Smith, 1833)
Mops (*Chaerephon*) *leucogaster* (A. Grandidier, 1869)
Mops (*Chaerephon*) *jobimena* (Goodman and Cardiff, 2004)
Mops (*Chaerephon*) *chapini* (J.A. Allen, 1917)
Mops (*Mops*) *leucostigma* (G.M. Allen, 1918)
Mops (*Chaerephon*) *nigeriæ* (Thomas, 1913)
Mops (*Chaerephon*) *nigeriæ nigeriæ* (Thomas, 1913)
Mops (*Chaerephon*) *pumilus* (Cretzschmar, 1826)
Mops (*Chaerephon*) *russatus* (J.A. Allen, 1917)
Mops (*Chaerephon*) *leucogaster* (A. Grandidier, 1869)
Mops (*Chaerephon*) *pumilus* (Cretzschmar, 1826)

Glauconycteris Dobson, 1875
Glauconycteris argentata (Dobson, 1875)
Glauconycteris argentata (Dobson, 1875)
Chambinycteris Ravel, 2016
Chambinycteris pusilli Ravel, 2016
Chamtwaria Butler, 1984
Chamtwaria pickfordi Butler, 1984
Molossinae Gervais, 1856
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
Mops (*Chaerephon*) *pumilus* (Cretzschmar, 1826)
Mops (*Chaerephon*) *leucogaster* (A. Grandidier, 1869)
Hipposideros Gray, 1831
Myotis Kaup, 1829
Cistugo Thomas, 1912

Original Name

Cistugo lesueuri Roberts, 1919
Cistugo seabrae Thomas, 1912
Clæotis Thomas, 1901
Clæotis Percivali Thomas, 1901
Clæotis percivali australis Roberts, 1917
Cnephaeus Kaup, 1829
Coeloephyllus Peters, 1867
Coelophyllus Peters, 1867
Coelopinae Tate, 1941
Colëura Peters, 1867
Coleura kibomalandy Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012
Coleura kummeri Monard, 1939
Coleura muthokai Wesselman, 1984
Coleura seychellensis Peters, 1868
Comastes Fitzinger, 1870
Cyclorhina Peters, 1871
Cynonycteris Peters, 1852
Cynonycteris Angolensis Bocage, 1898

Cynonycteris Angolensis Bocage, 1898
Cynonycteris brachycephala Bocage, 1889
Cynonycteris torquata Dobson, 1878
Cynonycteris torquata Dobson, 1878
Cynopterinae K. Andersen, 1912
Cynopterini Koopman and J.K. Jones Jr., 1970
Cynopterus collaris Gray, 1870
DERMAPTERA Aristotle, -330
Desmaplex Miller, 1907
Dhofarella Sigé, Thomas, Sen, Gheerbrant, Roger and Al-Sulaimani, 1994
Dhofarella sigei Gunnell, Simons and Seiffert, 2008
Dhofarella thaleri Sigé, Thomas, Sen, Gheerbrant, Roger and Al-Sulaimani, 1994
Dichromyotis Bianchi, 1917
Diclidurina Gray, 1866
Diclidurinae Miller, 1907
Dinops Savi, 1825
Dinops Cestonii Savi, 1825
Dizya Sigé, 1991
Dizya exsultans Sigé, 1991
Doryrhina Peters, 1871
Drakonycteris Ravel, 2016
Drakonycteris glibzegdouensis Ravel, 2016
Dysopes Cretzschmar, 1830-1831
Dysopes brachypterus Peters, 1852

Dysopes dubius Peters, 1852

Dysopes geoffroyi Temminck, 1827

Dysopes hepaticus Heuglin, 1864
Dysopes limbatus Peters, 1852

Dysopes midas Sundevall, 1843
Dysopes midas Sundevall, 1843
Dysopes natalensis A. Smith, 1847

Dysopes pumilus Cretzschmar, 1826

Dysopes rüpelii Temminck, 1827
Dysopes Savii Schinz, 1840
Dysopes talpinus Heuglin, 1877

E[uryalus] *atlanticus* K. Andersen and Matschie, 1904
E[uryalus] *barbarus* K. Andersen and Matschie, 1904
E[uryalus] *Cabrerae* K. Andersen and Matschie, 1904
E[uryalus] *meridionalis* K. Andersen and Matschie, 1904

Current Name

Cistugo lesueuri Roberts, 1919
Cistugo seabrae Thomas, 1912
Cloeotis Thomas, 1901
Cloeotis percivali Thomas, 1901
Cloeotis percivali Thomas, 1901
Eptesicus (Cnephaeus) Kaup, 1829
Rhinolophus Lacépède, 1799
Rhinolophus Lacépède, 1799
HIPPOSIDERIDAE Lydekker, 1891
Coleura Peters, 1867
Coleura kibomalandy Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012
Coleura afra (Peters, 1852)
Coleura muthokai Wesselman, 1984
Coleura seychellensis Peters, 1868
Myotis Kaup, 1829
Hipposideros Gray, 1831
Rousettus Gray, 1821
Myonycteris angolensis angolensis (Bocage, 1898)
Myonycteris angolensis (Bocage, 1898)
Myonycteris brachycephala (Bocage, 1889)
Myonycteris torquata (Dobson, 1878)
Myonycteris torquata torquata (Dobson, 1878)
Pteropodinae Gray, 1821
Pteropodinae Gray, 1821
Myonycteris torquata (Dobson, 1878)
CHIROPTERA Blumenbach, 1779
Pteropus Brisson, 1762
Dhofarella Sigé, Thomas, Sen, Gheerbrant, Roger and Al-Sulaimani, 1985
Dhofarella sigei Gunnell, Simons and Seiffert, 2008
Dhofarella thaleri Sigé et al., 1994

Myotis Kaup, 1829
EMBALLONURIDAE Gervais, 1855
EMBALLONURIDAE Gervais, 1855
Tadarida Rafinesque, 1814
Tadarida teniotis (Rafinesque, 1814)
Dizya Sigé, 1991
Dizya exsultans Sigé, 1991
Doryrhina Peters, 1871
Drakonycteris Ravel, 2016
Drakonycteris glibzegdouensis Ravel, 2016
Tadarida Rafinesque, 1814
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Tadarida aegyptiaca aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Mops (Mops) condylurus (A. Smith, 1833)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Mops (Mops) midas (Sundevall, 1843)
Mops (Mops) midas midas (Sundevall, 1843)
Mormopterus acetabulosus natalensis (A. Smith, 1847)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Tadarida teniotis (Rafinesque, 1814)
Tadarida teniotis (Rafinesque, 1814)
Tadarida aegyptiaca aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Rhinolophus euryale Blasius, 1853

Rhinolophus mehelyi Matschie, 1901

Rhinolophus euryale Blasius, 1853

Rhinolophus mehelyi Matschie, 1901

Original Name

Eidolinae Almeida, Giannini and Simmons, 2016
Eidolon Rafinesque, 1815
Eidolon helvum annobonensis Juste, Ibáñez and Machordom, 2000
Eleutherura Gray, 1844
Eleutherura unicolor Gray, 1870

Emballonura afra Peters, 1852
Emballonura atrata Peters, 1874
Emballonura tiavato Goodman, Cardiff, Ranivo, Russell and Yoder, 2006
Emballonuridae Dobson, 1875
Emballonuridae Gervais, 1855
Emballonurina Gray, 1866
Emballonurinae Gervais, 1855
Emballonurini Winge, 1893
Emballonurini Gervais, 1855
Emballonuroidea Teeling, Springer, Madsen, Bates, O'Brien and Murphy, 2005
Emballonuroidea Weber, 1928
Entomophaga A. Murray, 1866

Eomops Thomas, 1905
Ep[omophorus (Epomophorus)] büttikoferi Matschie, 1899
Ep[omophorus (Epomophorus)] doriae Matschie, 1899
Ep[omophorus (Epomophorus)] neumanni Matschie, 1899
Ep[omophorus (Epomophorus)] stuhlmanni Matschie, 1899
Ep[omophorus (Epomophorus)] zechi Matschie, 1899
Ep[omophorus (Epomophorus)] zenkeri Matschie, 1899
Epomophorina Gray, 1866
Epomophorinae K. Andersen, 1912
Epomophorini Gray, 1866
Epomophorus Bennett, 1836
Epomophorus anchietæ Seabra, 1900
Epomophorus anseli Bergmans and Van Strien, 2004
Epomophorus anurus Heuglin, 1864
Epomophorus comptus H. Allen, 1862
Epomophorus crypturus Peters, 1852
Epomophorus Dobsonii Bocage, 1889
Epomophorus franqueti Tomes, 1860
Epomophorus gambianus parvus Ansell, 1960
Epomophorus guineensis Bocage, 1898
Epomophorus guineensis Bocage, 1898

Epomophorus labiatus anurus
Epomophorus macrocephalus Peters, 1876
Epomophorus macrocephalus var. *unicolor* Gray, 1870
Epomophorus minimus Claessen and De Vree, 1991
Epomophorus minor Dobson, 1880
Epomophorus pousarguesi Trouessart, 1904

Epomophorus pusillus Peters, 1868
Epomophorus reii Aellen, 1950

Epomophorus reii Aellen, 1950
Epomophorus sp. Hill and Morris, 1971
Epomophorus veldkampii Jentink, 1888
Epomops Gray, 1870
Epomops franqueti strepitans K. Andersen, 1910
Eptesicops Roberts, 1926
Eptesicus Rafinesque, 1820
Eptesicus ater J.A. Allen, 1917

Current Name

Eidolinae Almeida, Giannini and Simmons, 2016
Eidolon Rafinesque, 1815
Eidolon helvum helvum (Kerr, 1792)

Rousettus Gray, 1821
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Coleura afra (Peters, 1852)
Paremballonura atrata (Peters, 1874)
Paremballonura tiavato (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)
EMBALLONURIDAE Gervais, 1855
EMBALLONURIDAE Gervais, 1855
EMBALLONURIDAE Gervais, 1855
Emballonurinae Gervais, 1855
EMBALLONURIDAE Gervais, 1855
Emballonurini Gervais, 1855
NYCTEROIDEA Van der Hoeven, 1855

EMBALLONURIDAE Gervais, 1855
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
Myopterus E. Geoffroy St.-Hilaire, 1818
Epomops buettikoferi (Matschie, 1899)

Epomophorus labiatus (Temminck, 1837)

Epomophorus wahlbergi (Sundevall, 1846)

Epomophorus wahlbergi (Sundevall, 1846)

Epomophorus gambianus gambianus (Ogilby, 1835)
Epomophorus wahlbergi (Sundevall, 1846)

Pteropodinae Gray, 1821
Rousettinae Andersen, 1912
Epomophorini Gray, 1866
Epomophorus Bennett, 1836
Plerotes anchietæ (Seabra, 1900)
Epomophorus anseli Bergmans and Van Strien, 2004
Epomophorus labiatus (Temminck, 1837)
Epomops franqueti (Tomes, 1860)
Epomophorus crypturus Peters, 1852
Epomophorus dobsonii Bocage, 1889
Epomops franqueti (Tomes, 1860)
Epomophorus crypturus Peters, 1852
Epomophorus gambianus (Ogilby, 1835)
Epomophorus gambianus gambianus (Ogilby, 1835)
Epomophorus minor Dobson, 1880
Hypsignathus monstrosus H. Allen, 1862
Epomophorus wahlbergi (Sundevall, 1846)

Epomophorus minimus Claessen & De Vree, 1991

Epomophorus minor Dobson, 1880
Epomophorus gambianus pousarguesi Trouessart, 1904

Epomophorus pusillus Peters, 1868
Epomophorus gambianus gambianus (Ogilby, 1835)

Epomophorus labiatus (Temminck, 1837)
Epomophorus labiatus (Temminck, 1837)
Nanonycteris veldkampii (Jentink, 1888)
Epomops Gray, 1866
Epomops franqueti (Tomes, 1860)

Pipistrellus Kaup, 1829
Eptesicus Rafinesque, 1820
Pseudoromicia tenuipinnis (Peters, 1872)

Original Name

Eptesicus capensis angolensis J.Eric Hill, 1937
Eptesicus capensis nkatiensis Roberts, 1932
Eptesicus faradjius J.A. Allen, 1917
Eptesicus garambæ J.A. Allen, 1917
Eptesicus hottentotus bensoni Roberts, 1946

Eptesicus hottentotus portavernus Schlitter and Aggundey, 1986
Eptesicus loveni Granvik, 1924
Eptesicus megalurus pallidior Shortridge, 1942

Eptesicus melckorum Roberts, 1919
Eptesicus phasma G.M. Allen, 1911
Eptesicus rectitragus Wettstein, 1916
Eptesicus somalicus malagasyensis Peterson, Eger and Mitchell, 1995
Eptesicus ugandæ Hollister, 1916
Eptesicus zuluensis Roberts, 1924
Eptesiformes Menu, 1987
Eucheira Hodgson, 1847
Eunyeris Gray, 1866
Euryalus Matschie, 1901
Euvespertilio Acloque, 1899
Euvesperugo Acloque, 1899
Exochurus Fitzinger, 1870
FRUCTIVORAE Gray, 1821

FRUGIVORA Giebel, 1855

Glauconycteris Dobson, 1875
Glauconycteris alboguttatus J.A. Allen, 1917
Glauconycteris atra Hassanin, Colombo, Gembu, Merle, Tu, Görföl, Musaba Akawa, Csorba, Kearney, Monadjem and Ing, 2017
Glauconycteris beatrix Thomas, 1901
Glauconycteris curryi Eger and Schlitter, 2001

Glauconycteris egeria Thomas, 1913
Glauconycteris Floweri de Winton, 1901

Glauconycteris gleni Peterson and Smith, 1973
Glauconycteris humeralis J.A. Allen, 1917
Glauconycteris kenyacola Peterson, 1982
Glauconycteris machadoi Hayman, 1963
Glauconycteris papilio Thomas, 1905
Glauconycteris phalæna Thomas, 1915
Glauconycteris superba Hayman, 1939
Glauconycteris superba sheila Hayman, 1947
Gloionycteris Gray, 1866
Gymnorhina Giebel, 1855

Gymnorhina Wagner, 1843
Gymnorhinina Fatio, 1869
Gymnuridae Ameghino, 1889
H[ipposiderus]. gigas gambiensis K. Andersen, 1906
H[ypsignathus] monstrosus H. Allen, 1862
Harpyidae C. H. Smith, 1842
Hesperomyotis Cabrera, 1958
Hipposideridae Miller, 1907
Hipposideridae Lydekker, 1891
Hipposiderina Koopman, 1994
Hipposiderinae Lydekker, 1891
Hipposiderini Koopman and J.K. Jones Jr., 1970
Hipposideros Gray, 1831
Hipposideros (Pseudorhinolophus) africanum Ravel, 2016
Hipposideros (Pseudorhinolophus) amenhotepos Gunnell, Winkler, Miller, Head, El-Barkooky, Gawad, Sanders and Gingerich, 2015

Current Name

Nycticeinops grandidieri (Dobson, 1876)

Laephotis capensis (A. Smith, 1829)
Pseudoromicia rendalli (Thomas, 1889)
Laephotis capensis (A. Smith, 1829)
Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
Myotis tricolor (Temminck, 1832)
Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
Laephotis capensis (A. Smith, 1829)
Pseudoromicia rendalli (Thomas, 1889)
Neoromicia guineensis (Bocage, 1889)
Laephotis malagasyensis (Peterson, Eger and Mitchell, 1995)
Neoromicia somalica (Thomas, 1901)
Neoromicia zuluensis (Roberts, 1924)
Vespertilioninae Gray, 1821
Megaderma E. Geoffroy St.-Hilaire, 1810
Pteropus Brisson, 1762
Rhinolophus Lacépède, 1799
Myotis Kaup, 1829
Pipistrellus Kaup, 1829
Myotis Kaup, 1829
PTEROPODIFORMI Van Cakenberghe, Kearney and Seamark, 2007
PTEROPODIFORMI Van Cakenberghe, Kearney and Seamark, 2007
Glauconycteris Dobson, 1875
Glauconycteris alboguttata J.A. Allen, 1917
Glauconycteris atra Hassanin, Colombo, Gembu, Merle, Tu, Görföl, Musaba Akawa, Csorba, Kearney, Monadjem and Ing, 2017
Glauconycteris beatrix Thomas, 1901
Glauconycteris curryae Eger and Schlitter, 2001
Glauconycteris egeria Thomas, 1913
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Glauconycteris gleni Peterson and Smith, 1973

Glauconycteris humeralis J.A. Allen, 1917
Glauconycteris kenyacola Peterson, 1982
Glauconycteris machadoi Hayman, 1963
Glauconycteris variegata (Tomes, 1861)
Glauconycteris variegata (Tomes, 1861)
Glauconycteris superba Hayman, 1939
Glauconycteris superba Hayman, 1939
Hipposideros Gray, 1831
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
VESPERTILIONIDAE Gray, 1821
VESPERTILIONIDAE Gray, 1821
MOLOSSIDAE Gervais, 1856
Macronycteris gigas (Wagner, 1845)

Hypsignathus monstrosus H. Allen, 1862
PTEROPODIDAE Gray, 1821
Myotis Kaup, 1829
HIPPOSIDERIDAE Lydekker, 1891
HIPPOSIDERIDAE Lydekker, 1891
HIPPOSIDERIDAE Lydekker, 1891
HIPPOSIDERIDAE Lydekker, 1891
HIPPOSIDERIDAE Lydekker, 1891

Hipposideros Gray, 1831
Hipposideros africanum Ravel, 2016

Hipposideros amenhotepos Gunnell, Winkler, Miller, Head, El-Barkooky, Gawad, Sanders and Gingerich, 2015

Original Name

Hipposideros abae J.A. Allen, 1917
Hipposideros beatus maximus Verschuren, 1957
Hipposideros besaoka Samonds, 2007
Hipposideros braima Monard, 1939
Hipposideros caffer aurantiaca de Beaux, 1924
Hipposideros caffer niapu J.A. Allen, 1917
Hipposideros camerunensis Eisentraut, 1956
Hipposideros cf. caffer Monadjem, Richards and Denys, 2016
Hipposideros cf. ruber (lineage C1) Monadjem, Richards and Denys, 2016
Hipposideros cf. ruber (lineage D1) Mamba, Dalton, Mahlaba, Kropff and Monadjem, 2020
Hipposideros cf. ruber (lineage E1) Monadjem, Richards and Denys, 2016
Hipposideros Commersoni mostellum Thomas, 1904
Hipposideros cryptovalorona Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016
Hipposideros curtus G.M. Allen, 1921
Hipposideros gigas niangaræ J.A. Allen, 1917
Hipposideros gigas viegasi Monard, 1939
Hipposideros jonesi Hayman, 1947
Hipposideros kaumbului Wesselman, 1984
Hipposideros lamottei Brosset, 1985
Hipposideros langi J.A. Allen, 1917
Hipposideros marisae Aellen, 1954
Hipposideros nanus J.A. Allen, 1917
Hipposideros sandersoni Sanderson, 1937
Hipposiderus Gray, 1834
Hipposiderus beatus K. Andersen, 1906
Hipposiderus caffer centralis K. Andersen, 1906
Hipposiderus caffer guineensis K. Andersen, 1906
Hipposiderus tephros Cabrera, 1906
Histiorrhina Van der Hoeven, 1855
Histiorrhina Haeckel, 1866

Hypodermida Haeckel, 1866
Hypsignathus H. Allen, 1862
Hypsignathus haldemani Matschie, 1899
Hypsugo Kolenati, 1856
Hypsugo bemaity Goodman, Rakotondramanana, Ramasindrazana, Kearney, Monadjem, Schoeman, Taylor, Naughton and Appleton, 2015
Indorhinolophus Guillén-Servent, Francis and Ricklefs, 2003
INSECTIVORAE Gray, 1821

Isotus Kolenati, 1856
Kehelvoulha Jentink, 1904
Kerivoula Gray, 1842
Kerivoula africana Dobson, 1878
Kerivoula argentata Tomes, 1861
Kerivoula brunnea Dobson, 1878
Kerivoula cuprosa Thomas, 1912
Kerivoula harrisoni Thomas, 1901
Kerivoula harrisoni bellula Aellen, 1959
Kerivoula lucia Hinton, 1920
Kerivoula lueia Kershaw, 1922
Kerivoula muscilla Thomas, 1906
Kerivoula nidicola zuluensis Roberts, 1924
Kerivoula phalaena Thomas, 1912
Kerivoula poensis Gray, 1842
Kerivoula Smithii Thomas, 1880
Kerivoulinae Miller, 1907
Khonsunycteris Gunnell, Simons and Seiffert, 2008

Current Name

Hipposideros abae J.A. Allen, 1917
Hipposideros beatus (K. Andersen, 1906)

Macronycteris besaoka (Samonds, 2007)
Hipposideros tephros (Cabrera, 1906)
Hipposideros caffer (Sundevall, 1846)

Hipposideros ruber (Noack, 1893)
Doryrhina camerunensis (Eisentraut, 1956)
Hipposideros caffer (Sundevall, 1846)

Hipposideros ruber (Noack, 1893)

Hipposideros ruber (Noack, 1893)

Macronycteris vittatus (Peters, 1852)

Macronycteris cryptovalorona (Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016)
Hipposideros curtus G.M. Allen, 1921
Macronycteris gigas (Wagner, 1845)
Macronycteris gigas (Wagner, 1845)
Hipposideros jonesi Hayman, 1947
Hipposideros kaumbului Wesselman, 1984
Hipposideros lamottei Brosset, 1985
Doryrhina cyclops (Temminck, 1853)
Hipposideros marisae Aellen, 1954
Hipposideros caffer (Sundevall, 1846)
Hipposideros curtus G.M. Allen, 1921
Hipposideros Gray, 1831
Hipposideros beatus (K. Andersen, 1906)
Hipposideros ruber (Noack, 1893)

Hipposideros ruber (Noack, 1893)

Hipposideros tephros (Cabrera, 1906)
RHINOLOPHIDAE Gray, 1825
RHINOLOPHIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
PTEROPODIDAE Gray, 1821
Hypsignathus H. Allen, 1862
Hypsignathus monstrosus H. Allen, 1862
Hypsugo Kolenati, 1856
Neoromicia bemaity (Goodman, Rakotondramanana, Ramasindrazana, Kearney, Monadjem, Schoeman, Taylor, Naughton and Appleton, 2015)
Rhinolophus Lacépède, 1799

VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
Myotis Kaup, 1829
Kerivoula Gray, 1842
Kerivoula Gray, 1842
Kerivoula africana Dobson, 1878
Kerivoula argentata Tomes, 1861
Kerivoula lanosa (A. Smith, 1847)
Kerivoula cuprosa Thomas, 1912
Kerivoula lanosa (A. Smith, 1847)
Kerivoula lanosa (A. Smith, 1847)
Kerivoula lanosa (A. Smith, 1847)
Kerivoula lanosa (A. Smith, 1847)
Kerivoula lanosa (A. Smith, 1847)
Kerivoula argentata Tomes, 1861
Kerivoula phalaena Thomas, 1912
Glauconycteris poensis (Gray, 1842)
Kerivoula smithii Thomas, 1880
Kerivoulinae Miller, 1907
Khonsunycteris Gunnell, Simons and Seiffert, 2008

Original Name

Khonsunyeris aegypticus Gunnell, Simons and Seiffert, 2008
Khoufechia Ravel, 2016
Khoufechia gunnelli Ravel, 2016
Kirivoula Gervais, 1849
Laephotis Thomas, 1901
Laephotis angolensis Monard, 1935
Laephotis botswanae Setzer, 1971
Laephotis cf. angolensis
Laephotis kirinyaga Monadjem, Patterson, Webala and Demos, 2020
Laephotis namibensis Setzer, 1971
Laephotis Wintoni Thomas, 1901
Lavia Gray, 1838
Lavia frons affinis K. Andersen and Wroughton, 1907
Lavia rex Miller, 1905
Leiponyx Jentink, 1881
Leiponyx büttikoferi Jentink, 1881
Leuconoe Boie, 1830
Leuconoformes Menu, 1987
Leuconoïdes Menu, 1987
Liponycteris Thomas, 1922
Liponyx Forbes, 1882
Lissonycteris angolensis goliath Bergmans, 1997
Lissonycteris angolensis petraea Bergmans, 1997
Lyroderma Peters, 1872
M[acronycteris] cryptovalorona Foley, Goodman, Whelan, Puechmaille and Teeling, 2017
Macronycteris Gray, 1866
Macrotus Leach, 1816
major E. Geoffroy Saint-Hilaire, 1803
MEGACHIROPTERA Dobson, 1875

Megachiropteramorpha Simmons and Geisler, 1998
Megaderma E. Geoffroy St.-Hilaire, 1810
Megaderma cor Peters, 1872
Megaderma frons E. Geoffroy Saint-Hilaire, 1810
Megaderma jaegeri Sigé, 1976
Megadermata Peters, 1866
Megadermatidae H. Allen, 1864
Megadermatini Winge, 1893
Megadermidae Gill, 1872
Megadermina Gray, 1866
Megaloglossus Pagenstecher, 1885
Megaloglossus azagnyi Nesi, Kadjo and Hassanin, 2012
Megaloglossus woermanni Pagenstecher, 1885
Megaloglossus woermanni prigoginei Hayman, 1966
Megapipistrellus Bianchi, 1917
Meteoros Kolenati, 1856
MICROCHIROPTERA Dobson, 1875
Microchiropteraformes Simmons and Geisler, 1998
Microchiropteramorpha Simmons and Geisler, 1998
Micropteropus grandis Sanborn, 1950
Micropteropus intermedius Hayman, 1963
midas Schulze, 1897
Mimetillus Thomas, 1904
Mimetillus bernerii Monard, 1933
Mimetillus thomasi Hinton, 1920
Miniopterae Trouessart, 1898
Miniopteri Dobson, 1875
Miniopteridae Dobson, 1875
Miniopterus Bonaparte, 1837

Current Name

Khonsunyeris aegypticus Gunnell, Simons and Seiffert, 2008
Khoufechia Ravel, 2016
Khoufechia gunnelli Ravel, 2016
Kerivoula Gray, 1842
Laephotis Thomas, 1901
Laephotis angolensis Monard, 1935
Laephotis botswanae Setzer, 1971
Laephotis angolensis Monard, 1935
Laephotis kirinyaga Monadjem, Patterson, Webala and Demos, 2020
Laephotis namibensis Setzer, 1971
Laephotis wintoni Thomas, 1901
Lavia Gray, 1838
Lavia frons (E. Geoffroy St.-Hilaire, 1810)

Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Eidolon Rafinesque, 1815
Eidolon helvum helvum (Kerr, 1792)
Myotis Kaup, 1829
Myotinae Tate, 1942
Vespertilioninae Gray, 1821
Taphozous E. Geoffroy St.-Hilaire, 1818
Eidolon Rafinesque, 1815
Myonycteris angolensis goliath (Bergmans, 1997)
Myonycteris angolensis petraea (Bergmans, 1997)
Megaderma E. Geoffroy St.-Hilaire, 1810
Macronycteris cryptovalorona (Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016)
Macronycteris Gray, 1866
Plecotus E. Geoffroy St.-Hilaire, 1818
Rhinolophus ferrumequinum (Schreber, 1774)
PTEROPODIFORMI Van Cakenberghe, Kearney and Seamark, 2007
RHINOLOPHIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
Megaderma E. Geoffroy St.-Hilaire, 1810
Cardioderma cor (Peters, 1872)
Lavia frons (E. Geoffroy St.-Hilaire, 1810)

Megaderma jaegeri Sigé, 1976
MEGADERMATIDAE H. Allen, 1864
MEGADERMATIDAE H. Allen, 1864
MEGADERMATIDAE H. Allen, 1864
MEGADERMATIDAE H. Allen, 1864
MEGADERMATIDAE H. Allen, 1864
Megaloglossus Pagenstecher, 1885
Megaloglossus azagnyi Nesi, Kadjo and Hassanin, 2012
Megaloglossus woermanni Pagenstecher, 1885
Megaloglossus woermanni Pagenstecher, 1885
Myotis Kaup, 1829
Eptesicus Rafinesque, 1820
CHIROPTERA Blumenbach, 1779
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
Epomophorus grandis (Sanborn, 1950)
Epomophorus intermedius (Hayman, 1963)
Tadarida teniotis (Rafinesque, 1814)
Mimetillus Thomas, 1904
Mimetillus thomasi Hinton, 1920
Mimetillus thomasi Hinton, 1920
MINIOPTERIDAE Dobson, 1875
MINIOPTERIDAE Dobson, 1875
MINIOPTERIDAE Dobson, 1875
Miniopterus Bonaparte, 1837

Original Name

Miniopterus aelleni Goodman, Maminirina, Weyeneth, Bradman, Christidis, Ruedi and Appleton, 2009
Miniopterus africanus Sanborn, 1936
Miniopterus ambohitrensis Goodman, Ramasindrazana, Naughton and Appleton, 2015
Miniopterus brachytragos Goodman, Maminirina, Bradman, Christidis and Appleton, 2009
Miniopterus breyeri Jameson, 1909
Miniopterus breyeri vicinior J.A. Allen, 1917

Miniopterus cf. inflatus Monadjem, Shapiro, Richards, Karabulut, Crawley, Broman Nielsen, Hansen, Bohmann and Mourier, 2020
Miniopterus egeri Goodman, Ramasindrazana, Maminirina, Schoeman and Appleton, 2011
Miniopterus fraterculus Thomas and Schwann, 1906
Miniopterus gleni Peterson, Eger and Mitchell, 1995
Miniopterus griffithsi Goodman, Maminirina, Bradman, Christidis and Appleton, 2009
Miniopterus horaceki Gunnell, Eiting and Geraads, 2011
Miniopterus inflatus Thomas, 1903
Miniopterus inflatus Thomas, 1903
Miniopterus inflatus villiersi Aellen, 1956
Miniopterus maghrebensis Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014
Miniopterus mahafaliensis Goodman, Bradman, Christides and Appleton, 2009
Miniopterus Majori Thomas, 1906
Miniopterus manavi Thomas, 1906
Miniopterus minor Peters, 1867
Miniopterus minor griveaudi Harrison, 1959
Miniopterus minor occidentalis Juste and Ibañez, 1992
Miniopterus mossambicus Monadjem, Goodman, Stanley and Appleton, 2013
Miniopterus natalensis arenarius Heller, 1912
Miniopterus Newtoni Bocage, 1889
Miniopterus nimbae Monadjem, Shapiro, Richards, Karabulut, Crawley, Nielsen, Hansen, Bohmann and Mourier, 2020
Miniopterus petersoni Goodman, Bradman, Maminirina, Ryan, Christidis and Appleton, 2008
Miniopterus rufus Sanborn, 1936
Miniopterus smitianus Thomas, 1927
Miniopterus sororculus Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton, 2007
Miniopterus sp. , 9999
Miniopterus wilsoni Monadjem, Guyton, Naskrecki, Richards, Kropff and Dalton, 2020
minor Kerr, 1792
Miomegaderma gaillardi Trouessart, 1898
Molossi Peters, 1865
Molossidae Gill, 1872
Molossidae Gervais, 1856
Molossina Gray, 1866
Molossina Gray, 1866
Molossinae Gervais, 1856
Molossini Winge, 1893
Molossoidae Gervais, 1856
Mops Lesson, 1842
Mops Lesson, 1842
Mops (Allomops) faradjius J.A. Allen, 1917
Mops (Allomops) nanulus J.A. Allen, 1917
Mops (Allomops) occipitalis J.A. Allen, 1917

Current Name

Miniopterus aelleni Goodman, Maminirina, Weyeneth, Bradman, Christidis, Ruedi and Appleton, 2009
Miniopterus africanus Sanborn, 1936
Miniopterus ambohitrensis Goodman, Ramasindrazana, Naughton and Appleton, 2015
Miniopterus brachytragos Goodman, Maminirina, Bradman, Christidis and Appleton, 2009
Miniopterus natalensis (A. Smith, 1833)
Miniopterus fraterculus Thomas and Schwann, 1906
Miniopterus cf. inflatus Monadjem, Shapiro, Richards, Karabulut, Crawley, Broman Nielsen, Hansen, Bohmann and Mourier, 2020
Miniopterus egeri Goodman, Ramasindrazana, Maminirina, Schoeman and Appleton, 2011

Miniopterus fraterculus Thomas and Schwann, 1906
Miniopterus gleni Peterson, Eger and Mitchell, 1995
Miniopterus griffithsi Goodman, Maminirina, Bradman, Christidis and Appleton, 2009
Miniopterus horaceki Gunnell, Eiting and Geraads, 2011
Miniopterus inflatus Thomas, 1903
Miniopterus inflatus inflatus Thomas, 1903
Miniopterus villiersi Aellen, 1956
Miniopterus maghrebensis Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014

Miniopterus mahafaliensis Goodman, Bradman, Christides and Appleton, 2009
Miniopterus majori Thomas, 1906
Miniopterus manavi Thomas, 1906
Miniopterus minor Peters, 1867
Miniopterus griveaudi Harrison, 1959
Miniopterus minor Peters, 1867

Miniopterus mossambicus Monadjem, Goodman, Stanley and Appleton, 2013
Miniopterus arenarius Heller, 1912
Miniopterus newtoni Bocage, 1889
Miniopterus nimbae Monadjem, Shapiro, Richards, Karabulut, Crawley, Nielsen, Hansen, Bohmann and Mourier, 2020
Miniopterus petersoni Goodman, Bradman, Maminirina, Ryan, Christidis & Appleton, 2008

Miniopterus inflatus rufus Sanborn, 1936
Miniopterus natalensis (A. Smith, 1833)
Miniopterus sororculus Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton, 2007
Miniopterus Bonaparte, 1837
Miniopterus wilsoni Monadjem, Guyton, Naskrecki, Richards, Kropff and Dalton, 2020
Rhinolophus hipposideros (Bechstein, 1800)
Megaderma gaillardi (Trouessart, 1898)
MOLOSSIDAE Gervais, 1856
MOLOSSIDAE Gervais, 1856
MOLOSSIDAE Gervais, 1856
Molossinae Gervais, 1856
MOLOSSIDAE Gervais, 1856
Molossinae Gervais, 1856
MOLOSSIDAE Gervais, 1856
MOLOSSOIDEA Gervais, 1856
Mops Lesson, 1842
Mops (Mops) Lesson, 1842
Mops (Mops) demonstrator (Thomas, 1903)
Mops (Xiphonycteris) nanulus J.A. Allen, 1917
Mops (Xiphonycteris) thersites (Thomas, 1903)

Original Name

Mops (Allomops) osborni J.A. Allen, 1917
Mops angolensis niveiventer Cabrera and Ruxton, 1926
Mops angolensis orientis G.M. Allen and Loveridge, 1942
Mops angolensis wonderi Sanborn, 1936
Mops bakarii Stanley, 2008
Mops bemmeleni Simmons and Cirranello, 2020
Mops calabarensis Hayman, 1940
Mops chitauensis J.Eric Hill, 1937

Mops conigicus J.A. Allen, 1917
Mops kerio Gunnell and Manthi, 2018
Mops niangaræ J.A. Allen, 1917
Mops osborni occidentalis Monard, 1939
Mops osborni occidentalis f. fulva Monard, 1939
Mops rüppellii Allen, 1939
Mops russatus Simmons and Cirranello, 2020
Mops trevori J.A. Allen, 1917
Mops turkwellensis Gunnell and Manthi, 2018
Mormopterus Peters, 1865
Mormopterus francoismoutoui Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst and Bowie, 2008
Mormopterus setiger Peters, 1878
Mormopterus setiger Peters, 1878
Mormopterus Whiteleyi Scharff, 1900
Myonycteris Matschie, 1899
Myonycterini Lawrence and Novick, 1963
Myonycteris leptodon K. Andersen, 1908
Myonycteris relicta Bergmans, 1980
Myonycteris wroughtoni K. Andersen, 1908

Myoptera de Blainville, 1837
Myopteris Desmarest, 1820
Myopterus E. Geoffroy Saint-Hilaire, 1818
Myopterus albatu Thomas, 1915
Myopterus daubentonii Desmarest, 1820
Myopterus senegalensis Oken, 1816
Myotinae Tate, 1942
Myotini Tate, 1942
Myotini Tate, 1942
Myotis Kaup, 1829
Myotis anjouanensis
Myotis blythii punicus Felten, 1977
Myotis Bocagei cupreolus Thomas, 1904
Myotis cf. punicus Castella, Ruedi, Excoffier, Ibanez, Arlettaz and Hausser, 2000
Myotis darelbeidensis Gunnell, Eiting and Geraads, 2011
Myotis dieteri M. Happold, 2005
Myotis escalerae cabreræ Çoraman, Mayer and Dietz, 2019
Myotis Goudoti anjouanensis Dorst, 1960
Myotis Hildegardæ Thomas, 1904
Myotis morrisi Hill, 1971
Myotis scotti Thomas, 1927
Myotis zenatius Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019
Myzopoda A. Milne-Edwards and A. Grandidier, 1878
Myzopoda africana Gunnell, Butler, Greenwood and Simmons, 2015
Myzopoda aurita A. Milne-Edwards and A. Grandidier, 1878
Myzopoda schliemanni Goodman, Rakotondraparany and Kofoky, 2007
Myzopodidae Thomas, 1904
N(yctinomus) midas de Winton, 1901
N(yctinomus) taeniotis Thomas, 1891
N[ycteris] albiventer Wagner, 1840
N[ycteris] discolor Wagner, 1840

Current Name

Mops (Mops) condylurus (A. Smith, 1833)
Mops (Mops) niveiventer Cabrera and Ruxton, 1926
Mops (Mops) condylurus (A. Smith, 1833)

Mops (Mops) condylurus (A. Smith, 1833)
Mops (Xiphonycteris) bakarii Stanley, 2009
Mops (Chaerephon) bemmeleni (Jentink, 1879)

Mops (Xiphonycteris) nanulus J.A. Allen, 1917
Mops (Mops) niveiventer Cabrera and Ruxton, 1926
Mops (Mops) conigicus J.A. Allen, 1917
Mops kerio Gunnell and Manthi, 2018
Mops (Mops) niangaræ J.A. Allen, 1917
Mops (Mops) condylurus (A. Smith, 1833)
Mops (Mops) condylurus (A. Smith, 1833)

Tadarida ventralis (Heuglin, 1861)
Mops (Chaerephon) russatus (J.A. Allen, 1917)
Mops (Mops) trevori J.A. Allen, 1917
Mops turkwellensis Gunnell and Manthi, 2018
Mormopterus Peters, 1865
Mormopterus francoismoutoui Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst, Bowie, 2008
Platymops setiger (Peters, 1878)
Platymops setiger setiger (Peters, 1878)
Myopterus whiteleyi (Scharff, 1900)
Rousettus Gray, 1821
Myonycterini Lawrence and Novick, 1963
Myonycteris leptodon K. Andersen, 1908
Myonycteris relicta Bergmans, 1980
Myonycteris torquata wroughtoni K. Andersen, 1908

Myopterus E. Geoffroy St.-Hilaire, 1818
Myopterus E. Geoffroy St.-Hilaire, 1818
Myopterus E. Geoffroy St.-Hilaire, 1818
Myopterus daubentonii Desmarest, 1820
Myopterus daubentonii Desmarest, 1820
Myopterus daubentonii Desmarest, 1820
Myotinae Tate, 1942
Myotinae Tate, 1942
VESPERTILIONIDAE Gray, 1821
Myotis Kaup, 1829
Myotis anjouanensis Dorst, 1960
Myotis punicus Felten, 1977
Myotis bocagii cupreolus Thomas, 1904
Myotis punicus Felten, 1977

Myotis darelbeidensis Gunnell, Eiting and Geraads, 2011
Myotis dieteri M. Happold, 2005
Myotis zenatius Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019
Myotis anjouanensis Dorst, 1960
Myotis bocagii bocagii (Peters, 1870)
Myotis morrisi Hill, 1971
Myotis scotti Thomas, 1927
Myotis zenatius Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019
Myzopoda Milne-Edwards and A. Grandidier, 1878
Myzopoda africana Gunnell, Butler, Greenwood and Simmons, 2015
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Myzopoda schliemanni Goodman, Rakotondraparany and Kofoky, 2007
MYZOPODIDAE Thomas, 1904
Tadarida ventralis (Heuglin, 1861)
Tadarida ventralis (Heuglin, 1861)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818

Original Name

N[ycticejus] adovanus Heuglin, 1877
Nannugo Kolenati, 1856
Nannugo Kolenatii Müller, 1858
Necromanter Lydekker, 1888
Necromantidae Sigé, 2011
Necromantis Weithofer, 1888
Necronycteris Palmer, 1903
Néoptésiformes Menu, 1987
Neoromicia Roberts, 1926
Neoromicia "incertae-sedis", 9999
Neoromicia cf. capensis, 9999
Neoromicia isabella Decher, Hutterer and Monadjem, 2016
Neoromicia robertsi Goodman, Taylor, Ratrimomanarivo and Hoofer, 2012
Neoromicia roseveari Monadjem, Richards, Taylor and Stoffberg, 2013
Neoromicia stanleyi Goodman, Kearney, Ratsimbazafy and Hassanin, 2017
Neoromicia vansoni Roberts, 1932
Nicticejus Rüppell, 1842
Numbaha Reeder, Helgen, Vodzak, Lunde and Ejotre, 2013
NOCTILIONIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
Noctilionoidea
Noctula Bonaparte, 1837
Noctulina Macalister, 9999
Noctulinia Cabrera, 1914
Noctulinia Gray, 1842
Nyctaliformes Menu, 1987
Nyctaloïdes Menu, 1987
Nyctalus Bowdich, 1825
Nyctalus verrucosus Bowdich, 1825
Nyctericina Gray, 1866
Nycteridae Van der Hoeven, 1855
Nycterides Haeckel, 1866
NYCTERIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
Nycterikaupius Menu, 1987
Nycterikaupius Menu, 1987
Nycterikaupius Menu, 1987
Nycteris G. Cuvier and E. Geoffroy, 1795
Nycteris aethiopica Dobson, 1878
Nycteris aethiopica guineensis Monard, 1939
Nycteris aethiopica guineensis f. aurantiaca Monard, 1939
Nycteris aethiopica luteola Thomas, 1901
Nycteris affinis A. Smith, 1829
Nycteris angolensis Peters, 1871
Nycteris arge Thomas, 1903
Nycteris avakubia J.A. Allen, 1917
Nycteris Baikii Gray, 1866
Nycteris benuensis Aellen, 1952
Nycteris Capensis A. Smith, 1829
Nycteris cf. parisii Kruskop, Benda, Vasenkov and Lavrenchenko, 2016
Nycteris damarensis Peters, 1871
Nycteris Daubentonii E. Geoffroy Saint-Hilaire, 1813
Nycteris fuliginosa Peters, 1852
Nycteris Geoffroyi Desmarest, 1820
Nycteris Geoffroyi Var. *Senegalensis* Hartmann, 1868
Nycteris grandis Peters, 1865
Nycteris intermedia Aellen, 1959
Nycteris labiata Heuglin, 1861
Nycteris macrotis Dobson, 1876
Nycteris madagascariensis G. Grandidier, 1937
Nycteris major J.A. Allen, 1917
Nycteris marica Kershaw, 1923

Current Name

Nycticeinops schlieffenii (Peters, 1859)
Pipistrellus Kaup, 1829
Pipistrellus (Pipistrellus) "incertae-sedis"
Necromantis Weithofer, 1887
NECROMANTIDAE Sigé, 2011
Necromantis Weithofer, 1887
Necromantis Weithofer, 1887
Vespertilioninae Gray, 1821
Neoromicia Roberts, 1926
Laephotis "incertae-sedis"
Laephotis capensis (A. Smith, 1829)
Pseudoromicia isabella (Decher, Hutterer and Monadjem, 2016)
Laephotis robertsi (Goodman, Taylor, Ratrimomanarivo and Hoofer, 2012)
Pseudoromicia roseveari (Monadjem, Richards, Taylor and Stoffberg, 2013)
Laephotis stanleyi (Goodman, Kearney, Ratsimbazafy and Hassanin, 2017)
Neoromicia zuluensis (Roberts, 1924)
Scotophilus Leach, 1821
Glauconycteris Dobson, 1875

NOCTILIONIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
NOCTILIONOIDEA Gray, 1821
Eptesicus Rafinesque, 1820
Nyctalus Bowdich, 1825
Nyctalus Bowdich, 1825
Nyctalus Bowdich, 1825
Vespertilioninae Gray, 1821
Vespertilioninae Gray, 1821
Nyctalus Bowdich, 1825
Nyctalus leisleri (Kuhl, 1817)
NYCTERIDAE Van der Hoeven, 1855
NYCTERIDAE Van der Hoeven, 1855
CHIROPTERA Blumenbach, 1779
NYCTERIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
Laephotis Thomas, 1901
Pseudoromicia Monadjem, Patterson, Webala and Demos, 2020
Neoromicia Roberts, 1926
Pipistrellus Kaup, 1829
Nycteris G. Cuvier and E. Geoffroy, 1795
Nycteris macrotis Dobson, 1876
Nycteris macrotis Dobson, 1876
Nycteris macrotis Dobson, 1876

Nycteris macrotis Dobson, 1876
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris arge Thomas, 1903
Nycteris major (K. Andersen, 1912)
Nycteris grandis Peters, 1865
Nycteris parisii (de Beaux, 1924)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris cf. parisii De Beaux, 1924

Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris hispida (Schreber, 1774)

Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818

Nycteris grandis Peters, 1865
Nycteris intermedia Aellen, 1959
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris macrotis Dobson, 1876
Nycteris macrotis Dobson, 1876

Nycteris macrotis Dobson, 1876
Nycteris grandis Peters, 1865

Original Name

Nycteris nana tristis G.M. Allen and Lawrence, 1936
Nycteris oriana Kershaw, 1922
Nycteris pallida J.A. Allen, 1917
Nycteris poensis Gray, 1843
Nycteris proxima Lönnberg and Gyldenstolpe, 1925
Nycteris Revoilii Robin, 1881
Nycteris Thebaicus E. Geoffroy Saint-Hilaire, 1818
Nycteris villosa Peters, 1852
Nycteris vinsoni Dalquest, 1965
Nycteris woodi K. Andersen, 1914
Nycteris woodi sabiensis Roberts, 1946
Nycteroides Van der Hoeven, 1855
Nycterops Gray, 1866
Nycterops pilosa Gray, 1866
Nycticeiini Gervais, 1855
Nycticeina Gervais, 1855
Nycticeina Gervais, 1855
Nycticeiini Tate, 1942
Nycticeinops Hill and Harrison, 1987
Nycticeinops bellieri Monadjem, Demos, Dalton, Webala, Musila, Kerbis Peterhans and Patterson, 2020
Nycticeinops serengetiensis Gunnell, Butler, Greenwood and Simmons, 2015
Nycticeius africanus G.M. Allen, 1911
Nycticejinae Gill, 1872
Nycticejus eriophorus Heuglin, 1877
Nycticejus flavigaster Heuglin, 1861
Nycticejus leucogaster Cretzschmar, 1826
Nycticejus murino-flavus Heuglin, 1861
Nycticejus nidicola Kirk, 1865
Nycticejus planirostris Peters, 1852
Nycticejus Schlieffenii Peters, 1859
Nycticejus serratus Heuglin, 1877
Nycticejus viridis Peters, 1852
Nyctimones Gray, 1821
Nyctinoma Bowdich, 1821
Nyctinomia Fleming, 1822
Nyctinomus E. Geoffroy Saint-Hilaire, 1818
Nyctinomus (Chaerephon) Dobson, 1874
Nyctinomus (Dysopes) ventralis Heuglin, 1861
Nyctinomus (Mormopterus) jugularis Peters, 1865
Nyctinomus (Nyctinomus) tongaënsis Wettstein, 1916
Nyctinomus [(Mormopterus)] albiventer Dobson, 1877
Nyctinomus [(Nyctinomus)] cestoni Dobson, 1877
Nyctinomus aegyptiacus E. Geoffroy Saint-Hilaire, 1818
Nyctinomus aegyptiacus E. Geoffroy Saint-Hilaire, 1818
Nyctinomus africanus Dobson, 1876
Nyctinomus aloysii-sabaudiae Festa, 1907

Nyctinomus Anchietæ Seabra, 1900
Nyctinomus angolensis Peters, 1870
Nyctinomus ansorgei Thomas, 1913
Nyctinomus Bemmeleni Jentink, 1879
Nyctinomus bivittatus Heuglin, 1861
Nyctinomus Bocagei Seabra, 1900
Nyctinomus brunneus Seabra, 1900
Nyctinomus cisturus Thomas, 1903
Nyctinomus Condylurus A. Smith, 1833
Nyctinomus demonstrator Thomas, 1903
Nyctinomus dubius A. Smith, 1833

Nyctinomus Emini de Winton, 1901
Nyctinomus fulminans Thomas, 1903

Current Name

Nycteris nana (K. Andersen, 1912)

Nycteris macrotis Dobson, 1876
Nycteris hispida (Schreber, 1774)
Nycteris hispida (Schreber, 1774)
Nycteris grandis Peters, 1865

Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818

Nycteris hispida (Schreber, 1774)
Nycteris vinsoni Dalquest, 1965
Nycteris woodi K. Andersen, 1914
Nycteris woodi K. Andersen, 1914
NYCTEROIDEA Van der Hoeven, 1855
Nycteris G. Cuvier and E. Geoffroy, 1795
Nycteris hispida (Schreber, 1774)
Nycticeiini Gervais, 1855
VESPERTILIONIDAE Gray, 1821
Nycticeiini Gervais, 1855
Vespertilioninae Gray, 1821
Nycticeinops Hill & Harrison, 1987
Nycticeinops bellieri (De Vree, 1972)

Nycticeinops serengetiensis Gunnell, Butler, Greenwood and Simmons, 2015
Nycticeinops schlieffenii (Peters, 1859)
Nycticeiini Gervais, 1855
Kerivoula eriophora (Heuglin, 1877)
Scotophilus leucogaster (Cretzschmar, 1826)
Scotophilus leucogaster (Cretzschmar, 1826)
Scotophilus leucogaster (Cretzschmar, 1826)
Kerivoula argentata Tomes, 1861
Scotophilus dinganii (A. Smith, 1833)
Nycticeinops schlieffenii (Peters, 1859)
Taphozous nudiventris Cretzschmar, 1830
Scotophilus viridis (Peters, 1852)
Tadarida Rafinesque, 1814
Tadarida Rafinesque, 1814
Tadarida Rafinesque, 1814
Tadarida Rafinesque, 1814
Mops (Chaerephon) Dobson, 1874
Tadarida ventralis (Heuglin, 1861)
Mormopterus jugularis (Peters, 1865)

Tadarida aegyptiaca aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Mormopterus jugularis (Peters, 1865)

Tadarida ventralis (Heuglin, 1861)

Tadarida aegyptiaca aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Tadarida ventralis (Heuglin, 1861)
Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
Tadarida aegyptiaca bocagei (Seabra, 1900)
Mops (Mops) condylurus (A. Smith, 1833)
Mops (Chaerephon) ansorgei (Thomas, 1913)
Mops (Chaerephon) bemmeleni (Jentink, 1879)
Mops (Chaerephon) bivittatus (Heuglin, 1861)
Tadarida aegyptiaca bocagei (Seabra, 1900)
Tadarida aegyptiaca bocagei (Seabra, 1900)
Mops (Chaerephon) bemmeleni (Jentink, 1879)
Mops (Mops) condylurus (A. Smith, 1833)
Mops (Mops) demonstrator (Thomas, 1903)
Mormopterus acetabulosus natalensis (A. Smith, 1847)
Mops (Chaerephon) major (Trouessart, 1897)
Tadarida fulminans (Thomas, 1903)

Original Name**Current Name***Nyctinomus gambianus* de Winton, 1901*Mops (Chaerephon) pumilus* (Cretzschmar, 1826)*Nyctinomus hindei* Thomas, 1904*Mops (Chaerephon) pumilus* (Cretzschmar, 1826)*Nyctinomus leonis* Thomas, 1908*Mops (Xiphonycteris) brachypterus* (Peters, 1852)*Nyctinomus leucogaster* A. Grandidier, 1869*Mops (Chaerephon) leucogaster* (A. Grandidier, 1869)*Nyctinomus lobatus* Thomas, 1891*Tadarida lobata* (Thomas, 1891)*Nyctinomus martiensseni* Matschie, 1897*Otomops martiensseni martiensseni* (Matschie, 1897)*Nyctinomus martiensseni* Matschie, 1897*Otomops martiensseni* (Matschie, 1897)*Nyctinomus mastersoni* Roberts, 1946*Tadarida fulminans* (Thomas, 1903)*Nyctinomus mauritanus* Horsfield, 1823-1824*Mormopterus acetabulosus acetabulosus* (Hermann, 1804)*Nyctinomus Miarensis* A. Grandidier, 1869*Mops (Mops) midas miarensis* (A. Grandidier, 1869)*Nyctinomus ochraceus* J.A. Allen, 1917*Mops (Xiphonycteris) brachypterus* (Peters, 1852)*Nyctinomus pusillus* Miller, 1902*Mops (Chaerephon) pusillus* (Miller, 1902)*Nyctinomus rhodesiae* Roberts, 1946*Mops (Chaerephon) ansorgei* (Thomas, 1913)*Nyctinomus spillmanni* Monard, 1933*Mops (Chaerephon) nigeriae spillmanni* (Monard, 1933)*Nyctinomus thersites* Thomas, 1903*Mops (Xiphonycteris) thersites* (Thomas, 1903)*Nyctinomus unicolor* A. Grandidier, 1870*Mops (Mops) midas miarensis* (A. Grandidier, 1869)*Nyctiptennis* Hall and Kelson, 1959*Eptesicus Rafinesque, 1820**Nyctiptenus* Fitzinger, 1870*Eptesicus Rafinesque, 1820**Nyctophilina* Gray, 1866*VESPERTILIONIDAE* Gray, 1821*Nyctophilinae* Peters, 1865*Vespertilioninae* Gray, 1821*Nyctophilinae* Miller, 1907*VESPERTILIONIDAE* Gray, 1821*Nyctophylax* Fitzinger, 1861*Kerivoula* Gray, 1842*Nystactes* Kaup, 1829*Myotis* Kaup, 1829*Otomops* Thomas, 1913*Otomops* Thomas, 1913*Otomops harrisoni* Ralph, Richards, Taylor, Napier and Lamb, 2015*Otomops harrisoni* Ralph, Richards, Taylor, Napier and Lamb, 2015*Otomops icarus* Chubb, 1917*Otomops martiensseni icarus* Chubb, 1917*Otomops madagascariensis* Dorst, 1953*Otomops madagascariensis* Dorst, 1953*Otonycteris* Peters, 1859*Otonycteris* Peters, 1859*Otonycteris cinereus* Satunin, 1909*Otonycteris hemprichii* Peters, 1859*Otonycteris Hemprichii* Peters, 1859*Otonycteris hemprichii* Peters, 1859*Otonycteris jin* Cheesman and Hinton, 1924*Otonycteris hemprichii* Peters, 1859*Otonycteris petersi* Anderson, 1902*Otonycteris hemprichii* Peters, 1859*Pipistrellus kuhlii pallidus* Heim de Balsac, 1936*Pipistrellus (Pipistrellus) kuhlii* (Kuhl, 1817)*Pachyomus* Gray, 1866*Eptesicus Rafinesque, 1820**Pachyotus* Gray, 1831*Scotophilus* Leach, 1821*Pachysoma* Temminck, 1853*Epomophorus* Bennett, 1836*Palaeophyllophora* Revilliod, 1917*Palaeophyllophora* Revilliod, 1917*Panugo* Kolenati, 1856*Nyctalus* Bowdich, 1825*Parahypsugo* Hutterer, Decher, Monadjem and Astrin, 2019*Nycticeinops* Hill & Harrison, 1987*Parahypsugo happoldorum* Hutterer, Decher, Monadjem and Astrin, 2019*Nycticeinops happoldorum* (Hutterer, Decher, Monadjem and Astrin, 2019)*Parahypsugo macrocephalus* Hutterer and Kerbis Peterhans, 2020*Nycticeinops macrocephalus* (Hutterer and Kerbis Peterhans, 2020)*Paramyotis* Bianchi, 1917*Myotis* Kaup, 1829*Paratriaenops* Benda and Vallo, 2009*Paratriaenops* Benda and Vallo, 2009*Paremballonura* Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012*Paremballonura* Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012*Paremballonura atrata* Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012*Paremballonura atrata* (Peters, 1874)*Paremballonura tiavato* Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012*Paremballonura tiavato* (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)*Paremballonura atrata* Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012*Paremballonura tiavato* Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012*Paremballonura tiavato* Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012*Paremballonura tiavato* Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012*Pareptesicus* Bianchi, 1917*Eptesicus Rafinesque, 1820**Petalia* Gray, 1838*Nycteris* G. Cuvier and E. Geoffroy, 1795*Petalia (Nycteris) thebaica aurantiaca* de Beaux, 1923*Nycteris thebaica* E. Geoffroy St.-Hilaire, 1818*Petalia aurita* K. Andersen, 1912*Nycteris aurita* (K. Andersen, 1912)*Petalia damarensis brockmani* K. Andersen, 1912*Nycteris thebaica* E. Geoffroy St.-Hilaire, 1818

Original Name

Petalia damarensis media K. Andersen, 1912
Petalia gambiensis K. Andersen, 1912
Petalia major K. Andersen, 1912
Petalia nana K. Andersen, 1912
Petalia parisii de Beaux, 1924
Petaliidae Miller, 1910
Phasmatonycteris Gunnell, Simmons and Seiffert, 2014
Phasmatonycteris butleri Gunnell, Simmons and Seiffert, 2014
Phasmatonycteris phiomensis Gunnell, Simmons and Seiffert, 2014
Philisidae Sigé, 1985
Philisinae Horáček, Fejfar and Hulva, 2006

Philisis Sigé, 1985
Philisis sphingis Sigé, 1985
Phyllorhina Leach, 1816
Phyllorhina Koch, 1860
Phyllorhina angolensis Seabra, 1898
Phyllorhina commersoni var. *thomensis* Bocage, 1891
Phyllorhina commersonii var. *marungensis* Noack, 1887
Phyllorhina rubra Noack, 1893
Phyllorhina tridens murraiana Anderson, 1881
Phyllorhinidae de Rochebrune, 1883
Phyllorhina Bonaparte, 1837
Phyllorhina Bonaparte, 1831
Phyllorhina Koch, 1860
Phyllorhina bicornis Heuglin, 1861
Phyllorhina caffa Peters, 1852
Phyllorhina cyclops Temminck, 1853
Phyllorhina fuliginosa Temminck, 1853
Phyllorhina gracilis Peters, 1852
Phyllorhina megalotis Heuglin, 1861
Phyllorhina patellifera Peters, 1852
Phyllorhina vittata Peters, 1852
PHYLLOSTOMATIA Van Valen, 1979

Phyllotis Gray, 1866
Pipistrella Bonaparte, 1837
Pipistrellini Tate, 1842
Pipistrellus Kaup, 1829
Pipistrellus (Pipistrellus) permixtus Aellen, 1957
Pipistrellus (Romicia) kuhlii broomi Roberts, 1948
Pipistrellus abaensis J.A. Allen, 1917
Pipistrellus aero Heller, 1912
Pipistrellus africanus meesteri Kock, 2001
Pipistrellus ariel Thomas, 1904
Pipistrellus cf kuhlii Volleth, Bronner, Göpfert, Heller, von Helversen and Yong, 2001
Pipistrellus crassulus Thomas, 1904
Pipistrellus culex Thomas, 1911
Pipistrellus deserti Thomas, 1902
Pipistrellus eisenrauti Hill, 1968
Pipistrellus eisenrauti bellieri De Vree, 1972
Pipistrellus fouriei Thomas, 1926
Pipistrellus fuscipes Thomas, 1913
Pipistrellus hanaki Hulva and Benda, 2004

Pipistrellus helios Heller, 1912
Pipistrellus inexpectatus Aellen, 1959

Pipistrellus krugeri Herkt, Skidmore and Fahr, 2017
Pipistrellus Kuhlii fuscatus Thomas, 1901

Pipistrellus leucomelas Monard, 1933
Pipistrellus marrensis Thomas and Hinton, 1923
Pipistrellus minusculus Miller, 1900

Current Name

Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris gambiensis (K. Andersen, 1912)
Nycteris major (K. Andersen, 1912)
Nycteris nana (K. Andersen, 1912)
Nycteris parisii (de Beaux, 1924)
NYCTERIDAE Van der Hoeven, 1855
Phasmatonycteris Gunnell, Simmons and Seiffert, 2014
Phasmatonycteris butleri Gunnell, Simmons and Seiffert, 2014
Phasmatonycteris phiomensis Gunnell, Simmons and Seiffert, 2014
PHILISIDAE Sigé, 1985
Scotophilinae Van Cakenberghe and Seamark, 2008
Philisis Sigé, 1985
Philisis sphingis Sigé, 1985
Rhinolophus Lacépède, 1799
HIPPOSIDERIDAE Lydekker, 1891
Hipposideros caffer (Sundevall, 1846)
Macronycteris vittatus (Peters, 1852)

Macronycteris vittatus (Peters, 1852)

Hipposideros ruber (Noack, 1893)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
RHINOLOPHIDAE Gray, 1825
Hipposideros Gray, 1831
Hipposideros Gray, 1831
HIPPOSIDERIDAE Lydekker, 1891
Hipposideros tephros (Cabrera, 1906)
Hipposideros caffer (Sundevall, 1846)
Doryrhina cyclops (Temminck, 1853)
Hipposideros fuliginosus (Temminck, 1853)
Hipposideros caffer (Sundevall, 1846)
Hipposideros megalotis (Heuglin, 1861)
Hipposideros caffer (Sundevall, 1846)
Macronycteris vittatus (Peters, 1852)
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
Rhinolophus Lacépède, 1799
Pipistrellus Kaup, 1829
Pipistrellini Tate, 1942
Pipistrellus Kaup, 1829
Pipistrellus (Pipistrellus) permixtus Aellen, 1957
Pipistrellus (Pipistrellus) hesperidus broomi Roberts, 1948
Afronycteris nana (Peters, 1852)
Pipistrellus (Pipistrellus) aero Heller, 1912
Afronycteris nana (Peters, 1852)
Hypsugo ariel (Thomas, 1904)
*Pipistrellus *(Pipistrellus)* sp. aff. *kuhlii** Kuhl, 1817
Nycticeinops crassulus (Thomas, 1904)
Afronycteris nana (Peters, 1852)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Nycticeinops eisenrauti (Hill, 1968)
Nycticeinops bellieri (De Vree, 1972)
Afronycteris nana (Peters, 1852)
Vansonia rueppellii fuscipes (Thomas, 1913)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Afronycteris helios (Heller, 1912)
Pipistrellus (Pipistrellus) inexpectatus Aellen, 1959
Laephotis stanleyi (Goodman, Kearney, Ratsimbazafy and Hassanin, 2017)
Pipistrellus (Pipistrellus) hesperidus hesperidus (Temminck, 1840)
Vansonia rueppellii vernayi (Roberts, 1932)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Afronycteris nana (Peters, 1852)

Original Name

Pipistrellus musciculus Thomas, 1913
Pipistrellus nanulus Thomas, 1904

Pipistrellus nanus australis Roberts, 1913
Pipistrellus raceyi Bates, Ratrimomanarivo, Harrison and Goodman, 2006

Pipistrellus Rüppelli senegalensis Dorst, 1960
Pipistrellus simandouensis Monadjem, Richards, Decher and Hutterer, 2020

Pipistrellus vernayi Roberts, 1932
Pizonyx Miller, 1906
Platymops Roberts, 1917
Platymops Thomas, 1906
Platymops (Sauromys) Roberts, 1917
Platymops barbatogularis Harrison, 1956
Platymops barbatogularis parkeri Harrison and Fleetwood, 1960
Platymops haagneri Roberts, 1917

Platymops haagneri umbratus Shortridge and Carter, 1938
Platymops Macmillani Thomas, 1906
Platymops petrophilus Roberts, 1917
Platymops petrophilus erongensis Roberts, 1946
Platymops petrophilus fitzsimonsi Roberts, 1946
Plecotina Gray, 1866
Plecotinæ Miller, 1897
Plecotini Gray, 1866
Plecotus E. Geoffroy Saint-Hilaire, 1818
Plecotus ægyptiacus Dobson, 1878
Plecotus æthiopicus Heuglin and Fitzinger, 1866
Plecotus æthiopicus Heuglin, 1866
Plecotus auritus saharae Laurent, 1936
Plecotus austriacus kolombatovici Dulic, 1980
Plecotus balensis Kruskop & Lavrenchenko, 2000
Plecotus cf. gaisleri Ancillotto, Bosso, Smeraldo, Mori, Mazza, Herkt, Galimberti, Ramazzotti and Russo, 2020
Plecotus christii Gray, 1838
Plecotus christii petraeus Benda, 2008
Plecotus teneriffae Barrett-Hamilton, 1907
Plecotus teneriffae gaisleri Benda, Kiefer, Hanák and Veith, 2004
Plecotus ustus Fitzinger and Heuglin, 1866
Plerotes K. Andersen, 1910
Plerotini Bergmans, 1997
Propottinae Butler, 1984
Propotto Simpson, 1967
Propotto leakeyi Simpson, 1967
Provampyrus Schlosser, 1911
Provampyrus orientalis Schlosser, 1911
Pselaphon Gray, 1870
Pseudoromicia Monadjem, Patterson, Webala and Demos, 2020
Pseudoromicia kityoi Monadjem, Kerbis Peterhans, Nalikka, Waswa, Demos and Patterson, 2020
Pseudoromicia nyanza Monadjem, Patterson, Webala and Demos, 2020
Pseudovespertiliavus Ravel, 2016
Pseudovespertiliavus parva Ravel, 2016
Pteropterus Peters, 1868
Pterocarpus Mishra, Rout and Panda, 2011
Pterocynes Haeckel, 1866

Pterocyon Peters, 1862
Pterocyon paleaceus Peters, 1862
Pteropi Winge, 1893

Current Name

Hypsugo musciculus (Thomas, 1913)
Pipistrellus (Pipistrellus) nanulus Thomas, 1904

Afronycteris nana (Peters, 1852)
Pipistrellus (Pipistrellus) raceyi Bates, Ratrimomanarivo, Harrison and Goodman, 2006

Vansonia rueppellii senegalensis (Dorst, 1960)
Pipistrellus (Pipistrellus) simandouensis Monadjem, Richards, Decher and Hutterer, 2020

Vansonia rueppellii vernayi (Roberts, 1932)
Myotis Kaup, 1829
Sauromys Roberts, 1917
Platymops Thomas, 1906
Sauromys Roberts, 1917
Platymops setiger macmillani Thomas, 1906
Platymops setiger macmillani Thomas, 1906

Sauromys petrophilus haagneri (Roberts, 1917)
Sauromys petrophilus umbratus (Shortridge & Carter, 1938)
Platymops setiger macmillani Thomas, 1906
Sauromys petrophilus (Roberts, 1917)
Sauromys petrophilus erongensis (Roberts, 1946)
Sauromys petrophilus umbratus (Shortridge & Carter, 1938)
VESPERTILIONIDAE Gray, 1821
Plecotini Gray, 1866
Plecotini Gray, 1866
Plecotus E. Geoffroy St.-Hilaire, 1818
Plecotus christii Gray, 1838
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818

Plecotus christii Gray, 1838
Otonycteris hemprichii Peters, 1859
Plecotus kolombatovici Dulic, 1980
Plecotus balensis Kruskop & Lavrenchenko, 2000
Plecotus gaisleri Benda, Kiefer, Hanak & Veith, 2004

Plecotus christii Gray, 1838
Plecotus christii Gray, 1838
Plecotus teneriffae Barrett-Hamilton, 1907
Plecotus gaisleri Benda, Kiefer, Hanak & Veith, 2004

Otonycteris hemprichii Peters, 1859
Plerotes K. Andersen, 1910
Plerotini Bergmans, 1997
Propottinae Butler, 1984
Propotto Simpson, 1967
Propotto leakeyi Simpson, 1967
Vampyravus Schlosser, 1910
Vampyravus orientalis Schlosser, 1910
Pteropus Brisson, 1762
Pseudoromicia Monadjem, Patterson, Webala and Demos, 2020
Pseudoromicia kityoi Monadjem, Kerbis Peterhans, Nalikka, Waswa, Demos and Patterson, 2020
Pseudoromicia nyanza Monadjem, Patterson, Webala and Demos, 2020
Pseudovespertiliavus Ravel, 2016
Pseudovespertiliavus parva Ravel, 2016
Myotis Kaup, 1829
Pteropus Brisson, 1762
PTEROPODIFORMI Van Cakenberghe, Kearney and Seamark, 2007
Eidolon Rafinesque, 1815
Eidolon helvum helvum (Kerr, 1792)
PTEROPODIDAE Gray, 1821

Original Name

Pteropi Peters, 1866
Pteropidae Gray, 1821
Pteropina Gray, 1825
Pteropinae Trouessart, 1897
Pteropodae Bonaparte, 1835
Pteropodida Haeckel, 1866
PTEROPODIDAE Bonaparte, 1838
PTEROPODIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
PTEROPODIFORMES Hutcheon and Kirsch, 2004
PTEROPODIFORMI Van Cakenberghe, Kearney and Seamark, 2007
Pteropodimorpha Zagorodniuk, 2014

Pteropodina Bonaparte, 1831
Pteropodinae Gray, 1821
Pteropodini Gray, 1821
PTEROPODOIDEA Gray, 1821
Pteropoidea Serra-Cobo, López-Roig, Bayer, Amengual and Guasch, 2008
Pteropus Brisson, 1762
Pteropus (Spectrum) voeltzkowi Matschie, 1909
Pteropus aegyptiacus E. Geoffroy Saint-Hilaire, 1818
Pteropus aegyptiacus, 9999

Pteropus aldabrensis True, 1893
Pteropus collaris Lichtenstein, 1823

Pteropus collaris Illiger, 1815
Pteropus comorensis Wallace, 1880

Pteropus comorensis Nicoll, 1908

Pteropus dupréanus Schlegel, 1867
Pteropus Edwardsii E. Geoffroy Saint-Hilaire, 1810
Pteropus Egyptiacus E. Geoffroy Saint-Hilaire, 1810
Pteropus epomophorus Bennett, 1836

Pteropus flavus Illiger, 1815
Pteropus fuscus E. Geoffroy Saint-Hilaire, 1803
Pteropus Gambianus Ogilby, 1835
Pteropus Gambianus Ogilby, 1835

Pteropus geoffroyi Temminck, 1825

Pteropus Haldemani Hallowell, 1846
Pteropus hottentottus Temminck, 1832

Pteropus labiatus Temminck, 1837
Pteropus Leachii A. Smith, 1829

Pteropus livingstonii Gray, 1866
Pteropus macrocephalus Ogilby, 1835

Pteropus madagascariensis Oken, 1816
Pteropus mascarinus Mason, 1907
Pteropus megacephalus Swainson, 1835

Pteropus mollipilosus H. Allen, 1862
Pteropus palmarum Heuglin, 1865
Pteropus phæops Oken, 1838
Pteropus phaiops Temminck, 1825
Pteropus pteropus Merriam, 1895
Pteropus rodricensis Dobson, 1878
Pteropus ruber E. Geoffroy Saint-Hilaire, 1803
Pteropus rubicollis E. Geoffroy Saint-Hilaire, 1810
Pteropus rufus E. Geoffroy Saint-Hilaire, 1803

Current Name

PTEROPODIDAE Gray, 1821
PTEROPODIDAE Gray, 1821
PTEROPODIDAE Gray, 1821
Pteropodinae Gray, 1821
Pteropodinae Gray, 1821
PTEROPODIDAE Gray, 1821
PTEROPODIDAE Gray, 1821
PTEROPODIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
PTEROPODIFORMI Van Cakenberghe, Kearney and Seamark, 2007
PTEROPODIFORMI Van Cakenberghe, Kearney and Seamark, 2007
PTEROPODIFORMI Van Cakenberghe, Kearney and Seamark, 2007
Pteropodinae Gray, 1821
Pteropodinae Gray, 1821
Pteropodini Gray, 1821
PTEROPODOIDEA Gray, 1821
PTEROPODOIDEA Gray, 1821

Pteropus Brisson, 1762
Pteropus voeltzkowi Matschie, 1909

Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pteropus aldabrensis True, 1893
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pteropus subniger (Kerr, 1792)
Pteropus seychellensis A. Milne-Edwards, 1877
Pteropus seychellensis A. Milne-Edwards, 1877
Eidolon dupreanum (Schegel, 1867)
Pteropus rufus E. Geoffroy St.-Hilaire, 1803

Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Epomophorus gambianus gambianus (Ogilby, 1835)
Eidolon helvum helvum (Kerr, 1792)
Pteropus niger (Kerr, 1792)

Epomophorus gambianus (Ogilby, 1835)
Epomophorus gambianus gambianus (Ogilby, 1835)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Epomophorus wahlbergi (Sundevall, 1846)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Epomophorus labiatus (Temminck, 1837)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pteropus livingstonii Gray, 1866
Epomophorus gambianus gambianus (Ogilby, 1835)
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Pteropus rodricensis Dobson, 1878
Epomophorus gambianus gambianus (Ogilby, 1835)
Eidolon helvum helvum (Kerr, 1792)
Eidolon helvum helvum (Kerr, 1792)
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Pteropus niger (Kerr, 1792)
Pteropus rodricensis Dobson, 1878
Pteropus subniger (Kerr, 1792)
Pteropus subniger (Kerr, 1792)

Pteropus rufus E. Geoffroy St.-Hilaire, 1803

Original Name

Pteropus rufus princeps K. Andersen, 1908
Pteropus Schoënsii Rüppell, 1842
Pteropus seychellensis A. Milne-Edwards, 1877
Pteropus stramineus E. Geoffroy Saint-Hilaire, 1803
Pteropus torquatus G. Fischer, 1814
Pteropus vulgaris Temminck, 1837
Pteropus vulgaris E. Geoffroy Saint-Hilaire, 1810
Pteropus Wahlbergi Sundevall, 1846
Pteropus Whitei Bennett, 1836

Pterygistes Kaup, 1829
Pterygistes azureum Thomas, 1901
Pterygistes madeiræ Barrett-Hamilton, 1906
Ptética Ameghino, 1889
Ptychorhina Peters, 1871
Qarunycteris Gunnell, Simons and Seiffert, 2008
Qarunycteris moerisae Gunnell, Simons and Seiffert, 2008
R[hinopoma] cystops macinnesi Hayman, 1937
R[oussettus] pattersoni Gunnell and Manthi, 2018
Rhinocrepis Gervais, 1836
Rhinolophi Peters, 1865
Rhinolophidae Gray, 1825
Rhinolophides Gervais, 1854
RHINOLOPHIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
Rhinolophina Gray, 1825
Rhinolophina Gray, 1825
Rhinolophina Lesson, 1827
Rhinolophini Winge, 1893
Rhinolophoidea Gray, 1825
Rhinolophus Lacépède, 1799
Rhinolophus abæ J.A. Allen, 1917
Rhinolophus acrotis Heuglin, 1861
Rhinolophus acrotis brachygnatus K. Andersen, 1905
Rhinolophus acrotis schwarzi Heim de Balsac, 1934
Rhinolophus adami Aellen and Brosset, 1968
Rhinolophus æthiops Peters, 1869
Rhinolophus æthiops diversus Sanborn, 1939
Rhinolophus alcyone Temminck, 1853
Rhinolophus alcyone alticolus Sanborn, 1936
Rhinolophus algirus Loche, 1867
Rhinolophus Andersoni Thomas, 1904
Rhinolophus andreinii Senna, 1905
Rhinolophus angolensis Seabra, 1898
Rhinolophus antinorii Dobson, 1885
Rhinolophus augur K. Andersen, 1904
Rhinolophus augur zambesiensis K. Andersen, 1904
Rhinolophus augur zuluensis K. Andersen, 1904
Rhinolophus auritus Sundevall, 1860
Rhinolophus axillaris Allen, 1917
Rhinolophus bembanicus Senna, 1914
Rhinolophus blasii Peters, 1867
Rhinolophus blasiusi Trouessart, 1910
Rhinolophus brockmani Thomas, 1910
Rhinolophus caffer Sundevall, 1846
Rhinolophus capensis Lichtenstein, 1823
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus clivosus hillorum Koopman, 1989
Rhinolophus cohenae Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Rhinolophus Commersonii E. Geoffroy Saint-Hilaire, 1813

Current Name

Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Epomophorus labiatus (Temminck, 1837)
Pteropus seychellensis A. Milne-Edwards, 1877
Eidolon helvum helvum (Kerr, 1792)

Pteropus subniger (Kerr, 1792)
Pteropus subniger (Kerr, 1792)
Pteropus niger (Kerr, 1792)

Epomophorus wahlbergi (Sundevall, 1846)
Epomophorus gambianus gambianus (Ogilby, 1835)
Nyctalus Bowdich, 1825
Nyctalus azureum (Thomas, 1901)
Nyctalus leisleri (Kuhl, 1817)
CHIROPTERA Blumenbach, 1779
Hipposideros Gray, 1831
Qarunycteris Gunnell, Simons and Seiffert, 2008
Qarunycteris moerisae Gunnell, Simons and Seiffert, 2008
Rhinopoma macinnesi Hayman, 1937

Roussettus pattersoni Gunnell and Manthi, 2018

Rhinolophus Lacépède, 1799
RHINOLOPHIDAE Gray, 1825
RHINOLOPHIDAE Gray, 1825
RHINOLOPHIDAE Gray, 1825
RHINOLOPHIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
RHINOLOPHIDAE Gray, 1825
RHINOLOPHOIDEA J. E. Gray, 1825
RHINOLOPHIDAE Gray, 1825
RHINOLOPHIDAE Gray, 1825
RHINOLOPHOIDEA J. E. Gray, 1825
Rhinolophus Lacépède, 1799
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus clivosus Cretzschmar, 1828

Rhinolophus clivosus Cretzschmar, 1828

Rhinolophus adami Aellen and Brosset, 1968
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus alcyone Temminck, 1853
Rhinolophus simulator K. Andersen, 1904
Rhinolophus euryale Blasius, 1853
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus blasii Peters, 1867
Rhinolophus lobatus Peters, 1852
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus clivosus Cretzschmar, 1828

Rhinolophus clivosus Cretzschmar, 1828

Rhinolophus capensis Lichtenstein, 1823
Rhinolophus landeri Martin, 1838
Rhinolophus simulator K. Andersen, 1904
Rhinolophus blasii Peters, 1867
Rhinolophus blasii Peters, 1867
Rhinolophus blasii Peters, 1867
Hipposideros caffer (Sundevall, 1846)
Rhinolophus capensis Lichtenstein, 1823
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus hillorum Koopman, 1989
Rhinolophus cohenae Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Macronycteris commersoni (E. Geoffroy St.-Hilaire, 1813)

Original Name

Rhinolophus Darlingi K. Andersen, 1905
Rhinolophus darlingi barbertonensis Roberts, 1924
Rhinolophus darlingi damarensis Roberts, 1946
Rhinolophus Deckenii Peters, 1868
Rhinolophus Denti Thomas, 1904
Rhinolophus denti knorri Eisentraut, 1960
Rhinolophus Dobsoni Thomas, 1904
Rhinolophus Eggenhöffner Fitzinger, 1870
Rhinolophus empusa K. Andersen, 1904
Rhinolophus euryale Blasius, 1853
Rhinolophus euryale tuneti Deleuil and Labbé, 1955
Rhinolophus ferrumequinum mellali Lavocat, 1961
Rhinolophus foxi Thomas, 1913
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus fumigatus exsul K. Andersen, 1905
Rhinolophus Geoffroyii A. Smith, 1829
Rhinolophus gigas Wagner, 1845
Rhinolophus gorongosae Taylor, MacDonald, Goodman, Kearney, Cotterill, Stoffberg, Monadjem, Schoeman, Guyton, Naskrecki and Richards, 2018
Rhinolophus Hildebrandti eloquens K. Andersen, 1905
Rhinolophus hildebrandti perauritus de Beaux, 1922
Rhinolophus Hildebrandtii Peters, 1878
Rhinolophus hilli Aellen, 1973
Rhinolophus hipposideros vespa Laurent, 1937
Rhinolophus hipposiderus typicus K. Andersen, 1905
Rhinolophus horaceki Benda and Vallo, 2012
Rhinolophus kahuzi Fahr and Kerbis Peterhans, 2013
Rhinolophus keniensis Hollister, 1916
Rhinolophus landeri Martin, 1838
Rhinolophus landeri guineensis Eisentraut, 1960
Rhinolophus lobatus Peters, 1852
Rhinolophus mabuensis Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Rhinolophus Maclaudi Pousargues, 1898
Rhinolophus macrocephalus Heuglin, 1877
Rhinolophus maendeleo Kock, Csorba and Howell, 2000
Rhinolophus maghrebensis Gunnell, Eiting and Geraads, 2011
Rhinolophus Martini Fraser, 1843
Rhinolophus mehelyi Matschie, 1901
Rhinolophus micaceus de Winton, 1897
Rhinolophus mimus Heuglin, 1861
Rhinolophus mossambicus Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Rhinolophus ruwenzorii J. Eric Hill, 1942
Rhinolophus sakeiensis Cotterill, 2002
Rhinolophus silvestris Aellen, 1959
Rhinolophus simulator K. Andersen, 1904
Rhinolophus smithersi Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Rhinolophus swinnyi Gough, 1908
Rhinolophus swinnyi piriensis Hewitt, 1913
Rhinolophus swinnyi rhodesiae Roberts, 1946
Rhinolophus tridens E. Geoffroy Saint-Hilaire, 1813
Rhinolophus willardi Kerbis Peterhans and Fahr, 2013

Current Name

Rhinolophus darlingi K. Andersen, 1905
Rhinolophus darlingi K. Andersen, 1905
Rhinolophus damarensis Roberts, 1946
Rhinolophus deckenii Peters, 1868
Rhinolophus denti Thomas, 1904
Rhinolophus denti Thomas, 1904
Rhinolophus landeri Martin, 1838
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus blasii Peters, 1867
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Rhinolophus mellali Lavocat, 1961
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus clivosus Cretzschmar, 1828
Macronycteris gigas (Wagner, 1845)
Rhinolophus gorongosae Taylor, MacDonald, Goodman, Kearney, Cotterill, Stoffberg, Monadjem, Schoeman, Guyton, Naskrecki and Richards, 2018
Rhinolophus eloquens K. Andersen, 1905
Rhinolophus eloquens K. Andersen, 1905
Rhinolophus hildebrandtii Peters, 1878
Rhinolophus hilli Aellen, 1973
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus horaceki Benda and Vallo, 2012
Rhinolophus kahuzi Fahr and Kerbis Peterhans, 2013
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus landeri Martin, 1838
Rhinolophus guineensis Eisentraut, 1960
Rhinolophus lobatus Peters, 1852
Rhinolophus mabuensis Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Rhinolophus maclaudi Pousargues, 1898
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus maendeleo Kock, Csorba and Howell, 2000
Rhinolophus maghrebensis Gunnell, Eiting and Geraads, 2011
Nycteris hispida (Schreber, 1774)
Rhinolophus mehelyi Matschie, 1901
Doryrhina cyclops (Temminck, 1853)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus mossambicus Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Rhinolophus ruwenzorii J. Eric Hill, 1942
Rhinolophus sakeiensis Cotterill, 2002
Rhinolophus silvestris Aellen, 1959
Rhinolophus simulator K. Andersen, 1904
Rhinolophus smithersi Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Rhinolophus swinnyi Gough, 1908
Rhinolophus swinnyi Gough, 1908
Rhinolophus rhodesiae Roberts, 1946
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Rhinolophus willardi Kerbis Peterhans and Fahr, 2013

Original Name

Rhinolophus ziama Fahr, Vierhaus, Hutterer and Kock, 2002
Rhinomegalophus Bourret, 1951
Rhinonycteridae, 9999
Rhinonycteridae Gray, 1866
Rhinonycterina Gray, 1866
Rhinophylla Gray, 1866
Rhinophyllotis Troughton, 1941
Rhinopoma E. Geoffroy Saint-Hilaire, 1818
Rhinopoma brevicaudatum Oken, 1816
Rhinopoma cordofanicum Heuglin, 1877
Rhinopoma cystops Thomas, 1903
Rhinopoma hardwickei sennaariense Kock, 1969
Rhinopoma Lepsianum Peters, 1859
Rhinopoma longicaudatum Fitzinger, 1866
Rhinopoma microphyllum tropicalis Kock, 1969
Rhinopoma sennaariense Fitzinger, 1866
Rhinopoma sennarens Hartmann, 1868
Rhinopomatidae Dobson, 1872
Rhinopomatoidea Dobson, 1872
Rhinopomidae Miller, 1907
Rhinopomina Bonaparte, 1838
Rhinopterus Miller, 1906
Rhinopterus lowei Thomas, 1915

Rhizomops Legendre, 1984
Rhyneptesicus Bianchi, 1917
Rhynopoma Bowdich, 1821
Rickettia Bianchi, 1917
Romicia Gray, 1838
Romiciana Gray, 1866
Romicus Blyth, 1840
Rousettina Koopman and J.K. Jones Jr., 1970
Rousettinae K. Andersen, 1912
Rousettini K. Andersen, 1912
Rousettus Gray, 1821
Rousettus (Lissonycteris) crypticola Cabrera, 1920
Rousettus (Rousettus) obliviosus Kock, 1978
Rousettus aegyptiacus occidentalis Eisentraut, 1960
Rousettus aegyptiacus princeps Juste and Ibáñez, 1993
Rousettus aegyptiacus thomensis Feiler, Haft and Widmann, 1993
Rousettus aegyptiacus tomensis Juste and Ibáñez, 1993
Rousettus angolensis ruwenzorii Eisentraut, 1965
Rousettus kemp Thomas, 1909
Rousettus lanosus Thomas, 1906
Rousettus madagascariensis G. Grandidier, 1929
Rousettus sjöstedti Lönnberg, 1908

Rousettus smithii Thomas, 1908
Rousettus sp. cf. R. madagascariensis

rueppellii Qumsiyeh, 1985
Saccolaimus Temminck, 1838
Saccolaimus incognita Butler and Hopwood, 1957
Saccolaimus kenyensis Gunnell and Manthi, 2018
Saharaderma Gunnell, Simons and Seiffert, 2008
Saharaderma pseudovampyrus Gunnell, Simons and Seiffert, 2008
Scabrifer G.M. Allen, 1908
Scabrifer notius G.M. Allen, 1908
Scoteinus schlieffeni albiventer Thomas and Wroughton, 1908

Current Name

Rhinolophus ziama Fahr, Vierhaus, Hutterer and Kock, 2002
Rhinolophus Lacépède, 1799
RHINONYCTERIDAE J. E. Gray, 1866
RHINONYCTERIDAE J. E. Gray, 1866
RHINONYCTERIDAE J. E. Gray, 1866
Hipposideros Gray, 1831
Rhinolophus Lacépède, 1799
Rhinopoma E. Geoffroy St.-Hilaire, 1818
Rhinopoma cystops Thomas, 1903
Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma cystops Thomas, 1903
Rhinopoma cystops Thomas, 1903

Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma cystops Thomas, 1903
Rhinopoma microphyllum (Brünnich, 1782)

Rhinopoma cystops Thomas, 1903
Rhinopoma cystops Thomas, 1903
RHINOPOMATIDAE Dobson, 1872
RHINOPOMATOIDEA Dobson, 1872
RHINOPOMATIDAE Dobson, 1872
RHINOPOMATIDAE Dobson, 1872
Eptesicus (Rhinopterus) Miller, 1906
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Tadarida Rafinesque, 1814
Eptesicus Rafinesque, 1820
Rhinopoma E. Geoffroy St.-Hilaire, 1818
Myotis Kaup, 1829
Pipistrellus Kaup, 1829
VESPERTILIONIDAE Gray, 1821
Pipistrellus Kaup, 1829
Rousettinae Andersen, 1912
Rousettinae Andersen, 1912
Rousettini Andersen, 1912
Rousettus Gray, 1821
Myonycteris angolensis (Bocage, 1898)

Rousettus obliviosus Kock, 1978
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Myonycteris angolensis ruwenzorii (Eisentraut, 1965)
Stenonycteris lanosus (Thomas, 1906)
Stenonycteris lanosus (Thomas, 1906)
Rousettus madagascariensis G. Grandidier, 1929
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Myonycteris angolensis smithii (Thomas, 1908)
Rousettus madagascariensis G. Grandidier, 1929
Tadarida teniotis (Rafinesque, 1814)
Saccolaimus Temminck, 1838
Taphozous incognita (Butler & Hopwood, 1957)

Saccolaimus kenyensis Gunnell and Manthi, 2018
Saharaderma Gunnell, Simons and Seiffert, 2008
Saharaderma pseudovampyrus Gunnell, Simons and Seiffert, 2008
Laephotis Thomas, 1901
Laephotis capensis (A. Smith, 1829)
Nycticeinops schlieffenii (Peters, 1859)

Original Name

Scoteinus schlieffeni australis Thomas and Wroughton, 1908
Scoteinus schlieffeni bedouin Thomas and Wroughton, 1908
Scoteinus schlieffenii fitzsimonsi Roberts, 1932
Scotoecus Thomas, 1901
Scotoecus albigula Thomas, 1909
Scotoecus artinii de Beaux, 1923
Scotoecus cinnamomeus Wettstein, 1916
Scotoecus falabæ Thomas, 1915
Scotoecus Hindei Thomas, 1901
Scotoecus olduvensis Gunnell, Butler, Greenwood and Simmons, 2015
Scotoecus woodi Thomas, 1917
Scotonycterini Bergmans, 1997
Scotonycteris Matschie, 1894
Scotonycteris bedfordi Thomas, 1904
Scotonycteris bergmansi Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015
Scotonycteris bergmansi bergmansi Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015
Scotonycteris bergmansi congoensis Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015
Scotonycteris ophiodon Pohle, 1943
Scotonycteris ophiodon cansdalei Hayman, 1946
Scotonycteris zenkeri Matschie, 1894
Scotonycteris zenkeri occidentalis Hayman, 1947
Scotophilini A. Murray, 1866

Scotophilina Gray, 1866

Scotophilinae Van Cakenberghe and Seamark, 2008
Scotophilini Hill and Harrison, 1987

Scotophilis Horáček, Fejfar and Hulva, 2006
Scotophilis libycus Horáček, Fejfar and Hulva, 2006
Scotophilus Leach, 1821
Scotophilus albofuscus Thomas, 1890
Scotophilus altilis G.M. Allen, 1914
Scotophilus alvenslebeni Dalquest, 1965
Scotophilus andrewreborii Brooks and Bickham, 2014
Scotophilus angusticeps Shortridge and Carter, 1938
Scotophilus cf. viridis, 9999
Scotophilus damarensis Thomas, 1906
Scotophilus darwini Tomes, 1859
Scotophilus ejetai Brooks and Bickham, 2014
Scotophilus gigas Dobson, 1875
Scotophilus hirundo de Winton, 1899
Scotophilus livingstonii Brooks and Bickham, 2014
Scotophilus marovaza Goodman, Ratrimomanarivo and Randrianandrianina, 2006
Scotophilus minimus Noack, 1887
Scotophilus nigrita colias Thomas, 1904
Scotophilus nigrita herero Thomas, 1906
Scotophilus nigrita nux Thomas, 1904
Scotophilus nigrita pondoensis Roberts, 1946
Scotophilus nigritellus de Winton, 1899
Scotophilus nucella Robbins, 1983
Scotophilus robustus A. Milne-Edwards, 1881
Scotophilus rusticus Tomes, 1861

Current Name

Nycticeinops schlieffenii (Peters, 1859)
Nycticeinops schlieffenii (Peters, 1859)
Nycticeinops schlieffenii (Peters, 1859)

Scotoecus Thomas, 1901
Scotoecus albigula Thomas, 1909
Scotoecus hirundo (de Winton, 1899)
Nycticeinops schlieffenii (Peters, 1859)
Scotoecus hindei Thomas, 1901
Scotoecus hindei Thomas, 1901
Scotoecus olduvensis Gunnell, Butler, Greenwood and Simmons, 2015
Scotoecus albofuscus (Thomas, 1890)
Scotonycterini Bergmans, 1997
Scotonycteris Matschie, 1894
Scotonycteris zenkeri Matschie, 1894
Scotonycteris bergmansi Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015
Scotonycteris bergmansi bergmansi Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015
Scotonycteris bergmansi congoensis Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015

Casinycteris ophiodon (Pohle, 1943)
Casinycteris ophiodon (Pohle, 1943)

Scotonycteris zenkeri Matschie, 1894
Scotonycteris occidentalis Hayman, 1947

Scotophilinae Van Cakenberghe and Seamark, 2008
Scotophilinae Van Cakenberghe and Seamark, 2008
Scotophilinae Van Cakenberghe and Seamark, 2008
Scotophilinae Van Cakenberghe and Seamark, 2008
Scotophilis Horáček, Fejfar and Hulva, 2006
Scotophilis libycus Horáček, Fejfar and Hulva, 2006
Scotophilus Leach, 1821
Scotoecus albofuscus (Thomas, 1890)
Scotophilus altilis G.M. Allen, 1914
Scotophilus nigrita (Schreber, 1774)
Scotophilus andrewreborii Brooks and Bickham, 2014
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Scotophilus viridis (Peters, 1852)
Scotophilus leucogaster (Cretzschmar, 1826)
Hypsugo savii (Bonaparte, 1837)
Scotophilus ejetai Brooks and Bickham, 2014
Scotophilus nigrita (Schreber, 1774)
Scotoecus hirundo (de Winton, 1899)
Scotophilus livingstonii Brooks and Bickham, 2014
Scotophilus marovaza Goodman, Ratrimomanarivo and Randrianandrianina, 2006
Nycticeinops schlieffenii (Peters, 1859)
Scotophilus dinganii (A. Smith, 1833)
Scotophilus dinganii (A. Smith, 1833)
Scotophilus nux Thomas, 1904
Scotophilus dinganii (A. Smith, 1833)
Scotophilus nigritellus de Winton, 1899
Scotophilus nucella Robbins, 1983
Scotophilus robustus A. Milne-Edwards, 1881
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)

Original Name

Scotophilus tandrefana Goodman, Jenkins and Ratrimomanarivo, 2005
Scotophilus trujilloi Brooks and Bickham, 2014
Scotophilus variegatus Tomes, 1861
Scotophilus welwitschii Gray, 1866
Selysius Bonaparte, 1841
Senonycteris Gray, 1870
Sericonycteris Matschie, 1899
Sideroderma Peters, 1871
Spasma Gray, 1866
Spectrum Lacépède, 1799
Spectrum rubidum Daudin, 1802
Spectrum vulgare Gray, 1870
Speorifera Gray, 1866
Sphyrocephalus A. Murray, 1862
Sphyrocephalus labrosus A. Murray, 1862
Stenonycterini Nesi, Kadjo, Pourrut, Leroy, Pongombo Shongo, Cruaud and Hassanin, 2012
Stenonycteris Gray, 1870
Sybdesmotus Peters, 1871
Syndesmotis Peters, 1871
Syndesmotus C.O. Waterhouse, 1902
Synotus Keyserling and Blasius, 1839
Tadarida Rafinesque, 1814
Tadarida (Chaerephon) cyclotis Brosset, 1966

Tadarida (Chaerephon) faini Hayman, 1951

Tadarida (Chaerephon) gallagheri Harrison, 1975
Tadarida (Chaerephon) tomensis Juste and Ibáñez, 1993
Tadarida [(Xiphonycteris)] petersoni El Rayah, 1981
Tadarida engesseri Rachl, 1983
Tadarida rusingae Arroyo-Cabrales, Gregorin, Schlitter and Walker, 2002
Tanzanycterididae Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003
Tanzanycteris Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003
Tanzanycteris mannardi Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003
Taphonycteris Dobson, 1876
Taphozoinae Jerdon, 1867
Taphozoini Robbins and Sarich, 1988
Taphozous E. Geoffroy Saint-Hilaire, 1818
Taphozous abitus Wesselman, 1984
Taphozous Dobsoni Jentink, 1879

Taphozous hamiltoni Thomas, 1920
Taphozous hildegardeae Thomas, 1909
Taphozous leucopterus Temminck, 1835

Taphozous maritimus Heuglin, 1877

Taphozous mauritanus E. Geoffroy Saint-Hilaire, 1818
Taphozous nudiventris Cretzschmar, 1830-1831
Taphozous peli Temminck, 1853
Taphozous perforatus E. Geoffroy Saint-Hilaire, 1818
Taphozous perforatus E. Geoffroy Saint-Hilaire, 1818
Taphozous perforatus hædinus Thomas, 1915

Current Name

Scotophilus tandrefana Goodman, Jenkins and Ratrimomanarivo, 2005
Scotophilus trujilloi Brooks and Bickham, 2014
Glauconycteris variegata (Tomes, 1861)
Myotis welwitschii (Gray, 1866)
Myotis Kaup, 1829
Rousettus Gray, 1821
Pteropus Brisson, 1762
Hipposideros Gray, 1831
Megaderma E. Geoffroy St.-Hilaire, 1810
Pteropus Brisson, 1762
Pteropus subniger (Kerr, 1792)
Pteropus niger (Kerr, 1792)
Hipposideros Gray, 1831
Hypsignathus H. Allen, 1862
Hypsignathus monstrosus H. Allen, 1862
Stenonycterini Nesi, Kadjo, Pourrut, Leroy, Pongombo Shongo, Cruaud and Hassanin, 2012
Stenonycteris Gray, 1870
Hipposideros Gray, 1831
Hipposideros Gray, 1831
Hipposideros Gray, 1831
Barbastella Gray, 1821
Tadarida Rafinesque, 1814
Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Mops (Chaerephon) gallagheri (Harrison, 1975)

Mops (Chaerephon) tomensis (Juste and Ibáñez, 1993)
Mops (Xiphonycteris) petersoni (El Rayah, 1981)
Tadarida engesseri Rachl, 1983
Mops rusingae (Arroyo-Cabrales, Gregorin, Schlitter and Walker, 2002)
TANZANYCTERIDIDAE Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003
Tanzanycteris Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003
Tanzanycteris mannardi Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003
Saccolaimus Temminck, 1838
Taphozoinae Jerdon, 1867
Taphozoinae Jerdon, 1867
Taphozous E. Geoffroy St.-Hilaire, 1818
Saccolaimus abitus (Wesselman, 1984)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Taphozous hamiltoni Thomas, 1920
Taphozous hildegardeae Thomas, 1909
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Taphozous nudiventris Cretzschmar, 1830

Saccolaimus peli (Temminck, 1853)
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus perforatus E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818

Original Name

Taphozous perforatus swirae Harrison, 1958
Taphozous perforatus var. *assabensis* Monticelli, 1885
Taphozous senegalensis Desmarest, 1820
Taphozous *Sudani* Thomas, 1915
Taphozous sudani australis Harrison, 1962
Taphozous sudani rhodesiae Harrison, 1964
Thyreorhina Peters, 1871
Triaenopini Benda and Vallo, 2009
Triaenops Dobson, 1871
Triaenops afer Peters, 1877
Triaenops aurita G. Grandidier, 1912
Triaenops furcula Trouessart, 1907
Triaenops goodmani Samonds, 2007
Triaenops Humbloti A. Milne-Edwards, 1881
Triaenops menamena Goodman and Ranivo, 2009
Triaenops pauliani Goodman and Ranivo, 2008
Triaenops persicus majusculus Aellen and Brosset, 1968
Triaenops rufus A. Milne-Edwards, 1881
Trilatitus Gray, 1842
Trygenycteris Lydekker, 1891
Tuitatus Kishida and Mori, 1931
Turkanycteris Gunnell and Manthi, 2018
Turkanycteris harrisi Gunnell and Manthi, 2018
V[espertilio noctula] var. *sicula* Mina-Palumbo, 1868
V[espertilio] acetabulosus Hermann, 1804
V[espertilio] acetabulosus Hermann, 1804
V[espertilio] mauritanus Hermann, 1804
V[espertilio] Rüppelii J.B. Fischer, 1829
V[espertilio] Smithii Wagner, 1855
V[espertilio] tricolor Temminck, 1832
Vampyravus Schlosser, 1910
Vampyravus orientalis Schlosser, 1910
Vansonia Roberts, 1946
Vansonia Roberts, 1946
Vespertilia Rafinesque, 1815
Vespertiliavus Schlosser, 1887
VESPERTILIFORMES Zagorodniuk, 1997
VESPERTILIIFORMES Zagorodniuk, Godovanets, Pokynchereda and Kyseliuk, 1995
Vespertilio Gray, 1843
Vespertilio (Pteropterus) Peters, 1867
Vespertilio barbastellus Schreber, 1774
Vespertilio Bocagii Peters, 1870
Vespertilio Bocagii Peters, 1870
Vespertilio borbonicus E. Geoffroy Saint-Hilaire, 1803
Vespertilio Boscai Cabrera, 1904
Vespertilio brevicauda Stresemann, 1954
Vespertilio caninus Blumenbach, 1797
Vespertilio caninus var. *b* Goldfuss, 1809
Vespertilio Capaccinii Bonaparte, 1837
Vespertilio Capensis A. Smith, 1829
Vespertilio capensis gracilior Thomas and Schwann, 1905
Vespertilio dasykarpos Kuhl, 1819
Vespertilio d'asythrix Temminck, 1840
Vespertilio Dinganii A. Smith, 1833
Vespertilio emarginatus E. Geoffroy Saint-Hilaire, 1806

Current Name

Taphozous perforatus perforatus E. Geoffroy St.-Hilaire, 1818
Taphozous nudiventris Cretzschmar, 1830
Taphozous perforatus perforatus E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus sudani Thomas, 1915
Taphozous perforatus sudani Thomas, 1915
Taphozous perforatus sudani Thomas, 1915
Hipposideros Gray, 1831
RHINONYCTERIDAE J. E. Gray, 1866
Triaenops Dobson, 1871
Triaenops afer Peters, 1877
Paratriaenops auritus (G. Grandidier, 1912)
Paratriaenops furculus (Trouessart, 1907)
Triaenops goodmani Samonds, 2007
Triaenops menamena Goodman and Ranivo, 2009
Triaenops menamena Goodman and Ranivo, 2009
Paratriaenops pauliani (Goodman and Ranivo, 2008)
Triaenops afer Peters, 1877
Triaenops menamena Goodman and Ranivo, 2009
Myotis Kaup, 1829
Megaloglossus Pagenstecher, 1885
Eptesicus Rafinesque, 1820
Turkanycteris Gunnell and Manthi, 2018
Turkanycteris harrisi Gunnell and Manthi, 2018
Nyctalus lasiopterus (Schreber, 1780)
Mormopterus acetabulosus (Hermann, 1804)
Mormopterus acetabulosus acetabulosus (Hermann, 1804)
Pteropus niger (Kerr, 1792)
Vansonia rueppelii (J.B. Fischer, 1829)
Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
Myotis tricolor (Temminck, 1832)
Vampyravus Schlosser, 1910
Vampyravus orientalis Schlosser, 1910
Vansonia Roberts, 1946
Pipistrellus Kaup, 1829
VESPERTILIONIDAE Gray, 1821
Vespertiliavus Schlosser, 1887
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
Myotis Kaup, 1829
Myotis Kaup, 1829
Barbastella barbastellus (Schreber, 1774)
Myotis bocagii (Peters, 1870)
Myotis bocagii bocagii (Peters, 1870)
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Rhinopoma cystops Thomas, 1903
Pteropus niger (Kerr, 1792)
Eidolon helvum helvum (Kerr, 1792)
Myotis capaccinii (Bonaparte, 1837)
Laephotis capensis (A. Smith, 1829)
Laephotis capensis (A. Smith, 1829)
Nyctalus leisleri (Kuhl, 1817)
Miniopterus natalensis (A. Smith, 1833)
Scotophilus dinganii (A. Smith, 1833)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)

Original Name

Vespertilio epichrysus Temminck, 1832
Vespertilio ferox Stresemann, 1954
Vespertilio ferrugineus Brehm, 1827
Vespertilio ferrum-equinum Schreber, 1774
Vespertilio ferrum-equinum major Kerr, 1792
Vespertilio goudoti A. Smith, 1834
Vespertilio hesperida Temminck, 1840

Vespertilio hesperida Temminck, 1840

Vespertilio hipposideros Bechstein, 1800
Vespertilio hispidus Schreber, 1774
Vespertilio hottentota A. Smith, 1833

Vespertilio isabellinus Temminck, 1835-1841

Vespertilio Kuhlii Kuhl, 1817
Vespertilio lanosus A. Smith, 1847
Vespertilio lasiopterus Schreber, 1780
Vespertilio Leisleri Kuhl, 1817
Vespertilio leucomelas Cretzschmar, 1826
Vespertilio Madagascariensis Tomes, 1858
Vespertilio marginatus Cretzschmar, 1830
Vespertilio matroka Thomas and Schwann, 1905
Vespertilio megalotis Bechstein, 1800
Vespertilio megalurus Temminck, 1835-1841

Vespertilio Microphyllum Brünnich, 1782
Vespertilio minuta Temminck, 1840
Vespertilio minutus A. Smith, 1849

Vespertilio minutus somalicus Thomas, 1901
Vespertilio murinus africanus Dobson, 1875
Vespertilio mystacinus Kuhl, 1817
Vespertilio nanus Peters, 1852
Vespertilio Natalensis A. Smith, 1833
Vespertilio Nigrita Schreber, 1774
Vespertilio ochromixtus Cabrera, 1904
Vespertilio pachygnathus Michahelles, 1839
Vespertilio pipistrellus Schreber, 1774

Vespertilio Pipistrellus varietas africana
 Rüppell, 1842
Vespertilio platycephalus Temminck, 1832

Vespertilio platyops Thomas, 1901

Vespertilio pusillus Leconte, 1857
Vespertilio Pygmæus Leach, 1825

Vespertilio ruppellii J.B. Fischer, 1829

Vespertilio Savii Bonaparte, 1837
Vespertilio schreibersii Kuhl, 1817
Vespertilio scotinus Sundevall, 1846
Vespertilio smithii Wagner, 1855

Vespertilio subtilis Sundevall, 1846

Vespertilio sylvicola A. Grandidier, 1870
Vespertilio temminckii Cretzschmar, 1827

Vespertilio temminckii Cretzschmar, 1826
Vespertilio vampirus niger Kerr, 1792
Vespertilio vampyrus helvus Kerr, 1792
Vespertilio vampyrus helvus Kerr, 1792
Vespertilio vampyrus subniger Kerr, 1792
Vespertilio venustus Matschie, 1899
Vespertiliones Dobson, 1878
Vespertiliones Pallas, 1767
Vespertiliones Peters, 1866
VESPERTILIONIA Van Valen, 1979

Current Name

Scotophilus dinganii (A. Smith, 1833)
Rhinopoma cystops Thomas, 1903
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Myotis goudoti (A. Smith, 1834)
Pipistrellus (Pipistrellus) hesperidus hesperidus
 (Temminck, 1840)
Pipistrellus (Pipistrellus) hesperidus
 (Temminck, 1840)
Rhinolophus hipposideros (Bechstein, 1800)
Nycteris hispida (Schreber, 1774)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Kerivoula lanosa (A. Smith, 1847)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Barbastella leucomelas (Cretzschmar, 1826)
Myotis goudoti (A. Smith, 1834)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Laephotis matroka (Thomas and Schwann, 1905)
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Rhinopoma microphyllum (Brünnich, 1782)
Laephotis capensis (A. Smith, 1829)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Neoromicia somalica (Thomas, 1901)
Myotis punicus Felten, 1977
Myotis mystacinus (Kuhl, 1817)
Afronycteris nana (Peters, 1852)
Miniopterus natalensis (A. Smith, 1833)
Scotophilus nigrita (Schreber, 1774)
Hypsugo savii (Bonaparte, 1837)
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Afronycteris nana (Peters, 1852)
Pipistrellus (Pipistrellus) hesperidus
 (Temminck, 1840)
Eptesicus (Cnephaeus) platyops (Thomas, 1901)
Laephotis "incertae-sedis"
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Vansonia rueppellii rueppellii (J.B. Fischer, 1829)
Hypsugo savii (Bonaparte, 1837)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus natalensis (A. Smith, 1833)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Pipistrellus (Pipistrellus) hesperidus subtilis
 (Sundevall, 1846)
Myotis goudoti (A. Smith, 1834)
Vansonia rueppellii rueppellii (J.B. Fischer, 1829)
Vansonia rueppellii (J.B. Fischer, 1829)
Pteropus niger (Kerr, 1792)
Eidolon helvum (Kerr, 1792)
Eidolon helvum helvum (Kerr, 1792)
Pteropus subniger (Kerr, 1792)
Myotis welwitschii (Gray, 1866)
Vespertilioninae Gray, 1821
CHIROPTERA Blumenbach, 1779
VESPERTILIONIDAE Gray, 1821
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007

Original Name

Vespertilionidae Gray, 1821
VESPERTILIONIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
Vespertilioniformes Zagorodniuk, 2014
VESPERTILIONIFORMES Zagorodnyuk, 1998
VESPERTILIONIFORMES Hutcheon and Kirsch, 2004
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
Vespertilionimorpha Zagorodniuk, 2014

Vespertilionina Gray, 1825
Vespertilioninae Miller, 1897
Vespertilioninae Gray, 1821
Vespertilionini Gray, 1821
Vespertilionini Gray, 1821
Vespertilionoidea Gray, 1821
Vespertiliavus kasserinensis Ravel, 2016
Vesperugo Keyserling and Blasius, 1839
Vesperugo Keyserling and Blasius, 1839
Vesperugo Keyserling and Blasius, 1839
Vesperugo (*Vesperus*) *brunneus* Thomas, 1880
Vesperugo (*Vesperus*) *flavescens* Seabra, 1900
Vesperugo (*Vesperus*) *grandidieri* Dobson, 1876
Vesperugo (*Vesperus*) *Innesi* Lataste, 1887
Vesperugo (*Vesperus*) *Moloneyi* Thomas, 1891
Vesperugo (*Vesperus*) *Rendalli* Thomas, 1889
Vesperugo [(*Vesperugo*)] *maderensis* Dobson, 1878
Vesperugo *Anchieta* Seabra, 1900
Vesperugo *hypoleucus* Heuglin and Fitzinger, 1866
Vesperugo noctula var. *maxima* Fatio, 1869
Vesperugo *pagenstecheri* Noack, 1889
Vesperugo pulcher Dobson, 1875
Vesperugo pusillulus Peters, 1870
Vesperugo senarensis Heuglin, 1877

Vesperugo sennaariensis Heuglin and Fitzinger, 1866
Vesperugo stampflii Jentink, 1888
Vesperus Keyserling and Blasius, 1839
Vesperus (*Hesperoptenus*) *kraussii* Peters, 1868
Vesperus bicolor Bocage, 1889
Vesperus Bottae Peters, 1869
Vesperus damarensis Noack, 1889
Vesperus guineensis Bocage, 1889
Vesperus hösemanni Turni and Kock, 2008
Vesperus Humbloti A. Milne-Edwards, 1881
Vesperus pusillus Noack, 1889
Vesperus tenuipinnis Peters, 1872
Witwatia Gunnell, Simons and Seiffert, 2008
Witwatia eremicus Gunnell, Simons and Seiffert, 2008
Witwatia schlosseri Gunnell, Simons and Seiffert, 2008
Witwatia sigei Ravel, Marivaux, Tabuce, Bel Haj Ali, Essid and Vianey-Liaud, 2012
Xantharpyia Gray, 1843
Xantharpyia leucomelas Heuglin, 1866
Xantharpyia leucomelas Fitzinger, 1866
Xiphonycteris Dollman, 1911
Xiphonycteris spurrelli Dollman, 1911
YANGOCHIROPTERA Koopman, 1985

Yangochiroptera

Yinochiroptera Koopman, 1985

Current Name

VESPERTILIONIDAE Gray, 1821
VESPERTILIONIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
CHIROPTERA Blumenbach, 1779
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
VESPERTILIONIDAE Gray, 1821
Vespertilioninae Gray, 1821
Vespertilioninae Gray, 1821
Vespertilionini Gray, 1821
Vespertilioninae Gray, 1821
VESPERTILIONOIDEA Gray, 1821
Vespertiliavus kasserinensis (Ravel, 2016)
Eptesicus Rafinesque, 1820
Nyctalus Bowdich, 1825
Pipistrellus Kaup, 1829
Pseudoromicia brunnea (Thomas, 1880)

Nycticeinops grandidieri (Dobson, 1876)

Nycticeinops grandidieri (Dobson, 1876)

Eptesicus (*Cnephaeus*) *bottae* (Peters, 1869)
Mimetillus moloneyi (Thomas, 1891)

Pseudoromicia rendalli (Thomas, 1889)
Pipistrellus (*Pipistrellus*) *maderensis* (Dobson, 1878)
Neoromicia anchietae (Seabra, 1900)
Vansonia rueppellii rueppellii (J.B. Fischer, 1829)
Nyctalus lasiopterus (Schreber, 1780)
Afronycteris nana (Peters, 1852)
Vansonia rueppellii pulcher (Dobson, 1875)
Afronycteris nana (Peters, 1852)
Vansonia rueppellii rueppellii (J.B. Fischer, 1829)
Vansonia rueppellii rueppellii (J.B. Fischer, 1829)
Afronycteris nana (Peters, 1852)
Eptesicus Rafinesque, 1820
Glauconycteris poensis (Gray, 1842)

Pseudoromicia tenuipinnis (Peters, 1872)
Eptesicus (*Cnephaeus*) *bottae* (Peters, 1869)
Laephotis capensis (A. Smith, 1829)
Neoromicia guineensis (Bocage, 1889)
Pseudoromicia rendalli (Thomas, 1889)
Laephotis humbloti (A. Milne-Edwards, 1881)
Afronycteris nana (Peters, 1852)
Pseudoromicia tenuipinnis (Peters, 1872)
Witwatia Gunnell, Simons and Seiffert, 2008
Witwatia eremicus Gunnell, Simons and Seiffert, 2008
Witwatia schlosseri Gunnell, Simons and Seiffert, 2008
Witwatia sigei Ravel, Marivaux, Tabuce, Bel Haj Ali, Essid and Vianey-Liaud, 2012
Rousettus Gray, 1821
Eidolon helvum (Kerr, 1792)
Eidolon helvum helvum (Kerr, 1792)
Mops (*Xiphonycteris*) *Dollman*, 1911
Mops (*Xiphonycteris*) *spurrelli* (Dollman, 1911)
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
VESPERTILIONIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
CHIROPTERA Blumenbach, 1779

Original Name

YINPTEROCHIROPTERA Springer, Teeling,
Madsen, Stanhope and De Jong, 2001
YINPTEROCHIROPTERA Springer, Teeling,
Madsen, Stanhope and De Jong, 2001
Zygænocephalus A. Murray, 1862

Current Name

VESPERTILIONIFORMI Van Cakenberghe,
Kearney and Seamark, 2007
PTEROPODIFORMI Van Cakenberghe,
Kearney and Seamark, 2007
Hypsignathus H. Allen, 1862

Appendix 3b: Synonyms by Author

This appendix contains all of the synonyms covered in this report (excluding new name combinations and lapsi calami). The synonyms are sorted by author and year of description and are linked to the current name.

Some of the synonyms might be linked to several current names, indicating that they can only be partly assigned to these current names, e.g. *Vesperugo* Keyserling and Blasius, 1839 in part covers *Eptesicus* Rafinesque, 1820, *Nyctalus* Bowdich, 1825, and *Pipistrellus* Kaup, 1829.

Unknown Author

9999	<i>Epomophorus labiatus anurus</i>	<i>Epomophorus minor</i> Dobson, 1880
9999	<i>Laephotis cf. angolensis</i>	<i>Laephotis angolensis</i> Monard, 1935
9999	<i>Myotis anjouanensis</i>	<i>Myotis anjouanensis</i> Dorst, 1960
9999	<i>Noctilionoidea</i>	<i>NOCTILIONOIDEA</i> Gray, 1821
9999	<i>Rousettus sp. cf. R. madagascariensis</i>	<i>Rousettus madagascariensis</i> G. Grandidier, 1929
9999	<i>Yangochiroptera</i>	<i>VESPERTILIONIFORMACEI</i> Van Cakenberghe, Kearney and Seemark, 2007
9999	<i>Miniopterus sp.</i>	<i>Miniopterus</i> Bonaparte, 1837
9999	<i>Neoromicia "incertae-sedis"</i>	<i>Laephotis "incertae-sedis"</i>
9999	<i>Neoromicia cf. capensis</i>	<i>Laephotis capensis</i> (A. Smith, 1829)
9999	<i>Pteropus aegyptiacus</i>	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
9999	<i>Rhinonictidae</i>	<i>RHINONYCTERIDAE</i> J. E. Gray, 1866
9999	<i>Scotophilus cf. viridis</i>	<i>Scotophilus viridis</i> (Peters, 1852)

Acloque

1899	<i>Euvespertilio</i>	<i>Myotis</i> Kaup, 1829
1899	<i>Euvesperugo</i>	<i>Pipistrellus</i> Kaup, 1829

Aellen

1950	<i>Epomophorus reii</i>	<i>Epomophorus labiatus</i> (Temminck, 1837)
1950	<i>Epomophorus reii</i>	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
1952	<i>Nycteris benuensis</i>	<i>Nycteris parisii</i> (de Beaux, 1924)
1954	<i>Hipposideros marisae</i>	<i>Hipposideros marisae</i> Aellen, 1954
1956	<i>Miniopterus inflatus villiersi</i>	<i>Miniopterus villiersi</i> Aellen, 1956
1957	<i>Pipistrellus (Pipistrellus) permixtus</i>	<i>Pipistrellus (Pipistrellus) permixtus</i> Aellen, 1957
1959	<i>Kerivoula harrisoni bellula</i>	<i>Kerivoula lanosa</i> (A. Smith, 1847)
1959	<i>Nycteris intermedia</i>	<i>Nycteris intermedia</i> Aellen, 1959
1959	<i>Pipistrellus inexpectatus</i>	<i>Pipistrellus (Pipistrellus) inexpectatus</i> Aellen, 1959
1959	<i>Rhinolophus silvestris</i>	<i>Rhinolophus silvestris</i> Aellen, 1959
1973	<i>Rhinolophus hilli</i>	<i>Rhinolophus hilli</i> Aellen, 1973

Aellen and Brosset

1968	<i>Rhinolophus adami</i>	<i>Rhinolophus adami</i> Aellen and Brosset, 1968
1968	<i>Triaenops persicus majusculus</i>	<i>Triaenops afer</i> Peters, 1877

Allen

1917	<i>Rhinolophus axillaris</i>	<i>Rhinolophus landeri</i> Martin, 1838
1939	<i>Mops rüppellii</i>	<i>Tadarida ventralis</i> (Heuglin, 1861)

G.M. Allen and Lawrence

1936	<i>Nycteris nana tristis</i>	<i>Nycteris nana</i> (K. Andersen, 1912)

G.M. Allen and Loveridge

1942	<i>Mops angolensis orientis</i>	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)

G.M. Allen

1908	<i>Scabrifer</i>	<i>Laephotis</i> Thomas, 1901
1908	<i>Scabrifer notius</i>	<i>Laephotis capensis</i> (A. Smith, 1829)
1911	<i>Eptesicus phasma</i>	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
1911	<i>Nycticeius africanus</i>	<i>Nycticeinops schlieffenii</i> (Peters, 1859)

1914	<i>Scotophilus altilis</i>	<i>Scotophilus altilis</i> G.M. Allen, 1914
1914	<i>acrotis</i>	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1918	<i>Chaerephon leucostigma</i>	<i>Mops (Mops) leucostigma</i> (G.M. Allen, 1918)
1921	<i>Hipposideros curtus</i>	<i>Hipposideros curtus</i> G.M. Allen, 1921

H. Allen

1862	<i>Epomophorus comptus</i>	<i>Epomops franqueti</i> (Tomes, 1860)
1862	<i>H[ypsignathus] monstrosus</i>	<i>Hypsignathus monstrosus</i> H. Allen, 1862
1862	<i>Hypsignathus</i>	<i>Hypsignathus</i> H. Allen, 1862
1862	<i>Pteropus mollipilosus</i>	<i>Eidolon helvum helvum</i> (Kerr, 1792)
1864	<i>Megadermatidae</i>	<i>MEGADERMATIDAE</i> H. Allen, 1864
1892	<i>Adelonycteris</i>	<i>Eptesicus Rafinesque</i> , 1820

J.A. Allen

1917	<i>Allomops</i>	<i>Mops (Mops)</i> Lesson, 1842
1917	<i>Chærephon (Lophomops)</i>	<i>Mops (Chaerephon)</i> Dobson, 1874
1917	<i>Chærephon (Lophomops) abæ</i>	<i>Mops (Chaerephon) major</i> (Trouessart, 1897)
1917	<i>Chærephon (Lophomops) chapini</i>	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
1917	<i>Chærephon (Lophomops) cristatus</i>	<i>Mops (Chaerephon) leucogaster</i> (A. Grandidier, 1869)
1917	<i>Chærephon frater</i>	<i>Mops (Chaerephon) leucogaster</i> (A. Grandidier, 1869)
1917	<i>Chærephon russatus</i>	<i>Mops (Chaerephon) russatus</i> (J.A. Allen, 1917)
1917	<i>Eptesicus ater</i>	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
1917	<i>Eptesicus faradjius</i>	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
1917	<i>Eptesicus garambæ</i>	<i>Laephotis capensis</i> (A. Smith, 1829)
1917	<i>Glauconycteris alboguttatus</i>	<i>Glauconycteris alboguttata</i> J.A. Allen, 1917
1917	<i>Glauconycteris humeralis</i>	<i>Glauconycteris humeralis</i> J.A. Allen, 1917
1917	<i>Hipposideros abæ</i>	<i>Hipposideros abae</i> J.A. Allen, 1917
1917	<i>Hipposideros caffer niapu</i>	<i>Hipposideros ruber</i> (Noack, 1893)
1917	<i>Hipposideros gigas niangaræ</i>	<i>Macronycteris gigas</i> (Wagner, 1845)
1917	<i>Hipposideros langi</i>	<i>Doryrhina cyclops</i> (Temminck, 1853)
1917	<i>Hipposideros nanus</i>	<i>Hipposideros caffer</i> (Sundevall, 1846)
1917	<i>Miniopterus breyeri vicinior</i>	<i>Miniopterus fraterculus</i> Thomas and Schwann, 1906
1917	<i>Mops (Allomops) faradjius</i>	<i>Mops (Mops) demonstrator</i> (Thomas, 1903)
1917	<i>Mops (Allomops) nanulus</i>	<i>Mops (Xiphonycteris) nanulus</i> J.A. Allen, 1917
1917	<i>Mops (Allomops) occipitalis</i>	<i>Mops (Xiphonycteris) thersites</i> (Thomas, 1903)
1917	<i>Mops (Allomops) osborni</i>	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
1917	<i>Mops congcicus</i>	<i>Mops (Mops) congcicus</i> J.A. Allen, 1917
1917	<i>Mops niangaræ</i>	<i>Mops (Mops) niangaræ</i> J.A. Allen, 1917
1917	<i>Mops trevori</i>	<i>Mops (Mops) trevori</i> J.A. Allen, 1917
1917	<i>Nycteris avakubia</i>	<i>Nycteris major</i> (K. Andersen, 1912)
1917	<i>Nycteris pallida</i>	<i>Nycteris hispida</i> (Schreber, 1774)
1917	<i>Nyctinomus ochraceus</i>	<i>Mops (Xiphonycteris) brachypterus</i> (Peters, 1852)
1917	<i>Pipistrellus abaensis</i>	<i>Afronycteris nana</i> (Peters, 1852)
1917	<i>Rhinolophus abæ</i>	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1917	<i>Nycteris major</i>	<i>Nycteris macrotis</i> Dobson, 1876

Almeida, Giannini and Simmons

2016	<i>Eidolinae</i>	<i>Eidolinae</i> Almeida, Giannini and Simmons, 2016

Ameghino

1889	<i>Gymnuridae</i>	<i>MOLOSSIDAE</i> Gervais, 1856
1889	<i>Ptética</i>	<i>CHIROPTERA</i> Blumenbach, 1779

Ancillotto, Bosso, Smeraldo, Mori, Mazza, Herkt, Galimberti, Ramazzotti and Russo

2020	<i>Plecotus cf. gaisleri</i>	<i>Plecotus gaisleri</i> Benda, Kiefer, Hanak & Veith, 2004

K. Andersen and Matschie

1904	<i>E[uryalus] atlanticus</i>	<i>Rhinolophus euryale</i> Blasius, 1853
1904	<i>E[uryalus] barbarus</i>	<i>Rhinolophus mehelyi</i> Matschie, 1901
1904	<i>E[uryalus] Cabrera</i>	<i>Rhinolophus euryale</i> Blasius, 1853
1904	<i>E[uryalus] meridionalis</i>	<i>Rhinolophus mehelyi</i> Matschie, 1901

K. Andersen and Wroughton

1907	<i>Lavia frons affinis</i>	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)

K. Andersen

1904	<i>Rhinolophus augur</i>	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1904	<i>Rhinolophus augur zambesiensis</i>	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1904	<i>Rhinolophus augur zuluensis</i>	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1904	<i>Rhinolophus simulator</i>	<i>Rhinolophus simulator</i> K. Andersen, 1904
1904	<i>Rhinolophus empusa</i>	<i>Rhinolophus blasii</i> Peters, 1867
1905	<i>Rhinolophus acrotis brachygnatus</i>	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1905	<i>Rhinolophus Darlingi</i>	<i>Rhinolophus darlingi</i> K. Andersen, 1905
1905	<i>Rhinolophus fumigatus exsul</i>	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1905	<i>Rhinolophus Hildebrandti eloquens</i>	<i>Rhinolophus eloquens</i> K. Andersen, 1905
1905	<i>Rhinolophus hipposideros typicus</i>	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1906	<i>H[ipposiderus]. gigas gambiensis</i>	<i>Macronycteris gigas</i> (Wagner, 1845)
1906	<i>Hipposiderus beatus</i>	<i>Hipposideros beatus</i> (K. Andersen, 1906)
1906	<i>Hipposiderus caffer centralis</i>	<i>Hipposideros ruber</i> (Noack, 1893)
1906	<i>Hipposiderus caffer guineensis</i>	<i>Hipposideros ruber</i> (Noack, 1893)
1908	<i>Pteropus rufus princeps</i>	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
1908	<i>Myonycteris leptodon</i>	<i>Myonycteris leptodon</i> K. Andersen, 1908
1908	<i>Myonycteris wroughtoni</i>	<i>Myonycteris torquata wroughtoni</i> K. Andersen, 1908
1910	<i>Epomops franqueti strepitans</i>	<i>Epomops franqueti</i> (Tomes, 1860)
1910	<i>Plerotes</i>	<i>Plerotes</i> K. Andersen, 1910
1912	<i>Cynopterinae</i>	<i>Pteropodinae</i> Gray, 1821
1912	<i>Epomophorinae</i>	<i>Rousettinae</i> Andersen, 1912
1912	<i>Rousettinae</i>	<i>Rousettinae</i> Andersen, 1912
1912	<i>Rousettini</i>	<i>Rousettini</i> Andersen, 1912
1912	<i>Petalia aurita</i>	<i>Nycteris aurita</i> (K. Andersen, 1912)
1912	<i>Petalia damarensis brockmani</i>	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1912	<i>Petalia damarensis media</i>	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1912	<i>Petalia gambiensis</i>	<i>Nycteris gambiensis</i> (K. Andersen, 1912)
1912	<i>Petalia major</i>	<i>Nycteris major</i> (K. Andersen, 1912)
1912	<i>Petalia nana</i>	<i>Nycteris nana</i> (K. Andersen, 1912)
1914	<i>Nycteris woodi</i>	<i>Nycteris woodi</i> K. Andersen, 1914
1918	<i>[Rhinolophus hipposideros] escalerae</i>	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1918	<i>Asellia tridens diluta</i>	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)

Anderson

1881	<i>Phyllorhina tridens murreana</i>	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
1902	<i>Otonycteris petersi</i>	<i>Otonycteris hemprichii</i> Peters, 1859

Ansell

1960	<i>Epomophorus gambianus parvus</i>	<i>Epomophorus crypturus</i> Peters, 1852

Aristotle

-330	<i>DERMAPTERA</i>	<i>CHIROPTERA</i> Blumenbach, 1779

Arroyo-Cabrales, Gregorin, Schlitter and Walker

2002	<i>Tadarida rusingae</i>	<i>Mops rusingae</i> (Arroyo-Cabrales, Gregorin, Schlitter and Walker, 2002)

Barrett-Hamilton

1906	<i>Pterygistes madeirae</i>	<i>Nyctalus leisleri</i> (Kuhl, 1817)
1907	<i>Plecotus teneriffae</i>	<i>Plecotus teneriffae</i> Barrett-Hamilton, 1907

Bates, Ratrimomanarivo, Harrison and Goodman

2006	<i>Pipistrellus raceyi</i>	<i>Pipistrellus (Pipistrellus) raceyi</i> Bates, Ratrimomanarivo, Harrison and Goodman, 2006

Bechstein

1800	<i>Vespertilio hipposideros</i>	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1800	<i>Vespertilio megalotis</i>	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)

Benda

2008	<i>Plecotus christii petraeus</i>	<i>Plecotus christii</i> Gray, 1838

Benda and Vallo

2009	<i>Paratriaenops</i>	<i>Paratriaenops</i> Benda and Vallo, 2009
2009	<i>Triaenopini</i>	<i>RHINONYCTERIDAE</i> J. E. Gray, 1866
2012	<i>Rhinolophus horaceki</i>	<i>Rhinolophus horaceki</i> Benda and Vallo, 2012

Benda, Kiefer, Hanák and Veith

2004	<i>Plecotus teneriffae gaisleri</i>	<i>Plecotus gaisleri</i> Benda, Kiefer, Hanak & Veith, 2004

Bennett

1836	<i>Epomophorus</i>	<i>Epomophorus</i> Bennett, 1836
1836	<i>Pteropus epomophorus</i>	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
1836	<i>Pteropus Whitei</i>	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)

Bergmans

1980	<i>Myonycteris relict</i>	<i>Myonycteris relict</i> Bergmans, 1980
1997	<i>Lissonycteris angolensis goliath</i>	<i>Myonycteris angolensis goliath</i> (Bergmans, 1997)
1997	<i>Lissonycteris angolensis petraea</i>	<i>Myonycteris angolensis petraea</i> (Bergmans, 1997)
1997	<i>Plerotini</i>	<i>Plerotini</i> Bergmans, 1997
1997	<i>Scotonycterini</i>	<i>Scotonycterini</i> Bergmans, 1997

Bergmans and Van Strien

2004	<i>Epomophorus anelli</i>	<i>Epomophorus anelli</i> Bergmans and Van Strien, 2004

Bianchi

1917	<i>Dichromyotis</i>	<i>Myotis</i> Kaup, 1829
1917	<i>Paramyotis</i>	<i>Myotis</i> Kaup, 1829
1917	<i>Rickettia</i>	<i>Myotis</i> Kaup, 1829
1917	<i>Megapipistrellus</i>	<i>Myotis</i> Kaup, 1829
1917	<i>Pareptesicus</i>	<i>Eptesicus</i> Rafinesque, 1820
1917	<i>Rhynptesicus</i>	<i>Eptesicus</i> Rafinesque, 1820

Blanford

1891	<i>Cerivoula</i>	<i>Kerivoula</i> Gray, 1842

Blasius

1853	<i>Rhinolophus euryale</i>	<i>Rhinolophus euryale</i> Blasius, 1853

Blumenbach

1779	<i>CHIROPTERA</i>	<i>CHIROPTERA</i> Blumenbach, 1779
1797	<i>Vespertilio caninus</i>	<i>Pteropus niger</i> (Kerr, 1792)

Blyth

1840	<i>Romicius</i>	<i>Pipistrellus</i> Kaup, 1829

Bocage

1889	<i>Epomophorus Dobsonii</i>	<i>Epomophorus dobsonii</i> Bocage, 1889
1889	<i>Vesperus bicolor</i>	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
1889	<i>Vesperus guineensis</i>	<i>Neoromicia guineensis</i> (Bocage, 1889)
1889	<i>Cynonycteris brachycephala</i>	<i>Myonycteris brachycephala</i> (Bocage, 1889)
1889	<i>Miniopterus Newtoni</i>	<i>Miniopterus newtoni</i> Bocage, 1889
1891	<i>Phyllorhina commersoni</i> var. <i>thomensis</i>	<i>Macronycteris vittatus</i> (Peters, 1852)
1898	<i>Cynonycteris Angolensis</i>	<i>Myonycteris angolensis angolensis</i> (Bocage, 1898)
1898	<i>Cynonycteris Angolensis</i>	<i>Myonycteris angolensis</i> (Bocage, 1898)
1898	<i>Epomophorus guineensis</i>	<i>Epomophorus gambianus</i> (Ogilby, 1835)
1898	<i>Epomophorus guineensis</i>	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)

Boie

1830	<i>Leuconoe</i>	<i>Myotis</i> Kaup, 1829

Bonaparte

1831	<i>Phyllorrhina</i>	<i>Hipposideros</i> Gray, 1831
1831	<i>Pteropodina</i>	<i>Pteropodinae</i> Gray, 1821
1835	<i>Pteropodae</i>	<i>Pteropodinae</i> Gray, 1821
1837	<i>Miniopterus</i>	<i>Miniopterus</i> Bonaparte, 1837
1837	<i>Noctula</i>	<i>Eptesicus</i> Rafinesque, 1820
1837	<i>Phyllorrhina</i>	<i>Hipposideros</i> Gray, 1831
1837	<i>Pipistrella</i>	<i>Pipistrellus</i> Kaup, 1829
1837	<i>Vespertilio Capaccinii</i>	<i>Myotis capaccinii</i> (Bonaparte, 1837)
1837	<i>Vespertilio Savii</i>	<i>Hypsugo savii</i> (Bonaparte, 1837)
1838	<i>PTEROPODIDAE</i>	<i>PTEROPODIDAE</i> Gray, 1821
1838	<i>Rhinopomina</i>	<i>RHINOPOMATIDAE</i> Dobson, 1872
1841	<i>Capaccinius</i>	<i>Myotis</i> Kaup, 1829
1841	<i>Selysius</i>	<i>Myotis</i> Kaup, 1829

Bourret

1951	<i>Rhinomegalophus</i>	<i>Rhinolophus</i> Lacépède, 1799

Bowdich

1821	<i>Nyctinoma</i>	<i>Tadarida</i> Rafinesque, 1814
1821	<i>Rhinopoma</i>	<i>Rhinopoma</i> E. Geoffroy St.-Hilaire, 1818
1825	<i>Nyctalus</i>	<i>Nyctalus</i> Bowdich, 1825
1825	<i>Nyctalus verrucosus</i>	<i>Nyctalus leisleri</i> (Kuhl, 1817)

Brehm

1827	<i>Vespertilio ferrugineus</i>	<i>Nyctalus lasiopterus</i> (Schreber, 1780)

Brisson

1762	<i>Pteropus</i>	<i>Pteropus</i> Brisson, 1762

Brooks and Bickham

2014	<i>Scotophilus andrewreborii</i>	<i>Scotophilus andrewreborii</i> Brooks and Bickham, 2014
2014	<i>Scotophilus livingstonii</i>	<i>Scotophilus livingstonii</i> Brooks and Bickham, 2014
2014	<i>Scotophilus ejetai</i>	<i>Scotophilus ejetai</i> Brooks and Bickham, 2014
2014	<i>Scotophilus trujilloi</i>	<i>Scotophilus trujilloi</i> Brooks and Bickham, 2014

Brosset

1966	<i>Tadarida (Chaerephon) cyclotis</i>	<i>Mops (Chaerephon) aloysiisabaudiae</i> (Festa, 1907)
1985	<i>Hipposideros lamottei</i>	<i>Hipposideros lamottei</i> Brosset, 1985

Brünnich

1782	<i>Vespertilio Microphyllus</i>	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)

Burnett

1829	<i>Cercopteropus</i>	<i>Rousettus</i> Gray, 1821

Butler

1984	<i>Chamtwaria</i>	<i>Chamtwaria</i> Butler, 1984
1984	<i>Chamtwaria pickfordi</i>	<i>Chamtwaria pickfordi</i> Butler, 1984
1984	<i>Propottininae</i>	<i>Propottininae</i> Butler, 1984

Butler and Hopwood

1957	<i>Saccolaimus incognita</i>	<i>Taphozous incognita</i> (Butler & Hopwood, 1957)

Cabrera

1904	<i>Vespertilio Boscai</i>	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
1904	<i>Vespertilio ochromixtus</i>	<i>Hypsugo savii</i> (Bonaparte, 1837)
1906	<i>Hipposiderus tephurus</i>	<i>Hipposideros tephurus</i> (Cabrera, 1906)

1914	<i>Noctulinia</i>	<i>Nyctalus</i> Bowdich, 1825
1920	<i>Rousettus (Lissonycteris) crypticola</i>	<i>Myonycteris angolensis</i> (Bocage, 1898)
1958	<i>Hesperomyotis</i>	<i>Myotis</i> Kaup, 1829

Cabrera and Ruxton

1926	<i>Mops angolensis niveiventer</i>	<i>Mops (Mops) niveiventer</i> Cabrera and Ruxton, 1926

Castella, Ruedi, Excoffier, Ibanez, Arlettaz and Hausser

2000	<i>Myotis cf. punicus</i>	<i>Myotis punicus</i> Felten, 1977

Cheesman and Hinton

1924	<i>Otonycteris jin</i>	<i>Otonycteris hemprichii</i> Peters, 1859

Chubb

1917	<i>Otomops icarus</i>	<i>Otomops martiensseni icarus</i> Chubb, 1917

Claessen and De Vree

1991	<i>Epomophorus minimus</i>	<i>Epomophorus minimus</i> Claessen & De Vree, 1991

Çoraman, Mayer and Dietz

2019	<i>Myotis escalerae cabrerai</i>	<i>Myotis zenatus</i> Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019

Cotterill

2002	<i>Rhinolophus sakejiensis</i>	<i>Rhinolophus sakejiensis</i> Cotterill, 2002

Cretzschmar

1826	<i>Dysopes pumilus</i>	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1826	<i>Nycticejus leucogaster</i>	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
1826	<i>Vespertilio leucomelas</i>	<i>Barbastella leucomelas</i> (Cretzschmar, 1826)
1826	<i>Vespertilio temminckii</i>	<i>Vansonia rueppellii</i> (J.B. Fischer, 1829)
1827	<i>Vespertilio temminckii</i>	<i>Vansonia rueppellii rueppellii</i> (J.B. Fischer, 1829)
1828	<i>Rhinolophus clivosus</i>	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1830	<i>Vespertilio marginatus</i>	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
1830-1831	<i>Dysopes</i>	<i>Tadarida Rafinesque</i> , 1814
1830-1831	<i>Taphozous nudiventris</i>	<i>Taphozous nudiventris</i> Cretzschmar, 1830

G. Cuvier and E. Geoffroy

1795	<i>Nycteris</i>	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795

G. Cuvier

1816	<i>Cheiroptères</i>	<i>CHIROPTERA</i> Blumenbach, 1779

Dalquest

1965	<i>Nycteris vinsoni</i>	<i>Nycteris vinsoni</i> Dalquest, 1965
1965	<i>Scotophilus alvenslebeni</i>	<i>Scotophilus nigrita</i> (Schreber, 1774)

Daudin

1802	<i>Spectrum rubidum</i>	<i>Pteropus subniger</i> (Kerr, 1792)

de Beaux

1922	<i>Rhinolophus hildebrandti perauritus</i>	<i>Rhinolophus eloquens</i> K. Andersen, 1905
1923	<i>Petalia (Nycteris) thebaica aurantiaca</i>	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1923	<i>Scotoecus artinii</i>	<i>Scotoecus hirundo</i> (de Winton, 1899)
1924	<i>Petalia parisii</i>	<i>Nycteris parisii</i> (de Beaux, 1924)
1924	<i>Hipposideros caffer aurantiaca</i>	<i>Hipposideros caffer</i> (Sundevall, 1846)
1931	<i>Asellia patrizii</i>	<i>Asellia patrizii</i> de Beaux, 1931
1931	<i>Asellia tridens italo-somalica</i>	<i>Asellia italosomalica</i> de Beaux, 1931

de Blainville

1837 *Myoptera* *Myopterus* E. Geoffroy St.-Hilaire, 1818

de Rochebrune

1883 *Chiropteri* *CHIROPTERA* Blumenbach, 1779
 1883 *Phyllorhinidae* *RHINOLOPHIDAE* Gray, 1825

De Vree

1972 *Pipistrellus eisentrauti bellieri* *Nycticeinops bellieri* (De Vree, 1972)

de Winton

1897 *Rhinolophus micaceus* *Doryrhina cyclops* (Temminck, 1853)
 1899 *Scotophilus hirundo* *Scotoecus hirundo* (de Winton, 1899)
 1899 *Scotophilus nigrnellus* *Scotophilus nigrnellus* de Winton, 1899
 1901 *Glauconycteris Floweri* *Eptesicus (Rhinopterus) floweri* (de Winton, 1901)
 1901 *N(yctinomus) midas* *Tadarida ventralis* (Heuglin, 1861)
 1901 *Nyctinomus Emini* *Mops (Chaerephon) major* (Trouessart, 1897)
 1901 *Nyctinomus gambianus* *Mops (Chaerephon) pumilus* (Cretzschmar, 1826)

Decher, Hutterer and Monadjem

2016 *Neoromicia isabella* *Pseudoromicia isabella* (Decher, Hutterer and Monadjem, 2016)

Deleuil and Labbé

1955 *Rhinolophus euryale tuneti* *Rhinolophus mehelyi* Matschie, 1901

Desmarest

1820 *Myopterus* *Myopterus* E. Geoffroy St.-Hilaire, 1818
 1820 *Myopterus daubentonii* *Myopterus daubentonii* Desmarest, 1820
 1820 *Nycteris Geoffroyi* *Nycteris thebaica* E. Geoffroy St.-Hilaire, 1818
 1820 *Taphozous senegalensis* *Taphozous perforatus perforatus* E. Geoffroy St.-Hilaire, 1818

Dobson

1871 *Triaenops* *Triaenops* Dobson, 1871
 1872 *Rhinopomatidae* *RHINOPOMATIDAE* Dobson, 1872
 1872 *Rhinopomatoidea* *RHINOPOMATOIDEA* Dobson, 1872
 1874 *Nyctinomus (Chaerephon)* *Mops (Chaerephon)* Dobson, 1874
 1875 *Scotophilus gigas* *Scotophilus nigrita* (Schreber, 1774)
 1875 *Vespertilio murinus africanus* *Myotis punicus* Felten, 1977
 1875 *Chalinolobus argentatus* *Glauconycteris argentata* (Dobson, 1875)
 1875 *Glauconycteris* *Glauconycteris* Dobson, 1875
 1875 *Vesperugo pulcher* *Vansonia rueppellii pulcher* (Dobson, 1875)
 1875 *Emballonuridae* *EMBALLONURIDAE* Gervais, 1855
 1875 *Miniopteri* *MINIOPTERIDAE* Dobson, 1875
 1875 *Miniopteridae* *MINIOPTERIDAE* Dobson, 1875
 1875 *MEGACHIROPTERA* *PTEROPODIFORMI* Van Cakenberghe, Kearney and Seemark, 2007
 1875 *MICROCHIROPTERA* *CHIROPTERA* Blumenbach, 1779
 1876 *Taphonycteris* *Saccolaimus* Temminck, 1838
 1876 *Nyctinomus africanus* *Tadarida ventralis* (Heuglin, 1861)
 1876 *Vesperugo (Vesperus) grandidieri* *Nycticeinops grandidieri* (Dobson, 1876)
 1876 *Nycteris macrotis* *Nycteris macrotis* Dobson, 1876
 1877 *Nyctinomus [(Mormopterus)] albiventer* *Mormopterus jugularis* (Peters, 1865)
 1877 *Nyctinomus [(Nyctinomus)] cestoni* *Tadarida ventralis* (Heuglin, 1861)
 1878 *Cynonycteris torquata* *Myonycteris torquata torquata* (Dobson, 1878)
 1878 *Cynonycteris torquata* *Myonycteris torquata* (Dobson, 1878)
 1878 *Kerivoula africana* *Kerivoula africana* Dobson, 1878
 1878 *Kerivoula brunnea* *Kerivoula lanosa* (A. Smith, 1847)
 1878 *Nycteris aethiopica* *Nycteris macrotis* Dobson, 1876
 1878 *Plecotus aegyptiacus* *Plecotus christii* Gray, 1838
 1878 *Pteropus rodricensis* *Pteropus rodricensis* Dobson, 1878
 1878 *Vesperugo [(Vesperugo)] maderensis* *Pipistrellus (Pipistrellus) maderensis* (Dobson, 1878)
 1878 *Vespertiliones* *Vespertilioninae* Gray, 1821

1880	<i>Epomophorus minor</i>	<i>Epomophorus minor</i> Dobson, 1880
1885	<i>Rhinolophus antinorii</i>	<i>Rhinolophus fumigatus</i> Rüppell, 1842
- Dollman
- | | | |
|------|--------------------------------|--|
| 1908 | <i>Chœrephon websteri</i> | <i>Mops (Chaerephon) leucogaster</i> (A. Grandidier, 1869) |
| 1911 | <i>Xiphonycteris</i> | <i>Mops (Xiphonycteris)</i> Dollman, 1911 |
| 1911 | <i>Xiphonycteris spurrelli</i> | <i>Mops (Xiphonycteris) spurrelli</i> (Dollman, 1911) |
- Dorst
- | | | |
|------|---|---|
| 1953 | <i>Otomops madagascariensis</i> | <i>Otomops madagascariensis</i> Dorst, 1953 |
| 1960 | <i>Myotis Goudoti anjouanensis</i> | <i>Myotis anjouanensis</i> Dorst, 1960 |
| 1960 | <i>Pipistrellus Rüppelli senegalensis</i> | <i>Vansonia rueppellii senegalensis</i> (Dorst, 1960) |
- Dulic
- | | | |
|------|--|---|
| 1980 | <i>Plecotus austriacus kolombatovici</i> | <i>Plecotus kolombatovici</i> Dulic, 1980 |
|------|--|---|
- Duméril
- | | | |
|------|--------------------|------------------------------------|
| 1806 | <i>Chiropteren</i> | <i>CHIROPTERA</i> Blumenbach, 1779 |
|------|--------------------|------------------------------------|
- Eger and Schlitter
- | | | |
|------|------------------------------|--|
| 2001 | <i>Glauconycteris curryi</i> | <i>Glauconycteris curryae</i> Eger and Schlitter, 2001 |
|------|------------------------------|--|
- Eisentraut
- | | | |
|------|---|--|
| 1956 | <i>Hipposideros camerunensis</i> | <i>Doryrhina camerunensis</i> (Eisentraut, 1956) |
| 1960 | <i>Rousettus aegyptiacus occidentalis</i> | <i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810) |
| 1960 | <i>Rhinolophus denti knorri</i> | <i>Rhinolophus denti</i> Thomas, 1904 |
| 1960 | <i>Rhinolophus landeri guineensis</i> | <i>Rhinolophus guineensis</i> Eisentraut, 1960 |
| 1965 | <i>Rousettus angolensis ruwenzorii</i> | <i>Myonycteris angolensis ruwenzorii</i> (Eisentraut, 1965) |
- El Rayah
- | | | |
|------|---|--|
| 1981 | <i>Tadarida [(Xiphonycteris)] petersoni</i> | <i>Mops (Xiphonycteris) petersoni</i> (El Rayah, 1981) |
|------|---|--|
- J. Eric Hill
- | | | |
|------|-------------------------------|--|
| 1942 | <i>Rhinolophus ruwenzorii</i> | <i>Rhinolophus ruwenzorii</i> J. Eric Hill, 1942 |
|------|-------------------------------|--|
- Fahr and Kerbis Peterhans
- | | | |
|------|---------------------------|---|
| 2013 | <i>Rhinolophus kahuzi</i> | <i>Rhinolophus kahuzi</i> Fahr and Kerbis Peterhans, 2013 |
|------|---------------------------|---|
- Fahr, Vierhaus, Hutterer and Kock
- | | | |
|------|--------------------------|--|
| 2002 | <i>Rhinolophus ziama</i> | <i>Rhinolophus ziama</i> Fahr, Vierhaus, Hutterer and Kock, 2002 |
|------|--------------------------|--|
- Fatio
- | | | |
|------|--------------------------------------|--|
| 1869 | <i>Gymnorhinida</i> | <i>VESPERTILIONIDAE</i> Gray, 1821 |
| 1869 | <i>Vesperugo noctula var. maxima</i> | <i>Nyctalus lasiopterus</i> (Schreber, 1780) |
- Feiler, Haft and Widmann
- | | | |
|------|--|--|
| 1993 | <i>Rousettus aegyptiacus thomensis</i> | <i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810) |
|------|--|--|
- Felten
- | | | |
|------|-------------------------------|------------------------------------|
| 1977 | <i>Myotis blythii punicus</i> | <i>Myotis punicus</i> Felten, 1977 |
|------|-------------------------------|------------------------------------|
- Festa
- | | | |
|------|-------------------------------------|---|
| 1907 | <i>Nyctinomus aloysii-sabaudiae</i> | <i>Mops (Chaerephon) aloysiisabaudiae</i> (Festa, 1907) |
|------|-------------------------------------|---|

G. Fischer

1814 *Pteropus torquatus* *Pteropus subniger* (Kerr, 1792)

J.B. Fischer

1829 [*Vespertilio auritus*] *β aegyptius* *Plecotus christii* Gray, 1838
 1829 [*Vespertilio pipistrellus* var.] *β Aegyptius* *Pipistrellus (Pipistrellus) kuhlii* (Kuhl, 1817)
 1829 [*Vespertilio*] *Rüppelii* *Vansonia rueppellii* (J.B. Fischer, 1829)
 1829 *Vespertilio ruppellii* *Vansonia rueppellii rueppellii* (J.B. Fischer, 1829)

Fitzinger

1861 *Nyctophylax* *Kerivoula* Gray, 1842
 1866 *Rhinopoma longicaudatum* *Rhinopoma cystops* Thomas, 1903
 1866 *Rhinopoma sennaariense* *Rhinopoma cystops* Thomas, 1903
 1866 *Xantharpyia leucomelas* *Eidolon helvum helvum* (Kerr, 1792)
 1870 *Aëorestes* *Myotis* Kaup, 1829
 1870 *Comastes* *Myotis* Kaup, 1829
 1870 *Exochurus* *Myotis* Kaup, 1829
 1870 *Nyctiptenus* *Eptesicus* Rafinesque, 1820
 1870 *Rhinolophus Eggenhöffner* *Rhinolophus hipposideros* (Bechstein, 1800)

Fitzinger and Heuglin

1866 *Plecotus ustus* *Otonycteris hemprichii* Peters, 1859

Fleming

1822 *CHEIROPTERA* *CHIROPTERA* Blumenbach, 1779
 1822 *Nyctinomia* *Tadarida* Rafinesque, 1814

Foley, Goodman, Whelan, Puechmaille and Teeling

2017 *M[acronycteris] cryptovalorona* *Macronycteris cryptovalorona* (Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016)

Forbes

1882 *Liponyx* *Eidolon* Rafinesque, 1815

Fraser

1843 *Rhinolophus Martini* *Nycteris hispida* (Schreber, 1774)

E. Geoffroy Saint-Hilaire

1803 *major* *Rhinolophus ferrumequinum* (Schreber, 1774)
 1803 *Pteropus fuscus* *Pteropus niger* (Kerr, 1792)
 1803 *Pteropus ruber* *Pteropus subniger* (Kerr, 1792)
 1803 *Pteropus rufus* *Pteropus rufus* E. Geoffroy St.-Hilaire, 1803
 1803 *Pteropus stramineus* *Eidolon helvum helvum* (Kerr, 1792)
 1803 *Vespertilio borbonicus* *Scotophilus borbonicus* (E. Geoffroy St.-Hilaire, 1803)
 1806 *Vespertilio emarginatus* *Myotis emarginatus* (E. Geoffroy St.-Hilaire, 1806)
 1810 *Megaderma frons* *Lavia frons* (E. Geoffroy St.-Hilaire, 1810)
 1810 *Pteropus Edwardsii* *Pteropus rufus* E. Geoffroy St.-Hilaire, 1803
 1810 *Pteropus Egyptiacus* *Rousettus aegyptiacus* (E. Geoffroy St.-Hilaire, 1810)
 1810 *Pteropus rubricollis* *Pteropus subniger* (Kerr, 1792)
 1810 *Pteropus vulgaris* *Pteropus niger* (Kerr, 1792)
 1813 *Nycteris Daubentonii* *Nycteris hispida* (Schreber, 1774)
 1813 *Rhinolophus Commersonii* *Macronycteris commersoni* (E. Geoffroy St.-Hilaire, 1813)
 1813 *Rhinolophus tridens* *Asellia tridens* (E. Geoffroy St.-Hilaire, 1813)
 1818 *Myopterus* *Myopterus* E. Geoffroy St.-Hilaire, 1818
 1818 *Nycteris Thebaicus* *Nycteris thebaica* E. Geoffroy St.-Hilaire, 1818
 1818 *Nyctinomus* *Tadarida* Rafinesque, 1814
 1818 *Nyctinomus aegyptiacus* *Tadarida aegyptiaca aegyptiaca* (E. Geoffroy St.-Hilaire, 1818)
 1818 *Nyctinomus aegyptiacus* *Tadarida aegyptiaca* (E. Geoffroy St.-Hilaire, 1818)
 1818 *Plecotus* *Plecotus* E. Geoffroy St.-Hilaire, 1818
 1818 *Pteropus aegyptiacus* *Rousettus aegyptiacus* (E. Geoffroy St.-Hilaire, 1810)
 1818 *Rhinopoma* *Rhinopoma* E. Geoffroy St.-Hilaire, 1818

1818	<i>Taphozous</i>	<i>Taphozous</i> E. Geoffroy St.-Hilaire, 1818
1818	<i>Taphozous mauritianus</i>	<i>Taphozous mauritianus</i> E. Geoffroy St.-Hilaire, 1818
1818	<i>Taphozous perforatus</i>	<i>Taphozous perforatus perforatus</i> E. Geoffroy St.-Hilaire, 1818
1818	<i>Taphozous perforatus</i>	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818

E. Geoffroy St.-Hilaire

1810	<i>Megaderma</i>	<i>Megaderma</i> E. Geoffroy St.-Hilaire, 1810

Gervais

1836	<i>Rhinocrepis</i>	<i>Rhinolophus</i> Lacépède, 1799
1849	<i>Kirivoula</i>	<i>Kirivoula</i> Gray, 1842
1854	<i>Rhinolophides</i>	<i>RHINOLOPHIDAE</i> Gray, 1825
1855	<i>Emballonuridae</i>	<i>EMBALLONURIDAE</i> Gervais, 1855
1855	<i>Emballonurinae</i>	<i>Emballonurinae</i> Gervais, 1855
1855	<i>Nycticeiini</i>	<i>Nycticeiini</i> Gervais, 1855
1855	<i>Nycticeina</i>	<i>Nycticeiini</i> Gervais, 1855
1855	<i>Emballonurini</i>	<i>Emballonurini</i> Gervais, 1855
1856	<i>Molossidae</i>	<i>MOLOSSIDAE</i> Gervais, 1856
1856	<i>Molossinae</i>	<i>Molossinae</i> Gervais, 1856
1855	<i>Nycticeina</i>	<i>VESPERTILIONIDAE</i> Gray, 1821
1856	<i>Molossoidea</i>	<i>MOLOSSOIDEA</i> Gervais, 1856

Giebel

1855	<i>FRUGIVORA</i>	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
1855	<i>Gymnorhina</i>	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007

Gill

1872	<i>Nycticeijinae</i>	<i>Nycticeiini</i> Gervais, 1855
1872	<i>ANIMALIVORA</i>	<i>CHIROPTERA</i> Blumenbach, 1779
1872	<i>Megadermidae</i>	<i>MEGADERMATIDAE</i> H. Allen, 1864
1872	<i>Molossidae</i>	<i>MOLOSSIDAE</i> Gervais, 1856

Goldfuss

1809	<i>Vespertilio caninus</i> var. <i>b</i>	<i>Eidolon helvum helvum</i> (Kerr, 1792)

Goodman and Cardiff

2004	<i>Chaerephon jobimena</i>	<i>Mops (Chaerephon) jobimena</i> (Goodman and Cardiff, 2004)

Goodman and Ranivo

2008	<i>Triaenops pauliani</i>	<i>Paratriaenops pauliani</i> (Goodman and Ranivo, 2008)
2009	<i>Triaenops menamena</i>	<i>Triaenops menamena</i> Goodman and Ranivo, 2009

Goodman, Bradman, Christides and Appleton

2009	<i>Miniopterus mahafaliensis</i>	<i>Miniopterus mahafaliensis</i> Goodman, Bradman, Christides and Appleton, 2009

Goodman, Bradman, Maminirina, Ryan, Christidis and Appleton

2008	<i>Miniopterus petersoni</i>	<i>Miniopterus petersoni</i> Goodman, Bradman, Maminirina, Ryan, Christidis & Appleton, 2008

Goodman, Buccas, Naidoo, Ratrimomanarivo, Taylor and Lamb

2010	<i>Chaerephon atsinanana</i>	<i>Mops (Chaerephon) atsinanana</i> (Goodman, Buccas, Naidoo, Ratrimomanarivo, Taylor and Lamb, 2010)

Goodman, Cardiff, Ranivo, Russell and Yoder

- 2006 *Emballonura tiavato* *Paremballonura tiavato* (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)

Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst and Bowie

- 2008 *Mormopterus francoismoutoui* *Mormopterus francoismoutoui* Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst, Bowie, 2008

Goodman, Jenkins and Ratrimomanarivo

- 2005 *Scotophilus tandrefana* *Scotophilus tandrefana* Goodman, Jenkins and Ratrimomanarivo, 2005

Goodman, Kearney, Ratsimbazafy and Hassanin

- 2017 *Neoromicia stanleyi* *Laephotis stanleyi* (Goodman, Kearney, Ratsimbazafy and Hassanin, 2017)

Goodman, Maminirina, Bradman, Christidis and Appleton

- 2009 *Miniopterus griffithsi* *Miniopterus griffithsi* Goodman, Maminirina, Bradman, Christidis and Appleton, 2009
 2009 *Miniopterus brachytragos* *Miniopterus brachytragos* Goodman, Maminirina, Bradman, Christidis and Appleton, 2009

Goodman, Maminirina, Weyeneth, Bradman, Christidis, Ruedi and Appleton

- 2009 *Miniopterus aelleni* *Miniopterus aelleni* Goodman, Maminirina, Weyeneth, Bradman, Christidis, Ruedi and Appleton, 2009

Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling

- 2012 *Coleura kibomalandy* *Coleura kibomalandy* Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012
 2012 *Paremballonura* *Paremballonura* Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012
 2012 *Paremballonura atrata* *Paremballonura atrata* (Peters, 1874)
 2012 *Paremballonura tiavato* *Paremballonura tiavato* (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)

Goodman, Rakotondramanana, Ramasindrazana, Kearney, Monadjem, Schoeman, Taylor, Naughton and Appleton

- 2015 *Hypsugo bemaity* *Neoromicia bemaity* (Goodman, Rakotondramanana, Ramasindrazana, Kearney, Monadjem, Schoeman, Taylor, Naughton and Appleton, 2015)

Goodman, Rakotondraparany and Kofoky

- 2007 *Myzopoda schliemanni* *Myzopoda schliemanni* Goodman, Rakotondraparany and Kofoky, 2007

Goodman, Ramasindrazana, Maminirina, Schoeman and Appleton

- 2011 *Miniopterus egeri* *Miniopterus egeri* Goodman, Ramasindrazana, Maminirina, Schoeman and Appleton, 2011

Goodman, Ramasindrazana, Naughton and Appleton

- 2015 *Miniopterus ambohitrensis* *Miniopterus ambohitrensis* Goodman, Ramasindrazana, Naughton and Appleton, 2015

Goodman, Ratrimomanarivo and Randrianandrianina

- 2006 *Scotophilus marovaza* *Scotophilus marovaza* Goodman, Ratrimomanarivo and Randrianandrianina, 2006

Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton

- 2007 *Miniopterus sororculus* *Miniopterus sororculus* Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton, 2007

Goodman, Schoeman, Rakotoarivelo and Willows-Munro

- 2016 *Hipposideros cryptovalorona* *Macronycteris cryptovalorona* (Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016)

Goodman, Taylor, Ratrimomanarivo and Hoofer

- 2012 *Neoromicia robertsi* *Laephotis robertsi* (Goodman, Taylor, Ratrimomanarivo and Hoofer, 2012)

Gough

- 1908 *Rhinolophus swinnyi* *Rhinolophus swinnyi* Gough, 1908

A. Grandidier

- 1869 *Nyctinomus leucogaster* *Mops (Chaerephon) leucogaster* (A. Grandidier, 1869)
 1869 *Nyctinomus Miarensis* *Mops (Mops) midas miarensis* (A. Grandidier, 1869)
 1870 *Nyctinomus unicolor* *Mops (Mops) midas miarensis* (A. Grandidier, 1869)
 1870 *Vespertilio sylvicola* *Myotis goudoti* (A. Smith, 1834)

G. Grandidier

- 1912 *Triænopops aurita* *Paratriaenops auritus* (G. Grandidier, 1912)
 1929 *Rousettus madagascariensis* *Rousettus madagascariensis* G. Grandidier, 1929
 1937 *Nycteris madagascariensis* *Nycteris macrotis* Dobson, 1876

Granvik

- 1924 *Eptesicus loveni* *Myotis tricolor* (Temminck, 1832)

Gray

- 1821 *Barbastella* *Barbastella* Gray, 1821
 1821 *Nyctimones* *Tadarida* Rafinesque, 1814
 1821 *Pteropidæ* *PTEROPODIDAE* Gray, 1821
 1821 *Vespertilionidæ* *VESPERTILIONIDAE* Gray, 1821
 1821 *Vespertilioninae* *Vespertilioninae* Gray, 1821
 1821 *Vespertilionini* *Vespertilionini* Gray, 1821
 1821 *Vespertilionoidea* *VESPERTILIONOIDEA* Gray, 1821
 1821 *Cephalotidæ* *PTEROPODIDAE* Gray, 1821
 1821 *CHEIROPTERA* *CHIROPTERA* Blumenbach, 1779
 1821 *FRUCTIVORAE* *PTEROPODIFORMI* Van Cakenberghe, Kearney and Seamark, 2007
 1821 *INSECTIVORAE* *VESPERTILIONIFORMI* Van Cakenberghe, Kearney and Seamark, 2007
 1821 *Pteropodinae* *Pteropodinae* Gray, 1821
 1821 *Pteropodini* *Pteropodini* Gray, 1821
 1821 *PTEROPODOIDEA* *PTEROPODOIDEA* Gray, 1821
 1821 *Vespertilionini* *Vespertilioninae* Gray, 1821
 1825 *Pteropina* *PTEROPODIDAE* Gray, 1821
 1825 *Vespertilionina* *VESPERTILIONIDAE* Gray, 1821
 1825 *Rhinolophidae* *RHINOLOPHIDAE* Gray, 1825
 1825 *Rhinolophina* *RHINOLOPHOIDEA* J. E. Gray, 1825
 1825 *Rhinolophina* *RHINOLOPHIDAE* Gray, 1825
 1825 *Rhinolophoidea* *RHINOLOPHOIDEA* J. E. Gray, 1825
 1831 *Hipposideros* *Hipposideros* Gray, 1831
 1831 *Pachyotus* *Scotophilus* Leach, 1821
 1834 *Hipposiderus* *Hipposideros* Gray, 1831
 1838 *Asellia* *Asellia* Gray, 1838
 1838 *Barbastellus communis* *Barbastella barbastellus* (Schreber, 1774)
 1838 *Lavia* *Lavia* Gray, 1838
 1838 *Petalia* *Nycteris* G. Cuvier and E. Geoffroy, 1795
 1838 *Plecotus christii* *Plecotus christii* Gray, 1838
 1838 *Romicia* *Pipistrellus* Kaup, 1829
 1842 *Kerivoula* *Kerivoula* Gray, 1842
 1842 *Kerivoula poensis* *Glauconycteris poensis* (Gray, 1842)

1842	<i>Noctulinia</i>	<i>Nyctalus</i> Bowdich, 1825
1842	<i>Trilatitus</i>	<i>Myotis</i> Kaup, 1829
1843	<i>Nycteris poensis</i>	<i>Nycteris hispida</i> (Schreber, 1774)
1843	<i>Vespertilio</i>	<i>Myotis</i> Kaup, 1829
1843	<i>Xantharpyia</i>	<i>Rousettus</i> Gray, 1821
1844	<i>Eleutherura</i>	<i>Rousettus</i> Gray, 1821
1847	<i>Aquias</i>	<i>Rhinolophus</i> Lacépède, 1799
1866	<i>Nyctericina</i>	NYCTERIDAE Van der Hoeven, 1855
1866	<i>Diclidurina</i>	EMBALLONURIDAE Gervais, 1855
1866	<i>Emballonurina</i>	EMBALLONURIDAE Gervais, 1855
1866	<i>Molossina</i>	MOLOSSIDAE Gervais, 1856
1866	<i>Molossina</i>	<i>Molossinae</i> Gervais, 1856
1866	<i>Nyctophilina</i>	VESPERTILIONIDAE Gray, 1821
1866	<i>Pachyomus</i>	<i>Eptesicus</i> Rafinesque, 1820
1866	<i>Plecotina</i>	VESPERTILIONIDAE Gray, 1821
1866	<i>Plecotini</i>	<i>Plecotini</i> Gray, 1866
1866	<i>Romiciana</i>	VESPERTILIONIDAE Gray, 1821
1866	<i>Chrysonycteris</i>	<i>Hipposideros</i> Gray, 1831
1866	<i>Epomophorina</i>	<i>Pteropodinae</i> Gray, 1821
1866	<i>Epomophorini</i>	<i>Epomophorini</i> Gray, 1866
1866	<i>Eunycteris</i>	<i>Pteropus</i> Brisson, 1762
1866	<i>Gloionycteris</i>	<i>Hipposideros</i> Gray, 1831
1866	<i>Macronycteris</i>	<i>Macronycteris</i> Gray, 1866
1866	<i>Megaderma</i>	MEGADERMATIDAE H. Allen, 1864
1866	<i>Nycterops</i>	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795
1866	<i>Nycterops pilosa</i>	<i>Nycteris hispida</i> (Schreber, 1774)
1866	<i>Phyllotis</i>	<i>Rhinolophus</i> Lacépède, 1799
1866	<i>Pteropus livingstonii</i>	<i>Pteropus livingstonii</i> Gray, 1866
1866	<i>Rhinonycterina</i>	RHINONYCTERIDAE J. E. Gray, 1866
1866	<i>Rhinophylla</i>	<i>Hipposideros</i> Gray, 1831
1866	<i>Speorifera</i>	<i>Hipposideros</i> Gray, 1831
1866	<i>Scotophilus welwitschii</i>	<i>Myotis welwitschii</i> (Gray, 1866)
1866	<i>Nycteris Baikii</i>	<i>Nycteris grandis</i> Peters, 1865
1866	<i>Rhinonycteridae</i>	RHINONYCTERIDAE J. E. Gray, 1866
1866	<i>Scotophilina</i>	<i>Scotophilinae</i> Van Cakenberghe and Seamark, 2008
1866	<i>Spasma</i>	<i>Megaderma</i> E. Geoffroy St.-Hilaire, 1810
1870	[<i>Epomophorus macrocephalus</i>] var. <i>angolensis</i>	<i>Epomophorus angolensis</i> Gray, 1870
1870	[<i>Epomophorus macrocephalus</i>] var. <i>unicolor</i>	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
1870	<i>Cynopterus collaris</i>	<i>Myonycteris torquata</i> (Dobson, 1878)
1870	<i>Eleutherura unicolor</i>	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1870	<i>Epomophorus macrocephalus</i> var. <i>unicolor</i>	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
1870	<i>Epomops</i>	<i>Epomops</i> Gray, 1866
1870	<i>Pselaphon</i>	<i>Pteropus</i> Brisson, 1762
1870	<i>Senonycteris</i>	<i>Rousettus</i> Gray, 1821
1870	<i>Spectrum vulgare</i>	<i>Pteropus niger</i> (Kerr, 1792)
1870	<i>Stenonycteris</i>	<i>Stenonycteris</i> Gray, 1870
1821	<i>Rousettus</i>	<i>Rousettus</i> Gray, 1821

Guillén-Servent, Francis and Ricklefs

2003	<i>Indorhinolophus</i>	<i>Rhinolophus</i> Lacépède, 1799

Gunnell and Manthi

2018	<i>Mops kerio</i>	<i>Mops kerio</i> Gunnell and Manthi, 2018
2018	<i>Mops turkwellensis</i>	<i>Mops turkwellensis</i> Gunnell and Manthi, 2018
2018	<i>R[ousettus] pattersoni</i>	<i>Rousettus pattersoni</i> Gunnell and Manthi, 2018
2018	<i>Saccolaimus kenyensis</i>	<i>Saccolaimus kenyensis</i> Gunnell and Manthi, 2018
2018	<i>Turkanycteris</i>	<i>Turkanycteris</i> Gunnell and Manthi, 2018
2018	<i>Turkanycteris harrisi</i>	<i>Turkanycteris harrisi</i> Gunnell and Manthi, 2018

Gunnell, Butler, Greenwood and Simmons

2015	<i>Cardioderma leakeyi</i>	<i>Cardioderma leakeyi</i> Gunnell, Butler, Greenwood and Simmons, 2015
2015	<i>Myzopoda africana</i>	<i>Myzopoda africana</i> Gunnell, Butler, Greenwood and Simmons, 2015
2015	<i>Nycticeinops serengetiensis</i>	<i>Nycticeinops serengetiensis</i> Gunnell, Butler, Greenwood and Simmons, 2015
2015	<i>Scotoecus olduvensis</i>	<i>Scotoecus olduvensis</i> Gunnell, Butler, Greenwood and Simmons, 2015

Gunnell, Eiting and Geraads

2011	<i>Miniopterus horaceki</i>	<i>Miniopterus horaceki</i> Gunnell, Eiting and Geraads, 2011
2011	<i>Myotis darelbeidensis</i>	<i>Myotis darelbeidensis</i> Gunnell, Eiting and Geraads, 2011
2011	<i>Rhinolophus maghrebensis</i>	<i>Rhinolophus maghrebensis</i> Gunnell, Eiting and Geraads, 2011

Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch

2003	<i>Tanzanycterididae</i>	<i>TANZANYCTERIDIDAE</i> Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003
2003	<i>Tanzanycteris</i>	<i>Tanzanycteris</i> Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003
2003	<i>Tanzanycteris mannardi</i>	<i>Tanzanycteris mannardi</i> Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003

Gunnell, Simmons and Seiffert

2014	<i>Phasmatonycteris</i>	<i>Phasmatonycteris</i> Gunnell, Simmons and Seiffert, 2014
2014	<i>Phasmatonycteris butleri</i>	<i>Phasmatonycteris butleri</i> Gunnell, Simmons and Seiffert, 2014
2014	<i>Phasmatonycteris phiomensis</i>	<i>Phasmatonycteris phiomensis</i> Gunnell, Simmons and Seiffert, 2014

Gunnell, Simons and Seiffert

2008	<i>Dhofarella sigei</i>	<i>Dhofarella sigei</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Khonsunysteris</i>	<i>Khonsunysteris</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Khonsunysteris aegypticus</i>	<i>Khonsunysteris aegypticus</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Qarunysteris</i>	<i>Qarunysteris</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Qarunysteris moerisae</i>	<i>Qarunysteris moerisae</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Saharaderma</i>	<i>Saharaderma</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Saharaderma pseudovampyrus</i>	<i>Saharaderma pseudovampyrus</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Witwatia</i>	<i>Witwatia</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Witwatia eremicus</i>	<i>Witwatia eremicus</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Witwatia schlosseri</i>	<i>Witwatia schlosseri</i> Gunnell, Simons and Seiffert, 2008

Gunnell, Winkler, Miller, Head, El-Barkooky, Gawad, Sanders and Gingerich

2015	<i>Hipposideros (Pseudorhinolophus) amenhotepos</i>	<i>Hipposideros amenhotepos</i> Gunnell, Winkler, Miller, Head, El-Barkooky, Gawad, Sanders and Gingerich, 2015

C. H. Smith

1842	<i>Harpyidae</i>	<i>PTEROPODIDAE</i> Gray, 1821

Haeckel

1866	<i>Histiorrhina</i>	<i>RHINOLOPHIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
1866	<i>Hypodermida</i>	<i>PTEROPODIDAE</i> Gray, 1821
1866	<i>Nycterides</i>	<i>CHIROPTERA</i> Blumenbach, 1779
1866	<i>Pterocynes</i>	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
1866	<i>Pteropodida</i>	<i>PTEROPODIDAE</i> Gray, 1821

Hall and Kelson

1959	<i>Nyctiptennis</i>	<i>Eptesicus Rafinesque</i> , 1820

Hallowell

1846	<i>Pteropus Haldemani</i>	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)

Hand and Archer

2005 *Brevipalatus* *Brevipalatus* Hand and Archer, 2005

M. Happold

2005 *Myotis dieteri* *Myotis dieteri* M. Happold, 2005

Harrison

1956 *Platymops barbatogularis* *Platymops setiger macmillani* Thomas, 1906
 1958 *Taphozous perforatus swirae* *Taphozous perforatus perforatus* E. Geoffroy St.-Hilaire, 1818
 1959 *Miniopterus minor griveaudi* *Miniopterus griveaudi* Harrison, 1959
 1962 *Taphozous sudani australis* *Taphozous perforatus sudani* Thomas, 1915
 1964 *Taphozous sudani rhodesiae* *Taphozous perforatus sudani* Thomas, 1915
 1975 *Tadarida (Chaerephon) gallagheri* *Mops (Chaerephon) gallagheri* (Harrison, 1975)

Harrison and Fleetwood

1960 *Platymops barbatogularis parkeri* *Platymops setiger macmillani* Thomas, 1906

Hartmann

1868 *Nycteris Geoffroyi* Var. *Senegalensis* *Nycteris thebaica* E. Geoffroy St.-Hilaire, 1818
 1868 *Rhinopoma sennarens* *Rhinopoma cystops* Thomas, 1903

Hassanin

2014 *Casinycteris campomaanensis* *Casinycteris campomaanensis* Hassanin, 2014

Hassanin, Colombo, Gembu, Merle, Tu, Görföl, Musaba Akawa, Csorba, Kearney, Monadjem and Ing

2017 *Glauconycteris atra* *Glauconycteris atra* Hassanin, Colombo, Gembu, Merle, Tu, Görföl, Musaba Akawa, Csorba, Kearney, Monadjem and Ing, 2017

Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo

2015 *Scotonycteris bergmansi* *Scotonycteris bergmansi* Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015
 2015 *Scotonycteris bergmansi bergmansi* *Scotonycteris bergmansi bergmansi* Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015
 2015 *Scotonycteris bergmansi congoensis* *Scotonycteris bergmansi congoensis* Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015

Hatt

1928 *Chærophon (Lophomops) nigri* *Mops (Chaerephon) leucogaster* (A. Grandidier, 1869)

Hayman

1937 *R[hinopoma] cystops macinnesi* *Rhinopoma macinnesi* Hayman, 1937
 1938 *Chærephon lancasteri* *Mops (Chaerephon) chapini* (J.A. Allen, 1917)
 1939 *Glauconycteris superba* *Glauconycteris superba* Hayman, 1939
 1940 *Mops calabarensis* *Mops (Xiphonycteris) nanulus* J.A. Allen, 1917
 1946 *Scotonycteris ophiodon cansdalei* *Casinycteris ophiodon* (Pohle, 1943)
 1947 *Scotonycteris zenkeri occidentalis* *Scotonycteris occidentalis* Hayman, 1947
 1947 *Glauconycteris superba sheila* *Glauconycteris superba* Hayman, 1939
 1947 *Hipposideros jonesi* *Hipposideros jonesi* Hayman, 1947
 1951 *Tadarida (Chaerephon) faini* *Mops (Chaerephon) pumilus* (Cretzschmar, 1826)
 1963 *Glauconycteris machadoi* *Glauconycteris machadoi* Hayman, 1963
 1963 *Micropteropus intermedius* *Epomophorus intermedius* (Hayman, 1963)
 1966 *Megaloglossus woermanni prigoginei* *Megaloglossus woermanni* Pagenstecher, 1885

Heim de Balsac

1934	<i>Rhinolophus acrotis schwarzi</i>	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1936	<i>Pipistrellus kuhlii pallidus</i>	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)

Heller

1912	<i>Miniopterus natalensis arenarius</i>	<i>Miniopterus arenarius</i> Heller, 1912
1912	<i>Pipistrellus aero</i>	<i>Pipistrellus (Pipistrellus) aero</i> Heller, 1912
1912	<i>Pipistrellus helios</i>	<i>Afronycteris helios</i> (Heller, 1912)

Herkt, Skidmore and Fahr

2017	<i>Pipistrellus krugeri</i>	<i>Laephotis stanleyi</i> (Goodman, Kearney, Ratsimbazafy and Hassanin, 2017)

Hermann

1804	<i>V[espertilio] acetabulosus</i>	<i>Mormopterus acetabulosus</i> (Hermann, 1804)
1804	<i>V[espertilio] acetabulosus</i>	<i>Mormopterus acetabulosus acetabulosus</i> (Hermann, 1804)
1804	<i>V[espertilio] mauritanus</i>	<i>Pteropus niger</i> (Kerr, 1792)

Heuglin

1861	<i>Nycteris labiata</i>	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1861	<i>Nycticejus flavigaster</i>	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
1861	<i>Nycticejus murino-flavus</i>	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
1861	<i>Nyctinomus (Dysopes) ventralis</i>	<i>Tadarida ventralis</i> (Heuglin, 1861)
1861	<i>Nyctinomus bivittatus</i>	<i>Mops (Chaerephon) bivittatus</i> (Heuglin, 1861)
1861	<i>Phyllorrhina bicornis</i>	<i>Hipposideros tephurus</i> (Cabrera, 1906)
1861	<i>Phyllorrhina megalotis</i>	<i>Hipposideros megalotis</i> (Heuglin, 1861)
1861	<i>Rhinolophus acrotis</i>	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1861	<i>Rhinolophus miminus</i>	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1864	<i>Dysopes hepaticus</i>	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
1864	<i>Epomophorus anurus</i>	<i>Epomophorus labiatus</i> (Temminck, 1837)
1865	<i>Pteropus palmarum</i>	<i>Eidolon helvum helvum</i> (Kerr, 1792)
1866	<i>Plecotus aethiopicus</i>	<i>Plecotus christii</i> Gray, 1838
1866	<i>Xantharpyia leucomelas</i>	<i>Eidolon helvum</i> (Kerr, 1792)
1877	<i>Dysopes talpinus</i>	<i>Tadarida aegyptiaca aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
1877	<i>N[yc]ticejus adovanus</i>	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
1877	<i>Nycticejus eriophorus</i>	<i>Kerivoula eriophora</i> (Heuglin, 1877)
1877	<i>Nycticejus serratus</i>	<i>Taphozous nudiventris</i> Cretzschmar, 1830
1877	<i>Rhinolophus macrocephalus</i>	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1877	<i>Rhinopoma cordofanicum</i>	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
1877	<i>Taphozous maritimus</i>	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
1877	<i>Vesperugo senarensis</i>	<i>Vansonia rueppellii rueppellii</i> (J.B. Fischer, 1829)

Heuglin and Fitzinger

1866	<i>Plecotus aethiopicus</i>	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1866	<i>Vesperugo hypoleucus</i>	<i>Vansonia rueppellii rueppellii</i> (J.B. Fischer, 1829)
1866	<i>Vesperugo sennaariensis</i>	<i>Vansonia rueppellii rueppellii</i> (J.B. Fischer, 1829)

Hewitt

1913	<i>Rhinolophus swinnyi piriensis</i>	<i>Rhinolophus swinnyi</i> Gough, 1908

Hill

1968	<i>Pipistrellus eisentrauti</i>	<i>Nycticeinops eisentrauti</i> (Hill, 1968)
1971	<i>Myotis morrisi</i>	<i>Myotis morrisi</i> Hill, 1971

Hill and Harrison

1987	<i>Scotophilini</i>	<i>Scotophilinae</i> Van Cakenberghe and Seamark, 2008
1987	<i>Nycticeinops</i>	<i>Nycticeinops</i> Hill & Harrison, 1987

Hill and Morris

1971	<i>Epomophorus</i> sp.	<i>Epomophorus labiatus</i> (Temminck, 1837)

J.Eric Hill

1937	<i>Eptesicus capensis angolensis</i>	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
1937	<i>Mops chitauensis</i>	<i>Mops (Mops) niveiventer</i> Cabrera and Ruxton, 1926

Hinton

1920	<i>Kerivoula lucia</i>	<i>Kerivoula lanosa</i> (A. Smith, 1847)
1920	<i>Mimetillus thomasi</i>	<i>Mimetillus thomasi</i> Hinton, 1920

Hodgson

1847	<i>Eucheira</i>	<i>Megaderma</i> E. Geoffroy St.-Hilaire, 1810

Hollister

1916	<i>Chaerephon pumilus naivashæ</i>	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1916	<i>Eptesicus ugandæ</i>	<i>Neoromicia somalica</i> (Thomas, 1901)
1916	<i>Rhinolophus kenienensis</i>	<i>Rhinolophus clivosus</i> Cretzschmar, 1828

Horáček, Fejfar and Hulva

2006	<i>Philisinae</i>	<i>Scotophilinae</i> Van Cakenberghe and Seamark, 2008
2006	<i>Scotophilis</i>	<i>Scotophilis</i> Horáček, Fejfar and Hulva, 2006
2006	<i>Scotophilis libycus</i>	<i>Scotophilis libycus</i> Horáček, Fejfar and Hulva, 2006

Horsfield

1823-1824	<i>Nyctinomus mauritanus</i>	<i>Mormopterus acetabulosus acetabulosus</i> (Hermann, 1804)

Hulva and Benda

2004	<i>Pipistrellus hanaki</i>	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004

Hutcheon and Kirsch

2004	<i>PTEROPODIFORMES</i>	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
2004	<i>VESPERTILIONIFORMES</i>	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007

Hutterer and Kerbis Peterhans

2020	<i>Parahypsugo macrocephalus</i>	<i>Nycticeinops macrocephalus</i> (Hutterer and Kerbis Peterhans, 2020)

Hutterer, Decher, Monadjem and Astrin

2019	<i>Parahypsugo</i>	<i>Nycticeinops</i> Hill & Harrison, 1987
2019	<i>Parahypsugo happoldorum</i>	<i>Nycticeinops happoldorum</i> (Hutterer, Decher, Monadjem and Astrin, 2019)

Ibáñez, Juste, Salicini, Puechmaille and Ruedi

2019	<i>Myotis zenatius</i>	<i>Myotis zenatius</i> Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019

Illiger

1815	<i>Pteropus collaris</i>	<i>Pteropus subniger</i> (Kerr, 1792)
1815	<i>Pteropus flavus</i>	<i>Eidolon helvum helvum</i> (Kerr, 1792)

Iredale and Troughton

1934	<i>Austronomus</i>	<i>Tadarida</i> Rafinesque, 1814

Jameson

1909	<i>Miniopterus breyeri</i>	<i>Miniopterus natalensis</i> (A. Smith, 1833)

Jentink

1879	<i>Nyctinomus Bemmeleni</i>	<i>Mops (Chaerephon) bemmeleni</i> (Jentink, 1879)
1879	<i>Taphozous Dobsoni</i>	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
1881	<i>Leiponyx</i>	<i>Eidolon Rafinesque</i> , 1815
1881	<i>Leiponyx büttikoferi</i>	<i>Eidolon helvum helvum</i> (Kerr, 1792)
1888	<i>Epomophorus veldkampii</i>	<i>Nanonycteris veldkampii</i> (Jentink, 1888)
1888	<i>Vesperugo stampflii</i>	<i>Afronycteris nana</i> (Peters, 1852)
1904	<i>Kehelvoulha</i>	<i>Kerivoula</i> Gray, 1842
1910	<i>Chrysopteron</i>	<i>Myotis</i> Kaup, 1829

Jerdon

1867	<i>Taphozoinae</i>	<i>Taphozoinae</i> Jerdon, 1867

Juste and Ibáñez

1992	<i>Miniopterus minor occidentalis</i>	<i>Miniopterus minor</i> Peters, 1867
1993	<i>Rousettus aegyptiacus princeps</i>	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1993	<i>Tadarida (Chaerephon) tomensis</i>	<i>Mops (Chaerephon) tomensis</i> (Juste and Ibáñez, 1993)
1993	<i>Rousettus aegyptiacus tomensis</i>	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)

Juste, Ibáñez and Machordom

2000	<i>Eidolon helvum annobonensis</i>	<i>Eidolon helvum helvum</i> (Kerr, 1792)

Kaup

1829	<i>Barbastellus</i>	<i>Barbastella</i> Gray, 1821
1829	<i>Cnephaeus</i>	<i>Eptesicus (Cnephaeus)</i> Kaup, 1829
1829	<i>Myotis</i>	<i>Myotis</i> Kaup, 1829
1829	<i>Nystactes</i>	<i>Myotis</i> Kaup, 1829
1829	<i>Pipistrellus</i>	<i>Pipistrellus</i> Kaup, 1829
1829	<i>Pterygistes</i>	<i>Nyctalus</i> Bowdich, 1825

Kerbis Peterhans and Fahr

2013	<i>Rhinolophus willardi</i>	<i>Rhinolophus willardi</i> Kerbis Peterhans and Fahr, 2013

Kerr

1792	<i>minor</i>	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1792	<i>Vespertilio vampirus niger</i>	<i>Pteropus niger</i> (Kerr, 1792)
1792	<i>Vespertilio vampyrus helvus</i>	<i>Eidolon helvum helvum</i> (Kerr, 1792)
1792	<i>Vespertilio vampyrus helvus</i>	<i>Eidolon helvum</i> (Kerr, 1792)
1792	<i>Vespertilio vampyrus subniger</i>	<i>Pteropus subniger</i> (Kerr, 1792)
1792	<i>Vespertilio ferrum-equinum major</i>	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)

Kershaw

1922	<i>Kerivoula lueia</i>	<i>Kerivoula lanosa</i> (A. Smith, 1847)
1922	<i>Nycteris oriana</i>	<i>Nycteris macrotis</i> Dobson, 1876
1923	<i>Nycteris marica</i>	<i>Nycteris grandis</i> Peters, 1865

Keyserling and Blasius

1839	<i>Synotus</i>	<i>Barbastella</i> Gray, 1821
1839	<i>Vesperugo</i>	<i>Eptesicus</i> Rafinesque, 1820
1839	<i>Vesperugo</i>	<i>Nyctalus</i> Bowdich, 1825
1839	<i>Vesperugo</i>	<i>Pipistrellus</i> Kaup, 1829
1839	<i>Vesperus</i>	<i>Eptesicus</i> Rafinesque, 1820

Kinman

1994	<i>Chiropteriformes</i>	<i>CHIROPTERA</i> Blumenbach, 1779

Kirk

1865	<i>Nycticejus nidicola</i>	<i>Kerivoula argentata</i> Tomes, 1861

Kishida and Mori

1931 *Tuitatus* *Eptesicus Rafinesque, 1820*

Koch

1860 *Phyllorhina* *HIPPOSIDERIDAE Lydekker, 1891*
 1860 *Phyllorrhina* *HIPPOSIDERIDAE Lydekker, 1891*

Kock

1969 *Rhinopoma hardwickei sennaariense* *Rhinopoma cystops Thomas, 1903*
 1969 *Rhinopoma microphyllum tropicalis* *Rhinopoma microphyllum (Brünnich, 1782)*
 1978 *Rousettus (Rousettus) obliviosus* *Rousettus obliviosus Kock, 1978*
 2001 *Pipistrellus africanus meesteri* *Afronycteris nana (Peters, 1852)*

Kock, Csorba and Howell

2000 *Rhinolophus maendeleo* *Rhinolophus maendeleo Kock, Csorba and Howell, 2000*

Kolenati

1856 *Brachyotus* *Myotis Kaup, 1829*
 1856 *Cateorus* *Eptesicus Rafinesque, 1820*
 1856 *Hypsugo* *Hypsugo Kolenati, 1856*
 1856 *Isotus* *Myotis Kaup, 1829*
 1856 *Meteoros* *Eptesicus Rafinesque, 1820*
 1856 *Nannugo* *Pipistrellus Kaup, 1829*
 1856 *Panugo* *Nyctalus Bowdich, 1825*
 1858 *Amblyotus* *Eptesicus Rafinesque, 1820*
 1863 *Aristippe* *Eptesicus Rafinesque, 1820*

Koopman

1985 *YANGOCHIROPTERA* *VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007*
 1985 *Yinochiroptera* *CHIROPTERA Blumenbach, 1779*
 1989 *Rhinolophus clivosus hillorum* *Rhinolophus hillorum Koopman, 1989*
 1994 *Hipposiderina* *HIPPOSIDERIDAE Lydekker, 1891*

Koopman and J.K. Jones Jr.

1970 *Cynopterini* *Pteropodinae Gray, 1821*
 1970 *Hipposiderini* *HIPPOSIDERIDAE Lydekker, 1891*
 1970 *Rousettina* *Rousettinae Andersen, 1912*

Kruskop and Lavrenchenko

2000 *Plecotus balensis* *Plecotus balensis Kruskop & Lavrenchenko, 2000*

Kruskop, Benda, Vasenkov and Lavrenchenko

2016 *Nycteris cf. parisii* *Nycteris cf. parisii De Beaux, 1924*

Kuhl

1817 *Vespertilio Kuhlii* *Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)*
 1817 *Vespertilio Leisleri* *Nyctalus leisleri (Kuhl, 1817)*
 1817 *Vespertilio mystacinus* *Myotis mystacinus (Kuhl, 1817)*
 1817 *Vespertilio schreibersii* *Miniopterus schreibersii (Kuhl, 1817)*
 1819 *Vespertilio dasykarpus* *Nyctalus leisleri (Kuhl, 1817)*

Lacépède

1799 *Rhinolophus* *Rhinolophus Lacépède, 1799*
 1799 *Spectrum* *Pteropus Brisson, 1762*

Lataste

1887 *Vesperugo (Vesperus) Innesi* *Eptesicus (Cnephaeus) bottae (Peters, 1869)*

Laurent

1936	<i>Plecotus auritus saharæ</i>	<i>Otonycteris hemprichii</i> Peters, 1859
1937	<i>Rhinolophus hipposideros vespa</i>	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1937	<i>[Pipistrellus Kuhl] latastei</i>	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
1937	<i>Asellia tridens pallida</i>	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)

Lavocat

1961	<i>Afropterus</i>	<i>Megaderma</i> E. Geoffroy St.-Hilaire, 1810
1961	<i>Afropterus gigas</i>	<i>Megaderma gigas</i> (Lavocat, 1961)
1961	<i>Afropterus gigas</i>	<i>Megaderma gaillardi</i> (Trouessart, 1898)
1961	<i>Asellia vetus</i>	<i>Hipposideros vetus</i> (Lavocat, 1961)
1961	<i>Rhinolophus ferrumequinum mellali</i>	<i>Rhinolophus mellali</i> Lavocat, 1961

Lawrence and Novick

1963	<i>Myonycterini</i>	<i>Myonycterini</i> Lawrence and Novick, 1963

Leach

1816	<i>Macrotus</i>	<i>Plecotus</i> E. Geoffroy St.-Hilaire, 1818
1816	<i>Phyllorhina</i>	<i>Rhinolophus</i> Lacépède, 1799
1821	<i>Scotophilus</i>	<i>Scotophilus</i> Leach, 1821
1825	<i>Vespertilio Pygmæus</i>	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)

Leconte

1857	<i>Vespertilio pusillus</i>	<i>Laephotis "incertae-sedis"</i>

Legendre

1984	<i>Cheiomelinae</i>	<i>Molossinae</i> Gervais, 1856
1984	<i>Rhizomops</i>	<i>Tadarida</i> Rafinesque, 1814

Lesson

1827	<i>Rhinolophina</i>	<i>RHINOLOPHIDAE</i> Gray, 1825
1842	<i>Mops</i>	<i>Mops (Mops)</i> Lesson, 1842
1842	<i>Mops</i>	<i>Mops</i> Lesson, 1842

Lichtenstein

1823	<i>Pteropus collaris</i>	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1823	<i>Rhinolophus capensis</i>	<i>Rhinolophus capensis</i> Lichtenstein, 1823

Loche

1867	<i>Rhinolophus algirus</i>	<i>Rhinolophus euryale</i> Blasius, 1853

Lönnberg

1908	<i>Rousettus sjöstedti</i>	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)

Lönnberg and Gyldenstolpe

1925	<i>Nycteris proxima</i>	<i>Nycteris grandis</i> Peters, 1865

Lydekker

1888	<i>Necromanter</i>	<i>Necromantis</i> Weithofer, 1887
1891	<i>Hipposideridae</i>	<i>HIPPOSIDERIDAE</i> Lydekker, 1891
1891	<i>Hipposiderinae</i>	<i>HIPPOSIDERIDAE</i> Lydekker, 1891
1891	<i>Trygenycteris</i>	<i>Megaloglossus</i> Pagenstecher, 1885

Macalister

9999	<i>Noctulina</i>	<i>Nyctalus</i> Bowdich, 1825

Mamba, Dalton, Mahlaba, Kropff and Monadjem

2020	<i>Hipposideros cf. ruber</i> (lineage D1)	<i>Hipposideros ruber</i> (Noack, 1893)
Martin		
1838	<i>Rhinolophus landeri</i>	<i>Rhinolophus landeri</i> Martin, 1838
Mason		
1907	<i>Pteropus mascarinus</i>	<i>Pteropus rodricensis</i> Dobson, 1878
Matschie		
1894	<i>Scotonycteris</i>	<i>Scotonycteris</i> Matschie, 1894
1894	<i>Scotonycteris zenkeri</i>	<i>Scotonycteris zenkeri</i> Matschie, 1894
1897	<i>Nyctinomus martiensseni</i>	<i>Otomops martiensseni martiensseni</i> (Matschie, 1897)
1897	<i>Nyctinomus martiensseni</i>	<i>Otomops martiensseni</i> (Matschie, 1897)
1899	[<i>Epomophorus</i> (<i>Nanonycteris</i>)]	<i>Nanonycteris</i> Matschie, 1899
1899	[<i>Xantharpyia</i> (<i>Myonycteris</i>)]	<i>Myonycteris</i> Matschie, 1899
1899	<i>Ep[omophorus</i> (<i>Epomophorus</i>)] <i>büttikoferi</i>	<i>Epomops buettikoferi</i> (Matschie, 1899)
1899	<i>Ep[omophorus</i> (<i>Epomophorus</i>)] <i>doriae</i>	<i>Epomophorus labiatus</i> (Temminck, 1837)
1899	<i>Ep[omophorus</i> (<i>Epomophorus</i>)] <i>neumanni</i>	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
1899	<i>Ep[omophorus</i> (<i>Epomophorus</i>)] <i>stuhlmanni</i>	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
1899	<i>Ep[omophorus</i> (<i>Epomophorus</i>)] <i>zechi</i>	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
1899	<i>Ep[omophorus</i> (<i>Epomophorus</i>)] <i>zenkeri</i>	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
1899	<i>Hypsignathus haldemani</i>	<i>Hypsignathus monstrosus</i> H. Allen, 1862
1899	<i>Mynonycteris</i>	<i>Rousettus</i> Gray, 1821
1899	<i>Sericonycteris</i>	<i>Pteropus</i> Brisson, 1762
1899	<i>Vespertilio venustus</i>	<i>Myotis welwitschii</i> (Gray, 1866)
1901	<i>Euryalus</i>	<i>Rhinolophus</i> Lacépède, 1799
1901	<i>Rhinolophus mehelyi</i>	<i>Rhinolophus mehelyi</i> Matschie, 1901
1909	<i>Pteropus (Spectrum) voeltzkowi</i>	<i>Pteropus voeltzkowi</i> Matschie, 1909
Menu		
1987	<i>Eptésiformes</i>	<i>Vespertilioninae</i> Gray, 1821
1987	<i>Leuconoformes</i>	<i>Myotinae</i> Tate, 1942
1987	<i>Leuconoïdes</i>	<i>Vespertilioninae</i> Gray, 1821
1987	<i>Néoeptésiformes</i>	<i>Vespertilioninae</i> Gray, 1821
1987	<i>Nyctaliiformes</i>	<i>Vespertilioninae</i> Gray, 1821
1987	<i>Nyctaloïdes</i>	<i>Vespertilioninae</i> Gray, 1821
1987	<i>Attalepharca</i>	<i>Pipistrellus</i> Kaup, 1829
1987	<i>Nycterikaupius</i>	<i>Pseudoromicia</i> Monadjem, Patterson, Webala and Demos, 2020
1987	<i>Nycterikaupius</i>	<i>Neoromicia</i> Roberts, 1926
1987	<i>Nycterikaupius</i>	<i>Laephotis</i> Thomas, 1901
1987	<i>Nycterikaupius</i>	<i>Pipistrellus</i> Kaup, 1829
Merriam		
1895	<i>Pteropus pteropus</i>	<i>Pteropus niger</i> (Kerr, 1792)
Michahelles		
1839	<i>Vespertilio pachygnathus</i>	<i>Nyctalus leisleri</i> (Kuhl, 1817)
Miller		
1897	<i>Plecotinæ</i>	<i>Plecotini</i> Gray, 1866
1897	<i>Vespertilioninæ</i>	<i>Vespertilioninae</i> Gray, 1821
1900	<i>Pipistrellus minusculus</i>	<i>Afronycteris nana</i> (Peters, 1852)
1902	<i>Nyctinomus pusillus</i>	<i>Mops (Chaerephon) pusillus</i> (Miller, 1902)
1905	<i>Lavia rex</i>	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
1906	<i>Pizonyx</i>	<i>Myotis</i> Kaup, 1829
1906	<i>Rhinopterus</i>	<i>Eptesicus (Rhinopterus)</i> Miller, 1906
1907	<i>Desmaplex</i>	<i>Pteropus</i> Brisson, 1762
1907	<i>Diclidurinae</i>	<i>EMBALLONURIDAE</i> Gervais, 1855
1907	<i>Hipposideridae</i>	<i>HIPPOSIDERIDAE</i> Lydekker, 1891
1907	<i>Kerivoulinae</i>	<i>Kerivoulinae</i> Miller, 1907
1907	<i>Nyctophilinae</i>	<i>VESPERTILIONIDAE</i> Gray, 1821
1907	<i>Rhinopomidae</i>	<i>RHINOPOMATIDAE</i> Dobson, 1872

- 1910 *Petaliidæ* NYCTERIDAE Van der Hoeven, 1855
- A. Milne-Edwards and A. Grandidier
- 1878 *Myzopoda* *Myzopoda* Milne-Edwards and A. Grandidier, 1878
 1878 *Myzopoda aurita* *Myzopoda aurita* Milne-Edwards and A. Grandidier, 1878
- A. Milne-Edwards
- 1877 *Pteropus seychellensis* *Pteropus seychellensis* A. Milne-Edwards, 1877
 1881 *Scotophilus robustus* *Scotophilus robustus* A. Milne-Edwards, 1881
 1881 *Triænopis Humbloti* *Triænopis menamena* Goodman and Ranivo, 2009
 1881 *Triænopis rufus* *Triænopis menamena* Goodman and Ranivo, 2009
 1881 *Vesperus Humbloti* *Laephotis humbloti* (A. Milne-Edwards, 1881)
- Mina-Palumbo
- 1868 *V[espertilio noctula] var. sicula* *Nyctalus lasiopterus* (Schreber, 1780)
- Mishra, Rout and Panda
- 2011 *Pterocarpus* *Pteropus* Brisson, 1762
- Monadjem, Demos, Dalton, Webala, Musila, Kerbis Peterhans and Patterson
- 2020 *Nycticeinops bellieri* *Nycticeinops bellieri* (De Vree, 1972)
- Monadjem, Goodman, Stanley and Appleton
- 2013 *Miniopterus mossambicus* *Miniopterus mossambicus* Monadjem, Goodman, Stanley and Appleton, 2013
- Monadjem, Guyton, Naskrecki, Richards, Kropff and Dalton
- 2020 *Miniopterus wilsoni* *Miniopterus wilsoni* Monadjem, Guyton, Naskrecki, Richards, Kropff and Dalton, 2020
- Monadjem, Kerbis Peterhans, Nalikka, Waswa, Demos and Patterson
- 2020 *Pseudoromicia kityoi* *Pseudoromicia kityoi* Monadjem, Kerbis Peterhans, Nalikka, Waswa, Demos and Patterson, 2020
- Monadjem, Patterson and Demos
- 2020 *Afronycteris* *Afronycteris* Monadjem, Patterson and Demos, 2020
- Monadjem, Patterson, Webala and Demos
- 2020 *Laephotis kirinyaga* *Laephotis kirinyaga* Monadjem, Patterson, Webala and Demos, 2020
 2020 *Pseudoromicia* *Pseudoromicia* Monadjem, Patterson, Webala and Demos, 2020
 2020 *Pseudoromicia nyanza* *Pseudoromicia nyanza* Monadjem, Patterson, Webala and Demos, 2020
- Monadjem, Richards and Denys
- 2016 *Hipposideros cf. caffer* *Hipposideros caffer* (Sundevall, 1846)
 2016 *Hipposideros cf. ruber (lineage C1)* *Hipposideros ruber* (Noack, 1893)
 2016 *Hipposideros cf. ruber (lineage E1)* *Hipposideros ruber* (Noack, 1893)
- Monadjem, Richards, Decher and Hutterer
- 2020 *Pipistrellus simandouensis* *Pipistrellus (Pipistrellus) simandouensis* Monadjem, Richards, Decher and Hutterer, 2020
- Monadjem, Richards, Taylor and Stoffberg
- 2013 *Neoromicia roseveari* *Pseudoromicia roseveari* (Monadjem, Richards, Taylor and Stoffberg, 2013)

Monadjem, Shapiro, Richards, Karabulut, Crawley, Broman Nielsen, Hansen, Bohmann and Mourier

2020	<i>Miniopterus cf. inflatus</i>	<i>Miniopterus cf. inflatus</i> Monadjem, Shapiro, Richards, Karabulut, Crawley, Broman Nielsen, Hansen, Bohmann and Mourier, 2020

Monadjem, Shapiro, Richards, Karabulut, Crawley, Nielsen, Hansen, Bohmann and Mourier

2020	<i>Miniopterus nimbae</i>	<i>Miniopterus nimbae</i> Monadjem, Shapiro, Richards, Karabulut, Crawley, Nielsen, Hansen, Bohmann and Mourier, 2020

Monard

1933	<i>Mimetillus bernerii</i>	<i>Mimetillus thomasi</i> Hinton, 1920
1933	<i>Nyctinomus spillmanni</i>	<i>Mops (Chaerephon) nigeriae spillmanni</i> (Monard, 1933)
1933	<i>Pipistrellus leucomelas</i>	<i>Vansonia rueppellii vernayi</i> (Roberts, 1932)
1935	<i>Laephotis angolensis</i>	<i>Laephotis angolensis</i> Monard, 1935
1939	<i>Coleura kummeri</i>	<i>Coleura afra</i> (Peters, 1852)
1939	<i>Hipposideros braima</i>	<i>Hipposideros tephros</i> (Cabrera, 1906)
1939	<i>Hipposideros gigas viegasi</i>	<i>Macronycteris gigas</i> (Wagner, 1845)
1939	<i>Mops osborni occidentalis</i>	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
1939	<i>Mops osborni occidentalis f. fulva</i>	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
1939	<i>Nycteris aethiopica guineensis</i>	<i>Nycteris macrotis</i> Dobson, 1876
1939	<i>Nycteris aethiopica guineensis f. aurantiaca</i>	<i>Nycteris macrotis</i> Dobson, 1876

Monticelli

1885	<i>Taphozous perforatus</i> var. <i>assabensis</i>	<i>Taphozous nudiventris</i> Cretzschmar, 1830

Müller

1858	<i>Nannugo Kolenatii</i>	<i>Pipistrellus (Pipistrellus) "incertae-sedis"</i>

A. Murray

1862	<i>Sphyrocephalus</i>	<i>Hypsignathus</i> H. Allen, 1862
1862	<i>Sphyrocephalus labrosus</i>	<i>Hypsignathus monstrosus</i> H. Allen, 1862
1862	<i>Zygænocephalus</i>	<i>Hypsignathus</i> H. Allen, 1862
1866	<i>Entomophaga</i>	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
1866	<i>Scotophilii</i>	<i>Scotophilinae</i> Van Cakenberghe and Seamark, 2008

Nesi, Kadjo and Hassanin

2012	<i>Megaloglossus azagnyi</i>	<i>Megaloglossus azagnyi</i> Nesi, Kadjo and Hassanin, 2012

Nesi, Kadjo, Pourrut, Leroy, Pongombo Shongo, Cruaud and Hassanin

2012	<i>Stenonycterini</i>	<i>Stenonycterini</i> Nesi, Kadjo, Pourrut, Leroy, Pongombo Shongo, Cruaud and Hassanin, 2012

Nicoll

1908	<i>Pteropus comorensis</i>	<i>Pteropus seychellensis</i> A. Milne-Edwards, 1877

Noack

1887	<i>Phyllorhina commersonii</i> var. <i>marungensis</i>	<i>Macronycteris vittatus</i> (Peters, 1852)
1887	<i>Scotophilus minimus</i>	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
1889	<i>Chalinolobus conigicus</i>	<i>Glauconycteris argentata</i> (Dobson, 1875)
1889	<i>Vesperugo pagenstecheri</i>	<i>Afronycteris nana</i> (Peters, 1852)
1889	<i>Vesperus damarensis</i>	<i>Laephotis capensis</i> (A. Smith, 1829)
1889	<i>Vesperus pusillus</i>	<i>Afronycteris nana</i> (Peters, 1852)
1893	<i>Phyllorhina rubra</i>	<i>Hipposideros ruber</i> (Noack, 1893)

Ogilby

1835	<i>Pteropus Gambianus</i>	<i>Epomophorus gambianus</i> (Ogilby, 1835)
1835	<i>Pteropus Gambianus</i>	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)

- 1835 *Pteropus macrocephalus* *Epomophorus gambianus gambianus* (Ogilby, 1835)
- Oken
- 1816 *Myopterus senegalensis* *Myopterus daubentonii* Desmarest, 1820
 1816 *Pteropus madagascariensis* *Pteropus rufus* E. Geoffroy St.-Hilaire, 1803
 1816 *Rhinopoma brevicaudatum* *Rhinopoma cystops* Thomas, 1903
 1838 *Pteropus phæops* *Pteropus rufus* E. Geoffroy St.-Hilaire, 1803
- Pagenstecher
- 1885 *Megaloglossus* *Megaloglossus* Pagenstecher, 1885
 1885 *Megaloglossus woermanni* *Megaloglossus woermanni* Pagenstecher, 1885
- Pallas
- 1767 *Vespertiliones* *CHIROPTERA* Blumenbach, 1779
- Palmer
- 1903 *Necronycteris* *Necromantis* Weithofer, 1887
- Peters
- 1852 *Cynonycteris* *Rousettus* Gray, 1821
 1852 *Dysopes brachypterus* *Mops (Xiphonycteris) brachypterus* (Peters, 1852)
 1852 *Dysopes dubius* *Mops (Chaerephon) pumilus* (Cretzschmar, 1826)
 1852 *Dysopes limbatus* *Mops (Chaerephon) pumilus* (Cretzschmar, 1826)
 1852 *Emballonura afra* *Coleura afra* (Peters, 1852)
 1852 *Epomophorus crypturus* *Epomophorus crypturus* Peters, 1852
 1852 *Nycteris fuliginosa* *Nycteris thebaica* E. Geoffroy St.-Hilaire, 1818
 1852 *Nycteris villosa* *Nycteris hispida* (Schreber, 1774)
 1852 *Nycticejus planirostris* *Scotophilus dinganii* (A. Smith, 1833)
 1852 *Nycticejus viridis* *Scotophilus viridis* (Peters, 1852)
 1852 *Phyllorrhina caffa* *Hipposideros caffer* (Sundevall, 1846)
 1852 *Phyllorrhina gracilis* *Hipposideros caffer* (Sundevall, 1846)
 1852 *Phyllorrhina patellifera* *Hipposideros caffer* (Sundevall, 1846)
 1852 *Phyllorrhina vittata* *Macronycteris vittatus* (Peters, 1852)
 1852 *Rhinolophus lobatus* *Rhinolophus lobatus* Peters, 1852
 1852 *Vespertilio nanus* *Afronycteris nana* (Peters, 1852)
 1859 *Nycticejus Schlieffenii* *Nycticeinops schlieffenii* (Peters, 1859)
 1859 *Otonycteris* *Otonycteris* Peters, 1859
 1859 *Otonycteris Hemprichii* *Otonycteris hemprichii* Peters, 1859
 1859 *Rhinopoma Lepsianum* *Rhinopoma microphyllum* (Brünnich, 1782)
 1862 *Pterocyon* *Eidolon Rafinesque*, 1815
 1862 *Pterocyon paleaceus* *Eidolon helvum helvum* (Kerr, 1792)
 1865 *Nyctinomus (Mormopterus) jugularis* *Mormopterus jugularis* (Peters, 1865)
 1865 *Brachyura* *EMBALLONURIDAE* Gervais, 1855
 1865 *Molossi* *MOLOSSIDAE* Gervais, 1856
 1865 *Mormopterus* *Mormopterus* Peters, 1865
 1865 *Nycteris grandis* *Nycteris grandis* Peters, 1865
 1865 *Nyctophilinae* *Vespertilioninae* Gray, 1821
 1865 *Rhinolophi* *RHINOLOPHIDAE* Gray, 1825
 1866 *Megadermata* *MEGADERMATIDAE* H. Allen, 1864
 1866 *Pteropi* *PTEROPODIDAE* Gray, 1821
 1866 *Vespertiliones* *VESPERTILIONIDAE* Gray, 1821
 1867 *Coeloephyllus* *Rhinolophus Lacépède*, 1799
 1867 *Coelophyllus* *Rhinolophus Lacépède*, 1799
 1867 *Chalinolobus* *Glauconycteris* Dobson, 1875
 1867 *Colëura* *Coleura* Peters, 1867
 1867 *Miniopterus minor* *Miniopterus minor* Peters, 1867
 1867 *Rhinolophus blasii* *Rhinolophus blasii* Peters, 1867
 1867 *Vespertilio (Pternopterus)* *Myotis* Kaup, 1829
 1868 *Alobus* *Pipistrellus* Kaup, 1829
 1868 *Epomophorus pusillus* *Epomophorus pusillus* Peters, 1868
 1868 *Pternopterus* *Myotis* Kaup, 1829
 1868 *Rhinolophus Deckenii* *Rhinolophus deckenii* Peters, 1868
 1868 *Vesperus (Hesperoptenus) kraussii* *Glauconycteris poensis* (Gray, 1842)
 1868 *Coleura seychellensis* *Coleura seychellensis* Peters, 1868
 1869 *Rhinolophus æthiops* *Rhinolophus fumigatus* Rüppell, 1842
 1869 *Vesperus Bottae* *Eptesicus (Cnephæus) bottae* (Peters, 1869)
 1870 *Nyctinomus angolensis* *Mops (Mops) condylurus* (A. Smith, 1833)
 1870 *Vespertilio Bocagii* *Myotis bocagii bocagii* (Peters, 1870)
 1870 *Vespertilio Bocagii* *Myotis bocagii* (Peters, 1870)

1870	<i>Vesperugo pusillulus</i>	<i>Afronycteris nana</i> (Peters, 1852)
1871	<i>Cyclorhina</i>	<i>Hipposideros</i> Gray, 1831
1871	<i>Doryrhina</i>	<i>Doryrhina</i> Peters, 1871
1871	<i>Nycteris angolensis</i>	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1871	<i>Nycteris damarensis</i>	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1871	<i>Ptychorhina</i>	<i>Hipposideros</i> Gray, 1831
1871	<i>Sideroderma</i>	<i>Hipposideros</i> Gray, 1831
1871	<i>Sybdesmotus</i>	<i>Hipposideros</i> Gray, 1831
1871	<i>Syndesmotis</i>	<i>Hipposideros</i> Gray, 1831
1871	<i>Thyreorhina</i>	<i>Hipposideros</i> Gray, 1831
1872	<i>Lyroderma</i>	<i>Megaderma</i> E. Geoffroy St.-Hilaire, 1810
1872	<i>Megaderma cor</i>	<i>Cardioderma cor</i> (Peters, 1872)
1872	<i>Vesperus tenuipinnis</i>	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
1873	<i>Cardioderma</i>	<i>Cardioderma</i> Peters, 1873
1874	<i>Emballonura atrata</i>	<i>Paremballonura atrata</i> (Peters, 1874)
1876	<i>Epomophorus macrocephalus</i>	<i>Hypsignathus monstrosus</i> H. Allen, 1862
1877	<i>Triænops afer</i>	<i>Triænops afer</i> Peters, 1877
1878	<i>Mormopterus setiger</i>	<i>Platymops setiger setiger</i> (Peters, 1878)
1878	<i>Mormopterus setiger</i>	<i>Platymops setiger</i> (Peters, 1878)
1878	<i>Rhinolophus Hildebrandtii</i>	<i>Rhinolophus hildebrandtii</i> Peters, 1878

Peterson

1982	<i>Glauconycteris kenyacola</i>	<i>Glauconycteris kenyacola</i> Peterson, 1982

Peterson and Smith

1973	<i>Glauconycteris gleni</i>	<i>Glauconycteris gleni</i> Peterson and Smith, 1973

Peterson, Eger and Mitchell

1995	<i>Eptesicus somalicus malagasyensis</i>	<i>Laephotis malagasyensis</i> (Peterson, Eger and Mitchell, 1995)
1995	<i>Miniopterus gleni</i>	<i>Miniopterus gleni</i> Peterson, Eger and Mitchell, 1995

Pohle

1943	<i>Scotonycteris ophiodon</i>	<i>Casinycteris ophiodon</i> (Pohle, 1943)

Pousargues

1898	<i>Rhinolophus Maclaudi</i>	<i>Rhinolophus maclaudi</i> Pousargues, 1898

Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste

2014	<i>Miniopterus maghrebensis</i>	<i>Miniopterus maghrebensis</i> Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014

Qumsiyeh

1985	<i>rueppellii</i>	<i>Tadarida teniotis</i> (Rafinesque, 1814)

Rachl

1983	<i>Tadarida engesseri</i>	<i>Tadarida engesseri</i> Rachl, 1983

Rafinesque

1814	<i>Cephalotes teniotis</i>	<i>Tadarida teniotis</i> (Rafinesque, 1814)
1814	<i>Tadarida</i>	<i>Tadarida</i> Rafinesque, 1814
1815	<i>Chiroptera</i>	<i>CHIROPTERA</i> Blumenbach, 1779
1815	<i>Eidolon</i>	<i>Eidolon</i> Rafinesque, 1815
1815	<i>Vespertilia</i>	<i>VESPERTILIONIDAE</i> Gray, 1821
1820	<i>Eptesicus</i>	<i>Eptesicus</i> Rafinesque, 1820

Ralph, Richards, Taylor, Napier and Lamb

2015	<i>Otomops harrisoni</i>	<i>Otomops harrisoni</i> Ralph, Richards, Taylor, Napier and Lamb, 2015

Ravel

2016	<i>?Necromantis fragmentum</i>	<i>Necromantis fragmentum</i> (Ravel, 2016)
2016	<i>?Palaeophyllophora tunisiensis</i>	<i>Palaeophyllophora tunisiensis</i> (Ravel, 2016)
2016	<i>?Vespertiliavus aenigma</i>	<i>Vespertiliavus aenigma</i> (Ravel, 2016)
2016	<i>Chambinycteris</i>	<i>Chambinycteris</i> Ravel, 2016
2016	<i>Chambinycteris pusilli</i>	<i>Chambinycteris pusilli</i> Ravel, 2016
2016	<i>Drakonycteris</i>	<i>Drakonycteris</i> Ravel, 2016
2016	<i>Drakonycteris glibzegdouensis</i>	<i>Drakonycteris glibzegdouensis</i> Ravel, 2016
2016	<i>Hipposideros (Pseudorhinolophus) africanum</i>	<i>Hipposideros africanum</i> Ravel, 2016
2016	<i>Khoufechia</i>	<i>Khoufechia</i> Ravel, 2016
2016	<i>Khoufechia gunnelli</i>	<i>Khoufechia gunnelli</i> Ravel, 2016
2016	<i>Pseudovespertiliavus</i>	<i>Pseudovespertiliavus</i> Ravel, 2016
2016	<i>Pseudovespertiliavus parva</i>	<i>Pseudovespertiliavus parva</i> Ravel, 2016
2016	<i>Vespertiliavus kasserinensis</i>	<i>Vespertiliavus kasserinensis</i> (Ravel, 2016)

Ravel, Marivaux, Tabuce, Bel Haj Ali, Essid and Vianey-Liaud

2012	<i>Witwatia sigei</i>	<i>Witwatia sigei</i> Ravel, Marivaux, Tabuce, Bel Haj Ali, Essid and Vianey-Liaud, 2012

Reeder, Helgen, Vodzak, Lunde and Ejotre

2013	<i>Niumbaha</i>	<i>Glauconycteris</i> Dobson, 1875

Revilliod

1917	<i>Palaeophyllophora</i>	<i>Palaeophyllophora</i> Revilliod, 1917

Robbins

1983	<i>Scotophilus nucella</i>	<i>Scotophilus nucella</i> Robbins, 1983

Robbins and Sarich

1988	<i>Taphozoini</i>	<i>Taphozoinae</i> Jerdon, 1867

Roberts

1913	<i>Pipistrellus nanus australis</i>	<i>Afronycteris nana</i> (Peters, 1852)
1917	<i>Cloeotis percivali australis</i>	<i>Cloeotis percivali</i> Thomas, 1901
1917	<i>Platymops</i>	<i>Sauromys</i> Roberts, 1917
1917	<i>Platymops (Sauromys)</i>	<i>Sauromys</i> Roberts, 1917
1917	<i>Platymops haagneri</i>	<i>Sauromys petrophilus haagneri</i> (Roberts, 1917)
1917	<i>Platymops petrophilus</i>	<i>Sauromys petrophilus</i> (Roberts, 1917)
1919	<i>Cistugo lesueuri</i>	<i>Cistugo lesueuri</i> Roberts, 1919
1919	<i>Eptesicus melckorum</i>	<i>Laephotis capensis</i> (A. Smith, 1829)
1924	<i>Eptesicus zuluensis</i>	<i>Neoromicia zuluensis</i> (Roberts, 1924)
1924	<i>Kerivoula nidicola zuluensis</i>	<i>Kerivoula argentata</i> Tomes, 1861
1924	<i>Rhinolophus darlingi barbertonensis</i>	<i>Rhinolophus darlingi</i> K. Andersen, 1905
1926	<i>Chaerephon pumilus elphicki</i>	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1926	<i>Eptesicops</i>	<i>Pipistrellus</i> Kaup, 1829
1926	<i>Neoromicia</i>	<i>Neoromicia</i> Roberts, 1926
1932	<i>Choerephon (Lophomops) langi</i>	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1932	<i>Eptesicus capensis nkatiensis</i>	<i>Laephotis capensis</i> (A. Smith, 1829)
1932	<i>Neoromicia vansonii</i>	<i>Neoromicia zuluensis</i> (Roberts, 1924)
1932	<i>Pipistrellus vernayi</i>	<i>Vansonia rueppellii vernayi</i> (Roberts, 1932)
1932	<i>Scoteinus schlieffenii fitzsimonsi</i>	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
1946	<i>Eptesicus hottentotus bensoni</i>	<i>Eptesicus (Cnephæus) hottentotus</i> (A. Smith, 1833)
1946	<i>Nycteris woodi sabiensis</i>	<i>Nycteris woodi</i> K. Andersen, 1914
1946	<i>Nyctinomus masoni</i>	<i>Tadarida fulminans</i> (Thomas, 1903)
1946	<i>Nyctinomus rhodesiae</i>	<i>Mops (Chaerephon) ansorgei</i> (Thomas, 1913)
1946	<i>Platymops petrophilus erongensis</i>	<i>Sauromys petrophilus erongensis</i> (Roberts, 1946)
1946	<i>Platymops petrophilus fitzsimonsi</i>	<i>Sauromys petrophilus umbratus</i> (Shortridge & Carter, 1938)
1946	<i>Rhinolophus darlingi damarensis</i>	<i>Rhinolophus damarensis</i> Roberts, 1946
1946	<i>Rhinolophus swinnyi rhodesiae</i>	<i>Rhinolophus rhodesiae</i> Roberts, 1946
1946	<i>Scotophilus nigrita pondoensis</i>	<i>Scotophilus dinganii</i> (A. Smith, 1833)
1946	<i>Vansonia</i>	<i>Vansonia</i> Roberts, 1946
1946	<i>Vansonia</i>	<i>Pipistrellus</i> Kaup, 1829
1948	<i>Pipistrellus (Romicia) kuhlii broomi</i>	<i>Pipistrellus (Pipistrellus) hesperidus broomi</i> Roberts, 1948

Robin

1881 *Nycteris Revoilii* *Nycteris thebaica* E. Geoffroy St.-Hilaire, 1818

Rosina and Pickford

2021 *Afrillonura* *Afrillonura* Rosina and Pickford, 2021
 2021 *Afrillonura namibensis* *Afrillonura namibensis* Rosina and Pickford, 2021

Rüppell

1842 *Nicticejus* *Scotophilus* Leach, 1821
 1842 *Pteropus Schoënsis* *Epomophorus labiatus* (Temminck, 1837)
 1842 *Rhinolophus fumigatus* *Rhinolophus fumigatus* Rüppell, 1842
 1842 *Vespertilio Pipistrellus varietas africana* *Afronycteris nana* (Peters, 1852)

Samonds

2007 *Hipposideros besaoka* *Macronycteris besaoka* (Samonds, 2007)
 2007 *Triaenops goodmani* *Triaenops goodmani* Samonds, 2007

Sanborn

1936 *Miniopterus africanus* *Miniopterus africanus* Sanborn, 1936
 1936 *Miniopterus rufus* *Miniopterus inflatus rufus* Sanborn, 1936
 1936 *Mops angolensis wonderi* *Mops (Mops) condylurus* (A. Smith, 1833)
 1936 *Rhinolophus alcyone alticolus* *Rhinolophus simulator* K. Andersen, 1904
 1939 *Rhinolophus aethiops diversus* *Rhinolophus fumigatus* Rüppell, 1842
 1950 *Micropteropus grandis* *Epomophorus grandis* (Sanborn, 1950)

Sanderson

1937 *Hipposideros sandersoni* *Hipposideros curtus* G.M. Allen, 1921

Satunin

1909 *Otonycteris cinereus* *Otonycteris hemprichii* Peters, 1859

Savi

1825 *Dinops* *Tadarida Rafinesque*, 1814
 1825 *Dinops Cestonii* *Tadarida teniotis* (Rafinesque, 1814)

Scharff

1900 *Mormopterus Whitleyi* *Myopterus whitleyi* (Scharff, 1900)

Schinz

1840 *Dysopes Savii* *Tadarida teniotis* (Rafinesque, 1814)

Schlegel

1867 *Pteropus dupréanus* *Eidolon dupreanum* (Schegel, 1867)

Schlitter and Aggundey

1986 *Eptesicus hottentotus portavernus* *Eptesicus (Cnephaeus) hottentotus* (A. Smith, 1833)

Schlosser

1887 *Vespertiliavus* *Vespertiliavus* Schlosser, 1887
 1910 *Vampyravus* *Vampyravus* Schlosser, 1910
 1910 *Vampyravus orientalis* *Vampyravus orientalis* Schlosser, 1910
 1911 *Provampyrus* *Vampyravus* Schlosser, 1910
 1911 *Provampyrus orientalis* *Vampyravus orientalis* Schlosser, 1910

Schreber

1774	<i>Vespertilio barbastellus</i>	<i>Barbastella barbastellus</i> (Schreber, 1774)
1774	<i>Vespertilio ferrum-equinum</i>	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
1774	<i>Vespertilio hispidus</i>	<i>Nycteris hispida</i> (Schreber, 1774)
1774	<i>Vespertilio Nigrita</i>	<i>Scotophilus nigrita</i> (Schreber, 1774)
1774	<i>Vespertilio pipistrellus</i>	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
1780	<i>Vespertilio lasiopterus</i>	<i>Nyctalus lasiopterus</i> (Schreber, 1780)

Schulze

1897	<i>midas</i>	<i>Tadarida teniotis</i> (Rafinesque, 1814)

Seabra

1898	<i>Phyllorhina angolensis</i>	<i>Hipposideros caffer</i> (Sundevall, 1846)
1898	<i>Rhinolophus angolensis</i>	<i>Rhinolophus lobatus</i> Peters, 1852
1900	<i>Epomophorus anchietæ</i>	<i>Plerotes anchietæ</i> (Seabra, 1900)
1900	<i>Nyctinomus Anchietæ</i>	<i>Tadarida aegyptiaca bocagei</i> (Seabra, 1900)
1900	<i>Nyctinomus Bocagei</i>	<i>Tadarida aegyptiaca bocagei</i> (Seabra, 1900)
1900	<i>Nyctinomus brunneus</i>	<i>Tadarida aegyptiaca bocagei</i> (Seabra, 1900)
1900	<i>Vesperugo Anchietæ</i>	<i>Neoromicia anchietæ</i> (Seabra, 1900)
1900	<i>Vesperugo (Vesperus) flavescens</i>	<i>Nycticeinops grandidieri</i> (Dobson, 1876)

Senna

1905	<i>Rhinolophus andreinii</i>	<i>Rhinolophus blasii</i> Peters, 1867
1914	<i>Rhinolophus bembanicus</i>	<i>Rhinolophus simulator</i> K. Andersen, 1904

Serra-Cobo, López-Roig, Bayer, Amengual and Guasch

2008	<i>Pteropoidea</i>	<i>PTEROPODOIDEA</i> Gray, 1821

Setzer

1971	<i>Laephotis botswanae</i>	<i>Laephotis botswanae</i> Setzer, 1971
1971	<i>Laephotis namibensis</i>	<i>Laephotis namibensis</i> Setzer, 1971

Shortridge

1942	<i>Eptesicus megalurus pallidior</i>	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)

Shortridge and Carter

1938	<i>Platymops haagneri umbratus</i>	<i>Sauromys petrophilus umbratus</i> (Shortridge & Carter, 1938)
1938	<i>Scotophilus angusticeps</i>	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)

Sigé

1968	<i>Brachipposideros</i>	<i>Brachipposideros</i> Sigé, 1968
1976	<i>Megaderma jaegeri</i>	<i>Megaderma jaegeri</i> Sigé, 1976
1985	<i>Philisidae</i>	<i>PHILISIDAE</i> Sigé, 1985
1985	<i>Philisis</i>	<i>Philisis</i> Sigé, 1985
1985	<i>Philisis sphingis</i>	<i>Philisis sphingis</i> Sigé, 1985
1991	<i>Dizzya</i>	<i>Dizzya</i> Sigé, 1991
1991	<i>Dizzya exsultans</i>	<i>Dizzya exsultans</i> Sigé, 1991
2011	<i>Necromantidae</i>	<i>NECROMANTIDAE</i> Sigé, 2011

Sigé, Thomas, Sen, Gheerbrant, Roger and Al-Sulaimani

1994	<i>Dhofarella</i>	<i>Dhofarella</i> Sigé, Thomas, Sen, Gheerbrant, Roger and Al-Sulaimani, 1985
1994	<i>Dhofarella thaleri</i>	<i>Dhofarella thaleri</i> Sigé et al., 1994

Simmons and Cirranello

2020	<i>Mops bemmeleni</i>	<i>Mops (Chaerephon) bemmeleni</i> (Jentink, 1879)
2020	<i>Mops russatus</i>	<i>Mops (Chaerephon) russatus</i> (J.A. Allen, 1917)

Simmons and Geisler

1998	<i>Megachiroptera</i>	<i>RHINOLOPHIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
1998	<i>Microchiroptera</i>	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
1998	<i>Microchiroptera</i>	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007

Simmons, Seiffert and Gunnell

2016	<i>Aegyptonycteridae</i>	<i>AEGYPTONYCTERIDAE</i> Simmons, Seiffert and Gunnell, 2016
2016	<i>Aegyptonycteris</i>	<i>Aegyptonycteris</i> Simmons, Seiffert and Gunnell, 2016
2016	<i>Aegyptonycteris knightae</i>	<i>Aegyptonycteris knightae</i> Simmons, Seiffert and Gunnell, 2016

Simpson

1967	<i>Propotto</i>	<i>Propotto</i> Simpson, 1967
1967	<i>Propotto leakeyi</i>	<i>Propotto leakeyi</i> Simpson, 1967

A. Smith

1829	<i>Nycteris affinis</i>	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1829	<i>Nycteris Capensis</i>	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1829	<i>Pteropus Leachii</i>	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1829	<i>Rhinolophus Geoffroyii</i>	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1829	<i>Vespertilio Capensis</i>	<i>Laephotis capensis</i> (A. Smith, 1829)
1833	<i>Nyctinomus Condylurus</i>	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
1833	<i>Nyctinomus dubius</i>	<i>Mormopterus acetabulosus natalensis</i> (A. Smith, 1847)
1833	<i>Vespertilio Danganii</i>	<i>Scotophilus danganii</i> (A. Smith, 1833)
1833	<i>Vespertilio Hottentota</i>	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
1833	<i>Vespertilio Natalensis</i>	<i>Miniopterus natalensis</i> (A. Smith, 1833)
1834	<i>Vespertilio goudoti</i>	<i>Myotis goudoti</i> (A. Smith, 1834)
1847	<i>Dysopes natalensis</i>	<i>Mormopterus acetabulosus natalensis</i> (A. Smith, 1847)
1847	<i>Vespertilio lanosus</i>	<i>Kerivoula lanosa</i> (A. Smith, 1847)
1849	<i>Vespertilio minutus</i>	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)

Springer, Teeling, Madsen, Stanhope and De Jong

2001	<i>YINPTEROCHIROPTERA</i>	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
2001	<i>YINPTEROCHIROPTERA</i>	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007

Stanley

2008	<i>Mops bakarii</i>	<i>Mops (Xiphonycteris) bakarii</i> Stanley, 2009

Stresemann

1954	<i>Vespertilio brevicauda</i>	<i>Rhinopoma cystops</i> Thomas, 1903
1954	<i>Vespertilio ferox</i>	<i>Rhinopoma cystops</i> Thomas, 1903

Sundevall

1843	<i>Dysopes midas</i>	<i>Mops (Mops) midas</i> (Sundevall, 1843)
1843	<i>Dysopes midas</i>	<i>Mops (Mops) midas midas</i> (Sundevall, 1843)
1846	<i>Pteropus Wahlbergi</i>	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
1846	<i>Rhinolophus caffer</i>	<i>Hipposideros caffer</i> (Sundevall, 1846)
1846	<i>Vespertilio scotinus</i>	<i>Miniopterus natalensis</i> (A. Smith, 1833)
1846	<i>Vespertilio subtilis</i>	<i>Pipistrellus (Pipistrellus) hesperidus subtilis</i> (Sundevall, 1846)
1860	<i>Rhinolophus auritus</i>	<i>Rhinolophus capensis</i> Lichtenstein, 1823

Swainson

1835	<i>Pteropus megacephalus</i>	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)

Tate

1941	<i>Coelopinae</i>	<i>HIPPOSIDERIDAE</i> Lydekker, 1891
1941	<i>[Triaenops] furinea</i>	<i>Paratriaenops pauliani</i> (Goodman and Ranivo, 2008)
1942	<i>Myotinae</i>	<i>Myotinae</i> Tate, 1942
1942	<i>Myotini</i>	<i>VESPERTILIONIDAE</i> Gray, 1821
1942	<i>Myotini</i>	<i>Myotinae</i> Tate, 1942
1942	<i>Nycticeini</i>	<i>Vespertilioninae</i> Gray, 1821
1842	<i>Pipistrellini</i>	<i>Pipistrellini</i> Tate, 1942

Taylor, MacDonald, Goodman, Kearney, Cotterill, Stoffberg, Monadjem, Schoeman, Guyton, Naskrecki and Richards

2018	<i>Rhinolophus gorongosae</i>	<i>Rhinolophus gorongosae</i> Taylor, MacDonald, Goodman, Kearney, Cotterill, Stoffberg, Monadjem, Schoeman, Guyton, Naskrecki and Richards, 2018

Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill

2012	<i>Rhinolophus cohenae</i>	<i>Rhinolophus cohenae</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
2012	<i>Rhinolophus mabuensis</i>	<i>Rhinolophus mabuensis</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
2012	<i>Rhinolophus mossambicus</i>	<i>Rhinolophus mossambicus</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
2012	<i>Rhinolophus smithersi</i>	<i>Rhinolophus smithersi</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012

Teeling, Springer, Madsen, Bates, O'Brien and Murphy

2005	<i>Emballonuroidea</i>	<i>NYCTEROIDEA</i> Van der Hoeven, 1855

Temminck

1825	<i>Pteropus geoffroyi</i>	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1825	<i>Pteropus phaiops</i>	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
1827	<i>Dysopes geoffroyi</i>	<i>Tadarida aegyptiaca aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
1827	<i>Dysopes rüpelii</i>	<i>Tadarida teniotis</i> (Rafinesque, 1814)
1832	<i>Pteropus hottentottus</i>	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1832	<i>V[espertilio] tricolor</i>	<i>Myotis tricolor</i> (Temminck, 1832)
1832	<i>Vespertilio epichrysus</i>	<i>Scotophilus dinganii</i> (A. Smith, 1833)
1832	<i>Vespertilio platycephalus</i>	<i>Pipistrellus (Pipistrellus) hesperidus</i> (Temminck, 1840)
1835	<i>Taphozous leucopterus</i>	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
1837	<i>Pteropus labiatus</i>	<i>Epomophorus labiatus</i> (Temminck, 1837)
1837	<i>Pteropus vulgaris</i>	<i>Pteropus subniger</i> (Kerr, 1792)
1838	<i>Saccolaimus</i>	<i>Saccolaimus</i> Temminck, 1838
1840	<i>Vespertilio d'asythrix</i>	<i>Miniopterus natalensis</i> (A. Smith, 1833)
1840	<i>Vespertilio hesperida</i>	<i>Pipistrellus (Pipistrellus) hesperidus hesperidus</i> (Temminck, 1840)
1840	<i>Vespertilio hesperida</i>	<i>Pipistrellus (Pipistrellus) hesperidus</i> (Temminck, 1840)
1840	<i>Vespertilio minuta</i>	<i>Laephotis capensis</i> (A. Smith, 1829)
1835-1841	<i>Vespertilio isabellinus</i>	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
1835-1841	<i>Vespertilio megalurus</i>	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
1853	<i>Pachysoma</i>	<i>Epomophorus</i> Bennett, 1836
1853	<i>Phyllorrhina cyclops</i>	<i>Doryrhina cyclops</i> (Temminck, 1853)
1853	<i>Phyllorrhina fuliginosa</i>	<i>Hipposideros fuliginosus</i> (Temminck, 1853)
1853	<i>Rhinolophus alcyone</i>	<i>Rhinolophus alcyone</i> Temminck, 1853
1853	<i>Taphozous peli</i>	<i>Saccolaimus peli</i> (Temminck, 1853)

Thomas

1880	<i>Kerivoula Smithii</i>	<i>Kerivoula smithii</i> Thomas, 1880
1880	<i>Vesperugo (Vesperus) brunneus</i>	<i>Pseudoromicia brunnea</i> (Thomas, 1880)
1889	<i>Vesperugo (Vesperus) Rendalli</i>	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
1890	<i>Scotophilus albofuscus</i>	<i>Scotoecus albofuscus</i> (Thomas, 1890)
1891	<i>Nyctinomus lobatus</i>	<i>Tadarida lobata</i> (Thomas, 1891)
1891	<i>Vesperugo (Vesperus) Moloneyi</i>	<i>Mimetillus moloneyi</i> (Thomas, 1891)
1891	<i>N(yctinomus) taeniotis</i>	<i>Tadarida ventralis</i> (Heuglin, 1861)
1901	<i>Scotæcus</i>	<i>Scotoecus</i> Thomas, 1901
1901	<i>Scotæcus Hindei</i>	<i>Scotoecus hindei</i> Thomas, 1901
1901	<i>Kerivoula harrisoni</i>	<i>Kerivoula lanosa</i> (A. Smith, 1847)

1901	<i>Laephotis</i>	<i>Laephotis</i> Thomas, 1901
1901	<i>Laephotis Wintoni</i>	<i>Laephotis wintoni</i> Thomas, 1901
1901	<i>Cloeotis</i>	<i>Cloeotis</i> Thomas, 1901
1901	<i>Cloeotis Percivali</i>	<i>Cloeotis percivali</i> Thomas, 1901
1901	<i>Nycteris aethiopica luteola</i>	<i>Nycteris macrotis</i> Dobson, 1876
1901	<i>Pipistrellus KuhlII fuscatus</i>	<i>Pipistrellus (Pipistrellus) hesperidus hesperidus</i> (Temminck, 1840)
1901	<i>Pterygistes azureum</i>	<i>Nyctalus azureum</i> (Thomas, 1901)
1901	<i>Vespertilio minutus somalicus</i>	<i>Neoromicia somalica</i> (Thomas, 1901)
1901	<i>Vespertilio platyops</i>	<i>Eptesicus (Cnephæus) platyops</i> (Thomas, 1901)
1901	<i>Glauconycteris beatrix</i>	<i>Glauconycteris beatrix</i> Thomas, 1901
1902	<i>Pipistrellus deserti</i>	<i>Pipistrellus (Pipistrellus) kuhlII</i> (Kuhl, 1817)
1903	<i>Rhinopoma cystops</i>	<i>Rhinopoma cystops</i> Thomas, 1903
1903	<i>Nyctinomus cisturus</i>	<i>Mops (Chaerephon) bemmeleni</i> (Jentink, 1879)
1903	<i>Nyctinomus demonstrator</i>	<i>Mops (Mops) demonstrator</i> (Thomas, 1903)
1903	<i>Nyctinomus fulminans</i>	<i>Tadarida fulminans</i> (Thomas, 1903)
1903	<i>Miniopterus inflatus</i>	<i>Miniopterus inflatus inflatus</i> Thomas, 1903
1903	<i>Miniopterus inflatus</i>	<i>Miniopterus inflatus</i> Thomas, 1903
1903	<i>Nycteris arge</i>	<i>Nycteris arge</i> Thomas, 1903
1903	<i>Nyctinomus thersites</i>	<i>Mops (Xiphonycteris) thersites</i> (Thomas, 1903)
1904	<i>Myotis Hildegardæ</i>	<i>Myotis bocagii bocagii</i> (Peters, 1870)
1904	<i>Nyctinomus hindei</i>	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1904	<i>Pipistrellus crassulus</i>	<i>Nycticeinops crassulus</i> (Thomas, 1904)
1904	<i>Scotophilus nigrila colias</i>	<i>Scotophilus dinganii</i> (A. Smith, 1833)
1904	<i>Scotophilus nigrila nux</i>	<i>Scotophilus nux</i> Thomas, 1904
1904	<i>Scotonycteris bedfordi</i>	<i>Scotonycteris zenkeri</i> Matschie, 1894
1904	<i>Hipposideros Commersoni mostellum</i>	<i>Macronycteris vittatus</i> (Peters, 1852)
1904	<i>Rhinolophus Denti</i>	<i>Rhinolophus denti</i> Thomas, 1904
1904	<i>Myotis Bocagei cupreolus</i>	<i>Myotis bocagii cupreolus</i> Thomas, 1904
1904	<i>Pipistrellus ariel</i>	<i>Hypsugo ariel</i> (Thomas, 1904)
1904	<i>Rhinolophus Andersoni</i>	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1904	<i>Rhinolophus Dobsoni</i>	<i>Rhinolophus landeri</i> Martin, 1838
1904	<i>Pipistrellus nanulus</i>	<i>Pipistrellus (Pipistrellus) nanulus</i> Thomas, 1904
1904	<i>Myzopodidae</i>	<i>MYZOPODIDAE</i> Thomas, 1904
1904	<i>Mimetillus</i>	<i>Mimetillus</i> Thomas, 1904
1905	<i>Glauconycteris papilio</i>	<i>Glauconycteris variegata</i> (Tomes, 1861)
1905	<i>Eomops</i>	<i>Myopterus</i> E. Geoffroy St.-Hilaire, 1818
1906	<i>Miniopterus Majori</i>	<i>Miniopterus majori</i> Thomas, 1906
1906	<i>Miniopterus manavi</i>	<i>Miniopterus manavi</i> Thomas, 1906
1906	<i>Scotophilus damarensis</i>	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
1906	<i>Scotophilus nigrila herero</i>	<i>Scotophilus dinganii</i> (A. Smith, 1833)
1906	<i>Platymops</i>	<i>Platymops</i> Thomas, 1906
1906	<i>Platymops Macmillani</i>	<i>Platymops setiger macmillani</i> Thomas, 1906
1906	<i>Rousettus lanosus</i>	<i>Stenonycteris lanosus</i> (Thomas, 1906)
1906	<i>Kerivoula muscilla</i>	<i>Kerivoula lanosa</i> (A. Smith, 1847)
1908	<i>Nyctinomus leonis</i>	<i>Mops (Xiphonycteris) brachypterus</i> (Peters, 1852)
1908	<i>Rousettus smithii</i>	<i>Myonycteris angolensis smithii</i> (Thomas, 1908)
1909	<i>Taphozous hildegardeæ</i>	<i>Taphozous hildegardeæ</i> Thomas, 1909
1909	<i>Rousettus kempfi</i>	<i>Stenonycteris lanosus</i> (Thomas, 1906)
1909	<i>Scotoecus albigula</i>	<i>Scotoecus albigula</i> Thomas, 1909
1910	<i>Rhinolophus brockmani</i>	<i>Rhinolophus blasii</i> Peters, 1867
1910	<i>Casinycteris</i>	<i>Casinycteris</i> Thomas, 1910
1910	<i>Casinycteris argynnis</i>	<i>Casinycteris argynnis</i> Thomas, 1910
1911	<i>Pipistrellus culex</i>	<i>Afronycteris nana</i> (Peters, 1852)
1912	<i>Kerivoula cuprosa</i>	<i>Kerivoula cuprosa</i> Thomas, 1912
1912	<i>Cistugo</i>	<i>Cistugo</i> Thomas, 1912
1912	<i>Cistugo seabrae</i>	<i>Cistugo seabrae</i> Thomas, 1912
1912	<i>Kerivoula phalaena</i>	<i>Kerivoula phalaena</i> Thomas, 1912
1913	<i>Glauconycteris egeria</i>	<i>Glauconycteris egeria</i> Thomas, 1913
1913	<i>Chærephon nigeriæ</i>	<i>Mops (Chaerephon) nigeriæ nigeriæ</i> (Thomas, 1913)
1913	<i>Chærephon nigeriæ</i>	<i>Mops (Chaerephon) nigeriæ</i> (Thomas, 1913)
1913	<i>Nyctinomus ansorgei</i>	<i>Mops (Chaerephon) ansorgei</i> (Thomas, 1913)
1913	<i>Pipistrellus fuscipes</i>	<i>Vansonia rueppellii fuscipes</i> (Thomas, 1913)
1913	<i>Pipistrellus musciculus</i>	<i>Hypsugo musciculus</i> (Thomas, 1913)
1913	<i>Rhinolophus foxi</i>	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1913	<i>Otomops</i>	<i>Otomops</i> Thomas, 1913
1915	<i>C[oleura] g[allarum] nilosa</i>	<i>Coleura afra</i> (Peters, 1852)
1915	<i>C[oleura] gallarum</i>	<i>Coleura gallarum</i> Thomas, 1915
1915	<i>C[oleura] silhouettæ</i>	<i>Coleura seychellensis</i> Peters, 1868
1915	<i>Glauconycteris phalaena</i>	<i>Glauconycteris variegata</i> (Tomes, 1861)
1915	<i>Rhinopterus lowei</i>	<i>Eptesicus (Rhinopterus) floweri</i> (de Winton, 1901)
1915	<i>Taphozous Sudani</i>	<i>Taphozous perforatus sudani</i> Thomas, 1915
1915	<i>Taphozous perforatus hædinus</i>	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
1915	<i>Scotoecus falabæ</i>	<i>Scotoecus hindei</i> Thomas, 1901
1915	<i>Myopterus albatu</i>	<i>Myopterus daubentonii</i> Desmarest, 1820
1917	<i>Scotoecus woodi</i>	<i>Scotoecus albofuscus</i> (Thomas, 1890)

1920	<i>Taphozous hamiltoni</i>	<i>Taphozous hamiltoni</i> Thomas, 1920
1922	<i>Liponycteris</i>	<i>Taphozous</i> E. Geoffroy St.-Hilaire, 1818
1926	<i>Chaerephon (Lophomops) shortridgei</i>	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
1926	<i>Pipistrellus fouriei</i>	<i>Afronycteris nana</i> (Peters, 1852)
1927	<i>Myotis scotti</i>	<i>Myotis scotti</i> Thomas, 1927
1927	<i>Miniopterus smitianus</i>	<i>Miniopterus natalensis</i> (A. Smith, 1833)

Thomas and Hinton

1923	<i>Pipistrellus marrensis</i>	<i>Pipistrellus (Pipistrellus) rusticus</i> (Tomes, 1861)

Thomas and Schwann

1905	<i>Vespertilio capensis gracilior</i>	<i>Laephotis capensis</i> (A. Smith, 1829)
1905	<i>Vespertilio matroka</i>	<i>Laephotis matroka</i> (Thomas and Schwann, 1905)
1906	<i>Miniopterus fraterculus</i>	<i>Miniopterus fraterculus</i> Thomas and Schwann, 1906

Thomas and Wroughton

1908	<i>Scoteinus schlieffeni albiventer</i>	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
1908	<i>Scoteinus schlieffeni australis</i>	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
1908	<i>Scoteinus schlieffeni bedouin</i>	<i>Nycticeinops schlieffenii</i> (Peters, 1859)

Thorn, Kock and Cuisin

2007	<i>Afropipistrellus</i>	<i>Nycticeinops</i> Hill & Harrison, 1987

Tomes

1858	<i>Vespertilio Madagascariensis</i>	<i>Myotis goudoti</i> (A. Smith, 1834)
1859	<i>Scotophilus darwini</i>	<i>Hypsugo savii</i> (Bonaparte, 1837)
1860	<i>Epomophorus franqueti</i>	<i>Epomops franqueti</i> (Tomes, 1860)
1861	<i>Kerivoula argentata</i>	<i>Kerivoula argentata</i> Tomes, 1861
1861	<i>Scotophilus rusticus</i>	<i>Pipistrellus (Pipistrellus) rusticus</i> (Tomes, 1861)
1861	<i>Scotophilus variegatus</i>	<i>Glauconycteris variegata</i> (Tomes, 1861)

Trouessart

1897	<i>[Nyctinomus (Nyctinomus) pumilus] Var. Major</i>	<i>Mops (Chaerephon) major</i> (Trouessart, 1897)
1897	<i>[Vesperugo (Vesperus) serotinus] Var. Gabonensis</i>	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
1897	<i>Pteropinae</i>	<i>Pteropodinae</i> Gray, 1821
1898	<i>Miniopterae</i>	<i>MINIOPTERIDAE</i> Dobson, 1875
1898	<i>Miomegaderma gaillardi</i>	<i>Megaderma gaillardi</i> (Trouessart, 1898)
1904	<i>Epomophorus pousarguesi</i>	<i>Epomophorus gambianus pousarguesi</i> Trouessart, 1904
1907	<i>Triaenops furcula</i>	<i>Paratriaenops furculus</i> (Trouessart, 1907)
1910	<i>Rhinolophus blasiusi</i>	<i>Rhinolophus blasii</i> Peters, 1867

Troughton

1929	<i>Anamygdon</i>	<i>Myotis</i> Kaup, 1829
1941	<i>Austronomus</i>	<i>Tadarida</i> Rafinesque, 1814
1941	<i>Rhinophyllotis</i>	<i>Rhinolophus</i> Lacépède, 1799

True

1893	<i>Pteropus aldabrensis</i>	<i>Pteropus aldabrensis</i> True, 1893

Trujillo, Ibáñez and Juste

2002	<i>Barbastella barbastellus guanchae</i>	<i>Barbastella barbastellus</i> (Schreber, 1774)

Turni and Kock

2008	<i>Vesperus hösemanni</i>	<i>Pseudoromicia rendalli</i> (Thomas, 1889)

Van Cakenberghe and Seamark

2008	<i>Scotophilinae</i>	<i>Scotophilinae</i> Van Cakenberghe and Seamark, 2008

Van Cakenberghe, Kearney and Seamark

2007	<i>NOCTILIONIFORMACEI</i>	<i>NOCTILIONIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
2007	<i>NYCTERIFORMACEI</i>	<i>NYCTERIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
2007	<i>PTEROPODIFORMACEI</i>	<i>PTEROPODIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
2007	<i>PTEROPODIFORMI</i>	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
2007	<i>RHINOLOPHIFORMACEI</i>	<i>RHINOLOPHIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
2007	<i>VESPERTILIONIFORMACEI</i>	<i>VESPERTILIONIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
2007	<i>VESPERTILIONIFORMI</i>	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007

Van der Hoeven

1855	<i>Histiorhina</i>	<i>RHINOLOPHIDAE</i> Gray, 1825
1855	<i>Nycteridae</i>	<i>NYCTERIDAE</i> Van der Hoeven, 1855
1855	<i>Nycterioidea</i>	<i>NYCTEROIDEA</i> Van der Hoeven, 1855

Van Valen

1979	<i>PHYLLOSTOMATIA</i>	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
1979	<i>VESPERTILIONIA</i>	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007

Verschuren

1957	<i>Hipposideros beatus maximus</i>	<i>Hipposideros beatus</i> (K. Andersen, 1906)

Volleth, Bronner, Göpfert, Heller, von Helversen and Yong

2001	<i>Pipistrellus cf kuhlii</i>	<i>Pipistrellus</i> <i>*(Pipistrellus)*</i> sp. aff. <i>*kuhlii*</i> Kuhl, 1817

Wagner

1840	<i>N[ycteris] albiventer</i>	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1840	<i>N[ycteris] discolor</i>	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1843	<i>Gymnorhina</i>	<i>VESPERTILIONIDAE</i> Gray, 1821
1845	<i>Rhinolophus gigas</i>	<i>Macronycteris gigas</i> (Wagner, 1845)
1855	<i>V[espertilio] Smithii</i>	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
1855	<i>Vespertilio smithii</i>	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)

Wallace

1880	<i>Pteropus comorensis</i>	<i>Pteropus seychellensis</i> A. Milne-Edwards, 1877

C.O. Waterhouse

1902	<i>Syndesmotus</i>	<i>Hipposideros</i> Gray, 1831

Weber

1928	<i>Emballonuroidea</i>	<i>EMBALLONURIDAE</i> Gervais, 1855

Weithofer

1888	<i>Necromantis</i>	<i>Necromantis</i> Weithofer, 1887

Wesselman

1984	<i>Coleura muthokai</i>	<i>Coleura muthokai</i> Wesselman, 1984
1984	<i>Hipposideros kaumbului</i>	<i>Hipposideros kaumbului</i> Wesselman, 1984
1984	<i>Tapozous abitus</i>	<i>Saccolaimus abitus</i> (Wesselman, 1984)

Wettstein

1916	<i>Eptesicus rectitragus</i>	<i>Neoromicia guineensis</i> (Bocage, 1889)
1916	<i>Nyctinomus (Nyctinomus) tongaënsis</i>	<i>Tadarida aegyptiaca aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
1916	<i>Scotoecus cinnamomeus</i>	<i>Nycticeinops schlieffenii</i> (Peters, 1859)

Winge

1893	<i>Emballonurini</i>	<i>EMBALLONURIDAE</i> Gervais, 1855
1893	<i>Megadermatini</i>	<i>MEGADERMATIDAE</i> H. Allen, 1864
1893	<i>Molossini</i>	<i>MOLOSSIDAE</i> Gervais, 1856
1893	<i>Pteropi</i>	<i>PTEROPODIDAE</i> Gray, 1821
1893	<i>Rhinolophini</i>	<i>RHINOLOPHIDAE</i> Gray, 1825

Wroughton

1911	<i>Chærephon emini</i>	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)

Zagorodniuk

1997	<i>VESPERTILIFORMES</i>	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
2014	<i>Pteropodimorpha</i>	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
2014	<i>Vespertilioniformes</i>	<i>CHIROPTERA</i> Blumenbach, 1779
2014	<i>Vespertilionimorpha</i>	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007

Zagorodniuk, Godovanets, Pokynchereda and Kyseliuk

1995	<i>VESPERTILIIFORMES</i>	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007

Zagorodnyuk

1998	<i>VESPERTILIONIFORMES</i>	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007

Appendix 3c: Synonyms by Publication Date

In this appendix, the synonyms are sorted according to the date they were published.

If a complete date could be found, this date is given. In most cases, however, only the year of publication is known, in which case (according to the International Code of Zoological Nomenclature) the last day of the year is considered to be the date of publication (i.e. 31 December). If a month is known too, the final date of the month is entered.

In a large number of cases the publication date is taken from the (cover of the) publication, and as such the date might not be 100 % correct, since these dates tend to be earlier than the exact date the publication was published.

For names published in "*Annals and Magazine of Natural History*" and "*Journal of Natural History*" the publication dates are taken from Evenhuis (2003). For names published in the "*Proceedings of the Zoological Society of London*", the publication dates are taken from Duncan (1937). The publication dates from Schreber's "*Die Säugethiere*" are taken from Sherborn (1892). For E. Geoffroy St.-Hilaire's "*Description de l'Egypte*", the dates are taken from Sherborn (1897). For Kerr's "*Animal Kingdom*" see Allen (1895).

Publication date	Original Name	Current Name
31 Dec 1762	<i>Pteropus</i> Brisson	<i>Pteropus</i> Brisson, 1762
31 Dec 1767	<i>Vespertilio</i> Pallas	<i>CHIROPTERA</i> Blumenbach, 1779
31 Dec 1774	<i>Vespertilio barbastellus</i> Schreber	<i>Barbastella barbastellus</i> (Schreber, 1774)
31 Dec 1774	<i>Vespertilio ferrum-equinum</i> Schreber	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
31 Dec 1774	<i>Vespertilio hispidus</i> Schreber	<i>Nycteris hispida</i> (Schreber, 1774)
31 Dec 1774	<i>Vespertilio Nigrita</i> Schreber	<i>Scotophilus nigrita</i> (Schreber, 1774)
31 Dec 1774	<i>Vespertilio pipistrellus</i> Schreber	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
31 Dec 1779	<i>CHIROPTERA</i> Blumenbach	<i>CHIROPTERA</i> Blumenbach, 1779
31 Dec 1780	<i>Vespertilio lasiopterus</i> Schreber	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
31 Dec 1782	<i>Vespertilio Microphyllus</i> Brännich	<i>Rhinopoma microphyllum</i> (Brännich, 1782)
28 Feb 1792	<i>minor</i> Kerr	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
28 Feb 1792	<i>Vespertilio vampirus niger</i> Kerr	<i>Pteropus niger</i> (Kerr, 1792)
28 Feb 1792	<i>Vespertilio vampyrus helvus</i> Kerr	<i>Eidolon helvum</i> (Kerr, 1792)
28 Feb 1792	<i>Vespertilio vampyrus helvus</i> Kerr	<i>Eidolon helvum helvum</i> (Kerr, 1792)
28 Feb 1792	<i>Vespertilio vampyrus subniger</i> Kerr	<i>Pteropus subniger</i> (Kerr, 1792)
31 Dec 1792	<i>Vespertilio ferrum-equinum major</i> Kerr	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
31 Dec 1795	<i>Nycteris</i> G. Cuvier and E. Geoffroy	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795
31 Dec 1797	<i>Vespertilio caninus</i> Blumenbach	<i>Pteropus niger</i> (Kerr, 1792)
31 Dec 1799	<i>Rhinolophus</i> Lacépède	<i>Rhinolophus</i> Lacépède, 1799
31 Dec 1799	<i>Spectrum</i> Lacépède	<i>Pteropus</i> Brisson, 1762
31 Dec 1800	<i>Vespertilio hipposideros</i> Bechstein	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
31 Dec 1800	<i>Vespertilio megalotis</i> Bechstein	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
31 Dec 1802	<i>Spectrum rubidum</i> Daudin	<i>Pteropus subniger</i> (Kerr, 1792)
31 Dec 1803	<i>major</i> E. Geoffroy Saint-Hilaire	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
31 Dec 1803	<i>Pteropus fuscus</i> E. Geoffroy Saint-Hilaire	<i>Pteropus niger</i> (Kerr, 1792)
31 Dec 1803	<i>Pteropus ruber</i> E. Geoffroy Saint-Hilaire	<i>Pteropus subniger</i> (Kerr, 1792)
31 Dec 1803	<i>Pteropus rufus</i> E. Geoffroy Saint-Hilaire	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
31 Dec 1803	<i>Pteropus stramineus</i> E. Geoffroy Saint-Hilaire	<i>Eidolon helvum helvum</i> (Kerr, 1792)
31 Dec 1803	<i>Vespertilio borbonicus</i> E. Geoffroy Saint-Hilaire	<i>Scotophilus borbonicus</i> (E. Geoffroy St.-Hilaire, 1803)
31 Aug 1804	<i>V[espertilio] acetabulosus</i> Hermann	<i>Mormopterus acetabulosus acetabulosus</i> (Hermann, 1804)
31 Aug 1804	<i>V[espertilio] acetabulosus</i> Hermann	<i>Mormopterus acetabulosus</i> (Hermann, 1804)
31 Aug 1804	<i>V[espertilio] mauritanus</i> Hermann	<i>Pteropus niger</i> (Kerr, 1792)
31 Dec 1806	<i>Chiropteren</i> Duméril	<i>CHIROPTERA</i> Blumenbach, 1779
31 Dec 1806	<i>Vespertilio emarginatus</i> E. Geoffroy Saint-Hilaire	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
31 Dec 1809	<i>Vespertilio caninus</i> var. <i>b</i> Goldfuss	<i>Eidolon helvum helvum</i> (Kerr, 1792)
31 Dec 1810	<i>Megaderma frons</i> E. Geoffroy Saint-Hilaire	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
31 Dec 1810	<i>Pteropus Edwardsii</i> E. Geoffroy Saint-Hilaire	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
31 Dec 1810	<i>Pteropus Egyptiacus</i> E. Geoffroy Saint-Hilaire	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
31 Dec 1810	<i>Pteropus rubricollis</i> E. Geoffroy Saint-Hilaire	<i>Pteropus subniger</i> (Kerr, 1792)
31 Dec 1810	<i>Pteropus vulgaris</i> E. Geoffroy Saint-Hilaire	<i>Pteropus niger</i> (Kerr, 1792)
31 Dec 1810	<i>Megaderma</i> E. Geoffroy St.-Hilaire	<i>Megaderma</i> E. Geoffroy St.-Hilaire, 1810

Publication date	Original Name	Current Name
31 Aug 1813	<i>Nycteris Daubentonii</i> E. Geoffroy Saint-Hilaire	<i>Nycteris hispida</i> (Schreber, 1774)
31 Dec 1813	<i>Rhinolophus Commersonii</i> E. Geoffroy Saint-Hilaire	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
31 Dec 1813	<i>Rhinolophus tridens</i> E. Geoffroy Saint-Hilaire	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
3 Jun 1814	<i>Cephalotes teniotis</i> Rafinesque	<i>Tadarida teniotis</i> (Rafinesque, 1814)
3 Jun 1814	<i>Tadarida</i> Rafinesque	<i>Tadarida</i> Rafinesque, 1814
31 Dec 1814	<i>Pteropus torquatus</i> G. Fischer	<i>Pteropus subniger</i> (Kerr, 1792)
31 Dec 1815	<i>Pteropus collaris</i> Illiger	<i>Pteropus subniger</i> (Kerr, 1792)
31 Dec 1815	<i>Pteropus flavus</i> Illiger	<i>Eidolon helvum helvum</i> (Kerr, 1792)
31 Dec 1815	<i>Chiroptera</i> Rafinesque	<i>CHIROPTERA</i> Blumenbach, 1779
31 Dec 1815	<i>Eidolon</i> Rafinesque	<i>Eidolon</i> Rafinesque, 1815
31 Dec 1815	<i>Vespertilia</i> Rafinesque	<i>VESPERTILIONIDAE</i> Gray, 1821
31 Dec 1816	<i>Cheiroptères</i> G. Cuvier	<i>CHIROPTERA</i> Blumenbach, 1779
31 Dec 1816	<i>Macrotus</i> Leach	<i>Plecotus</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1816	<i>Phyllorhina</i> Leach	<i>Rhinolophus</i> Lacépède, 1799
31 Dec 1816	<i>Myopterus senegalensis</i> Oken	<i>Myopterus daubentonii</i> Desmarest, 1820
31 Dec 1816	<i>Pteropus madagascariensis</i> Oken	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
31 Dec 1816	<i>Rhinopoma breviceaudatum</i> Oken	<i>Rhinopoma cystops</i> Thomas, 1903
31 Dec 1817	<i>Vespertilio Kuhlii</i> Kuhl	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
31 Dec 1817	<i>Vespertilio Leisleri</i> Kuhl	<i>Nyctalus leisleri</i> (Kuhl, 1817)
31 Dec 1817	<i>Vespertilio mystacinus</i> Kuhl	<i>Myotis mystacinus</i> (Kuhl, 1817)
31 Dec 1817	<i>Vespertilio schreibersii</i> Kuhl	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
31 Dec 1818	<i>Myopterus</i> E. Geoffroy Saint-Hilaire	<i>Myopterus</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1818	<i>Nycteris Thebaicus</i> E. Geoffroy Saint-Hilaire	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1818	<i>Nyctinomus</i> E. Geoffroy Saint-Hilaire	<i>Tadarida</i> Rafinesque, 1814
31 Dec 1818	<i>Nyctinomus aegyptiacus</i> E. Geoffroy Saint-Hilaire	<i>Tadarida aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
31 Dec 1818	<i>Nyctinomus aegyptiacus</i> E. Geoffroy Saint-Hilaire	<i>Tadarida aegyptiaca aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
31 Dec 1818	<i>Plecotus</i> E. Geoffroy Saint-Hilaire	<i>Plecotus</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1818	<i>Pteropus aegyptiacus</i> E. Geoffroy Saint-Hilaire	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
31 Dec 1818	<i>Rhinopoma</i> E. Geoffroy Saint-Hilaire	<i>Rhinopoma</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1818	<i>Taphozous</i> E. Geoffroy Saint-Hilaire	<i>Taphozous</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1818	<i>Taphozous mauritanus</i> E. Geoffroy Saint-Hilaire	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1818	<i>Taphozous perforatus</i> E. Geoffroy Saint-Hilaire	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1818	<i>Taphozous perforatus</i> E. Geoffroy Saint-Hilaire	<i>Taphozous perforatus perforatus</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1819	<i>Vespertilio dasykarpos</i> Kuhl	<i>Nyctalus leisleri</i> (Kuhl, 1817)
1 Mar 1820	<i>Eptesicus</i> Rafinesque	<i>Eptesicus</i> Rafinesque, 1820
31 Dec 1820	<i>Myopterus</i> Desmarest	<i>Myopterus</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1820	<i>Myopterus daubentonii</i> Desmarest	<i>Myopterus daubentonii</i> Desmarest, 1820
31 Dec 1820	<i>Nycteris Geoffroyi</i> Desmarest	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1820	<i>Taphozous senegalensis</i> Desmarest	<i>Taphozous perforatus perforatus</i> E. Geoffroy St.-Hilaire, 1818
1 Apr 1821	<i>Barbastella</i> Gray	<i>Barbastella</i> Gray, 1821
1 Apr 1821	<i>Nyctimones</i> Gray	<i>Tadarida</i> Rafinesque, 1814
1 Apr 1821	<i>Pteropidae</i> Gray	<i>PTEROPODIDAE</i> Gray, 1821
1 Apr 1821	<i>Vespertilionidae</i> Gray	<i>VESPERTILIONIDAE</i> Gray, 1821
1 Apr 1821	<i>Vespertilioninae</i> Gray	<i>Vespertilioninae</i> Gray, 1821
1 Apr 1821	<i>Vespertilionini</i> Gray	<i>Vespertilionini</i> Gray, 1821
1 Apr 1821	<i>Vespertilionoidea</i> Gray	<i>VESPERTILIONOIDEA</i> Gray, 1821
23 Jun 1821	<i>Scotophilus</i> Leach	<i>Scotophilus</i> Leach, 1821
31 Dec 1821	<i>Nyctinoma</i> Bowdich	<i>Tadarida</i> Rafinesque, 1814
31 Dec 1821	<i>Rhynopoma</i> Bowdich	<i>Rhinopoma</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1821	<i>Cephalotidae</i> Gray	<i>PTEROPODIDAE</i> Gray, 1821
31 Dec 1821	<i>CHEIROPTERA</i> Gray	<i>CHIROPTERA</i> Blumenbach, 1779
31 Dec 1821	<i>FRUCTIVORAE</i> Gray	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 1821	<i>INSECTIVORAE</i> Gray	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 1821	<i>Pteropodinae</i> Gray	<i>Pteropodinae</i> Gray, 1821
31 Dec 1821	<i>Pteropodini</i> Gray	<i>Pteropodini</i> Gray, 1821
31 Dec 1821	<i>PTEROPODOIDEA</i> Gray	<i>PTEROPODOIDEA</i> Gray, 1821
31 Dec 1821	<i>Vespertilionini</i> Gray	<i>Vespertilioninae</i> Gray, 1821
31 Dec 1822	<i>CHEIROPTERA</i> Fleming	<i>CHIROPTERA</i> Blumenbach, 1779
31 Dec 1822	<i>Nyctinomia</i> Fleming	<i>Tadarida</i> Rafinesque, 1814
31 Dec 1823	<i>Pteropus collaris</i> Lichtenstein	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
31 Dec 1823	<i>Rhinolophus capensis</i> Lichtenstein	<i>Rhinolophus capensis</i> Lichtenstein, 1823

Publication date	Original Name	Current Name
31 Dec 1824	<i>Nyctinomus mauritanus</i> Horsfield	<i>Mormopterus acetabulosus acetabulosus</i> (Hermann, 1804)
30 Nov 1825	<i>Pteropina</i> Gray	<i>PTEROPODIDAE</i> Gray, 1821
30 Nov 1825	<i>Vespertilionina</i> Gray	<i>VESPERTILIONIDAE</i> Gray, 1821
31 Dec 1825	<i>Nyctalus</i> Bowdich	<i>Nyctalus</i> Bowdich, 1825
31 Dec 1825	<i>Nyctalus verrucosus</i> Bowdich	<i>Nyctalus leisleri</i> (Kuhl, 1817)
31 Dec 1825	<i>Rhinolophidae</i> Gray	<i>RHINOLOPHIDAE</i> Gray, 1825
31 Dec 1825	<i>Rhinolophina</i> Gray	<i>RHINOLOPHOIDEA</i> J. E. Gray, 1825
31 Dec 1825	<i>Rhinolophina</i> Gray	<i>RHINOLOPHIDAE</i> Gray, 1825
31 Dec 1825	<i>Rhinolophoidea</i> Gray	<i>RHINOLOPHOIDEA</i> J. E. Gray, 1825
31 Dec 1825	<i>Vespertilio Pygmaeus</i> Leach	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
31 Dec 1825	<i>Dinops</i> Savi	<i>Tadarida Rafinesque</i> , 1814
31 Dec 1825	<i>Dinops Cestonii</i> Savi	<i>Tadarida teniotis</i> (Rafinesque, 1814)
31 Dec 1825	<i>Pteropus geoffroyi</i> Temminck	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
31 Dec 1825	<i>Pteropus phaiops</i> Temminck	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
31 Dec 1826	<i>Dysopes pumilus</i> Cretzschmar	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
31 Dec 1826	<i>Nycticejus leucogaster</i> Cretzschmar	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
31 Dec 1826	<i>Vespertilio leucomelas</i> Cretzschmar	<i>Barbastella leucomelas</i> (Cretzschmar, 1826)
31 Dec 1826	<i>Vespertilio temminckii</i> Cretzschmar	<i>Vansonia rueppellii</i> (J.B. Fischer, 1829)
31 Dec 1827	<i>Vespertilio ferrugineus</i> Brehm	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
31 Dec 1827	<i>Vespertilio temminckii</i> Cretzschmar	<i>Vansonia rueppellii rueppellii</i> (J.B. Fischer, 1829)
31 Dec 1827	<i>Rhinolophina</i> Lesson	<i>RHINOLOPHIDAE</i> Gray, 1825
31 Dec 1827	<i>Dysopes geoffroyi</i> Temminck	<i>Tadarida aegyptiaca aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
31 Dec 1827	<i>Dysopes rüppelii</i> Temminck	<i>Tadarida teniotis</i> (Rafinesque, 1814)
31 Dec 1828	<i>Rhinolophus clivus</i> Cretzschmar	<i>Rhinolophus clivus</i> Cretzschmar, 1828
31 May 1829	<i>Nycteris affinis</i> A. Smith	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
31 May 1829	<i>Nycteris Capensis</i> A. Smith	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
31 May 1829	<i>Pteropus Leachii</i> A. Smith	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
31 May 1829	<i>Rhinolophus Geoffroyii</i> A. Smith	<i>Rhinolophus clivus</i> Cretzschmar, 1828
31 May 1829	<i>Vespertilio Capensis</i> A. Smith	<i>Laephotis capensis</i> (A. Smith, 1829)
30 Jun 1829	<i>Cercopteropus</i> Burnett	<i>Rousettus</i> Gray, 1821
31 Dec 1829	<i>[Vespertilio auritus] β aegyptius</i> J.B. Fischer	<i>Plecotus christii</i> Gray, 1838
31 Dec 1829	<i>[Vespertilio pipistrellus var.] β Aegyptius</i> J.B. Fischer	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
31 Dec 1829	<i>V[espertilio] Rüppelii</i> J.B. Fischer	<i>Vansonia rueppellii</i> (J.B. Fischer, 1829)
31 Dec 1829	<i>Vespertilio ruppelii</i> J.B. Fischer	<i>Vansonia rueppellii rueppellii</i> (J.B. Fischer, 1829)
31 Dec 1829	<i>Barbastellus</i> Kaup	<i>Barbastella</i> Gray, 1821
31 Dec 1829	<i>Cnephæus</i> Kaup	<i>Eptesicus (Cnephæus)</i> Kaup, 1829
31 Dec 1829	<i>Myotis</i> Kaup	<i>Myotis</i> Kaup, 1829
31 Dec 1829	<i>Nystactes</i> Kaup	<i>Myotis</i> Kaup, 1829
31 Dec 1829	<i>Pipistrellus</i> Kaup	<i>Pipistrellus</i> Kaup, 1829
31 Dec 1829	<i>Pterygistes</i> Kaup	<i>Nyctalus</i> Bowdich, 1825
31 Dec 1830	<i>Leuconoe</i> Boie	<i>Myotis</i> Kaup, 1829
31 Dec 1830	<i>Vespertilio marginatus</i> Cretzschmar	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
31 Dec 1831	<i>Phyllorrhina</i> Bonaparte	<i>Hipposideros</i> Gray, 1831
31 Dec 1831	<i>Pteropodina</i> Bonaparte	<i>Pteropodinae</i> Gray, 1821
31 Dec 1831	<i>Dysopes</i> Cretzschmar	<i>Tadarida Rafinesque</i> , 1814
31 Dec 1831	<i>Taphozous nudiventris</i> Cretzschmar	<i>Taphozous nudiventris</i> Cretzschmar, 1830
31 Dec 1831	<i>Hipposideros</i> Gray	<i>Hipposideros</i> Gray, 1831
31 Dec 1831	<i>Pachyotus</i> Gray	<i>Scotophilus</i> Leach, 1821
31 Dec 1832	<i>Pteropus hottentottus</i> Temminck	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
31 Dec 1832	<i>V[espertilio] tricolor</i> Temminck	<i>Myotis tricolor</i> (Temminck, 1832)
31 Dec 1832	<i>Vespertilio epichrysalis</i> Temminck	<i>Scotophilus dinganii</i> (A. Smith, 1833)
31 Dec 1832	<i>Vespertilio platycephalus</i> Temminck	<i>Pipistrellus (Pipistrellus) hesperidus</i> (Temminck, 1840)
30 Nov 1833	<i>Nyctinomus Condylurus</i> A. Smith	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
30 Nov 1833	<i>Nyctinomus dubius</i> A. Smith	<i>Mormopterus acetabulosus natalensis</i> (A. Smith, 1847)
30 Nov 1833	<i>Vespertilio Dinganii</i> A. Smith	<i>Scotophilus dinganii</i> (A. Smith, 1833)
30 Nov 1833	<i>Vespertilio Hottentota</i> A. Smith	<i>Eptesicus (Cnephæus) hottentotus</i> (A. Smith, 1833)
30 Nov 1833	<i>Vespertilio Natalensis</i> A. Smith	<i>Miniopterus natalensis</i> (A. Smith, 1833)
30 Jun 1834	<i>Vespertilio goudoti</i> A. Smith	<i>Myotis goudoti</i> (A. Smith, 1834)
26 Sep 1834	<i>Hipposideros</i> Gray	<i>Hipposideros</i> Gray, 1831
9 Oct 1835	<i>Pteropus Gambianus</i> Ogilby	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
9 Oct 1835	<i>Pteropus Gambianus</i> Ogilby	<i>Epomophorus gambianus</i> (Ogilby, 1835)
9 Oct 1835	<i>Pteropus macrocephalus</i> Ogilby	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
31 Dec 1835	<i>Pteropodae</i> Bonaparte	<i>Pteropodinae</i> Gray, 1821
31 Dec 1835	<i>Pteropus megacephalus</i> Swainson	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
31 Dec 1835	<i>Taphozous leucopterus</i> Temminck	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
12 Feb 1836	<i>Epomophorus</i> Bennett	<i>Epomophorus</i> Bennett, 1836
12 Feb 1836	<i>Pteropus epomophorus</i> Bennett	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
2 Oct 1836	<i>Pteropus Whitei</i> Bennett	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
31 Dec 1836	<i>Rhinocrepis</i> Gervais	<i>Rhinolophus Lacépède</i> , 1799
31 Dec 1837	<i>Miniopterus</i> Bonaparte	<i>Miniopterus</i> Bonaparte, 1837

Publication date	Original Name	Current Name
31 Dec 1837	<i>Noctula</i> Bonaparte	<i>Eptesicus</i> Rafinesque, 1820
31 Dec 1837	<i>Phyllorrhina</i> Bonaparte	<i>Hipposideros</i> Gray, 1831
31 Dec 1837	<i>Pipistrella</i> Bonaparte	<i>Pipistrellus</i> Kaup, 1829
31 Dec 1837	<i>Vespertilio Capaccinii</i> Bonaparte	<i>Myotis capaccinii</i> (Bonaparte, 1837)
31 Dec 1837	<i>Vespertilio Savii</i> Bonaparte	<i>Hypsugo savii</i> (Bonaparte, 1837)
31 Dec 1837	<i>Myoptera</i> de Blainville	<i>Myopterus</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1837	<i>Pteropus labiatus</i> Temminck	<i>Epomophorus labiatus</i> (Temminck, 1837)
31 Dec 1837	<i>Pteropus vulgaris</i> Temminck	<i>Pteropus subniger</i> (Kerr, 1792)
1 Feb 1838	<i>Asellia</i> Gray	<i>Asellia</i> Gray, 1838
1 Feb 1838	<i>Barbastella communis</i> Gray	<i>Barbastella barbastellus</i> (Schreber, 1774)
1 Feb 1838	<i>Lavia</i> Gray	<i>Lavia</i> Gray, 1838
1 Feb 1838	<i>Petalia</i> Gray	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795
1 Feb 1838	<i>Plecotus christii</i> Gray	<i>Plecotus christii</i> Gray, 1838
1 Feb 1838	<i>Romicia</i> Gray	<i>Pipistrellus</i> Kaup, 1829
25 May 1838	<i>Rhinolophus landeri</i> Martin	<i>Rhinolophus landeri</i> Martin, 1838
31 Dec 1838	PTEROPODIDAE Bonaparte	PTEROPODIDAE Gray, 1821
31 Dec 1838	<i>Rhinopomina</i> Bonaparte	RHINOPOMATIDAE Dobson, 1872
31 Dec 1838	<i>Pteropus phæops</i> Oken	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
31 Dec 1838	<i>Saccolaimus</i> Temminck	<i>Saccolaimus</i> Temminck, 1838
31 Dec 1839	<i>Synotus</i> Keyserling and Blasius	<i>Barbastella</i> Gray, 1821
31 Dec 1839	<i>Vesperugo</i> Keyserling and Blasius	<i>Pipistrellus</i> Kaup, 1829
31 Dec 1839	<i>Vesperugo</i> Keyserling and Blasius	<i>Eptesicus</i> Rafinesque, 1820
31 Dec 1839	<i>Vesperugo</i> Keyserling and Blasius	<i>Nyctalus</i> Bowdich, 1825
31 Dec 1839	<i>Vesperus</i> Keyserling and Blasius	<i>Eptesicus</i> Rafinesque, 1820
31 Dec 1839	<i>Vespertilio pachygnathus</i> Michahelles	<i>Nyctalus leisleri</i> (Kuhl, 1817)
31 Dec 1840	<i>Romicius</i> Blyth	<i>Pipistrellus</i> Kaup, 1829
31 Dec 1840	<i>Dysopes Savii</i> Schinz	<i>Tadarida teniotis</i> (Rafinesque, 1814)
31 Dec 1840	<i>Vespertilio dasythrix</i> Temminck	<i>Miniopterus natalensis</i> (A. Smith, 1833)
31 Dec 1840	<i>Vespertilio hesperida</i> Temminck	<i>Pipistrellus (Pipistrellus) hesperidus hesperidus</i> (Temminck, 1840)
31 Dec 1840	<i>Vespertilio hesperida</i> Temminck	<i>Pipistrellus (Pipistrellus) hesperidus</i> (Temminck, 1840)
31 Dec 1840	<i>Vespertilio minuta</i> Temminck	<i>Laephotis capensis</i> (A. Smith, 1829)
31 Dec 1840	<i>N[ycteris] albiventer</i> Wagner	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1840	<i>N[ycteris] discolor</i> Wagner	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1841	<i>Capaccinius</i> Bonaparte	<i>Myotis</i> Kaup, 1829
31 Dec 1841	<i>Selysius</i> Bonaparte	<i>Myotis</i> Kaup, 1829
31 Dec 1841	<i>Vespertilio isabellinus</i> Temminck	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
31 Dec 1841	<i>Vespertilio megalurus</i> Temminck	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
1 Dec 1842	<i>Kerivoula</i> Gray	<i>Kerivoula</i> Gray, 1842
1 Dec 1842	<i>Kerivoula poensis</i> Gray	<i>Glauconycteris poensis</i> (Gray, 1842)
1 Dec 1842	<i>Noctulinia</i> Gray	<i>Nyctalus</i> Bowdich, 1825
1 Dec 1842	<i>Trilatitus</i> Gray	<i>Myotis</i> Kaup, 1829
31 Dec 1842	Harpyidae C. H. Smith	PTEROPODIDAE Gray, 1821
31 Dec 1842	<i>Mops</i> Lesson	<i>Mops (Mops) Lesson</i> , 1842
31 Dec 1842	<i>Mops</i> Lesson	<i>Mops</i> Lesson, 1842
31 Dec 1842	<i>Nicticejus</i> Rüppell	<i>Scotophilus</i> Leach, 1821
31 Dec 1842	<i>Pteropus Schoënsis</i> Rüppell	<i>Epomophorus labiatus</i> (Temminck, 1837)
31 Dec 1842	<i>Rhinolophus fumigatus</i> Rüppell	<i>Rhinolophus fumigatus</i> Rüppell, 1842
31 Dec 1842	<i>Vespertilio Pipistrellus varietas africana</i> Rüppell	<i>Afronycteris nana</i> (Peters, 1852)
13 May 1843	<i>Nycteris poensis</i> Gray	<i>Nycteris hispida</i> (Schreber, 1774)
13 May 1843	<i>Vespertilio</i> Gray	<i>Myotis</i> Kaup, 1829
13 May 1843	<i>Xantharpyia</i> Gray	<i>Rousettus</i> Gray, 1821
31 Jul 1843	<i>Rhinolophus Martini</i> Fraser	<i>Nycteris hispida</i> (Schreber, 1774)
31 Dec 1843	<i>Dysopes midas</i> Sundevall	<i>Mops (Mops) midas midas</i> (Sundevall, 1843)
31 Dec 1843	<i>Dysopes midas</i> Sundevall	<i>Mops (Mops) midas</i> (Sundevall, 1843)
31 Dec 1843	<i>Gymnorhina</i> Wagner	VESPERTILIONIDAE Gray, 1821
31 Dec 1844	<i>Eleutherura</i> Gray	<i>Rousettus</i> Gray, 1821
31 Dec 1845	<i>Rhinolophus gigas</i> Wagner	<i>Macronycteris gigas</i> (Wagner, 1845)
31 Jul 1846	<i>Pteropus Haldemani</i> Hallowell	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
31 Dec 1846	<i>Pteropus Wahlbergi</i> Sundevall	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
31 Dec 1846	<i>Rhinolophus caffer</i> Sundevall	<i>Hipposideros caffer</i> (Sundevall, 1846)
31 Dec 1846	<i>Vespertilio scotinus</i> Sundevall	<i>Miniopterus natalensis</i> (A. Smith, 1833)
31 Dec 1846	<i>Vespertilio subtilis</i> Sundevall	<i>Pipistrellus (Pipistrellus) hesperidus subtilis</i> (Sundevall, 1846)
13 Apr 1847	<i>Aquias</i> Gray	<i>Rhinolophus</i> Lacépède, 1799
31 Dec 1847	<i>Eucheira</i> Hodgson	<i>Megaderma</i> E. Geoffroy St.-Hilaire, 1810
31 Dec 1847	<i>Dysopes natalensis</i> A. Smith	<i>Mormopterus acetabulosus natalensis</i> (A. Smith, 1847)
31 Dec 1847	<i>Vespertilio lanosus</i> A. Smith	<i>Kerivoula lanosa</i> (A. Smith, 1847)
31 Dec 1849	<i>Kirivoula</i> Gervais	<i>Kerivoula</i> Gray, 1842
31 Dec 1849	<i>Vespertilio minutus</i> A. Smith	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
31 Dec 1852	<i>Cynonycteris</i> Peters	<i>Rousettus</i> Gray, 1821
31 Dec 1852	<i>Dysopes brachypterus</i> Peters	<i>Mops (Xiphonycteris) brachypterus</i> (Peters, 1852)
31 Dec 1852	<i>Dysopes dubius</i> Peters	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)

Publication date	Original Name	Current Name
31 Dec 1852	<i>Dysopes limbatus</i> Peters	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
31 Dec 1852	<i>Emballonura afra</i> Peters	<i>Coleura afra</i> (Peters, 1852)
31 Dec 1852	<i>Epomophorus crypturus</i> Peters	<i>Epomophorus crypturus</i> Peters, 1852
31 Dec 1852	<i>Nycteris fuliginosa</i> Peters	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1852	<i>Nycteris villosa</i> Peters	<i>Nycteris hispida</i> (Schreber, 1774)
31 Dec 1852	<i>Nycticejus planirostris</i> Peters	<i>Scotophilus dinganii</i> (A. Smith, 1833)
31 Dec 1852	<i>Nycticejus viridis</i> Peters	<i>Scotophilus viridis</i> (Peters, 1852)
31 Dec 1852	<i>Phyllorhina caffa</i> Peters	<i>Hipposideros caffer</i> (Sundevall, 1846)
31 Dec 1852	<i>Phyllorhina gracilis</i> Peters	<i>Hipposideros caffer</i> (Sundevall, 1846)
31 Dec 1852	<i>Phyllorhina patellifera</i> Peters	<i>Hipposideros caffer</i> (Sundevall, 1846)
31 Dec 1852	<i>Phyllorhina vittata</i> Peters	<i>Macronycteris vittatus</i> (Peters, 1852)
31 Dec 1852	<i>Rhinolophus lobatus</i> Peters	<i>Rhinolophus lobatus</i> Peters, 1852
31 Dec 1852	<i>Vespertilio nanus</i> Peters	<i>Afronycteris nana</i> (Peters, 1852)
31 Dec 1853	<i>Rhinolophus euryale</i> Blasius	<i>Rhinolophus euryale</i> Blasius, 1853
31 Dec 1853	<i>Pachysoma</i> Temminck	<i>Epomophorus</i> Bennett, 1836
31 Dec 1853	<i>Phyllorhina cyclops</i> Temminck	<i>Doryrhina cyclops</i> (Temminck, 1853)
31 Dec 1853	<i>Phyllorhina fuliginosa</i> Temminck	<i>Hipposideros fuliginosus</i> (Temminck, 1853)
31 Dec 1853	<i>Rhinolophus alcyone</i> Temminck	<i>Rhinolophus alcyone</i> Temminck, 1853
31 Dec 1853	<i>Taphozous peli</i> Temminck	<i>Saccolaimus peli</i> (Temminck, 1853)
31 Dec 1854	<i>Rhinolophides</i> Gervais	<i>RHINOLOPHIDAE</i> Gray, 1825
23 Jul 1855	<i>Emballonuridae</i> Gervais	<i>EMBALLONURIDAE</i> Gervais, 1855
23 Jul 1855	<i>Emballonurinae</i> Gervais	<i>Emballonurinae</i> Gervais, 1855
23 Jul 1855	<i>Nycticeiini</i> Gervais	<i>Nycticeiini</i> Gervais, 1855
23 Jul 1855	<i>Nycticeina</i> Gervais	<i>Nycticeiini</i> Gervais, 1855
28 Sep 1855	<i>V[espertilio] Smithii</i> Wagner	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
23 Dec 1855	<i>Emballonurini</i> Gervais	<i>Emballonurini</i> Gervais, 1855
31 Dec 1855	<i>FRUGIVORA</i> Giebel	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 1855	<i>Gymnorhina</i> Giebel	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 1855	<i>Histiorhina</i> Van der Hoeven	<i>RHINOLOPHIDAE</i> Gray, 1825
31 Dec 1855	<i>Nycteridae</i> Van der Hoeven	<i>NYCTERIDAE</i> Van der Hoeven, 1855
31 Dec 1855	<i>Nycterioidea</i> Van der Hoeven	<i>NYCTEROIDEA</i> Van der Hoeven, 1855
31 Dec 1855	<i>Vespertilio smithii</i> Wagner	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
23 Jul 1856	<i>Molossidae</i> Gervais	<i>MOLOSSIDAE</i> Gervais, 1856
23 Jul 1856	<i>Molossinae</i> Gervais	<i>Molossinae</i> Gervais, 1856
23 Jul 1856	<i>Nycticeina</i> Gervais	<i>VESPERTILIONIDAE</i> Gray, 1821
31 Dec 1856	<i>Molossoidae</i> Gervais	<i>MOLOSSOIDEA</i> Gervais, 1856
31 Dec 1856	<i>Brachyotus Kolenati</i>	<i>Myotis</i> Kaup, 1829
31 Dec 1856	<i>Cateorus Kolenati</i>	<i>Eptesicus Rafinesque</i> , 1820
31 Dec 1856	<i>Hypsugo Kolenati</i>	<i>Hypsugo Kolenati</i> , 1856
31 Dec 1856	<i>Isotus Kolenati</i>	<i>Myotis</i> Kaup, 1829
31 Dec 1856	<i>Meteorus Kolenati</i>	<i>Eptesicus Rafinesque</i> , 1820
31 Dec 1856	<i>Nannugo Kolenati</i>	<i>Pipistrellus</i> Kaup, 1829
31 Dec 1856	<i>Panugo Kolenati</i>	<i>Nyctalus Bowdich</i> , 1825
31 Dec 1857	<i>Vespertilio pusillus</i> Leconte	<i>Laephotis "incertae-sedis"</i>
27 Apr 1858	<i>Vespertilio Madagascariensis</i> Tomes	<i>Myotis goudoti</i> (A. Smith, 1834)
31 Dec 1858	<i>Amblyotus Kolenati</i>	<i>Eptesicus Rafinesque</i> , 1820
31 Dec 1858	<i>Nannugo Kolenatii</i> Müller	<i>Pipistrellus (Pipistrellus) "incertae-sedis"</i>
30 Jun 1859	<i>Scotophilus darwini</i> Tomes	<i>Hypsugo savii</i> (Bonaparte, 1837)
31 Dec 1859	<i>Nycticejus Schlieffenii</i> Peters	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
31 Dec 1859	<i>Otonycteris Peters</i>	<i>Otonycteris Peters</i> , 1859
31 Dec 1859	<i>Otonycteris Hemprichii</i> Peters	<i>Otonycteris hemprichii</i> Peters, 1859
31 Dec 1859	<i>Rhinopoma Lepsianum</i> Peters	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
31 May 1860	<i>Epomophorus franqueti</i> Tomes	<i>Epomops franqueti</i> (Tomes, 1860)
31 Dec 1860	<i>Phyllorhina Koch</i>	<i>HIPPOSIDERIDAE</i> Lydekker, 1891
31 Dec 1860	<i>Phyllorhina Koch</i>	<i>HIPPOSIDERIDAE</i> Lydekker, 1891
31 Dec 1860	<i>Rhinolophus auritus</i> Sundevall	<i>Rhinolophus capensis</i> Lichtenstein, 1823
31 May 1861	<i>Kerivoula argentata</i> Tomes	<i>Kerivoula argentata</i> Tomes, 1861
31 May 1861	<i>Scotophilus rusticus</i> Tomes	<i>Pipistrellus (Pipistrellus) rusticus</i> (Tomes, 1861)
31 May 1861	<i>Scotophilus variegatus</i> Tomes	<i>Glauconycteris variegata</i> (Tomes, 1861)
31 Dec 1861	<i>Nyctophylax Fitzinger</i>	<i>Kerivoula</i> Gray, 1842
31 Dec 1861	<i>Nycteris labiata</i> Heuglin	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1861	<i>Nycticejus flavigaster</i> Heuglin	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
31 Dec 1861	<i>Nycticejus murino-flavus</i> Heuglin	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
31 Dec 1861	<i>Nyctinomus (Dysopes) ventralis</i> Heuglin	<i>Tadarida ventralis</i> (Heuglin, 1861)
31 Dec 1861	<i>Nyctinomus bivittatus</i> Heuglin	<i>Mops (Chaerephon) bivittatus</i> (Heuglin, 1861)
31 Dec 1861	<i>Phyllorhina bicornis</i> Heuglin	<i>Hipposideros tephurus</i> (Cabrera, 1906)
31 Dec 1861	<i>Phyllorhina megalotis</i> Heuglin	<i>Hipposideros megalotis</i> (Heuglin, 1861)
31 Dec 1861	<i>Rhinolophus acrotis</i> Heuglin	<i>Rhinolophus clivus</i> Cretzschmar, 1828
31 Dec 1861	<i>Rhinolophus minimus</i> Heuglin	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
30 Jun 1862	<i>Sphyrocephalus A. Murray</i>	<i>Hypsignathus</i> H. Allen, 1862
30 Jun 1862	<i>Sphyrocephalus labrosus</i> A. Murray	<i>Hypsignathus monstrosus</i> H. Allen, 1862
30 Jun 1862	<i>Zygænocephalus A. Murray</i>	<i>Hypsignathus</i> H. Allen, 1862

Publication date	Original Name	Current Name
31 Dec 1862	<i>Epomophorus comptus</i> H. Allen	<i>Epomops franqueti</i> (Tomes, 1860)
31 Dec 1862	<i>H[y]psignathus</i> <i>monstrosus</i> H. Allen	<i>Hypsignathus monstrosus</i> H. Allen, 1862
31 Dec 1862	<i>Hypsignathus</i> H. Allen	<i>Hypsignathus</i> H. Allen, 1862
31 Dec 1862	<i>Pteropus mollipilosus</i> H. Allen	<i>Eidolon helvum helvum</i> (Kerr, 1792)
31 Dec 1862	<i>Pterocyon</i> Peters	<i>Eidolon</i> Rafinesque, 1815
31 Dec 1862	<i>Pterocyon paleaceus</i> Peters	<i>Eidolon helvum helvum</i> (Kerr, 1792)
31 Dec 1863	<i>Aristippe</i> Kolenati	<i>Eptesicus</i> Rafinesque, 1820
31 Dec 1864	<i>Megadermatidae</i> H. Allen	<i>MEGADERMATIDAE</i> H. Allen, 1864
31 Dec 1864	<i>Dysopes hepaticus</i> Heuglin	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
31 Dec 1864	<i>Epomophorus anurus</i> Heuglin	<i>Epomophorus labiatus</i> (Temminck, 1837)
31 May 1865	<i>Nycticejus nidicola</i> Kirk	<i>Kerivoula argentata</i> Tomes, 1861
30 Jun 1865	<i>Pteropus palmarum</i> Heuglin	<i>Eidolon helvum helvum</i> (Kerr, 1792)
31 Oct 1865	<i>Nyctinomus (Mormopterus) jugularis</i> Peters	<i>Mormopterus jugularis</i> (Peters, 1865)
31 Dec 1865	<i>Brachyura</i> Peters	<i>EMBALLONURIDAE</i> Gervais, 1855
31 Dec 1865	<i>Molossi</i> Peters	<i>MOLOSSIDAE</i> Gervais, 1856
31 Dec 1865	<i>Mormopterus</i> Peters	<i>Mormopterus</i> Peters, 1865
31 Dec 1865	<i>Nycteris grandis</i> Peters	<i>Nycteris grandis</i> Peters, 1865
31 Dec 1865	<i>Nyctophilinae</i> Peters	<i>Vespertilioninae</i> Gray, 1821
31 Dec 1865	<i>Rhinolophi</i> Peters	<i>RHINOLOPHIDAE</i> Gray, 1825
2 Feb 1866	<i>Nyctericina</i> Gray	<i>NYCTERIDAE</i> Van der Hoeven, 1855
1 Feb 1866	<i>Diclidurina</i> Gray	<i>EMBALLONURIDAE</i> Gervais, 1855
1 Feb 1866	<i>Emballonurina</i> Gray	<i>EMBALLONURIDAE</i> Gervais, 1855
1 Feb 1866	<i>Molossina</i> Gray	<i>Molossinae</i> Gervais, 1856
1 Feb 1866	<i>Molossina</i> Gray	<i>MOLOSSIDAE</i> Gervais, 1856
1 Feb 1866	<i>Nyctophilina</i> Gray	<i>VESPERTILIONIDAE</i> Gray, 1821
1 Feb 1866	<i>Pachyomus</i> Gray	<i>Eptesicus</i> Rafinesque, 1820
1 Feb 1866	<i>Plecotina</i> Gray	<i>VESPERTILIONIDAE</i> Gray, 1821
1 Feb 1866	<i>Plecotini</i> Gray	<i>Plecotini</i> Gray, 1866
1 Feb 1866	<i>Romiciana</i> Gray	<i>VESPERTILIONIDAE</i> Gray, 1821
31 May 1866	<i>Chrysonycteris</i> Gray	<i>Hipposideros</i> Gray, 1831
31 May 1866	<i>Epomophorina</i> Gray	<i>Pteropodinae</i> Gray, 1821
31 May 1866	<i>Epomophorini</i> Gray	<i>Epomophorini</i> Gray, 1866
31 May 1866	<i>Eunycteris</i> Gray	<i>Pteropus</i> Brisson, 1762
31 May 1866	<i>Gloionycteris</i> Gray	<i>Hipposideros</i> Gray, 1831
31 May 1866	<i>Macronycteris</i> Gray	<i>Macronycteris</i> Gray, 1866
31 May 1866	<i>Megadermina</i> Gray	<i>MEGADERMATIDAE</i> H. Allen, 1864
31 May 1866	<i>Nycterops</i> Gray	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795
31 May 1866	<i>Nycterops pilosa</i> Gray	<i>Nycteris hispida</i> (Schreber, 1774)
31 May 1866	<i>Phyllotis</i> Gray	<i>Rhinolophus</i> Lacépède, 1799
31 May 1866	<i>Pteropus livingstonii</i> Gray	<i>Pteropus livingstonii</i> Gray, 1866
31 May 1866	<i>Rhinonycterina</i> Gray	<i>RHINONYCTERIDAE</i> J. E. Gray, 1866
31 May 1866	<i>Rhinophylla</i> Gray	<i>Hipposideros</i> Gray, 1831
31 May 1866	<i>Speorifera</i> Gray	<i>Hipposideros</i> Gray, 1831
30 Sep 1866	<i>Scotophilus welwitschii</i> Gray	<i>Myotis welwitschii</i> (Gray, 1866)
31 Dec 1866	<i>Rhinopoma longicaudatum</i> Fitzinger	<i>Rhinopoma cystops</i> Thomas, 1903
31 Dec 1866	<i>Rhinopoma sennaariense</i> Fitzinger	<i>Rhinopoma cystops</i> Thomas, 1903
31 Dec 1866	<i>Xantharpyia leucomelas</i> Fitzinger	<i>Eidolon helvum helvum</i> (Kerr, 1792)
31 Dec 1866	<i>Plecotus ustus</i> Fitzinger and Heuglin	<i>Otonycteris hemprichii</i> Peters, 1859
31 Dec 1866	<i>Nycteris Baikii</i> Gray	<i>Nycteris grandis</i> Peters, 1865
31 Dec 1866	<i>Rhinonycteridae</i> Gray	<i>RHINONYCTERIDAE</i> J. E. Gray, 1866
31 Dec 1866	<i>Scotophilina</i> Gray	<i>Scotophilinae</i> Van Cakenberghe and Seamark, 2008
31 Dec 1866	<i>Spasma</i> Gray	<i>Megaderma</i> E. Geoffroy St.-Hilaire, 1810
31 Dec 1866	<i>Histiorrhina</i> Haeckel	<i>RHINOLOPHIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 1866	<i>Hypodermida</i> Haeckel	<i>PTEROPODIDAE</i> Gray, 1821
31 Dec 1866	<i>Nycterides</i> Haeckel	<i>CHIROPTERA</i> Blumenbach, 1779
31 Dec 1866	<i>Pterocynes</i> Haeckel	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 1866	<i>Pteropodida</i> Haeckel	<i>PTEROPODIDAE</i> Gray, 1821
31 Dec 1866	<i>Plecotus aethiopicus</i> Heuglin	<i>Plecotus christii</i> Gray, 1838
31 Dec 1866	<i>Xantharpyia leucomelas</i> Heuglin	<i>Eidolon helvum</i> (Kerr, 1792)
31 Dec 1866	<i>Plecotus aethiopicus</i> Heuglin and Fitzinger	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1866	<i>Vesperugo hypoleucus</i> Heuglin and Fitzinger	<i>Vansonia rueppellii rueppellii</i> (J.B. Fischer, 1829)
31 Dec 1866	<i>Vesperugo sennaariensis</i> Heuglin and Fitzinger	<i>Vansonia rueppellii rueppellii</i> (J.B. Fischer, 1829)
31 Dec 1866	<i>Entomophaga</i> A. Murray	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 1866	<i>Scotophili</i> A. Murray	<i>Scotophilinae</i> Van Cakenberghe and Seamark, 2008
31 Dec 1866	<i>Megadermata</i> Peters	<i>MEGADERMATIDAE</i> H. Allen, 1864
31 Dec 1866	<i>Pteropi</i> Peters	<i>PTEROPODIDAE</i> Gray, 1821
31 Dec 1866	<i>Vespertiliones</i> Peters	<i>VESPERTILIONIDAE</i> Gray, 1821
30 Apr 1867	<i>Cœloephyllus</i> Peters	<i>Rhinolophus</i> Lacépède, 1799

Publication date	Original Name	Current Name
30 Apr 1867	<i>Coelophyllus</i> Peters	<i>Rhinolophus</i> Lacépède, 1799
30 Apr 1867	<i>Pteropus dupréanus</i> Schlegel	<i>Eidolon dupreanum</i> (Schegel, 1867)
31 Dec 1867	<i>Taphozoinae</i> Jerdon	<i>Taphozoinae</i> Jerdon, 1867
31 Dec 1867	<i>Rhinolophus algius</i> Loche	<i>Rhinolophus euryale</i> Blasius, 1853
31 Dec 1867	<i>Chalinolobus</i> Peters	<i>Glauconycteris</i> Dobson, 1875
31 Dec 1867	<i>Colëura</i> Peters	<i>Coleura</i> Peters, 1867
31 Dec 1867	<i>Miniopterus minor</i> Peters	<i>Miniopterus minor</i> Peters, 1867
31 Dec 1867	<i>Rhinolophus blasii</i> Peters	<i>Rhinolophus blasii</i> Peters, 1867
31 Dec 1867	<i>Vespertilio</i> (<i>Pteropterus</i>) Peters	<i>Myotis</i> Kaup, 1829
31 Dec 1868	<i>Nycteris Geoffroyi</i> Var. <i>Senegalensis</i> Hartmann	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1868	<i>Rhinopoma sennarensense</i> Hartmann	<i>Rhinopoma cystops</i> Thomas, 1903
31 Dec 1868	<i>V[espertilio noctula] var. sicula</i> Mina-Palumbo	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
31 Dec 1868	<i>Alobus</i> Peters	<i>Pipistrellus</i> Kaup, 1829
31 Dec 1868	<i>Epomophorus pusillus</i> Peters	<i>Epomophorus pusillus</i> Peters, 1868
31 Dec 1868	<i>Pteropterus</i> Peters	<i>Myotis</i> Kaup, 1829
31 Dec 1868	<i>Rhinolophus Deckenii</i> Peters	<i>Rhinolophus deckenii</i> Peters, 1868
31 Dec 1868	<i>Vesperus</i> (<i>Hesperoptenus</i>) <i>kraussii</i> Peters	<i>Glauconycteris poensis</i> (Gray, 1842)
30 Sep 1869	<i>Nyctinomus leucogaster</i> A. Grandidier	<i>Mops</i> (<i>Chaerephon</i>) <i>leucogaster</i> (A. Grandidier, 1869)
30 Sep 1869	<i>Nyctinomus Miarensis</i> A. Grandidier	<i>Mops</i> (<i>Mops</i>) <i>midas miarensis</i> (A. Grandidier, 1869)
31 Dec 1869	<i>Gymnorhinida</i> Fatio	<i>VESPERTILIONIDAE</i> Gray, 1821
31 Dec 1869	<i>Vesperugo noctula var. maxima</i> Fatio	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
31 Dec 1869	<i>Coleura seychellensis</i> Peters	<i>Coleura seychellensis</i> Peters, 1868
31 Dec 1869	<i>Rhinolophus aethiops</i> Peters	<i>Rhinolophus fumigatus</i> Rüppell, 1842
31 Dec 1869	<i>Vesperus Bottae</i> Peters	<i>Eptesicus</i> (<i>Cnephaeus</i>) <i>bottae</i> (Peters, 1869)
28 Feb 1870	<i>Nyctinomus unicolor</i> A. Grandidier	<i>Mops</i> (<i>Mops</i>) <i>midas miarensis</i> (A. Grandidier, 1869)
28 Feb 1870	<i>Vespertilio sylvicola</i> A. Grandidier	<i>Myotis goudoti</i> (A. Smith, 1834)
31 Oct 1870	<i>Aëorestes</i> Fitzinger	<i>Myotis</i> Kaup, 1829
31 Dec 1870	<i>Comastes</i> Fitzinger	<i>Myotis</i> Kaup, 1829
31 Dec 1870	<i>Exochurus</i> Fitzinger	<i>Myotis</i> Kaup, 1829
31 Dec 1870	<i>Nyctiptenus</i> Fitzinger	<i>Eptesicus Rafinesque</i> , 1820
31 Dec 1870	<i>Rhinolophus Eggenhöfner</i> Fitzinger	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
31 Dec 1870	<i>[Epomophorus macrocephalus] var. angolensis</i> Gray	<i>Epomophorus angolensis</i> Gray, 1870
31 Dec 1870	<i>[Epomophorus macrocephalus] var. unicolor</i> Gray	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
31 Dec 1870	<i>Cynopterus collaris</i> Gray	<i>Myonycteris torquata</i> (Dobson, 1878)
31 Dec 1870	<i>Eleutherura unicolor</i> Gray	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
31 Dec 1870	<i>Epomophorus macrocephalus var. unicolor</i> Gray	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
31 Dec 1870	<i>Epomops</i> Gray	<i>Epomops</i> Gray, 1866
31 Dec 1870	<i>Pselaphon</i> Gray	<i>Pteropus</i> Brisson, 1762
31 Dec 1870	<i>Senonycteris</i> Gray	<i>Rousettus</i> Gray, 1821
31 Dec 1870	<i>Spectrum vulgare</i> Gray	<i>Pteropus niger</i> (Kerr, 1792)
31 Dec 1870	<i>Stenonycteris</i> Gray	<i>Stenonycteris</i> Gray, 1870
31 Dec 1870	<i>Nyctinomus angolensis</i> Peters	<i>Mops</i> (<i>Mops</i>) <i>condylurus</i> (A. Smith, 1833)
31 Dec 1870	<i>Vespertilio Bocagii</i> Peters	<i>Myotis bocagii</i> (Peters, 1870)
31 Dec 1870	<i>Vespertilio Bocagii</i> Peters	<i>Myotis bocagii bocagii</i> (Peters, 1870)
31 Dec 1870	<i>Vesperugo pusillulus</i> Peters	<i>Afronycteris nana</i> (Peters, 1852)
28 Dec 1871	<i>Triaenops</i> Dobson	<i>Triaenops</i> Dobson, 1871
31 Dec 1871	<i>Cyclorhina</i> Peters	<i>Hipposideros</i> Gray, 1831
31 Dec 1871	<i>Doryrhina</i> Peters	<i>Doryrhina</i> Peters, 1871
31 Dec 1871	<i>Nycteris angolensis</i> Peters	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1871	<i>Nycteris damarensis</i> Peters	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1871	<i>Ptychorhina</i> Peters	<i>Hipposideros</i> Gray, 1831
31 Dec 1871	<i>Sideroderma</i> Peters	<i>Hipposideros</i> Gray, 1831
31 Dec 1871	<i>Sybdesmotus</i> Peters	<i>Hipposideros</i> Gray, 1831
31 Dec 1871	<i>Syndesmotis</i> Peters	<i>Hipposideros</i> Gray, 1831
31 Dec 1871	<i>Thyreorhina</i> Peters	<i>Hipposideros</i> Gray, 1831
1 May 1872	<i>Noctulina</i> Macalister	<i>Nyctalus Bowdich</i> , 1825
30 Nov 1872	<i>Nycticejinae</i> Gill	<i>Nycticeiini</i> Gervais, 1855
31 Dec 1872	<i>Rhinopomatidae</i> Dobson	<i>RHINOPOMATIDAE</i> Dobson, 1872
31 Dec 1872	<i>Rhinopomatoidea</i> Dobson	<i>RHINOPOMATOIDEA</i> Dobson, 1872
31 Dec 1872	<i>ANIMALIVORA</i> Gill	<i>CHIROPTERA</i> Blumenbach, 1779
31 Dec 1872	<i>Megadermidae</i> Gill	<i>MEGADERMATIDAE</i> H. Allen, 1864
31 Dec 1872	<i>Molossidae</i> Gill	<i>MOLOSSIDAE</i> Gervais, 1856
31 Dec 1872	<i>Lyroderma</i> Peters	<i>Megaderma</i> E. Geoffroy St.-Hilaire, 1810
31 Dec 1872	<i>Megaderma cor</i> Peters	<i>Cardiaderma cor</i> (Peters, 1872)
31 Dec 1872	<i>Vesperus tenuipinnis</i> Peters	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
23 Jun 1873	<i>Cardiaderma</i> Peters	<i>Cardiaderma</i> Peters, 1873
17 Oct 1874	<i>Nyctinomus</i> (<i>Chaerephon</i>) Dobson	<i>Mops</i> (<i>Chaerephon</i>) Dobson, 1874
31 Dec 1874	<i>Emballonura atrata</i> Peters	<i>Paremballonura atrata</i> (Peters, 1874)
1 Aug 1875	<i>Scotophilus gigas</i> Dobson	<i>Scotophilus nigrita</i> (Schreber, 1774)

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1 Oct 1875	<i>Vespertilio murinus africanus</i> Dobson	<i>Myotis punicus</i> Felten, 1977
31 Oct 1875	<i>Chalinolobus argentatus</i> Dobson	<i>Glauconycteris argentata</i> (Dobson, 1875)
31 Oct 1875	<i>Glauconycteris</i> Dobson	<i>Glauconycteris</i> Dobson, 1875
31 Oct 1875	<i>Vesperugo pulcher</i> Dobson	<i>Vansonia rueppellii pulcher</i> (Dobson, 1875)
1 Nov 1875	<i>Emballonuridae</i> Dobson	<i>EMBALLONURIDAE</i> Gervais, 1855
1 Nov 1875	<i>Miniopteri</i> Dobson	<i>MINIOPTERIDAE</i> Dobson, 1875
1 Nov 1875	<i>Miniopteridae</i> Dobson	<i>MINIOPTERIDAE</i> Dobson, 1875
31 Dec 1875	<i>MEGACHIROPTERA</i> Dobson	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 1875	<i>MICROCHIROPTERA</i> Dobson	<i>CHIROPTERA</i> Blumenbach, 1779
30 Apr 1876	<i>Taphonycteris</i> Dobson	<i>Saccolaimus</i> Temminck, 1838
1 May 1876	<i>Nyctinomus africanus</i> Dobson	<i>Tadarida ventralis</i> (Heuglin, 1861)
1 Dec 1876	<i>Vesperugo (Vesperus) grandidieri</i> Dobson	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
31 Dec 1876	<i>Nycteris macrotis</i> Dobson	<i>Nycteris macrotis</i> Dobson, 1876
31 Dec 1876	<i>Epomophorus macrocephalus</i> Peters	<i>Hypsiphatius monstrosus</i> H. Allen, 1862
30 Apr 1877	<i>Nyctinomus [(Mormopterus)] albiventer</i> Dobson	<i>Mormopterus jugularis</i> (Peters, 1865)
30 Apr 1877	<i>Nyctinomus [(Nyctinomus)] cestoni</i> Dobson	<i>Tadarida ventralis</i> (Heuglin, 1861)
31 Dec 1877	<i>Dysopes talpinus</i> Heuglin	<i>Tadarida aegyptiaca aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
31 Dec 1877	<i>Nycticejus] adovanus</i> Heuglin	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
31 Dec 1877	<i>Nycticejus eriophorus</i> Heuglin	<i>Kerivoula eriophora</i> (Heuglin, 1877)
31 Dec 1877	<i>Nycticejus serratus</i> Heuglin	<i>Taphozous nudiventris</i> Cretzschmar, 1830
31 Dec 1877	<i>Rhinolophus macrocephalus</i> Heuglin	<i>Rhinolophus fumigatus</i> Rüppell, 1842
31 Dec 1877	<i>Rhinopoma cordofanicum</i> Heuglin	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
31 Dec 1877	<i>Taphozous maritimus</i> Heuglin	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1877	<i>Vesperugo senarensis</i> Heuglin	<i>Vansonia rueppellii rueppellii</i> (J.B. Fischer, 1829)
31 Dec 1877	<i>Pteropus seychellensis</i> A. Milne-Edwards	<i>Pteropus seychellensis</i> A. Milne-Edwards, 1877
31 Dec 1877	<i>Triænopis afer</i> Peters	<i>Triænopis afer</i> Peters, 1877
22 Jun 1878	<i>Myzopoda</i> A. Milne-Edwards and A. Grandidier	<i>Myzopoda</i> Milne-Edwards and A. Grandidier, 1878
22 Jun 1878	<i>Myzopoda aurita</i> A. Milne-Edwards and A. Grandidier	<i>Myzopoda aurita</i> Milne-Edwards and A. Grandidier, 1878
30 Jun 1878	<i>Cynonycteris torquata</i> Dobson	<i>Myonycteris torquata</i> (Dobson, 1878)
30 Jun 1878	<i>Cynonycteris torquata</i> Dobson	<i>Myonycteris torquata torquata</i> (Dobson, 1878)
30 Jun 1878	<i>Kerivoula africana</i> Dobson	<i>Kerivoula africana</i> Dobson, 1878
30 Jun 1878	<i>Kerivoula brunnea</i> Dobson	<i>Kerivoula lanosa</i> (A. Smith, 1847)
30 Jun 1878	<i>Nycteris æthiopica</i> Dobson	<i>Nycteris macrotis</i> Dobson, 1876
30 Jun 1878	<i>Plecotus ægyptiacus</i> Dobson	<i>Plecotus christii</i> Gray, 1838
30 Jun 1878	<i>Pteropus rodricensis</i> Dobson	<i>Pteropus rodricensis</i> Dobson, 1878
30 Jun 1878	<i>Vesperugo [(Vesperugo)] maderensis</i> Dobson	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
31 Dec 1878	<i>Vespertiliones</i> Dobson	<i>Vespertilioninae</i> Gray, 1821
31 Dec 1878	<i>Mormopterus setiger</i> Peters	<i>Platymops setiger setiger</i> (Peters, 1878)
31 Dec 1878	<i>Mormopterus setiger</i> Peters	<i>Platymops setiger</i> (Peters, 1878)
31 Dec 1878	<i>Rhinolophus Hildebrandtii</i> Peters	<i>Rhinolophus hildebrandtii</i> Peters, 1878
30 Apr 1879	<i>Nyctinomus Bemmeleni</i> Jentink	<i>Mops (Chaerephon) bemmeleni</i> (Jentink, 1879)
30 Apr 1879	<i>Taphozous Dobsoni</i> Jentink	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
30 Apr 1880	<i>Epomophorus minor</i> Dobson	<i>Epomophorus minor</i> Dobson, 1880
1 Aug 1880	<i>Kerivoula Smithii</i> Thomas	<i>Kerivoula smithii</i> Thomas, 1880
1 Aug 1880	<i>Vesperugo (Vesperus) brunneus</i> Thomas	<i>Pseudoromicia brunnea</i> (Thomas, 1880)
31 Dec 1880	<i>Pteropus comorensis</i> Wallace	<i>Pteropus seychellensis</i> A. Milne-Edwards, 1877
30 Apr 1881	<i>Leiponyx</i> Jentink	<i>Eidolon Rafinesque</i> , 1815
30 Apr 1881	<i>Leiponyx büttikoferi</i> Jentink	<i>Eidolon helvum helvum</i> (Kerr, 1792)
31 Dec 1881	<i>Phyllorhina tridens murraiana</i> Anderson	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
31 Dec 1881	<i>Scotophilus robustus</i> A. Milne-Edwards	<i>Scotophilus robustus</i> A. Milne-Edwards, 1881
31 Dec 1881	<i>Triænopis Humbloti</i> A. Milne-Edwards	<i>Triænopis menamena</i> Goodman and Ranivo, 2009
31 Dec 1881	<i>Triænopis rufus</i> A. Milne-Edwards	<i>Triænopis menamena</i> Goodman and Ranivo, 2009
31 Dec 1881	<i>Vesperus Humbloti</i> A. Milne-Edwards	<i>Laephotis humbloti</i> (A. Milne-Edwards, 1881)
31 Dec 1881	<i>Nycteris Revouillii</i> Robin	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1882	<i>Liponyx</i> Forbes	<i>Eidolon Rafinesque</i> , 1815
31 Dec 1883	<i>Chiropteri</i> de Rochebrune	<i>CHIROPTERA</i> Blumenbach, 1779
31 Dec 1883	<i>Phyllorhinidae</i> de Rochebrune	<i>RHINOLOPHIDAE</i> Gray, 1825
27 Apr 1885	<i>Megaloglossus Pagenstecher</i>	<i>Megaloglossus Pagenstecher</i> , 1885
27 Apr 1885	<i>Megaloglossus woermanni</i> Pagenstecher	<i>Megaloglossus woermanni</i> Pagenstecher, 1885
22 May 1885	<i>Rhinolophus antinorii</i> Dobson	<i>Rhinolophus fumigatus</i> Rüppell, 1842
31 Dec 1885	<i>Taphozous perforatus</i> var. <i>assabensis</i> Monticelli	<i>Taphozous nudiventris</i> Cretzschmar, 1830
7 May 1887	<i>Phyllorhina commersonii</i> var. <i>marungensis</i> Noack	<i>Macronycteris vittatus</i> (Peters, 1852)
7 May 1887	<i>Scotophilus minimus</i> Noack	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
25 Jul 1887	<i>Vesperugo (Vesperus) Innesi</i> Lataste	<i>Eptesicus (Cnephaeus) bottae</i> (Peters, 1869)
31 Dec 1887	<i>Vespertiliavus</i> Schlosser	<i>Vespertiliavus</i> Schlosser, 1887

Publication date	Original Name	Current Name
30 Apr 1888	<i>Epomophorus veldkampii</i> Jentink	<i>Nanonycteris veldkampii</i> (Jentink, 1888)
30 Apr 1888	<i>Vesperugo stamplii</i> Jentink	<i>Afronycteris nana</i> (Peters, 1852)
31 Dec 1888	<i>Necromanter</i> Lydekker	<i>Necromantis</i> Weithofer, 1887
31 Dec 1888	<i>Necromantis</i> Weithofer	<i>Necromantis</i> Weithofer, 1887
28 Feb 1889	<i>Chalinolobus conigicus</i> Noack	<i>Glauconycteris argentata</i> (Dobson, 1875)
28 Feb 1889	<i>Vesperugo pagenstecheri</i> Noack	<i>Afronycteris nana</i> (Peters, 1852)
28 Feb 1889	<i>Vesperus damarensis</i> Noack	<i>Laephotis capensis</i> (A. Smith, 1829)
28 Feb 1889	<i>Vesperus pusillus</i> Noack	<i>Afronycteris nana</i> (Peters, 1852)
31 Mar 1889	<i>Epomophorus Dobsonii</i> Bocage	<i>Epomophorus dobsonii</i> Bocage, 1889
31 Mar 1889	<i>Vesperus bicolor</i> Bocage	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
31 Mar 1889	<i>Vesperus guineensis</i> Bocage	<i>Neoromicia guineensis</i> (Bocage, 1889)
1 Apr 1889	<i>Vesperugo (Vesperus) Rendalli</i> Thomas	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
31 Dec 1889	<i>Gymnuridae</i> Ameghino	MOLOSSIDAE Gervais, 1856
31 Dec 1889	<i>Ptética</i> Ameghino	CHIROPTERA Blumenbach, 1779
31 Dec 1889	<i>Cynonycteris brachycephala</i> Bocage	<i>Myonycteris brachycephala</i> (Bocage, 1889)
31 Dec 1889	<i>Miniopterus Newtoni</i> Bocage	<i>Miniopterus newtoni</i> Bocage, 1889
3 Feb 1890	<i>Scotophilus albobfuscus</i> Thomas	<i>Scotoecus albobfuscus</i> (Thomas, 1890)
1 Mar 1891	<i>Nyctinomus lobatus</i> Thomas	<i>Tadarida lobata</i> (Thomas, 1891)
1 Jun 1891	<i>Vesperugo (Vesperus) Moloneyi</i> Thomas	<i>Mimetillus moloneyi</i> (Thomas, 1891)
31 Aug 1891	<i>N(yctinomus) taeniotis</i> Thomas	<i>Tadarida ventralis</i> (Heuglin, 1861)
30 Sep 1891	<i>Phyllorhina commersoni</i> var. <i>thomensis</i> Bocage	<i>Macronycteris vittatus</i> (Peters, 1852)
31 Dec 1891	<i>Cerivoula</i> Blanford	<i>Kerivoula</i> Gray, 1842
31 Dec 1891	<i>Hipposideridae</i> Lydekker	HIPPOSIDERIDAE Lydekker, 1891
31 Dec 1891	<i>Hipposiderinae</i> Lydekker	HIPPOSIDERIDAE Lydekker, 1891
31 Dec 1891	<i>Trygenycteris</i> Lydekker	<i>Megaloglossus</i> Pagenstecher, 1885
19 Jan 1892	<i>Adelonycteris</i> H. Allen	<i>Eptesicus</i> Rafinesque, 1820
14 Jul 1893	<i>Pteropus aldabrensis</i> True	<i>Pteropus aldabrensis</i> True, 1893
23 Dec 1893	<i>Phyllorhina rubra</i> Noack	<i>Hipposideros ruber</i> (Noack, 1893)
31 Dec 1893	<i>Emballonurini</i> Winge	EMBALLONURIDAE Gervais, 1855
31 Dec 1893	<i>Megadermatini</i> Winge	MEGADERMATIDAE H. Allen, 1864
31 Dec 1893	<i>Molossini</i> Winge	MOLOSSIDAE Gervais, 1856
31 Dec 1893	<i>Pteropi</i> Winge	PTEROPODIDAE Gray, 1821
31 Dec 1893	<i>Rhinolophini</i> Winge	RHINOLOPHIDAE Gray, 1825
31 Dec 1894	<i>Scotonycteris</i> Matschie	<i>Scotonycteris</i> Matschie, 1894
31 Dec 1894	<i>Scotonycteris zenkeri</i> Matschie	<i>Scotonycteris zenkeri</i> Matschie, 1894
5 Apr 1895	<i>Pteropus pteropus</i> Merriam	<i>Pteropus niger</i> (Kerr, 1792)
16 Oct 1897	<i>Plecotinæ</i> Miller	<i>Plecotini</i> Gray, 1866
16 Oct 1897	<i>Vespertilioninæ</i> Miller	<i>Vespertilioninae</i> Gray, 1821
31 Oct 1897	<i>Nyctinomus martiensseni</i> Matschie	<i>Otomops martiensseni</i> (Matschie, 1897)
31 Oct 1897	<i>Nyctinomus martiensseni</i> Matschie	<i>Otomops martiensseni martiensseni</i> (Matschie, 1897)
1 Dec 1897	<i>Rhinolophus micaceus</i> de Winton	<i>Doryrhina cyclops</i> (Temminck, 1853)
31 Dec 1897	<i>midas</i> Schulze	<i>Tadarida teniotis</i> (Rafinesque, 1814)
31 Dec 1897	<i>[Nyctinomus (Nyctinomus) pumilus]</i> Var. <i>Major</i> Trouessart	<i>Mops (Chaerephon) major</i> (Trouessart, 1897)
31 Dec 1897	<i>[Vesperugo (Vesperus) serotinus]</i> Var. <i>Gabonensis</i> Trouessart	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
31 Dec 1897	<i>Pteropinae</i> Trouessart	<i>Pteropodinae</i> Gray, 1821
22 Jan 1898	<i>Rhinolophus Maclaudi</i> Pousargues	<i>Rhinolophus maclaudi</i> Pousargues, 1898
30 Jun 1898	<i>Cynonycteris Angolensis</i> Bocage	<i>Myonycteris angolensis</i> (Bocage, 1898)
30 Jun 1898	<i>Cynonycteris Angolensis</i> Bocage	<i>Myonycteris angolensis angolensis</i> (Bocage, 1898)
30 Jun 1898	<i>Epomophorus guineensis</i> Bocage	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
30 Jun 1898	<i>Epomophorus guineensis</i> Bocage	<i>Epomophorus gambianus</i> (Ogilby, 1835)
31 Dec 1898	<i>Phyllorhina angolensis</i> Seabra	<i>Hipposideros caffer</i> (Sundevall, 1846)
31 Dec 1898	<i>Rhinolophus angolensis</i> Seabra	<i>Rhinolophus lobatus</i> Peters, 1852
31 Dec 1898	<i>Miniopterae</i> Trouessart	MINIOPTERIDAE Dobson, 1875
31 Dec 1898	<i>Miomegaderma gaillardi</i> Trouessart	<i>Megaderma gaillardi</i> (Trouessart, 1898)
1 Nov 1899	<i>Scotophilus hirundo</i> de Winton	<i>Scotoecus hirundo</i> (de Winton, 1899)
1 Nov 1899	<i>Scotophilus nigrnellus</i> de Winton	<i>Scotophilus nigrnellus</i> de Winton, 1899
31 Dec 1899	<i>Euvespertilio</i> Acloque	<i>Myotis</i> Kaup, 1829
31 Dec 1899	<i>Euvesperugo</i> Acloque	<i>Pipistrellus</i> Kaup, 1829
31 Dec 1899	<i>[Epomophorus ([Nanonycteris])] Matschie</i>	<i>Nanonycteris</i> Matschie, 1899
31 Dec 1899	<i>[Xantharpyia ([Myonycteris])] Matschie</i>	<i>Myonycteris</i> Matschie, 1899
31 Dec 1899	<i>Ep[omophorus (Epomophorus)] büttikoferi</i> Matschie	<i>Epomops buettikoferi</i> (Matschie, 1899)
31 Dec 1899	<i>Ep[omophorus (Epomophorus)] doriae</i> Matschie	<i>Epomophorus labiatus</i> (Temminck, 1837)
31 Dec 1899	<i>Ep[omophorus (Epomophorus)] neumanni</i> Matschie	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
31 Dec 1899	<i>Ep[omophorus (Epomophorus)] stuhlmanni</i> Matschie	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
31 Dec 1899	<i>Ep[omophorus (Epomophorus)] zechi</i> Matschie	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)

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31 Dec 1899	<i>Ep[omophorus (Epomophorus)] zenkeri</i> Matschie	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
31 Dec 1899	<i>Hypsignathus haldemani</i> Matschie	<i>Hypsignathus monstrosus</i> H. Allen, 1862
31 Dec 1899	<i>Mynonycteris</i> Matschie	<i>Rousettus</i> Gray, 1821
31 Dec 1899	<i>Sericonycteris</i> Matschie	<i>Pteropus</i> Brisson, 1762
31 Dec 1899	<i>Vespertilio venustus</i> Matschie	<i>Myotis welwitschii</i> (Gray, 1866)
31 Aug 1900	<i>Epomophorus anchietæ</i> Seabra	<i>Plerotes anchietæ</i> (Seabra, 1900)
31 Aug 1900	<i>Nyctinomus Anchietæ</i> Seabra	<i>Tadarida aegyptiaca bocagei</i> (Seabra, 1900)
31 Aug 1900	<i>Nyctinomus Bocagei</i> Seabra	<i>Tadarida aegyptiaca bocagei</i> (Seabra, 1900)
31 Aug 1900	<i>Nyctinomus brunneus</i> Seabra	<i>Tadarida aegyptiaca bocagei</i> (Seabra, 1900)
31 Aug 1900	<i>Vesperugo Anchietæ</i> Seabra	<i>Neoromicia anchietæ</i> (Seabra, 1900)
1 Dec 1900	<i>Mormopterus Whitleyi</i> Scharff	<i>Myopterus whitleyi</i> (Scharff, 1900)
28 Dec 1900	<i>Pipistrellus minusculus</i> Miller	<i>Afronycteris nana</i> (Peters, 1852)
31 Dec 1900	<i>Vesperugo (Vesperus) flavescens</i> Seabra	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
1 Jan 1901	<i>Glauconycteris Floweri</i> de Winton	<i>Eptesicus (Rhinopterus) floweri</i> (de Winton, 1901)
1 Jan 1901	<i>N(yctinomus) midas</i> de Winton	<i>Tadarida ventralis</i> (Heuglin, 1861)
1 Jan 1901	<i>Nyctinomus Emini</i> de Winton	<i>Mops (Chaerephon) major</i> (Trouessart, 1897)
1 Jan 1901	<i>Nyctinomus gambianus</i> de Winton	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1 Mar 1901	<i>Scotæcus</i> Thomas	<i>Scotoecus</i> Thomas, 1901
1 Mar 1901	<i>Scotæcus Hindei</i> Thomas	<i>Scotoecus hindei</i> Thomas, 1901
30 Apr 1901	<i>Kerivoula harrisoni</i> Thomas	<i>Kerivoula lanosa</i> (A. Smith, 1847)
1 May 1901	<i>Læphotis</i> Thomas	<i>Laephotis</i> Thomas, 1901
1 May 1901	<i>Læphotis Wintoni</i> Thomas	<i>Laephotis wintoni</i> Thomas, 1901
1 Jul 1901	<i>Clæotis</i> Thomas	<i>Cloeotis</i> Thomas, 1901
1 Jul 1901	<i>Clæotis Percivali</i> Thomas	<i>Cloeotis percivali</i> Thomas, 1901
1 Jul 1901	<i>Nycteris aethiopica luteola</i> Thomas	<i>Nycteris macrotis</i> Dobson, 1876
1 Jul 1901	<i>Pipistrellus Kuhlii fuscatus</i> Thomas	<i>Pipistrellus (Pipistrellus) hesperidus hesperidus</i> (Temminck, 1840)
1 Jul 1901	<i>Pterygistes azureum</i> Thomas	<i>Nyctalus azureum</i> (Thomas, 1901)
1 Jul 1901	<i>Vespertilio minutus somalicus</i> Thomas	<i>Neoromicia somalica</i> (Thomas, 1901)
1 Jul 1901	<i>Vespertilio platyops</i> Thomas	<i>Eptesicus (Cnephaeus) platyops</i> (Thomas, 1901)
1 Sep 1901	<i>Glauconycteris beatrix</i> Thomas	<i>Glauconycteris beatrix</i> Thomas, 1901
31 Dec 1901	<i>Euryalus</i> Matschie	<i>Rhinolophus</i> Lacépède, 1799
31 Dec 1901	<i>Rhinolophus mehelyi</i> Matschie	<i>Rhinolophus mehelyi</i> Matschie, 1901
31 Oct 1902	<i>Pipistrellus deserti</i> Thomas	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
16 Dec 1902	<i>Nyctinomus pusillus</i> Miller	<i>Mops (Chaerephon) pusillus</i> (Miller, 1902)
31 Dec 1902	<i>Otonycteris petersi</i> Anderson	<i>Otonycteris hemprichii</i> Peters, 1859
31 Dec 1902	<i>Syndesmotus</i> C.O. Waterhouse	<i>Hipposideros</i> Gray, 1831
1 May 1903	<i>Rhinopoma cystops</i> Thomas	<i>Rhinopoma cystops</i> Thomas, 1903
29 May 1903	<i>Necronycteris</i> Palmer	<i>Necromantis</i> Weithofer, 1887
1 Nov 1903	<i>Nyctinomus cisturus</i> Thomas	<i>Mops (Chaerephon) bemmeleni</i> (Jentink, 1879)
1 Nov 1903	<i>Nyctinomus demonstrator</i> Thomas	<i>Mops (Mops) demonstrator</i> (Thomas, 1903)
1 Nov 1903	<i>Nyctinomus fulminans</i> Thomas	<i>Tadarida fulminans</i> (Thomas, 1903)
1 Dec 1903	<i>Miniopterus inflatus</i> Thomas	<i>Miniopterus inflatus</i> Thomas, 1903
1 Dec 1903	<i>Miniopterus inflatus</i> Thomas	<i>Miniopterus inflatus inflatus</i> Thomas, 1903
1 Dec 1903	<i>Nycteris arge</i> Thomas	<i>Nycteris arge</i> Thomas, 1903
1 Dec 1903	<i>Nyctinomus thersites</i> Thomas	<i>Mops (Xiphonycteris) thersites</i> (Thomas, 1903)
1 Mar 1904	<i>Myotis Hildegardeæ</i> Thomas	<i>Myotis bocagii bocagii</i> (Peters, 1870)
1 Mar 1904	<i>Nyctinomus hindei</i> Thomas	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1 Mar 1904	<i>Pipistrellus crassulus</i> Thomas	<i>Nycticeinops crassulus</i> (Thomas, 1904)
1 Mar 1904	<i>Scotophilus nigrita colias</i> Thomas	<i>Scotophilus dinganii</i> (A. Smith, 1833)
1 Mar 1904	<i>Scotophilus nigrita nux</i> Thomas	<i>Scotophilus nux</i> Thomas, 1904
8 Mar 1904	<i>Scotonycteris bedfordi</i> Thomas	<i>Scotonycteris zenkeri</i> Matschie, 1894
1 May 1904	<i>Hipposideros Commersoni mostellum</i> Thomas	<i>Macronycteris vittatus</i> (Peters, 1852)
1 May 1904	<i>Rhinolophus Denti</i> Thomas	<i>Rhinolophus denti</i> Thomas, 1904
1 Jun 1904	<i>Myotis Bocagei cupreolus</i> Thomas	<i>Myotis bocagii cupreolus</i> Thomas, 1904
1 Aug 1904	<i>Pipistrellus ariel</i> Thomas	<i>Hypsugo ariel</i> (Thomas, 1904)
1 Aug 1904	<i>Rhinolophus Andersoni</i> Thomas	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1 Aug 1904	<i>Rhinolophus Dobsoni</i> Thomas	<i>Rhinolophus landeri</i> Martin, 1838
31 Aug 1904	<i>Pipistrellus nanulus</i> Thomas	<i>Pipistrellus (Pipistrellus) nanulus</i> Thomas, 1904
1 Oct 1904	<i>Myzopodidae</i> Thomas	<i>MYZOPODIDAE</i> Thomas, 1904
1 Nov 1904	<i>Rhinolophus augur</i> K. Andersen	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1 Nov 1904	<i>Rhinolophus augur zambesiensis</i> K. Andersen	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1 Nov 1904	<i>Rhinolophus augur zuluensis</i> K. Andersen	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1 Nov 1904	<i>Rhinolophus simulator</i> K. Andersen	<i>Rhinolophus simulator</i> K. Andersen, 1904
22 Nov 1904	<i>Mimetillus</i> Thomas	<i>Mimetillus</i> Thomas, 1904
1 Dec 1904	<i>Rhinolophus empusa</i> K. Andersen	<i>Rhinolophus blasii</i> Peters, 1867
15 Dec 1904	<i>Kehelvoulha</i> Jentink	<i>Kerivoula</i> Gray, 1842
31 Dec 1904	<i>E[uryalus] atlanticus</i> K. Andersen and Matschie	<i>Rhinolophus euryale</i> Blasius, 1853
31 Dec 1904	<i>E[uryalus] barbarus</i> K. Andersen and Matschie	<i>Rhinolophus mehelyi</i> Matschie, 1901

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31 Dec 1904	<i>E[uryalus] Cabreræ</i> K. Andersen and Matschie	<i>Rhinolophus euryale</i> Blasius, 1853
31 Dec 1904	<i>E[uryalus] meridionalis</i> K. Andersen and Matschie	<i>Rhinolophus mehelyi</i> Matschie, 1901
31 Dec 1904	<i>Vespertilio Boscai</i> Cabrera	<i>Eptesicus (Cnephæus) isabellinus</i> (Temminck, 1840)
31 Dec 1904	<i>Vespertilio ochromixtus</i> Cabrera	<i>Hypsugo savii</i> (Bonaparte, 1837)
31 Dec 1904	<i>Epomophorus pousarguesi</i> Trouessart	<i>Epomophorus gambianus pousarguesi</i> Trouessart, 1904
1 Jan 1905	<i>Rhinolophus acrotis brachygnatus</i> K. Andersen	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1 Jan 1905	<i>Rhinolophus Darlingi</i> K. Andersen	<i>Rhinolophus darlingi</i> K. Andersen, 1905
1 Jan 1905	<i>Rhinolophus fumigatus exsul</i> K. Andersen	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1 Jan 1905	<i>Rhinolophus Hildebrandti eloquens</i> K. Andersen	<i>Rhinolophus eloquens</i> K. Andersen, 1905
1 Jan 1905	<i>Glauconycteris papilio</i> Thomas	<i>Glauconycteris variegata</i> (Tomes, 1861)
10 Aug 1905	<i>Vespertilio capensis gracilior</i> Thomas and Schwann	<i>Laephotis capensis</i> (A. Smith, 1829)
10 Aug 1905	<i>Vespertilio matroka</i> Thomas and Schwann	<i>Laephotis matroka</i> (Thomas and Schwann, 1905)
30 Sep 1905	<i>Rhinolophus andreinii</i> Senna	<i>Rhinolophus blasii</i> Peters, 1867
7 Oct 1905	<i>Rhinolophus hipposiderus typicus</i> K. Andersen	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1 Nov 1905	<i>Eomops</i> Thomas	<i>Myotis</i> E. Geoffroy St.-Hilaire, 1818
9 Dec 1905	<i>Lavia rex</i> Miller	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
1 Jan 1906	<i>H[ipposiderus]. gigas gambiensis</i> K. Andersen	<i>Macronycteris gigas</i> (Wagner, 1845)
1 Jan 1906	<i>Pterygistes madeiræ</i> Barrett-Hamilton	<i>Nyctalus leisleri</i> (Kuhl, 1817)
1 Feb 1906	<i>Miniopterus Majori</i> Thomas	<i>Miniopterus majori</i> Thomas, 1906
1 Feb 1906	<i>Miniopterus manavi</i> Thomas	<i>Miniopterus manavi</i> Thomas, 1906
1 Feb 1906	<i>Scotophilus damarensis</i> Thomas	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
1 Feb 1906	<i>Scotophilus nigrita herero</i> Thomas	<i>Scotophilus dinganii</i> (A. Smith, 1833)
1 Mar 1906	<i>Hipposiderus beatus</i> K. Andersen	<i>Hipposideros beatus</i> (K. Andersen, 1906)
1 Mar 1906	<i>Hipposiderus caffer centralis</i> K. Andersen	<i>Hipposideros ruber</i> (Noack, 1893)
1 Mar 1906	<i>Hipposiderus caffer guineensis</i> K. Andersen	<i>Hipposideros ruber</i> (Noack, 1893)
1 May 1906	<i>Platymops</i> Thomas	<i>Platymops</i> Thomas, 1906
1 May 1906	<i>Platymops Macmillani</i> Thomas	<i>Platymops setiger macmillani</i> Thomas, 1906
4 Jun 1906	<i>Pizonyx</i> Miller	<i>Myotis</i> Kaup, 1829
4 Jun 1906	<i>Rhinopterus</i> Miller	<i>Eptesicus (Rhinopterus)</i> Miller, 1906
7 Jun 1906	<i>Miniopterus fraterculus</i> Thomas and Schwann	<i>Miniopterus fraterculus</i> Thomas and Schwann, 1906
30 Jun 1906	<i>Hipposiderus tephros</i> Cabrera	<i>Hipposideros tephros</i> (Cabrera, 1906)
1 Aug 1906	<i>Rousettus lanosus</i> Thomas	<i>Stenonycteris lanosus</i> (Thomas, 1906)
1 Oct 1906	<i>Kerivoula muscilla</i> Thomas	<i>Kerivoula lanosa</i> (A. Smith, 1847)
21 Jan 1907	<i>Nyctinomus aloysii-sabaudiae</i> Festa	<i>Mops (Chaerephon) aloysiisabaudiae</i> (Festa, 1907)
1 Feb 1907	<i>Lavia frons affinis</i> K. Andersen and Wroughton	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
26 Feb 1907	<i>Triaenops furcula</i> Trouessart	<i>Paratriaenops furculus</i> (Trouessart, 1907)
29 Jun 1907	<i>Desmaplex</i> Miller	<i>Pteropus</i> Brisson, 1762
29 Jun 1907	<i>Diclidurinae</i> Miller	<i>EMBALLONURIDAE</i> Gervais, 1855
29 Jun 1907	<i>Hipposideridae</i> Miller	<i>HIPPOSIDERIDAE</i> Lydekker, 1891
29 Jun 1907	<i>Kerivoulinae</i> Miller	<i>Kerivoulinae</i> Miller, 1907
29 Jun 1907	<i>Nyctophilinae</i> Miller	<i>VESPERTILIONIDAE</i> Gray, 1821
29 Jun 1907	<i>Rhinopomidae</i> Miller	<i>RHINOPOMATIDAE</i> Dobson, 1872
1 Sep 1907	<i>Pteropus mascarinus</i> Mason	<i>Pteropus rodricensis</i> Dobson, 1878
1 Dec 1907	<i>Plecotus teneriffae</i> Barrett-Hamilton	<i>Plecotus teneriffae</i> Barrett-Hamilton, 1907
30 Apr 1908	<i>Rhinolophus swinnyi</i> Gough	<i>Rhinolophus swinnyi</i> Gough, 1908
31 Jul 1908	<i>Scabrifer</i> G.M. Allen	<i>Laephotis</i> Thomas, 1901
31 Jul 1908	<i>Scabrifer notius</i> G.M. Allen	<i>Laephotis capensis</i> (A. Smith, 1829)
1 Oct 1908	<i>Pteropus rufus princeps</i> K. Andersen	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
1 Oct 1908	<i>Nyctinomus leonis</i> Thomas	<i>Mops (Xiphonycteris) brachypterus</i> (Peters, 1852)
1 Oct 1908	<i>Rousettus smithii</i> Thomas	<i>Myonycteris angolensis smithii</i> (Thomas, 1908)
30 Oct 1908	<i>Scoteinus schlieffeni albiventer</i> Thomas and Wroughton	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
30 Oct 1908	<i>Scoteinus schlieffeni australis</i> Thomas and Wroughton	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
30 Oct 1908	<i>Scoteinus schlieffeni bedouin</i> Thomas and Wroughton	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
1 Nov 1908	<i>Myonycteris leptodon</i> K. Andersen	<i>Myonycteris leptodon</i> K. Andersen, 1908
1 Nov 1908	<i>Myonycteris wroughtoni</i> K. Andersen	<i>Myonycteris torquata wroughtoni</i> K. Andersen, 1908
1 Dec 1908	<i>Chœrephon websteri</i> Dollman	<i>Mops (Chaerephon) leucogaster</i> (A. Grandidier, 1869)
31 Dec 1908	<i>Rousettus sjöstedti</i> Lönnberg	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
31 Dec 1908	<i>Pteropus comorensis</i> Nicoll	<i>Pteropus seychellensis</i> A. Milne-Edwards, 1877
1 Aug 1909	<i>Taphozous hildegardeæ</i> Thomas	<i>Taphozous hildegardeæ</i> Thomas, 1909
31 Oct 1909	<i>Pteropus (Spectrum) voeltzkowi</i> Matschie	<i>Pteropus voeltzkowi</i> Matschie, 1909
1 Nov 1909	<i>Miniopterus breyeri</i> Jameson	<i>Miniopterus natalensis</i> (A. Smith, 1833)

Publication date	Original Name	Current Name
1 Dec 1909	<i>Rousettus kempfi</i> Thomas	<i>Stenonycteris lanosus</i> (Thomas, 1906)
1 Dec 1909	<i>Scotoecus albigula</i> Thomas	<i>Scotoecus albigula</i> Thomas, 1909
31 Dec 1909	<i>Otonycteris cinereus</i> Satunin	<i>Otonycteris hemprichii</i> Peters, 1859
1 Jan 1910	<i>Epomops franqueti strepitans</i> K. Andersen	<i>Epomops franqueti</i> (Tomes, 1860)
1 Jan 1910	<i>Plerotes</i> K. Andersen	<i>Plerotes</i> K. Andersen, 1910
31 Jan 1910	<i>Chrysopteron</i> Jentink	<i>Myotis</i> Kaup, 1829
1 Feb 1910	<i>Rhinolophus brockmani</i> Thomas	<i>Rhinolophus blasii</i> Peters, 1867
1 Jul 1910	<i>Casinycteris</i> Thomas	<i>Casinycteris</i> Thomas, 1910
1 Jul 1910	<i>Casinycteris argynnis</i> Thomas	<i>Casinycteris argynnis</i> Thomas, 1910
31 Dec 1910	<i>Petalidæ</i> Miller	<i>NYCTERIDAE</i> Van der Hoeven, 1855
31 Dec 1910	<i>Vampyravus</i> Schlosser	<i>Vampyravus</i> Schlosser, 1910
31 Dec 1910	<i>Vampyravus orientalis</i> Schlosser	<i>Vampyravus orientalis</i> Schlosser, 1910
31 Dec 1910	<i>Rhinolophus blasiusi</i> Trouessart	<i>Rhinolophus blasii</i> Peters, 1867
1 Feb 1911	<i>Xiphonycteris</i> Dollman	<i>Mops (Xiphonycteris)</i> Dollman, 1911
1 Feb 1911	<i>Xiphonycteris spurrelli</i> Dollman	<i>Mops (Xiphonycteris) spurrelli</i> (Dollman, 1911)
29 Apr 1911	<i>Pipistrellus culex</i> Thomas	<i>Afronycteris nana</i> (Peters, 1852)
1 Oct 1911	<i>Chærephon emini</i> Wroughton	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
31 Dec 1911	<i>Eptesicus phasma</i> G.M. Allen	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
31 Dec 1911	<i>Nycticeius africanus</i> G.M. Allen	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
31 Dec 1911	<i>Provampyrus</i> Schlosser	<i>Vampyravus</i> Schlosser, 1910
31 Dec 1911	<i>Provampyrus orientalis</i> Schlosser	<i>Vampyravus orientalis</i> Schlosser, 1910
25 Jan 1912	<i>Triænonops aurita</i> G. Grandidier	<i>Paratriaenops auritus</i> (G. Grandidier, 1912)
23 Mar 1912	<i>Cynopterinae</i> K. Andersen	<i>Pteropodinae</i> Gray, 1821
23 Mar 1912	<i>Epomophorinae</i> K. Andersen	<i>Rousettinae</i> Andersen, 1912
23 Mar 1912	<i>Rousettinae</i> K. Andersen	<i>Rousettinae</i> Andersen, 1912
23 Mar 1912	<i>Rousettini</i> K. Andersen	<i>Rousettini</i> Andersen, 1912
1 Jul 1912	<i>Kerivoula cuprosa</i> Thomas	<i>Kerivoula cuprosa</i> Thomas, 1912
1 Aug 1912	<i>Cistugo</i> Thomas	<i>Cistugo</i> Thomas, 1912
1 Aug 1912	<i>Cistugo seabrae</i> Thomas	<i>Cistugo seabrae</i> Thomas, 1912
1 Sep 1912	<i>Kerivoula phalaena</i> Thomas	<i>Kerivoula phalaena</i> Thomas, 1912
1 Nov 1912	<i>Petalia aurita</i> K. Andersen	<i>Nycteris aurita</i> (K. Andersen, 1912)
1 Nov 1912	<i>Petalia damarensis brockmani</i> K. Andersen	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1 Nov 1912	<i>Petalia damarensis media</i> K. Andersen	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1 Nov 1912	<i>Petalia gambiensis</i> K. Andersen	<i>Nycteris gambiensis</i> (K. Andersen, 1912)
1 Nov 1912	<i>Petalia major</i> K. Andersen	<i>Nycteris major</i> (K. Andersen, 1912)
1 Nov 1912	<i>Petalia nana</i> K. Andersen	<i>Nycteris nana</i> (K. Andersen, 1912)
4 Nov 1912	<i>Miniopterus natalensis arenarius</i> Heller	<i>Miniopterus arenarius</i> Heller, 1912
4 Nov 1912	<i>Pipistrellus aero</i> Heller	<i>Pipistrellus (Pipistrellus) aero</i> Heller, 1912
4 Nov 1912	<i>Pipistrellus helios</i> Heller	<i>Afronycteris helios</i> (Heller, 1912)
1 Jan 1913	<i>Glauconycteris egeria</i> Thomas	<i>Glauconycteris egeria</i> Thomas, 1913
6 Feb 1913	<i>Rhinolophus swinnyi piriensis</i> Hewitt	<i>Rhinolophus swinnyi</i> Gough, 1908
1 Mar 1913	<i>Chærephon nigeriæ</i> Thomas	<i>Mops (Chærephon) nigeriæ nigeriæ</i> (Thomas, 1913)
1 Mar 1913	<i>Chærephon nigeriæ</i> Thomas	<i>Mops (Chærephon) nigeriæ</i> (Thomas, 1913)
1 Mar 1913	<i>Nyctinomus ansorgei</i> Thomas	<i>Mops (Chærephon) ansorgei</i> (Thomas, 1913)
1 Mar 1913	<i>Pipistrellus fuscipes</i> Thomas	<i>Vansonia rueppellii fuscipes</i> (Thomas, 1913)
1 Mar 1913	<i>Pipistrellus musciculus</i> Thomas	<i>Hypsugo musciculus</i> (Thomas, 1913)
1 Mar 1913	<i>Rhinolophus foxi</i> Thomas	<i>Rhinolophus fumigatus</i> Rüppell, 1842
30 Apr 1913	<i>Otomops</i> Thomas	<i>Otomops</i> Thomas, 1913
20 Oct 1913	<i>Pipistrellus nanus australis</i> Roberts	<i>Afronycteris nana</i> (Peters, 1852)
27 Mar 1914	<i>Rhinolophus bembanicus</i> Senna	<i>Rhinolophus simulator</i> K. Andersen, 1904
1 Jun 1914	<i>Nycteris woodi</i> K. Andersen	<i>Nycteris woodi</i> K. Andersen, 1914
31 Jul 1914	<i>Scotophilus altilis</i> G.M. Allen	<i>Scotophilus altilis</i> G.M. Allen, 1914
31 Dec 1914	<i>acrotis</i> G.M. Allen	<i>Rhinolophus fumigatus</i> Rüppell, 1842
31 Dec 1914	<i>Noctulinia</i> Cabrera	<i>Nyctalus</i> Bowdich, 1825
1 Jun 1915	<i>C[oleura] g[allarum] nilosa</i> Thomas	<i>Coleura afra</i> (Peters, 1852)
1 Jun 1915	<i>C[oleura] gallarum</i> Thomas	<i>Coleura gallarum</i> Thomas, 1915
1 Jun 1915	<i>C[oleura] silhouettæ</i> Thomas	<i>Coleura seychellensis</i> Peters, 1868
1 Jun 1915	<i>Glauconycteris phalaena</i> Thomas	<i>Glauconycteris variegata</i> (Tomes, 1861)
1 Jun 1915	<i>Rhinopterus lowei</i> Thomas	<i>Eptesicus (Rhinopterus) floweri</i> (de Winton, 1901)
1 Jun 1915	<i>Taphozous sudani</i> Thomas	<i>Taphozous perforatus sudani</i> Thomas, 1915
30 Sep 1915	<i>Taphozous perforatus hædinus</i> Thomas	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
1 Nov 1915	<i>Scotoecus falabæ</i> Thomas	<i>Scotoecus hindei</i> Thomas, 1901
1 Dec 1915	<i>Myopterus albatu</i> Thomas	<i>Myopterus daubentonii</i> Desmarest, 1820
10 Feb 1916	<i>Chaerephon pumilus naivashæ</i> Hollister	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
10 Feb 1916	<i>Eptesicus ugandæ</i> Hollister	<i>Neoromicia somalica</i> (Thomas, 1901)
10 Feb 1916	<i>Rhinolophus keniensis</i> Hollister	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
31 Dec 1916	<i>Eptesicus rectitragus</i> Wettstein	<i>Neoromicia guineensis</i> (Bocage, 1889)
31 Dec 1916	<i>Nyctinomus (Nyctinomus) tongaënsis</i> Wettstein	<i>Tadarida aegyptiaca aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
31 Dec 1916	<i>Scotoecus cinnamomeus</i> Wettstein	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
1 Mar 1917	<i>Scotoecus woodi</i> Thomas	<i>Scotoecus albofuscus</i> (Thomas, 1890)
14 May 1917	<i>Dichromyotis</i> Bianchi	<i>Myotis</i> Kaup, 1829
14 May 1917	<i>Paramyotis</i> Bianchi	<i>Myotis</i> Kaup, 1829

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14 May 1917	<i>Rickettia</i> Bianchi	<i>Myotis</i> Kaup, 1829
16 May 1917	<i>Cloeotis percivali australis</i> Roberts	<i>Cloeotis percivali</i> Thomas, 1901
21 May 1917	<i>Otomops icarus</i> Chubb	<i>Otomops martiensseni icarus</i> Chubb, 1917
14 Jun 1917	<i>Megapipistrellus</i> Bianchi	<i>Myotis</i> Kaup, 1829
28 Jun 1917	<i>Platymops</i> Roberts	<i>Sauromys</i> Roberts, 1917
28 Jun 1917	<i>Platymops (Sauromys)</i> Roberts	<i>Sauromys</i> Roberts, 1917
28 Jun 1917	<i>Platymops haagneri</i> Roberts	<i>Sauromys petrophilus haagneri</i> (Roberts, 1917)
28 Jun 1917	<i>Platymops petrophilus</i> Roberts	<i>Sauromys petrophilus</i> (Roberts, 1917)
29 Sep 1917	<i>Rhinolophus axillaris</i> Allen	<i>Rhinolophus landeri</i> Martin, 1838
29 Sep 1917	<i>Allomops</i> J.A. Allen	<i>Mops (Mops)</i> Lesson, 1842
29 Sep 1917	<i>Chærephon (Lophomops)</i> J.A. Allen	<i>Mops (Chaerephon)</i> Dobson, 1874
29 Sep 1917	<i>Chærephon (Lophomops) abæ</i> J.A. Allen	<i>Mops (Chaerephon) major</i> (Trouessart, 1897)
29 Sep 1917	<i>Chærephon (Lophomops) chapini</i> J.A. Allen	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
29 Sep 1917	<i>Chærephon (Lophomops) cristatus</i> J.A. Allen	<i>Mops (Chaerephon) leucogaster</i> (A. Grandidier, 1869)
29 Sep 1917	<i>Chærephon frater</i> J.A. Allen	<i>Mops (Chaerephon) leucogaster</i> (A. Grandidier, 1869)
29 Sep 1917	<i>Chærephon russatus</i> J.A. Allen	<i>Mops (Chaerephon) russatus</i> (J.A. Allen, 1917)
29 Sep 1917	<i>Eptesicus ater</i> J.A. Allen	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
29 Sep 1917	<i>Eptesicus faradjus</i> J.A. Allen	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
29 Sep 1917	<i>Eptesicus garambæ</i> J.A. Allen	<i>Laephotis capensis</i> (A. Smith, 1829)
29 Sep 1917	<i>Glauconycteris alboguttatus</i> J.A. Allen	<i>Glauconycteris alboguttata</i> J.A. Allen, 1917
29 Sep 1917	<i>Glauconycteris humeralis</i> J.A. Allen	<i>Glauconycteris humeralis</i> J.A. Allen, 1917
29 Sep 1917	<i>Hipposideros abæ</i> J.A. Allen	<i>Hipposideros abae</i> J.A. Allen, 1917
29 Sep 1917	<i>Hipposideros caffer niapu</i> J.A. Allen	<i>Hipposideros ruber</i> (Noack, 1893)
29 Sep 1917	<i>Hipposideros gigas niangaræ</i> J.A. Allen	<i>Macronycteris gigas</i> (Wagner, 1845)
29 Sep 1917	<i>Hipposideros langi</i> J.A. Allen	<i>Doryrhina cyclops</i> (Temminck, 1853)
29 Sep 1917	<i>Hipposideros nanus</i> J.A. Allen	<i>Hipposideros caffer</i> (Sundevall, 1846)
29 Sep 1917	<i>Miniopterus breyeri vicinior</i> J.A. Allen	<i>Miniopterus fraterculus</i> Thomas and Schwann, 1906
29 Sep 1917	<i>Mops (Allomops) faradjus</i> J.A. Allen	<i>Mops (Mops) demonstrator</i> (Thomas, 1903)
29 Sep 1917	<i>Mops (Allomops) nanulus</i> J.A. Allen	<i>Mops (Xiphonycteris) nanulus</i> J.A. Allen, 1917
29 Sep 1917	<i>Mops (Allomops) occipitalis</i> J.A. Allen	<i>Mops (Xiphonycteris) thersites</i> (Thomas, 1903)
29 Sep 1917	<i>Mops (Allomops) osborni</i> J.A. Allen	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
29 Sep 1917	<i>Mops conigicus</i> J.A. Allen	<i>Mops (Mops) conigicus</i> J.A. Allen, 1917
29 Sep 1917	<i>Mops niangaræ</i> J.A. Allen	<i>Mops (Mops) niangaræ</i> J.A. Allen, 1917
29 Sep 1917	<i>Mops trevori</i> J.A. Allen	<i>Mops (Mops) trevori</i> J.A. Allen, 1917
29 Sep 1917	<i>Nycteris avakubia</i> J.A. Allen	<i>Nycteris major</i> (K. Andersen, 1912)
29 Sep 1917	<i>Nycteris pallida</i> J.A. Allen	<i>Nycteris hispida</i> (Schreber, 1774)
29 Sep 1917	<i>Nyctinomus ochraceus</i> J.A. Allen	<i>Mops (Xiphonycteris) brachypterus</i> (Peters, 1852)
29 Sep 1917	<i>Pipistrellus abaensis</i> J.A. Allen	<i>Afronycteris nana</i> (Peters, 1852)
29 Sep 1917	<i>Rhinolophus abæ</i> J.A. Allen	<i>Rhinolophus fumigatus</i> Rüppell, 1842
31 Dec 1917	<i>Nycteris major</i> J.A. Allen	<i>Nycteris macrotis</i> Dobson, 1876
31 Dec 1917	<i>Pareptesicus</i> Bianchi	<i>Eptesicus Rafinesque</i> , 1820
31 Dec 1917	<i>Rhyneptesicus</i> Bianchi	<i>Eptesicus Rafinesque</i> , 1820
31 Dec 1917	<i>Palaeophyllophora</i> Revilliod	<i>Palaeophyllophora</i> Revilliod, 1917
27 Feb 1918	<i>Chaerephon leucostigma</i> G.M. Allen	<i>Mops (Mops) leucostigma</i> (G.M. Allen, 1918)
1 Oct 1918	<i>[Rhinolophus hipposideros] escalæræ</i> K. Andersen	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1 Oct 1918	<i>Asellia tridens diluta</i> K. Andersen	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
31 Dec 1919	<i>Cistugo lesueuri</i> Roberts	<i>Cistugo lesueuri</i> Roberts, 1919
31 Dec 1919	<i>Eptesicus melckorum</i> Roberts	<i>Laephotis capensis</i> (A. Smith, 1829)
1 Jan 1920	<i>Taphozous hamiltoni</i> Thomas	<i>Taphozous hamiltoni</i> Thomas, 1920
1 Aug 1920	<i>Kerivoula lucia</i> Hinton	<i>Kerivoula lanosa</i> (A. Smith, 1847)
1 Aug 1920	<i>Mimetillus thomasi</i> Hinton	<i>Mimetillus thomasi</i> Hinton, 1920
31 Dec 1920	<i>Rousettus (Lissonycteris) crypticola</i> Cabrera	<i>Myonycteris angolensis</i> (Bocage, 1898)
1 Apr 1921	<i>Rousettus</i> Gray	<i>Rousettus</i> Gray, 1821
31 Dec 1921	<i>Hipposideros curtus</i> G.M. Allen	<i>Hipposideros curtus</i> G.M. Allen, 1921
28 Feb 1922	<i>Rhinolophus hildebrandti perauritus</i> de Beaux	<i>Rhinolophus eloquens</i> K. Andersen, 1905
1 Mar 1922	<i>Liponycteris</i> Thomas	<i>Taphozous</i> E. Geoffroy St.-Hilaire, 1818
1 Jul 1922	<i>Kerivoula lueia</i> Kershaw	<i>Kerivoula lanosa</i> (A. Smith, 1847)
1 Aug 1922	<i>Nycteris oriana</i> Kershaw	<i>Nycteris macrotis</i> Dobson, 1876
6 Jul 1923	<i>Pipistrellus marrensis</i> Thomas and Hinton	<i>Pipistrellus (Pipistrellus) rusticus</i> (Tomes, 1861)
31 Jul 1923	<i>Petalia (Nycteris) thebaica aurantiaca</i> de Beaux	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
31 Jul 1923	<i>Scotoecus artinii</i> de Beaux	<i>Scotoecus hirundo</i> (de Winton, 1899)
1 Oct 1923	<i>Nycteris marica</i> Kershaw	<i>Nycteris grandis</i> Peters, 1865
31 Jan 1924	<i>Eptesicus zuluensis</i> Roberts	<i>Neoromicia zuluensis</i> (Roberts, 1924)
31 Jan 1924	<i>Kerivoula nidicola zuluensis</i> Roberts	<i>Kerivoula argentata</i> Tomes, 1861
31 Jan 1924	<i>Rhinolophus darlingi barbertonensis</i> Roberts	<i>Rhinolophus darlingi</i> K. Andersen, 1905
28 Feb 1924	<i>Petalia parisii</i> de Beaux	<i>Nycteris parisii</i> (de Beaux, 1924)
31 Dec 1924	<i>Otonycteris jin</i> Cheesman and Hinton	<i>Otonycteris hemprichii</i> Peters, 1859

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31 Dec 1924	<i>Hipposideros caffer aurantiaca</i> de Beaux	<i>Hipposideros caffer</i> (Sundevall, 1846)
31 Dec 1924	<i>Eptesicus loveni</i> Granvik	<i>Myotis tricolor</i> (Temminck, 1832)
14 May 1925	<i>Nycteris proxima</i> Lönnberg and Gyldestolpe	<i>Nycteris grandis</i> Peters, 1865
29 Apr 1926	<i>Chaerephon (Lophomops) shortridgei</i> Thomas	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
29 Apr 1926	<i>Pipistrellus fouriei</i> Thomas	<i>Afronycteris nana</i> (Peters, 1852)
1 May 1926	<i>Mops angolensis niveiventer</i> Cabrera and Ruxton	<i>Mops (Mops) niveiventer</i> Cabrera and Ruxton, 1926
14 Sep 1926	<i>Chaerephon pumilus elphicki</i> Roberts	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
14 Sep 1926	<i>Eptesicops</i> Roberts	<i>Pipistrellus</i> Kaup, 1829
14 Sep 1926	<i>Neoromicia</i> Roberts	<i>Neoromicia</i> Roberts, 1926
1 May 1927	<i>Myotis scotti</i> Thomas	<i>Myotis scotti</i> Thomas, 1927
12 Jul 1927	<i>Miniopterus smitanius</i> Thomas	<i>Miniopterus natalensis</i> (A. Smith, 1833)
10 Dec 1928	<i>Chærophon (Lophomops) nigri</i> Hatt	<i>Mops (Chaerephon) leucogaster</i> (A. Grandidier, 1869)
31 Dec 1928	<i>Emballonuroidea</i> Weber	<i>EMBALLONURIDAE</i> Gervais, 1855
26 Dec 1929	<i>Anamygdon</i> Troughton	<i>Myotis</i> Kaup, 1829
31 Dec 1929	<i>Rousettus madagascariensis</i> G. Grandidier	<i>Rousettus madagascariensis</i> G. Grandidier, 1929
6 Jun 1931	<i>Asellia patrizii</i> de Beaux	<i>Asellia patrizii</i> de Beaux, 1931
6 Jun 1931	<i>Asellia tridens italo-somalica</i> de Beaux	<i>Asellia italosomalica</i> de Beaux, 1931
31 Dec 1931	<i>Tuitatus</i> Kishida and Mori	<i>Eptesicus</i> Rafinesque, 1820
1 Oct 1932	<i>Choerephon (Lophomops) langi</i> Roberts	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1 Oct 1932	<i>Eptesicus capensis nkatiensis</i> Roberts	<i>Laephotis capensis</i> (A. Smith, 1829)
1 Oct 1932	<i>Neoromicia vansonii</i> Roberts	<i>Neoromicia zuluensis</i> (Roberts, 1924)
1 Oct 1932	<i>Pipistrellus vernayi</i> Roberts	<i>Vansonia rueppellii vernayi</i> (Roberts, 1932)
1 Oct 1932	<i>Scoteinus schlieffenii fitzsimonsi</i> Roberts	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
31 Dec 1933	<i>Mimetillus bernerii</i> Monard	<i>Mimetillus thomasi</i> Hinton, 1920
31 Dec 1933	<i>Nyctinomus spillmanni</i> Monard	<i>Mops (Chaerephon) nigeriae spillmanni</i> (Monard, 1933)
31 Dec 1933	<i>Pipistrellus leucomelas</i> Monard	<i>Vansonia rueppellii vernayi</i> (Roberts, 1932)
30 Nov 1934	<i>Rhinolophus acrotis schwarzi</i> Heim de Balsac	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
31 Dec 1934	<i>Austronomus</i> Iredale and Troughton	<i>Tadarida</i> Rafinesque, 1814
31 Dec 1935	<i>Laephotis angolensis</i> Monard	<i>Laephotis angolensis</i> Monard, 1935
30 Jan 1936	<i>Nycteris nana tristis</i> G.M. Allen and Lawrence	<i>Nycteris nana</i> (K. Andersen, 1912)
15 Aug 1936	<i>Miniopterus africanus</i> Sanborn	<i>Miniopterus africanus</i> Sanborn, 1936
15 Aug 1936	<i>Miniopterus rufus</i> Sanborn	<i>Miniopterus inflatus rufus</i> Sanborn, 1936
15 Aug 1936	<i>Mops angolensis wonderi</i> Sanborn	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
15 Aug 1936	<i>Rhinolophus alcyone alticolus</i> Sanborn	<i>Rhinolophus simulator</i> K. Andersen, 1904
31 Dec 1936	<i>Pipistrellus k[uhlii] pallidus</i> Heim de Balsac	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
31 Dec 1936	<i>Plecotus auritus saharæ</i> Laurent	<i>Otonycteris hemprichii</i> Peters, 1859
28 Feb 1937	<i>Rhinolophus hipposideros vespa</i> Laurent	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
17 Apr 1937	<i>Eptesicus capensis angolensis</i> J.Eric Hill	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
17 Apr 1937	<i>Mops chitauensis</i> J.Eric Hill	<i>Mops (Mops) niveiventer</i> Cabrera and Ruxton, 1926
1 May 1937	<i>Rhinopoma cystops macinnesi</i> Hayman	<i>Rhinopoma macinnesi</i> Hayman, 1937
30 Sep 1937	<i>Hipposideros sandersoni</i> Sanderson	<i>Hipposideros curtus</i> G.M. Allen, 1921
30 Nov 1937	<i>Nycteris madagascariensis</i> G. Grandidier	<i>Nycteris macrotis</i> Dobson, 1876
31 Dec 1937	<i>Pipistrellus K[uhlii] latastei</i> Laurent	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
31 Dec 1937	<i>Asellia tridens pallida</i> Laurent	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
1 Apr 1938	<i>Chaerephon lancasteri</i> Hayman	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
31 Jul 1938	<i>Platymops haagneri umbratus</i> Shortridge and Carter	<i>Sauromys petrophilus umbratus</i> (Shortridge & Carter, 1938)
31 Jul 1938	<i>Scotophilus angusticeps</i> Shortridge and Carter	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
1 Feb 1939	<i>Glauconycteris superba</i> Hayman	<i>Glauconycteris superba</i> Hayman, 1939
31 Mar 1939	<i>Coleura kummeri</i> Monard	<i>Coleura afra</i> (Peters, 1852)
31 Mar 1939	<i>Hipposideros braima</i> Monard	<i>Hipposideros tephurus</i> (Cabrera, 1906)
31 Mar 1939	<i>Hipposideros gigas viegasi</i> Monard	<i>Macronycteris gigas</i> (Wagner, 1845)
31 Mar 1939	<i>Mops osborni occidentalis</i> Monard	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
31 Mar 1939	<i>Mops osborni occidentalis f. fulva</i> Monard	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
31 Mar 1939	<i>Nycteris æthiopica guineensis</i> Monard	<i>Nycteris macrotis</i> Dobson, 1876
31 Mar 1939	<i>Nycteris æthiopica guineensis f. aurantiaca</i> Monard	<i>Nycteris macrotis</i> Dobson, 1876
19 Sep 1939	<i>Rhinolophus aethiops diversus</i> Sanborn	<i>Rhinolophus fumigatus</i> Rüppell, 1842
31 Dec 1939	<i>Mops rüppellii</i> Allen	<i>Tadarida ventralis</i> (Heuglin, 1861)
31 Jan 1940	<i>Mops calabarensis</i> Hayman	<i>Mops (Xiphonycteris) nanulus</i> J.A. Allen, 1917
20 Aug 1941	<i>Coelopinae</i> Tate	<i>HIPPOSIDERIDAE</i> Lydekker, 1891
31 Dec 1941	<i>[Triaenops] furinea</i> Tate	<i>Paratriaenops pauliani</i> (Goodman and Ranivo, 2008)
31 Dec 1941	<i>Austronomus</i> Troughton	<i>Tadarida</i> Rafinesque, 1814
31 Dec 1941	<i>Rhinophyllotis</i> Troughton	<i>Rhinolophus</i> Lacépède, 1799
28 Feb 1942	<i>Mops angolensis orientis</i> G.M. Allen and Loveridge	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)

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28 Jul 1942	<i>Rhinolophus ruwenzorii</i> J. Eric Hill	<i>Rhinolophus ruwenzorii</i> J. Eric Hill, 1942
27 Nov 1942	<i>Myotinae</i> Tate	<i>Myotinae</i> Tate, 1942
27 Nov 1942	<i>Myotini</i> Tate	<i>VESPERTILIONIDAE</i> Gray, 1821
27 Nov 1942	<i>Myotini</i> Tate	<i>Myotinae</i> Tate, 1942
27 Nov 1942	<i>Nycticeini</i> Tate	<i>Vespertilioninae</i> Gray, 1821
27 Nov 1942	<i>Pipistrellini</i> Tate	<i>Pipistrellini</i> Tate, 1942
31 Dec 1942	<i>Eptesicus megalurus pallidior</i> Shortridge	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
31 Dec 1943	<i>Scotonycteris ophiodon</i> Pohle	<i>Casinonycteris ophiodon</i> (Pohle, 1943)
19 Sep 1946	<i>Scotonycteris ophiodon cansdalei</i> Hayman	<i>Casinonycteris ophiodon</i> (Pohle, 1943)
31 Dec 1946	<i>Eptesicus hottentotus bensoni</i> Roberts	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
31 Dec 1946	<i>Nycteris woodi sabiensis</i> Roberts	<i>Nycteris woodi</i> K. Andersen, 1914
31 Dec 1946	<i>Nyctinomus masoni</i> Roberts	<i>Tadarida fulminans</i> (Thomas, 1903)
31 Dec 1946	<i>Nyctinomus rhodesiae</i> Roberts	<i>Mops (Chaerephon) ansorgei</i> (Thomas, 1913)
31 Dec 1946	<i>Platymops petrophilus erongensis</i> Roberts	<i>Sauromys petrophilus erongensis</i> (Roberts, 1946)
31 Dec 1946	<i>Platymops petrophilus fitzsimonsi</i> Roberts	<i>Sauromys petrophilus umbratus</i> (Shortridge & Carter, 1938)
31 Dec 1946	<i>Rhinolophus darlingi damarensis</i> Roberts	<i>Rhinolophus damarensis</i> Roberts, 1946
31 Dec 1946	<i>Rhinolophus swinnyi rhodesiae</i> Roberts	<i>Rhinolophus rhodesiae</i> Roberts, 1946
31 Dec 1946	<i>Scotophilus nigrita pondoensis</i> Roberts	<i>Scotophilus dinganii</i> (A. Smith, 1833)
31 Dec 1946	<i>Vansonia</i> Roberts	<i>Vansonia</i> Roberts, 1946
31 Dec 1946	<i>Vansonia</i> Roberts	<i>Pipistrellus</i> Kaup, 1829
25 Jun 1947	<i>Scotonycteris zenkeri occidentalis</i> Hayman	<i>Scotonycteris occidentalis</i> Hayman, 1947
7 Aug 1947	<i>Glauconycteris superba sheila</i> Hayman	<i>Glauconycteris superba</i> Hayman, 1939
19 Nov 1947	<i>Hipposideros jonesi</i> Hayman	<i>Hipposideros jonesi</i> Hayman, 1947
31 Dec 1948	<i>Pipistrellus (Romicia) kuhlii broomi</i> Roberts	<i>Pipistrellus (Pipistrellus) hesperidus broomi</i> Roberts, 1948
29 Nov 1950	<i>Micropteropus grandis</i> Sanborn	<i>Epomophorus grandis</i> (Sanborn, 1950)
31 Dec 1950	<i>Epomophorus reii</i> Aellen	<i>Epomophorus labiatus</i> (Temminck, 1837)
31 Dec 1950	<i>Epomophorus reii</i> Aellen	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
22 Dec 1951	<i>Tadarida (Chaerephon) faini</i> Hayman	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
31 Dec 1951	<i>Rhinomegalophus</i> Bourret	<i>Rhinolophus</i> Lacépède, 1799
31 Dec 1952	<i>Nycteris benuensis</i> Aellen	<i>Nycteris parisii</i> (de Beaux, 1924)
31 Dec 1953	<i>Otomops madagascariensis</i> Dorst	<i>Otomops madagascariensis</i> Dorst, 1953
31 Dec 1954	<i>Hipposideros marisae</i> Aellen	<i>Hipposideros marisae</i> Aellen, 1954
31 Dec 1954	<i>Vespertilio brevicauda</i> Stresemann	<i>Rhinopoma cystops</i> Thomas, 1903
31 Dec 1954	<i>Vespertilio ferox</i> Stresemann	<i>Rhinopoma cystops</i> Thomas, 1903
31 Dec 1955	<i>Rhinolophus euryale tuneti</i> Deleuil and Labbé	<i>Rhinolophus mehelyi</i> Matschie, 1901
18 Dec 1956	<i>Platymops barbatogularis</i> Harrison	<i>Platymops setiger macmillani</i> Thomas, 1906
27 Dec 1956	<i>Hipposideros camerunensis</i> Eisentraut	<i>Doryrhina camerunensis</i> (Eisentraut, 1956)
31 Dec 1956	<i>Miniopterus inflatus villiersi</i> Aellen	<i>Miniopterus villiersi</i> Aellen, 1956
31 Dec 1957	<i>Pipistrellus (Pipistrellus) permixtus</i> Aellen	<i>Pipistrellus (Pipistrellus) permixtus</i> Aellen, 1957
31 Dec 1957	<i>Saccolaimus incognita</i> Butler and Hopwood	<i>Taphozous incognita</i> (Butler & Hopwood, 1957)
31 Dec 1957	<i>Hipposideros beatus maximus</i> Verschuren	<i>Hipposideros beatus</i> (K. Andersen, 1906)
15 Dec 1958	<i>Taphozous perforatus swirae</i> Harrison	<i>Taphozous perforatus perforatus</i> E. Geoffroy St.-Hilaire, 1818
31 Dec 1958	<i>Hesperomyotis</i> Cabrera	<i>Myotis</i> Kaup, 1829
30 Apr 1959	<i>Miniopterus minor griveaudi</i> Harrison	<i>Miniopterus griveaudi</i> Harrison, 1959
31 Dec 1959	<i>Kerivoula harrisoni bellula</i> Aellen	<i>Kerivoula lanosa</i> (A. Smith, 1847)
31 Dec 1959	<i>Nycteris intermedia</i> Aellen	<i>Nycteris intermedia</i> Aellen, 1959
31 Dec 1959	<i>Pipistrellus inexpectatus</i> Aellen	<i>Pipistrellus (Pipistrellus) inexpectatus</i> Aellen, 1959
31 Dec 1959	<i>Rhinolophus silvestris</i> Aellen	<i>Rhinolophus silvestris</i> Aellen, 1959
31 Dec 1959	<i>Nyctiophis</i> Hall and Kelson	<i>Eptesicus Rafinesque</i> , 1820
10 Jan 1960	<i>Rousettus aegyptiacus occidentalis</i> Eisentraut	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
30 Apr 1960	<i>Platymops barbatogularis parkeri</i> Harrison and Fleetwood	<i>Platymops setiger macmillani</i> Thomas, 1906
1 Jun 1960	<i>Rhinolophus denti</i> Knorri Eisentraut	<i>Rhinolophus denti</i> Thomas, 1904
1 Jun 1960	<i>Rhinolophus landeri guineensis</i> Eisentraut	<i>Rhinolophus guineensis</i> Eisentraut, 1960
31 Dec 1960	<i>Epomophorus gambianus parvus</i> Ansell	<i>Epomophorus crypturus</i> Peters, 1852
31 Dec 1960	<i>Myotis Goudoti anjouanensis</i> Dorst	<i>Myotis anjouanensis</i> Dorst, 1960
31 Dec 1960	<i>Pipistrellus Rüppelli senegalensis</i> Dorst	<i>Vansonia rueppellii senegalensis</i> (Dorst, 1960)
31 Dec 1961	<i>Afropteris Lavocat</i>	<i>Megaderma</i> E. Geoffroy St.-Hilaire, 1810
31 Dec 1961	<i>Afropteris gigas</i> Lavocat	<i>Megaderma gigas</i> (Lavocat, 1961)
31 Dec 1961	<i>Afropteris gigas</i> Lavocat	<i>Megaderma gaillardi</i> (Trouessart, 1898)
31 Dec 1961	<i>Asellia vetus</i> Lavocat	<i>Hipposideros vetus</i> (Lavocat, 1961)
31 Dec 1961	<i>Rhinolophus ferrumequinum mellali</i> Lavocat	<i>Rhinolophus mellali</i> Lavocat, 1961
31 Dec 1962	<i>Taphozous sudani australis</i> Harrison	<i>Taphozous perforatus sudani</i> Thomas, 1915
18 Apr 1963	<i>Myonycterini</i> Lawrence and Novick	<i>Myonycterini</i> Lawrence and Novick, 1963
31 Dec 1963	<i>Glauconycteris machadoi</i> Hayman	<i>Glauconycteris machadoi</i> Hayman, 1963
31 Dec 1963	<i>Micropteropus intermedius</i> Hayman	<i>Epomophorus intermedius</i> (Hayman, 1963)

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31 Dec 1964	<i>Taphozous sudani rhodesiae</i> Harrison	<i>Taphozous perforatus sudani</i> Thomas, 1915
31 May 1965	<i>Nycteris vinsoni</i> Dalquest	<i>Nycteris vinsoni</i> Dalquest, 1965
31 May 1965	<i>Scotophilus alvenslebeni</i> Dalquest	<i>Scotophilus nigrata</i> (Schreber, 1774)
31 Dec 1965	<i>Rousettus angolensis ruwenzorii</i> Eisentraut	<i>Myonycteris angolensis ruwenzorii</i> (Eisentraut, 1965)
31 Dec 1966	<i>Tadarida (Chaerephon) cyclotis</i> Brosset	<i>Mops (Chaerephon) aloysiisabaudiae</i> (Festa, 1907)
31 Dec 1966	<i>Megaloglossus woermanni prigoginei</i> Hayman	<i>Megaloglossus woermanni</i> Pagenstecher, 1885
31 Dec 1967	<i>Propotto</i> Simpson	<i>Propotto</i> Simpson, 1967
31 Dec 1967	<i>Propotto leakeyi</i> Simpson	<i>Propotto leakeyi</i> Simpson, 1967
10 Apr 1968	<i>Pipistrellus eisentrauti</i> Hill	<i>Nycticeinops eisentrauti</i> (Hill, 1968)
31 Dec 1968	<i>Rhinolophus adami</i> Aellen and Brosset	<i>Rhinolophus adami</i> Aellen and Brosset, 1968
31 Dec 1968	<i>Trienops persicus majusculus</i> Aellen and Brosset	<i>Trienops afer</i> Peters, 1877
31 Dec 1968	<i>Brachipposideros</i> Sigé	<i>Brachipposideros</i> Sigé, 1968
31 Dec 1969	<i>Rhinopoma hardwickei sennaariense</i> Kock	<i>Rhinopoma cystops</i> Thomas, 1903
31 Dec 1969	<i>Rhinopoma microphyllum tropicalis</i> Kock	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
31 Dec 1970	<i>Cynopterini</i> Koopman and J.K. Jones Jr.	<i>Pteropodinae</i> Gray, 1821
31 Dec 1970	<i>Hipposiderini</i> Koopman and J.K. Jones Jr.	<i>HIPPOSIDERIDAE</i> Lydekker, 1891
31 Dec 1970	<i>Rousettina</i> Koopman and J.K. Jones Jr.	<i>Rousettinae</i> Andersen, 1912
28 Apr 1971	<i>Myotis morrisi</i> Hill	<i>Myotis morrisi</i> Hill, 1971
4 Nov 1971	<i>Laephotis botswanae</i> Setzer	<i>Laephotis botswanae</i> Setzer, 1971
4 Nov 1971	<i>Laephotis namibensis</i> Setzer	<i>Laephotis namibensis</i> Setzer, 1971
31 Dec 1971	<i>Epomophorus</i> sp. Hill and Morris	<i>Epomophorus labiatus</i> (Temminck, 1837)
30 Jun 1972	<i>Pipistrellus eisentrauti bellieri</i> De Vree	<i>Nycticeinops bellieri</i> (De Vree, 1972)
17 Aug 1973	<i>Glauconycteris gleni</i> Peterson and Smith	<i>Glauconycteris gleni</i> Peterson and Smith, 1973
31 Dec 1973	<i>Rhinolophus hilli</i> Aellen	<i>Rhinolophus hilli</i> Aellen, 1973
31 Dec 1975	<i>Tadarida (Chaerephon) gallagheri</i> Harrison	<i>Mops (Chaerephon) gallagheri</i> (Harrison, 1975)
31 Dec 1976	<i>Megaderma jaegeri</i> Sigé	<i>Megaderma jaegeri</i> Sigé, 1976
22 Aug 1977	<i>Myotis blythii punicus</i> Felten	<i>Myotis punicus</i> Felten, 1977
31 Dec 1978	<i>Rousettus (Rousettus) obliviosus</i> Kock	<i>Rousettus obliviosus</i> Kock, 1978
31 Jul 1979	<i>PHYLLOSTOMATIA</i> Van Valen	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Jul 1979	<i>VESPERTILIONIA</i> Van Valen	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
4 Mar 1980	<i>Myonycteris relicta</i> Bergmans	<i>Myonycteris relicta</i> Bergmans, 1980
31 Dec 1980	<i>Plecotus austriacus kolombatovici</i> Dulic	<i>Plecotus kolombatovici</i> Dulic, 1980
11 Dec 1981	<i>Tadarida [(Xiphonycteris)] petersoni</i> El Rayah	<i>Mops (Xiphonycteris) petersoni</i> (El Rayah, 1981)
31 Dec 1982	<i>Glauconycteris kenyacola</i> Peterson	<i>Glauconycteris kenyacola</i> Peterson, 1982
31 Dec 1983	<i>Tadarida engesseri</i> Rachl	<i>Tadarida engesseri</i> Rachl, 1983
31 Dec 1983	<i>Scotophilus nucella</i> Robbins	<i>Scotophilus nucella</i> Robbins, 1983
30 Jun 1984	<i>Cheiramelinae</i> Legendre	<i>Molossinae</i> Gervais, 1856
30 Jun 1984	<i>Rhizomops</i> Legendre	<i>Tadarida Rafinesque</i> , 1814
15 Nov 1984	<i>Chamtvaria</i> Butler	<i>Chamtvaria</i> Butler, 1984
15 Nov 1984	<i>Chamtvaria pickfordi</i> Butler	<i>Chamtvaria pickfordi</i> Butler, 1984
15 Nov 1984	<i>Propottinae</i> Butler	<i>Propottinae</i> Butler, 1984
31 Dec 1984	<i>Coleura muthokai</i> Wesselman	<i>Coleura muthokai</i> Wesselman, 1984
31 Dec 1984	<i>Hipposideros kaumbului</i> Wesselman	<i>Hipposideros kaumbului</i> Wesselman, 1984
31 Dec 1984	<i>Taphozous abitus</i> Wesselman	<i>Saccolaimus abitus</i> (Wesselman, 1984)
31 Dec 1985	<i>Hipposideros lamottei</i> Brosset	<i>Hipposideros lamottei</i> Brosset, 1985
31 Dec 1985	<i>YANGOCHIROPTERA</i> Koopman	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 1985	<i>Yinochiroptera</i> Koopman	<i>CHIROPTERA</i> Blumenbach, 1779
31 Dec 1985	<i>rueppellii</i> Qumsiyeh	<i>Tadarida teniotis</i> (Rafinesque, 1814)
31 Dec 1985	<i>Philisidae</i> Sigé	<i>PHILISIDAE</i> Sigé, 1985
31 Dec 1985	<i>Philisis</i> Sigé	<i>Philisis</i> Sigé, 1985
31 Dec 1985	<i>Philisis sphingis</i> Sigé	<i>Philisis sphingis</i> Sigé, 1985
14 Feb 1986	<i>Eptesicus hottentotus portavernus</i> Schlitter and Aggundey	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
31 Mar 1987	<i>Eptésiformes</i> Menu	<i>Vespertilioninae</i> Gray, 1821
31 Mar 1987	<i>Leuconoformes</i> Menu	<i>Myotinae</i> Tate, 1942
31 Mar 1987	<i>Leuconoïdes</i> Menu	<i>Vespertilioninae</i> Gray, 1821
31 Mar 1987	<i>Néoeptésiformes</i> Menu	<i>Vespertilioninae</i> Gray, 1821
31 Mar 1987	<i>Nyctaliformes</i> Menu	<i>Vespertilioninae</i> Gray, 1821
31 Mar 1987	<i>Nyctaloïdes</i> Menu	<i>Vespertilioninae</i> Gray, 1821
30 Jul 1987	<i>Scotophilini</i> Hill and Harrison	<i>Scotophilinae</i> Van Cakenberghe and Seamark, 2008
15 Nov 1987	<i>Attalepharca</i> Menu	<i>Pipistrellus</i> Kaup, 1829
15 Nov 1987	<i>Nycterikaupius</i> Menu	<i>Pseudoromicia</i> Monadjem, Patterson, Webala and Demos, 2020
15 Nov 1987	<i>Nycterikaupius</i> Menu	<i>Pipistrellus</i> Kaup, 1829
15 Nov 1987	<i>Nycterikaupius</i> Menu	<i>Neoromicia</i> Roberts, 1926
15 Nov 1987	<i>Nycterikaupius</i> Menu	<i>Laephotis</i> Thomas, 1901

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31 Dec 1987	<i>Nycticeinops</i> Hill and Harrison	<i>Nycticeinops</i> Hill & Harrison, 1987
25 Feb 1988	<i>Taphozoini</i> Robbins and Sarich	<i>Taphozoinae</i> Jerdon, 1867
28 Jun 1989	<i>Rhinolophus clivosus hillorum</i> Koopman	<i>Rhinolophus hillorum</i> Koopman, 1989
31 Dec 1991	<i>Epomophorus minimus</i> Claessen and De Vree	<i>Epomophorus minimus</i> Claessen & De Vree, 1991
31 Dec 1991	<i>Dizzya</i> Sigé	<i>Dizzya</i> Sigé, 1991
31 Dec 1991	<i>Dizzya exsultans</i> Sigé	<i>Dizzya exsultans</i> Sigé, 1991
21 Oct 1992	<i>Miniopterus minor occidentalis</i> Juste and Ibáñez	<i>Miniopterus minor</i> Peters, 1867
28 Feb 1993	<i>Rousettus aegyptiacus princeps</i> Juste and Ibáñez	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1 Jul 1993	<i>Rousettus aegyptiacus thomensis</i> Feiler, Haft and Widmann	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
30 Nov 1993	<i>Tadarida (Chaerephon) tomensis</i> Juste and Ibáñez	<i>Mops (Chaerephon) tomensis</i> (Juste and Ibáñez, 1993)
31 Dec 1993	<i>Rousettus aegyptiacus tomensis</i> Juste and Ibáñez	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
31 Dec 1994	<i>Chiropteriformes</i> Kinman	<i>CHIROPTERA</i> Blumenbach, 1779
31 Dec 1994	<i>Hipposiderina</i> Koopman	<i>HIPPOSIDERIDAE</i> Lydekker, 1891
31 Dec 1994	<i>Dhofarella</i> Sigé, Thomas, Sen, Gheerbrant, Roger and Al-Sulaimani	<i>Dhofarella</i> Sigé, Thomas, Sen, Gheerbrant, Roger and Al-Sulaimani, 1985
31 Dec 1994	<i>Dhofarella thaleri</i> Sigé, Thomas, Sen, Gheerbrant, Roger and Al-Sulaimani	<i>Dhofarella thaleri</i> Sigé et al., 1994
31 Dec 1995	<i>Eptesicus somalicus malagasyensis</i> Peterson, Eger and Mitchell	<i>Laephotis malagasyensis</i> (Peterson, Eger and Mitchell, 1995)
31 Dec 1995	<i>Miniopterus gleni</i> Peterson, Eger and Mitchell	<i>Miniopterus gleni</i> Peterson, Eger and Mitchell, 1995
31 Dec 1995	<i>VESPERTILIFORMES</i> Zagorodniuk, Godovanets, Pokynchereda and Kyseliuk	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
20 Jun 1997	<i>Lissonycteris angolensis goliath</i> Bergmans	<i>Myonycteris angolensis goliath</i> (Bergmans, 1997)
20 Jun 1997	<i>Lissonycteris angolensis petraea</i> Bergmans	<i>Myonycteris angolensis petraea</i> (Bergmans, 1997)
20 Jun 1997	<i>Plerotini</i> Bergmans	<i>Plerotini</i> Bergmans, 1997
30 Jun 1997	<i>Scotonycterini</i> Bergmans	<i>Scotonycterini</i> Bergmans, 1997
31 Dec 1997	<i>VESPERTILIFORMES</i> Zagorodniuk	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 1998	<i>Megachiropteramorpha</i> Simmons and Geisler	<i>RHINOLOPHIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 1998	<i>Microchiropteraformes</i> Simmons and Geisler	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 1998	<i>Microchiropteramorpha</i> Simmons and Geisler	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 1998	<i>VESPERTILIONIFORMES</i> Zagorodnyuk	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
22 Dec 2000	<i>Rhinolophus maendeleo</i> Kock, Csorba and Howell	<i>Rhinolophus maendeleo</i> Kock, Csorba and Howell, 2000
31 Dec 2000	<i>Myotis cf. punicus</i> Castella, Ruedi, Excoffier, Ibanez, Arlettaz and Hausser	<i>Myotis punicus</i> Felten, 1977
31 Dec 2000	<i>Eidolon helvum annobonensis</i> Juste, Ibáñez and Machordom	<i>Eidolon helvum helvum</i> (Kerr, 1792)
31 Dec 2000	<i>Plecotus balensis</i> Kruskop and Lavrenchenko	<i>Plecotus balensis</i> Kruskop & Lavrenchenko, 2000
31 Dec 2001	<i>Glauconycteris curryi</i> Eger and Schlitter	<i>Glauconycteris curryae</i> Eger and Schlitter, 2001
31 Dec 2001	<i>Pipistrellus africanus meesteri</i> Kock	<i>Afronycteris nana</i> (Peters, 1852)
31 Dec 2001	<i>YINPTEROCHIROPTERA</i> Springer, Teeling, Madsen, Stanhope and De Jong	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 2001	<i>YINPTEROCHIROPTERA</i> Springer, Teeling, Madsen, Stanhope and De Jong	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 2001	<i>Pipistrellus cf. kuhlii</i> Volleth, Bronner, Göpfert, Heller, von Helversen and Yong	<i>Pipistrellus</i> <i>*(Pipistrellus)*</i> sp. aff. <i>*kuhlii*</i> Kuhl, 1817
30 Sep 2002	<i>Barbastella barbastellus guanchae</i> Trujillo, Ibáñez and Juste	<i>Barbastella barbastellus</i> (Schreber, 1774)
31 Dec 2002	<i>Tadarida rusingae</i> Arroyo-Cabrales, Gregorin, Schlitter and Walker	<i>Mops rusingae</i> (Arroyo-Cabrales, Gregorin, Schlitter and Walker, 2002)
31 Dec 2002	<i>Rhinolophus sakejiensis</i> Cotterill	<i>Rhinolophus sakejiensis</i> Cotterill, 2002
31 Dec 2002	<i>Rhinolophus ziama</i> Fahr, Vierhaus, Hutterer and Kock	<i>Rhinolophus ziama</i> Fahr, Vierhaus, Hutterer and Kock, 2002
31 Dec 2003	<i>Indorhinolophus</i> Guillén-Servent, Francis and Ricklefs	<i>Rhinolophus</i> Lacépède, 1799
31 Dec 2003	<i>Tanzanycterididae</i> Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch	<i>TANZANYCTERIDIDAE</i> Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003

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31 Dec 2003	<i>Tanzanycteris</i> Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch	<i>Tanzanycteris</i> Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003
31 Dec 2003	<i>Tanzanycteris mannardi</i> Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch	<i>Tanzanycteris mannardi</i> Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003
31 Dec 2004	<i>Plecotus teneriffae gaisleri</i> Benda, Kiefer, Hanák and Veith	<i>Plecotus gaisleri</i> Benda, Kiefer, Hanak & Veith, 2004
31 Dec 2004	<i>Epomophorus anseli</i> Bergmans and Van Strien	<i>Epomophorus anseli</i> Bergmans and Van Strien, 2004
31 Dec 2004	<i>Chaerephon jobimena</i> Goodman and Cardiff	<i>Mops (Chaerephon) jobimena</i> (Goodman and Cardiff, 2004)
31 Dec 2004	<i>Pipistrellus hanaki</i> Hulva and Benda	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
31 Dec 2004	<i>PTEROPODIFORMES</i> Hutcheon and Kirsch	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 2004	<i>VESPERTILIONIFORMES</i> Hutcheon and Kirsch	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 2005	<i>Scotophilus tandrefana</i> Goodman, Jenkins and Ratrimomanarivo	<i>Scotophilus tandrefana</i> Goodman, Jenkins and Ratrimomanarivo, 2005
31 Dec 2005	<i>Brevipalatus</i> Hand and Archer	<i>Brevipalatus</i> Hand and Archer, 2005
31 Dec 2005	<i>Myotis dieteri</i> M. Happold	<i>Myotis dieteri</i> M. Happold, 2005
31 Dec 2005	<i>Emballonuroidea</i> Teeling, Springer, Madsen, Bates, O'Brien and Murphy	<i>NYCTEROIDEA</i> Van der Hoeven, 1855
19 Oct 2006	<i>Emballonura tiavato</i> Goodman, Cardiff, Ranivo, Russell and Yoder	<i>Paremballonura tiavato</i> (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)
31 Dec 2006	<i>Pipistrellus raceyi</i> Bates, Ratrimomanarivo, Harrison and Goodman	<i>Pipistrellus (Pipistrellus) raceyi</i> Bates, Ratrimomanarivo, Harrison and Goodman, 2006
31 Dec 2006	<i>Scotophilus marovaza</i> Goodman, Ratrimomanarivo and Randrianandrianina	<i>Scotophilus marovaza</i> Goodman, Ratrimomanarivo and Randrianandrianina, 2006
31 Dec 2006	<i>Philisinae</i> Horáček, Fejfar and Hulva	<i>Scotophilinae</i> Van Cakenberghe and Seamark, 2008
31 Dec 2006	<i>Scotophilis</i> Horáček, Fejfar and Hulva	<i>Scotophilis</i> Horáček, Fejfar and Hulva, 2006
31 Dec 2006	<i>Scotophilis libycus</i> Horáček, Fejfar and Hulva	<i>Scotophilis libycus</i> Horáček, Fejfar and Hulva, 2006
26 Mar 2007	<i>Myzopoda schliemanni</i> Goodman, Rakotondraparany and Kofoky	<i>Myzopoda schliemanni</i> Goodman, Rakotondraparany and Kofoky, 2007
31 Jul 2007	<i>NOCTILIONIFORMACEI</i> Van Cakenberghe, Kearney and Seamark	<i>NOCTILIONIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Jul 2007	<i>NYCTERIFORMACEI</i> Van Cakenberghe, Kearney and Seamark	<i>NYCTERIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Jul 2007	<i>PTEROPODIFORMACEI</i> Van Cakenberghe, Kearney and Seamark	<i>PTEROPODIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Jul 2007	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Jul 2007	<i>RHINOLOPHIFORMACEI</i> Van Cakenberghe, Kearney and Seamark	<i>RHINOLOPHIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Jul 2007	<i>VESPERTILIONIFORMACEI</i> Van Cakenberghe, Kearney and Seamark	<i>VESPERTILIONIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Jul 2007	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 2007	<i>Miniopterus sororculus</i> Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton	<i>Miniopterus sororculus</i> Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton, 2007
31 Dec 2007	<i>Hipposideros besaoka</i> Samonds	<i>Macronycteris besaoka</i> (Samonds, 2007)
31 Dec 2007	<i>Trienops goodmani</i> Samonds	<i>Trienops goodmani</i> Samonds, 2007
31 Dec 2007	<i>Afropipistrellus</i> Thorn, Kock and Cuisin	<i>Nycticeinops</i> Hill & Harrison, 1987
1 Jul 2008	<i>Scotophilinae</i> Van Cakenberghe and Seamark	<i>Scotophilinae</i> Van Cakenberghe and Seamark, 2008
31 Dec 2008	<i>Plecotus christii petraeus</i> Benda	<i>Plecotus christii</i> Gray, 1838
31 Dec 2008	<i>Trienops pauliani</i> Goodman and Ranivo	<i>Paratrienops pauliani</i> (Goodman and Ranivo, 2008)
31 Dec 2008	<i>Miniopterus petersoni</i> Goodman, Bradman, Maminirina, Ryan, Christidis and Appleton	<i>Miniopterus petersoni</i> Goodman, Bradman, Maminirina, Ryan, Christidis & Appleton, 2008
31 Dec 2008	<i>Mormopterus francoismoutoui</i> Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst and Bowie	<i>Mormopterus francoismoutoui</i> Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst, Bowie, 2008
31 Dec 2008	<i>Dhofarella sigei</i> Gunnell, Simons and Seiffert	<i>Dhofarella sigei</i> Gunnell, Simons and Seiffert, 2008
31 Dec 2008	<i>Khonsunycteris</i> Gunnell, Simons and Seiffert	<i>Khonsunycteris</i> Gunnell, Simons and Seiffert, 2008
31 Dec 2008	<i>Khonsunycteris aegypticus</i> Gunnell, Simons and Seiffert	<i>Khonsunycteris aegypticus</i> Gunnell, Simons and Seiffert, 2008
31 Dec 2008	<i>Qarunycteris</i> Gunnell, Simons and Seiffert	<i>Qarunycteris</i> Gunnell, Simons and Seiffert, 2008

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31 Dec 2008	<i>Qarunycteris moerisae</i> Gunnell, Simons and Seiffert	<i>Qarunycteris moerisae</i> Gunnell, Simons and Seiffert, 2008
31 Dec 2008	<i>Saharaderma</i> Gunnell, Simons and Seiffert	<i>Saharaderma</i> Gunnell, Simons and Seiffert, 2008
31 Dec 2008	<i>Saharaderma pseudovampyrus</i> Gunnell, Simons and Seiffert	<i>Saharaderma pseudovampyrus</i> Gunnell, Simons and Seiffert, 2008
31 Dec 2008	<i>Witwatia</i> Gunnell, Simons and Seiffert	<i>Witwatia</i> Gunnell, Simons and Seiffert, 2008
31 Dec 2008	<i>Witwatia eremicus</i> Gunnell, Simons and Seiffert	<i>Witwatia eremicus</i> Gunnell, Simons and Seiffert, 2008
31 Dec 2008	<i>Witwatia schlosseri</i> Gunnell, Simons and Seiffert	<i>Witwatia schlosseri</i> Gunnell, Simons and Seiffert, 2008
31 Dec 2008	<i>Pteropoidea</i> Serra-Cobo, López-Roig, Bayer, Amengual and Guasch	<i>PTEROPODOIDEA</i> Gray, 1821
31 Dec 2008	<i>Mops bakarii</i> Stanley	<i>Mops (Xiphonycteris) bakarii</i> Stanley, 2009
31 Dec 2008	<i>Vesperus hösemanni</i> Turni and Kock	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
31 Mar 2009	<i>Triaenops menamena</i> Goodman and Ranivo	<i>Triaenops menamena</i> Goodman and Ranivo, 2009
22 Jun 2009	<i>Miniopterus aelleni</i> Goodman, Maminirina, Weyeneth, Bradman, Christidis, Ruedi and Appleton	<i>Miniopterus aelleni</i> Goodman, Maminirina, Weyeneth, Bradman, Christidis, Ruedi and Appleton, 2009
31 Oct 2009	<i>Miniopterus griffithsi</i> Goodman, Maminirina, Bradman, Christidis and Appleton	<i>Miniopterus griffithsi</i> Goodman, Maminirina, Bradman, Christidis and Appleton, 2009
30 Nov 2009	<i>Miniopterus mahafaliensis</i> Goodman, Bradman, Christides and Appleton	<i>Miniopterus mahafaliensis</i> Goodman, Bradman, Christides and Appleton, 2009
30 Nov 2009	<i>Miniopterus brachytragos</i> Goodman, Maminirina, Bradman, Christidis and Appleton	<i>Miniopterus brachytragos</i> Goodman, Maminirina, Bradman, Christidis and Appleton, 2009
31 Dec 2009	<i>Paratriaenops</i> Benda and Vallo	<i>Paratriaenops</i> Benda and Vallo, 2009
31 Dec 2009	<i>Triaenopini</i> Benda and Vallo	<i>RHINONYCTERIDAE</i> J. E. Gray, 1866
28 Jul 2010	<i>Chaerephon atsinanana</i> Goodman, Buccas, Naidoo, Ratrimomanarivo, Taylor and Lamb	<i>Mops (Chaerephon) atsinanana</i> (Goodman, Buccas, Naidoo, Ratrimomanarivo, Taylor and Lamb, 2010)
31 Jan 2011	<i>Miniopterus horaceki</i> Gunnell, Eiting and Geraads	<i>Miniopterus horaceki</i> Gunnell, Eiting and Geraads, 2011
31 Jan 2011	<i>Myotis darelbeidensis</i> Gunnell, Eiting and Geraads	<i>Myotis darelbeidensis</i> Gunnell, Eiting and Geraads, 2011
31 Jan 2011	<i>Rhinolophus maghrebensis</i> Gunnell, Eiting and Geraads	<i>Rhinolophus maghrebensis</i> Gunnell, Eiting and Geraads, 2011
31 Jan 2011	<i>Pterocarpus</i> Mishra, Rout and Panda	<i>Pteropus</i> Brisson, 1762
17 May 2011	<i>Miniopterus egeri</i> Goodman, Ramasindrazana, Maminirina, Schoeman and Appleton	<i>Miniopterus egeri</i> Goodman, Ramasindrazana, Maminirina, Schoeman and Appleton, 2011
31 Dec 2011	<i>Necromantidae</i> Sigé	<i>NECROMANTIDAE</i> Sigé, 2011
29 Mar 2012	<i>Neoromicia robertsi</i> Goodman, Taylor, Ratrimomanarivo and Hoofer	<i>Laephotis robertsi</i> (Goodman, Taylor, Ratrimomanarivo and Hoofer, 2012)
12 Sep 2012	<i>Rhinolophus cohenae</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill	<i>Rhinolophus cohenae</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
12 Sep 2012	<i>Rhinolophus mabuensis</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill	<i>Rhinolophus mabuensis</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
12 Sep 2012	<i>Rhinolophus mossambicus</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill	<i>Rhinolophus mossambicus</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
12 Sep 2012	<i>Rhinolophus smithersi</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill	<i>Rhinolophus smithersi</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
18 Sep 2012	<i>Witwatia sigei</i> Ravel, Marivaux, Tabuce, Bel Haj Ali, Essid and Vianey-Liaud	<i>Witwatia sigei</i> Ravel, Marivaux, Tabuce, Bel Haj Ali, Essid and Vianey-Liaud, 2012
10 Oct 2012	<i>Stenonycterini</i> Nesi, Kadjo, Pourrut, Leroy, Pongombo Shongo, Cruaud and Hassanin	<i>Stenonycterini</i> Nesi, Kadjo, Pourrut, Leroy, Pongombo Shongo, Cruaud and Hassanin, 2012
31 Dec 2012	<i>Rhinolophus horaceki</i> Benda and Vallo	<i>Rhinolophus horaceki</i> Benda and Vallo, 2012
31 Dec 2012	<i>Coleura kibomalandy</i> Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling	<i>Coleura kibomalandy</i> Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012
31 Dec 2012	<i>Paremballonura</i> Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling	<i>Paremballonura</i> Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012
31 Dec 2012	<i>Paremballonura atrata</i> Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling	<i>Paremballonura atrata</i> (Peters, 1874)

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31 Dec 2012	<i>Paremballonura tiavato</i> Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling	<i>Paremballonura tiavato</i> (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)
31 Dec 2012	<i>Megaloglossus azagnyi</i> Nesi, Kadio and Hassanin	<i>Megaloglossus azagnyi</i> Nesi, Kadio and Hassanin, 2012
18 Jan 2013	<i>Neoromicia roseveari</i> Monadjem, Richards, Taylor and Stoffberg	<i>Pseudoromicia roseveari</i> (Monadjem, Richards, Taylor and Stoffberg, 2013)
30 Nov 2013	<i>Rhinolophus kahuzi</i> Fahr and Kerbis Peterhans	<i>Rhinolophus kahuzi</i> Fahr and Kerbis Peterhans, 2013
30 Nov 2013	<i>Rhinolophus willardi</i> Kerbis Peterhans and Fahr	<i>Rhinolophus willardi</i> Kerbis Peterhans and Fahr, 2013
10 Dec 2013	<i>Miniopterus mossambicus</i> Monadjem, Goodman, Stanley and Appleton	<i>Miniopterus mossambicus</i> Monadjem, Goodman, Stanley and Appleton, 2013
31 Dec 2013	<i>Niumbaha</i> Reeder, Helgen, Vozzak, Lunde and Ejotre	<i>Glauconycteris</i> Dobson, 1875
4 Feb 2014	<i>Phasmatonycteris</i> Gunnell, Simmons and Seiffert	<i>Phasmatonycteris</i> Gunnell, Simmons and Seiffert, 2014
4 Feb 2014	<i>Phasmatonycteris butleri</i> Gunnell, Simmons and Seiffert	<i>Phasmatonycteris butleri</i> Gunnell, Simmons and Seiffert, 2014
4 Feb 2014	<i>Phasmatonycteris phiomensis</i> Gunnell, Simmons and Seiffert	<i>Phasmatonycteris phiomensis</i> Gunnell, Simmons and Seiffert, 2014
5 Feb 2014	<i>Casinycteris campomaanensis</i> Hassanin	<i>Casinycteris campomaanensis</i> Hassanin, 2014
5 May 2014	<i>Miniopterus maghrebensis</i> Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste	<i>Miniopterus maghrebensis</i> Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014
26 Jun 2014	<i>Scotophilus andrewreborii</i> Brooks and Bickham	<i>Scotophilus andrewreborii</i> Brooks and Bickham, 2014
26 Jun 2014	<i>Scotophilus livingstonii</i> Brooks and Bickham	<i>Scotophilus livingstonii</i> Brooks and Bickham, 2014
26 Dec 2014	<i>Scotophilus ejetai</i> Brooks and Bickham	<i>Scotophilus ejetai</i> Brooks and Bickham, 2014
26 Dec 2014	<i>Scotophilus trujilloi</i> Brooks and Bickham	<i>Scotophilus trujilloi</i> Brooks and Bickham, 2014
31 Dec 2014	<i>Pteropodimorpha</i> Zagorodniuk	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
31 Dec 2014	<i>Vespertilioniformes</i> Zagorodniuk	<i>CHIROPTERA</i> Blumenbach, 1779
31 Dec 2014	<i>Vespertilionimorpha</i> Zagorodniuk	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
27 Jan 2015	<i>Scotonycteris bergmansi</i> Hassanin, Khouider, Gembu, Goodman, Kadio, Nesi, Pourrut, Nakouné and Bonillo	<i>Scotonycteris bergmansi</i> Hassanin, Khouider, Gembu, Goodman, Kadio, Nesi, Pourrut, Nakouné and Bonillo, 2015
27 Jan 2015	<i>Scotonycteris bergmansi bergmansi</i> Hassanin, Khouider, Gembu, Goodman, Kadio, Nesi, Pourrut, Nakouné and Bonillo	<i>Scotonycteris bergmansi bergmansi</i> Hassanin, Khouider, Gembu, Goodman, Kadio, Nesi, Pourrut, Nakouné and Bonillo, 2015
27 Jan 2015	<i>Scotonycteris bergmansi congoensis</i> Hassanin, Khouider, Gembu, Goodman, Kadio, Nesi, Pourrut, Nakouné and Bonillo	<i>Scotonycteris bergmansi congoensis</i> Hassanin, Khouider, Gembu, Goodman, Kadio, Nesi, Pourrut, Nakouné and Bonillo, 2015
23 Mar 2015	<i>Miniopterus ambohitrensis</i> Goodman, Ramasindrazana, Naughton and Appleton	<i>Miniopterus ambohitrensis</i> Goodman, Ramasindrazana, Naughton and Appleton, 2015
1 Oct 2015	<i>Hipposideros (Pseudorhinolophus) amenhotepos</i> Gunnell, Winkler, Miller, Head, El-Barkooky, Gawad, Sanders and Gingerich	<i>Hipposideros amenhotepos</i> Gunnell, Winkler, Miller, Head, El-Barkooky, Gawad, Sanders and Gingerich, 2015
9 Dec 2015	<i>Otomops harrisoni</i> Ralph, Richards, Taylor, Napier and Lamb	<i>Otomops harrisoni</i> Ralph, Richards, Taylor, Napier and Lamb, 2015
16 Dec 2015	<i>Cardioderma leakeyi</i> Gunnell, Butler, Greenwood and Simmons	<i>Cardioderma leakeyi</i> Gunnell, Butler, Greenwood and Simmons, 2015
16 Dec 2015	<i>Myzopoda africana</i> Gunnell, Butler, Greenwood and Simmons	<i>Myzopoda africana</i> Gunnell, Butler, Greenwood and Simmons, 2015
16 Dec 2015	<i>Nycticeinops serengetiensis</i> Gunnell, Butler, Greenwood and Simmons	<i>Nycticeinops serengetiensis</i> Gunnell, Butler, Greenwood and Simmons, 2015
16 Dec 2015	<i>Scotoecus olduvensis</i> Gunnell, Butler, Greenwood and Simmons	<i>Scotoecus olduvensis</i> Gunnell, Butler, Greenwood and Simmons, 2015
31 Dec 2015	<i>Hypsugo bemainty</i> Goodman, Rakotondramanana, Ramasindrazana, Kearney, Monadjem, Schoeman, Taylor, Naughton and Appleton	<i>Neoromicia bemainty</i> (Goodman, Rakotondramanana, Ramasindrazana, Kearney, Monadjem, Schoeman, Taylor, Naughton and Appleton, 2015)
27 Jan 2016	<i>Neoromicia isabella</i> Decher, Hutterer and Monadjem	<i>Pseudoromicia isabella</i> (Decher, Hutterer and Monadjem, 2016)
9 May 2016	<i>Aegyptonycteridae</i> Simmons, Seiffert and Gunnell	<i>AEGYPTONYCTERIDAE</i> Simmons, Seiffert and Gunnell, 2016
9 May 2016	<i>Aegyptonycteris</i> Simmons, Seiffert and Gunnell	<i>Aegyptonycteris</i> Simmons, Seiffert and Gunnell, 2016
9 May 2016	<i>Aegyptonycteris knightae</i> Simmons, Seiffert and Gunnell	<i>Aegyptonycteris knightae</i> Simmons, Seiffert and Gunnell, 2016

Publication date	Original Name	Current Name
20 May 2016	<i>Hipposideros cryptovalorona</i> Goodman, Schoeman, Rakotoarivelo and Willows-Munro	<i>Macronycteris cryptovalorona</i> (Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016)
30 Jun 2016	<i>Eidolinae</i> Almeida, Giannini and Simmons	<i>Eidolinae</i> Almeida, Giannini and Simmons, 2016
30 Sep 2016	? <i>Necromantis fragmentum</i> Ravel	<i>Necromantis fragmentum</i> (Ravel, 2016)
30 Sep 2016	? <i>Palaeophyllophora tunisiensis</i> Ravel	<i>Palaeophyllophora tunisiensis</i> (Ravel, 2016)
30 Sep 2016	? <i>Vespertiliavus aenigma</i> Ravel	<i>Vespertiliavus aenigma</i> (Ravel, 2016)
30 Sep 2016	<i>Chambinycteris</i> Ravel	<i>Chambinycteris</i> Ravel, 2016
30 Sep 2016	<i>Chambinycteris pusilli</i> Ravel	<i>Chambinycteris pusilli</i> Ravel, 2016
30 Sep 2016	<i>Drakonycteris</i> Ravel	<i>Drakonycteris</i> Ravel, 2016
30 Sep 2016	<i>Drakonycteris glibzegdouensis</i> Ravel	<i>Drakonycteris glibzegdouensis</i> Ravel, 2016
30 Sep 2016	<i>Hipposideros</i> (<i>Pseudorhinolophus</i>) <i>africanum</i> Ravel	<i>Hipposideros africanum</i> Ravel, 2016
30 Sep 2016	<i>Khoufechia</i> Ravel	<i>Khoufechia</i> Ravel, 2016
30 Sep 2016	<i>Khoufechia gunnelli</i> Ravel	<i>Khoufechia gunnelli</i> Ravel, 2016
30 Sep 2016	<i>Pseudovespertiliavus</i> Ravel	<i>Pseudovespertiliavus</i> Ravel, 2016
30 Sep 2016	<i>Pseudovespertiliavus parva</i> Ravel	<i>Pseudovespertiliavus parva</i> Ravel, 2016
30 Sep 2016	<i>Vespertiliavus kasserinensis</i> Ravel	<i>Vespertiliavus kasserinensis</i> (Ravel, 2016)
31 Dec 2016	<i>Nycteris cf. parisii</i> Kruskop, Benda, Vasenkov and Lavrenchenko	<i>Nycteris cf. parisii</i> De Beaux, 1924
31 Dec 2016	<i>Hipposideros cf. caffer</i> Monadjem, Richards and Denys	<i>Hipposideros caffer</i> (Sundevall, 1846)
31 Dec 2016	<i>Hipposideros cf. ruber</i> (lineage C1) Monadjem, Richards and Denys	<i>Hipposideros ruber</i> (Noack, 1893)
31 Dec 2016	<i>Hipposideros cf. ruber</i> (lineage E1) Monadjem, Richards and Denys	<i>Hipposideros ruber</i> (Noack, 1893)
16 Jun 2017	<i>Pipistrellus krugeri</i> Herkt, Skidmore and Fahr	<i>Laephotis stanleyi</i> (Goodman, Kearney, Ratsimbazafy and Hassanin, 2017)
30 Jun 2017	<i>M[acronycteris] cryptovalorona</i> Foley, Goodman, Whelan, Puechmaille and Teeling	<i>Macronycteris cryptovalorona</i> (Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016)
22 Aug 2017	<i>Glauconycteris atra</i> Hassanin, Colombo, Gembu, Merle, Tu, Görföl, Musaba Akawa, Csorba, Kearney, Monadjem and Ing	<i>Glauconycteris atra</i> Hassanin, Colombo, Gembu, Merle, Tu, Görföl, Musaba Akawa, Csorba, Kearney, Monadjem and Ing, 2017
31 Dec 2017	<i>Neoromicia stanleyi</i> Goodman, Kearney, Ratsimbazafy and Hassanin	<i>Laephotis stanleyi</i> (Goodman, Kearney, Ratsimbazafy and Hassanin, 2017)
6 Apr 2018	<i>Mops kerio</i> Gunnell and Manthi	<i>Mops kerio</i> Gunnell and Manthi, 2018
6 Apr 2018	<i>Mops turkwellensis</i> Gunnell and Manthi	<i>Mops turkwellensis</i> Gunnell and Manthi, 2018
6 Apr 2018	<i>R[oussettus] pattersoni</i> Gunnell and Manthi	<i>Roussettus pattersoni</i> Gunnell and Manthi, 2018
6 Apr 2018	<i>Saccolaimus kenyensis</i> Gunnell and Manthi	<i>Saccolaimus kenyensis</i> Gunnell and Manthi, 2018
6 Apr 2018	<i>Turkanycteris</i> Gunnell and Manthi	<i>Turkanycteris</i> Gunnell and Manthi, 2018
6 Apr 2018	<i>Turkanycteris harrisi</i> Gunnell and Manthi	<i>Turkanycteris harrisi</i> Gunnell and Manthi, 2018
24 Apr 2018	<i>Rhinolophus gorongosae</i> Taylor, MacDonald, Goodman, Kearney, Cotterill, Stoffberg, Monadjem, Schoeman, Guyton, Naskrecki and Richards	<i>Rhinolophus gorongosae</i> Taylor, MacDonald, Goodman, Kearney, Cotterill, Stoffberg, Monadjem, Schoeman, Guyton, Naskrecki and Richards, 2018
28 Jan 2019	<i>Myotis escalerae cabrerai</i> Çoraman, Mayer and Dietz	<i>Myotis zenatius</i> Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019
30 Jan 2019	<i>Myotis zenatius</i> Ibáñez, Juste, Salicini, Puechmaille and Ruedi	<i>Myotis zenatius</i> Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019
5 Oct 2019	<i>Parahypsugo</i> Hutterer, Decher, Monadjem and Astrin	<i>Nycticeinops</i> Hill & Harrison, 1987
5 Oct 2019	<i>Parahypsugo happoldorum</i> Hutterer, Decher, Monadjem and Astrin	<i>Nycticeinops happoldorum</i> (Hutterer, Decher, Monadjem and Astrin, 2019)
28 Feb 2020	<i>Parahypsugo macrocephalus</i> Hutterer and Kerbis Peterhans	<i>Nycticeinops macrocephalus</i> (Hutterer and Kerbis Peterhans, 2020)
29 Apr 2020	<i>Plecotus cf. gaisleri</i> Ancillotto, Bosso, Smeraldo, Mori, Mazza, Herkt, Galimberti, Ramazzotti and Russo	<i>Plecotus gaisleri</i> Benda, Kiefer, Hanak & Veith, 2004
3 Sep 2020	<i>Miniopterus wilsoni</i> Monadjem, Guyton, Naskrecki, Richards, Kropff and Dalton	<i>Miniopterus wilsoni</i> Monadjem, Guyton, Naskrecki, Richards, Kropff and Dalton, 2020
4 Sep 2020	<i>Pipistrellus simandouensis</i> Monadjem, Richards, Decher and Hutterer	<i>Pipistrellus (Pipistrellus) simandouensis</i> Monadjem, Richards, Decher and Hutterer, 2020
10 Sep 2020	<i>Nycticeinops bellieri</i> Monadjem, Demos, Dalton, Webala, Musila, Kerbis Peterhans and Patterson	<i>Nycticeinops bellieri</i> (De Vree, 1972)
10 Sep 2020	<i>Pseudoromicia kityoi</i> Monadjem, Kerbis Peterhans, Nalikka, Waswa, Demos and Patterson	<i>Pseudoromicia kityoi</i> Monadjem, Kerbis Peterhans, Nalikka, Waswa, Demos and Patterson, 2020
10 Sep 2020	<i>Afronycteris</i> Monadjem, Patterson and Demos	<i>Afronycteris</i> Monadjem, Patterson and Demos, 2020

Publication date	Original Name	Current Name
10 Sep 2020	<i>Laephotis kirinyaga</i> Monadjem, Patterson, Webala and Demos	<i>Laephotis kirinyaga</i> Monadjem, Patterson, Webala and Demos, 2020
10 Sep 2020	<i>Pseudoromicia</i> Monadjem, Patterson, Webala and Demos	<i>Pseudoromicia</i> Monadjem, Patterson, Webala and Demos, 2020
10 Sep 2020	<i>Pseudoromicia nyanza</i> Monadjem, Patterson, Webala and Demos	<i>Pseudoromicia nyanza</i> Monadjem, Patterson, Webala and Demos, 2020
8 Dec 2020	<i>Hipposideros cf. ruber (lineage D1)</i> Mamba, Dalton, Mahlaba, Kropff and Monadjem	<i>Hipposideros ruber</i> (Noack, 1893)
31 Dec 2020	<i>Miniopterus cf. inflatus</i> Monadjem, Shapiro, Richards, Karabulut, Crawley, Broman Nielsen, Hansen, Bohmann and Mourier	<i>Miniopterus cf. inflatus</i> Monadjem, Shapiro, Richards, Karabulut, Crawley, Broman Nielsen, Hansen, Bohmann and Mourier, 2020
31 Dec 2020	<i>Miniopterus nimbae</i> Monadjem, Shapiro, Richards, Karabulut, Crawley, Nielsen, Hansen, Bohmann and Mourier	<i>Miniopterus nimbae</i> Monadjem, Shapiro, Richards, Karabulut, Crawley, Nielsen, Hansen, Bohmann and Mourier, 2020
31 Dec 2020	<i>Mops bemmeleni</i> Simmons and Cirranello	<i>Mops (Chaerephon) bemmeleni</i> (Jentink, 1879)
31 Dec 2020	<i>Mops russatus</i> Simmons and Cirranello	<i>Mops (Chaerephon) russatus</i> (J.A. Allen, 1917)
14 Sep 2021	<i>Afrillonura</i> Rosina and Pickford	<i>Afrillonura</i> Rosina and Pickford, 2021
14 Sep 2021	<i>Afrillonura namibensis</i> Rosina and Pickford	<i>Afrillonura namibensis</i> Rosina and Pickford, 2021
31 Dec -330	<i>DERMAPTERA</i> Aristotle	<i>CHIROPTERA</i> Blumenbach, 1779
?	<i>Epomophorus labiatus anurus</i>	<i>Epomophorus minor</i> Dobson, 1880
?	<i>Laephotis cf. angolensis</i>	<i>Laephotis angolensis</i> Monard, 1935
?	<i>Myotis anjouanensis</i>	<i>Myotis anjouanensis</i> Dorst, 1960
?	<i>Noctilionoidea</i>	<i>NOCTILIONOIDEA</i> Gray, 1821
?	<i>Rousettus sp. cf. R. madagascariensis</i>	<i>Rousettus madagascariensis</i> G. Grandidier, 1929
?	<i>Yangochiroptera</i>	<i>VESPERTILIONIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
?	<i>Miniopterus sp.</i>	<i>Miniopterus</i> Bonaparte, 1837
?	<i>Neoromicia "incertae-sedis"</i>	<i>Laephotis "incertae-sedis"</i>
?	<i>Neoromicia cf. capensis</i>	<i>Laephotis capensis</i> (A. Smith, 1829)
?	<i>Pteropus aegyptiacus</i>	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
?	<i>Rhinonictidae</i>	<i>RHINONYCTERIDAE</i> J. E. Gray, 1866
?	<i>Scotophilus cf. viridis</i>	<i>Scotophilus viridis</i> (Peters, 1852)

Appendix 3d: Synonyms by Country of Type Specimen

In this appendix the synonyms are ordered by country from which they were described and by year or publication.

Synonyms for which "Null" is mentioned as author simply indicate incomplete entries, for which this information could not be found (yet).

Unknown Country

1762	<i>Pteropus</i> Brisson	<i>Pteropus</i> Brisson, 1762
1767	<i>Vespertiliones</i> Pallas	<i>CHIROPTERA</i> Blumenbach, 1779
1779	<i>CHIROPTERA</i> Blumenbach	<i>CHIROPTERA</i> Blumenbach, 1779
1795	<i>Nycteris</i> G. Cuvier and E. Geoffroy	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795
1797	<i>Vespertilio caninus</i> Blumenbach	<i>Pteropus niger</i> (Kerr, 1792)
1799	<i>Rhinolophus</i> Lacépède	<i>Rhinolophus</i> Lacépède, 1799
1799	<i>Spectrum</i> Lacépède	<i>Pteropus</i> Brisson, 1762
1803	<i>major</i> E. Geoffroy Saint-Hilaire	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
1806	<i>Chiropteren</i> Duméril	<i>CHIROPTERA</i> Blumenbach, 1779
1809	<i>Vespertilio caninus</i> var. <i>b</i> Goldfuss	<i>Eidolon helvum helvum</i> (Kerr, 1792)
1810	<i>Megaderma</i> E. Geoffroy St.-Hilaire	<i>Megaderma</i> E. Geoffroy St.-Hilaire, 1810
1814	<i>Tadarida</i> Rafinesque	<i>Tadarida</i> Rafinesque, 1814
1814	<i>Pteropus torquatus</i> G. Fischer	<i>Pteropus subniger</i> (Kerr, 1792)
1815	<i>Chiropteria</i> Rafinesque	<i>CHIROPTERA</i> Blumenbach, 1779
1815	<i>Eidolon</i> Rafinesque	<i>Eidolon</i> Rafinesque, 1815
1815	<i>Pteropus flavus</i> Illiger	<i>Eidolon helvum helvum</i> (Kerr, 1792)
1815	<i>Vespertilia</i> Rafinesque	<i>VESPERTILIONIDAE</i> Gray, 1821
1816	<i>Cheiroptères</i> G. Cuvier	<i>CHIROPTERA</i> Blumenbach, 1779
1816	<i>Macrotus</i> Leach	<i>Plecotus</i> E. Geoffroy St.-Hilaire, 1818
1816	<i>Phyllorhina</i> Leach	<i>Rhinolophus</i> Lacépède, 1799
1818	<i>Myopterus</i> E. Geoffroy Saint-Hilaire	<i>Myopterus</i> E. Geoffroy St.-Hilaire, 1818
1818	<i>Nyctinomus</i> E. Geoffroy Saint-Hilaire	<i>Tadarida</i> Rafinesque, 1814
1818	<i>Plecotus</i> E. Geoffroy Saint-Hilaire	<i>Plecotus</i> E. Geoffroy St.-Hilaire, 1818
1818	<i>Pteropus aegyptiacus</i> E. Geoffroy Saint-Hilaire	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1818	<i>Rhinopoma</i> E. Geoffroy Saint-Hilaire	<i>Rhinopoma</i> E. Geoffroy St.-Hilaire, 1818
1818	<i>Taphozous</i> E. Geoffroy Saint-Hilaire	<i>Taphozous</i> E. Geoffroy St.-Hilaire, 1818
1819	<i>Vespertilio dasykarpos</i> Kuhl	<i>Nyctalus leisleri</i> (Kuhl, 1817)
1820	<i>Eptesicus</i> Rafinesque	<i>Eptesicus</i> Rafinesque, 1820
1820	<i>Myopteris</i> Desmarest	<i>Myopterus</i> E. Geoffroy St.-Hilaire, 1818
1821	<i>Barbastella</i> Gray	<i>Barbastella</i> Gray, 1821
1821	<i>Nyctimones</i> Gray	<i>Tadarida</i> Rafinesque, 1814
1821	<i>Pteropidae</i> Gray	<i>PTEROPODIDAE</i> Gray, 1821
1821	<i>Vespertilionidae</i> Gray	<i>VESPERTILIONIDAE</i> Gray, 1821
1821	<i>Vespertilioninae</i> Gray	<i>Vespertilioninae</i> Gray, 1821
1821	<i>Vespertilionini</i> Gray	<i>Vespertilionini</i> Gray, 1821
1821	<i>Vespertilionoidea</i> Gray	<i>VESPERTILIONOIDEA</i> Gray, 1821
1821	<i>Scotophilus</i> Leach	<i>Scotophilus</i> Leach, 1821
1821	<i>Cephalotidae</i> Gray	<i>PTEROPODIDAE</i> Gray, 1821
1821	<i>CHEIROPTERA</i> Gray	<i>CHIROPTERA</i> Blumenbach, 1779
1821	<i>FRUCTIVORAE</i> Gray	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
1821	<i>INSECTIVORAE</i> Gray	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
1821	<i>Nyctinoma</i> Bowdich	<i>Tadarida</i> Rafinesque, 1814
1821	<i>Pteropodinae</i> Gray	<i>Pteropodinae</i> Gray, 1821
1821	<i>Pteropodini</i> Gray	<i>Pteropodini</i> Gray, 1821
1821	<i>PTEROPODOIDEA</i> Gray	<i>PTEROPODOIDEA</i> Gray, 1821
1821	<i>Rhinopoma</i> Bowdich	<i>Rhinopoma</i> E. Geoffroy St.-Hilaire, 1818
1821	<i>Vespertilionini</i> Gray	<i>Vespertilioninae</i> Gray, 1821
1822	<i>CHEIROPTERA</i> Fleming	<i>CHIROPTERA</i> Blumenbach, 1779
1822	<i>Nyctinomia</i> Fleming	<i>Tadarida</i> Rafinesque, 1814
1825	<i>Pteropina</i> Gray	<i>PTEROPODIDAE</i> Gray, 1821
1825	<i>Vespertilionina</i> Gray	<i>VESPERTILIONIDAE</i> Gray, 1821
1825	<i>Dinops</i> Savi	<i>Tadarida</i> Rafinesque, 1814
1825	<i>Nyctalus</i> Bowdich	<i>Nyctalus</i> Bowdich, 1825
1825	<i>Rhinolophidae</i> Gray	<i>RHINOLOPHIDAE</i> Gray, 1825
1825	<i>Rhinolophina</i> Gray	<i>RHINOLOPHIDAE</i> Gray, 1825
1825	<i>Rhinolophina</i> Gray	<i>RHINOLOPHOIDEA</i> J. E. Gray, 1825
1825	<i>Rhinolophoidea</i> Gray	<i>RHINOLOPHOIDEA</i> J. E. Gray, 1825
1826	<i>Vespertilio leucomelas</i> Cretzschmar	<i>Barbastella leucomelas</i> (Cretzschmar, 1826)

1827	<i>Dysopes geoffroyi</i> Temminck	<i>Tadarida aegyptiaca aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
1827	<i>Rhinolophina</i> Lesson	<i>RHINOLOPHIDAE</i> Gray, 1825
1829	<i>Cercopteropus</i> Burnett	<i>Rousettus</i> Gray, 1821
1829	<i>[Vespertilio auritus] β aegyptius</i> J.B. Fischer	<i>Plecotus christii</i> Gray, 1838
1829	<i>Barbastellus</i> Kaup	<i>Barbastella</i> Gray, 1821
1829	<i>Cnephæus</i> Kaup	<i>Eptesicus (Cnephæus)</i> Kaup, 1829
1829	<i>Myotis</i> Kaup	<i>Myotis</i> Kaup, 1829
1829	<i>Nystactes</i> Kaup	<i>Myotis</i> Kaup, 1829
1829	<i>Pipistrellus</i> Kaup	<i>Pipistrellus</i> Kaup, 1829
1829	<i>Pterygistes</i> Kaup	<i>Nyctalus</i> Bowdich, 1825
1830	<i>Leuconoe</i> Boie	<i>Myotis</i> Kaup, 1829
1830-1831	<i>Dysopes Cretzschmar</i>	<i>Tadarida</i> Rafinesque, 1814
1831	<i>Hipposideros</i> Gray	<i>Hipposideros</i> Gray, 1831
1831	<i>Pachyotus</i> Gray	<i>Scotophilus</i> Leach, 1821
1831	<i>Phyllorhina</i> Bonaparte	<i>Hipposideros</i> Gray, 1831
1831	<i>Pteropodina</i> Bonaparte	<i>Pteropodinae</i> Gray, 1821
1834	<i>Hipposiderus</i> Gray	<i>Hipposideros</i> Gray, 1831
1835	<i>Pteropodae</i> Bonaparte	<i>Pteropodinae</i> Gray, 1821
1836	<i>Epomophorus</i> Bennett	<i>Epomophorus</i> Bennett, 1836
1836	<i>Rhinocrepis</i> Gervais	<i>Rhinolophus</i> Lacépède, 1799
1837	<i>Miniopterus</i> Bonaparte	<i>Miniopterus</i> Bonaparte, 1837
1837	<i>Myoptera</i> de Blainville	<i>Myopterus</i> E. Geoffroy St.-Hilaire, 1818
1837	<i>Noctula</i> Bonaparte	<i>Eptesicus</i> Rafinesque, 1820
1837	<i>Phyllorhina</i> Bonaparte	<i>Hipposideros</i> Gray, 1831
1837	<i>Pipistrella</i> Bonaparte	<i>Pipistrellus</i> Kaup, 1829
1837	<i>Pteropus vulgaris</i> Temminck	<i>Pteropus subniger</i> (Kerr, 1792)
1838	<i>Asellia</i> Gray	<i>Asellia</i> Gray, 1838
1838	<i>Barbastellus communis</i> Gray	<i>Barbastella barbastellus</i> (Schreber, 1774)
1838	<i>Lavia</i> Gray	<i>Lavia</i> Gray, 1838
1838	<i>Petalia</i> Gray	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795
1838	<i>Romicia</i> Gray	<i>Pipistrellus</i> Kaup, 1829
1838	<i>PTEROPODIDAE</i> Bonaparte	<i>PTEROPODIDAE</i> Gray, 1821
1838	<i>Rhinopomina</i> Bonaparte	<i>RHINOPOMATIDAE</i> Dobson, 1872
1838	<i>Saccolaimus</i> Temminck	<i>Saccolaimus</i> Temminck, 1838
1839	<i>Synotus</i> Keyserling and Blasius	<i>Barbastella</i> Gray, 1821
1839	<i>Vesperugo</i> Keyserling and Blasius	<i>Eptesicus</i> Rafinesque, 1820
1839	<i>Vesperugo</i> Keyserling and Blasius	<i>Nyctalus</i> Bowdich, 1825
1839	<i>Vesperugo</i> Keyserling and Blasius	<i>Pipistrellus</i> Kaup, 1829
1839	<i>Vesperus</i> Keyserling and Blasius	<i>Eptesicus</i> Rafinesque, 1820
1840	<i>Dysopes Savii</i> Schinz	<i>Tadarida teniotis</i> (Rafinesque, 1814)
1840	<i>Romicius</i> Blyth	<i>Pipistrellus</i> Kaup, 1829
1841	<i>Capaccinius</i> Bonaparte	<i>Myotis</i> Kaup, 1829
1841	<i>Selysius</i> Bonaparte	<i>Myotis</i> Kaup, 1829
1842	<i>Kerivoula</i> Gray	<i>Kerivoula</i> Gray, 1842
1842	<i>Noctulinia</i> Gray	<i>Nyctalus</i> Bowdich, 1825
1842	<i>Trilatitus</i> Gray	<i>Myotis</i> Kaup, 1829
1842	<i>Harpyidae</i> C. H. Smith	<i>PTEROPODIDAE</i> Gray, 1821
1842	<i>Mops</i> Lesson	<i>Mops (Mops)</i> Lesson, 1842
1842	<i>Mops</i> Lesson	<i>Mops</i> Lesson, 1842
1842	<i>Nicticejus</i> Rüppell	<i>Scotophilus</i> Leach, 1821
1843	<i>Vespertilio</i> Gray	<i>Myotis</i> Kaup, 1829
1843	<i>Xantharpyia</i> Gray	<i>Rousettus</i> Gray, 1821
1843	<i>Gymnorhina</i> Wagner	<i>VESPERTILIONIDAE</i> Gray, 1821
1844	<i>Eleutherura</i> Gray	<i>Rousettus</i> Gray, 1821
1847	<i>Aquias</i> Gray	<i>Rhinolophus</i> Lacépède, 1799
1847	<i>Eucheira</i> Hodgson	<i>Megaderma</i> E. Geoffroy St.-Hilaire, 1810
1849	<i>Kirivoula</i> Gervais	<i>Kerivoula</i> Gray, 1842
1849	<i>Vespertilio minutus</i> A. Smith	<i>Eptesicus (Cnephæus) hottentotus</i> (A. Smith, 1833)
1852	<i>Cynonycteris</i> Peters	<i>Rousettus</i> Gray, 1821
1853	<i>Pachysoma</i> Temminck	<i>Epomophorus</i> Bennett, 1836
1854	<i>Rhinolophides</i> Gervais	<i>RHINOLOPHIDAE</i> Gray, 1825
1855	<i>Emballonuridae</i> Gervais	<i>EMBALLONURIDAE</i> Gervais, 1855
1855	<i>Emballonurinae</i> Gervais	<i>Emballonurinae</i> Gervais, 1855
1855	<i>Nycticeiini</i> Gervais	<i>Nycticeiini</i> Gervais, 1855
1855	<i>Nycticeina</i> Gervais	<i>Nycticeiini</i> Gervais, 1855
1855	<i>Emballonurini</i> Gervais	<i>Emballonurini</i> Gervais, 1855
1855	<i>FRUGIVORA</i> Giebel	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
1855	<i>Gymnorhina</i> Giebel	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
1855	<i>Histiorhina</i> Van der Hoeven	<i>RHINOLOPHIDAE</i> Gray, 1825
1855	<i>Nycteridae</i> Van der Hoeven	<i>NYCTERIDAE</i> Van der Hoeven, 1855
1855	<i>Nycteroides</i> Van der Hoeven	<i>NYCTEROIDEA</i> Van der Hoeven, 1855
1855	<i>Vespertilio smithii</i> Wagner	<i>Eptesicus (Cnephæus) hottentotus</i> (A. Smith, 1833)
1856	<i>Molossidae</i> Gervais	<i>MOLOSSIDAE</i> Gervais, 1856

1856	<i>Molossinae</i> Gervais	<i>Molossinae</i> Gervais, 1856
1855	<i>Nycticeina</i> Gervais	VESPERTILIONIDAE Gray, 1821
1856	<i>Brachyotus</i> Kolenati	<i>Myotis</i> Kaup, 1829
1856	<i>Cateorus</i> Kolenati	<i>Eptesicus</i> Rafinesque, 1820
1856	<i>Hypsugo</i> Kolenati	<i>Hypsugo</i> Kolenati, 1856
1856	<i>Isotus</i> Kolenati	<i>Myotis</i> Kaup, 1829
1856	<i>Meteorus</i> Kolenati	<i>Eptesicus</i> Rafinesque, 1820
1856	<i>Molossoidea</i> Gervais	MOLOSSOIDEA Gervais, 1856
1856	<i>Nannugo</i> Kolenati	<i>Pipistrellus</i> Kaup, 1829
1856	<i>Panugo</i> Kolenati	<i>Nyctalus</i> Bowdich, 1825
1858	<i>Amblyotus</i> Kolenati	<i>Eptesicus</i> Rafinesque, 1820
1859	<i>Otonycteris</i> Peters	<i>Otonycteris</i> Peters, 1859
1860	<i>Phyllorhina</i> Koch	HIPPOSIDERIDAE Lydekker, 1891
1860	<i>Phyllorrhina</i> Koch	HIPPOSIDERIDAE Lydekker, 1891
1861	<i>Nyctophylax</i> Fitzinger	<i>Kerivoula</i> Gray, 1842
1862	<i>Sphyrocephalus</i> A. Murray	<i>Hypsignathus</i> H. Allen, 1862
1862	<i>Zygænocephalus</i> A. Murray	<i>Hypsignathus</i> H. Allen, 1862
1862	<i>Hypsignathus</i> H. Allen	<i>Hypsignathus</i> H. Allen, 1862
1862	<i>Pterocyon</i> Peters	<i>Eidolon</i> Rafinesque, 1815
1863	<i>Aristippe</i> Kolenati	<i>Eptesicus</i> Rafinesque, 1820
1864	<i>Megadermatidae</i> H. Allen	MEGADERMATIDAE H. Allen, 1864
1865	<i>Brachyura</i> Peters	EMBALLONURIDAE Gervais, 1855
1865	<i>Molossi</i> Peters	MOLOSSIDAE Gervais, 1856
1865	<i>Mormopterus</i> Peters	<i>Mormopterus</i> Peters, 1865
1865	<i>Nyctophilinae</i> Peters	Vespertilioninae Gray, 1821
1865	<i>Rhinolophi</i> Peters	RHINOLOPHIDAE Gray, 1825
1866	<i>Nycterica</i> Gray	NYCTERIDAE Van der Hoeven, 1855
1866	<i>Diclidurina</i> Gray	EMBALLONURIDAE Gervais, 1855
1866	<i>Emballonurina</i> Gray	EMBALLONURIDAE Gervais, 1855
1866	<i>Molossina</i> Gray	MOLOSSIDAE Gervais, 1856
1866	<i>Molossina</i> Gray	<i>Molossinae</i> Gervais, 1856
1866	<i>Nyctophilina</i> Gray	VESPERTILIONIDAE Gray, 1821
1866	<i>Pachyomus</i> Gray	<i>Eptesicus</i> Rafinesque, 1820
1866	<i>Plecotina</i> Gray	VESPERTILIONIDAE Gray, 1821
1866	<i>Plecotini</i> Gray	<i>Plecotini</i> Gray, 1866
1866	<i>Romiciana</i> Gray	VESPERTILIONIDAE Gray, 1821
1866	<i>Chrysonycteris</i> Gray	<i>Hipposideros</i> Gray, 1831
1866	<i>Epomophorina</i> Gray	<i>Pteropodinae</i> Gray, 1821
1866	<i>Epomophorini</i> Gray	<i>Epomophorini</i> Gray, 1866
1866	<i>Eunycteris</i> Gray	<i>Pteropus</i> Brisson, 1762
1866	<i>Gloionycteris</i> Gray	<i>Hipposideros</i> Gray, 1831
1866	<i>Macronycteris</i> Gray	<i>Macronycteris</i> Gray, 1866
1866	<i>Megadermina</i> Gray	MEGADERMATIDAE H. Allen, 1864
1866	<i>Nycterops</i> Gray	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795
1866	<i>Phyllotis</i> Gray	<i>Rhinolophus</i> Lacépède, 1799
1866	<i>Rhinonycterina</i> Gray	RHINONYCTERIDAE J. E. Gray, 1866
1866	<i>Rhinophylla</i> Gray	<i>Hipposideros</i> Gray, 1831
1866	<i>Speorifera</i> Gray	<i>Hipposideros</i> Gray, 1831
1866	<i>Entomophaga</i> A. Murray	VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
1866	<i>Histiorrhina</i> Haeckel	RHINOLOPHIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
1866	<i>Hypodermida</i> Haeckel	PTEROPODIDAE Gray, 1821
1866	<i>Megadermata</i> Peters	MEGADERMATIDAE H. Allen, 1864
1866	<i>Nycterides</i> Haeckel	CHIROPTERA Blumenbach, 1779
1866	<i>Pterocynes</i> Haeckel	PTEROPODIFORMI Van Cakenberghe, Kearney and Seamark, 2007
1866	<i>Pteropi</i> Peters	PTEROPODIDAE Gray, 1821
1866	<i>Pteropodida</i> Haeckel	PTEROPODIDAE Gray, 1821
1866	<i>Rhinonycteridae</i> Gray	RHINONYCTERIDAE J. E. Gray, 1866
1866	<i>Scotophilii</i> A. Murray	<i>Scotophilinae</i> Van Cakenberghe and Seamark, 2008
1866	<i>Scotophilina</i> Gray	<i>Scotophilinae</i> Van Cakenberghe and Seamark, 2008
1866	<i>Spasma</i> Gray	<i>Megaderma</i> E. Geoffroy St.-Hilaire, 1810
1866	<i>Vespertiones</i> Peters	VESPERTILIONIDAE Gray, 1821
1867	<i>Coeloephyllus</i> Peters	<i>Rhinolophus</i> Lacépède, 1799
1867	<i>Coelophyllus</i> Peters	<i>Rhinolophus</i> Lacépède, 1799
1867	<i>Chalinolobus</i> Peters	<i>Glauconycteris</i> Dobson, 1875
1867	<i>Colëura</i> Peters	<i>Coleura</i> Peters, 1867
1867	<i>Taphozoinae</i> Jerdon	<i>Taphozoinae</i> Jerdon, 1867
1867	<i>Vespertilio</i> (<i>Pteropterus</i>) Peters	<i>Myotis</i> Kaup, 1829
1868	<i>Alobus</i> Peters	<i>Pipistrellus</i> Kaup, 1829
1868	<i>Pteropterus</i> Peters	<i>Myotis</i> Kaup, 1829
1868	<i>Rhinopoma sennarensense</i> Hartmann	<i>Rhinopoma cystops</i> Thomas, 1903
1869	<i>Gymnorhinida</i> Fatio	VESPERTILIONIDAE Gray, 1821
1870	<i>Aëorestes</i> Fitzinger	<i>Myotis</i> Kaup, 1829

1870	<i>[Epomophorus macrocephalus]</i> var. <i>unicolor</i> Gray	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
1870	<i>Comastes</i> Fitzinger	<i>Myotis</i> Kaup, 1829
1870	<i>Epomops</i> Gray	<i>Epomops</i> Gray, 1866
1870	<i>Exochurus</i> Fitzinger	<i>Myotis</i> Kaup, 1829
1870	<i>Nyctiptenus</i> Fitzinger	<i>Eptesicus</i> Rafinesque, 1820
1870	<i>Pselaphon</i> Gray	<i>Pteropus</i> Brisson, 1762
1870	<i>Rhinolophus Eggenhöfner</i> Fitzinger	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1870	<i>Senonycteris</i> Gray	<i>Rousettus</i> Gray, 1821
1870	<i>Spectrum vulgare</i> Gray	<i>Pteropus niger</i> (Kerr, 1792)
1870	<i>Stenonycteris</i> Gray	<i>Stenonycteris</i> Gray, 1870
1871	<i>Triænops</i> Dobson	<i>Triænops</i> Dobson, 1871
1871	<i>Cyclorhina</i> Peters	<i>Hipposideros</i> Gray, 1831
1871	<i>Doryrhina</i> Peters	<i>Doryrhina</i> Peters, 1871
1871	<i>Ptychorhina</i> Peters	<i>Hipposideros</i> Gray, 1831
1871	<i>Sideroderma</i> Peters	<i>Hipposideros</i> Gray, 1831
1871	<i>Sybdesmotus</i> Peters	<i>Hipposideros</i> Gray, 1831
1871	<i>Syndesmotis</i> Peters	<i>Hipposideros</i> Gray, 1831
1871	<i>Thyreorhina</i> Peters	<i>Hipposideros</i> Gray, 1831
9999	<i>Noctulina</i> Macalister	<i>Nyctalus</i> Bowdich, 1825
1872	<i>Nycticejinae</i> Gill	<i>Nycticeiini</i> Gervais, 1855
1872	ANIMALIVORA Gill	CHIROPTERA Blumenbach, 1779
1872	<i>Lyroderma</i> Peters	<i>Megaderma</i> E. Geoffroy St.-Hilaire, 1810
1872	<i>Megadermidae</i> Gill	MEGADERMATIDAE H. Allen, 1864
1872	<i>Molossidae</i> Gill	MOLOSSIDAE Gervais, 1856
1872	<i>Rhinopomatidae</i> Dobson	RHINOPOMATIDAE Dobson, 1872
1872	<i>Rhinopomatoidea</i> Dobson	RHINOPOMATOIDEA Dobson, 1872
1873	<i>Cardioderma</i> Peters	<i>Cardioderma</i> Peters, 1873
1874	<i>Nyctinomus (Chaerephon)</i> Dobson	<i>Mops (Chaerephon)</i> Dobson, 1874
1875	<i>Glauconycteris</i> Dobson	<i>Glauconycteris</i> Dobson, 1875
1875	<i>Emballonuridae</i> Dobson	EMBALLONURIDAE Gervais, 1855
1875	<i>Miniopteri</i> Dobson	MINIOPTERIDAE Dobson, 1875
1875	<i>Miniopteridae</i> Dobson	MINIOPTERIDAE Dobson, 1875
1875	MEGACHIROPTERA Dobson	PTEROPODIFORMI Van Cakenberghe, Kearney and Seamark, 2007
1875	MICROCHIROPTERA Dobson	CHIROPTERA Blumenbach, 1779
1876	<i>Taphonycteris</i> Dobson	<i>Saccolaimus</i> Temminck, 1838
1877	<i>Nyctinomus [(Nyctinomus)] cestoni</i> Dobson	<i>Tadarida ventralis</i> (Heuglin, 1861)
1877	<i>Vesperugo senarensis</i> Heuglin	<i>Vansonia rueppellii rueppellii</i> (J.B. Fischer, 1829)
1878	<i>Myzopoda</i> A. Milne-Edwards and A. Grandidier	<i>Myzopoda</i> Milne-Edwards and A. Grandidier, 1878
1878	<i>Vespertiliones</i> Dobson	<i>Vespertilioninae</i> Gray, 1821
1881	<i>Leiponyx</i> Jentink	<i>Eidolon</i> Rafinesque, 1815
1881	<i>Triænops Humbloti</i> A. Milne-Edwards	<i>Triænops menamena</i> Goodman and Ranivo, 2009
1882	<i>Liponyx</i> Forbes	<i>Eidolon</i> Rafinesque, 1815
1883	<i>Chiropteri de Rochebrune</i>	CHIROPTERA Blumenbach, 1779
1883	<i>Phyllorhinidae de Rochebrune</i>	RHINOLOPHIDAE Gray, 1825
1885	<i>Megaloglossus Pagenstecher</i>	<i>Megaloglossus</i> Pagenstecher, 1885
1887	<i>Vespertiliavus Schlosser</i>	<i>Vespertiliavus</i> Schlosser, 1887
1888	<i>Necromanter Lydekker</i>	<i>Necromantis</i> Weithofer, 1887
1888	<i>Necromantis</i> Weithofer	<i>Necromantis</i> Weithofer, 1887
1889	<i>Gymnuridae Ameghino</i>	MOLOSSIDAE Gervais, 1856
1889	<i>Ptética Ameghino</i>	CHIROPTERA Blumenbach, 1779
1891	<i>N(yctinomus) taeniotis</i> Thomas	<i>Tadarida ventralis</i> (Heuglin, 1861)
1891	<i>Cerivoula Blanford</i>	<i>Kerivoula</i> Gray, 1842
1891	<i>Hipposideridae Lydekker</i>	HIPPOSIDERIDAE Lydekker, 1891
1891	<i>Hipposiderinae Lydekker</i>	HIPPOSIDERIDAE Lydekker, 1891
1891	<i>Trygenycteris Lydekker</i>	<i>Megaloglossus</i> Pagenstecher, 1885
1892	<i>Adelonycteris</i> H. Allen	<i>Eptesicus</i> Rafinesque, 1820
1893	<i>Emballonurini Winge</i>	EMBALLONURIDAE Gervais, 1855
1893	<i>Megadermatini Winge</i>	MEGADERMATIDAE H. Allen, 1864
1893	<i>Molossini Winge</i>	MOLOSSIDAE Gervais, 1856
1893	<i>Pteropi Winge</i>	PTEROPODIDAE Gray, 1821
1893	<i>Rhinolophini Winge</i>	RHINOLOPHIDAE Gray, 1825
1894	<i>Scotonycteris Matschie</i>	<i>Scotonycteris</i> Matschie, 1894
1897	<i>Plecotinæ Miller</i>	<i>Plecotini</i> Gray, 1866
1897	<i>Vespertilioninæ Miller</i>	<i>Vespertilioninae</i> Gray, 1821
1897	<i>midas</i> Schulze	<i>Tadarida teniotis</i> (Rafinesque, 1814)
1897	<i>Pteropinae Trouessart</i>	<i>Pteropodinae</i> Gray, 1821
1898	<i>Miniopterae Trouessart</i>	MINIOPTERIDAE Dobson, 1875
1899	<i>[Epomophorus (Nanonycteris)]</i> Matschie	<i>Nanonycteris</i> Matschie, 1899
1899	<i>[Xantharpyia (Myonycteris)]</i> Matschie	<i>Myonycteris</i> Matschie, 1899
1899	<i>Euvespertilio Aclouque</i>	<i>Myotis</i> Kaup, 1829
1899	<i>Euvesperugo Aclouque</i>	<i>Pipistrellus</i> Kaup, 1829
1899	<i>Hypsognathus haldemani</i> Matschie	<i>Hypsognathus monstrosus</i> H. Allen, 1862
1899	<i>Mynonycteris</i> Matschie	<i>Rousettus</i> Gray, 1821

1899	<i>Sericonycteris</i> Matschie	<i>Pteropus</i> Brisson, 1762
1901	<i>N(yctinomus) midas</i> de Winton	<i>Tadarida ventralis</i> (Heuglin, 1861)
1901	<i>Scotæcus</i> Thomas	<i>Scotoecus</i> Thomas, 1901
1901	<i>Læphotis</i> Thomas	<i>Laephotis</i> Thomas, 1901
1901	<i>Clæotis</i> Thomas	<i>Cloeotis</i> Thomas, 1901
1901	<i>Glauconycteris beatrix</i> Thomas	<i>Glauconycteris beatrix</i> Thomas, 1901
1901	<i>Euryalus</i> Matschie	<i>Rhinolophus</i> Lacépède, 1799
1902	<i>Syndesmotus</i> C.O. Waterhouse	<i>Hipposideros</i> Gray, 1831
1903	<i>Necronycteris</i> Palmer	<i>Necromantis</i> Weithofer, 1887
1904	<i>Myzopodidae</i> Thomas	<i>MYZOPODIDAE</i> Thomas, 1904
1904	<i>Mimetillus</i> Thomas	<i>Mimetillus</i> Thomas, 1904
1904	<i>Kehelvoulha</i> Jentink	<i>Kerivoula</i> Gray, 1842
1905	<i>Rhinolophus hipposiderus typicus</i> K. Andersen	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1905	<i>Eomops</i> Thomas	<i>Myopterus</i> E. Geoffroy St.-Hilaire, 1818
1906	<i>Platymops</i> Thomas	<i>Platymops</i> Thomas, 1906
1906	<i>Pizonyx</i> Miller	<i>Myotis</i> Kaup, 1829
1906	<i>Rhinopterus</i> Miller	<i>Eptesicus (Rhinopterus)</i> Miller, 1906
1907	<i>Desmaplex</i> Miller	<i>Pteropus</i> Brisson, 1762
1907	<i>Diclidurinae</i> Miller	<i>EMBALLONURIDAE</i> Gervais, 1855
1907	<i>Hipposideridae</i> Miller	<i>HIPPOSIDERIDAE</i> Lydekker, 1891
1907	<i>Kerivoulinae</i> Miller	<i>Kerivoulinae</i> Miller, 1907
1907	<i>Nyctophilinae</i> Miller	<i>VESPERTILIONIDAE</i> Gray, 1821
1907	<i>Rhinopomidae</i> Miller	<i>RHINOPOMATIDAE</i> Dobson, 1872
1908	<i>Scabrifer</i> G.M. Allen	<i>Laephotis</i> Thomas, 1901
1908	<i>Scoteinus schlieffeni bedouin</i> Thomas and Wroughton	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
1909	<i>Otonycteris cinereus</i> Satunin	<i>Otonycteris hemprichii</i> Peters, 1859
1910	<i>Plerotes</i> K. Andersen	<i>Plerotes</i> K. Andersen, 1910
1910	<i>Chrysopteron</i> Jentink	<i>Myotis</i> Kaup, 1829
1910	<i>Casinycteris</i> Thomas	<i>Casinycteris</i> Thomas, 1910
1910	<i>Petaliidae</i> Miller	<i>NYCTERIDAE</i> Van der Hoeven, 1855
1910	<i>Rhinolophus blasiusi</i> Trouessart	<i>Rhinolophus blasii</i> Peters, 1867
1910	<i>Vampyravus</i> Schlosser	<i>Vampyravus</i> Schlosser, 1910
1911	<i>Xiphonycteris</i> Dollman	<i>Mops (Xiphonycteris)</i> Dollman, 1911
1911	<i>Provampyrus</i> Schlosser	<i>Vampyravus</i> Schlosser, 1910
1911	<i>Provampyrus orientalis</i> Schlosser	<i>Vampyravus orientalis</i> Schlosser, 1910
1912	<i>Cynopterinae</i> K. Andersen	<i>Pteropodinae</i> Gray, 1821
1912	<i>Epomophorinae</i> K. Andersen	<i>Rousettinae</i> Andersen, 1912
1912	<i>Rousettinae</i> K. Andersen	<i>Rousettinae</i> Andersen, 1912
1912	<i>Rousettini</i> K. Andersen	<i>Rousettini</i> Andersen, 1912
1912	<i>Cistugo</i> Thomas	<i>Cistugo</i> Thomas, 1912
1913	<i>Otomops</i> Thomas	<i>Otomops</i> Thomas, 1913
1914	<i>acrotis</i> G.M. Allen	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1914	<i>Noctulinia</i> Cabrera	<i>Nyctalus</i> Bowdich, 1825
1917	<i>Dichromyotis</i> Bianchi	<i>Myotis</i> Kaup, 1829
1917	<i>Paramyotis</i> Bianchi	<i>Myotis</i> Kaup, 1829
1917	<i>Rickettia</i> Bianchi	<i>Myotis</i> Kaup, 1829
1917	<i>Megapipistrellus</i> Bianchi	<i>Myotis</i> Kaup, 1829
1917	<i>Platymops</i> Roberts	<i>Sauromys</i> Roberts, 1917
1917	<i>Platymops (Sauromys)</i> Roberts	<i>Sauromys</i> Roberts, 1917
1917	<i>Allomops</i> J.A. Allen	<i>Mops (Mops)</i> Lesson, 1842
1917	<i>Chærephon (Lophomops)</i> J.A. Allen	<i>Mops (Chærephon)</i> Dobson, 1874
1917	<i>Nycteris major</i> J.A. Allen	<i>Nycteris macrotis</i> Dobson, 1876
1917	<i>Palaeophyllophora</i> Revilliod	<i>Palaeophyllophora</i> Revilliod, 1917
1917	<i>Pareptesicus</i> Bianchi	<i>Eptesicus</i> Rafinesque, 1820
1917	<i>Rhynptesicus</i> Bianchi	<i>Eptesicus</i> Rafinesque, 1820
1821	<i>Rousettus</i> Gray	<i>Rousettus</i> Gray, 1821
1922	<i>Liponycteris</i> Thomas	<i>Taphozous</i> E. Geoffroy St.-Hilaire, 1818
1922	<i>Kerivoula lueia</i> Kershaw	<i>Kerivoula lanosa</i> (A. Smith, 1847)
1926	<i>Eptesicops</i> Roberts	<i>Pipistrellus</i> Kaup, 1829
1926	<i>Neoromicia</i> Roberts	<i>Neoromicia</i> Roberts, 1926
1928	<i>Emballonuroidea</i> Weber	<i>EMBALLONURIDAE</i> Gervais, 1855
1929	<i>Anamygdon</i> Troughton	<i>Myotis</i> Kaup, 1829
1931	<i>Tuitatus</i> Kishida and Mori	<i>Eptesicus</i> Rafinesque, 1820
1934	<i>Austronomus</i> Iredale and Troughton	<i>Tadarida</i> Rafinesque, 1814
1939	<i>Mops rüppellii</i> Allen	<i>Tadarida ventralis</i> (Heuglin, 1861)
1941	<i>Coelopinae</i> Tate	<i>HIPPOSIDERIDAE</i> Lydekker, 1891
1941	<i>Austronomus</i> Troughton	<i>Tadarida</i> Rafinesque, 1814
1941	<i>Rhinophyllotis</i> Troughton	<i>Rhinolophus</i> Lacépède, 1799
1942	<i>Myotinae</i> Tate	<i>Myotinae</i> Tate, 1942
1942	<i>Myotini</i> Tate	<i>VESPERTILIONIDAE</i> Gray, 1821
1942	<i>Myotini</i> Tate	<i>Myotinae</i> Tate, 1942
1942	<i>Nycticeini</i> Tate	<i>Vespertilioninae</i> Gray, 1821
1842	<i>Pipistrellini</i> Tate	<i>Pipistrellini</i> Tate, 1942
1946	<i>Vansonia</i> Roberts	<i>Vansonia</i> Roberts, 1946

- 1946 *Vansonia* Roberts
 1951 *Rhinomegalophus* Bourret
 1958 *Hesperomyotis* Cabrera
 1959 *Nyctiptennis* Hall and Kelson
 1961 *Afropterus* Lavocat
 1961 *Asellia vetus* Lavocat
 1963 *Myonycterini* Lawrence and Novick
 1964 *Taphozous sudani rhodesiae* Harrison
 1967 *Propotto* Simpson
 1967 *Propotto leakeyi* Simpson
 1968 *Brachipposideros* Sigé
 1970 *Cynopterini* Koopman and J.K. Jones Jr.
 1970 *Hipposiderini* Koopman and J.K. Jones Jr.
 1970 *Rousettina* Koopman and J.K. Jones Jr.
 1971 *Epomophorus* sp. Hill and Morris
 1979 *PHYLLOSTOMATIA* Van Valen
 1979 *VESPERTILIONIA* Van Valen
 1984 *Cheiromelinae* Legendre
 1984 *Rhizomops* Legendre
 1984 *Chamtwarinae* Butler
 1984 *Propottininae* Butler
 1984 *Coleura muthokai* Wesselman
 1985 *Philisidae* Sigé
 1985 *Philisis* Sigé
 1985 *Philisis sphingis* Sigé
 1985 *rupepelli* Qumsiyeh
 1985 *YANGOCHIROPTERA* Koopman
 1985 *Yinochiroptera* Koopman
 1987 *Eptésiformes* Menu
 1987 *Leuconoformes* Menu
 1987 *Leuconoïdes* Menu
 1987 *Néoeptésiformes* Menu
 1987 *Nyctaliformes* Menu
 1987 *Nyctaloïdes* Menu
 1987 *Scotophilini* Hill and Harrison
 1987 *Attalepharca* Menu
 1987 *Nycterikaupius* Menu
 1987 *Nycterikaupius* Menu
 1987 *Nycterikaupius* Menu
 1987 *Nycterikaupius* Menu
 1987 *Nycticeinops* Hill and Harrison
 1988 *Taphozoini* Robbins and Sarich
 1991 *Dizya* Sigé
 1994 *Chiropteriformes* Kinman
 1994 *Dhofarella* Sigé, Thomas, Sen, Gheerbrant, Roger and Al-Sulaimani
 1994 *Hipposiderina* Koopman
 1995 *VESPERTILIFORMES* Zagorodniuk, Godovanets, Pokynchereda and Kyseliuk
 1997 *Plerotini* Bergmans
 1997 *Scotonycterini* Bergmans
 1997 *VESPERTILIFORMES* Zagorodniuk
 1998 *Megachiropteromorpha* Simmons and Geisler
 1998 *Microchiropteraformes* Simmons and Geisler
 1998 *Microchiropteromorpha* Simmons and Geisler
 1998 *VESPERTILIFORMES* Zagorodnyuk
 2000 *Myotis cf. punicus* Castella, Ruedi, Excoffier, Ibanez, Arlettaz and Hausser
 2001 *Pipistrellus cf. kuhlii* Volleth, Bronner, Göpfert, Heller, von Helversen and Yong
 2001 *YINPTEROCHIROPTERA* Springer, Teeling, Madsen, Stanhope and De Jong
 2001 *YINPTEROCHIROPTERA* Springer, Teeling, Madsen, Stanhope and De Jong
 2003 *Indorhinolophus* Guillén-Servent, Francis and Ricklefs
Pipistrellus Kaup, 1829
Rhinolophus Lacépède, 1799
Myotis Kaup, 1829
Eptesicus Rafinesque, 1820
Megaderma E. Geoffroy St.-Hilaire, 1810
Hipposideros vetus (Lavocat, 1961)
Myonycterini Lawrence and Novick, 1963
Taphozous perforatus sudani Thomas, 1915
Propotto Simpson, 1967
Propotto leakeyi Simpson, 1967
Brachipposideros Sigé, 1968
Pteropodinae Gray, 1821
HIPPOSIDERIDAE Lydekker, 1891
Rousettinae Andersen, 1912
Epomophorus labiatus (Temminck, 1837)
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
Molossinae Gervais, 1856
Tadarida Rafinesque, 1814
Chamtwarinae Butler, 1984
Propottininae Butler, 1984
Coleura muthokai Wesselman, 1984
PHILISIDAE Sigé, 1985
Philisis Sigé, 1985
Philisis sphingis Sigé, 1985
Tadarida teniotis (Rafinesque, 1814)
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
CHIROPTERA Blumenbach, 1779
Vespertilioninae Gray, 1821
Myotinae Tate, 1942
Vespertilioninae Gray, 1821
Vespertilioninae Gray, 1821
Vespertilioninae Gray, 1821
Vespertilioninae Gray, 1821
Scotophilinae Van Cakenberghe and Seamark, 2008
Pipistrellus Kaup, 1829
Laephotis Thomas, 1901
Neoromicia Roberts, 1926
Pseudoromicia Monadjem, Patterson, Webala and Demos, 2020
Pipistrellus Kaup, 1829
Nycticeinops Hill & Harrison, 1987
Taphozoinae Jerdon, 1867
Dizya Sigé, 1991
CHIROPTERA Blumenbach, 1779
Dhofarella Sigé, Thomas, Sen, Gheerbrant, Roger and Al-Sulaimani, 1985
HIPPOSIDERIDAE Lydekker, 1891
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
Plerotini Bergmans, 1997
Scotonycterini Bergmans, 1997
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
RHINOLOPHIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
Myotis punicus Felten, 1977
Pipistrellus **(Pipistrellus)** sp. aff. **kuhlii** Kuhl, 1817
VESPERTILIONIFORMI Van Cakenberghe, Kearney and Seamark, 2007
PTEROPODIFORMI Van Cakenberghe, Kearney and Seamark, 2007
Rhinolophus Lacépède, 1799

2003	<i>Tanzanycterididae</i> Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch	<i>TANZANYCTERIDIDAE</i> Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003
2003	<i>Tanzanycteris</i> Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch	<i>Tanzanycteris</i> Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003
2004	<i>PTEROPODIFORMES</i> Hutcheon and Kirsch	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
2004	<i>VESPERTILIONIFORMES</i> Hutcheon and Kirsch	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
2005	<i>Brevipalatus</i> Hand and Archer	<i>Brevipalatus</i> Hand and Archer, 2005
2005	<i>Emballonuroidea</i> Teeling, Springer, Madsen, Bates, O'Brien and Murphy	<i>NYCTEROIDEA</i> Van der Hoeven, 1855
2006	<i>Philisinae</i> Horáček, Fejfar and Hulva	<i>Scotophilinae</i> Van Cakenberghe and Seamark, 2008
2006	<i>Scotophilis</i> Horáček, Fejfar and Hulva	<i>Scotophilis</i> Horáček, Fejfar and Hulva, 2006
2007	<i>NOCTILIONIFORMACEI</i> Van Cakenberghe, Kearney and Seamark	<i>NOCTILIONIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
2007	<i>NYCTERIFORMACEI</i> Van Cakenberghe, Kearney and Seamark	<i>NYCTERIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
2007	<i>PTEROPODIFORMACEI</i> Van Cakenberghe, Kearney and Seamark	<i>PTEROPODIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
2007	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
2007	<i>RHINOLOPHIFORMACEI</i> Van Cakenberghe, Kearney and Seamark	<i>RHINOLOPHIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
2007	<i>VESPERTILIONIFORMACEI</i> Van Cakenberghe, Kearney and Seamark	<i>VESPERTILIONIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
2007	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
2007	<i>Afropipistrellus</i> Thorn, Kock and Cuisin	<i>Nycticeinops</i> Hill & Harrison, 1987
2008	<i>Scotophilinae</i> Van Cakenberghe and Seamark	<i>Scotophilinae</i> Van Cakenberghe and Seamark, 2008
2008	<i>Khonsunyeris</i> Gunnell, Simons and Seiffert	<i>Khonsunyeris</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Pteropoidea</i> Serra-Cobo, López-Roig, Bayer, Amengual and Guasch	<i>PTEROPODOIDEA</i> Gray, 1821
2008	<i>Qarunyeris</i> Gunnell, Simons and Seiffert	<i>Qarunyeris</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Saharaderma</i> Gunnell, Simons and Seiffert	<i>Saharaderma</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Vesperus hösemanni</i> Turni and Kock	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
2008	<i>Witwatia</i> Gunnell, Simons and Seiffert	<i>Witwatia</i> Gunnell, Simons and Seiffert, 2008
2009	<i>Paratriaenops</i> Benda and Vallo	<i>Paratriaenops</i> Benda and Vallo, 2009
2009	<i>Triaenopini</i> Benda and Vallo	<i>RHINONYCTERIDAE</i> J. E. Gray, 1866
2011	<i>Pterocarpus</i> Mishra, Rout and Panda	<i>Pteropus</i> Brisson, 1762
2011	<i>Necromantidae</i> Sigé	<i>NECROMANTIDAE</i> Sigé, 2011
2012	<i>Stenonycterini</i> Nesi, Kadjo, Pourrut, Leroy, Pongombo Shongo, Cruaud and Hassanin	<i>Stenonycterini</i> Nesi, Kadjo, Pourrut, Leroy, Pongombo Shongo, Cruaud and Hassanin, 2012
2012	<i>Paremballonura</i> Goodman, Puechmaile, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling	<i>Paremballonura</i> Goodman, Puechmaile, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012
2012	<i>Paremballonura atrata</i> Goodman, Puechmaile, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling	<i>Paremballonura atrata</i> (Peters, 1874)
2012	<i>Paremballonura tiavato</i> Goodman, Puechmaile, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling	<i>Paremballonura tiavato</i> (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)
2012	<i>Rhinolophus horaceki</i> Benda and Vallo	<i>Rhinolophus horaceki</i> Benda and Vallo, 2012
2013	<i>Niumbaha</i> Reeder, Helgen, Vodzak, Lunde and Ejotre	<i>Glauconycteris</i> Dobson, 1875
2014	<i>Phasmatonycteris</i> Gunnell, Simmons and Seiffert	<i>Phasmatonycteris</i> Gunnell, Simmons and Seiffert, 2014
2014	<i>Pteropodimorpha</i> Zagorodniuk	<i>PTEROPODIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
2014	<i>Vespertilioniformes</i> Zagorodniuk	<i>CHIROPTERA</i> Blumenbach, 1779
2014	<i>Vespertilionimorpha</i> Zagorodniuk	<i>VESPERTILIONIFORMI</i> Van Cakenberghe, Kearney and Seamark, 2007
2015	<i>Nycticeinops serengetiensis</i> Gunnell, Butler, Greenwood and Simmons	<i>Nycticeinops serengetiensis</i> Gunnell, Butler, Greenwood and Simmons, 2015
2016	<i>Aegyptonycteridae</i> Simmons, Seiffert and Gunnell	<i>AEGYPTONYCTERIDAE</i> Simmons, Seiffert and Gunnell, 2016
2016	<i>Aegyptonycteris</i> Simmons, Seiffert and Gunnell	<i>Aegyptonycteris</i> Simmons, Seiffert and Gunnell, 2016
2016	<i>Eidolinae</i> Almeida, Giannini and Simmons	<i>Eidolinae</i> Almeida, Giannini and Simmons, 2016
2016	<i>Chambinycteris</i> Ravel	<i>Chambinycteris</i> Ravel, 2016
2016	<i>Chambinycteris pusilli</i> Ravel	<i>Chambinycteris pusilli</i> Ravel, 2016
2016	<i>Drakonycteris</i> Ravel	<i>Drakonycteris</i> Ravel, 2016
2016	<i>Khoufechia</i> Ravel	<i>Khoufechia</i> Ravel, 2016

2016	<i>Pseudovespertiliavus</i> Ravel	<i>Pseudovespertiliavus</i> Ravel, 2016
2016	<i>Hipposideros cf. caffer</i> Monadjem, Richards and Denys	<i>Hipposideros caffer</i> (Sundevall, 1846)
2016	<i>Hipposideros cf. ruber</i> (lineage C1) Monadjem, Richards and Denys	<i>Hipposideros ruber</i> (Noack, 1893)
2016	<i>Hipposideros cf. ruber</i> (lineage E1) Monadjem, Richards and Denys	<i>Hipposideros ruber</i> (Noack, 1893)
2016	<i>Nycteris cf. parisii</i> Kruskop, Benda, Vasenkov and Lavrenchenko	<i>Nycteris cf. parisii</i> De Beaux, 1924
2017	<i>Pipistrellus krugeri</i> Herkt, Skidmore and Fahr	<i>Laephotis stanleyi</i> (Goodman, Kearney, Ratsimbazafy and Hassanin, 2017)
2017	<i>M[acronycteris] cryptovalorona</i> Foley, Goodman, Whelan, Puechmaille and Teeling	<i>Macronycteris cryptovalorona</i> (Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016)
2018	<i>Turkanycteris</i> Gunnell and Manthi	<i>Turkanycteris</i> Gunnell and Manthi, 2018
2019	<i>Parahypsugo</i> Hutterer, Decher, Monadjem and Astrin	<i>Nycticeinops</i> Hill & Harrison, 1987
2020	<i>Plecotus cf. gaisleri</i> Ancillotto, Bosso, Smeraldo, Mori, Mazza, Herkt, Galimberti, Ramazzotti and Russo	<i>Plecotus gaisleri</i> Benda, Kiefer, Hanak & Veith, 2004
2020	<i>Afronycteris</i> Monadjem, Patterson and Demos	<i>Afronycteris</i> Monadjem, Patterson and Demos, 2020
2020	<i>Nycticeinops bellieri</i> Monadjem, Demos, Dalton, Webala, Musila, Kerbis Peterhans and Patterson	<i>Nycticeinops bellieri</i> (De Vree, 1972)
2020	<i>Pseudoromicia</i> Monadjem, Patterson, Webala and Demos	<i>Pseudoromicia</i> Monadjem, Patterson, Webala and Demos, 2020
2020	<i>Hipposideros cf. ruber</i> (lineage D1) Mamba, Dalton, Mahlaba, Kropff and Monadjem	<i>Hipposideros ruber</i> (Noack, 1893)
2020	<i>Miniopterus cf. inflatus</i> Monadjem, Shapiro, Richards, Karabulut, Crawley, Broman Nielsen, Hansen, Bohmann and Mourier	<i>Miniopterus cf. inflatus</i> Monadjem, Shapiro, Richards, Karabulut, Crawley, Broman Nielsen, Hansen, Bohmann and Mourier, 2020
2020	<i>Mops bemmeleni</i> Simmons and Cirranello	<i>Mops (Chaerephon) bemmeleni</i> (Jentink, 1879)
2020	<i>Mops russatus</i> Simmons and Cirranello	<i>Mops (Chaerephon) russatus</i> (J.A. Allen, 1917)
2021	<i>Afrillonura</i> Rosina and Pickford	<i>Afrillonura</i> Rosina and Pickford, 2021
-330	DERMAPTERA Aristotle	CHIROPTERA Blumenbach, 1779
9999	<i>Epomophorus labiatus anurus</i>	<i>Epomophorus minor</i> Dobson, 1880
9999	<i>Laephotis cf. angolensis</i>	<i>Laephotis angolensis</i> Monard, 1935
9999	<i>Miniopterus</i> sp.	<i>Miniopterus</i> Bonaparte, 1837
9999	<i>Myotis anjouanensis</i>	<i>Myotis anjouanensis</i> Dorst, 1960
9999	<i>Neoromicia "incertae-sedis"</i>	<i>Laephotis "incertae-sedis"</i>
9999	<i>Neoromicia cf. capensis</i>	<i>Laephotis capensis</i> (A. Smith, 1829)
9999	<i>Noctilionoidea</i>	NOCTILIONOIDEA Gray, 1821
9999	<i>Pteropus aegyptiacus</i>	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
9999	<i>Rhinonictidae</i>	RHINONYCTERIDAE J. E. Gray, 1866
9999	<i>Rousettus sp. cf. R. madagascariensis</i>	<i>Rousettus madagascariensis</i> G. Grandidier, 1929
9999	<i>Scotophilus cf. viridis</i>	<i>Scotophilus viridis</i> (Peters, 1852)
9999	<i>Yangochiroptera</i>	VESPERTILIONIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007

"Africa"

1830	<i>Vespertilio marginatus</i> Cretzschmar	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
1857	<i>Vespertilio pusillus</i> Leconte	<i>Laephotis "incertae-sedis"</i>
1862	<i>Epomophorus comptus</i> H. Allen	<i>Epomops franqueti</i> (Tomes, 1860)
1862	<i>H[ypsognathus] monstrosus</i> H. Allen	<i>Hypsognathus monstrosus</i> H. Allen, 1862
1866	<i>Nycterops pilosa</i> Gray	<i>Nycteris hispida</i> (Schreber, 1774)
1866	<i>Nycteris Baikii</i> Gray	<i>Nycteris grandis</i> Peters, 1865
1878	<i>Pteropus rodricensis</i> Dobson	<i>Pteropus rodricensis</i> Dobson, 1878
1879	<i>Nyctinomus Bemmeleni</i> Jentink	<i>Mops (Chaerephon) bemmeleni</i> (Jentink, 1879)
1881	<i>Triænopis rufus</i> A. Milne-Edwards	<i>Triænopis menamena</i> Goodman and Ranivo, 2009
1937	<i>[Pipistrellus Kuhl] latastei</i> Laurent	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)

Algeria

1867	<i>Rhinolophus algirus</i> Loche	<i>Rhinolophus euryale</i> Blasius, 1853
1904	<i>E[uryalus] meridionalis</i> K. Andersen and Matschie	<i>Rhinolophus mehelyi</i> Matschie, 1901
1918	<i>Asellia tridens diluta</i> K. Andersen	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
1934	<i>Rhinolophus acrotis schwarzi</i> Heim de Balsac	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1936	<i>P[ipistrellus] k[uhlii] pallidus</i> Heim de Balsac	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
1936	<i>Plecotus auritus saharæ</i> Laurent	<i>Otonycteris hemprichii</i> Peters, 1859
2016	<i>Drakonycteris glibzegdouensis</i> Ravel	<i>Drakonycteris glibzegdouensis</i> Ravel, 2016
2016	<i>Pseudovespertiliavus parva</i> Ravel	<i>Pseudovespertiliavus parva</i> Ravel, 2016

Angola

1845	<i>Rhinolophus gigas</i> Wagner	<i>Macronycteris gigas</i> (Wagner, 1845)
1866	<i>Scotophilus welwitschii</i> Gray	<i>Myotis welwitschii</i> (Gray, 1866)
1870	<i>[Epomophorus macrocephalus]</i> var. <i>angolensis</i> Gray	<i>Epomophorus angolensis</i> Gray, 1870
1870	<i>Cynopterus collaris</i> Gray	<i>Myonycteris torquata</i> (Dobson, 1878)
1870	<i>Nyctinomus angolensis</i> Peters	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
1870	<i>Vespertilio Bocagii</i> Peters	<i>Myotis bocagii</i> (Peters, 1870)
1870	<i>Vespertilio Bocagii</i> Peters	<i>Myotis bocagii bocagii</i> (Peters, 1870)
1870	<i>Vesperugo pusillulus</i> Peters	<i>Afronycteris nana</i> (Peters, 1852)
1871	<i>Nycteris angolensis</i> Peters	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1878	<i>Cynonycteris torquata</i> Dobson	<i>Myonycteris torquata torquata</i> (Dobson, 1878)
1878	<i>Cynonycteris torquata</i> Dobson	<i>Myonycteris torquata</i> (Dobson, 1878)
1889	<i>Epomophorus Dobsonii</i> Bocage	<i>Epomophorus dobsonii</i> Bocage, 1889
1889	<i>Vesperus bicolor</i> Bocage	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
1898	<i>Cynonycteris Angolensis</i> Bocage	<i>Myonycteris angolensis angolensis</i> (Bocage, 1898)
1898	<i>Cynonycteris Angolensis</i> Bocage	<i>Myonycteris angolensis</i> (Bocage, 1898)
1898	<i>Phyllorhina angolensis</i> Seabra	<i>Hipposideros caffer</i> (Sundevall, 1846)
1898	<i>Rhinolophus angolensis</i> Seabra	<i>Rhinolophus lobatus</i> Peters, 1852
1899	<i>Ep[omophorus (Epomophorus)] zenkeri</i> Matschie	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
1900	<i>Epomophorus anchietæ</i> Seabra	<i>Plerotes anchietæ</i> (Seabra, 1900)
1900	<i>Nyctinomus Anchietæ</i> Seabra	<i>Tadarida aegyptiaca bocagei</i> (Seabra, 1900)
1900	<i>Nyctinomus Bocagei</i> Seabra	<i>Tadarida aegyptiaca bocagei</i> (Seabra, 1900)
1900	<i>Nyctinomus brunneus</i> Seabra	<i>Tadarida aegyptiaca bocagei</i> (Seabra, 1900)
1900	<i>Vesperugo Anchietæ</i> Seabra	<i>Neoromicia anchietæ</i> (Seabra, 1900)
1900	<i>Vesperugo (Vesperus) flavescens</i> Seabra	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
1912	<i>Cistugo seabrae</i> Thomas	<i>Cistugo seabrae</i> Thomas, 1912
1913	<i>Nyctinomus ansorgei</i> Thomas	<i>Mops (Chaerephon) ansorgei</i> (Thomas, 1913)
1933	<i>Mimetillus bernerii</i> Monard	<i>Mimetillus thomasi</i> Hinton, 1920
1933	<i>Nyctinomus spillmanni</i> Monard	<i>Mops (Chaerephon) nigeriae spillmanni</i> (Monard, 1933)
1933	<i>Pipistrellus leucomelas</i> Monard	<i>Vansonia rueppellii vernayi</i> (Roberts, 1932)
1935	<i>Laephotis angolensis</i> Monard	<i>Laephotis angolensis</i> Monard, 1935
1937	<i>Eptesicus capensis angolensis</i> J.Eric Hill	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
1937	<i>Mops chitauensis</i> J.Eric Hill	<i>Mops (Mops) niveiventer</i> Cabrera and Ruxton, 1926
1950	<i>Micropteropus grandis</i> Sanborn	<i>Epomophorus grandis</i> (Sanborn, 1950)
1963	<i>Glauconycteris machadoi</i> Hayman	<i>Glauconycteris machadoi</i> Hayman, 1963
1963	<i>Micropteropus intermedius</i> Hayman	<i>Epomophorus intermedius</i> (Hayman, 1963)

Azores (Portugal)

1901	<i>Pterygistes azureum</i> Thomas	<i>Nyctalus azureum</i> (Thomas, 1901)

Benin

1899	<i>Ep[omophorus (Epomophorus)] zechi</i> Matschie	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)

Botswana

1932	<i>Choerephon (Lophomops) langi</i> Roberts	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1932	<i>Eptesicus capensis nkatiensis</i> Roberts	<i>Laephotis capensis</i> (A. Smith, 1829)
1932	<i>Neoromicia vansonii</i> Roberts	<i>Neoromicia zuluensis</i> (Roberts, 1924)
1932	<i>Pipistrellus vernayi</i> Roberts	<i>Vansonia rueppellii vernayi</i> (Roberts, 1932)
1932	<i>Scoteinus schlieffenii fitzsimonsi</i> Roberts	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
1971	<i>Laephotis botswanae</i> Setzer	<i>Laephotis botswanae</i> Setzer, 1971
2017	<i>Neoromicia stanleyi</i> Goodman, Kearney, Ratsimbazafy and Hassanin	<i>Laephotis stanleyi</i> (Goodman, Kearney, Ratsimbazafy and Hassanin, 2017)

Cameroon

1875	<i>Chalinolobus argentatus</i> Dobson	<i>Glauconycteris argentata</i> (Dobson, 1875)
1894	<i>Scotonycteris zenkeri</i> Matschie	<i>Scotonycteris zenkeri</i> Matschie, 1894
1903	<i>Miniopterus inflatus</i> Thomas	<i>Miniopterus inflatus</i> Thomas, 1903
1903	<i>Miniopterus inflatus</i> Thomas	<i>Miniopterus inflatus inflatus</i> Thomas, 1903
1903	<i>Nycteris arge</i> Thomas	<i>Nycteris arge</i> Thomas, 1903
1903	<i>Nyctinomus thersites</i> Thomas	<i>Mops (Xiphonycteris) thersites</i> (Thomas, 1903)
1904	<i>Pipistrellus crassulus</i> Thomas	<i>Nycticeinops crassulus</i> (Thomas, 1904)
1904	<i>Scotophilus nigrita nux</i> Thomas	<i>Scotophilus nux</i> Thomas, 1904
1904	<i>Myotis Bocagei cupreolus</i> Thomas	<i>Myotis bocagii cupreolus</i> Thomas, 1904
1904	<i>Pipistrellus nanulus</i> Thomas	<i>Pipistrellus (Pipistrellus) nanulus</i> Thomas, 1904
1906	<i>Kerivoula muscilla</i> Thomas	<i>Kerivoula lanosa</i> (A. Smith, 1847)

1910	<i>Casinonycteris argynnis</i> Thomas	<i>Casinonycteris argynnis</i> Thomas, 1910
1912	<i>Kerivoula cuprosa</i> Thomas	<i>Kerivoula cuprosa</i> Thomas, 1912
1912	<i>Petalia major</i> K. Andersen	<i>Nycteris major</i> (K. Andersen, 1912)
1913	<i>Glauconycteris egeria</i> Thomas	<i>Glauconycteris egeria</i> Thomas, 1913
1913	<i>Pipistrellus musciculus</i> Thomas	<i>Hypsugo musciculus</i> (Thomas, 1913)
1921	<i>Hipposideros curtus</i> G.M. Allen	<i>Hipposideros curtus</i> G.M. Allen, 1921
1936	<i>Rhinolophus alcyone alticolus</i> Sanborn	<i>Rhinolophus simulator</i> K. Andersen, 1904
1937	<i>Hipposideros sandersoni</i> Sanderson	<i>Hipposideros curtus</i> G.M. Allen, 1921
1943	<i>Scotonycteris ophiodon</i> Pohle	<i>Casinonycteris ophiodon</i> (Pohle, 1943)
1950	<i>Epomophorus reii</i> Aellen	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
1950	<i>Epomophorus reii</i> Aellen	<i>Epomophorus labiatus</i> (Temminck, 1837)
1952	<i>Nycteris benuensis</i> Aellen	<i>Nycteris parisii</i> (de Beaux, 1924)
1956	<i>Hipposideros camerunensis</i> Eisentraut	<i>Doryrhina camerunensis</i> (Eisentraut, 1956)
1959	<i>Pipistrellus inexpectatus</i> Aellen	<i>Pipistrellus (Pipistrellus) inexpectatus</i> Aellen, 1959
1960	<i>Rousettus aegyptiacus occidentalis</i> Eisentraut	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1968	<i>Pipistrellus eisentrauti</i> Hill	<i>Nycticeinops eisentrauti</i> (Hill, 1968)
1973	<i>Glauconycteris gleni</i> Peterson and Smith	<i>Glauconycteris gleni</i> Peterson and Smith, 1973
1981	<i>Tadarida [(Xiphonycteris)] petersoni</i> El Rayah	<i>Mops (Xiphonycteris) petersoni</i> (El Rayah, 1981)
2001	<i>Glauconycteris curryi</i> Eger and Schlitter	<i>Glauconycteris curryae</i> Eger and Schlitter, 2001
2014	<i>Casinonycteris campomaanensis</i> Hassanin	<i>Casinonycteris campomaanensis</i> Hassanin, 2014

Canary Islands (Spain)

1859	<i>Scotophilus darwini</i> Tomes	<i>Hypsugo savii</i> (Bonaparte, 1837)
1907	<i>Plecotus teneriffae</i> Barrett-Hamilton	<i>Plecotus teneriffae</i> Barrett-Hamilton, 1907
2002	<i>Barbastella barbastellus guanchae</i> Trujillo, Ibáñez and Juste	<i>Barbastella barbastellus</i> (Schreber, 1774)

Central African Republic

1904	<i>Epomophorus pousarguesi</i> Trouessart	<i>Epomophorus gambianus pousarguesi</i> Trouessart, 1904
2015	<i>Scotonycteris bergmansi</i> Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo	<i>Scotonycteris bergmansi</i> Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015
2015	<i>Scotonycteris bergmansi bergmansi</i> Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo	<i>Scotonycteris bergmansi bergmansi</i> Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015

Comoros

1866	<i>Pteropus livingstonii</i> Gray	<i>Pteropus livingstonii</i> Gray, 1866
1880	<i>Pteropus comorensis</i> Wallace	<i>Pteropus seychellensis</i> A. Milne-Edwards, 1877
1959	<i>Miniopterus minor griveaudi</i> Harrison	<i>Miniopterus griveaudi</i> Harrison, 1959
1960	<i>Myotis Goudoti anjouanensis</i> Dorst	<i>Myotis anjouanensis</i> Dorst, 1960
1978	<i>Rousettus (Rousettus) obliviosus</i> Kock	<i>Rousettus obliviosus</i> Kock, 1978

Congo

1912	<i>Petalia nana</i> K. Andersen	<i>Nycteris nana</i> (K. Andersen, 1912)
1968	<i>Rhinolophus adami</i> Aellen and Brosset	<i>Rhinolophus adami</i> Aellen and Brosset, 1968
1968	<i>Triaenops persicus majusculus</i> Aellen and Brosset	<i>Triaenops afer</i> Peters, 1877
1992	<i>Miniopterus minor occidentalis</i> Juste and Ibáñez	<i>Miniopterus minor</i> Peters, 1867
2005	<i>Myotis dieteri</i> M. Happold	<i>Myotis dieteri</i> M. Happold, 2005

Congo (Democratic Republic of the)

1887	<i>Phyllorhina commersonii</i> var. <i>marungensis</i> Noack	<i>Macronycteris vittatus</i> (Peters, 1852)
1889	<i>Chalinolobus conigicus</i> Noack	<i>Glauconycteris argentata</i> (Dobson, 1875)
1889	<i>Vesperugo pagenstecheri</i> Noack	<i>Afronycteris nana</i> (Peters, 1852)
1889	<i>Vesperus pusillus</i> Noack	<i>Afronycteris nana</i> (Peters, 1852)
1908	<i>Myonycteris wroughtoni</i> K. Andersen	<i>Myonycteris torquata wroughtoni</i> K. Andersen, 1908
1915	<i>Myopterus albatrus</i> Thomas	<i>Myopterus daubentonii</i> Desmarest, 1820
1917	<i>Chærephon (Lophomops) abæ</i> J.A. Allen	<i>Mops (Chærephon) major</i> (Trouessart, 1897)
1917	<i>Chærephon (Lophomops) chapini</i> J.A. Allen	<i>Mops (Chærephon) chapini</i> (J.A. Allen, 1917)
1917	<i>Chærephon (Lophomops) cristatus</i> J.A. Allen	<i>Mops (Chærephon) leucogaster</i> (A. Grandidier, 1869)
1917	<i>Chærephon frater</i> J.A. Allen	<i>Mops (Chærephon) leucogaster</i> (A. Grandidier, 1869)
1917	<i>Chærephon russatus</i> J.A. Allen	<i>Mops (Chærephon) russatus</i> (J.A. Allen, 1917)
1917	<i>Eptesicus ater</i> J.A. Allen	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)

1917	<i>Eptesicus faradjius</i> J.A. Allen	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
1917	<i>Eptesicus garambæ</i> J.A. Allen	<i>Laephotis capensis</i> (A. Smith, 1829)
1917	<i>Glauconycteris alboguttatus</i> J.A. Allen	<i>Glauconycteris alboguttata</i> J.A. Allen, 1917
1917	<i>Glauconycteris humeralis</i> J.A. Allen	<i>Glauconycteris humeralis</i> J.A. Allen, 1917
1917	<i>Hipposideros abae</i> J.A. Allen	<i>Hipposideros abae</i> J.A. Allen, 1917
1917	<i>Hipposideros caffer niapu</i> J.A. Allen	<i>Hipposideros ruber</i> (Noack, 1893)
1917	<i>Hipposideros gigas niangaræ</i> J.A. Allen	<i>Macronycteris gigas</i> (Wagner, 1845)
1917	<i>Hipposideros langi</i> J.A. Allen	<i>Doryrhina cyclops</i> (Temminck, 1853)
1917	<i>Hipposideros nanus</i> J.A. Allen	<i>Hipposideros caffer</i> (Sundevall, 1846)
1917	<i>Miniopterus breyeri vicinior</i> J.A. Allen	<i>Miniopterus fraterculus</i> Thomas and Schwann, 1906
1917	<i>Mops (Allomops) faradjius</i> J.A. Allen	<i>Mops (Mops) demonstrator</i> (Thomas, 1903)
1917	<i>Mops (Allomops) nanulus</i> J.A. Allen	<i>Mops (Xiphonycteris) nanulus</i> J.A. Allen, 1917
1917	<i>Mops (Allomops) occipitalis</i> J.A. Allen	<i>Mops (Xiphonycteris) thersites</i> (Thomas, 1903)
1917	<i>Mops (Allomops) osborni</i> J.A. Allen	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
1917	<i>Mops congicus</i> J.A. Allen	<i>Mops (Mops) congicus</i> J.A. Allen, 1917
1917	<i>Mops niangaræ</i> J.A. Allen	<i>Mops (Mops) niangaræ</i> J.A. Allen, 1917
1917	<i>Mops trevori</i> J.A. Allen	<i>Mops (Mops) trevori</i> J.A. Allen, 1917
1917	<i>Nycteris avakubia</i> J.A. Allen	<i>Nycteris major</i> (K. Andersen, 1912)
1917	<i>Nycteris pallida</i> J.A. Allen	<i>Nycteris hispida</i> (Schreber, 1774)
1917	<i>Nyctinomus ochraceus</i> J.A. Allen	<i>Mops (Xiphonycteris) brachypterus</i> (Peters, 1852)
1917	<i>Pipistrellus abaensis</i> J.A. Allen	<i>Afronycteris nana</i> (Peters, 1852)
1917	<i>Rhinolophus abae</i> J.A. Allen	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1917	<i>Rhinolophus axillaris</i> Allen	<i>Rhinolophus landeri</i> Martin, 1838
1925	<i>Nycteris proxima</i> Lönnberg and Gyldestolpe	<i>Nycteris grandis</i> Peters, 1865
1926	<i>Mops angolensis niveiventer</i> Cabrera and Ruxton	<i>Mops (Mops) niveiventer</i> Cabrera and Ruxton, 1926
1936	<i>Miniopterus rufus</i> Sanborn	<i>Miniopterus inflatus rufus</i> Sanborn, 1936
1939	<i>Glauconycteris superba</i> Hayman	<i>Glauconycteris superba</i> Hayman, 1939
1942	<i>Rhinolophus ruwenzorii</i> J. Eric Hill	<i>Rhinolophus ruwenzorii</i> J. Eric Hill, 1942
1951	<i>Tadarida (Chaerephon) faini</i> Hayman	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1957	<i>Hipposideros beatus maximus</i> Verschuren	<i>Hipposideros beatus</i> (K. Andersen, 1906)
1966	<i>Megaloglossus woermanni prigoginei</i> Hayman	<i>Megaloglossus woermanni</i> Pagenstecher, 1885
1975	<i>Tadarida (Chaerephon) gallagheri</i> Harrison	<i>Mops (Chaerephon) gallagheri</i> (Harrison, 1975)
2013	<i>Rhinolophus kahuzi</i> Fahr and Kerbis Peterhans	<i>Rhinolophus kahuzi</i> Fahr and Kerbis Peterhans, 2013
2013	<i>Rhinolophus willardi</i> Kerbis Peterhans and Fahr	<i>Rhinolophus willardi</i> Kerbis Peterhans and Fahr, 2013
2015	<i>Scotonycteris bergmansi congoensis</i> Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo	<i>Scotonycteris bergmansi congoensis</i> Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015
2017	<i>Glauconycteris atra</i> Hassanin, Colombo, Gembu, Merle, Tu, Görföl, Musaba Akawa, Csorba, Kearney, Monadjem and Ing	<i>Glauconycteris atra</i> Hassanin, Colombo, Gembu, Merle, Tu, Görföl, Musaba Akawa, Csorba, Kearney, Monadjem and Ing, 2017
2020	<i>Parahypsugo macrocephalus</i> Hutterer and Kerbis Peterhans	<i>Nycticeinops macrocephalus</i> (Hutterer and Kerbis Peterhans, 2020)

Côte d'Ivoire

1954	<i>Hipposideros marisae</i> Aellen	<i>Hipposideros marisae</i> Aellen, 1954
1959	<i>Kerivoula harrisoni bellula</i> Aellen	<i>Kerivoula lanosa</i> (A. Smith, 1847)
1959	<i>Nycteris intermedia</i> Aellen	<i>Nycteris intermedia</i> Aellen, 1959
1972	<i>Pipistrellus eisentrauti bellieri</i> De Vree	<i>Nycticeinops bellieri</i> (De Vree, 1972)
2012	<i>Megaloglossus azagnyi</i> Nesi, Kadjo and Hassanin	<i>Megaloglossus azagnyi</i> Nesi, Kadjo and Hassanin, 2012

Croatia

1839	<i>Vespertilio pachygnathus</i> Michahelles	<i>Nyctalus leisleri</i> (Kuhl, 1817)
1980	<i>Plecotus austriacus kolombatovici</i> Dulic	<i>Plecotus kolombatovici</i> Dulic, 1980

Egypt

1782	<i>Vespertilio Microphyllum</i> Brünnich	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
1810	<i>Pteropus Egyptiacus</i> E. Geoffroy Saint-Hilaire	<i>Roussettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1813	<i>Rhinolophus tridens</i> E. Geoffroy Saint-Hilaire	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
1816	<i>Rhinopoma brevicaudatum</i> Oken	<i>Rhinopoma cystops</i> Thomas, 1903
1818	<i>Nycteris Thebaicus</i> E. Geoffroy Saint-Hilaire	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1818	<i>Nyctinomus aegyptiacus</i> E. Geoffroy Saint-Hilaire	<i>Tadarida aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)

1818	<i>Nyctinomus aegyptiacus</i> E. Geoffroy Saint-Hilaire	<i>Tadarida aegyptiaca aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
1818	<i>Taphozous perforatus</i> E. Geoffroy Saint-Hilaire	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
1818	<i>Taphozous perforatus</i> E. Geoffroy Saint-Hilaire	<i>Taphozous perforatus perforatus</i> E. Geoffroy St.-Hilaire, 1818
1827	<i>Dysopes rüpelii</i> Temminck	<i>Tadarida teniotis</i> (Rafinesque, 1814)
1829	<i>[Vespertilio pipistrellus var.] β Aegyptius</i> J.B. Fischer	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
1830-1831	<i>Taphozous nudiventris</i> Cretzschmar	<i>Taphozous nudiventris</i> Cretzschmar, 1830
1838	<i>Plecotus christii</i> Gray	<i>Plecotus christii</i> Gray, 1838
1858	<i>Nannugo Kolenatii</i> Müller	<i>Pipistrellus (Pipistrellus) "incertae-sedis"</i>
1859	<i>Nycticejus Schlieffenii</i> Peters	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
1859	<i>Otonycteris Hemprichii</i> Peters	<i>Otonycteris hemprichii</i> Peters, 1859
1878	<i>Plecotus aegyptiacus</i> Dobson	<i>Plecotus christii</i> Gray, 1838
1887	<i>Vesperugo (Vesperus) Innesi</i> Lataste	<i>Eptesicus (Cnephaeus) bottae</i> (Peters, 1869)
1903	<i>Rhinopoma cystops</i> Thomas	<i>Rhinopoma cystops</i> Thomas, 1903
1904	<i>Rhinolophus Andersoni</i> Thomas	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1905	<i>Rhinolophus acrotis brachygnatus</i> K. Andersen	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1910	<i>Vampyravus orientalis</i> Schlosser	<i>Vampyravus orientalis</i> Schlosser, 1910
1954	<i>Vespertilio brevicauda</i> Stresemann	<i>Rhinopoma cystops</i> Thomas, 1903
1954	<i>Vespertilio ferox</i> Stresemann	<i>Rhinopoma cystops</i> Thomas, 1903
2008	<i>Dhofarella sigei</i> Gunnell, Simons and Seiffert	<i>Dhofarella sigei</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Khonsunyeris aegypticus</i> Gunnell, Simons and Seiffert	<i>Khonsunyeris aegypticus</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Plecotus christii petraeus</i> Benda	<i>Plecotus christii</i> Gray, 1838
2008	<i>Qarunyeris moerisae</i> Gunnell, Simons and Seiffert	<i>Qarunyeris moerisae</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Saharaderma pseudovampyrus</i> Gunnell, Simons and Seiffert	<i>Saharaderma pseudovampyrus</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Witwatia eremicus</i> Gunnell, Simons and Seiffert	<i>Witwatia eremicus</i> Gunnell, Simons and Seiffert, 2008
2008	<i>Witwatia schlosseri</i> Gunnell, Simons and Seiffert	<i>Witwatia schlosseri</i> Gunnell, Simons and Seiffert, 2008
2014	<i>Phasmatonycteris butleri</i> Gunnell, Simmons and Seiffert	<i>Phasmatonycteris butleri</i> Gunnell, Simmons and Seiffert, 2014
2014	<i>Phasmatonycteris phiomensis</i> Gunnell, Simmons and Seiffert	<i>Phasmatonycteris phiomensis</i> Gunnell, Simmons and Seiffert, 2014
2015	<i>Hipposideros (Pseudorhinolophus) amehotepos</i> Gunnell, Winkler, Miller, Head, El-Barkooky, Gawad, Sanders and Gingerich	<i>Hipposideros amehotepos</i> Gunnell, Winkler, Miller, Head, El-Barkooky, Gawad, Sanders and Gingerich, 2015
2016	<i>Aegyptonycteris knightae</i> Simmons, Seiffert and Gunnell	<i>Aegyptonycteris knightae</i> Simmons, Seiffert and Gunnell, 2016

Equatorial Guinea

1838	<i>Rhinolophus landeri</i> Martin	<i>Rhinolophus landeri</i> Martin, 1838
1843	<i>Nycteris poensis</i> Gray	<i>Nycteris hispida</i> (Schreber, 1774)
1843	<i>Rhinolophus Martini</i> Fraser	<i>Nycteris hispida</i> (Schreber, 1774)
1904	<i>Scotonycteris bedfordi</i> Thomas	<i>Scotonycteris zenkeri</i> Matschie, 1894
1906	<i>Hipposiderus beatus</i> K. Andersen	<i>Hipposideros beatus</i> (K. Andersen, 1906)
1920	<i>Rousettus (Lissonycteris) crypticola</i> Cabrera	<i>Myonycteris angolensis</i> (Bocage, 1898)
2000	<i>Eidolon helvum annobonensis</i> Juste, Ibáñez and Machordom	<i>Eidolon helvum helvum</i> (Kerr, 1792)

Eritrea

1826	<i>Dysopes pumilus</i> Cretzschmar	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1840	<i>Vespertilio hesperida</i> Temminck	<i>Pipistrellus (Pipistrellus) hesperidus hesperidus</i> (Temminck, 1840)
1840	<i>Vespertilio hesperida</i> Temminck	<i>Pipistrellus (Pipistrellus) hesperidus</i> (Temminck, 1840)
1861	<i>Nycteris labiata</i> Heuglin	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1861	<i>Nycticejus flavigaster</i> Heuglin	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
1861	<i>Nyctinomus (Dysopes) ventralis</i> Heuglin	<i>Tadarida ventralis</i> (Heuglin, 1861)
1861	<i>Nyctinomus bivittatus</i> Heuglin	<i>Mops (Chaerephon) bivittatus</i> (Heuglin, 1861)
1861	<i>Phyllorhina bicornis</i> Heuglin	<i>Hipposideros tephros</i> (Cabrera, 1906)
1861	<i>Phyllorhina megalotis</i> Heuglin	<i>Hipposideros megalotis</i> (Heuglin, 1861)
1861	<i>Rhinolophus acrotis</i> Heuglin	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1861	<i>Rhinolophus miminus</i> Heuglin	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1885	<i>Taphozous perforatus</i> var. <i>assabensis</i> Monticelli	<i>Taphozous nudiventris</i> Cretzschmar, 1830

1899	<i>Ep[omophorus (Epomophorus)] doriae</i> Matschie	<i>Epomophorus labiatus</i> (Temminck, 1837)
1905	<i>Rhinolophus andreinii</i> Senna	<i>Rhinolophus blasii</i> Peters, 1867
1931	<i>Asellia patrizii</i> de Beaux	<i>Asellia patrizii</i> de Beaux, 1931

Ethiopia

1842	<i>Pteropus Schoënsis</i> Rüppell	<i>Epomophorus labiatus</i> (Temminck, 1837)
1842	<i>Rhinolophus fumigatus</i> Rüppell	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1842	<i>Vespertilio Pipistrellus varietas africana</i> Rüppell	<i>Afronycteris nana</i> (Peters, 1852)
1861	<i>Nycticejus murino-flavus</i> Heuglin	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
1872	<i>Megaderma cor</i> Peters	<i>Cardioderma cor</i> (Peters, 1872)
1877	<i>N[ycticejus] adovanus</i> Heuglin	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
1877	<i>Nycticejus eriophorus</i> Heuglin	<i>Kerivoula eriophora</i> (Heuglin, 1877)
1877	<i>Rhinolophus macrocephalus</i> Heuglin	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1885	<i>Rhinolophus antinorii</i> Dobson	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1901	<i>Kerivoula harrisoni</i> Thomas	<i>Kerivoula lanosa</i> (A. Smith, 1847)
1906	<i>Platymops Macmillani</i> Thomas	<i>Platymops setiger macmillani</i> Thomas, 1906
1912	<i>Petalia damarensis media</i> K. Andersen	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1927	<i>Myotis scotti</i> Thomas	<i>Myotis scotti</i> Thomas, 1927
1936	<i>Miniopterus africanus</i> Sanborn	<i>Miniopterus africanus</i> Sanborn, 1936
1971	<i>Myotis morrisi</i> Hill	<i>Myotis morrisi</i> Hill, 1971
1984	<i>Hipposideros kaumbului</i> Wesselman	<i>Hipposideros kaumbului</i> Wesselman, 1984
1984	<i>Taphozous abitus</i> Wesselman	<i>Saccolaimus abitus</i> (Wesselman, 1984)
1991	<i>Epomophorus minimus</i> Claessen and De Vree	<i>Epomophorus minimus</i> Claessen & De Vree, 1991
1997	<i>Lissonycteris angolensis petraea</i> Bergmans	<i>Myonycteris angolensis petraea</i> (Bergmans, 1997)
2000	<i>Plecotus balensis</i> Kruskop and Lavrenchenko	<i>Plecotus balensis</i> Kruskop & Lavrenchenko, 2000
2014	<i>Scotophilus ejetai</i> Brooks and Bickham	<i>Scotophilus ejetai</i> Brooks and Bickham, 2014
2015	<i>Otomops harrisoni</i> Ralph, Richards, Taylor, Napier and Lamb	<i>Otomops harrisoni</i> Ralph, Richards, Taylor, Napier and Lamb, 2015

France

1774	<i>Vespertilio barbastellus</i> Schreber	<i>Barbastella barbastellus</i> (Schreber, 1774)
1774	<i>Vespertilio ferrum-equinum</i> Schreber	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
1774	<i>Vespertilio pipistrellus</i> Schreber	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
1792	<i>minor</i> Kerr	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1792	<i>Vespertilio ferrum-equinum major</i> Kerr	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
1800	<i>Vespertilio hipposideros</i> Bechstein	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1806	<i>Vespertilio emarginatus</i> E. Geoffroy Saint-Hilaire	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
1898	<i>Miomegaderma gaillardi</i> Trouessart	<i>Megaderma gaillardi</i> (Trouessart, 1898)
1904	<i>E[uryalus] atlanticus</i> K. Andersen and Matschie	<i>Rhinolophus euryale</i> Blasius, 1853

Gabon

1860	<i>Epomophorus franqueti</i> Tomes	<i>Epomops franqueti</i> (Tomes, 1860)
1862	<i>Pteropus mollipilosus</i> H. Allen	<i>Eidolon helvum helvum</i> (Kerr, 1792)
1870	<i>Eleutherura unicolor</i> Gray	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1875	<i>Vespertilio murinus africanus</i> Dobson	<i>Myotis punicus</i> Felten, 1977
1876	<i>Epomophorus macrocephalus</i> Peters	<i>Hypsignathus monstrosus</i> H. Allen, 1862
1885	<i>Megaloglossus woermanni</i> Pagenstecher	<i>Megaloglossus woermanni</i> Pagenstecher, 1885
1897	<i>Rhinolophus micaceus</i> de Winton	<i>Doryrhina cyclops</i> (Temminck, 1853)
1897	<i>[Vesperugo (Vesperus) serotinus] Var. Gabonensis</i> Trouessart	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
1906	<i>Hipposideros caffer guineensis</i> K. Andersen	<i>Hipposideros ruber</i> (Noack, 1893)
1959	<i>Rhinolophus silvestris</i> Aellen	<i>Rhinolophus silvestris</i> Aellen, 1959
1966	<i>Tadarida (Chaerephon) cyclotis</i> Brosset	<i>Mops (Chaerephon) aloysiisabaudiae</i> (Festa, 1907)

Germany

1817	<i>Vespertilio Leisleri</i> Kuhl	<i>Nyctalus leisleri</i> (Kuhl, 1817)
1817	<i>Vespertilio mystacinus</i> Kuhl	<i>Myotis mystacinus</i> (Kuhl, 1817)
1827	<i>Vespertilio ferrugineus</i> Brehm	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
1983	<i>Tadarida engesseri</i> Rachl	<i>Tadarida engesseri</i> Rachl, 1983

Ghana

1853	<i>Phyllorhina cyclops</i> Temminck	<i>Doryrhina cyclops</i> (Temminck, 1853)
1853	<i>Phyllorhina fuliginosa</i> Temminck	<i>Hipposideros fuliginosus</i> (Temminck, 1853)

1853	<i>Rhinolophus alcyone</i> Temminck	<i>Rhinolophus alcyone</i> Temminck, 1853
1853	<i>Taphozous peli</i> Temminck	<i>Saccolaimus peli</i> (Temminck, 1853)
1865	<i>Nycteris grandis</i> Peters	<i>Nycteris grandis</i> Peters, 1865
1899	<i>Scotophilus hirundo</i> de Winton	<i>Scotoecus hirundo</i> (de Winton, 1899)
1899	<i>Scotophilus nigrnellus</i> de Winton	<i>Scotophilus nigrnellus</i> de Winton, 1899
1911	<i>Xiphonycteris spurrelli</i> Dollman	<i>Mops (Xiphonycteris) spurrelli</i> (Dollman, 1911)
1912	<i>Kerivoula phalaena</i> Thomas	<i>Kerivoula phalaena</i> Thomas, 1912
1946	<i>Scotoonycteris ophiodon cansdalei</i> Hayman	<i>Casinycteris ophiodon</i> (Pohle, 1943)
1947	<i>Scotoonycteris zenkeri occidentalis</i> Hayman	<i>Scotoonycteris occidentalis</i> Hayman, 1947
1947	<i>Glauconycteris superba sheila</i> Hayman	<i>Glauconycteris superba</i> Hayman, 1939
1983	<i>Scotophilus nucella</i> Robbins	<i>Scotophilus nucella</i> Robbins, 1983

Guinea

1872	<i>Vesperus tenuipinnis</i> Peters	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
1898	<i>Rhinolophus maclaudi</i> Pousargues	<i>Rhinolophus maclaudi</i> Pousargues, 1898
1956	<i>Miniopterus inflatus villiersi</i> Aellen	<i>Miniopterus villiersi</i> Aellen, 1956
1960	<i>Rhinolophus denti knorri</i> Eisentraut	<i>Rhinolophus denti</i> Thomas, 1904
1960	<i>Rhinolophus landeri guineensis</i> Eisentraut	<i>Rhinolophus guineensis</i> Eisentraut, 1960
1985	<i>Hipposideros lamottei</i> Brosset	<i>Hipposideros lamottei</i> Brosset, 1985
2002	<i>Rhinolophus ziama</i> Fahr, Vierhaus, Hutterer and Kock	<i>Rhinolophus ziama</i> Fahr, Vierhaus, Hutterer and Kock, 2002
2016	<i>Neoromicia isabella</i> Decher, Hutterer and Monadjem	<i>Pseudoromicia isabella</i> (Decher, Hutterer and Monadjem, 2016)
2019	<i>Parahypsugo happoldorum</i> Hutterer, Decher, Monadjem and Astrin	<i>Nycticeinops happoldorum</i> (Hutterer, Decher, Monadjem and Astrin, 2019)
2020	<i>Pipistrellus simandouensis</i> Monadjem, Richards, Decher and Hutterer	<i>Pipistrellus (Pipistrellus) simandouensis</i> Monadjem, Richards, Decher and Hutterer, 2020

Guinea-Bissau

1889	<i>Vesperus guineensis</i> Bocage	<i>Neoromicia guineensis</i> (Bocage, 1889)
1898	<i>Epomophorus guineensis</i> Bocage	<i>Epomophorus gambianus</i> (Ogilby, 1835)
1898	<i>Epomophorus guineensis</i> Bocage	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
1939	<i>Coleura kummeri</i> Monard	<i>Coleura afra</i> (Peters, 1852)
1939	<i>Hipposideros braima</i> Monard	<i>Hipposideros tephros</i> (Cabrera, 1906)
1939	<i>Hipposideros gigas viegasi</i> Monard	<i>Macronycteris gigas</i> (Wagner, 1845)
1939	<i>Mops osborni occidentalis</i> Monard	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
1939	<i>Mops osborni occidentalis f. fulva</i> Monard	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
1939	<i>Nycteris aethiopica guineensis</i> Monard	<i>Nycteris macrotis</i> Dobson, 1876
1939	<i>Nycteris aethiopica guineensis f. aurantiaca</i> Monard	<i>Nycteris macrotis</i> Dobson, 1876

Iraq

1902	<i>Otonycteris petersi</i> Anderson	<i>Otonycteris hemprichii</i> Peters, 1859

Italy

1780	<i>Vespertilio lasiopterus</i> Schreber	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
1814	<i>Cephalotes teniotis</i> Rafinesque	<i>Tadarida teniotis</i> (Rafinesque, 1814)
1817	<i>Vespertilio Kuhlii</i> Kuhl	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
1825	<i>Dinops Cestonii</i> Savi	<i>Tadarida teniotis</i> (Rafinesque, 1814)
1837	<i>Vespertilio Capaccinii</i> Bonaparte	<i>Myotis capaccinii</i> (Bonaparte, 1837)
1837	<i>Vespertilio Savii</i> Bonaparte	<i>Hypsugo savii</i> (Bonaparte, 1837)
1853	<i>Rhinolophus euryale</i> Blasius	<i>Rhinolophus euryale</i> Blasius, 1853
1867	<i>Rhinolophus blasii</i> Peters	<i>Rhinolophus blasii</i> Peters, 1867
1868	<i>V[espertilio noctula] var. sicula</i> Mina-Palumbo	<i>Nyctalus lasiopterus</i> (Schreber, 1780)

Kenya

1877	<i>Triænonops afer</i> Peters	<i>Triænonops afer</i> Peters, 1877
1878	<i>Mormopterus setiger</i> Peters	<i>Platymops setiger</i> (Peters, 1878)
1878	<i>Mormopterus setiger</i> Peters	<i>Platymops setiger setiger</i> (Peters, 1878)
1878	<i>Rhinolophus Hildebrandtii</i> Peters	<i>Rhinolophus hildebrandtii</i> Peters, 1878
1891	<i>Nyctinomus lobatus</i> Thomas	<i>Tadarida lobata</i> (Thomas, 1891)
1899	<i>Ep[omophorus (Epomophorus)] neumanni</i> Matschie	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
1901	<i>Scotoecus Hindei</i> Thomas	<i>Scotoecus hindei</i> Thomas, 1901
1901	<i>Læphotis Wintoni</i> Thomas	<i>Læphotis wintoni</i> Thomas, 1901
1901	<i>Cloetis Percivali</i> Thomas	<i>Cloetis percivali</i> Thomas, 1901
1901	<i>Nycteris aethiopica luteola</i> Thomas	<i>Nycteris macrotis</i> Dobson, 1876

1901	<i>Pipistrellus Kuhlī fuscatus</i> Thomas	<i>Pipistrellus (Pipistrellus) hesperidus hesperidus</i> (Temminck, 1840)
1904	<i>Myotis Hildegardae</i> Thomas	<i>Myotis bocagii bocagii</i> (Peters, 1870)
1904	<i>Nyctinomus hindei</i> Thomas	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1904	<i>Scotophilus nigrīta colias</i> Thomas	<i>Scotophilus dinganii</i> (A. Smith, 1833)
1904	<i>Hipposideros Commersoni mostellum</i> Thomas	<i>Macronycteris vittatus</i> (Peters, 1852)
1905	<i>Rhinolophus fumigatus exsul</i> K. Andersen	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1905	<i>Lavia rex</i> Miller	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
1909	<i>Taphozous hildegardae</i> Thomas	<i>Taphozous hildegardae</i> Thomas, 1909
1909	<i>Rousettus kempi</i> Thomas	<i>Stenonycteris lanosus</i> (Thomas, 1906)
1909	<i>Scotoecus albigula</i> Thomas	<i>Scotoecus albigula</i> Thomas, 1909
1911	<i>Eptesicus phasma</i> G.M. Allen	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
1911	<i>Nycticeius africanus</i> G.M. Allen	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
1912	<i>Petalia aurita</i> K. Andersen	<i>Nycteris aurita</i> (K. Andersen, 1912)
1912	<i>Miniopterus natalensis arenarius</i> Heller	<i>Miniopterus arenarius</i> Heller, 1912
1912	<i>Pipistrellus aero</i> Heller	<i>Pipistrellus (Pipistrellus) aero</i> Heller, 1912
1912	<i>Pipistrellus helios</i> Heller	<i>Afronycteris helios</i> (Heller, 1912)
1915	<i>Taphozous perforatus hædinus</i> Thomas	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
1916	<i>Chaerephon pumilus naivashæ</i> Hollister	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1916	<i>Rhinolophus keniensis</i> Hollister	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1923	<i>Petalia (Nycteris) thebaica aurantiaca</i> de Beaux	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1923	<i>Scotoecus artinii</i> de Beaux	<i>Scotoecus hirundo</i> (de Winton, 1899)
1924	<i>Eptesicus loveni</i> Granvik	<i>Myotis tricolor</i> (Temminck, 1832)
1936	<i>Nycteris nana tristis</i> G.M. Allen and Lawrence	<i>Nycteris nana</i> (K. Andersen, 1912)
1937	<i>R[hinopoma] cystops macinnesi</i> Hayman	<i>Rhinopoma macinnesi</i> Hayman, 1937
1957	<i>Saccolaimus incognita</i> Butler and Hopwood	<i>Taphozous incognita</i> (Butler & Hopwood, 1957)
1960	<i>Platymops barbatogularis parkeri</i> Harrison and Fleetwood	<i>Platymops setiger macmillani</i> Thomas, 1906
1980	<i>Myonycteris relicta</i> Bergmans	<i>Myonycteris relicta</i> Bergmans, 1980
1982	<i>Glauconycteris kenyacola</i> Peterson	<i>Glauconycteris kenyacola</i> Peterson, 1982
1984	<i>Chamtwaria pickfordi</i> Butler	<i>Chamtwaria pickfordi</i> Butler, 1984
1986	<i>Eptesicus hottentotus portavernus</i> Schlitter and Aggundey	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
2002	<i>Tadarida rusingae</i> Arroyo-Cabrales, Gregorin, Schlitter and Walker	<i>Mops rusingae</i> (Arroyo-Cabrales, Gregorin, Schlitter and Walker, 2002)
2014	<i>Scotophilus andrewreborii</i> Brooks and Bickham	<i>Scotophilus andrewreborii</i> Brooks and Bickham, 2014
2014	<i>Scotophilus livingstonii</i> Brooks and Bickham	<i>Scotophilus livingstonii</i> Brooks and Bickham, 2014
2014	<i>Scotophilus trujilloi</i> Brooks and Bickham	<i>Scotophilus trujilloi</i> Brooks and Bickham, 2014
2018	<i>Mops kerio</i> Gunnell and Manthi	<i>Mops kerio</i> Gunnell and Manthi, 2018
2018	<i>Mops turkwellensis</i> Gunnell and Manthi	<i>Mops turkwellensis</i> Gunnell and Manthi, 2018
2018	<i>R[ousettus] pattersoni</i> Gunnell and Manthi	<i>Rousettus pattersoni</i> Gunnell and Manthi, 2018
2018	<i>Saccolaimus kenyensis</i> Gunnell and Manthi	<i>Saccolaimus kenyensis</i> Gunnell and Manthi, 2018
2018	<i>Turkanycteris harrisi</i> Gunnell and Manthi	<i>Turkanycteris harrisi</i> Gunnell and Manthi, 2018
2020	<i>Laephotis kirinyaga</i> Monadjem, Patterson, Webala and Demos	<i>Laephotis kirinyaga</i> Monadjem, Patterson, Webala and Demos, 2020
2020	<i>Pseudoromicia nyanza</i> Monadjem, Patterson, Webala and Demos	<i>Pseudoromicia nyanza</i> Monadjem, Patterson, Webala and Demos, 2020

Liberia

1846	<i>Pteropus Haldemani</i> Hallowell	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
1881	<i>Leiponyx büttikoferi</i> Jentink	<i>Eidolon helvum helvum</i> (Kerr, 1792)
1888	<i>Epomophorus veldkampii</i> Jentink	<i>Nanonycteris veldkampii</i> (Jentink, 1888)
1888	<i>Vesperugo stampflii</i> Jentink	<i>Afronycteris nana</i> (Peters, 1852)
1899	<i>Ep[omophorus (Epomophorus)] büttikoferi</i> Matschie	<i>Epomops buettikoferi</i> (Matschie, 1899)
1900	<i>Pipistrellus minusculus</i> Miller	<i>Afronycteris nana</i> (Peters, 1852)
1989	<i>Rhinolophus clivosus hillorum</i> Koopman	<i>Rhinolophus hillorum</i> Koopman, 1989
2013	<i>Neoromicia roseveari</i> Monadjem, Richards, Taylor and Stoffberg	<i>Pseudoromicia roseveari</i> (Monadjem, Richards, Taylor and Stoffberg, 2013)
2020	<i>Miniopterus nimbae</i> Monadjem, Shapiro, Richards, Karabulut, Crawley, Nielsen, Hansen, Bohmann and Mourier	<i>Miniopterus nimbae</i> Monadjem, Shapiro, Richards, Karabulut, Crawley, Nielsen, Hansen, Bohmann and Mourier, 2020

Libya

1835-1841	<i>Vespertilio isabellinus</i> Temminck	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
1902	<i>Pipistrellus deserti</i> Thomas	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
2004	<i>Pipistrellus hanaki</i> Hulva and Benda	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
2004	<i>Plecotus teneriffae gaisleri</i> Benda, Kiefer, Hanák and Veith	<i>Plecotus gaisleri</i> Benda, Kiefer, Hanak & Veith, 2004

2006	<i>Scotophilis libycus</i> Horáček, Fejfar and Hulva	<i>Scotophilis libycus</i> Horáček, Fejfar and Hulva, 2006
Madagascar		
1803	<i>Pteropus rufus</i> E. Geoffroy Saint-Hilaire	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
1810	<i>Pteropus Edwardsii</i> E. Geoffroy Saint-Hilaire	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
1813	<i>Rhinolophus Commersonii</i> E. Geoffroy Saint-Hilaire	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
1816	<i>Pteropus madagascariensis</i> Oken	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
1825	<i>Pteropus phaiops</i> Temminck	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
1834	<i>Vespertilio goudoti</i> A. Smith	<i>Myotis goudoti</i> (A. Smith, 1834)
1838	<i>Pteropus phaeops</i> Oken	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
1858	<i>Vespertilio Madagascariensis</i> Tomes	<i>Myotis goudoti</i> (A. Smith, 1834)
1865	<i>Nyctinomus (Mormopterus) jugularis</i> Peters	<i>Mormopterus jugularis</i> (Peters, 1865)
1867	<i>Pteropus dupréanus</i> Schlegel	<i>Eidolon dupreanus</i> (Schegel, 1867)
1869	<i>Nyctinomus leucogaster</i> A. Grandidier	<i>Mops (Chaerephon) leucogaster</i> (A. Grandidier, 1869)
1869	<i>Nyctinomus Miarensis</i> A. Grandidier	<i>Mops (Mops) midas miarensis</i> (A. Grandidier, 1869)
1870	<i>Nyctinomus unicolor</i> A. Grandidier	<i>Mops (Mops) midas miarensis</i> (A. Grandidier, 1869)
1870	<i>Vespertilio sylvicola</i> A. Grandidier	<i>Myotis goudoti</i> (A. Smith, 1834)
1874	<i>Emballonura atrata</i> Peters	<i>Paremballonura atrata</i> (Peters, 1874)
1877	<i>Nyctinomus [(Mormopterus)] albiventer</i> Dobson	<i>Mormopterus jugularis</i> (Peters, 1865)
1878	<i>Myzopoda aurita</i> A. Milne-Edwards and A. Grandidier	<i>Myzopoda aurita</i> Milne-Edwards and A. Grandidier, 1878
1879	<i>Taphozous Dobsoni</i> Jentink	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
1881	<i>Scotophilus robustus</i> A. Milne-Edwards	<i>Scotophilus robustus</i> A. Milne-Edwards, 1881
1881	<i>Vesperus Humbloti</i> A. Milne-Edwards	<i>Laephotis humbloti</i> (A. Milne-Edwards, 1881)
1903	<i>Nyctinomus fulminans</i> Thomas	<i>Tadarida fulminans</i> (Thomas, 1903)
1905	<i>Vespertilio matroka</i> Thomas and Schwann	<i>Laephotis matroka</i> (Thomas and Schwann, 1905)
1906	<i>Miniopterus Majori</i> Thomas	<i>Miniopterus majori</i> Thomas, 1906
1906	<i>Miniopterus manavi</i> Thomas	<i>Miniopterus manavi</i> Thomas, 1906
1907	<i>Triaenops furcula</i> Trouessart	<i>Paratriaenops furculus</i> (Trouessart, 1907)
1908	<i>Pteropus rufus princeps</i> K. Andersen	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
1912	<i>Triaenops aurita</i> G. Grandidier	<i>Paratriaenops auritus</i> (G. Grandidier, 1912)
1918	<i>Chaerephon leucostigma</i> G.M. Allen	<i>Mops (Mops) leucostigma</i> (G.M. Allen, 1918)
1929	<i>Rousettus madagascariensis</i> G. Grandidier	<i>Rousettus madagascariensis</i> G. Grandidier, 1929
1937	<i>Nycteris madagascariensis</i> G. Grandidier	<i>Nycteris macrotis</i> Dobson, 1876
1953	<i>Otomops madagascariensis</i> Dorst	<i>Otomops madagascariensis</i> Dorst, 1953
1995	<i>Eptesicus somalicus malagasyensis</i> Peterson, Eger and Mitchell	<i>Laephotis malagasyensis</i> (Peterson, Eger and Mitchell, 1995)
1995	<i>Miniopterus gleni</i> Peterson, Eger and Mitchell	<i>Miniopterus gleni</i> Peterson, Eger and Mitchell, 1995
2004	<i>Chaerephon jobimena</i> Goodman and Cardiff	<i>Mops (Chaerephon) jobimena</i> (Goodman and Cardiff, 2004)
2005	<i>Scotophilus tandrefana</i> Goodman, Jenkins and Ratrimomanarivo	<i>Scotophilus tandrefana</i> Goodman, Jenkins and Ratrimomanarivo, 2005
2006	<i>Emballonura tiavato</i> Goodman, Cardiff, Ranivo, Russell and Yoder	<i>Paremballonura tiavato</i> (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)
2006	<i>Pipistrellus raceyi</i> Bates, Ratrimomanarivo, Harrison and Goodman	<i>Pipistrellus (Pipistrellus) raceyi</i> Bates, Ratrimomanarivo, Harrison and Goodman, 2006
2006	<i>Scotophilus marovaza</i> Goodman, Ratrimomanarivo and Randrianandrianina	<i>Scotophilus marovaza</i> Goodman, Ratrimomanarivo and Randrianandrianina, 2006
2007	<i>Myzopoda schliemanni</i> Goodman, Rakotondraparany and Kofoky	<i>Myzopoda schliemanni</i> Goodman, Rakotondraparany and Kofoky, 2007
2007	<i>Hipposideros besaoka</i> Samonds	<i>Macronycteris besaoka</i> (Samonds, 2007)
2007	<i>Miniopterus sororculus</i> Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton	<i>Miniopterus sororculus</i> Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton, 2007
2007	<i>Triaenops goodmani</i> Samonds	<i>Triaenops goodmani</i> Samonds, 2007
2008	<i>Miniopterus petersoni</i> Goodman, Bradman, Maminirina, Ryan, Christidis and Appleton	<i>Miniopterus petersoni</i> Goodman, Bradman, Maminirina, Ryan, Christidis & Appleton, 2008
2009	<i>Triaenops menamena</i> Goodman and Ranivo	<i>Triaenops menamena</i> Goodman and Ranivo, 2009
2009	<i>Miniopterus aelleni</i> Goodman, Maminirina, Weyeneth, Bradman, Christidis, Ruedi and Appleton	<i>Miniopterus aelleni</i> Goodman, Maminirina, Weyeneth, Bradman, Christidis, Ruedi and Appleton, 2009
2009	<i>Miniopterus griffithsi</i> Goodman, Maminirina, Bradman, Christidis and Appleton	<i>Miniopterus griffithsi</i> Goodman, Maminirina, Bradman, Christidis and Appleton, 2009
2009	<i>Miniopterus brachytragos</i> Goodman, Maminirina, Bradman, Christidis and Appleton	<i>Miniopterus brachytragos</i> Goodman, Maminirina, Bradman, Christidis and Appleton, 2009
2009	<i>Miniopterus mahafaliensis</i> Goodman, Bradman, Christides and Appleton	<i>Miniopterus mahafaliensis</i> Goodman, Bradman, Christides and Appleton, 2009
2010	<i>Chaerephon atsinanana</i> Goodman, Buccas, Naidoo, Ratrimomanarivo, Taylor and Lamb	<i>Mops (Chaerephon) atsinanana</i> (Goodman, Buccas, Naidoo, Ratrimomanarivo, Taylor and Lamb, 2010)

2011	<i>Miniopterus egeri</i> Goodman, Ramasindrazana, Maminirina, Schoeman and Appleton	<i>Miniopterus egeri</i> Goodman, Ramasindrazana, Maminirina, Schoeman and Appleton, 2011
2012	<i>Neoromicia robertsi</i> Goodman, Taylor, Ratrimomanarivo and Hofer	<i>Laephotis robertsi</i> (Goodman, Taylor, Ratrimomanarivo and Hofer, 2012)
2012	<i>Coleura kibomalandy</i> Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling	<i>Coleura kibomalandy</i> Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012
2015	<i>Miniopterus ambohitrensis</i> Goodman, Ramasindrazana, Naughton and Appleton	<i>Miniopterus ambohitrensis</i> Goodman, Ramasindrazana, Naughton and Appleton, 2015
2015	<i>Hypsignathus bemaity</i> Goodman, Rakotondramanana, Ramasindrazana, Kearney, Monadjem, Schoeman, Taylor, Naughton and Appleton	<i>Neoromicia bemaity</i> (Goodman, Rakotondramanana, Ramasindrazana, Kearney, Monadjem, Schoeman, Taylor, Naughton and Appleton, 2015)
2016	<i>Hipposideros cryptovalorona</i> Goodman, Schoeman, Rakotoarivelo and Willows-Munro	<i>Macronycteris cryptovalorona</i> (Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016)

Madeira (Portugal)

1825	<i>Nyctalus verrucosus</i> Bowdich	<i>Nyctalus leisleri</i> (Kuhl, 1817)
1878	<i>Vesperugo [(Vesperugo)] maderensis</i> Dobson	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
1906	<i>Pterygistes madeiræ</i> Barrett-Hamilton	<i>Nyctalus leisleri</i> (Kuhl, 1817)

Malawi

1904	<i>Rhinolophus augur zambesiensis</i> K. Andersen	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1904	<i>Rhinolophus empusa</i> K. Andersen	<i>Rhinolophus blasii</i> Peters, 1867
1917	<i>Scotoecus woodi</i> Thomas	<i>Scotoecus albofuscus</i> (Thomas, 1890)
1922	<i>Nycteris oriana</i> Kershaw	<i>Nycteris macrotis</i> Dobson, 1876
1946	<i>Eptesicus hottentotus bensoni</i> Roberts	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
2004	<i>Epomophorus ansellii</i> Bergmans and Van Strien	<i>Epomophorus ansellii</i> Bergmans and Van Strien, 2004

Mali

1928	<i>Chærophon (Lophomops) nigri</i> Hatt	<i>Mops (Chaerephon) leucogaster</i> (A. Grandidier, 1869)
1936	<i>Mops angolensis wonderi</i> Sanborn	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)

Mauritius

1802	<i>Spectrum rubidum</i> Daudin	<i>Pteropus subniger</i> (Kerr, 1792)
1803	<i>Pteropus ruber</i> E. Geoffroy Saint-Hilaire	<i>Pteropus subniger</i> (Kerr, 1792)
1804	<i>V[espertilio] acetabulosus</i> Hermann	<i>Mormopterus acetabulosus</i> (Hermann, 1804)
1804	<i>V[espertilio] acetabulosus</i> Hermann	<i>Mormopterus acetabulosus acetabulosus</i> (Hermann, 1804)
1804	<i>V[espertilio] mauritanus</i> Hermann	<i>Pteropus niger</i> (Kerr, 1792)
1815	<i>Pteropus collaris</i> Illiger	<i>Pteropus subniger</i> (Kerr, 1792)
1818	<i>Taphozous mauritanus</i> E. Geoffroy Saint-Hilaire	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
1823-1824	<i>Nyctinomus mauritanus</i> Horsfield	<i>Mormopterus acetabulosus acetabulosus</i> (Hermann, 1804)
1907	<i>Pteropus mascarinus</i> Mason	<i>Pteropus rodricensis</i> Dobson, 1878

Mayotte

1908	<i>Pteropus comorensis</i> Nicoll	<i>Pteropus seychellensis</i> A. Milne-Edwards, 1877

Morocco

1904	<i>E[uryalus] barbarus</i> K. Andersen and Matschie	<i>Rhinolophus mehelyi</i> Matschie, 1901
1906	<i>Hipposideros tephros</i> Cabrera	<i>Hipposideros tephros</i> (Cabrera, 1906)
1918	<i>[Rhinolophus hipposideros] escalerae</i> K. Andersen	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1937	<i>Rhinolophus hipposideros vespa</i> Laurent	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
1937	<i>Asellia tridens pallida</i> Laurent	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
1961	<i>Afroteropus gigas</i> Lavocat	<i>Megaderma gaillardi</i> (Trouessart, 1898)
1961	<i>Afroteropus gigas</i> Lavocat	<i>Megaderma gigas</i> (Lavocat, 1961)
1961	<i>Rhinolophus ferrumequinum mellali</i> Lavocat	<i>Rhinolophus mellali</i> Lavocat, 1961
1976	<i>Megaderma jaegeri</i> Sigé	<i>Megaderma jaegeri</i> Sigé, 1976
2011	<i>Miniopterus horaceki</i> Gunnell, Eiting and Geraads	<i>Miniopterus horaceki</i> Gunnell, Eiting and Geraads, 2011

2011	<i>Myotis darelbeidensis</i> Gunnell, Eiting and Geraads	<i>Myotis darelbeidensis</i> Gunnell, Eiting and Geraads, 2011
2011	<i>Rhinolophus maghrebensis</i> Gunnell, Eiting and Geraads	<i>Rhinolophus maghrebensis</i> Gunnell, Eiting and Geraads, 2011
2014	<i>Miniopterus maghrebensis</i> Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste	<i>Miniopterus maghrebensis</i> Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014
2019	<i>Myotis escalerae cabreræ</i> Çoraman, Mayer and Dietz	<i>Myotis zenatius</i> Ibañez, Juste, Salicini, Puechmaille and Ruedi, 2019
2019	<i>Myotis zenatius</i> Ibañez, Juste, Salicini, Puechmaille and Ruedi	<i>Myotis zenatius</i> Ibañez, Juste, Salicini, Puechmaille and Ruedi, 2019

Mozambique

1852	<i>Dysopes brachypterus</i> Peters	<i>Mops (Xiphonycteris) brachypterus</i> (Peters, 1852)
1852	<i>Dysopes dubius</i> Peters	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1852	<i>Dysopes limbatus</i> Peters	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1852	<i>Emballonura afra</i> Peters	<i>Coleura afra</i> (Peters, 1852)
1852	<i>Epomophorus crypturus</i> Peters	<i>Epomophorus crypturus</i> Peters, 1852
1852	<i>Nycteris fuliginosa</i> Peters	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1852	<i>Nycteris villosa</i> Peters	<i>Nycteris hispida</i> (Schreber, 1774)
1852	<i>Nycticejus planirostris</i> Peters	<i>Scotophilus dinganii</i> (A. Smith, 1833)
1852	<i>Nycticejus viridis</i> Peters	<i>Scotophilus viridis</i> (Peters, 1852)
1852	<i>Phyllorhina caffra</i> Peters	<i>Hipposideros caffer</i> (Sundevall, 1846)
1852	<i>Phyllorhina gracilis</i> Peters	<i>Hipposideros caffer</i> (Sundevall, 1846)
1852	<i>Phyllorhina patellifera</i> Peters	<i>Hipposideros caffer</i> (Sundevall, 1846)
1852	<i>Phyllorhina vittata</i> Peters	<i>Macronycteris vittatus</i> (Peters, 1852)
1852	<i>Rhinolophus lobatus</i> Peters	<i>Rhinolophus lobatus</i> Peters, 1852
1852	<i>Vespertilio nanus</i> Peters	<i>Afronycteris nana</i> (Peters, 1852)
1865	<i>Nycticejus nidicola</i> Kirk	<i>Kerivoula argentata</i> Tomes, 1861
1870	<i>Epomophorus macrocephalus</i> var. <i>unicolor</i> Gray	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
1908	<i>Scoteinus schlieffeni australis</i> Thomas and Wroughton	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
1965	<i>Nycteris vinsoni</i> Dalquest	<i>Nycteris vinsoni</i> Dalquest, 1965
1965	<i>Scotophilus alvenslebeni</i> Dalquest	<i>Scotophilus nigrita</i> (Schreber, 1774)
2012	<i>Rhinolophus mabuensis</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill	<i>Rhinolophus mabuensis</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
2012	<i>Rhinolophus mossambicus</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill	<i>Rhinolophus mossambicus</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
2013	<i>Miniopterus mossambicus</i> Monadjem, Goodman, Stanley and Appleton	<i>Miniopterus mossambicus</i> Monadjem, Goodman, Stanley and Appleton, 2013
2018	<i>Rhinolophus gorongosae</i> Taylor, MacDonald, Goodman, Kearney, Cotterill, Stoffberg, Monadjem, Schoeman, Guyton, Naskrecki and Richards	<i>Rhinolophus gorongosae</i> Taylor, MacDonald, Goodman, Kearney, Cotterill, Stoffberg, Monadjem, Schoeman, Guyton, Naskrecki and Richards, 2018
2020	<i>Miniopterus wilsoni</i> Monadjem, Guyton, Naskrecki, Richards, Kropff and Dalton	<i>Miniopterus wilsoni</i> Monadjem, Guyton, Naskrecki, Richards, Kropff and Dalton, 2020

Namibia

1861	<i>Kerivoula argentata</i> Tomes	<i>Kerivoula argentata</i> Tomes, 1861
1861	<i>Scotophilus rusticus</i> Tomes	<i>Pipistrellus (Pipistrellus) rusticus</i> (Tomes, 1861)
1861	<i>Scotophilus variegatus</i> Tomes	<i>Glauconycteris variegata</i> (Tomes, 1861)
1869	<i>Rhinolophus aethiops</i> Peters	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1871	<i>Nycteris damarensis</i> Peters	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1889	<i>Vesperugo damarensis</i> Noack	<i>Laephotis capensis</i> (A. Smith, 1829)
1906	<i>Scotophilus damarensis</i> Thomas	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
1906	<i>Scotophilus nigrita herero</i> Thomas	<i>Scotophilus dinganii</i> (A. Smith, 1833)
1917	<i>Platymops haagneri</i> Roberts	<i>Sauromys petrophilus haagneri</i> (Roberts, 1917)
1926	<i>Chaerephon (Lophomops) shortridgei</i> Thomas	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
1926	<i>Pipistrellus fouriei</i> Thomas	<i>Afronycteris nana</i> (Peters, 1852)
1927	<i>Miniopterus smitjanus</i> Thomas	<i>Miniopterus natalensis</i> (A. Smith, 1833)
1946	<i>Platymops petrophilus erongensis</i> Roberts	<i>Sauromys petrophilus erongensis</i> (Roberts, 1946)
1946	<i>Rhinolophus darlingi damarensis</i> Roberts	<i>Rhinolophus damarensis</i> Roberts, 1946
1971	<i>Laephotis namibensis</i> Setzer	<i>Laephotis namibensis</i> Setzer, 1971
2021	<i>Afrillonura namibensis</i> Rosina and Pickford	<i>Afrillonura namibensis</i> Rosina and Pickford, 2021

Nigeria

1842	<i>Kerivoula poensis</i> Gray	<i>Glauconycteris poensis</i> (Gray, 1842)
1862	<i>Sphyrocephalus labrosus</i> A. Murray	<i>Hypsignathus monstrosus</i> H. Allen, 1862
1868	<i>Epomophorus pusillus</i> Peters	<i>Epomophorus pusillus</i> Peters, 1868

1868	<i>Vesperus (Hesperoptenus) kraussii</i> Peters	<i>Glauconycteris poensis</i> (Gray, 1842)
1875	<i>Scotophilus gigas</i> Dobson	<i>Scotophilus nigrita</i> (Schreber, 1774)
1880	<i>Kerivoula Smithii</i> Thomas	<i>Kerivoula smithii</i> Thomas, 1880
1880	<i>Vesperugo (Vesperus) brunneus</i> Thomas	<i>Pseudoromicia brunnea</i> (Thomas, 1880)
1891	<i>Vesperugo (Vesperus) Moloneyi</i> Thomas	<i>Mimetillus moloneyi</i> (Thomas, 1891)
1900	<i>Mormopterus Whitleyi</i> Scharff	<i>Myopterus whitleyi</i> (Scharff, 1900)
1901	<i>Vespertilio platyops</i> Thomas	<i>Eptesicus (Cnephaeus) platyops</i> (Thomas, 1901)
1908	<i>Chærephon websteri</i> Dollman	<i>Mops (Chaerephon) leucogaster</i> (A. Grandidier, 1869)
1910	<i>Epomops franqueti strepitans</i> K. Andersen	<i>Epomops franqueti</i> (Tomes, 1860)
1911	<i>Pipistrellus culex</i> Thomas	<i>Afronycteris nana</i> (Peters, 1852)
1913	<i>Chærephon nigeriae</i> Thomas	<i>Mops (Chaerephon) nigeriae nigeriae</i> (Thomas, 1913)
1913	<i>Chærephon nigeriae</i> Thomas	<i>Mops (Chaerephon) nigeriae</i> (Thomas, 1913)
1913	<i>Rhinolophus foxi</i> Thomas	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1915	<i>Scotoecus falandæ</i> Thomas	<i>Scotoecus hindei</i> Thomas, 1901
1940	<i>Mops calabarensis</i> Hayman	<i>Mops (Xiphonycteris) nanulus</i> J.A. Allen, 1917
1958	<i>Taphozous perforatus swirae</i> Harrison	<i>Taphozous perforatus perforatus</i> E. Geoffroy St.-Hilaire, 1818

Oman

1994	<i>Dhofarella thaleri</i> Sigé, Thomas, Sen, Gheerbrant, Roger and Al-Sulaimani	<i>Dhofarella thaleri</i> Sigé et al., 1994

Pakistan

1881	<i>Phyllorhina tridens murraiana</i> Anderson	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)

Réunion

1792	<i>Vespertilio vampirus niger</i> Kerr	<i>Pteropus niger</i> (Kerr, 1792)
1792	<i>Vespertilio vampyrus subniger</i> Kerr	<i>Pteropus subniger</i> (Kerr, 1792)
1803	<i>Pteropus fuscus</i> E. Geoffroy Saint-Hilaire	<i>Pteropus niger</i> (Kerr, 1792)
1803	<i>Vespertilio borbonicus</i> E. Geoffroy Saint-Hilaire	<i>Scotophilus borbonicus</i> (E. Geoffroy St.-Hilaire, 1803)
1810	<i>Pteropus rubricollis</i> E. Geoffroy Saint-Hilaire	<i>Pteropus subniger</i> (Kerr, 1792)
1810	<i>Pteropus vulgaris</i> E. Geoffroy Saint-Hilaire	<i>Pteropus niger</i> (Kerr, 1792)
1895	<i>Pteropus pteropus</i> Merriam	<i>Pteropus niger</i> (Kerr, 1792)
2008	<i>Mormopterus francoismoutoui</i> Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst and Bowie	<i>Mormopterus francoismoutoui</i> Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst, Bowie, 2008

Romania

1817	<i>Vespertilio schreibersii</i> Kuhl	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
1901	<i>Rhinolophus mehelyi</i> Matschie	<i>Rhinolophus mehelyi</i> Matschie, 1901

Rwanda

1973	<i>Rhinolophus hilli</i> Aellen	<i>Rhinolophus hilli</i> Aellen, 1973

São Tomé and Príncipe

1889	<i>Cynonycteris brachycephala</i> Bocage	<i>Myonycteris brachycephala</i> (Bocage, 1889)
1889	<i>Miniopterus Newtoni</i> Bocage	<i>Miniopterus newtoni</i> Bocage, 1889
1891	<i>Phyllorhina commersoni</i> var. <i>thomensis</i> Bocage	<i>Macronycteris vittatus</i> (Peters, 1852)
1993	<i>Rousettus aegyptiacus princeps</i> Juste and Ibáñez	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1993	<i>Rousettus aegyptiacus thomensis</i> Feiler, Haft and Widmann	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1993	<i>Tadarida (Chaerephon) tomensis</i> Juste and Ibáñez	<i>Mops (Chaerephon) tomensis</i> (Juste and Ibáñez, 1993)
1993	<i>Rousettus aegyptiacus tomensis</i> Juste and Ibáñez	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)

Saudi Arabia

1828	<i>Rhinolophus clivosus</i> Cretzschmar	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1924	<i>Otonycteris jin</i> Cheesman and Hinton	<i>Otonycteris hemprichii</i> Peters, 1859

Senegal

1774	<i>Vespertilio hispidus</i> Schreber	<i>Nycteris hispida</i> (Schreber, 1774)

1774	<i>Vespertilio Nigrita</i> Schreber	<i>Scotophilus nigrita</i> (Schreber, 1774)
1792	<i>Vespertilio vampyrus helvus</i> Kerr	<i>Eidolon helvum helvum</i> (Kerr, 1792)
1792	<i>Vespertilio vampyrus helvus</i> Kerr	<i>Eidolon helvum</i> (Kerr, 1792)
1810	<i>Megaderma frons</i> E. Geoffroy Saint-Hilaire	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
1813	<i>Nycteris Daubentonii</i> E. Geoffroy Saint-Hilaire	<i>Nycteris hispida</i> (Schreber, 1774)
1816	<i>Myopterus senegalensis</i> Oken	<i>Myopterus daubentonii</i> Desmarest, 1820
1820	<i>Myopterus daubentonii</i> Desmarest	<i>Myopterus daubentonii</i> Desmarest, 1820
1820	<i>Nycteris Geoffroyi</i> Desmarest	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1820	<i>Taphozous senegalensis</i> Desmarest	<i>Taphozous perforatus perforatus</i> E. Geoffroy St.-Hilaire, 1818
1825	<i>Pteropus geoffroyi</i> Temminck	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1912	<i>Petalia gambiensis</i> K. Andersen	<i>Nycteris gambiensis</i> (K. Andersen, 1912)
1939	<i>Rhinolophus aethiops diversus</i> Sanborn	<i>Rhinolophus fumigatus</i> Rüppell, 1842
1960	<i>Pipistrellus Rüppelli senegalensis</i> Dorst	<i>Vansonia rueppellii senegalensis</i> (Dorst, 1960)

Seychelles

1868	<i>Coleura seychellensis</i> Peters	<i>Coleura seychellensis</i> Peters, 1868
1877	<i>Pteropus seychellensis</i> A. Milne-Edwards	<i>Pteropus seychellensis</i> A. Milne-Edwards, 1877
1893	<i>Pteropus aldabrensis</i> True	<i>Pteropus aldabrensis</i> True, 1893
1902	<i>Nyctinomus pusillus</i> Miller	<i>Mops (Chaerephon) pusillus</i> (Miller, 1902)
1915	<i>C[oleura] silhouettæ</i> Thomas	<i>Coleura seychellensis</i> Peters, 1868
1941	<i>[Triaenops] furinea</i> Tate	<i>Paratriaenops pauliani</i> (Goodman and Ranivo, 2008)
2008	<i>Triaenops pauliani</i> Goodman and Ranivo	<i>Paratriaenops pauliani</i> (Goodman and Ranivo, 2008)

Sierra Leone

1876	<i>Nycteris macrotis</i> Dobson	<i>Nycteris macrotis</i> Dobson, 1876
1908	<i>Nyctinomus leonis</i> Thomas	<i>Mops (Xiphonycteris) brachypterus</i> (Peters, 1852)
1908	<i>Rousettus smithii</i> Thomas	<i>Myonycteris angolensis smithii</i> (Thomas, 1908)
1908	<i>Myonycteris leptodon</i> K. Andersen	<i>Myonycteris leptodon</i> K. Andersen, 1908
1947	<i>Hipposideros jonesi</i> Hayman	<i>Hipposideros jonesi</i> Hayman, 1947

Somalia

1881	<i>Nycteris Revoulii</i> Robin	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1901	<i>Vespertilio minutus somalicus</i> Thomas	<i>Neoromicia somalica</i> (Thomas, 1901)
1910	<i>Rhinolophus brockmani</i> Thomas	<i>Rhinolophus blasii</i> Peters, 1867
1912	<i>Petalia damarensis brockmani</i> K. Andersen	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1915	<i>C[oleura] gallarum</i> Thomas	<i>Coleura gallarum</i> Thomas, 1915
1922	<i>Rhinolophus hildebrandti perauritus</i> de Beaux	<i>Rhinolophus eloquens</i> K. Andersen, 1905
1924	<i>Petalia parisii</i> de Beaux	<i>Nycteris parisii</i> (de Beaux, 1924)
1924	<i>Hipposideros caffer aurantiaca</i> de Beaux	<i>Hipposideros caffer</i> (Sundevall, 1846)
1931	<i>Asellia tridens italo-somalica</i> de Beaux	<i>Asellia italosomalica</i> de Beaux, 1931

South Africa

1800	<i>Vespertilio megalotis</i> Bechstein	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
1823	<i>Pteropus collaris</i> Lichtenstein	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1823	<i>Rhinolophus capensis</i> Lichtenstein	<i>Rhinolophus capensis</i> Lichtenstein, 1823
1829	<i>Nycteris affinis</i> A. Smith	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1829	<i>Nycteris Capensis</i> A. Smith	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1829	<i>Pteropus Leachii</i> A. Smith	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1829	<i>Rhinolophus Geoffroyii</i> A. Smith	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1829	<i>Vespertilio Capensis</i> A. Smith	<i>Laephotis capensis</i> (A. Smith, 1829)
1832	<i>Pteropus hottentottus</i> Temminck	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
1832	<i>V[espertilio] tricolor</i> Temminck	<i>Myotis tricolor</i> (Temminck, 1832)
1832	<i>Vespertilio epichrysurus</i> Temminck	<i>Scotophilus dinganii</i> (A. Smith, 1833)
1832	<i>Vespertilio platycephalus</i> Temminck	<i>Pipistrellus (Pipistrellus) hesperidus</i> (Temminck, 1840)
1833	<i>Nyctinomus Condylurus</i> A. Smith	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
1833	<i>Nyctinomus dubius</i> A. Smith	<i>Mormopterus acetabulosus natalensis</i> (A. Smith, 1847)
1833	<i>Vespertilio Dinganii</i> A. Smith	<i>Scotophilus dinganii</i> (A. Smith, 1833)
1833	<i>Vespertilio Hottentota</i> A. Smith	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
1833	<i>Vespertilio Natalensis</i> A. Smith	<i>Miniopterus natalensis</i> (A. Smith, 1833)
1835	<i>Taphozous leucopterus</i> Temminck	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
1840	<i>N[ycteris] discolor</i> Wagner	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1840	<i>Vespertilio d'asythrix</i> Temminck	<i>Miniopterus natalensis</i> (A. Smith, 1833)
1840	<i>Vespertilio minuta</i> Temminck	<i>Laephotis capensis</i> (A. Smith, 1829)
1835-1841	<i>Vespertilio megalurus</i> Temminck	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
1846	<i>Pteropus Wahlbergi</i> Sundevall	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
1846	<i>Rhinolophus caffer</i> Sundevall	<i>Hipposideros caffer</i> (Sundevall, 1846)
1846	<i>Vespertilio scotinus</i> Sundevall	<i>Miniopterus natalensis</i> (A. Smith, 1833)

1846	<i>Vespertilio subtilis</i> Sundevall	<i>Pipistrellus (Pipistrellus) hesperidus subtilis</i> (Sundevall, 1846)
1847	<i>Dysopes natalensis</i> A. Smith	<i>Mormopterus acetabulosus natalensis</i> (A. Smith, 1847)
1847	<i>Vespertilio lanosus</i> A. Smith	<i>Kerivoula lanosa</i> (A. Smith, 1847)
1855	<i>V[espertilio] Smithii</i> Wagner	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
1860	<i>Rhinolophus auritus</i> Sundevall	<i>Rhinolophus capensis</i> Lichtenstein, 1823
1876	<i>Nyctinomus africanus</i> Dobson	<i>Tadarida ventralis</i> (Heuglin, 1861)
1878	<i>Kerivoula brunnea</i> Dobson	<i>Kerivoula lanosa</i> (A. Smith, 1847)
1904	<i>Rhinolophus Denti</i> Thomas	<i>Rhinolophus denti</i> Thomas, 1904
1904	<i>Rhinolophus augur</i> K. Andersen	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1904	<i>Rhinolophus augur zuluensis</i> K. Andersen	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
1905	<i>Vespertilio capensis gracilior</i> Thomas and Schwann	<i>Laephotis capensis</i> (A. Smith, 1829)
1906	<i>Miniopterus fraterculus</i> Thomas and Schwann	<i>Miniopterus fraterculus</i> Thomas and Schwann, 1906
1908	<i>Rhinolophus swinnyi</i> Gough	<i>Rhinolophus swinnyi</i> Gough, 1908
1908	<i>Scabrifer notius</i> G.M. Allen	<i>Laephotis capensis</i> (A. Smith, 1829)
1909	<i>Miniopterus breyeri</i> Jameson	<i>Miniopterus natalensis</i> (A. Smith, 1833)
1913	<i>Rhinolophus swinnyi piriensis</i> Hewitt	<i>Rhinolophus swinnyi</i> Gough, 1908
1913	<i>Pipistrellus nanus australis</i> Roberts	<i>Afronycteris nana</i> (Peters, 1852)
1917	<i>Cloeotis percivali australis</i> Roberts	<i>Cloeotis percivali</i> Thomas, 1901
1917	<i>Otomops icarus</i> Chubb	<i>Otomops martiensseni icarus</i> Chubb, 1917
1917	<i>Platymops petrophilus</i> Roberts	<i>Sauromys petrophilus</i> (Roberts, 1917)
1919	<i>Cistugo lesueuri</i> Roberts	<i>Cistugo lesueuri</i> Roberts, 1919
1919	<i>Eptesicus melckorum</i> Roberts	<i>Laephotis capensis</i> (A. Smith, 1829)
1924	<i>Eptesicus zuluensis</i> Roberts	<i>Neoromicia zuluensis</i> (Roberts, 1924)
1924	<i>Kerivoula nidicola zuluensis</i> Roberts	<i>Kerivoula argentata</i> Tomes, 1861
1924	<i>Rhinolophus darlingi barbertonensis</i> Roberts	<i>Rhinolophus darlingi</i> K. Andersen, 1905
1926	<i>Chaerephon pumilus elphicki</i> Roberts	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1938	<i>Platymops haagneri umbratus</i> Shortridge and Carter	<i>Sauromys petrophilus umbratus</i> (Shortridge & Carter, 1938)
1938	<i>Scotophilus angusticeps</i> Shortridge and Carter	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
1942	<i>Eptesicus megalurus pallidior</i> Shortridge	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
1946	<i>Platymops petrophilus fitzsimonsi</i> Roberts	<i>Sauromys petrophilus umbratus</i> (Shortridge & Carter, 1938)
1946	<i>Scotophilus nigrita pondoensis</i> Roberts	<i>Scotophilus dinganii</i> (A. Smith, 1833)
1948	<i>Pipistrellus (Romicia) kuhlii broomi</i> Roberts	<i>Pipistrellus (Pipistrellus) hesperidus broomi</i> Roberts, 1948
2001	<i>Pipistrellus africanus meesteri</i> Kock	<i>Afronycteris nana</i> (Peters, 1852)
2012	<i>Rhinolophus cohenae</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill	<i>Rhinolophus cohenae</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012

South Sudan

1916	<i>Eptesicus ugandæ</i> Hollister	<i>Neoromicia somalica</i> (Thomas, 1901)

Spain

1904	<i>E[uryalus] Cabrerae</i> K. Andersen and Matschie	<i>Rhinolophus euryale</i> Blasius, 1853
1904	<i>Vespertilio Boscai</i> Cabrera	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
1904	<i>Vespertilio ochromixtus</i> Cabrera	<i>Hypsugo savii</i> (Bonaparte, 1837)

Sudan

1803	<i>Pteropus stramineus</i> E. Geoffroy Saint-Hilaire	<i>Eidolon helvum helvum</i> (Kerr, 1792)
1826	<i>Nycticejus leucogaster</i> Cretzschmar	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
1826	<i>Vespertilio temminckii</i> Cretzschmar	<i>Vansonia rueppellii</i> (J.B. Fischer, 1829)
1827	<i>Vespertilio temminckii</i> Cretzschmar	<i>Vansonia rueppellii rueppellii</i> (J.B. Fischer, 1829)
1829	<i>V[espertilio] Rüppellii</i> J.B. Fischer	<i>Vansonia rueppellii</i> (J.B. Fischer, 1829)
1829	<i>Vespertilio ruppellii</i> J.B. Fischer	<i>Vansonia rueppellii rueppellii</i> (J.B. Fischer, 1829)
1837	<i>Pteropus labiatus</i> Temminck	<i>Epomophorus labiatus</i> (Temminck, 1837)
1840	<i>N[ycteris] albiventer</i> Wagner	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1843	<i>Dysopes midas</i> Sundevall	<i>Mops (Mops) midas</i> (Sundevall, 1843)
1843	<i>Dysopes midas</i> Sundevall	<i>Mops (Mops) midas midas</i> (Sundevall, 1843)
1859	<i>Rhinopoma Lepsianum</i> Peters	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
1862	<i>Pterocyon paleaceus</i> Peters	<i>Eidolon helvum helvum</i> (Kerr, 1792)
1864	<i>Dysopes hepaticus</i> Heuglin	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
1864	<i>Epomophorus anurus</i> Heuglin	<i>Epomophorus labiatus</i> (Temminck, 1837)
1865	<i>Pteropus palmarum</i> Heuglin	<i>Eidolon helvum helvum</i> (Kerr, 1792)
1866	<i>Plecotus æthiopicus</i> Heuglin	<i>Plecotus christii</i> Gray, 1838
1866	<i>Plecotus æthiopicus</i> Heuglin and Fitzinger	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
1866	<i>Plecotus ustus</i> Fitzinger and Heuglin	<i>Otonycteris hemprichii</i> Peters, 1859

- 1866 *Rhinopoma longicaudatum* Fitzinger *Rhinopoma cystops* Thomas, 1903
 1866 *Rhinopoma sennaariense* Fitzinger *Rhinopoma cystops* Thomas, 1903
 1866 *Vesperugo hypoleucus* Heuglin and Fitzinger *Vansonia rueppellii rueppellii* (J.B. Fischer, 1829)
 1866 *Vesperugo sennaariensis* Heuglin and Fitzinger *Vansonia rueppellii rueppellii* (J.B. Fischer, 1829)
 1866 *Xantharpyia leucomelas* Fitzinger *Eidolon helvum helvum* (Kerr, 1792)
 1866 *Xantharpyia leucomelas* Heuglin *Eidolon helvum* (Kerr, 1792)
 1868 *Nycteris Geoffroyi* Var. *Senegalensis* Hartmann *Nycteris thebaica* E. Geoffroy St.-Hilaire, 1818
 1877 *Dysopes talpinus* Heuglin *Tadarida aegyptiaca aegyptiaca* (E. Geoffroy St.-Hilaire, 1818)
 1877 *Nycticejus serratus* Heuglin *Taphozous nudiventris* Cretzschmar, 1830
 1877 *Rhinopoma cordofanicum* Heuglin *Rhinopoma microphyllum* (Brünnich, 1782)
 1877 *Taphozous maritimus* Heuglin *Taphozous perforatus* E. Geoffroy St.-Hilaire, 1818
 1878 *Nycteris aethiopica* Dobson *Nycteris macrotis* Dobson, 1876
 1897 *[Nyctinomus (Nyctinomus) pumilus]* Var. Major Trouessart *Mops (Chaerephon) major* (Trouessart, 1897)
 1901 *Glauconycteris Floweri* de Winton *Eptesicus (Rhinopterus) floweri* (de Winton, 1901)
 1903 *Nyctinomus cisturus* Thomas *Mops (Chaerephon) bemmeleni* (Jentink, 1879)
 1903 *Nyctinomus demonstrator* Thomas *Mops (Mops) demonstrator* (Thomas, 1903)
 1904 *Pipistrellus ariel* Thomas *Hypsugo ariel* (Thomas, 1904)
 1904 *Rhinolophus Dobsoni* Thomas *Rhinolophus landeri* Martin, 1838
 1907 *Lavia frons affinis* K. Andersen and Wroughton *Lavia frons* (E. Geoffroy St.-Hilaire, 1810)
 1908 *Scoteinus schlieffeni albiventer* Thomas and Wroughton *Nycticeinops schlieffenii* (Peters, 1859)
 1911 *Chærephon emini* Wroughton *Mops (Mops) condylurus* (A. Smith, 1833)
 1914 *Scotophilus altilis* G.M. Allen *Scotophilus altilis* G.M. Allen, 1914
 1915 *C[oleura] g[allorum] nilosa* Thomas *Coleura afra* (Peters, 1852)
 1915 *Glauconycteris phalæna* Thomas *Glauconycteris variegata* (Tomes, 1861)
 1915 *Rhinopterus lowei* Thomas *Eptesicus (Rhinopterus) floweri* (de Winton, 1901)
 1915 *Taphozous Sudani* Thomas *Taphozous perforatus sudani* Thomas, 1915
 1916 *Eptesicus rectitragus* Wettstein *Neoromicia guineensis* (Bocage, 1889)
 1916 *Nyctinomus (Nyctinomus) tongaënsis* Wettstein *Tadarida aegyptiaca aegyptiaca* (E. Geoffroy St.-Hilaire, 1818)
 1916 *Scotoecus cinnamomeus* Wettstein *Nycticeinops schlieffenii* (Peters, 1859)
 1920 *Taphozous hamiltoni* Thomas *Taphozous hamiltoni* Thomas, 1920
 1923 *Pipistrellus marrensis* Thomas and Hinton *Pipistrellus (Pipistrellus) rusticus* (Tomes, 1861)
 1956 *Platymops barbatogularis* Harrison *Platymops setiger macmillani* Thomas, 1906
 1969 *Rhinopoma hardwickei sennaariense* Kock *Rhinopoma cystops* Thomas, 1903
 1969 *Rhinopoma microphyllum tropicalis* Kock *Rhinopoma microphyllum* (Brünnich, 1782)
- Switzerland
- 1869 *Vesperugo noctula* var. *maxima* Fatio *Nyctalus lasiopterus* (Schreber, 1780)
- Tanzania
- 1867 *Miniopterus minor* Peters *Miniopterus minor* Peters, 1867
 1868 *Rhinolophus Deckenii* Peters *Rhinolophus deckenii* Peters, 1868
 1875 *Vesperugo pulcher* Dobson *Vansonia rueppellii pulcher* (Dobson, 1875)
 1876 *Vesperugo (Vesperus) grandidieri* Dobson *Nycticeinops grandidieri* (Dobson, 1876)
 1878 *Kerivoula africana* Dobson *Kerivoula africana* Dobson, 1878
 1880 *Epomophorus minor* Dobson *Epomophorus minor* Dobson, 1880
 1887 *Scotophilus minimus* Noack *Nycticeinops schlieffenii* (Peters, 1859)
 1893 *Phyllorhina rubra* Noack *Hipposideros ruber* (Noack, 1893)
 1897 *Nyctinomus martiensseni* Matschie *Otomops martiensseni* (Matschie, 1897)
 1897 *Nyctinomus martiensseni* Matschie *Otomops martiensseni martiensseni* (Matschie, 1897)
 1899 *Ep[omophorus (Epomophorus)] stuhlmanni* Matschie *Epomophorus wahlbergi* (Sundevall, 1846)
 1899 *Vespertilio venustus* Matschie *Myotis welwitschii* (Gray, 1866)
 1901 *Nyctinomus Emini* de Winton *Mops (Chaerephon) major* (Trouessart, 1897)
 1908 *Rousettus sjöstedti* Lönnberg *Rousettus aegyptiacus* (E. Geoffroy St.-Hilaire, 1810)
 1909 *Pteropus (Spectrum) voeltzkowi* Matschie *Pteropus voeltzkowi* Matschie, 1909
 1923 *Nycteris marica* Kershaw *Nycteris grandis* Peters, 1865
 1942 *Mops angolensis orientis* G.M. Allen and Loveridge *Mops (Mops) condylurus* (A. Smith, 1833)
 1957 *Pipistrellus (Pipistrellus) permixtus* Aellen *Pipistrellus (Pipistrellus) permixtus* Aellen, 1957
 2000 *Rhinolophus maendeleo* Kock, Csorba and Howell *Rhinolophus maendeleo* Kock, Csorba and Howell, 2000
 2003 *Tanzanycteris mannardi* Gunnell, Jacobs, Herendeen, Head, Kowalski, Msuya, Mizambwa, Harrison, Habersetzer and Storch, 2003

2008	<i>Mops bakarii</i> Stanley	<i>Mops (Xiphonycteris) bakarii</i> Stanley, 2009
2015	<i>Cardioderma leakeyi</i> Gunnell, Butler, Greenwood and Simmons	<i>Cardioderma leakeyi</i> Gunnell, Butler, Greenwood and Simmons, 2015
2015	<i>Myzopoda africana</i> Gunnell, Butler, Greenwood and Simmons	<i>Myzopoda africana</i> Gunnell, Butler, Greenwood and Simmons, 2015
2015	<i>Scotoecus olduvensis</i> Gunnell, Butler, Greenwood and Simmons	<i>Scotoecus olduvensis</i> Gunnell, Butler, Greenwood and Simmons, 2015

The Gambia

1835	<i>Pteropus Gambianus</i> Ogilby	<i>Epomophorus gambianus</i> (Ogilby, 1835)
1835	<i>Pteropus Gambianus</i> Ogilby	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
1835	<i>Pteropus macrocephalus</i> Ogilby	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
1835	<i>Pteropus megacephalus</i> Swainson	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
1836	<i>Pteropus epomophorus</i> Bennett	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
1836	<i>Pteropus Whitei</i> Bennett	<i>Epomophorus gambianus gambianus</i> (Ogilby, 1835)
1889	<i>Vesperugo (Vesperus) Rendalli</i> Thomas	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
1890	<i>Scotophilus albobfuscus</i> Thomas	<i>Scotoecus albobfuscus</i> (Thomas, 1890)
1901	<i>Nyctinomus gambianus</i> de Winton	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
1906	<i>H[ipposiderus]. gigas gambiensis</i> K. Andersen	<i>Macronycteris gigas</i> (Wagner, 1845)

Tunisia

1955	<i>Rhinolophus euryale tuneti</i> Deleuil and Labbé	<i>Rhinolophus mehelyi</i> Matschie, 1901
1977	<i>Myotis blythii punicus</i> Felten	<i>Myotis punicus</i> Felten, 1977
1991	<i>Dizya exsultans</i> Sigé	<i>Dizya exsultans</i> Sigé, 1991
2012	<i>Witwatia sigei</i> Ravel, Marivaux, Tabuce, Bel Haj Ali, Essid and Vianey-Liaud	<i>Witwatia sigei</i> Ravel, Marivaux, Tabuce, Bel Haj Ali, Essid and Vianey-Liaud, 2012
2016	<i>?Necromantis fragmentum</i> Ravel	<i>Necromantis fragmentum</i> (Ravel, 2016)
2016	<i>?Palaeophyllophora tunisiensis</i> Ravel	<i>Palaeophyllophora tunisiensis</i> (Ravel, 2016)
2016	<i>?Vespertiliavus aenigma</i> Ravel	<i>Vespertiliavus aenigma</i> (Ravel, 2016)
2016	<i>Hipposideros (Pseudorhinolophus) africanum</i> Ravel	<i>Hipposideros africanum</i> Ravel, 2016
2016	<i>Khoufechia gunnelli</i> Ravel	<i>Khoufechia gunnelli</i> Ravel, 2016
2016	<i>Vespertiliavus kasserinensis</i> Ravel	<i>Vespertiliavus kasserinensis</i> (Ravel, 2016)

Uganda

1905	<i>Glauconycteris papilio</i> Thomas	<i>Glauconycteris variegata</i> (Tomes, 1861)
1905	<i>Rhinolophus Hildebrandti eloquens</i> K. Andersen	<i>Rhinolophus eloquens</i> K. Andersen, 1905
1906	<i>Hipposideros caffer centralis</i> K. Andersen	<i>Hipposideros ruber</i> (Noack, 1893)
1906	<i>Rousettus lanosus</i> Thomas	<i>Stenonycteris lanosus</i> (Thomas, 1906)
1907	<i>Nyctinomus aloysii-sabaudiae</i> Festa	<i>Mops (Chaerephon) aloysiisabaudiae</i> (Festa, 1907)
1913	<i>Pipistrellus fuscipes</i> Thomas	<i>Vansonia rueppellii fuscipes</i> (Thomas, 1913)
1965	<i>Rousettus angolensis ruwenzorii</i> Eisentraut	<i>Myonycteris angolensis ruwenzorii</i> (Eisentraut, 1965)
2020	<i>Pseudoromicia kityoi</i> Monadjem, Kerbis Peterhans, Nalikka, Waswa, Demos and Patterson	<i>Pseudoromicia kityoi</i> Monadjem, Kerbis Peterhans, Nalikka, Waswa, Demos and Patterson, 2020

United Kingdom

1825	<i>Vespertilio Pygmæus</i> Leach	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)

Yemen

1869	<i>Vesperus Bottae</i> Peters	<i>Eptesicus (Cnephæus) bottae</i> (Peters, 1869)

Zambia

1914	<i>Nycteris woodi</i> K. Andersen	<i>Nycteris woodi</i> K. Andersen, 1914
1920	<i>Mimetillus thomasi</i> Hinton	<i>Mimetillus thomasi</i> Hinton, 1920
1938	<i>Chærephon lancasteri</i> Hayman	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
1960	<i>Epomophorus gambianus parvus</i> Ansell	<i>Epomophorus crypturus</i> Peters, 1852
2002	<i>Rhinolophus sakejiensis</i> Cotterill	<i>Rhinolophus sakejiensis</i> Cotterill, 2002

Zimbabwe

1904	<i>Rhinolophus simulator</i> K. Andersen	<i>Rhinolophus simulator</i> K. Andersen, 1904
1905	<i>Rhinolophus Darlingi</i> K. Andersen	<i>Rhinolophus darlingi</i> K. Andersen, 1905
1914	<i>Rhinolophus bembanicus</i> Senna	<i>Rhinolophus simulator</i> K. Andersen, 1904

1920	<i>Kerivoula lucia</i> Hinton	<i>Kerivoula lanosa</i> (A. Smith, 1847)
1946	<i>Nycteris woodi sabiensis</i> Roberts	<i>Nycteris woodi</i> K. Andersen, 1914
1946	<i>Nyctinomus mastersoni</i> Roberts	<i>Tadarida fulminans</i> (Thomas, 1903)
1946	<i>Nyctinomus rhodesiae</i> Roberts	<i>Mops (Chaerephon) ansorgei</i> (Thomas, 1913)
1946	<i>Rhinolophus swinnyi rhodesiae</i> Roberts	<i>Rhinolophus rhodesiae</i> Roberts, 1946
1962	<i>Taphozous sudani australis</i> Harrison	<i>Taphozous perforatus sudani</i> Thomas, 1915
1997	<i>Lissonycteris angolensis goliath</i> Bergmans	<i>Myonycteris angolensis goliath</i> (Bergmans, 1997)
2012	<i>Rhinolophus smithersi</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill	<i>Rhinolophus smithersi</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012

Appendix 3e: Common Names in Alphabetical Order**Common Name****Taxon Name**

Họ dơi cánh (Vietnamese)	MINIOPTERIDAE Dobson, 1875
Họ dơi muỗi (Vietnamese)	VESPERTILIONIDAE Gray, 1821
Họ dơi quạ (Vietnamese)	PTEROPODIDAE Gray, 1821
Họ dơi thò đuôi (Vietnamese)	MOLOSSIDAE Gervais, 1856
Họ dơi bao (Vietnamese)	EMBALLONURIDAE Gervais, 1855
Họ dơi lá (Vietnamese)	RHINOLOPHIDAE Gray, 1825
Họ dơi ma (Vietnamese)	MEGADERMATIDAE H. Allen, 1864
Họ dơi mũi (Vietnamese)	HIPPOSIDERIDAE Lydekker, 1891
Обични пећински летипас [= Obični pećinski letipás] (Serbian)	Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
东非伏翼 (Chinese)	Pipistrellus (Pipistrellus) aero Heller, 1912
中非小伏翼 (Chinese)	Pipistrellus (Pipistrellus) nanulus Thomas, 1904
串耳蹄蝠 (Chinese)	Hipposideros megalotis (Heuglin, 1861)
乌干达菊头蝠 (Chinese)	Rhinolophus eloquens K. Andersen, 1905
任氏伏翼 (Chinese)	Pseudoromicia rendalli (Thomas, 1889)
佐氏菊头蝠 (Chinese)	Rhinolophus clivosus Cretzschmar, 1828
侏凹脸蝠 (Chinese)	Nycteris nana (K. Andersen, 1912)
侏犬吻蝠 (Chinese)	Mops (Xiphonycteris) nanulus J.A. Allen, 1917
侏长翼蝠 (Chinese)	Miniopterus minor Peters, 1867
侏颈囊果蝠 (Chinese)	Epomophorus minimus Claessen & De Vree, 1991
兰德菊头蝠 (Chinese)	Rhinolophus landeri Martin, 1838
利文斯顿黄蝠 (Chinese)	Scotophilus livingstonii Brooks and Bickham, 2014
北非果蝠 (Chinese)	Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
北非犬吻 (Chinese)	Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
南非伏翼 (Chinese)	Laephotis capensis (A. Smith, 1829)
南非墓蝠 (Chinese)	Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
南非鼠耳蝠 (Chinese)	Myotis tricolor (Temminck, 1832)
史密斯彩蝠 (Chinese)	Kerivoula smithii Thomas, 1880
吕氏伏翼 (Chinese)	Vansonia rueppellii (J.B. Fischer, 1829)
Ørkenlangøre (Norwegian)	Otonycteris hemprichii Peters, 1859
哈氏巨犬吻蝠 (Chinese)	Otomops harrisoni Ralph, Richards, Taylor, Napier and Lamb, 2015
喀麦隆蹄蝠 (Chinese)	Doryrhina camerunensis (Eisentraut, 1956)
喀麦隆黄蝠 (Chinese)	Scotophilus nux Thomas, 1904
垂耳犬吻蝠 (Chinese)	Mops (Chaerephon) major (Trouessart, 1897)
埃及墓蝠 (Chinese)	Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
大白纹蹄蝠 (Chinese)	Macronycteris vittatus (Peters, 1852)
大眼蹄蝠 (Chinese)	Doryrhina cyclops (Temminck, 1853)
大耳凹脸蝠 (Chinese)	Nycteris macrotis Dobson, 1876
大耳犬吻蝠 (Chinese)	Otomops martiensseni (Matschie, 1897)
大耳犬吻蝠 (Chinese)	Tadarida lobata (Thomas, 1891)
大长翼蝠 (Chinese)	Miniopterus inflatus Thomas, 1903
孤颌果蝠 (Chinese)	Myonycteris relicta Bergmans, 1980
安哥拉犬吻蝠 (Chinese)	Mops (Mops) condylurus (A. Smith, 1833)
安哥拉颌果蝠 (Chinese)	Myonycteris angolensis (Bocage, 1898)
安氏凹脸蝠 (Chinese)	Nycteris aurita (K. Andersen, 1912)
安氏犬吻蝠 (Chinese)	Mops (Chaerephon) ansorgei (Thomas, 1913)
安氏黄蝠 (Chinese)	Scotophilus andrewborii Brooks and Bickham, 2014
宽首伏翼 (Chinese)	Nycticeinops crassulus (Thomas, 1904)
小彩蝠 (Chinese)	Kerivoula lanosa (A. Smith, 1847)
小犬吻蝠 (Chinese)	Mops (Chaerephon) pumilus (Cretzschmar, 1826)
小长翼蝠 (Chinese)	Miniopterus fraterculus Thomas and Schwann, 1906
小颈囊果蝠 (Chinese)	Epomophorus labiatus (Temminck, 1837)
岛犬吻蝠 (Chinese)	Tadarida fulminans (Thomas, 1903)
巨蹄蝠 (Chinese)	Macronycteris gigas (Wagner, 1845)
布什维尔德菊头蝠 (Chinese)	Rhinolophus simulator K. Andersen, 1904
希氏菊头蝠 (Chinese)	Rhinolophus hildebrandtii Peters, 1878
彩蝶蝠 (Chinese)	Glauconycteris variegata (Tomes, 1861)
彼德犬吻蝠 (Chinese)	Platymops setiger (Peters, 1878)
德氏菊头蝠 (Chinese)	Rhinolophus deckenii Peters, 1868
斑犬吻蝠 (Chinese)	Mops (Chaerephon) bivittatus (Heuglin, 1861)
施氏墓蝠 (Chinese)	Nycticeinops schlieffenii (Peters, 1859)
无畏犬吻蝠 (Chinese)	Mops (Xiphonycteris) thersites (Thomas, 1903)

Common Name

暗翼宽吻蝠 (Chinese)
 暗黑伏翼 (Chinese)
 松氏蹄蝠 (Chinese)
 查平犬吻蝠 (Chinese)
 桑布鲁伏 (Chinese)
 棕红鼠耳蝠 (Chinese)
 Ēģiptes augšsikspārnis (Latvian)
 淡翼宽吻 (Chinese)
 淡色凹脸蝠 (Chinese)
 温氏长耳蝠 (Chinese)
 特鲁黄蝠 (Chinese)
 狭齿果蝠 (Chinese)
 珀氏三叉蝠 (Chinese)
 白翼伏翼 (Chinese)
 白腹黄 (Chinese)
 Īzabel gōnlūcā (Azerbaijani)
 短翼犬吻蝠 (Chinese)
 祖鲁伏翼 (Chinese)
 米达犬吻蝠 (Chinese)
 粗毛凹脸蝠 (Chinese)
 索马里伏翼 (Chinese)
 纳塔尔长翼蝠 (Chinese)
 肯尼亚墓蝠 (Chinese)
 肯尼亚蝶蝠 (Chinese)
 腺尾犬吻蝠 (Chinese)
 花蝶蝠 (Chinese)
 苏丹墓蝠 (Chinese)
 莫桑比克长翼蝠 (Chinese)
 裸腹墓蝠 (Chinese)
 Šikšniukas mažylis (Lithuanian)
 Šikšniukas nykštukas (Lithuanian)
 Širokouhi šišmilu0353\ (Croatian)
 Širokouhi šišmiš (Bosnian)
 Širokouhi mračnjak (Croatian)
 Širokouhi netopir (Slovenian)
 Škorpjonjedi pušcavež (Slovenian)
 Šrajberov šišmiš (Bosnian)
 Šrajberov šišmiš (Croatian)
 Šreiberio ilgapiŗtis (Lithuanian)
 Šreibersa sikspārnis (Latvian)
 Šumski noćnik (Montenegrin)
 šerowec senegalský (Czech)
 šerowec tébský (Czech)
 šerowecové (Czech)
 širokouší morousi (Czech)
 širokouší ušanové (Czech)
 širokouhi slijepi miš (Montenegrin)
 诺氏蹄蝠 (Chinese)
 贝尔墓蝠 (Chinese)
 赤犬吻蝠 (Chinese)
 Ūsuotasis pelėausis (Lithuanian)
 达马拉菊头蝠 (Chinese)
 银彩蝠 (Chinese)
 银蝶蝠 (Chinese)
 锈色伏翼 (Chinese)
 锤头果蝠 (Chinese)
 žlázokřídlec Lesueurův (Czech)
 žlázokřídlec Seabrův (Czech)
 žlázokřídlecové (Czech)
 žlázokřídlecovití (Czech)
 žlutokřídli lyronosi (Czech)
 长尾棕蝠 (Chinese)
 非洲三叉蝠 (Chinese)
 非洲假吸血蝠 (Chinese)
 非洲凹脸蝠 (Chinese)
 非洲大犬吻蝠 (Chinese)

Taxon Name

Scotoecus hirundo (de Winton, 1899)
Pipistrellus (Pipistrellus) hesperidus (Temminck, 1840)
Hipposideros caffer (Sundevall, 1846)
Mops (Chaerephon) chapini (J.A. Allen, 1917)
Afronycteris helios (Heller, 1912)
Myotis bocagii (Peters, 1870)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Scotoecus albofuscus (Thomas, 1890)
Nycteris arge Thomas, 1903
Laephotis wintoni Thomas, 1901
Scotophilus trujilloi Brooks and Bickham, 2014
Stenonycteris lanosus (Thomas, 1906)
Cloeotis percivali Thomas, 1901
Pseudoromicia tenuipinnis (Peters, 1872)
Scotophilus leucogaster (Cretzschmar, 1826)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Neoromicia zuluensis (Roberts, 1924)
Mops (Mops) midas (Sundevall, 1843)
Nycteris hispida (Schreber, 1774)
Neoromicia somalica (Thomas, 1901)
Miniopterus natalensis (A. Smith, 1833)
Taphozous hildegardeae Thomas, 1909
Glauconycteris kenyacola Peterson, 1982
Mops (Chaerephon) bemmeleni (Jentink, 1879)
Glauconycteris humeralis J.A. Allen, 1917
Taphozous hamiltoni Thomas, 1920
Miniopterus mossambicus Monadjem, Goodman, Stanley and Appleton, 2013
Taphozous nudiventris Cretzschmar, 1830
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Otonycteris hemprichii Peters, 1859
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Nycteris hispida (Schreber, 1774)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris G. Cuvier and E. Geoffroy, 1795
Tadarida Rafinesque, 1814
Barbastella Gray, 1821
Barbastella barbastellus (Schreber, 1774)
Hipposideros ruber (Noack, 1893)
Saccolaimus peli (Temminck, 1853)
Mops (Chaerephon) russatus (J.A. Allen, 1917)
Myotis mystacinus (Kuhl, 1817)
Rhinolophus fumigatus Rüppell, 1842
Kerivoula argentata Tomes, 1861
Glauconycteris argentata (Dobson, 1875)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Hypsignathus monstrosus H. Allen, 1862
Cistugo lesueurii Roberts, 1919
Cistugo seabrae Thomas, 1912
Cistugo Thomas, 1912
CISTUGONIDAE Lack, Roehrs, Stanley, Ruedi and Van den Bussche, 2010
Lavia Gray, 1838
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Triaenops afer Peters, 1877
Cardioderma cor (Peters, 1872)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Tadarida ventralis (Heuglin, 1861)

Common Name

非洲大黄蝠 (Chinese)
 非洲小狐蝠 (Chinese)
 非洲扁颅蝠 (Chinese)
 非洲长翼蝠 (Chinese)
 非洲鞘尾蝠 (Chinese)
 韦氏颈囊果蝠 (Chinese)
 香蕉伏翼 (Chinese)
 魁凹脸蝠 (Chinese)
 魏氏鼠耳蝠 (Chinese)
 麦氏鼠尾蝠 (Chinese)
 黄伏翼 (Chinese)
 黄毛果蝠 (Chinese)
 黄翼蝠 (Chinese)
 Ατρίχη ταφονυχτερίδα (Greek)
 Αιγυπτιακή φρουτονυχτερίδα (Greek)
 Ασιατική τρανονυχτερίδα (Greek)
 Βουνονυχτερίδα (Greek)
 Λευκονυχτερίδα (Greek)
 Μεγάλος νυκτοβάτης (Greek)
 Μεσορινόλοφος (Greek)
 Μεσορινόλοφος (Greek)
 Μικρονυκτοβάτης (Greek)
 Μικρονυχτερίδα (Greek)
 Μικρορινόλοφος (Greek)
 Μουστακονυχτερίδα (Greek)
 Μπαρμπαστέλλος (Greek)
 Νανονυχτερίδα (Greek)
 Νανονυχτερίδα του Hanak (Greek)
 Νυκτοβάτης των Αζόρων (Greek)
 Νυχτερίδας της Μαδέρας (Greek)
 Νυχτονόμος (Greek)
 Ποδαρομυτίδα (Greek)
 Πτερυγονυχτερίδα (Greek)
 Πυρρομυτίδα (Greek)
 Ρινόλοφος του Blasius (Greek)
 Ρινόλοφος του Mehely (Greek)
 Τρανομυτίδα της Καρχηδόνας (Greek)
 Τρανορινόλοφος (Greek)
 Ωτονυχτερίδα της Τενερίφης (Greek)
 Ωτονυχτερίδα της ερήμου (Greek)
 Ωχρή τρανονυχτερίδα (Greek)
 Јужни потковичар [= Južni potkovičar] (Serbian)
 Џуцест лилјак (Macedonian)
 Азиатская широкоушка [= Aeiatskaya shirokouchka] (Russian)
 Азорски вечерник (Bulgarian)
 Азорски вечерник (Macedonian)
 Азорски ноћник [= Azorski noćnik] (Serbian)
 Атлантски сл[и]јепи мишић [= Atlantski sl[ij]epi mišić] (Serbian)
 Б[и]јелоруби слепи мишић [= B[ij]elorubi slepi mišić] (Serbian)
 Белорабен лилјак [= Beloraben Liljak] (Macedonian)
 Бласиев потковичар [= Blasiev Potkovichar] (Macedonian)
 Блискоисточни поноћњак [= Bliskoistočni ponočnjak] (Serbian)
 Большой подковонос [= Bolshoj podkvonos] (Russian)
 Ботев лилјак (Macedonian)
 Булдогов прилеп (Bulgarian)
 Бульдоговые (Russian)
 Велики ноћник [= Veliki noćnik] (Serbian)
 Велики потковичар [= Veliki potkovičar] (Serbian)
 Вечерница азорская (Russian)
 Вечерница гигантская (Russian)
 Вечерница малая (Russian)
 Вечірниця азорська (Ukrainian)
 Вечірниця гігантська (Ukrainian)
 Вечірниця мала (Ukrainian)
 Вухань канарський (Ukrainian)
 Вушан канарскі (Belarusian)
 Вячэрніца азорская (Belarusian)
 Вячэрніца гіганцкая (Belarusian)
 Вячэрніца малая (Belarusian)

Taxon Name

Scotophilus nigrita (Schreber, 1774)
Epomophorus pusillus Peters, 1868
Mimetillus moloneyi (Thomas, 1891)
Miniopterus africanus Sanborn, 1936
Coleura afra (Peters, 1852)
Epomophorus wahlbergi (Sundevall, 1846)
Afronycteris nana (Peters, 1852)
Nycteris grandis Peters, 1865
Myotis welwitschii (Gray, 1866)
Rhinopoma macinnesi Hayman, 1937
Nycticeinops grandidieri (Dobson, 1876)
Eidolon helvum (Kerr, 1792)
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Taphozous nudiventris Cretzschmar, 1830
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus blasii Peters, 1867
Rhinolophus euryale Blasius, 1853
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus hipposideros (Bechstein, 1800)
Myotis mystacinus (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Nyctalus azoreum (Thomas, 1901)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Tadarida teniotis (Rafinesque, 1814)
Myotis capaccinii (Bonaparte, 1837)
Miniopterus schreibersii (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Rhinolophus blasii Peters, 1867
Rhinolophus mehelyi Matschie, 1901
Myotis punicus Felten, 1977
Rhinolophus ferrumequinum (Schreber, 1774)
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Rhinolophus blasii Peters, 1867
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Barbastella leucomelas (Cretzschmar, 1826)

Nyctalus azoreum (Thomas, 1901)
Nyctalus azoreum (Thomas, 1901)
Nyctalus azoreum (Thomas, 1901)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)

Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Rhinolophus blasii Peters, 1867
Eptesicus (Cnephæus) bottae (Peters, 1869)

Rhinolophus ferrumequinum (Schreber, 1774)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Tadarida teniotis (Rafinesque, 1814)
MOLOSSIDAE Gervais, 1856
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)
Nyctalus azoreum (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus azoreum (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Plecotus teneriffae Barrett-Hamilton, 1907
Plecotus teneriffae Barrett-Hamilton, 1907
Nyctalus azoreum (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)

Common Name

Гигантская вечерница [= Gigantskaya vechernicha] (Russian)
 Гладконосые (Russian)
 Голем вечерник (Macedonian)
 Голем потковичар [= Golem Potkovichar] (Macedonian)
 Голокореман прилеп (Bulgarian)
 Голотрби гробнички м[и]јешкокрилаш [=Golotrbi grobnički m[i]ješkokrilaš] (Serbian)
 Голям вечерник (Bulgarian)
 Голям подковонос (Bulgarian)
 Даўгакрыл звычайны (Belarusian)
 Длиннокрыл обыкновенный (Russian)
 Длиннокрыловые (Russian)
 Довгокрил звичайний (Ukrainian)
 Довгокрили [= Dohvokryly] (Ukrainian)
 Долгокрилест лилјак (Macedonian)
 Долгопрст ноћник [= Dolgoprst Nokjnik] (Macedonian)
 Дугодлаки сл[и]јепи мишић [= Dugodlaki sl[i]jepi mišić] (Serbian)
 Дугопрсти вечерњак [= Dugoprsti večernjak] (Serbian)
 Дългопръст ношник (Bulgarian)
 Европейская широкоушка [= Evropejskaya širokoushka] (Serbian)
 Европски дугокрилаш (Serbian)
 Европски широкоушан [= Evropski širokoušan] (Serbian)
 Египетски овошен лилјак (Macedonian)
 Египетски плодояден прилеп (Bulgarian)
 Звичайні листоноси [= Zvychayni lystonosy] (Ukrainian)
 Изабелинов прилеп (Bulgarian)
 Изабелов лилјак (Macedonian)
 иилик (Пергач) пустельный (Ukrainian)
 Кажан Бота (Belarusian)
 Кажан изабэлавы (Belarusian)
 Канарски дългоух прилеп (Bulgarian)
 Канарски сиви дугоушан (Serbian)
 Кафяво прилепче (Bulgarian)
 Кожан изабелловый (Russian)
 Кожан пустынный (Russian)
 Кожановидный нетопырь [= Kozhanovidny netopyr] (Russian)
 Крилан египетський (Ukrainian)
 Крылан египетский (Russian)
 Крылан егіпецкі (Belarusian)
 Крылановые (Russian)
 Лайслеров (малък) вечерник (Bulgarian)
 Летючі лисиці (Ukrainian)
 Лжевампиры (Russian)
 Либијски сл[и]јепи мишић [= Libijski sl[i]jepi mišić] (Serbian)
 Лилик изабеліновий (Ukrainian)
 Лилики [= Lylyky] (Ukrainian)
 Лиликові [= Lylykovi] (Ukrainian)
 Листоносы (Russian)
 Листоніс Коммерсона [= Lystonis Kommersona] (Ukrainian)
 Листоніс південноафриканський [= Lystonis pівденноафриканський] (Ukrainian)
 Ложные подковоносы (Russian)
 Магребски велики вечерњак [= Magrebski veliki večernjak] (Serbian)
 Магребски ношник (Bulgarian)
 Магребски ноћник (Macedonian)
 Магребски поноћњак [= Magrebski ponoć njak] (Serbian)
 Магільнік галабрухі (Belarusian)
 Мадейрско прилепче (Bulgarian)
 Мал лилјак (Macedonian)
 Мал потковичар [= Mal Potkovichar] (Macedonian)
 Малая вечерница [= Malaya vechernicha] (Russian)
 Мали ноћник [= Mali noćnik] (Serbian)
 Мали потковичар [= Mali potkovičar] (Serbian)
 Малий еполетовий крилан [=Malyy epoletovyy krylan] (Ukrainian)
 Малко кафяво прилепче (Bulgarian)
 Малък подковонос (Bulgarian)
 Малый нетопырь [=Malyj Neopyr] (Russian)

Taxon Name

Nyctalus lasiopterus (Schreber, 1780)
 VESPERTILIONIDAE Gray, 1821
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)
Taphozous nudiventris Cretzschmar, 1830
Taphozous nudiventris Cretzschmar, 1830
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
 MINIOPTERIDAE Dobson, 1875
Miniopterus schreibersii (Kuhl, 1817)
 MINIOPTERIDAE Dobson, 1875
Miniopterus schreibersii (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
Myotis capaccinii (Bonaparte, 1837)
Myotis capaccinii (Bonaparte, 1837)
Barbastella barbastellus (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
 HIPPOSIDERIDAE Lydekker, 1891
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Plecotus teneriffae Barrett-Hamilton, 1907
Plecotus teneriffae Barrett-Hamilton, 1907
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Hypsugo savii (Bonaparte, 1837)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
 PTEROPODIDAE Gray, 1821
Nyctalus leisleri (Kuhl, 1817)
 PTEROPODIDAE Gray, 1821
 MEGADERMATIDAE H. Allen, 1864
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
 Vespertilioninae Gray, 1821
 VESPERTILIONIDAE Gray, 1821
 HIPPOSIDERIDAE Lydekker, 1891
Macronycteris commersoni (E. Geoffroy St.-Hilaire, 1813)
Hipposideros caffer (Sundevall, 1846)
 HIPPOSIDERIDAE Lydekker, 1891
Myotis punicus Felten, 1977
Myotis punicus Felten, 1977
Myotis punicus Felten, 1977
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Taphozous nudiventris Cretzschmar, 1830
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus hipposideros (Bechstein, 1800)
Epomophorus labiatus (Temminck, 1837)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)

Common Name

Медеирски лилјак (Macedonian)
 Медитерански потковичар (Macedonian)
 Мехелов потковичар (Macedonian)
 Мешкокрыл голобрюхий (Russian)
 Мустакат ношник (Bulgarian)
 Мустакест ношник [= Mustakjest Nokjnik] (Macedonian)
 Мішкокрил голобрюхий (Ukrainian)
 Начніца вусатая (Belarusian)
 Начніца даўгапалая (Belarusian)
 Начніца маґрыбская (Belarusian)
 Начніца трохкаляровая (Belarusian)
 Начніца фінікійская (Belarusian)
 Неороміція [= Neoromitsiya] (Ukrainian)
 Нетапыр Ганака (Belarusian)
 Нетапыр Куля (Belarusian)
 Нетапыр Саві (Belarusian)
 Нетапыр мадэрскі (Belarusian)
 Нетапыр міжземнаморскі (Belarusian)
 Нетапыр-карлік (Belarusian)
 Нетапыр-пігмей (Belarusian)
 Нетопир Ганака (Ukrainian)
 Нетопир білосмугий (Ukrainian)
 Нетопир карлик (Ukrainian)
 Нетопир мадейський (Ukrainian)
 Нетопир малий [= Netopyr malyj] (Ukrainian)
 Нетопир пігмей (Ukrainian)
 Нетопир скельний (Casi) (Ukrainian)
 Нетопырь Ганака (Russian)
 Нетопырь Куля (Russian)
 Нетопырь кожановидный (Russian)
 Нетопырь мадейский (Russian)
 Нетопырь малый (пигмей) (Russian)
 Нетопырь средиземноморский (Russian)
 Нетопырь-карлик [= Netopyr karlik] (Russian)
 Ночница средиземная (итальянская) (Russian)
 Ночница трёхцветная (Russian)
 Ночница усатая (Russian)
 Ночница финикийская (Russian)
 Нічниця вусата (Ukrainian)
 Нічниця довгопала (Ukrainian)
 Нічниця маґрибська (Ukrainian)
 Нічниця триколірна (Ukrainian)
 Нічниця [= Nichnytsi] (Ukrainian)
 Обични сл[и]јепи мишић [= Obični sl[ij]epi mišić] (Serbian)
 Обыкновенный длиннокрыл [= Obyknoennyj dlinnokryl] (Russian)
 Опашест лилјак [= Opashest Liljak] (Macedonian)
 Падкаванос Блазіуса (Belarusian)
 Падкаванос Мегеля (Belarusian)
 Падкаванос вялікі (Belarusian)
 Падкаванос малы (Belarusian)
 Падкаванос міжземнаморскі (Belarusian)
 Патуљасті сл[и]јепи мишић [= Patuljasti sl[ij]epi mišić] (Serbian)
 Пещерен дългокрил (Bulgarian)
 Подковоугубы (Russian)
 Подковонос Мегели (очковый) (Russian)
 Подковонос большой (Russian)
 Подковонос малый [= Podkovonos malyj] (Russian)
 Подковонос на Мехели (Bulgarian)
 Подковонос средиземноморский (Russian)
 Подковонос южный (Russian)
 Подковоносые (Russian)
 Полунощен прилеп на Бот (Bulgarian)
 Прилепче на Ханак (Bulgarian)
 Пустинен дългоух прилеп (Bulgarian)
 Пустиньски дугоушан [= Pustinjski dugoušan] (Serbian)
 Підковик (Підковоніс) Блазіуса (Ukrainian)
 Підковик (Підковоніс) Мегеля (Ukrainian)
 Підковик (Підковоніс) великий [=Pidkovyk velykyj] (Ukrainian)
 Підковик (Підковоніс) південний (Ukrainian)
 Підковик (Підковоніс) малий [= Pidkovyk malyj] (Ukrainian)
 Підковикові [= Pidkovykovij] (Ukrainian)

Taxon Name

Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Taphozous nudiventris Cretzschmar, 1830
Myotis mystacinus (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Myotis mystacinus (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Myotis punicus Felten, 1977
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis punicus Felten, 1977
Neoromicia Roberts, 1926
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myotis capaccinii (Bonaparte, 1837)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis mystacinus (Kuhl, 1817)
Myotis punicus Felten, 1977
Myotis mystacinus (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Myotis punicus Felten, 1977
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis Tate, 1942
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)

Tadarida teniotis (Rafinesque, 1814)
Rhinolophus blasii Peters, 1867
Rhinolophus mehelyi Matschie, 1901
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus euryale Blasius, 1853
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)

Miniopterus schreibersii (Kuhl, 1817)
HIPPOSIDERIDAE Lydekker, 1891
Rhinolophus mehelyi Matschie, 1901
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus mehelyi Matschie, 1901
Rhinolophus blasii Peters, 1867
Rhinolophus euryale Blasius, 1853
RHINOLOPHIDAE Gray, 1825
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Otonycteris hemprichii Peters, 1859
Otonycteris hemprichii Peters, 1859
Rhinolophus blasii Peters, 1867
Rhinolophus mehelyi Matschie, 1901
Rhinolophus ferrumequinum (Schreber, 1774)

Rhinolophus euryale Blasius, 1853
Rhinolophus hipposideros (Bechstein, 1800)
RHINOLOPHIDAE Gray, 1825

Common Name

Риђи вечерњак [= Ridí večernjak] (Serbian)
 Савиев лилјак [= Saviev Liljak] (Macedonian)
 Савиево прилепче (Bulgarian)
 Свободнохвостые (Russian)
 Складчатогуб широкоухий (Russian)
 Средиземноморски подковонос (Bulgarian)
 Средиземноморский нетопыр [= Sredizemnomorskiy netopyr] (Russian)
 Средиземноморско прилепче (Bulgarian)
 Средоземни потковичар [= Sredozemni potkovičar] (Serbian)
 Средоземни репаш [= Sredozemni repaš] (Serbian)
 Стрелоух белоброхий (Russian)
 Стрѣлавух Хемрыха (Belarusian)
 Стріловух білочеревий (Ukrainian)
 Тадарида європейська (Ukrainian)
 Тадарыда еўрапейская (Belarusian)
 Тамнолики бркати вечерњак [= Tamnoliki brkati večernjak] (Serbian)
 Тамнооки потковичар [= Tamnooki potkovičar] (Serbian)
 Тенерифов ушест лилјак (Macedonian)
 Торбокрилест лилјак (Macedonian)
 Трехцветная ну1086\чница [= Tryokhtsvetnaya nochnitsa] (Russian)
 Трилистоніс африканський [= Trylystonis afrykans'kyy] (Ukrainian)
 Трицветен ношник (Bulgarian)
 Тробоен нојник [= Troboen Nokjnik] (Macedonian)
 Усатая ночница [= Usataya nochnitsa] (Russian)
 Ушан канарский (Russian)
 Футлярохвостые (Russian)
 Ханаков лилјак (Macedonian)
 Хемпричиев долгоушест лилјак (Macedonian)
 Широковух європейський [= Shyrovokukh zvychnyy] (Ukrainian)
 Широкоух прилеп (Bulgarian)
 Широкоухий складчатогуб (Serbian)
 Широкоушест лилјак (Macedonian)
 Широкоушка європейская (Russian)
 Шумски вечерник [= Shumski Vechernik] (Macedonian)
 Шыракавух еўрапейскі (Belarusian)
 Южен подковонос (Bulgarian)
 Южный подковонос [= Yuzhnyj podkovonos] (Russian)
 Ագորյան իրիկնաչոցիկ (Armenian)
 Անապատային մաշկեղ (Armenian)
 Բեղավոր գիշերաչոցիկ (Armenian)
 Բլազիի կամ (Armenian)
 Փաջաճ փոքրաչոցիկ (Armenian)
 Եգիպտական մեծաչոցիկ (Armenian)
 Եռագույն գիշերաչոցիկ (Armenian)
 Եվրոպական լայնական չոցիկ (Armenian)
 Թզուկ փոքրաչոցիկ (Armenian)
 Իզաբելլայի մաշկեղ (Armenian)
 Լայնական ծալքաշուրթ չոցիկ (Armenian)
 Կանարական կամ Տենեիֆյան ականջեղ (Armenian)
 Հանակի փոքրաչոցիկ (Armenian)
 Հարավային պայտաքիթ (Armenian)
 Հսկա իրիկնաչոցիկ (Armenian)
 Մադեյրական փոքրաչոցիկ (Armenian)
 Մաշեղանման փոքրաչոցիկ (Armenian)
 Մեծ պայտաքիթ (Armenian)
 Մեհելիի պայտաքիթ (Armenian)
 Մերկփոր պարկաթ (Armenian)
 Միջերկրածովային գիշերաչոցիկ (Armenian)
 Միջերկրածովային փոքրաչոցիկ (Armenian)
 Սովորական երկարաթև չոցիկ (Armenian)
 Սպիտակափոր սրականջ (Armenian)
 Փյունիկյան գիշերաչոցիկ (Armenian)
 Փոքր իրիկնաչոցիկ (Armenian)
 Փոքր պայտաքիթ (Armenian)
 Վիջերկրածովային պայտաքիթ (Armenian)
 טוּא (Hebrew)
 טוּא (Hebrew)
 מְצוּא (Hebrew)

Taxon Name

Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Hypsugo savii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
 MOLOSSIDAE Gervais, 1856
Tadarida teniotis (Rafinesque, 1814)
Rhinolophus blasii Peters, 1867
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)

Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Rhinolophus euryale Blasius, 1853

Tadarida teniotis (Rafinesque, 1814)
Otonycteris hemprichii Peters, 1859
Otonycteris hemprichii Peters, 1859
Otonycteris hemprichii Peters, 1859
Tadarida teniotis (Rafinesque, 1814)
Tadarida teniotis (Rafinesque, 1814)
Myotis mystacinus (Kuhl, 1817)

Rhinolophus mehelyi Matschie, 1901
Plecotus teneriffae Barrett-Hamilton, 1907
Taphozous nudiventris Cretzschmar, 1830
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)

Triaenops afer Peters, 1877

Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis mystacinus (Kuhl, 1817)
Plecotus teneriffae Barrett-Hamilton, 1907
 EMBALLONURIDAE Gervais, 1855
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Otonycteris hemprichii Peters, 1859
Barbastella barbastellus (Schreber, 1774)

Barbastella barbastellus (Schreber, 1774)
Tadarida teniotis (Rafinesque, 1814)
Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Nyctalus leisleri (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
Rhinolophus euryale Blasius, 1853
Nyctalus azoreum (Thomas, 1901)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Myotis mystacinus (Kuhl, 1817)
Rhinolophus blasii Peters, 1867
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Barbastella barbastellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Tadarida teniotis (Rafinesque, 1814)
Plecotus teneriffae Barrett-Hamilton, 1907
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Rhinolophus euryale Blasius, 1853
Nyctalus lasiopterus (Schreber, 1780)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Hypsugo savii (Bonaparte, 1837)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus mehelyi Matschie, 1901
Taphozous nudiventris Cretzschmar, 1830
Myotis capaccinii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Otonycteris hemprichii Peters, 1859
Myotis punicus Felten, 1977
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus blasii Peters, 1867
Otonycteris hemprichii Peters, 1859
Plecotus christii Gray, 1838
Eptesicus (Cnephæus) bottae (Peters, 1869)

Common Name

חורש ומוח (Hebrew)
 חנף (Hebrew)
 חיל (Hebrew)
 חשפון גדות (Hebrew)
 חטלפון אירופי (Hebrew)
 חטלפון סאבי (Hebrew)
 חרסיה (Hebrew)
 חרסיה גדולה (Hebrew)
 חרסיה גמדי (Hebrew)
 חרסיה גמדי (Hebrew)
 חרסיה חיוור (Hebrew)
 חרסיה מצוי (Hebrew)
 חרסיה (Hebrew)
 الوطاطر خفاش (Arabic)
 எகிப்திய வ வால் (Tamil)
 திறந்த ரம்பு கல்லறை வால் (Tamil)
 ஸ்திரப ர்ஸ்-ன் தீண்ட விரலுள்ள வால் (Tamil)
 აზორული მელამურა (Georgian)
 ბლაზის ცხვირნალა (Georgian)
 გიგანტური მელამურა (Georgian)
 დიდი ცხვირნალა (Georgian)
 ეგვიპტური მფრინავი ძაღლი (Georgian)
 ევროპული მაჩუკთელა (Georgian)
 ევროპული ტადარიდა (Georgian)
 კანარიული ყურა (Georgian)
 კაპაჩინის მლამიოზი (Georgian)
 კულის (Georgian)
 მადეირული ღამორი (Georgian)
 მალრიბული მლამიოზი (Georgian)
 მეჭელის ცხვირნალა (Georgian)
 მცირე მელამურა (Georgian)
 მცირე ცხვირნალა (Georgian)
 პაწია ღამორი (Georgian)
 სავის ღამორი (Georgian)
 სამფერი მლამიოზი (Georgian)
 სამხრეთული ცხვირნალა (Georgian)
 ტყის მეგვიანე (Georgian)
 უდაბნოს მეგვიანე (Georgian)
 ულვაშა მლამიოზი (Georgian)
 შიმშელმუცელა ქარქაშკუდა (Georgian)
 ჩვეულებრივი ფრთაგრძელი (Georgian)
 ხმელთაშუაზღვიური ღამორი (Georgian)
 ჯუჯა ღამორი (Georgian)
 ჰანაკის ღამორი (Georgian)
 ჰემპრიხის ისარყურა (Georgian)
 Sraipars-ṇ Nīṇṭa Virālūḷa Velavāl (Tamil)
 45 kHz pipistrelle (English)
 55 kHz pipistrelle (English)
 Ağqarın oxqulaq yarasa (Azerbaijani)
 Aalwyndakvlermuis; Aalwynvlermuis (Afrikaans)
 Aavikkolepakko (Finnish)
 Aba Leaf-nosed Bat (English)
 Aba Roundleaf Bat (English)
 Aba-Rundblattnase (German)
 Abendfledermäuse (German)
 Abendsegler (German)
 Abendstern-Pipistrelle (German)
 Abendstern-Zwergfledermaus (German)
 Abo Bat (English)
 Abo Butterfly Bat (English)
 Abo-Schmetterlingsfledermaus (German)
 Abruzzi's Wrinkle-lipped Bat (English)
 Abyssinian Horseshoe Bat (English)
 Adam's Horseshoe Bat (English)
 Adams Hufeisennase (German)
 Adi uzunqanad yarasa (Azerbaijani)
 Aellens Langflügel-fledermaus (German)
 Aellen's Leaf-nosed Bat (English)

Taxon Name

Barbastella leucomelas (Cretzschmar, 1826)
Miniopterus schreibersii (Kuhl, 1817)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Myotis capaccinii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Hypsugo savii (Bonaparte, 1837)
Rhinolophus euryale Blasius, 1853
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus mehelyi Matschie, 1901
Rhinolophus blasii Peters, 1867
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
CHIROPTERA Blumenbach, 1779
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Taphozous nudiventris Cretzschmar, 1830
Miniopterus schreibersii (Kuhl, 1817)
Nyctalus azoreum (Thomas, 1901)
Rhinolophus blasii Peters, 1867
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Barbastella barbastellus (Schreber, 1774)
Tadarida teniotis (Rafinesque, 1814)
Plecotus teneriffae Barrett-Hamilton, 1907
Myotis capaccinii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Myotis puniceus Felten, 1977
Rhinolophus mehelyi Matschie, 1901
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Hypsugo savii (Bonaparte, 1837)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Rhinolophus euryale Blasius, 1853
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Myotis mystacinus (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Miniopterus schreibersii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Otonycteris hemprichii Peters, 1859
Miniopterus schreibersii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Otonycteris hemprichii Peters, 1859
Neoromicia zuluensis (Roberts, 1924)
Otonycteris hemprichii Peters, 1859
Hipposideros abae J.A. Allen, 1917
Hipposideros abae J.A. Allen, 1917
Hipposideros abae J.A. Allen, 1917
Nycticeinops Hill & Harrison, 1987
Nyctalus Bowdichi, 1825
Pipistrellus (Pipistrellus) hesperidus (Temminck, 1840)
Pipistrellus (Pipistrellus) hesperidus (Temminck, 1840)
Glauconycteris poensis (Gray, 1842)
Glauconycteris poensis (Gray, 1842)
Glauconycteris poensis (Gray, 1842)
Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus adami Aellen and Brosset, 1968
Rhinolophus adami Aellen and Brosset, 1968
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus aelleni Goodman, Maminirina, Weyeneth, Bradman, Christidis, Ruedi and Appleton, 2009
Hipposideros marisae Aellen, 1954

Common Name	Taxon Name
Aellen's Long-fingered Bat (English)	<i>Miniopterus aelleni</i> Goodman, Maminirina, Weyeneth, Bradman, Christidis, Ruedi and Appleton, 2009
Aellen's Pipistrelle (English)	<i>Pipistrellus (Pipistrellus) inexpectatus</i> Aellen, 1959
Aellen's Pipistrelle Bat (English)	<i>Pipistrellus (Pipistrellus) inexpectatus</i> Aellen, 1959
Aellen's Roundleaf Bat (English)	<i>Hipposideros marisae</i> Aellen, 1954
Aequatorial Broad Winged Bat (English)	<i>Miniopterus arenarius</i> Heller, 1912
Aerial Pipistrelle Bat (English)	<i>Pipistrellus (Pipistrellus) aero</i> Heller, 1912
Aflul Matsui (Hebrew)	<i>Eptesicus (Cnephaeus) bottae</i> (Peters, 1869)
aflul matzuy (Hebrew)	<i>Eptesicus (Cnephaeus) bottae</i> (Peters, 1869)
African Epauletted bats (English)	<i>Epomops</i> Gray, 1866
African False Vampire Bat (English)	<i>Cardioderma cor</i> (Peters, 1872)
African False Vampire Bats (English)	<i>Cardioderma</i> Peters, 1873
African Free-tailed Bats (English)	<i>Myopterus</i> E. Geoffroy St.-Hilaire, 1818
African Giant Free-tailed Bat (English)	<i>Tadarida ventralis</i> (Heuglin, 1861)
African Heart-nosed Bat (English)	<i>Cardioderma cor</i> (Peters, 1872)
African Long-eared Bats (English)	<i>Laephotis</i> Thomas, 1901
African Long-fingered Bat (English)	<i>Miniopterus africanus</i> Sanborn, 1936
African Long-tongued Bat (English)	<i>Megaloglossus woermanni</i> Pagenstecher, 1885
African Long-tongued Fruit-bats (English)	<i>Megaloglossus</i> Pagenstecher, 1885
African Nectar Fruit-bats (English)	<i>Megaloglossus</i> Pagenstecher, 1885
African pipistrelle (English)	<i>Pipistrellus (Pipistrellus) hesperidus</i> (Temminck, 1840)
African Sheath-tailed Bat (English)	<i>Coleura afra</i> (Peters, 1852)
African Straw-coloured Fruit-bat (English)	<i>Eidolon helvum</i> (Kerr, 1792)
African Taphozous (English)	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
African Trident Bat (English)	<i>Cloeotis percivali</i> Thomas, 1901
African Trident Bat (English)	<i>Triaenops afer</i> Peters, 1877
African Trident Bats (English)	<i>Cloeotis</i> Thomas, 1901
African Trident-nosed Bats (English)	<i>Cloeotis</i> Thomas, 1901
African Woolly Bat (English)	<i>Kerivoula africana</i> Dobson, 1878
African Yellow Bat (English)	<i>Scotophilus dinganii</i> (A. Smith, 1833)
African Yellow House Bat (English)	<i>Scotophilus dinganii</i> (A. Smith, 1833)
African Yellow-winged Bat (English)	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
afrifraa (Tanoboase (Ghana))	MOLOSSIDAE Gervais, 1856
Afrikagrotteflygehund (Norwegian)	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Afrikanische Dreizahn-Blattnase (German)	<i>Triaenops afer</i> Peters, 1877
Afrikanische Langflügelfledermaus (German)	<i>Miniopterus africanus</i> Sanborn, 1936
Afrikanische Langohrfledermäuse (German)	<i>Laephotis</i> Thomas, 1901
Afrikanische Langzungen-Flughunde (German)	<i>Megaloglossus</i> Pagenstecher, 1885
Afrikanische Schiebeschwanz-Fledermaus (German)	<i>Coleura afra</i> (Peters, 1852)
Afrikanischer Langzungen-Flughund (German)	<i>Megaloglossus woermanni</i> Pagenstecher, 1885
Afrikanischer Palmenflughund (German)	<i>Eidolon helvum</i> (Kerr, 1792)
Afrikanischer strohfarbener Flughund (German)	<i>Eidolon helvum</i> (Kerr, 1792)
Aguto (Ewe (Ghana))	<i>Eidolon helvum</i> (Kerr, 1792)
Aguto (Ewe (Ghana))	<i>Epomophorus gambianus</i> (Ogilby, 1835)
Aguto (Ewe (Ghana))	<i>Epomops buettikoferi</i> (Matschie, 1899)
Aguto (Ewe (Ghana))	<i>Epomops franqueti</i> (Tomes, 1860)
Ägyptische Bulldoggfledermaus (German)	<i>Tadarida aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
Ägyptische Grabfledermaus (German)	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
Ägyptische Mausschwanz-Fledermaus (German)	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
Ägyptische Schlitznasen-Fledermaus (German)	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
ahwenekron (Tanoboase (Ghana))	<i>Hypsignathus monstrosus</i> H. Allen, 1862
Akdeniz Cüce Yarasası (Turkish)	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Akdeniz Nalburunlu Yarasası (Turkish)	<i>Rhinolophus euryale</i> Blasius, 1853
Aldabra- Dreizahnblattnase (German)	<i>Paratriaenops pauliani</i> (Goodman and Ranivo, 2008)
Aldabra Flying Fox (English)	<i>Pteropus aldabrensis</i> True, 1893
Aldabra Flying-fox (English)	<i>Pteropus aldabrensis</i> True, 1893
Aldabra-Flugfuchs (German)	<i>Pteropus aldabrensis</i> True, 1893
Allen's Butterfly Bat (English)	<i>Glauconycteris alboguttata</i> J.A. Allen, 1917
Allen's Spotted Bat (English)	<i>Glauconycteris humeralis</i> J.A. Allen, 1917
Allen's Striped Bat (English)	<i>Glauconycteris alboguttata</i> J.A. Allen, 1917
Aloe Bat (English)	<i>Neoromicia zuluensis</i> (Roberts, 1924)
Aloe Serotine (English)	<i>Neoromicia zuluensis</i> (Roberts, 1924)
Aloe Serotine Bat (English)	<i>Neoromicia zuluensis</i> (Roberts, 1924)
Alpendwergvleermuis (Dutch)	<i>Hypsugo savii</i> (Bonaparte, 1837)
Alpenfledermaus (German)	<i>Hypsugo savii</i> (Bonaparte, 1837)
Alpen-Fliedermaus (Luxembourgish)	<i>Hypsugo savii</i> (Bonaparte, 1837)
Alpen-Pipistrelle (German)	<i>Hypsugo savii</i> (Bonaparte, 1837)
Alpenvleermuis (Dutch)	<i>Hypsugo savii</i> (Bonaparte, 1837)
Alpenzwergfledermaus (German)	<i>Hypsugo savii</i> (Bonaparte, 1837)
Alpesi denevér (Hungarian)	<i>Hypsugo savii</i> (Bonaparte, 1837)
Alpfladdermus (Swedish)	<i>Hypsugo savii</i> (Bonaparte, 1837)
Alpi nahkhiir (Estonian)	<i>Hypsugo savii</i> (Bonaparte, 1837)
Altwelt-Rundblattnasen (German)	<i>Hipposideros</i> Grav. 1831

Common Name

Ambre-Langflügelfledermaus (German)
 ampane (singular) (Tanoboase (Ghana))
 ampane ankasa (Tanoboase (Ghana))
 ampane kronfo [= "thief or criminal bat"] (see Ohemeng *et al.*, 2017: 184) (Tanoboase (Ghana))
 Anchieta's Bat (English)
 Anchieta's Breitgesicht-Flughund (German)
 Anchieta's Fruit Bat (English)
 Anchieta's Pipistrelle (German)
 Anchieta's Pipistrelle (English)
 Anchieta's Plerote Fruit-bat (English)
 Anchieta's Zwergfledermaus (German)
 Anchieta-vlermuis (Afrikaans)
 Andersens Hufeisennase (German)
 Andersens Schlitznasen-Fledermaus (German)
 Andersen's Slit-faced Bat (English)
 Andrew Rebori's House Bat (English)
 Andrews Hausfledermaus (German)
 Angavo (Malagasy)
 Angola-Bulldogfledermaus (German)
 Angola-Epaulettenflughund (German)
 Angola-langhaarvlermuis (Afrikaans)
 Angola-Langohrfledermaus (German)
 Angola-losstertvlermuis (Afrikaans)
 Angola-Mausohr (German)
 Angolan Epauletted Fruit Bat (English)
 Angolan Epauletted Fruit-bat (English)
 Angolan Free-tailed Bat (English)
 Angolan Fruit Bat (English)
 Angolan Hairy Bat (English)
 Angolan Long-eared Bat (English)
 Angolan Long-haired Fruit-bat (English)
 Angolan Mops Bat (English)
 Angolan Rousette (English)
 Angolan Soft-furred Fruit-bat (English)
 Angolan Wing-gland Bat (English)
 Angolanischer Samtfell-Flughund (German)
 Angolase Losstertvlermuis (Afrikaans)
 Angolase Vlerkkliervlermuis (Afrikaans)
 Angola-Weichfellflughund (German)
 Angola-witkolvrugtevlermuis (Afrikaans)
 angon (Fang)
 Anjouan Mouse-eared Bat (English)
 Anjouan Myotis (English)
 Anjouan-Mausohr (German)
 Ansells Epaulettenflughund (German)
 Ansonge se losstertvlermuis (Afrikaans)
 Ansonge-losstertvlermuis (Afrikaans)
 Ansonges Bulldogfledermaus (German)
 Ansonge's Free-tailed Bat (English)
 Ansonge's Wrinkle-lipped Bat (English)
 apan (singular) (Twi (Ghana))
 Apopo (Kinande (DRC))
 Arabian barbastelle (English)
 Arabian Horseshoe Bat (English)
 Arabische mopsvleermuis (Dutch)
 Ariel Pipistrelle (English)
 Arohautalepakko (Finnish)
 Asellia (French)
 Asellia à trois endentures (French)
 Asellia d'Afrique orientale (French)
 Asellia de Patrizi (French)
 Asèllia di Patrizi (Italian)
 Asèllia italosòmala (Italian)
 Asèllie (Italian)
 Ashman Gadol (Hebrew)
 Ashman Katan (Hebrew)
 Asian Barbastelle (English)
 Asiatic Barbastelle (English)
 Asorflaggermus (Norwegian)
 Assoori videvlane (Estonian)
 Atalef Perot (Hebrew)
 Atalef Yaznuv Gadol (Hebrew)

Taxon Name

Miniopterus ambohitrensis Goodman, Ramasindrazana, Naughton and Appleton, 2015
CHIROPTERA Blumenbach, 1779
Eidolon helvum (Kerr, 1792)
Epomophorus gambianus (Ogilby, 1835)
Neoromicia anchietae (Seabra, 1900)
Plerotes anchietae (Seabra, 1900)
Plerotes anchietae (Seabra, 1900)
Neoromicia anchietae (Seabra, 1900)
Neoromicia anchietae (Seabra, 1900)
Plerotes anchietae (Seabra, 1900)
Neoromicia anchietae (Seabra, 1900)
Neoromicia anchietae (Seabra, 1900)
Rhinolophus eloquens K. Andersen, 1905
Nycteris aurita (K. Andersen, 1912)
Nycteris aurita (K. Andersen, 1912)
Scotophilus andrewreborii Brooks and Bickham, 2014
Scotophilus andrewreborii Brooks and Bickham, 2014
Eidolon dupreanum (Schegel, 1867)
Mops (Mops) condylurus (A. Smith, 1833)
Epomophorus angolensis Gray, 1870
Cistugo seabrae Thomas, 1912
Laephotis angolensis Monard, 1935
Mops (Mops) condylurus (A. Smith, 1833)
Cistugo seabrae Thomas, 1912
Epomophorus angolensis Gray, 1870
Epomophorus angolensis Gray, 1870
Mops (Mops) condylurus (A. Smith, 1833)
Myonycteris angolensis (Bocage, 1898)
Cistugo seabrae Thomas, 1912
Laephotis angolensis Monard, 1935
Myonycteris angolensis (Bocage, 1898)
Mops (Mops) condylurus (A. Smith, 1833)
Myonycteris angolensis (Bocage, 1898)
Myonycteris angolensis (Bocage, 1898)
Cistugo seabrae Thomas, 1912
Myonycteris angolensis (Bocage, 1898)
Mops (Mops) condylurus (A. Smith, 1833)
Cistugo seabrae Thomas, 1912
Myonycteris angolensis (Bocage, 1898)
Epomophorus angolensis Gray, 1870
Doryrhina cyclops (Temminck, 1853)
Myotis anjouanensis Dorst, 1960
Myotis anjouanensis Dorst, 1960
Myotis anjouanensis Dorst, 1960
Epomophorus anselli Bergmans and Van Strien, 2004
Mops (Chaerephon) ansorgei (Thomas, 1913)
Mops (Chaerephon) ansorgei (Thomas, 1913)
Mops (Chaerephon) ansorgei (Thomas, 1913)
Mops (Chaerephon) ansorgei (Thomas, 1913)
Mops (Chaerephon) ansorgei (Thomas, 1913)
CHIROPTERA Blumenbach, 1779
Epomophorus labiatus (Temminck, 1837)
Barbastella leucomelas (Cretzschmar, 1826)
Rhinolophus clivosus Cretzschmar, 1826
Barbastella leucomelas (Cretzschmar, 1826)
Hypsugo ariel (Thomas, 1904)
Taphozous nudiventris Cretzschmar, 1830
Asellia Gray, 1838
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Cloeotis percivali Thomas, 1901
Asellia patrizii de Beaux, 1931
Asellia patrizii de Beaux, 1931
Asellia italosomalica de Beaux, 1931
Asellia Gray, 1838
Taphozous nudiventris Cretzschmar, 1830
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Barbastella leucomelas (Cretzschmar, 1826)
Barbastella leucomelas (Cretzschmar, 1826)
Nyctalus azoreum (Thomas, 1901)
Nyctalus azoreum (Thomas, 1901)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rhinopoma microphyllum (Brünnich, 1782)

Common Name	Taxon Name
Barbastelle d'Allen (French)	<i>Glauconycteris albuguttata</i> J.A. Allen, 1917
Barbastelle de Béatrix (French)	<i>Glauconycteris beatrix</i> Thomas, 1901
Barbastelle de Glen (French)	<i>Glauconycteris gleni</i> Peterson and Smith, 1973
Barbastelle d'Europe (French)	<i>Barbastella barbastellus</i> (Schreber, 1774)
Barbastelle du Bibundi (French)	<i>Glauconycteris egeria</i> Thomas, 1913
Barbastelle du Kenya (French)	<i>Glauconycteris kenyacola</i> Peterson, 1982
Barbastelle occidentale (French)	<i>Barbastella barbastellus</i> (Schreber, 1774)
Barbastelle orientale (French)	<i>Barbastella leucomelas</i> (Cretzschmar, 1826)
Barbastelles (English)	<i>Barbastella</i> Gray, 1821
Barbastelles (French)	<i>Barbastella</i> Gray, 1821
Barbastello (Italian)	<i>Barbastella barbastellus</i> (Schreber, 1774)
Bartfledermaus (German)	<i>Myotis mystacinus</i> (Kuhl, 1817)
Basıkburunlu Yarasa (Turkish)	<i>Barbastella barbastellus</i> (Schreber, 1774)
Baso-saguzar (Basque)	<i>Barbastella barbastellus</i> (Schreber, 1774)
Baso-saguzar arrunt (Basque)	<i>Barbastella barbastellus</i> (Schreber, 1774)
Bates' Schlitznasen-Fledermaus (German)	<i>Nycteris arge</i> Thomas, 1903
Bates's Slit-faced Bat (English)	<i>Nycteris arge</i> Thomas, 1903
Bats (English)	CHIROPTERA Blumenbach, 1779
Beatrix Butterfly Bat (English)	<i>Glauconycteris beatrix</i> Thomas, 1901
Beatrix's Bat (English)	<i>Glauconycteris beatrix</i> Thomas, 1901
bejložraví netopýři (Czech)	PTEROPODIDAE Gray, 1821
Belarrihandi kanariar (Basque)	<i>Plecotus teneriffae</i> Barrett-Hamilton, 1907
Belorobi netopir (Slovenian)	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Bemmelens Bulldoggfledermaus (German)	<i>Mops (Chaerephon) bemmeleni</i> (Jentink, 1879)
Benguela Fruit-bat (English)	<i>Plerotes anchietae</i> (Seabra, 1900)
Benito Leaf-nosed Bat (English)	<i>Hipposideros beatus</i> (K. Andersen, 1906)
Benito Roundleaf Bat (English)	<i>Hipposideros beatus</i> (K. Andersen, 1906)
Benito-Blattnase (German)	<i>Hipposideros beatus</i> (K. Andersen, 1906)
Benito-Rundblattnase (German)	<i>Hipposideros beatus</i> (K. Andersen, 1906)
Bent-winged bat (English)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Bent-winged Bats (English)	<i>Miniopterus</i> Bonaparte, 1837
Bergmans Harlekin-Flughund (German)	<i>Scotonycteris bergmansi</i> Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015
Bergmans's Collared Fruit Bat (English)	<i>Myonycteris relicta</i> Bergmans, 1980
Bergmans's fruit bat (English)	<i>Scotonycteris bergmansi</i> Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015
Bergmans's Small Fruit Bat (English)	<i>Myonycteris relicta</i> Bergmans, 1980
Beyazşeritli Yarasa (Turkish)	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Bibundi Bat (English)	<i>Glauconycteris egeria</i> Thomas, 1913
Bibundi Butterfly Bat (English)	<i>Glauconycteris egeria</i> Thomas, 1913
Bibundi Schmetterlingsfledermaus (German)	<i>Glauconycteris egeria</i> Thomas, 1913
Big-brown bats (English)	<i>Eptesicus Rafinesque</i> , 1820
Big-eared Bats (English)	<i>Plecotus</i> E. Geoffroy St.-Hilaire, 1818
Big-eared Bulldog Bats (English)	<i>Otomops</i> Thomas, 1913
Big-eared Free-tailed Bat (English)	<i>Tadarida lobata</i> (Thomas, 1891)
Big-eared Free-tailed Bats (English)	<i>Otomops</i> Thomas, 1913
Big-eared Guano Bat (English)	<i>Tadarida lobata</i> (Thomas, 1891)
Big-eared Kenya Free-tailed Bat (English)	<i>Tadarida lobata</i> (Thomas, 1891)
Big-eared Kenyan Guano Bat (English)	<i>Tadarida lobata</i> (Thomas, 1891)
Big-eared leaf-nosed bat (English)	<i>Hipposideros megalotis</i> (Heuglin, 1861)
Big-eared slit-faced bat (English)	<i>Nycteris macrotis</i> Dobson, 1876
Bijelorubi slijepi mišić (Montenegrin)	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Bini Free-tailed Bat (English)	<i>Myopterus whiteleyi</i> (Scharff, 1900)
Bini Pergamentflügel-Fledermaus (German)	<i>Myopterus whiteleyi</i> (Scharff, 1900)
Bini Winged-mouse Bat (English)	<i>Myopterus whiteleyi</i> (Scharff, 1900)
Bjelorubi šišmiš (Croatian)	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Blāzija pakavdegunis (Latvian)	<i>Rhinolophus blasii</i> Peters, 1867
Black and Red Free-tailed Bat (English)	<i>Mops (Chaerephon) jobimena</i> (Goodman and Cardiff, 2004)
Black Clinging Bat (English)	<i>Miniopterus fraterculus</i> Thomas and Schwann, 1906
Black Flying- fox (English)	<i>Pteropus livingstonii</i> Gray, 1866
Blackish Butterfly Bat (English)	<i>Glauconycteris atra</i> Hassanin, Colombo, Gembu, Merle, Tu, Görföl, Musaba Akawa, Csorba, Kearney, Monadjem and Ing, 2017
Bladneusvleermuizen (Dutch)	HIPPOSIDERIDAE Lydekker, 1891
Blasijev podkovnjak (Slovenian)	<i>Rhinolophus blasii</i> Peters, 1867
Blasius ferra-saguzar (Basque)	<i>Rhinolophus blasii</i> Peters, 1867
Blasius' hestekonæse (Danish)	<i>Rhinolophus blasii</i> Peters, 1867
Blasius' hoefijzerneus (Dutch)	<i>Rhinolophus blasii</i> Peters, 1867
Blasius' hoefizernoas (Frisian)	<i>Rhinolophus blasii</i> Peters, 1867
Blasius Nalburunlu Yarasa (Turkish)	<i>Rhinolophus blasii</i> Peters, 1867
Blasius-hoefijzerneus (Dutch)	<i>Rhinolophus blasii</i> Peters, 1867

Common Name

Blasius-Hufeisennase (German)
 Blasius-Huffeisennues (Luxembourgish)
 Blasius's Horseshoe Bat (English)
 Blazijaus pasagnosis (Lithuanian)
 Blazijev potkovasti ššmiš (Bosnian)
 Blazijev potkovnjak (Croatian)
 Blazius nalburunu (Azerbaijani)
 Bleek losstertvlermuis (Afrikaans)
 Blek sydfladdermuis (Swedish)
 Blekflaggermus (Norwegian)
 Blue-grey Bats (English)
 Blumaf Shahor (Hebrew)
 Bocage Banana Bat (English)
 Bocage se vrugtevlermuis (Afrikaans)
 Bocage's Free-tailed Bat (English)
 Bocage's Fruit-bat (English)
 Bocage's Hairy Bat (English)
 Bocage's Mouse-eared Bat (English)
 Bocage's Losstertvlermuis (Afrikaans)
 Bocage-vrugtevlermuis (Afrikaans)
 Bonneted Bats (English)
 Bonneted Bats (English)
 Borowiaczek (Polish)
 Borowiec azorski (Polish)
 Borowiec olbrzymi (Polish)
 Boskflearmûs (Frisian)
 Bosveldsaalneusvlermuis (Afrikaans)
 Bosveldvlermuis (Afrikaans)
 Bosvleermuis (Dutch)
 Botas sikspārnis (Latvian)
 Botin šišmiš (Bosnian)
 Botswana Long-eared Bat (English)
 Botswana-Langohrfledermaus (German)
 Botswana-langoorvlermuis (Afrikaans)
 Botswanan Long-eared Bat (English)
 Botta serotino (Basque)
 Botta'nin Genişkanatlı Yarasası (Turkish)
 Botta-Fledermaus (Luxembourgish)
 Botta-késeidenevér (Hungarian)
 Bottas Breitflügelfledermaus (German)
 Bottas Fledermaus (German)
 Botta's lette flearmûs (Frisian)
 Botta's Serotine (English)
 Botta's Serotine Bat (English)
 Bottas sydflagermus (Danish)
 Botta's vleermuis (Dutch)
 Bottin noćnjak (Croatian)
 Bouchaara (Arabian)
 Boujlida (Arabian)
 Bourbon Bat (English)
 Bourbonische Schwirrfledermaus (German)
 Böyük nalburun (Azerbaijani)
 Bredøre (Norwegian)
 Bredøret flaggermus (Danish)
 Breitflügelfledermäuse (German)
 Breitgesicht-Flughunde (German)
 Breitköpfige Pipistrelle (German)
 Breitköpfige Zwergfledermaus (German)
 Brivastes sikspārnis (Latvian)
 Brkati šišmiš (Croatian)
 Brkati netopir (Slovenian)
 Brkati večernjak (Montenegrin)
 Broad-headed Pipistrelle (English)
 Broad-headed Pipistrelle Bat (English)
 Brown bats (English)
 Brown House Bat (English)
 Brown Pipistrelle Bat (English)
 Brunnich's Mouse-tailed Bat (English)
 Buettikofer's Clamorous Fruit-bat (English)
 Buettikofer's Epauletted Bat (English)
 Buettikofer's Epauletted Fruit Bat (English)
 Bükükdodaq enliqulaq (Azerbaijani)
 buldočí pochvovrepí (Czech)
 Buldoggflaggermus (Norwegian)

Taxon Name

Rhinolophus blasii Peters, 1867
Rhinolophus blasii Peters, 1867
Rhinolophus blasii Peters, 1867
Rhinolophus blasii Peters, 1867
Rhinolophus blasii Peters, 1867
Rhinolophus blasii Peters, 1867
Rhinolophus blasii Peters, 1867
Mops (Chaerephon) chapini (J.A. Allen, 1917)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Glauconycteris Dobson, 1875
Barbastella leucomelas (Cretzschmar, 1826)
Myotis bocagii (Peters, 1870)
Myonycteris angolensis (Bocage, 1898)
Tadarida aegyptiaca bocagei (Seabra, 1900)
Myonycteris angolensis (Bocage, 1898)
Myotis bocagii (Peters, 1870)
Myotis bocagii (Peters, 1870)
Tadarida aegyptiaca bocagei (Seabra, 1900)
Myonycteris angolensis (Bocage, 1898)
 MOLOSSIDAE Gervais, 1856
 Molossinae Gervais, 1856
Nyctalus leisleri (Kuhl, 1817)
Nyctalus azoreum (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus simulator K. Andersen, 1904
Rhinolophus simulator K. Andersen, 1904
Nyctalus leisleri (Kuhl, 1817)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Laephotis botswanae Setzer, 1971
Laephotis botswanae Setzer, 1971
Laephotis botswanae Setzer, 1971
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
 CHIROPTERA Blumenbach, 1779
 CHIROPTERA Blumenbach, 1779
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Rhinolophus ferrumequinum (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Eptesicus Rafinesque, 1820
Plerotes K. Andersen, 1910
Nycticeinops crassulus (Thomas, 1904)
Nycticeinops crassulus (Thomas, 1904)
Tadarida teniotis (Rafinesque, 1814)
Myotis mystacinus (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Nycticeinops crassulus (Thomas, 1904)
Nycticeinops crassulus (Thomas, 1904)
Scotophilus Leach, 1821
Scotophilus nigrita (Schreber, 1774)
Pseudoromicia brunnea (Thomas, 1880)
Rhinopoma microphyllum (Brünnich, 1782)
Epomops buettikoferi (Matschie, 1899)
Epomops buettikoferi (Matschie, 1899)
Epomops buettikoferi (Matschie, 1899)
Tadarida teniotis (Rafinesque, 1814)
Coleura Peters, 1867
 MOLOSSIDAE Gervais, 1856

Common Name

Bulldogvleermuisen (Dutch)
 Bulldog Bats (English)
 Bulldog Bats (English)
 Bulldoggfledermaus (German)
 Bulldoggfledermäuse (German)
 Bulldogg-Fledermäuse (German)
 Bulveermuis (Dutch)
 Bunte Wollfledermaus (German)
 Burdflearmûs (Frisian)
 Buschland-Rundblattnase (German)
 Buschveld-Hufeisennase (German)
 Bushveld Horseshoe Bat (English)
 Butterfly Bat (English)
 Butterfly Bats (English)
 Büttikofer's Epaulettenflughund (German)
 Büttikofer's Fruit Bat (English)
 Büyük Akşamcı Yarasa (Turkish)
 Büyük Nalburunlu Yarasa (Turkish)
 Çıplakkarınlı Mezar Yarasa (Turkish)
 Cırtan şəbperə (Azerbaijani)
 Cənub nalburunu (Azerbaijani)
 Cabeza Martillo (Castilian (Spain))
 Cabrera's rondbladnase (Dutch)
 Cameroon Leaf-nosed Bat (English)
 Campo-Ma'an fruit bat (English)
 Campomaan-Flughund (German)
 Canarische grootoorvleermuis (Dutch)
 Canary big-eared bat (English)
 Canary Long-eared Bat (English)
 Canon Smith's Rousette (English)
 Capaccinii's Mouse-eared Bat (English)
 Capaccinis flagermus (Danish)
 Capaccini's vleermuis (Dutch)
 Cape bat (English)
 Cape Hairy Bat (English)
 Cape Hairy Myotis (English)
 Cape Horseshoe Bat (English)
 Cape House Bat (English)
 Cape Leaf-nosed bat (English)
 Cape Long-eared Bat (English)
 Cape Myotis (English)
 Cape Pipistrelle Bat (English)
 Cape Serotine (English)
 Cape Serotine Bat (English)
 Capische Abendfledermaus (German)
 Capische Trugfledermaus (German)
 Capischen Kamnase (German)
 Cardioderma (Italian)
 Cardioderma nàso a cuore (Italian)
 Cardioderme à nez en cœur (French)
 Cardiodermes (French)
 Carlino brachittero (Italian)
 Carlino condiluro (Italian)
 Carlino Mida (Italian)
 Carlino setoloso (Italian)
 Casinyctère (French)
 Casinyctère (French)
 Casinyctère de Campo-Ma'an (French)
 Casinyctère dorée (French)
 Central Africa Pipistrelle (English)
 Centralafrikanische Doggengrämler (German)
 Chagrin Fledermaus (German)
 Chapins Bulldoggfledermaus (German)
 Chapin's Free-tailed Bat (English)
 Chapin's Wrinkle-lipped Bat (English)
 Chaun sgulant da l'Egipta (Rhaeto-Romance)
 Chauve-souris à 3 endentures (French)
 Chauve-souris à épaulettes de Gambie (Ewe (Ghana))
 Chauve-souris à face creuse (French)
 Chauve-souris à fer à cheval (French)
 Chauve-souris à longues oreilles d'Angola (French)
 Chauve-souris à longues oreilles de De Winton (French)
 Chauve-souris à longues oreilles de Namibie (French)
 Chauve-souris à longues oreilles du Botswana (French)

Taxon Name

MOLOSSIDAE Gervais, 1856
 MOLOSSIDAE Gervais, 1856
 Molossinae Gervais, 1856
 Tadarida teniotis (Rafinesque, 1814)
 MOLOSSIDAE Gervais, 1856
 MOLOSSIDAE Gervais, 1856
 Tadarida teniotis (Rafinesque, 1814)
 Kerivoula argentata Toms, 1861
 Myotis mystacinus (Kuhl, 1817)
 Rhinolophus simulator K. Andersen, 1904
 Rhinolophus simulator K. Andersen, 1904
 Rhinolophus simulator K. Andersen, 1904
 Glauconycteris variegata (Toms, 1861)
 Glauconycteris Dobson, 1875
 Epomops buettikoferi (Matschie, 1899)
 Epomops buettikoferi (Matschie, 1899)
 Nyctalus lasiopterus (Schreber, 1780)
 Rhinolophus ferrumequinum (Schreber, 1774)
 Taphozous nudiventris Cretzschmar, 1830
 Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
 Rhinolophus euryale Blasius, 1853
 Hypsignathus monstrosus H. Allen, 1862
 Hipposideros tephros (Cabrera, 1906)
 Doryrhina camerunensis (Eisentraut, 1956)
 Casinycteris campomaanensis Hassanin, 2014
 Casinycteris campomaanensis Hassanin, 2014
 Plecotus teneriffae Barrett-Hamilton, 1907
 Plecotus teneriffae Barrett-Hamilton, 1907
 Plecotus teneriffae Barrett-Hamilton, 1907
 Myonycteris angolensis (Bocage, 1898)
 Myotis capaccinii (Bonaparte, 1837)
 Myotis capaccinii (Bonaparte, 1837)
 Myotis capaccinii (Bonaparte, 1837)
 Laephotis capensis (A. Smith, 1829)
 Myotis tricolor (Temminck, 1832)
 Myotis tricolor (Temminck, 1832)
 Rhinolophus capensis Lichtenstein, 1823
 Laephotis capensis (A. Smith, 1829)
 Hipposideros caffer (Sundevall, 1846)
 Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
 Myotis tricolor (Temminck, 1832)
 Laephotis capensis (A. Smith, 1829)
 Laephotis capensis (A. Smith, 1829)
 Laephotis capensis (A. Smith, 1829)
 Laephotis capensis (A. Smith, 1829)
 Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
 Rhinolophus capensis Lichtenstein, 1823
 Cardioderma Peters, 1873
 Cardioderma cor (Peters, 1872)
 Cardioderma cor (Peters, 1872)
 Cardioderma Peters, 1873
 Mops (Xiphonycteris) brachypterus (Peters, 1852)
 Mops (Mops) condylurus (A. Smith, 1833)
 Mops (Mops) midas (Sundevall, 1843)
 Platymops setiger (Peters, 1878)
 Casinycteris Thomas, 1910
 Casinycteris argynnis Thomas, 1910
 Casinycteris campomaanensis Hassanin, 2014
 Casinycteris argynnis Thomas, 1910
 Pipistrellus (Pipistrellus) inexpectatus Aellen, 1959
 Mops (Mops) condylurus (A. Smith, 1833)
 Eptesicus (Rhinopterus) floweri (de Winton, 1901)
 Mops (Chaerephon) chapini (J.A. Allen, 1917)
 Mops (Chaerephon) chapini (J.A. Allen, 1917)
 Mops (Chaerephon) chapini (J.A. Allen, 1917)
 Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
 Asellia Gray, 1838
 Epomophorus gambianus (Ogilby, 1835)
 NYCTERIDAE Van der Hoeven, 1855
 RHINOLOPHIDAE Gray, 1825
 Laephotis angolensis Monard, 1935
 Laephotis wintoni Thomas, 1901
 Laephotis namibensis Setzer, 1971
 Laephotis botswanae Setzer, 1971

Common Name

Chauve-souris à oreilles en trompette de Spurell (French)
 Chauve-souris à oreilles en trompette de Spurell (French)
 Chauve-souris à oreilles pointues de Hemprich (French)
 Chauve-souris à pieds à ventouses de Madagascar (French)

Chauve-souris à queue de souris (French)
 Chauve-souris à queue en fourreau d'Afrique (French)
 Chauve-souris à queue en fourreau des Seychelles (French)
 Chauve-souris à queue gainée d'Afrique (French)
 Chauve-souris à queue gainée de Pel (French)
 Chauve-souris à queue gainée des Seychelles (French)
 chauve-souris à tête de marteau (French)
 Chauve-souris à tête plate (French)
 Chauve-souris à tête plate de Moloney (French)
 Chauve-souris à ventouses de Schliemann (French)

Chauve-souris bouledogue géante (French)
 Chauve-souris de Commerson à nez feuillu (French)

Chauve-souris de Commerson à nez feuillu (French)
 Chauve-souris de Schlieffen (French)
 Chauve-souris de Schreiber (French)
 Chauve-souris de Thomas (French)
 Chauve-souris de Veldkamp (French)
 chauve-souris des Hauts (French)
 Chauve-souris des Tombes (French)
 Chauve-souris des tombes (French)
 Chauve-souris des tombes égyptiennes (French)
 Chauve-souris domestique malgache (French)
 Chauve-souris du crépuscule (French)
 Chauve-souris frugivore à collier d'Afrique orientale (French)
 Chauve-souris frugivore à collier de São Tomé (French)
 Chauve-souris frugivore à palais court (French)
 Chauve-souris frugivore d'Anchieta (French)
 Chauve-souris jaune des maisons (French)
 Chauve-souris laineuse (French)
 Chauve-souris laineuse cuivrée (French)
 Chauve-souris laineuse d'Ethiopie (French)
 Chauve-souris laineuse du Damara (French)
 Chauve-souris malgache à oreille de souris (French)
 Chauve-souris malgache à pieds à ventouses (French)

Chauve-souris malgache à queue en fourreau (French)
 Chauve-souris mauritienne des tombeaux (French)
 chauve-souris paillée (Ewe (Ghana))
 Chauve-souris papillon (French)
 Chauve-souris peinte argentée (French)
 Chauve-souris peinte cuivrée (French)
 Chauve-souris peinte de Smith (French)
 Chauve-souris peinte de Tanzanie (French)
 Chauve-souris peinte d'Ethiopie (French)
 Chauve-souris peinte phalène (French)
 Chauve-souris réticulée (French)
 Chauve-souris trident (French)
 Chauve-souris vesper (French)
 Chauves-souris (French)
 Chauves-souris à épaulettes (French)
 Chauves-souris à grandes oreilles (French)
 Chauves-souris à longues oreilles (French)
 Chauves-souris à nez large (French)
 Chauves-souris à queue de souris (French)
 Chauves-souris à queue gainée (French)
 Chauves-souris à queue libre (French)
 Chauves-souris à ventouses (French)
 Chauves-souris des maisons (French)
 Chauves-souris frugivore de l'ancien monde (French)
 Chauves-souris peintes (French)
 Cherefónte bivittato (Italian)
 Cherefónte di Ansörge (Italian)
 Cherefónte nigeriàno (Italian)
 Cherefónte pigmèo (Italian)
 Cherefónti (Italian)
 Cherivoula di Smith (Italian)
 Cherivoule (Italian)

Taxon Name

Kerivoula phalaena Thomas, 1912
Kerivoula phalaena Thomas, 1912
Otonycteris hemprichii Peters, 1859
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Rhinopoma cystops Thomas, 1903
Coleura afra (Peters, 1852)
Coleura seychellensis Peters, 1868
Coleura afra (Peters, 1852)
Saccolaimus peli (Temminck, 1853)
Coleura seychellensis Peters, 1868
Hypsignathus monstrosus H. Allen, 1862
Mimetillus Thomas, 1904
Mimetillus moloneyi (Thomas, 1891)
Myzopoda schliemanni Goodman, Rakotondraparany and Kofoky, 2007
Otomops martiensseni (Matschie, 1897)
Macronycteris commersoni (E. Geoffroy St.-Hilaire, 1813)
Macronycteris gigas (Wagner, 1845)
Nycticeinops schlieffenii (Peters, 1859)
Miniopterus schreibersii (Kuhl, 1817)
Scotoecus albofuscus (Thomas, 1890)
Nanonycteris veldkampii (Jentink, 1888)
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Taphozous E. Geoffroy St.-Hilaire, 1818
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Scotophilus robustus A. Milne-Edwards, 1881
Nycticeinops schlieffenii (Peters, 1859)
Myonycteris relicta Bergmans, 1980
Myonycteris brachycephala (Bocage, 1889)
Casinycteris argynnis Thomas, 1910
Plerotes anchietae (Seabra, 1900)
Scotophilus dinganii (A. Smith, 1833)
Kerivoula lanosa (A. Smith, 1847)
Kerivoula cuprosa Thomas, 1912
Kerivoula eriophora (Heuglin, 1877)
Kerivoula argentata Tomes, 1861
Myotis goudoti (A. Smith, 1834)
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Paremballonura atrata (Peters, 1874)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Eidolon helvum (Kerr, 1792)
Glauconycteris variegata (Tomes, 1861)
Kerivoula argentata Tomes, 1861
Kerivoula cuprosa Thomas, 1912
Kerivoula smithii Thomas, 1880
Kerivoula africana Dobson, 1878
Kerivoula eriophora (Heuglin, 1877)
Kerivoula phalaena Thomas, 1912
Glauconycteris variegata (Tomes, 1861)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
 VESPERTILIONIDAE Gray, 1821
 CHIROPTERA Blumenbach, 1779
Epomophorus Bennett, 1836
Plecotus E. Geoffroy St.-Hilaire, 1818
Laephotis Thomas, 1901
Nyctalus Bowdich, 1825
 RHINOPOMATIDAE Dobson, 1872
Coleura Peters, 1867
 MOLOSSIDAE Gervais, 1856
Myzopoda Milne-Edwards and A. Grandidier, 1878
Scotoecus Thomas, 1901
 PTEROPODIDAE Gray, 1821
Kerivoula Gray, 1842
Mops (Chaerephon) bivittatus (Heuglin, 1861)
Mops (Chaerephon) ansorgei (Thomas, 1913)
Mops (Chaerephon) nigeriae (Thomas, 1913)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Mops (Chaerephon) Dobson, 1874
Kerivoula smithii Thomas, 1880
Kerivoula Gray, 1842

Common Name

Taxon Name

Chien volant à épaulettes du Congo (Ewe (Ghana))	<i>Epomops franqueti</i> (Tomes, 1860)
Chien volant à tête en marteau (French)	<i>Hypsignathus monstrosus</i> H. Allen, 1862
Chiròtteri (Italian)	CHIROPTERA Blumenbach, 1779
Christie's Long-eared Bat (English)	<i>Plecotus christii</i> Gray, 1838
Christis Graues Langohr (German)	<i>Plecotus christii</i> Gray, 1838
Chuchu (Manganja (Malawi))	<i>Afronycteris nana</i> (Peters, 1852)
cíponos africký (Czech)	<i>Triaenops afer</i> Peters, 1877
cíponos aldabránský (Czech)	<i>Paratriaenops pauliani</i> (Goodman and Ranivo, 2008)
cíponos jihoafrický (Czech)	<i>Cloeotis percivali</i> Thomas, 1901
cíponos madagaskarský (Czech)	<i>Triaenops menamena</i> Goodman and Ranivo, 2009
cíponos malý (Czech)	<i>Paratriaenops furculus</i> (Trouessart, 1907)
cíponos zlatý (Czech)	<i>Paratriaenops auritus</i> (G. Grandidier, 1912)
Clamorous Fruit-bats (English)	<i>Epomops</i> Gray, 1866
Cléothe de Percival (French)	<i>Cloeotis percivali</i> Thomas, 1901
Cléotheres (French)	<i>Cloeotis</i> Thomas, 1901
Cloeotide di Percival (Italian)	<i>Cloeotis percivali</i> Thomas, 1901
Cloeotidi (Italian)	<i>Cloeotis</i> Thomas, 1901
Cohen's Horseshoe Bat (English)	<i>Rhinolophus cohenae</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
	<i>Rhinolophus cohenae</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Cohens Hufeisennase (German)	<i>Coleura afra</i> (Peters, 1852)
Còleura africana (Italian)	<i>Myonycteris</i> Matschie, 1899
Collared Fruit-bats (English)	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Commerson se bladneusvlermuis (Afrikaans)	<i>Macronycteris gigas</i> (Wagner, 1845)
Commerson se bladneusvlermuis (Afrikaans)	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Commerson-bladneusvlermuis (Afrikaans)	<i>Macronycteris gigas</i> (Wagner, 1845)
Commerson's Leaf-nosed Bat (English)	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Commerson's Leaf-nosed Bat (English)	<i>Macronycteris gigas</i> (Wagner, 1845)
Commerson's Rhinoloph (English)	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Commerson's Rhinoloph (English)	<i>Macronycteris gigas</i> (Wagner, 1845)
Commerson's roundleaf bat (English)	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Commerson's roundleaf bat (English)	<i>Macronycteris gigas</i> (Wagner, 1845)
Commersonse Blaarneusvlermuis (Afrikaans)	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Commersonse Blaarneusvlermuis (Afrikaans)	<i>Macronycteris gigas</i> (Wagner, 1845)
Common African Leaf-nosed Bat (English)	<i>Hipposideros caffer</i> (Sundevall, 1846)
Common Barbastelle (English)	<i>Barbastella barbastellus</i> (Schreber, 1774)
Common Bent-winged Bat (English)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Common Butterfly Bat (English)	<i>Glauconycteris argentata</i> (Dobson, 1875)
Common Long-fingered Bat (English)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Common Pipistrelle (English)	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Common Pipistrelle Bat (English)	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Common Slit-faced Bat (English)	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
Comores Flying Fox (English)	<i>Pteropus livingstonii</i> Gray, 1866
Comoro Black Flying Fox (English)	<i>Pteropus livingstonii</i> Gray, 1866
Comoro Flying Fox (English)	<i>Pteropus livingstonii</i> Gray, 1866
Comoro Rousette (English)	<i>Rousettus obliviosus</i> Kock, 1978
Congo Horseshoe Bat (English)	<i>Rhinolophus adami</i> Aellen and Brosset, 1968
Copper Woolly Bat (English)	<i>Kerivoula cuprosa</i> Thomas, 1912
Crested Free-tail Bat (English)	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
Crested Wrinkle-lipped Bat (English)	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
Cretzschmar's Brown Bat (English)	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
Cretzschmar's Horseshoe bat (English)	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
Crudh-ialtag bheag (Scottish Gaelic)	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Crudh-ialtag Blasii (Scottish Gaelic)	<i>Rhinolophus blasii</i> Peters, 1867
Crudh-ialtag Meadhan-threach (Scottish Gaelic)	<i>Rhinolophus euryale</i> Blasius, 1853
Crudh-ialtag Mehelyi (Scottish Gaelic)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Crudh-ialtag mhòr (Scottish Gaelic)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Crú-ialtóg bheag (Irish Gaelic)	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Crú-ialtóg Blasius (Irish Gaelic)	<i>Rhinolophus blasii</i> Peters, 1867
Crú-ialtóg Mehelyi (Irish Gaelic)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Crú-ialtóg Mheánmhuirí (Irish Gaelic)	<i>Rhinolophus euryale</i> Blasius, 1853
Crú-ialtóg mhór (Irish Gaelic)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Csonkafülű denevér (Hungarian)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Csúcsosnyergű patkósdenevér (Hungarian)	<i>Rhinolophus blasii</i> Peters, 1867
Csupaszhasú sírlakó-denevér (Hungarian)	<i>Taphozous nudiventris</i> Cretzschmar, 1830
Cüce Yarasa (Turkish)	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Curry's Bat (English)	<i>Glauconycteris curryae</i> Eger and Schlitter, 2001

Common Name

Taxon Name

Curry's Butterfly Bat (English)	<i>Glauconycteris curryae</i> Eger and Schlitter, 2001
Currys Schmetterlingsfledermaus (German)	<i>Glauconycteris curryae</i> Eger and Schlitter, 2001
Cyclops Bat (English)	<i>Doryrhina cyclops</i> (Temminck, 1853)
Cyclops Leaf-nosed Bat (English)	<i>Doryrhina cyclops</i> (Temminck, 1853)
Cyclops Roundleaf Bat (English)	<i>Doryrhina cyclops</i> (Temminck, 1853)
Damara Woolly Bat (English)	<i>Kerivoula argentata</i> Tomes, 1861
Damara-Hufeisennase (German)	<i>Rhinolophus damarensis</i> Roberts, 1946
Damaralandse Wolhaarvlermuis (Afrikaans)	<i>Kerivoula argentata</i> Tomes, 1861
Damara-wolhaarvlermuis (Afrikaans)	<i>Kerivoula argentata</i> Tomes, 1861
D'Anchieta's Fruit Bat (English)	<i>Plerotes anchietae</i> (Seabra, 1900)
Dar-es-salaam Pipistrelle (English)	<i>Pipistrellus (Pipistrellus) permixtus</i> Aellen, 1957
Dar-es-Salaam Pipistrelle Bat (English)	<i>Pipistrellus (Pipistrellus) permixtus</i> Aellen, 1957
Dark Flying Fox (English)	<i>Pteropus subniger</i> (Kerr, 1792)
Dark House Bat (English)	<i>Scotophilus nux</i> Thomas, 1904
Dark-brown Pipistrelle Bat (English)	<i>Pseudoromicia brunnea</i> (Thomas, 1880)
Dark-brown Serotine (English)	<i>Pseudoromicia brunnea</i> (Thomas, 1880)
Dark-winged Lesser House Bat (English)	<i>Scotoecus hirundo</i> (de Winton, 1899)
Darling se saalneusvlermuis (Afrikaans)	<i>Rhinolophus darlingi</i> K. Andersen, 1905
Darling's Horseshoe Bat (English)	<i>Rhinolophus darlingi</i> K. Andersen, 1905
Darlings Hufeisennase (German)	<i>Rhinolophus darlingi</i> K. Andersen, 1905
Darling-saalneusvlermuis (Afrikaans)	<i>Rhinolophus darlingi</i> K. Andersen, 1905
dasíci (Czech)	<i>Barbastella</i> Gray, 1821
dasík arabský (netopýr skalní) (Czech)	<i>Barbastella leucomelas</i> (Cretzschmar, 1826)
Daubenton's Free-tailed Bat (English)	<i>Myotis daubentonii</i> Desmarest, 1820
Daubentons Pergamentflügel-Fledermaus (German)	<i>Myotis daubentonii</i> Desmarest, 1820
Daubenton's Winged-mouse Bat (English)	<i>Myotis daubentonii</i> Desmarest, 1820
De Beaux's Trident Bat (English)	<i>Asellia italosomalica</i> de Beaux, 1931
de Wintonovi netopýři (Czech)	<i>Laephotis</i> Thomas, 1901
De Wintons Langohrfledermaus (German)	<i>Laephotis wintoni</i> Thomas, 1901
De Winton's Long-eared Bat (English)	<i>Laephotis wintoni</i> Thomas, 1901
Decken's Horseshoe Bat (English)	<i>Rhinolophus deckenii</i> Peters, 1868
Deckens Hufeisennase (German)	<i>Rhinolophus deckenii</i> Peters, 1868
Dent se saalneusvlermuis (Afrikaans)	<i>Rhinolophus denti</i> Thomas, 1904
Dent's Horseshoe Bat (English)	<i>Rhinolophus denti</i> Thomas, 1904
Dents Hufeisennase (German)	<i>Rhinolophus denti</i> Thomas, 1904
Dent-saalneusvlermuis (Afrikaans)	<i>Rhinolophus denti</i> Thomas, 1904
Dentse Vlermuis (Afrikaans)	<i>Rhinolophus denti</i> Thomas, 1904
Desert Bats (English)	<i>Otonycteris</i> Peters, 1859
Desert Big-eared Bats (English)	<i>Otonycteris</i> Peters, 1859
Desert Long-eared bat (English)	<i>Otonycteris hemprichii</i> Peters, 1859
Desert Long-eared Bats (English)	<i>Otonycteris</i> Peters, 1859
Desert Pipistrelle (English)	<i>Hypsugo ariel</i> (Thomas, 1904)
Didysis nakviša (Lithuanian)	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Didysis pasagnosis (Lithuanian)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Dingaan's House Bat (English)	<i>Scotophilus dinganii</i> (A. Smith, 1833)
Dingaan's Yellow Bat (English)	<i>Scotophilus dinganii</i> (A. Smith, 1833)
Dingan's Bat (English)	<i>Scotophilus dinganii</i> (A. Smith, 1833)
Dja Schlitznasen-Fledermaus (German)	<i>Nycteris major</i> (K. Andersen, 1912)
Dja Slit-faced Bat; Large Slit-faced Bat (English)	<i>Nycteris major</i> (K. Andersen, 1912)
Dja-Schlitznasenfledermaus (German)	<i>Nycteris major</i> (K. Andersen, 1912)
dlouhojazyční kaloni (Czech)	<i>Megaloglossus</i> Pagenstecher, 1885
Dobson se vrugtevlermuis (Afrikaans)	<i>Epomophorus dobsonii</i> Bocage, 1889
Dobson's Bats (English)	<i>Glauconycteris</i> Dobson, 1875
Dobson's Epauletted Bat (English)	<i>Epomophorus dobsonii</i> Bocage, 1889
Dobson's Epauletted Fruit Bat (English)	<i>Epomophorus dobsonii</i> Bocage, 1889
Dobsons Epaulettenflughund (German)	<i>Epomophorus dobsonii</i> Bocage, 1889
Dobson's Fruit Bat (English)	<i>Epomophorus dobsonii</i> Bocage, 1889
Dobson-vrugtevlermuis (Afrikaans)	<i>Epomophorus dobsonii</i> Bocage, 1889
Doggilepakko (Finnish)	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Doggilepakot (Finnish)	<i>MOLOSSIDAE</i> Gervais, 1856
Dolgokrilest Liljak (Macedonian)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Dolgokrili netopir (Slovenian)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Dolgonogi netopir (Slovenian)	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Dolgorepi netopir (Slovenian)	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Dongola-Dämmerungsfledermaus (German)	<i>Vansonia rueppellii</i> (J.B. Fischer, 1829)
Dreifarb-Mausohr (German)	<i>Myotis tricolor</i> (Temminck, 1832)
Dreizackblattnase (German)	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
Dreizackblattnasen (German)	<i>Asellia</i> Gray, 1838
Dreizahn-Blattnasen (German)	<i>Triaenops</i> Dobson, 1871
Dreyzahnige Kammnase (German)	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
Drietand-blaarneusvlermuis (Afrikaans)	<i>Cloeotis percivali</i> Thomas, 1901
Drietandbladneusvlermuis (Dutch)	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
Drietand-bladneusvlermuis (Afrikaans)	<i>Cloeotis percivali</i> Thomas, 1901
Drobni netopir (Slovenian)	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)

Common Name

Dugokrili pršnjak (Croatian)
 Dugokrili slijepi miš (Bosnian)
 Dugonogi šišmiš (Croatian)
 Dugoprsti šišmiš (Bosnian)
 Dugoprsti šišmiš (Croatian)
 Dugoprsti večernjak (Montenegrin)
 Dugorepi slijepi miš (Montenegrin)
 Dugorili prstenjak (Montenegrin)
 Duke of Abruzzi's Free-tailed Bat (English)
 Duke of Abruzzi's Wrinkle-lipped Bat (English)
 Dunkelbraune Pipistrelle (German)
 Dunkelbraune Zwergfledermaus (German)
 Dunkle Schmetterlingsfledermaus (German)

Dusky leaf-nosed bat (English)
 dusky pipistrelle (English)
 dutonosec (Czech)
 dutonosecové (Czech)
 Dværgflagermus (Danish)
 Dværgflagermus (Danish)
 Dvaerghesteskonaes (Danish)
 Dvärgfladdermus (Swedish)
 Dvärghästskonäsa (Swedish)
 Dvärgpipistrell (Swedish)
 Dvergflaggermus (Norwegian)
 Dvergflaggermus (Norwegian)
 Dverghesteskonese (Norwegian)
 Dwarf Epauletted Bat (English)
 Dwarf Epauletted Fruit Bat (English)
 Dwarf Free-tailed Bat (English)
 Dwarf Fruit-bats (English)
 Dwarf Leaf-nosed Bat (English)
 Dwarf Mops Bat (English)
 Dwarf Pipistrelle Bat (English)
 Dwarf Slit-faced Bat (English)
 Dwarsoorvleermuis (Dutch)
 Dzudzest Liljak (Macedonian)
 East African Collared Fruit-bat (English)
 East African Epauletted Fruit Bat (English)
 East African Epauletted Fruit-bat (English)
 East African Flat-headed Bat (English)
 East African Flat-headed Bats (English)
 East African Little Collared Fruit Bat (English)
 East African Serotine (English)

East African Trident Bat (English)
 Eastern Africa Free-tailed Bat (English)
 Eastern Africa horseshoe bat (English)
 Eastern Barbastelle (English)
 Eastern Barbastelle Bat (English)
 Eastern Sucker-footed Bat (English)

Eger-Langflügelfledermaus (German)

Eger's long-fingered bat (English)

Egipč anski leteči pes (Slovenian)
 Egipatski letipis (Bosnian)
 Egipatski letipis (Montenegrin)
 Egipatski plodojedi šišmiš (Croatian)
 Egiptiese losstertvlermuis (Afrikaans)
 Egiptiese spleetneusvlermuis (Afrikaans)
 Egiptiese vrugtevlermuis (Afrikaans)
 Egiptiese witylvlermuis (Afrikaans)
 Egiptinis vaisėdis (Lithuanian)
 Egiptuse öötiibur (Estonian)
 egyptští kaloni (Czech)
 Egyptian Desert Pipistrelle (English)
 Egyptian Free-tailed Bat (English)
 Egyptian Fruit Bat (English)
 Egyptian Guano Bat (English)
 Egyptian Mouse-tailed bat (English)
 Egyptian Nyctinome (English)

Taxon Name

Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Myotis capaccinii (Bonaparte, 1837)
Myotis capaccinii (Bonaparte, 1837)
Myotis capaccinii (Bonaparte, 1837)
Myotis capaccinii (Bonaparte, 1837)
Tadarida teniotis (Rafinesque, 1814)
Miniopterus schreibersii (Kuhl, 1817)
Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
Pseudoromicia brunnea (Thomas, 1880)
Pseudoromicia brunnea (Thomas, 1880)
Glauconycteris atra Hassanin, Colombo, Gembu, Merle, Tu, Görföl, Musaba Akawa, Csorba, Kearney, Monadjem and Ing, 2017
Hipposideros fuliginosus (Temminck, 1853)
Pipistrellus (Pipistrellus) hesperidus (Temminck, 1840)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris G. Cuvier and E. Geoffroy, 1795
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus hipposideros (Bechstein, 1800)
Myonycteris Matschie, 1899
Epomophorus pusillus Peters, 1868
Mops (Xiphonycteris) nanulus J.A. Allen, 1917
Nanonycteris Matschie, 1899
Hipposideros beatus (K. Andersen, 1906)
Mops (Xiphonycteris) nanulus J.A. Allen, 1917
Pipistrellus (Pipistrellus) nanulus Thomas, 1904
Nycteris nana (K. Andersen, 1912)
Barbastella barbastellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myonycteris relicta Bergmans, 1980
Epomophorus minimus Claessen & De Vree, 1991
Epomophorus minimus Claessen & De Vree, 1991
Platymops setiger (Peters, 1878)
Platymops Thomas, 1906
Myonycteris relicta Bergmans, 1980
Laephotis kirinyaga Monadjem, Patterson, Webala and Demos, 2020
Cloeotis percivali Thomas, 1901
Tadarida lobata (Thomas, 1891)
Rhinolophus deckenii Peters, 1868
Barbastella leucomelas (Cretzschmar, 1826)
Barbastella leucomelas (Cretzschmar, 1826)
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Miniopterus egeri Goodman, Ramasindrazana, Maminirina, Schoeman and Appleton, 2011
Miniopterus egeri Goodman, Ramasindrazana, Maminirina, Schoeman and Appleton, 2011
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus Gray, 1821
Hypsugo ariel (Thomas, 1904)
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Rhinopoma cystops Thomas, 1903
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)

Common Name

Egyptian Rousette (English)
 Egyptian Rousette Bat (English)
 Egyptian sheath-tailed bat (English)
 Egyptian Slit-faced Bat (English)
 Egyptian Tomb Bat (English)
 Egyptian Xantharpy (English)
 Egyptische grafvleermuis (Dutch)
 Egyptische grootoorvleermuis (Dutch)
 Egyptischer fliegender Hund (German)
 Egyptische bulvleermuis (Dutch)
 Egyptisk flyvehund (Danish)
 Èidoli (Italian)
 Èidolo paglierino (Italian)
 Eidolon fruit bats (English)
 Eiropas platausis (Latvian)
 Eisentraut's Leaf-nosed Bat (English)
 Eisentrauts Pipistrelle (German)
 Eisentraut's pipistrelle (English)
 Eisentraut's Pipistrelle Bat (English)
 Eisentrauts Zwergfledermaus (German)
 Ejeta's House Bat (English)
 Ekiptiya Vāarra Velāvā (Tamil)
 Ekte hestekonese (Norwegian)
 Elfen-Pipistrelle (German)
 Elfen-Zwergfledermaus (German)
 Eloquent Horseshoe Bat (English)
 Emballonure d'Afrique (French)
 Emballonure de Madagascar (French)

 Emballonure de Madagascar (French)
 Emballonure des rochers (French)

 Emballonure des Seychelles (French)
 Emballonuridés (French)
 Emballonuridi (Italian)
 embalonuroviti (Czech)
 Epaulet bats (English)
 Epauletted bats (English)
 Epauletted Fruit-bats (English)
 Epaulettenflughund (German)
 Epaulettenflughunde (German)
 Epaulettenflughunde (German)
 Epomòfori (Italian)
 Epomòforo di Wählberg (Italian)
 Epomòforo labiàto (Italian)
 Epomòforo minimo (Italian)
 Epomophore d'Angola (French)
 Epomophore d'Ansell (French)
 Epomophore de Buettikofer (Ewe (Ghana))
 Epomophore de Dobson (French)
 Epomophore de Peters (French)
 Epomophore de Sanborn (French)
 Epomophore d'Ethiopie (French)
 Epomophore frugivore de Wahlberg (French)
 Epomophore nain (French)
 Epomophores (French)
 Epomophores de l'ouest (French)
 Ethiopian Big-eared Bat (English)
 Ethiopian Epauletted Fruit Bat (English)
 Ethiopian Epauletted Fruit-bat (English)
 Ethiopian Large-eared Leaf-nosed Bat (English)
 Ethiopian Large-eared Roundleaf Bat (English)
 Ethiopian Long-eared Bat (English)
 Ethiopian Woolly Bat (English)
 Eurasian Free-tailed Bat (English)
 Euroopa kurdmokk (Estonian)
 Euroopa laikõrv (Estonian)
 Euroopa pikktiib (Estonian)
 Euroopanisoherkko (Finnish)
 Euroopanpikkuherkko (Finnish)
 Europæisk bulldogflagermus (Danish)
 Europäesch Bulldogfledermaus (Luxembourgish)
 Europäesch Mopsfledermaus (Luxembourgish)

Taxon Name

Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Taphozous nudiventris Cretzschmar, 1830
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Plecotus christii Gray, 1838
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Eidolon Rafinesque, 1815
Eidolon helvum (Kerr, 1792)
Eidolon Rafinesque, 1815
Barbastella barbastellus (Schreber, 1774)
Doryrhina camerunensis (Eisentraut, 1956)
Nycticeinops eisentrauti (Hill, 1968)
Nycticeinops eisentrauti (Hill, 1968)
Nycticeinops eisentrauti (Hill, 1968)
Nycticeinops eisentrauti (Hill, 1968)
Scotophilus ejetai Brooks and Bickham, 2014
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
 RHINOLOPHIDAE Gray, 1825
Hypsugo ariel (Thomas, 1904)
Hypsugo ariel (Thomas, 1904)
Rhinolophus eloquens K. Andersen, 1905
Coleura afra (Peters, 1852)
Coleura kibomalandy Goodman, Puechmaile, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012
Paremballonura atrata (Peters, 1874)
Paremballonura tiavato (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)
Coleura seychellensis Peters, 1868
 EMBALLONURIDAE Gervais, 1855
 EMBALLONURIDAE Gervais, 1855
 EMBALLONURIDAE Gervais, 1855
Epomops Gray, 1866
Epomops Gray, 1866
Epomophorus Bennett, 1836
Epomophorus Bennett, 1836
Epomophorus Bennett, 1836
Epomops Gray, 1866
Epomophorus Bennett, 1836
Epomophorus wahlbergi (Sundevall, 1846)
Epomophorus labiatus (Temminck, 1837)
Epomophorus minimus Claessen & De Vree, 1991
Epomophorus angolensis Gray, 1870
Epomophorus anselli Bergmans and Van Strien, 2004
Epomops buettikoferi (Matschie, 1899)
Epomophorus dobsonii Bocage, 1889
Epomophorus crypturus Peters, 1852
Epomophorus grandis (Sanborn, 1950)
Epomophorus labiatus (Temminck, 1837)
Epomophorus wahlbergi (Sundevall, 1846)
Epomophorus minimus Claessen & De Vree, 1991
Epomophorus Bennett, 1836
Epomops Gray, 1866
Plecotus balensis Kruskop & Lavrenchenko, 2000
Epomophorus labiatus (Temminck, 1837)
Epomophorus labiatus (Temminck, 1837)
Hipposideros megalotis (Heuglin, 1861)
Hipposideros megalotis (Heuglin, 1861)
Plecotus balensis Kruskop & Lavrenchenko, 2000
Kerivoula eriphora (Heuglin, 1877)
Tadarida teniotis (Rafinesque, 1814)
Tadarida teniotis (Rafinesque, 1814)
Barbastella barbastellus (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Tadarida teniotis (Rafinesque, 1814)
Tadarida teniotis (Rafinesque, 1814)
Barbastella barbastellus (Schreber, 1774)

Common Name

Taxon Name

Európai hosszúszárnyú-denevér (Hungarian)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Európai szabadfarkú-denevér (Hungarian)	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Europäische Bulldoggfledermaus (German)	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Europäische Sackfledermaus (German)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
European Free-tailed Bat (English)	<i>Tadarida teniotis</i> (Rafinesque, 1814)
European Guano Bat (English)	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Europeisk vikvinge (Swedish)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Europese bulvleermuis (Dutch)	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Europinis plačiaausis (Lithuanian)	<i>Barbastella barbastellus</i> (Schreber, 1774)
Europinis raukšlėtaiŲpis (Lithuanian)	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Evening bats (English)	VESPERTILIONIDAE Gray, 1821
Evropski dugokrlaš (Serbian Latin)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Eynekli nalburun (Azerbaijani)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Fairy Pipistrelle (English)	<i>Hypsugo ariel</i> (Thomas, 1904)
Falsche Vampire (German)	MEGADERMATIDAE H. Allen, 1864
False Vampire Bats (English)	MEGADERMATIDAE H. Allen, 1864
Fàlsi vampiri (Italian)	MEGADERMATIDAE H. Allen, 1864
Fàlsi vampiri d'alle ali gialle (Italian)	<i>Lavia</i> Gray, 1838
falske hestekoneser (Norwegian)	HIPPOSIDERIDAE Lydekker, 1891
falske vampyrer (Norwegian)	MEGADERMATIDAE H. Allen, 1864
Fàlso vampire d'alle ali gialle (Italian)	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
Fàlso vampiro d'alle ali gialle (Italian)	<i>Cardioderma cor</i> (Peters, 1872)
Fanihy (Malagasy)	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
Farfett il-Lejl Daqni (Maltese)	<i>Myotis mystacinus</i> (Kuhl, 1817)
Farfett il-Lejl Ghieri (Maltese)	<i>Taphozous nudiventris</i> Cretzschmar, 1830
Farfett il-Lejl ta' Widnejh Imnaqqxa (Maltese)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Farfett il-Lejl ta' Xrajber (Maltese)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Farfett il-Lejl tal-Frott ta' L-Egittu (Maltese)	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Farfett il-Lejl tan-Nala (Maltese)	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Farfett il-Lejl tan-Nala kbir (Maltese)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Farfett il-Lejl tas-Swaba Twal (Maltese)	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Farfett il-Lejl Widnejh Twal tad-Deżert (Maltese)	<i>Otonycteris hemprichii</i> Peters, 1859
Farfett il-Lejl Widnejh Twal tal-Gżejjer Kanarji (Maltese)	<i>Plecotus teneriffae</i> Barrett-Hamilton, 1907
Farfett il-Lejl Widnet il-gurdien ta' Maghreb (Maltese)	<i>Myotis punicus</i> Felten, 1977
Farfett il-Lejl Widnet il-Gurdien (Maltese)	<i>Myotis punicus</i> Felten, 1977
Faux vampire de l'ancien monde (French)	MEGADERMATIDAE H. Allen, 1864
Fehérszélű törpedenevér (Hungarian)	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Felten's Myotis (English)	<i>Myotis punicus</i> Felten, 1977
Fenicische vale vleermuis (Dutch)	<i>Myotis punicus</i> Felten, 1977
Fer à cheval d'Hildebrandt (French)	<i>Rhinolophus hildebrandtii</i> Peters, 1878
Fer à cheval du Kenya (French)	<i>Rhinolophus hildebrandtii</i> Peters, 1878
Fer-à-Cheval d'Afrique orientale (French)	<i>Rhinolophus deckenii</i> Peters, 1868
Fer-à-Cheval de Darling (French)	<i>Rhinolophus darlingi</i> K. Andersen, 1905
Ferra-saguzar handi (Basque)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Ferra-saguzar mediterraneo (Basque)	<i>Rhinolophus euryale</i> Blasius, 1853
Fèrri di cavàllo (Italian)	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Fèrro di cavàllo affumicàto (Italian)	<i>Rhinolophus Lacépède</i> , 1799
Ferro di cavallo di Blàsius (Italian)	<i>Rhinolophus fumigatus</i> Rüppell, 1842
Fèrro di cavàllo di Cretzschmar (Italian)	<i>Rhinolophus blasii</i> Peters, 1867
Fèrro di cavàllo di Dècken (Italian)	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
Fèrro di cavàllo di Hildebrandt (Italian)	<i>Rhinolophus deckenii</i> Peters, 1868
Ferro di cavallo di Méhely (Italian)	<i>Rhinolophus hildebrandtii</i> Peters, 1878
Fèrro di cavàllo eloquente (Italian)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Ferro di cavallo Euriale (Italian)	<i>Rhinolophus eloquens</i> K. Andersen, 1905
Ferro di Cavallo Maggiore (Italian)	<i>Rhinolophus euryale</i> Blasius, 1853
ferro di cavallo mediterraneo (Italian)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Ferro di Cavallo Minore (Italian)	<i>Rhinolophus euryale</i> Blasius, 1853
Fèrro di cavàllo simulatòre (Italian)	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Fers-à-cheval (French)	<i>Rhinolophus simulator</i> K. Andersen, 1904
Fers-à-cheval (French)	RHINOLOPHIDAE Gray, 1825
Feuille de Daubenton (French)	<i>Rhinolophus Lacépède</i> , 1799
Fid Mer (Somalian)	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
Fidmair (Somalian)	CHIROPTERA Blumenbach, 1779
Fidmer (Somalian)	CHIROPTERA Blumenbach, 1779
Finikinis peléausis (Lithuanian)	CHIROPTERA Blumenbach, 1779
Flachkopf-Bulldoggfledermäuse (German)	<i>Myotis punicus</i> Felten, 1977
Flachkopf-Bulldoggfledermäuse (German)	<i>Platymops</i> Thomas, 1906
Flachköpfige Dämmerungsfledermaus (German)	<i>Sauromys</i> Roberts, 1917
Flachschnauzigen Schwirrfledermaus (German)	<i>Pipistrellus (Pipistrellus) hesperidus</i> (Temminck, 1840)
Flat-faced Serotine Bat (English)	<i>Scotophilus dinganii</i> (A. Smith, 1833)
Flat-headed Bats (English)	<i>Eptesicus (Cnephæus) platyops</i> (Thomas, 1901)
Flat-headed Bats (English)	<i>Mormopterus</i> Peters, 1865
Flat-headed Bulldog Bats (English)	<i>Mimetillus</i> Thomas, 1904
	<i>Mormopterus</i> Peters, 1865

Common Name

Flat-headed free-tailed bat (English)
 Flat-headed Free-tailed Bat (English)
 Flat-headed Vesper Bats (English)
 Flattnasenfreischwänze (German)
 Flederhunde (German)
 Fledermaus mit gerändelten Ohren (German)
 Fledermäuse (German)
 Flikörad fladdermus (Swedish)
 Flitter Mouse (English)
 Flower's Pipistrelle Bat (English)
 Flugfuchse (German)
 Flughunde (German)
 Flying Calf (English)
 Flying Foxes (English)
 Flying-foxes (English)
 Flyvende hunder (Norwegian)
 foldeleppede flaggermus (Norwegian)
 Forest Bats (English)
 Forest Horseshoe Bat (English)
 Franquet's Clamorous Fruit-bat (English)
 Franquet's Epauletted Fruit Bat (English)
 Franquet's Epaulettenflugfuchs (German)
 Franquets Epaulettenflughund (German)
 Franquet's Fruit Bat (English)
 frede frede (Tanoboase (Ghana))
 Free-tailed Bats (English)
 Free-tailed Bats (English)
 Free-tailed Bats (English)
 Freischeinige Stelfussfledermaus (German)
 French:: Epomophore de Büttikofer (Ewe (Ghana))
 French:: Epomophore de Franquet (Ewe (Ghana))
 French:: Epomophore de Gambie (Ewe (Ghana))
 French:: Roussette jaune (Ewe (Ghana))
 Frigibell Blasius (Breton)
 Frigibell greizdouarel (Breton)
 Frigibell Méhely (Breton)
 Frigibell vihan (Breton)
 Frigibell vras (Breton)
 Frihalete flaggermus (Norwegian)
 Fritu-saguzar egiptoar (Basque)
 Fulo (Azande (DRC))
 Fulo (Azande (DRC))
 Fulo (Azande (DRC))
 Fürst von Aburzen Bulldoggfledermaus (German)
 Gacek kanaryjski (Polish)
 Gaislers grootoorvleermuis (Dutch)
 Gaislers Langohrfledermaus (German)
 Gaisler's Long-eared Bat (English)
 Gallaghers Bulldoggfledermaus (German)
 Gallagher's Free-tailed Bat (English)
 Gallagher's Wrinkle-lipped Bat (English)
 Gambia Epaulettenflughund (German)
 Gambia Schlitznasen-Fledermaus (German)
 Gambiaanse witkolvrugtevlermuis (Afrikaans)
 Gambian Epauletted Bat (English)
 Gambian Epauletted Fruit bat (English)
 Gambian Slit-faced Bat (English)
 Garpirkstu naktssikspārnīs (Latvian)
 Gau-saguzar azoretar (Basque)
 Gau-saguzar handi (Basque)
 Geat Brown Bat (English)
 Gebirgs-Höhlenflughund (German)
 Geel Dakvlermuis (Afrikaans)
 Geel vrugtevlermuis (Afrikaans)
 Gefleckte Bulldoggfledermaus (German)
 Gefleckte Bulldoggfledermaus (German)
 Gefleckte Schmetterlingsfledermaus (German)
 Gelbbäuchige Hausfledermaus (German)
 Gelbbäuchige Schwirrfledermaus (German)
 Gelbe Pipistrelle (German)
 Gelbe Zwergfledermaus (German)
 Gelbflügel-Fledermaus (German)
 Gelbflügel-Fledermäuse (German)
 Gelbflügelige Großblattnase (German)

Taxon Name

Platymops setiger (Peters, 1878)
Sauromys petrophilus (Roberts, 1917)
Mimetillus Thomas, 1904
 EMBALLONURIDAE Gervais, 1855
 PTEROPODIDAE Gray, 1821
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
 CHIROPTERA Blumenbach, 1779
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Pteropus Brisson, 1762
 PTEROPODIDAE Gray, 1821
Nanonycteris veldkampii (Jentink, 1888)
Pteropus Brisson, 1762
Pteropus Brisson, 1762
 PTEROPODIDAE Gray, 1821
 MOLOSSIDAE Gervais, 1856
Kerivoulinae Miller, 1907
Rhinolophus silvestris Aellen, 1959
Epomops franqueti (Tomes, 1860)
Epomops franqueti (Tomes, 1860)
Epomops franqueti (Tomes, 1860)
Epomops franqueti (Tomes, 1860)
Epomops franqueti (Tomes, 1860)
 MOLOSSIDAE Gervais, 1856
 MOLOSSIDAE Gervais, 1856
Molossinae Gervais, 1856
Tadarida Rafinesque, 1814
Myotis capaccinii (Bonaparte, 1837)
Epomops buettikoferi (Matschie, 1899)
Epomops franqueti (Tomes, 1860)
Epomophorus gambianus (Ogilby, 1835)
Eidolon helvum (Kerr, 1792)
Rhinolophus blasii Peters, 1867
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus ferrumequinum (Schreber, 1774)
 EMBALLONURIDAE Gervais, 1855
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Mops (Chaerephon) nigeriae (Thomas, 1913)
Afronycteris nana (Peters, 1852)
Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
Plecotus teneriffae Barrett-Hamilton, 1907
Plecotus gaisleri Benda, Kiefer, Hanak & Veith, 2004
Plecotus gaisleri Benda, Kiefer, Hanak & Veith, 2004
Plecotus gaisleri Benda, Kiefer, Hanak & Veith, 2004
Mops (Chaerephon) gallagheri (Harrison, 1975)
Mops (Chaerephon) gallagheri (Harrison, 1975)
Mops (Chaerephon) gallagheri (Harrison, 1975)
Epomophorus gambianus (Ogilby, 1835)
Nycteris gambiensis (K. Andersen, 1912)
Epomophorus gambianus (Ogilby, 1835)
Epomophorus gambianus (Ogilby, 1835)
Epomophorus gambianus (Ogilby, 1835)
Nycteris gambiensis (K. Andersen, 1912)
Myotis capaccinii (Bonaparte, 1837)
Nyctalus azoreum (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1774)
Scotophilus nigrita (Schreber, 1774)
Stenonycteris lanosus (Thomas, 1906)
Scotophilus dinganii (A. Smith, 1833)
Eidolon helvum (Kerr, 1792)
Mops (Chaerephon) bivittatus (Heuglin, 1861)
Mops (Mops) leucostigma (G.M. Allen, 1918)
Glauconycteris humeralis J.A. Allen, 1917
Scotophilus dinganii (A. Smith, 1833)
Scotophilus leucogaster (Cretzschmar, 1826)
Nycticeinops grandidieri (Dobson, 1876)
Nycticeinops grandidieri (Dobson, 1876)
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Lavia Gray, 1838
Lavia frons (E. Geoffroy St.-Hilaire, 1810)

Common Name

Gemeine Langflügelfledermaus (German)
 Gemeine Schlitznasen-Fledermaus (German)
 Genetzte Schmetterlingsfledermaus (German)
 Geoffroy saguzar (Basque)
 Geoffroy se saalneusvlermuis (Afrikaans)
 Geoffroyflaggermus (Norwegian)
 Geoffroy's Bat (English)
 Geoffroys Dreizackblattnase (German)
 Geoffroys flagermus (Danish)
 Geoffroy's hoefijzermuis (Dutch)
 Geoffroy's Horseshoe Bat (English)
 Geoffroys Hufeisennase (German)
 Geoffroy's Lesser Tident Bat (English)
 Geoffroy's Myotis (English)
 Geoffroy's Nycteris (English)
 Geoffroy's Tomb Bat (English)
 Geoffroy's Trident Leaf-nosed Bat (English)
 Geoffroy-saalneusvlermuis (Afrikaans)
 Geoffroyse vlermuis (Afrikaans)
 Gesäumte Doggengrämmler (German)
 Gestreifte Schmetterlingsfledermaus (German)
 Gevlekte losstertvlermuis (Afrikaans)
 Gewoane dwerchflearmûs (Frisian)
 Gewöhnliche Bulldoggfledermäuse (German)
 Gewöhnliche Rundblattnase (German)
 Gewone dwergvleermuis (Dutch)
 Gewone pisang blad vleermuis (Dutch)
 Gewone-spleetneusvlermuis (Afrikaans)
 Giant African Free-tailed Bat (English)
 Giant African Guano Bat (English)
 Giant Brown Bat (English)
 Giant Brown House Bat (English)
 Giant Free-tailed Bat (English)
 Giant gland-tailed bat (English)
 Giant Guano Bat (English)
 Giant House Bat (English)
 Giant Leaf-nosed Bat (English)

Giant Leaf-nosed Bat (English)
 Giant Mastiff Bat (English)
 Giant noctule (English)
 Giant Noctule Bat (English)
 Giant Yellow House Bat (English)
 Gladneusvleermuizen (Dutch)
 Gland-tailed Free-tailed Bat (English)
 Gland-tailed Wrinkle-lipped Bat (English)
 Glattnasen (German)
 Glattnasen-Fledermäuse (German)
 Glattnasen-Freischwänze (German)
 Glattneseflaggermus (Norwegian)
 Glattneser (Norwegian)
 Glauconicteride argentata (Italian)
 Glauconicteride keniota (Italian)
 Glauconicteride variegata (Italian)
 Glauconitteride argentata (Italian)
 Glauconitteride keniota (Italian)
 Glauconitteride variegata (Italian)
 Glauconittèridi (Italian)
 Glauconyctère à gorge blanche (French)
 Glauconyctère argentée (French)
 Glauconyctère d'Abo (French)
 Glauconyctère de Béatrix (French)
 Glauconyctère de Bibundi (French)
 Glauconyctère de Curry (French)
 Glauconyctère de Glen (French)
 Glauconyctère de Machado (French)
 Glauconyctère du Kenya (French)
 Glauconyctère papillon (French)
 Glauconyctère pie (French)
 Glauconyctère sombre (French)

Glauconyctère tachetée (French)
 Glauconyctères (French)

Taxon Name

Miniopterus schreibersii (Kuhl, 1817)
Nycteris hispida (Schreber, 1774)
Glauconycteris variegata (Tomes, 1861)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Rhinolophus clivosus Cretzschmar, 1828
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus clivosus Cretzschmar, 1828
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus clivosus Cretzschmar, 1828
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Glauconycteris alboguttata J.A. Allen, 1917
Mops (Chaerephon) bivittatus (Heuglin, 1861)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Tadarida Rafinesque, 1814
Hipposideros caffer (Sundevall, 1846)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myotis mystacinus (Kuhl, 1817)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Tadarida ventralis (Heuglin, 1861)
Tadarida ventralis (Heuglin, 1861)
Scotophilus nigrita (Schreber, 1774)
Scotophilus nigrita (Schreber, 1774)
Tadarida ventralis (Heuglin, 1861)
Mops (Chaerephon) major (Trouessart, 1897)
Tadarida ventralis (Heuglin, 1861)
Scotophilus nigrita (Schreber, 1774)
Macronycteris commersoni (E. Geoffroy St.-Hilaire, 1813)
Macronycteris gigas (Wagner, 1845)
Otomops martiensseni (Matschie, 1897)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus lasiopterus (Schreber, 1780)
Scotophilus nigrita (Schreber, 1774)
 VESPERTILIONIDAE Gray, 1821
Mops (Chaerephon) bemmeleni (Jentink, 1879)
Mops (Chaerephon) bemmeleni (Jentink, 1879)
 VESPERTILIONIDAE Gray, 1821
 VESPERTILIONIDAE Gray, 1821
 EMBALLONURIDAE Gervais, 1855
 VESPERTILIONIDAE Gray, 1821
 VESPERTILIONIDAE Gray, 1821
Glauconycteris argentata (Dobson, 1875)
Glauconycteris kenyacola Peterson, 1982
Glauconycteris variegata (Tomes, 1861)
Glauconycteris argentata (Dobson, 1875)
Glauconycteris kenyacola Peterson, 1982
Glauconycteris variegata (Tomes, 1861)
Glauconycteris Dobson, 1875
Glauconycteris alboguttata J.A. Allen, 1917
Glauconycteris argentata (Dobson, 1875)
Glauconycteris poensis (Gray, 1842)
Glauconycteris beatrix Thomas, 1901
Glauconycteris egeria Thomas, 1913
Glauconycteris curryae Eger and Schlitter, 2001
Glauconycteris gleni Peterson and Smith, 1973
Glauconycteris machadoi Hayman, 1963
Glauconycteris kenyacola Peterson, 1982
Glauconycteris variegata (Tomes, 1861)
Glauconycteris superba Hayman, 1939
Glauconycteris atra Hassanin, Colombo, Gembu, Merle, Tu, Görföl, Musaba Akawa, Csorba, Kearney, Monadjem and Ing, 2017
Glauconycteris humeralis J.A. Allen, 1917
Glauconycteris Dobson, 1875

Common Name

Glauconycteris (English)
 Gleichsatteligen Kammnase (German)
 Glen's Butterfly Bat (English)
 Glens Langflügelfledermaus (German)
 Glen's Long-fingered Bat (English)
 Glens Schmetterlingsfledermaus (German)
 Glen's Wattled Bat (English)
 Goblin bats (English)
 Golden Bat (English)

 Golden Short-palate Fruit-bat (English)
 Golden Short-palated Fruit Bat (English)
 Goldener Kurzgaumen-Flughund (German)
 Golorepi šišmiš (Bosnian)
 Golotrbi grobni šišmiš (Croatian)
 Golotrbusni grobni šišmiš (Bosnian)
 Golotrbusni grobni slijepi miš (Montenegrin)
 Golotrebusni mavzolejnik (Slovenian)
 Goodman's Trident Bat (English)
 Gorongosa-Hufeisennase (German)

 Gousper Savi (Breton)
 Gousperell ar Magreb (Breton)
 Gousperell Capaccini (Breton)
 Gousperell Geoffroy (Breton)
 Gousperell varvek (Breton)
 Gozdni mračnik (Slovenian)
 Grabflatterer (German)
 Grabfledermäuse (German)
 Grand Fer-à-Cheval (French)
 Grand minioptère (French)
 Grand minioptère africain (French)
 Grand minioptère malgache (French)
 Grand molosse à glandes caudales (French)
 Grand molosse à grandes oreilles (French)
 Grand rhinolophe (French)
 Grand Rhinopome (French)
 Grand scotophile africain (French)
 Grande noctule (French)
 Grande nyctère (French)
 grande Roussette des Mascareignes (French)
 Grande tadaride africaine (French)
 Grande Tadaride de Madagascar (French)
 Grandidier's Pipistrelle (English)
 Grandidiars Bulldoggfledermaus (German)
 Grandidiars Dreizahn-Blattnase (German)
 Grandidier's Free-tailed Bat (English)
 Grandidier's trident bat (English)
 Grandidiars Zwergfledermaus (German)
 gravflaggermus (Norwegian)
 Grayovi kaloni (Czech)
 Great Brown House Bat (English)
 Great pipistrelle (English)
 Greater Bent-winged Bat (English)
 Greater Cyclops (English)
 Greater Free-tailed Bats (English)
 Greater Horseshoe Bat (English)
 Greater Long-fingered Bat (English)
 Greater Mascarene Flying Fox (English)
 Greater Mouse-tailed Bat (English)
 Greater noctule (English)
 Greater Noctule Bat (English)
 Greater Rat-tailed Bat (English)
 Greater Roundleaf Bat (English)
 Greater Serotine Bats (English)
 Greater Slit-faced Bat (English)
 Greater Trident Bats (English)
 Green House Bat (English)
 Greenish Yellow Bat (English)
 Griffiths Langflügelfledermaus (German)

 Griveaud's Long-fingered bat (English)
 Grobownik golobrzuchy (Polish)

Taxon Name

Glauconycteris Dobson, 1875
Rhinolophus euryale Blasius, 1853
Glauconycteris gleni Peterson and Smith, 1973
Miniopterus gleni Peterson, Eger and Mitchell, 1995
Miniopterus gleni Peterson, Eger and Mitchell, 1995
Glauconycteris gleni Peterson and Smith, 1973
Glauconycteris gleni Peterson and Smith, 1973
Mormopterus Peters, 1865
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Casinycteris argynnis Thomas, 1910
Casinycteris argynnis Thomas, 1910
Casinycteris argynnis Thomas, 1910
Tadarida teniotis (Rafinesque, 1814)
Taphozous nudiventris Cretzschmar, 1830
Taphozous nudiventris Cretzschmar, 1830
Taphozous nudiventris Cretzschmar, 1830
Taphozous nudiventris Cretzschmar, 1830
Triaenops goodmani Samonds, 2007
Rhinolophus gorongosae Taylor, MacDonald, Goodman, Kearney, Cotterill, Stoffberg, Monadjem, Schoeman, Guyton, Naskrecki and Richards, 2018
Hypsugo savii (Bonaparte, 1837)
Myotis punicus Felten, 1977
Myotis capaccinii (Bonaparte, 1837)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis mystacinus (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Taphozous E. Geoffroy St.-Hilaire, 1818
Taphozous E. Geoffroy St.-Hilaire, 1818
Rhinolophus ferrumequinum (Schreber, 1774)
Miniopterus inflatus Thomas, 1903
Miniopterus inflatus Thomas, 1903
Miniopterus majori Thomas, 1906
Mops (Chaerephon) major (Trouessart, 1897)
Otomops martiensseni (Matschie, 1897)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinopoma microphyllum (Brünnich, 1782)
Scotophilus nigrita (Schreber, 1774)
Nyctalus lasiopterus (Schreber, 1780)
Nycteris grandis Peters, 1865
Pteropus niger (Kerr, 1792)
Tadarida ventralis (Heuglin, 1861)
Tadarida fulminans (Thomas, 1903)
Nycticeinops grandidieri (Dobson, 1876)
Mops (Chaerephon) leucogaster (A. Grandidier, 1869)
Paratriaenops auritus (G. Grandidier, 1912)
Mops (Chaerephon) leucogaster (A. Grandidier, 1869)
Paratriaenops auritus (G. Grandidier, 1912)
Nycticeinops grandidieri (Dobson, 1876)
 EMBALLONURIDAE Gervais, 1855
Epomops Gray, 1866
Scotophilus nigrita (Schreber, 1774)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Doryrhina camerunensis (Eisentraut, 1956)
Mops Lesson, 1842
Rhinolophus ferrumequinum (Schreber, 1774)
Miniopterus inflatus Thomas, 1903
Pteropus niger (Kerr, 1792)
Rhinopoma microphyllum (Brünnich, 1782)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus lasiopterus (Schreber, 1780)
Rhinopoma microphyllum (Brünnich, 1782)
Doryrhina camerunensis (Eisentraut, 1956)
Eptesicus Rafinesque, 1820
Nycteris macrotis Dobson, 1876
Triaenops Dobson, 1871
Scotophilus viridis (Peters, 1852)
Scotophilus viridis (Peters, 1852)
Miniopterus griffithsi Goodman, Maminirina, Bradman, Christidis and Appleton, 2009
Miniopterus griveaudi Harrison, 1959
Taphozous nudiventris Cretzschmar, 1830

Common Name	Taxon Name
Groot geel dakvlermuis (Afrikaans)	<i>Scotophilus nigrata</i> (Schreber, 1774)
Groot grotvlermuis (Afrikaans)	<i>Miniopterus inflatus</i> Thomas, 1903
Groot spleetneusvlermuis (Afrikaans)	<i>Nycteris grandis</i> Peters, 1865
Grootoor-loosstervlermuis (Afrikaans)	<i>Tadarida lobata</i> (Thomas, 1891)
Großabendsegler (German)	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Grossblattnasen (German)	MEGADERMATIDAE H. Allen, 1864
Großblattnasen (German)	MEGADERMATIDAE H. Allen, 1864
Große Bulldoggfledermaus (German)	<i>Tadarida ventralis</i> (Heuglin, 1861)
Große Hufeisennase (German)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Große Schlitznase (German)	<i>Nycteris grandis</i> Peters, 1865
Große Schlitznasen-Fledermaus (German)	<i>Nycteris grandis</i> Peters, 1865
Große Schmetterlingsfledermaus (German)	<i>Glauconycteris superba</i> Hayman, 1939
Grosser Maskarenen-Flugfuchs (German)	<i>Pteropus niger</i> (Kerr, 1792)
Großhufeisennase (German)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Großohr-Bulldoggfledermaus (German)	<i>Tadarida lobata</i> (Thomas, 1891)
Großohrhohltnase (German)	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
Großohrige Riesen-Bulldoggfledermaus (German)	<i>Otomops martiensseni</i> (Matschie, 1897)
Großohrige Schlitznasen-Fledermaus (German)	<i>Nycteris macrotis</i> Dobson, 1876
Großohr-Rundblattnase (German)	<i>Hipposideros megalotis</i> (Heuglin, 1861)
Grote fruitvleermuizen (Dutch)	PTEROPODIDAE Gray, 1821
Grote hoefijzerneus (Dutch)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Grote klapneusvleermuis (Dutch)	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
Grote rosse vleermuis (Dutch)	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Grote Schreiber's vleermuis (Dutch)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Groter spleetneusvlermuis (Afrikaans)	<i>Nycteris macrotis</i> Dobson, 1876
Grouss Hufeisennues (Luxembourgish)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Grünliche Hausfledermaus (German)	<i>Scotophilus viridis</i> (Peters, 1852)
Grutftuottige flearmûs (Frisian)	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Grutte hoefizernoas (Frisian)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Grutte reade flearmûs (Frisian)	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Guano Bats (English)	<i>Tadarida Rafinesque</i> , 1814
Guinea Serotine (English)	<i>Neoromicia guineensis</i> (Bocage, 1889)
Guinea-Hufeisennase (German)	<i>Rhinolophus guineensis</i> Eisentraut, 1960
Guinean Horseshoe Bat (Csorba <i>et al.</i> , 2003: 59 - 61; Simmons, 2005) (English)	<i>Rhinolophus guineensis</i> Eisentraut, 1960
Guinean Pipistrelle Bat (English)	<i>Neoromicia guineensis</i> (Bocage, 1889)
Guinea-Pipistrelle (German)	<i>Neoromicia guineensis</i> (Bocage, 1889)
Guinea-Zwergfledermaus (German)	<i>Neoromicia guineensis</i> (Bocage, 1889)
Haagner's Flat-headed Bat (English)	<i>Sauromys petrophilus haagneri</i> (Roberts, 1917)
Haagnerse Platkopvlermuis (Afrikaans)	<i>Sauromys petrophilus haagneri</i> (Roberts, 1917)
Haarbindigen Dämmerungsfledermaus (German)	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Haarige Schlitznasenfledermaus (German)	<i>Nycteris hispida</i> (Schreber, 1774)
Habelendlane (Estonian)	<i>Myotis mystacinus</i> (Kuhl, 1817)
hadozubí kaloni (Czech)	<i>Scotonycteris Matschie</i> , 1894
Hairy Bats (English)	<i>Myotis Kaup</i> , 1829
Hairy Long-eared Bat (English)	<i>Nycteris hispida</i> (Schreber, 1774)
Hairy Slit-faced Bat (English)	<i>Nycteris hispida</i> (Schreber, 1774)
Hairy-armed bat (English)	<i>Nyctalus leisleri</i> (Kuhl, 1817)
Halcyon Horseshoe Bat (English)	<i>Rhinolophus alcyone</i> Temminck, 1853
Halcyon Rundblattnase (German)	<i>Rhinolophus alcyone</i> Temminck, 1853
Halskrausen-Flughunde (German)	<i>Myonycteris Matschie</i> , 1899
Hamiltons Grabfledermaus (German)	<i>Taphozous hamiltoni</i> Thomas, 1920
Hamilton's Naked-bellied Tomb Bat (English)	<i>Taphozous hamiltoni</i> Thomas, 1920
Hamilton's Naked-rumped Tomb Bat (English)	<i>Taphozous hamiltoni</i> Thomas, 1920
Hamilton's Tomb Bat (English)	<i>Taphozous hamiltoni</i> Thomas, 1920
Hammer-headed bat (English)	<i>Hypsignathus monstrosus</i> H. Allen, 1862
Hammer-headed bats (English)	<i>Hypsignathus H. Allen</i> , 1862
Hammer-headed Fruit Bat (English)	<i>Hypsignathus monstrosus</i> H. Allen, 1862
Hammerkopf (German)	<i>Hypsignathus monstrosus</i> H. Allen, 1862
Hammerkopf-Flughund (German)	<i>Hypsignathus monstrosus</i> H. Allen, 1862
Hammerkopf-Flughunde (German)	<i>Hypsignathus H. Allen</i> , 1862
Hanak pipistrello (Basque)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hanak'in Cüce Yarasası (Turkish)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hanaka sikspārnis (Latvian)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hanaki cırdan şəbpərəsi (Azerbaijani)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hanako šikšniukas (Lithuanian)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hanakov šišmiš (Bosnian)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hanakov patuljasti šišmiš (Croatian)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hanaks dwerchflearmûs (Frisian)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hanaks dwergvleermuis (Dutch)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hanak's dwergvleermuis (Dutch)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hanaks flagermus (Danish)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hanaks Pipistrelle (German)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hanak's Pipistrelle (English)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004

Common Name

Hanaks Zwergfledermaus (German)
 Hanák-törpedenevér (Hungarian)
 Happolds' Pipistrelle (English)
 Hardwickes Mausschwanz-Fledermaus (German)
 Hardwicke's Mouse-tailed Bat (English)
 Harige Langoorvlermuis (Afrikaans)
 Harige spleetneusvlermuis (Afrikaans)
 Harlekin-Flughunde (German)
 Harlequin Fruit-bats (English)
 Harrison's large-eared giant mastiff bat (English)
 Harrison-Riesen-Bulldoggfledermaus (German)
 Harrison's fruit bat (English)
 Harrisons Samtfell-Flughund (German)
 Hartneusvleermuis (Dutch)
 Hausfledermäuse (German)
 Hautalepakot (Finnish)
 Hayman's tear-drop fruit bat (English)
 Hayman's Dwarf Epauletten Fruit Bat (English)
 Hayman's Epauletten Fruit Bat (English)
 Haymans Harlekin-Flughund (German)
 Haymans Kleiner Epaulettenflughund (German)
 Hayman's Lesser Fruit Bat (English)
 Hayman's Lesser Fruit-bat (English)
 Heart-nosed Bat (English)
 Heart-nosed Bats (English)
 Heart-nosed Big-eared Bat (English)
 Hedelmälepakot (Finnish)
 Hele-sagarnina (Estonian)
 Helios Pipistrelle Bat (English)
 Heller's Pipistrelle (English)
 Heller's Pipistrelle Bat (English)
 Hemprich belarrihandi (Basque)
 Hemprich's arrow-eared bat (English)
 Hemprichov dugouhi šišmiš (Croatian)
 Hemprich's Arrow-headed Bat (English)
 Hemprich's Big-eared Bat (English)
 Hemprich's Desert Bat (English)
 Hemprich's Desert Long-eared Bat (English)
 Hemprich's Long-eared Bat (English)
 Hemprich's Long-eared Desert Bat (English)
 Hemprichs Wüstengroßohr (German)
 Hempriha sikspārnis (Latvian)
 Hemprihov dugouhi šišmiš (Bosnian)
 Herznasen-Fledermaus (German)
 Herznasen-Fledermäuse (German)
 herzhörige Abendfledermaus (German)
 Hesteskoneser (Norwegian)
 Heuglins Wollfledermaus (German)
 Hevoskenkäyököt (Finnish)
 High-crowned bat (English)
 Hiidvidevlane (Estonian)
 Hildebrandt se saalneusvlermuis (Afrikaans)
 Hildebrandt's Horseshoe Bat (English)
 Hildebrandts Hufeisennase (German)
 Hildebrandt-saalneusvlermuis (Afrikaans)
 Hildebrandtse Vlermuis (Afrikaans)
 Hildegardes Grabfledermaus (German)
 Hildegarde's Tomb Bat (English)
 Hill's Horseshoe Bat (English)
 Hill's Horseshoe bat (English)
 Hills Hufeisennase (German)
 Hilobi-saguzar gibelsoil (Basque)
 Hindes Hausfledermaus (German)
 Hinde's Lesser House Bat (English)
 Hipposidéridés (French)
 Hispid bats (English)
 Hladkonos holobruchý (Slovak)
 Hochland-Hufeisennase (German)
 Hoefijzerneusvleermuizen (Dutch)
 Hoefijzerneuzen (Dutch)
 Hoefijzerneuzen (Dutch)

Taxon Name

[illegible]

Common Name

Höhlenflughunde (German)
 Hollow-faced bats (English)
 Hollow-faced bats (English)
 Horacekcs hoefijzerneus (Dutch)
 Horn-skinned Bat (English)
 Horn-skinned Pipistrelle Bat (English)
 Horn-skinned Serotine (English)
 Horseshoe bats (English)
 Horseshoe bats (English)
 Hosszúlábú denevér (Hungarian)
 Hosszúszárnyú denevér (Hungarian)
 Hottentot bat (English)
 Hottentot Serotine Bat (English)
 Hottentotten-Breitflügelfledermaus (German)
 House bats (English)
 House Bats (English)
 hrobkovec egyptský (Czech)
 hrobkovec guinejský (Czech)
 Hrobkovec lysobřichý (Czech)
 hrobkovec mauricijský (Czech)
 hrobkovec východoafrický (Czech)
 hrobkovec východoafrický (Czech)
 Hroznohled pyskatý (Czech)
 hroznohledové (Czech)
 Hufeisennase (German)
 Hufeisennasen (German)
 Hufeisennasen (German)
 Hufeisennasen-Fledermäuse (German)
 Hufeisennasen-Fledermäuse (German)
 Hullneser (Norwegian)
 Hundkopf-Schwirrfledermaus (German)
 Hvitrandflaggermus (Norwegian)
 Hypsignates (French)
 Hypsignathe monstrueux (French)
 Iranta Ramptu Kallarai Velāl (Tamil)
 Ialtag Asorach (Scottish Gaelic)
 Ialtag bharbastail iarach (Scottish Gaelic)
 Ialtag Botta (Scottish Gaelic)
 Ialtag eàrr-shaor Eòrpach (Scottish Gaelic)
 Ialtag fhad-chluasach Chanaireach (Scottish Gaelic)
 Ialtag fhad-chluasach Hemprich (Scottish Gaelic)
 Ialtag fhad-mheurach (Scottish Gaelic)
 Ialtag fheasgarach (Scottish Gaelic)
 Ialtag Geoffroy (Scottish Gaelic)
 Ialtag ghibeach (Scottish Gaelic)
 Ialtag Hanak (Scottish Gaelic)
 Ialtag isibealach (Scottish Gaelic)
 Ialtag Kuhl (Scottish Gaelic)
 Ialtag Leisler (Scottish Gaelic)
 Ialtag luch-chluasach Mhaigribeach (Scottish Gaelic)
 Ialtag meas Éipheiteach (Scottish Gaelic)
 Ialtag Mhaideurach (Scottish Gaelic)
 Ialtag mheur-fhada Schreiber (Scottish Gaelic)
 Ialtag nochd-thònach thuaim (Scottish Gaelic)
 Ialtag Savi (Scottish Gaelic)
 Ialtag shoprànach (Scottish Gaelic)
 Ialtóg Asórach (Irish Gaelic)
 Ialtóg bharbastail Iartharach (Irish Gaelic)
 Ialtóg Botta (Irish Gaelic)
 Ialtóg earrshaor Eorpach (Irish Gaelic)
 Ialtóg fhadchluasach Chanárach (Irish Gaelic)
 Ialtóg fhadchluasach Hemprich (Irish Gaelic)
 Ialtóg fhadmhéarach (Irish Gaelic)
 Ialtóg fheascrach (Irish Gaelic)
 Ialtóg Geoffroy (Irish Gaelic)
 Ialtóg ghiobach (Irish Gaelic)
 Ialtóg Hanak (Irish Gaelic)
 Ialtóg isibéalach (Irish Gaelic)
 Ialtóg Kuhl (Irish Gaelic)
 Ialtóg Leisler (Irish Gaelic)
 Ialtóg luch-chluasach Mhaigribeach (Irish Gaelic)
 Ialtóg meas Éigipteach (Irish Gaelic)
 Ialtóg Mhaidéarach (Irish Gaelic)
 Ialtóg mhéarfhada Schreiber (Irish Gaelic)

Taxon Name

Rousettus Gray, 1821
 NYCTERIDAE Van der Hoeven, 1855
Nycteris G. Cuvier and E. Geoffroy, 1795
Rhinolophus horaceki Benda and Vallo, 2012
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
 RHINOLOPHIDAE Gray, 1825
Rhinolophus Lacépède, 1799
Myotis capaccinii (Bonaparte, 1837)
Miniopterus schreibersii (Kuhl, 1817)
Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
Scotophilus Leach, 1821
Scotoecus Thomas, 1901
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Saccolaimus peli (Temminck, 1853)
Taphozous nudiventris Cretzschmar, 1830
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Taphozous hamiltoni Thomas, 1920
Taphozous hildegardeae Thomas, 1909
Tadarida teniotis (Rafinesque, 1814)
Tadarida Rafinesque, 1814
Rhinolophus ferrumequinum (Schreber, 1774)
 RHINOLOPHIDAE Gray, 1825
Rhinolophus Lacépède, 1799
 RHINOLOPHIDAE Gray, 1825
Rhinolophus Lacépède, 1799
 NYCTERIDAE Van der Hoeven, 1855
Scotophilus nigrita (Schreber, 1774)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Hypsignathus H. Allen, 1862
Hypsignathus monstrosus H. Allen, 1862
Taphozous nudiventris Cretzschmar, 1830
Nyctalus azoreum (Thomas, 1901)
Barbastella barbastellus (Schreber, 1774)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Tadarida teniotis (Rafinesque, 1814)
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859
Myotis capaccinii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis mystacinus (Kuhl, 1817)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Myotis punicus Felten, 1977
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Miniopterus schreibersii (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Nyctalus azoreum (Thomas, 1901)
Barbastella barbastellus (Schreber, 1774)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Tadarida teniotis (Rafinesque, 1814)
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859
Myotis capaccinii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis mystacinus (Kuhl, 1817)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Myotis punicus Felten, 1977
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Miniopterus schreibersii (Kuhl, 1817)

Common Name

Ialtóg nochtphrumpach thuama (Irish Gaelic)
 Ialtóg Savi (Irish Gaelic)
 Ialtóg shopránach (Irish Gaelic)
 Ibeeria nahkhiir (Estonian)
 idludaka (isiXhosa)
 Ilgapiřtis pelėausis (Lithuanian)
 İlpəqarın məzar yarasası (Azerbaijani)
 ilulwane (isiXhosa)
 İmtchaghyeye (Kabyle (Algeria))
 Indur-lenji Chamchika (Bengali)
 Ingekorven vleermuis (Dutch)
 Intermediäre Schlitznasenfledermaus (German)
 Intermediäre Schlitznasen-Fledermaus (German)
 Intermediate Slit-faced Bat (English)
 Ipposidèri (Italian)
 Ipposidèridi (Italian)
 Ipposidero càfro (Italian)
 Ipposidero oreccchiuto (Italian)
 Ipposidero vittato (Italian)
 Ipsùghi (Italian)
 Isabel-Fledermaus (Luxembourgish)
 Isabelkleurde lette flearmûs (Frisian)
 Isabellaflaggermus (Norwegian)
 Isabellasydfloggermus (Danish)
 Isabell-Breitflügel-Fledermaus (German)
 isabellfarbenen Abendfledermaus (German)
 Isabellfladdermus (Swedish)
 Isabellfledermaus (German)
 Isabelline Serotine Bat (English)
 Isabelline White-winged Serotine (English)

 Isabell-Weissflügel Serotine (German)

 Isabelvleermuis (Dutch)
 Isalo Serotine (English)

 İzabel Genişkanatlı Yarasası (Turkish)
 Izabelin šišmiš (Bosnian)
 Izabelin noćnjak (Croatian)
 Izabella-késeidenevér (Hungarian)
 Izabellas sikspārnis (Latvian)
 Izabelos šikšnys (Lithuanian)
 Ja Slit-faced Bat (English)
 ještěf morousi (Czech)
 jihoafričtí cíponosi (Czech)
 Jones' Leaf-nosed Bat (English)
 Jones' Roundleaf Bat (English)
 Jones' Rundblattnase (German)
 Jones's Roundleaf Bat (English)
 Južni potkovnjak (Slovenian)
 Južni potkovičar (Montenegrin)
 Južni potkovnjak (Croatian)
 Juzhen Potkovichar (Macedonian)
 Kõrbe-nahkhiir (Estonian)
 Kūla sikspārnis (Latvian)
 Kääbus-nahkhiir (Estonian)
 Kaalbuikgrafvleermuis (Dutch)
 Kääpiölepakko (Finnish)
 Kaapse Blaarnesvlermuis (Afrikaans)
 Kaapse dakvlermuis (Afrikaans)
 Kaapse Langhaarvlermuis (Afrikaans)
 Kaapse Langoorvlermuis (Afrikaans)
 Kaapse saalnesvlermuis (Afrikaans)
 Kaapse Vlermuis (Afrikaans)
 Kafeav Dzudžest Liljak (Macedonian)
 Kaffern-Sackfledermaus (German)
 Kahazo-komule (Kinande (DRC))
 Kahlschienige Dämmerungsfledermaus (German)
 Kahuzi horseshoe bat (English)
 Kahuzi-Hufeisennase (German)
 KailvĒdera sikspārnis (Latvian)
 Kakoro kombe (Kinande (DRC))
 Kakorokombe (Kinande (DRC))
 Kakorokombe (Kiande (DRC))

Taxon Name

Taphozous nudiventris Cretzschmar, 1830
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
CHIROPTERA Blumenbach, 1779
Myotis capaccinii (Bonaparte, 1837)
Taphozous nudiventris Cretzschmar, 1830
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
Rhinopoma microphyllum (Brünnich, 1782)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Nycteris intermedia Aellen, 1959
Nycteris intermedia Aellen, 1959
Nycteris intermedia Aellen, 1959
Hipposideros Gray, 1831
HIPPOSIDERIDAE Lydekker, 1891
Hipposideros caffer (Sundevall, 1846)
Hipposideros megalotis (Heuglin, 1861)
Macronycteris vittatus (Peters, 1852)
Hypsugo Kolenati, 1856
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Pseudoromicia isabella (Decher, Hutterer and Monadjem, 2016)
Pseudoromicia isabella (Decher, Hutterer and Monadjem, 2016)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Laephotis malagasyensis (Peterson, Eger and Mitchell, 1995)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Nycteris major (K. Andersen, 1912)
Sauromys Roberts, 1917
Cloeotis Thomas, 1901
Hipposideros jonesi Hayman, 1947
Hipposideros jonesi Hayman, 1947
Hipposideros jonesi Hayman, 1947
Hipposideros jonesi Hayman, 1947
Rhinolophus euryale Blasius, 1853
Rhinolophus euryale Blasius, 1853
Rhinolophus euryale Blasius, 1853
Rhinolophus euryale Blasius, 1853
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Taphozous nudiventris Cretzschmar, 1830
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Hipposideros caffer (Sundevall, 1846)
Laephotis capensis (A. Smith, 1829)
Myotis tricolor (Temminck, 1832)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Rhinolophus capensis Lichtenstein, 1823
Rhinolophus capensis Lichtenstein, 1823
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Miniopterus natalensis (A. Smith, 1833)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rhinolophus kahuzi Fahr and Kerbis Peterhans, 2013
Rhinolophus kahuzi Fahr and Kerbis Peterhans, 2013
Taphozous nudiventris Cretzschmar, 1830
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Nycteris G. Cuvier and E. Geoffroy, 1795

Taxon Name

[illegible]

Common Name

Kanarski dugouhi šišmiš (Bosnian)
 Kanarski sivi dugoušan (Serbian Latin)
 Kanarski slijepi miš (Montenegrin)
 Kanarski uhatež (Slovenian)
 Kanaryarlar Uzunkulaklı Yarasası (Turkish)
 Kanaryske grutearflearmûs (Frisian)
 Kanfan (Hebrew)
 Kapearäpyläpiippa (Finnish)
 Kap-Hufeisennase (German)
 Kap-Pipistrelle (German)
 Kap-Zwergfledermaus (German)
 Karakonube (Kinande (DRC))
 Karlik średni (Polish)
 Karlik cyrenejski (Polish)
 Karlik drobny (Polish)
 Karlik Kuhla (Polish)
 Karlik maderski (Polish)
 Karlik malutki (Polish)
 Karuru (Chewa (Malawi))
 Kashusha (Misuku (Malawi))
 Kastanienrothe Kammnase (German)
 Kasusu (Kiluba (DRC))
 Kasusu (Kiluba (DRC))
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 Kasusu (Kiluba (DRC))
 Katutu (Kiluba (DRC))
 Katutu (Kiluba (DRC))
 Katutu (Kiluba (DRC))
 Kaviokuonoyököt (Finnish)
 Kealbûktimpelflearmûs (Frisian)
 Kelelawar ekor lembing (Indonesian)
 Kelelawar Ekor-tikus (Indonesian)
 Kelelawar mini biasa (Indonesian)
 Kelelawar mini schreiber (Indonesian)
 Kelelawar myotis pucuk (Indonesian)
 Keltapomimulepakko (Finnish)
 Kenya Barbastelle (English)
 Kenya Pipistrelle (English)
 Kenyan Big-eared Free-tailed Bat (English)
 Kenyan Butterfly Bat (English)
 Kenyan Wattle Bat (English)
 Kenya-Schmetterlingsfledermaus (German)
 Kerbohrige Ohrenfledermaus (German)
 Kereknyergű patkósdenevér (Hungarian)
 Kereknyergű patkósrú denevér (Hungarian)
 Kérivoule de Tanzanie (French)
 kerivula africká (Czech)
 kerivuly (Czech)
 Khafash abiad (Arabian)
 Khafash Abu Bouz (Arabian)
 Khafash abu bouz saghir (Arabian)
 Khafash abu danab kabir (Arabian)
 Khafash abu danab saghir (Arabian)
 Khafash abu hadwa kabir (Arabian)
 Khafash abu hadwa saghir (Arabian)
 Khafash abu hadweh arabi (Arabian)
 Khafash El Fawakeh (Arabian)
 Khafash Masri (Arabian)
 Khafash Teiba (Arabian)
 Khaffash (Arabian)
 Khaffash (Arabian)
 Khaffash (Arabian)
 Khaffash (Arabian)
 Khaffash (Arabian)
 Khaffash (Arabian)

Taxon Name

Plecotus teneriffae Barrett-Hamilton, 1907
Plecotus teneriffae Barrett-Hamilton, 1907
Plecotus teneriffae Barrett-Hamilton, 1907
Plecotus teneriffae Barrett-Hamilton, 1907
Plecotus teneriffae Barrett-Hamilton, 1907
Plecotus teneriffae Barrett-Hamilton, 1907
Miniopterus schreibersii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Rhinolophus capensis Lichtenstein, 1823
Laephotis capensis (A. Smith, 1829)
Laephotis capensis (A. Smith, 1829)
Stenonycteris lanosus (Thomas, 1906)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Afronycteris nana (Peters, 1852)
Rhinolophus landeri Martin, 1838
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Rhinolophus Lacépède, 1799
Hipposideros caffer (Sundevall, 1846)
Rhinolophus hildebrandtii Peters, 1878
Rhinolophus landeri Martin, 1838
Cloeotis percivali Thomas, 1901
Nycteris G. Cuvier and E. Geoffroy, 1795
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Kerivoula argentata Tomes, 1861
Afronycteris nana (Peters, 1852)
Laephotis capensis (A. Smith, 1829)
Nycticeinops schlieffenii (Peters, 1859)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Nycteris arge Thomas, 1903
Nycteris hispida (Schreber, 1774)
 RHINOLOPHIDAE Gray, 1825
Taphozous nudiventris Cretzschmar, 1830
Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma microphyllum (Brünnich, 1782)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Glauconycteris kenyacola Peterson, 1982
Pipistrellus (Pipistrellus) aero Heller, 1912
Tadarida lobata (Thomas, 1891)
Glauconycteris kenyacola Peterson, 1982
Glauconycteris kenyacola Peterson, 1982
Glauconycteris kenyacola Peterson, 1982
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Rhinolophus euryale Blasius, 1853
Rhinolophus euryale Blasius, 1853
Kerivoula africana Dobson, 1878
Kerivoula africana Dobson, 1878
Kerivoulinae Miller, 1907
Otonycteris hemprichii Peters, 1859
Taphozous nudiventris Cretzschmar, 1830
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma cystops Thomas, 1903
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus clivosus Cretzschmar, 1828
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rhinolophus blasii Peters, 1867
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus euryale Blasius, 1853
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Rhinolophus ferrumequinum (Schreber, 1774)

Common Name

Khaffash (Arabian)
 Khaffash (Arabian)
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 Khaffash (Arabian)
 Khaffash (Arabian)
 Khouchaf al matoussat (Arabian)
 Kibil (Somalian)
 Kibille (Somalian)
 Kibilile (Somalian)
 Kiçik şəbpərə (Azerbaijani)
 Kiçik axşam yarasası (Azerbaijani)
 Kiçik nalburun (Azerbaijani)
 Kirpikli Yarasa (Turkish)
 Kis patkósdenevér (Hungarian)
 Kis patkósorrú denevér (Hungarian)
 kitrotroke (Malagasy)

 Kityo's serotine (English)

 kladvohlaví kaloni (Czech)
 klaffneser (Norwegian)
 klaponos egyiptský (Czech)
 klaponosové (Czech)
 Klein dakvlermuis (Afrikaans)
 Klein geel dakvlermuis (Afrikaans)
 Klein grotvlermuis (Afrikaans)
 Klein losstertvlermuis (Afrikaans)
 Klein Vrugtevlermuis (Afrikaans)
 Klein wolhaarvlermuis (Afrikaans)
 Kleinabendsegler (German)
 Kleine Abendsegler (German)
 Kleine Bartfledermaus (German)
 Kleine Bulldoggfledermaus (German)
 Kleine dwergvlermuis (Dutch)
 Kleine Hausfledermäuse (German)
 Kleine hoefijzerneus (Dutch)
 Kleine Hufeisennase (German)
 Kleine klapneusvlermuis (Dutch)
 Kleine Langflügel-fledermaus (German)
 Kleine Mausschwanzfledermaus (German)
 Kleine Pipistrelle (German)
 Kleine Schmetterlingsfledermaus (German)
 Kleine Waldfledermaus (German)
 Kleine Wollfledermaus (German)
 Kleine Zwergfledermaus (German)
 Kleiner Abendsegler (German)
 Kleiner Halsbandflughund (German)
 Kleiner Halskrausen-Flughund (German)
 Kleiner Maskarenen-Flugfuchs (German)
 Kleinhufeisennase (German)
 Kleinohr-Dreizackblattnase (German)
 Kleinste Schlitznasen-Fledermaus (German)
 Kleng Baartfledermaus (Luxembourgish)
 Kleng Bëschrfliedermaus (Luxembourgish)
 Kleng Fledermaus (Luxembourgish)
 Kleng Hufeisennues (Luxembourgish)
 Klippeflaggermus (Norwegian)
 Knight's Egyptian bat (English)

Taxon Name

Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus mehelyi Matschie, 1901
Rhinopoma cystops Thomas, 1903
Rhinopoma microphyllum (Brünnich, 1782)
Taphozous nudiventris Cretzschmar, 1830
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Tadarida teniotis (Rafinesque, 1814)
Miniopterus schreibersii (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Vansonia rueppellii (J.B. Fischer, 1829)
Barbastella leucomelas (Cretzschmar, 1826)
Otonycteris hemprichii Peters, 1859
Plecotus christii Gray, 1838
Hypsugo ariel (Thomas, 1904)
Hypsugo savii (Bonaparte, 1837)
Afronycteris nana (Peters, 1852)
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus hipposideros (Bechstein, 1800)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus hipposideros (Bechstein, 1800)
Miniopterus mahafaliensis Goodman, Bradman, Christides and Appleton, 2009
Pseudoromicia kityoi Monadjem, Kerbis Peterhans, Nalikka, Waswa, Demos and Patterson, 2020
Hypsignathus H. Allen, 1862
RHINOPOMATIDAE Dobson, 1872
Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma E. Geoffroy St.-Hilaire, 1818
Nycticeinops schlieffenii (Peters, 1859)
Scotophilus viridis (Peters, 1852)
Miniopterus fraterculus Thomas and Schwann, 1906
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Epomophorus crypturus Peters, 1852
Kerivoula lanosa (A. Smith, 1847)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Scotoecus Thomas, 1901
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinopoma cystops Thomas, 1903
Miniopterus fraterculus Thomas and Schwann, 1906
Rhinopoma cystops Thomas, 1903
Pipistrellus (Pipistrellus) nanulus Thomas, 1904
Glauconycteris beatrix Thomas, 1901
Nyctalus leisleri (Kuhl, 1817)
Kerivoula lanosa (A. Smith, 1847)
Pipistrellus (Pipistrellus) nanulus Thomas, 1904
Nyctalus leisleri (Kuhl, 1817)
Myonycteris torquata (Dobson, 1878)
Myonycteris torquata (Dobson, 1878)
Pteropus subniger (Kerr, 1792)
Rhinolophus hipposideros (Bechstein, 1800)
Cloeotis Thomas, 1901
Nycteris nana (K. Andersen, 1912)
Myotis mystacinus (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Hypsugo savii (Bonaparte, 1837)
Aegyptonycteris knightae Simmons, Seiffert and Gunnell, 2016

Common Name

Knight's Egyptische vleermuis (Dutch)
 Knob-tailed Mops Bat (English)
 Knob-tailed Nyctinome (English)
 kožnatcovití (Czech)
 Kocks Mausohr (German)
 Kolombatovicev dugoušan (Croatian)
 Kolombatovic's Big-eared Bat (English)
 Kolombatovic's Long-eared Bat (English)
 Komoren-Höhlenflughund (German)
 Komoren-Langflügelfledermaus (German)
 Kongo-Bulldoggfledermaus (German)
 Kordofan Sheath-tail-bat (English)
 Közönséges törpedenevér (Hungarian)
 Kraljičin poznik (Slovenian)
 Kraljičin slijepi miš (Montenegrin)
 krátkonebí kaloni (Czech)
 Kretapipistrell (Swedish)
 Kruger serotine bat (Taylor, 2005: 305 - first use) (English)
 Kryptische Madagaskar-Rundblattnase (German)
 krysoví morousi (Czech)
 Kubukubo ("Congolese")
 Kubukubo ("Congolese")
 Küçük Ağaç Yarasası (Turkish)
 Küçük Nalburunlu Yarasa (Turkish)
 Kuhl pipistrello (Basque)
 Kuhl saguzar (Basque)
 Kuhl se vlermuis (Afrikaans)
 Kuhlische Fledermaus (German)
 Kuhl's Bat (English)
 Kuhls dwergvleermuis (Dutch)
 Kuhls dwergvleermuis (Dutch)
 Kuhls fladdermus (Swedish)
 Kuhls flagermus (Danish)
 Kuhl's pipistrelle (English)
 Kuhl's Pipistrelle Bat (English)
 Kuhl-vleermuis (Afrikaans)
 Küli şəbpərəsi (Azerbaijani)
 Kulijev šišmiš (Bosnian)
 Kulio šikšniukas (Lithuanian)
 Kupferfarbene Wollfledermaus (German)
 Kupferfarbenes Mausohr (German)
 Kurzflügel-Bulldoggfledermaus (German)
 Kurzflügeligen Doggengrämler (German)
 Kurzgaumen-Flughunde (German)
 Kurzmaulige Fledermaus (German)
 Kurzschwanz-Rundblattnase (German)
 Kusthåstskonäsa (Swedish)
 Kyrenaikaflaggermus (Norwegian)
 la petite roussette des Comores (French)
 La Phyllorhine cryptique de Madagascar (French)
 la roussette de Livingstone (French)
 Laangflillekefledermaus (Luxembourgish)
 Laangfoussfledermaus (Luxembourgish)
 Lagos Serotine (English)
 Lagos Serotine Bat (English)
 Lagos-Breitflügelfledermaus (German)
 Lakuriq nate bisht-lire (Albanian)
 Lakuriq nate bishtlirë (Albanian)
 Lakuriq nate egjiptian i frutave (Albanian)
 Lakuriq nate gishtgjatë (Albanian)
 Lakuriq nate hundëpatkua i Blasius-it (Albanian)
 Lakuriq nate hundëpatkua i madh (Albanian)
 Lakuriq nate hundëpatkua i Mehely-it (Albanian)
 Lakuriq nate hundëpatkua i Mesdheut (Albanian)
 Lakuriq nate hundëpatkua i vogël (Albanian)
 Lakuriq nate i Geoffroy-it (Albanian)
 Lakuriq nate i Schreiber-it (Albanian)
 Lakuriq nate maghrebien (Albanian)
 Lakuriq nate me mustaqe (Albanian)
 Lakuriq nate veshgjatë i Hemprich-it (Albanian)

Taxon Name

Aegyptonycteris knightae Simmons, Seiffert and Gunnell, 2016
Mops (Mops) condylurus (A. Smith, 1833)
Mops (Mops) condylurus (A. Smith, 1833)
 MEGADERMATIDAE H. Allen, 1864
Myotis dieteri M. Happold, 2005
Plecotus kolombatovici Dulic, 1980
Plecotus kolombatovici Dulic, 1980
Plecotus kolombatovici Dulic, 1980
Rousettus obliviosus Kock, 1978
Miniopterus griveaudi Harrison, 1959
Mops (Mops) congicus J.A. Allen, 1917
Taphozous hamiltoni Thomas, 1920
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Casinycteris Thomas, 1910
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Laephotis stanleyi (Goodman, Kearney, Ratsimbazafy and Hassanin, 2017)
Macronycteris cryptovalorona (Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016)
Myopterus E. Geoffroy St.-Hilaire, 1818
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Taphozous nudiventris Cretzschmar, 1830
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Kerivoula cuprosa Thomas, 1912
Myotis bocagii (Peters, 1870)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Casinycteris Thomas, 1910
Barbastella barbastellus (Schreber, 1774)
Hipposideros curtus G.M. Allen, 1921
Rhinolophus mehelyi Matschie, 1901
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Rousettus obliviosus Kock, 1978
Macronycteris cryptovalorona (Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016)
Pteropus livingstonii Gray, 1866
Miniopterus schreibersii (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Eptesicus (Cnephæus) platyops (Thomas, 1901)
Eptesicus (Cnephæus) platyops (Thomas, 1901)
Eptesicus (Cnephæus) platyops (Thomas, 1901)
Tadarida teniotis (Rafinesque, 1814)
Tadarida teniotis (Rafinesque, 1814)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Myotis capaccinii (Bonaparte, 1837)
Rhinolophus blasii Peters, 1867
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Rhinolophus hipposideros (Bechstein, 1800)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Miniopterus schreibersii (Kuhl, 1817)
Myotis punicus Felten, 1977
Myotis mystacinus (Kuhl, 1817)
Otonycteris hemprichii Peters, 1859

Common Name

Lakuriq nate veshgjatë i Teneriffae-ve (Albanian)
 Lamotte's Leaf-nosed Bat (English)
 Lamotte's Roundleaf Bat (English)
 Lamottes Rundblattnase (German)
 Lander se saalneusvlermuis (Afrikaans)
 Lander's Horseshoe Bat (English)
 Landers Hufeisennase (German)
 Lander's Rundblattnase (German)
 Lander-saaneusvlermuis (Afrikaans)
 Lanflügelfledermaus (German)
 Langflügelfledermaus (German)
 Langflügelfledermäuse (German)
 Langflügelfledermäuse (German)
 Langfotflaggermus (Norwegian)
 Langfußfledermaus (German)
 Langhalete flaggermus (Norwegian)
 Langohren (German)
 Langohrfledermäuse (German)
 Langoor-vlermuise (Afrikaans)
 Langstaartvleermuizen (Dutch)
 Langstert-dakvlermuis (Afrikaans)
 Langvleugelvleermuis (Dutch)
 Langvleugelvleermuizen (Dutch)
 Lappet-eared Free-tailed Bat (English)
 Lappet-eared Wrinkle-lipped Bat (English)
 Large Bentwing Bat (English)
 large bent-winged bat (English)
 Large Free-tailed Bat (English)
 Large Guano Bat (English)
 Large Headed Pipistrelle (English)
 Large Slit-faced Bat (English)
 Large Wrinkle-lipped Bat (English)
 Large-eared Free-tailed Bat (English)
 Large-eared Giant Mastiff Bat (English)
 Large-eared Leaf-nosed Bat (English)
 Large-eared Slit-faced Bat (English)
 Large-footed Bats (English)
 Larger Horseshoe Bat (English)
 Larger Mouse-tailed Bat (English)
 Larger Rat-tailed Bat (English)
 Large-winged Bats (English)
 Lavie à ailes jaunes (French)
 le grand rhinolophe fer à cheval (French)
 Le Megaderme feuille (French)
 le vespertilion de Kuhl (French)
 le vespertilion de Leisler (French)
 le vespertilion pygmée (French)
 Leafnosed Bats (English)
 Leaf-nosed bats (English)
 Leaf-nosed Bats (English)
 Leaf-winged bat (English)
 Least Epauletted Fruit-bat (English)
 Least horseshoe bat (English)
 Least Long-fingered Bat (English)
 Least Pipistrelle Bat (English)
 Lefôte di Winton (Italian)
 Lefôti (Italian)
 leilan (Hebrew)
 Leisler gau-saguzar (Basque)
 Leisler's Bat (English)
 Leislers fladdermus (Swedish)
 Leislers flagermus (Danish)
 Leisler's flagermus (Danish)
 Leisler's noctule (English)
 Leislersche Fledermaus (German)
 Leseuers Mausohr (German)
 lesní netopýři (lesní létači) (Czech)
 Lesser (Sundevall's) Leaf-nosed Bat (English)
 Lesser Angolan Epauletted Fruit Bat (English)
 Lesser bent-winged bat (English)
 Lesser Epauletted Bat (English)

Taxon Name

Plecotus teneriffae Barrett-Hamilton, 1907
Hipposideros lamottei Brosset, 1985
Hipposideros lamottei Brosset, 1985
Hipposideros lamottei Brosset, 1985
Rhinolophus landeri Martin, 1838
Rhinolophus landeri Martin, 1838
Rhinolophus landeri Martin, 1838
Rhinolophus landeri Martin, 1838
Rhinolophus landeri Martin, 1838
Miniopterus villiersi Aellen, 1956
Miniopterus schreibersii (Kuhl, 1817)
 MINIOPTERIDAE Dobson, 1875
Miniopterus Bonaparte, 1837
Myotis capaccinii (Bonaparte, 1837)
Myotis capaccinii (Bonaparte, 1837)
 RHINOPOMATIDAE Dobson, 1872
Plecotus E. Geoffroy St.-Hilaire, 1818
Plecotus E. Geoffroy St.-Hilaire, 1818
 NYCTERIDAE Van der Hoeven, 1855
 RHINOPOMATIDAE Dobson, 1872
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Miniopterus schreibersii (Kuhl, 1817)
 MINIOPTERIDAE Dobson, 1875
Mops (Chaerephon) major (Trouessart, 1897)
Mops (Chaerephon) major (Trouessart, 1897)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Tadarida fulminans (Thomas, 1903)
Tadarida fulminans (Thomas, 1903)
Nycticeinops macrocephalus (Hutterer and Kerbis Peterhans, 2020)
Nycteris grandis Peters, 1865
Mops (Chaerephon) major (Trouessart, 1897)
Otomops martiensseni (Matschie, 1897)
Otomops martiensseni (Matschie, 1897)
Hipposideros megalotis (Heuglin, 1861)
Nycteris macrotis Dobson, 1876
Myotis Kaup, 1829
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma microphyllum (Brünnich, 1782)
 MEGADERMATIDAE H. Allen, 1864
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Rhinolophus ferrumequinum (Schreber, 1774)
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Hipposideros Gray, 1831
 HIPPOSIDERIDAE Lydekker, 1891
Hipposideros Gray, 1831
Glauconycteris variegata (Tomes, 1861)
Epomophorus minimus Claessen & De Vree, 1991
Rhinolophus gorongosae Taylor, MacDonald, Goodman, Kearney, Cotterill, Stoffberg, Monadjem, Schoeman, Guyton, Naskrecki and Richards, 2018
Miniopterus minor Peters, 1867
Hypsugo musciculus (Thomas, 1913)
Laephotis wintoni Thomas, 1901
Laephotis Thomas, 1901
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Cistugo lesueuri Roberts, 1919
Nyctalus Bowdich, 1825
Hipposideros tephros (Cabrera, 1906)
Epomophorus grandis (Sanborn, 1950)
Miniopterus fraterculus Thomas and Schwann, 1906
Myonycteris Matschie, 1899

Common Name

Lesser Free-tailed Bat (English)
 Lesser Free-tailed Bats (English)
 Lesser Horseshoe Bat (English)
 Lesser House Bats (English)
 Lesser Leaf-nosed bat (English)
 Lesser leaf-nosed bat (English)
 Lesser Long-fingered Bat (English)
 Lesser Mascarene Flying Fox (English)
 Lesser Mascarene Flying-fox (English)
 Lesser Mastiff Bats (English)
 Lesser Mouse-tailed Bat (English)
 Lesser noctule (English)
 Lesser Noctule Bat (English)
 Lesser Nut-coloured House Bat (English)
 Lesser Rat-tailed Bat (English)
 Lesser Sudan Horn-skinned Bat (English)
 Lesser Trident Bats (English)
 Lesser Woolly Bat (English)
 Lesser Yellow Bat (English)
 Lesser Yellow House Bat (English)
 Lesser Yellow House Bat (English)
 Lesser Yellow House Bat (English)
 Lesueur se langhaarvlermuis (Afrikaans)
 Lesueur-langhaarvlermuis (Afrikaans)
 Lesueur's Hairy Bat (English)
 Lesueur's Myotis (English)
 Lesueur's Wing-gland Bat (English)
 Lesueurse Vlerkkiervlermuis (Afrikaans)
 létací lišky (Czech)
 létací psi (Czech)
 letaunowé (Czech)
 létavci (Czech)
 létavcovití (Czech)
 létavec Aellenův (Czech)

 létavec Glenův (Czech)
 létavec jihoafrický (Czech)
 létavec komorský (Czech)
 létavec maghrebský (Czech)

 létavec manavijský (Czech)
 létavec natalský (Czech)
 létavec nejmenší (Czech)
 létavec Petersonův (Czech)

 létavec pískový (Czech)
 létavec sesterský (Czech)

 Létavec stěhovavý (Czech)
 létavec tomášský (Czech)
 létavec východoafrický (Czech)
 létavec východomadagaskarský (Czech)
 létavec větší (Czech)
 Letouň lesní (Czech)
 Letouň zakrpený (Czech)
 letouni či netopýři (Czech)
 letouni lysoblaní (Czech)
 Leylan (Hebrew)
 Lgodenn-dall rous Egipt (Breton)
 Libesch Zwergfliedermaus (Luxembourgish)
 Libijski drobní netopir (Slovenian)
 Libijski patuljasti slijepi mišić (Montenegrin)
 Libyanlepakko (Finnish)
 Lichinjī (applied to all large bats) (Yao (Malawi))
 Lichinjī (applied to all largish bats) (Yao (Malawi))
 Lielais pakavdegūnis (Latvian)
 Lielais vakarsiksparnis (Latvian)
 Lietavec stahovavý (Slovak)
 Lightning Guano Bat (English)
 Liibüa nahkhiir (Estonian)
 Lilacul cu coadă liberă europeană (Romanian)
 Liliac-cârn (Romanian)
 Liliac-cu-aripi-lungi (Romanian)

Taxon Name

Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Mops (Chaerephon) Dobson, 1874
Rhinolophus hipposideros (Bechstein, 1800)
Scotoecus Thomas, 1901
Hipposideros caffer (Sundevall, 1846)
Hipposideros tephros (Cabrera, 1906)
Miniopterus fraterculus Thomas and Schwann, 1906
Pteropus subniger (Kerr, 1792)
Pteropus subniger (Kerr, 1792)
Mops (Chaerephon) Dobson, 1874
Rhinopoma cystops Thomas, 1903
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Scotophilus nucella Robbins, 1983
Rhinopoma cystops Thomas, 1903
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Asellia Gray, 1838
Kerivoula lanosa (A. Smith, 1847)
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Scotophilus leucogaster (Cretzschmar, 1826)
Scotophilus viridis (Peters, 1852)
Cistugo lesueuri Roberts, 1919
Cistugo lesueuri Roberts, 1919
Cistugo lesueuri Roberts, 1919
Cistugo lesueuri Roberts, 1919
Cistugo lesueuri Roberts, 1919
Cistugo lesueuri Roberts, 1919
Cistugo lesueuri Roberts, 1919
PTEROPODIDAE Gray, 1821
PTEROPODIDAE Gray, 1821
CHIROPTERA Blumenbach, 1779
Miniopterus Bonaparte, 1837
MINIOPTERIDAE Dobson, 1875
Miniopterus aelleni Goodman, Maminirina, Weyeneth, Bradman, Christidis, Ruedi and Appleton, 2009
Miniopterus gleni Peterson, Eger and Mitchell, 1995
Miniopterus fraterculus Thomas and Schwann, 1906
Miniopterus griveaudi Harrison, 1959
Miniopterus maghrebensis Puechmalle, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014
Miniopterus manavi Thomas, 1906
Miniopterus natalensis (A. Smith, 1833)
Miniopterus minor Peters, 1867
Miniopterus petersoni Goodman, Bradman, Maminirina, Ryan, Christidis & Appleton, 2008
Miniopterus arenarius Heller, 1912
Miniopterus sororculus Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton, 2007
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus newtoni Bocage, 1889
Miniopterus africanus Sanborn, 1936
Miniopterus majori Thomas, 1906
Miniopterus inflatus Thomas, 1903
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Epomophorus minor Dobson, 1880
Rhinolophus hildebrandtii Peters, 1878
Rhinolophus ferrumequinum (Schreber, 1774)
Nyctalus lasiopterus (Schreber, 1780)
Miniopterus schreibersii (Kuhl, 1817)
Tadarida fulminans (Thomas, 1903)
Scotoecus albobfuscus (Thomas, 1890)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Tadarida teniotis (Rafinesque, 1814)
Barbastella barbastellus (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)

Common Name

Liliac-cu-gene-lungi (Romanian)
 Liliac-cu-mustăți (Romanian)
 Liliac-cu-picioare-lungi (Romanian)
 Liliac-mare-cupotcoavă (Romanian)
 Liliac-mic-cupotcoavă (Romanian)
 Liliac-româneșc (Romanian)
 Liliac-sudic (Romanian)
 Liliacul cărămiziu (Romanian)
 Liliacul cârn (Romanian)
 Liliacul cu aripi late mediteranean (Romanian)
 Liliacul cu aripi lungi (Romanian)
 Liliacul cu burtă golașă (Romanian)
 Liliacul cu dungă albă (Romanian)
 Liliacul cu picioare lungi (Romanian)
 Liliacul cu potcoavă a lui Blasius (Romanian)
 Liliacul cu potcoavă a lui Mehely (Romanian)
 Liliacul de amurg din Azore (Romanian)
 Liliacul din Maghreb (Romanian)
 Liliacul fructivor egiptean (Romanian)
 Liliacul lui Botta (Romanian)
 Liliacul lui Savi (Romanian)
 Liliacul mare cu potcoavă (Romanian)
 Liliacul mare de amurg (Romanian)
 Liliacul mediteranean cu potcoavă (Romanian)
 Liliacul mic cu potcoavă (Romanian)
 Liliacul mic de amurg (Romanian)
 Liliacul mustăcio (Romanian)
 Liliacul pigmeu (Romanian)
 Liliacul pitic (Romanian)
 Liliacul pitic a lui Hanak (Romanian)
 Liliacul pitic al lui Kuhl (Romanian)
 Liliacul pitic din Madeira (Romanian)
 Liliacul urecheat de desert (Romanian)
 Liliacul urecheat din Tenerife (Romanian)
 Liliacul-lui-Blasius (Romanian)
 Liliacul-pitic mediteranean (Slovenian)
 Liliaci cu pliuri nazale (Romanian)
 Liliaci cu tragus (Romanian)
 Lille hestekonæse (Danish)
 Lilulwane (SiSwati)
 Lilulwane (SiSwati)
 Lilulwane (SiSwati)
 Ílmcoví kaloni (Czech)
 Liputiputi (applied to all small bats) (Yao (Malawi))
 Liputiputi (applied to all small bats) (Yao (Malawi))
 Lissonictère angolana (Italian)
 Lissonictère angolana (Italian)
 Lissonyctère d'Angola (French)
 Lissonyctère de Simth (French)
 Lissonyctère d'Ethiopie (French)
 Lissonyctère du Ruwenzori (French)
 Lissonyctère goliath (French)
 Lissonyctères (French)
 listonos africký (Czech)
 listonos trojzubcový (Czech)
 listonosovití (Czech)
 Liten hestekonesse (Norwegian)
 Little Brown Bats (English)
 Little Bulldog Bats (English)
 Little Collared Fruit Bat (English)
 Little Collared Fruit-bat (English)
 Little Collared Fruit-bats (English)
 Little Epauletted Fruit-bat (English)
 Little Flying Cow (English)
 Little Flying Cow (English)
 Little Free-tailed Bat (English)
 Little Goblin Bats (English)
 Little Mastiff Bats (English)
 Little Nut-coloured House Bat (English)
 Little Wrinkle-lipped Bat (English)
 Livingstone's House Bat (English)
 Livingstone-Hausfledermaus (German)
 Livingstones Flugfuchs (German)
 Livingstone's Flying Fox (English)

Taxon Name

Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis mystacinus (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Barbastella barbastellus (Schreber, 1774)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Miniopterus schreibersii (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Rhinolophus blasii Peters, 1867
Rhinolophus mehelyi Matschie, 1901
Nyctalus azoreum (Thomas, 1901)
Myotis punicus Felten, 1977
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Hypsugo savii (Bonaparte, 1837)
Rhinolophus ferrumequinum (Schreber, 1774)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus euryale Blasius, 1853
Rhinolophus hipposideros (Bechstein, 1800)
Nyctalus leisleri (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Otonycteris hemprichii Peters, 1859
Plecotus teneriffae Barrett-Hamilton, 1907
Rhinolophus blasii Peters, 1867
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
 RHINOLOPHIDAE Gray, 1825
 VESPERTILIONIDAE Gray, 1821
Rhinolophus hipposideros (Bechstein, 1800)
Cloeotis percivali Thomas, 1901
Miniopterus fraterculus Thomas and Schwann, 1906
Miniopterus natalensis (A. Smith, 1833)
Myonycteris Matschie, 1899
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Afronycteris nana (Peters, 1852)
Myonycteris angolensis (Bocage, 1898)
Myonycteris angolensis (Bocage, 1898)
Myonycteris angolensis (Bocage, 1898)
Myonycteris angolensis smithii (Thomas, 1908)
Myonycteris angolensis petraea (Bergmans, 1997)
Myonycteris angolensis ruwenzorii (Eisentraut, 1965)
Myonycteris angolensis goliath (Bergmans, 1997)
Myonycteris Matschie, 1899
Hipposideros caffer (Sundevall, 1846)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
 HIPPOSIDERIDAE Lydekker, 1891
Rhinolophus hipposideros (Bechstein, 1800)
Myotis Kaup, 1829
Mormopterus Peters, 1865
Myonycteris torquata (Dobson, 1878)
Myonycteris torquata (Dobson, 1878)
Myonycteris Matschie, 1899
Epomophorus labiatus (Temminck, 1837)
Nanonycteris Matschie, 1899
Nanonycteris veldkampii (Jentink, 1888)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Mormopterus Peters, 1865
Mormopterus Peters, 1865
Scotophilus nucella Robbins, 1983
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Scotophilus livingstonii Brooks and Bickham, 2014
Scotophilus livingstonii Brooks and Bickham, 2014
Pteropus livingstonii Gray, 1866
Pteropus livingstonii Gray, 1866

Common Name

Lobe-lipped Bats (English)
 Logodenn-dall kof noazh ar beziou (Breton)
 Lomba-leji Chamchika (Bengali)
 Long-crested Free-tailed Bat (English)
 Long-crested Gland-tailed Bat (English)
 Long-crested Wrinkle-lipped Bat (English)
 Long-eared bats (English)
 Long-eared bats (English)
 Long-eared Bats (English)
 Long-eared Desert Bat (English)
 Long-eared Desert Bats (English)
 Long-fingered Bat (English)
 Long-fingered Bats (English)
 Long-fingered Bats (English)
 Long-fingered Myotis (English)
 Long-haired Rousette (English)
 Long-haired Rousette (English)
 Long-tailed Bat (English)
 Long-tailed Greater Serotine Bat (English)
 Long-tailed House Bat (English)
 Long-tailed Serotine Bat (English)
 Long-wing Bat (English)
 Luftige Pipistrelle (German)
 Luftige Zwergfledermaus (German)
 Lybische Hufeisennase (German)
 lyronos žlutokřídý (Czech)
 lyronos srdcový (Czech)
 lyronosovití (Czech)
 Lytse Alpenflearmûs (Frisian)
 Lytse dwerchflearmûs (Frisian)
 Lytse hoefizernoas (Frisian)
 měkkosrstí kaloni (Czech)
 Mısır Meyve Yarasası (Turkish)
 Mažasis nakviša (Lithuanian)
 Mažasis pasagnosis (Lithuanian)
 Machado's Butterfly Bat (English)
 Machados Schmetterlingsfledermaus (German)
 MacInnes' Mausschwanz-Fledermaus (German)
 MacInnes's Mouse-tailed Bat (English)
 MacLaud's Horseshoe Bat (English)
 Macclauds Hufeisennase (German)
 Macroglosse de Woermann (French)
 Madagascan Flying Fox (English)
 Madagascan Flying-fox (English)
 Madagascan Free-tailed Bat (English)
 Madagascan Fruit Bat (English)
 Madagascan Large Free-tailed Bat (English)
 Madagascan Large Guano Bat (English)
 Madagascan Rousette (English)
 Madagascan Straw-coloured Fruit-bat (English)
 Madagascan white-bellied Free-tailed Bat (English)
 Madagascar cryptic leaf-nosed bat (English)

 Madagascar Flying Fox (English)
 Madagascar Free-tailed Bat (English)
 Madagascar Fruit Bat (English)
 Madagascar Large Free-tailed Bat (English)
 Madagascar Leaf-nosed Bat (English)
 Madagascar Mastiff Bat (English)
 Madagascar Rousette (English)
 Madagascar serotine (English)
 Madagascar Sheath-tailed Bat (English)

 Madagascar sheath-tailed bat (English)
 Madagascar Sucker-footed Bat (English)

 Madagascar White-bellied Free-tailed Bat (English)
 Madagascar Yellow House Bat (English)
 Madagaskar- Dreizack-Blattnasen (German)
 Madagaskar Mastino-Fledermaus (German)
 Madagaskar Pipistrelle (German)

 Madagaskar-Flugfuchs (German)

Taxon Name

Glauconycteris Dobson, 1875
Taphozous nudiventris Cretzschmar, 1830
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Mops (Chaerephon) chapini (J.A. Allen, 1917)
Mops (Chaerephon) chapini (J.A. Allen, 1917)
Mops (Chaerephon) chapini (J.A. Allen, 1917)
 NYCTERIDAE Van der Hoeven, 1855
Plecotus E. Geoffroy St.-Hilaire, 1818
Laephotis Thomas, 1901
Otonycteris hemprichii Peters, 1859
Otonycteris Peters, 1859
Myotis capaccinii (Bonaparte, 1837)
 MINIOPTERIDAE Dobson, 1875
Miniopterus Bonaparte, 1837
Myotis capaccinii (Bonaparte, 1837)
Myonycteris angolensis (Bocage, 1898)
Stenonycteris lanosus (Thomas, 1906)
Rhinopoma microphyllum (Brünnich, 1782)
Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
Miniopterus schreibersii (Kuhl, 1817)
Pipistrellus (Pipistrellus) aero Heller, 1912
Pipistrellus (Pipistrellus) aero Heller, 1912
Rhinolophus horaceki Benda and Vallo, 2012
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Cardioderma cor (Peters, 1872)
 MEGADERMATIDAE H. Allen, 1864
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus hipposideros (Bechstein, 1800)
Myonycteris Matschie, 1899
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus hipposideros (Bechstein, 1800)
Glauconycteris machadoi Hayman, 1963
Glauconycteris machadoi Hayman, 1963
Rhinopoma macinnesi Hayman, 1937
Rhinopoma macinnesi Hayman, 1937
Rhinolophus macclaudi Pousargues, 1898
Rhinolophus macclaudi Pousargues, 1898
Megaloglossus woermanni Pagenstecher, 1885
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Tadarida fulminans (Thomas, 1903)
Eidolon dupreanum (Schegel, 1867)
Tadarida fulminans (Thomas, 1903)
Tadarida fulminans (Thomas, 1903)
Rousettus madagascariensis G. Grandidier, 1929
Eidolon dupreanum (Schegel, 1867)
Mops (Chaerephon) leucogaster (A. Grandidier, 1869)
Macronycteris cryptovalorona (Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016)
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Otomops madagascariensis Dorst, 1953
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Tadarida fulminans (Thomas, 1903)
Paratriaenops furculus (Trouessart, 1907)
Otomops madagascariensis Dorst, 1953
Rousettus madagascariensis G. Grandidier, 1929
Laephotis matroka (Thomas and Schwann, 1905)
Coleura kibomalandy Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012
Paremballonura atrata (Peters, 1874)
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Mops (Mops) leucostigma (G.M. Allen, 1918)
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Paratriaenops Benda and Vallo, 2009
Mormopterus jugularis (Peters, 1865)
Laephotis malagasyensis (Peterson, Eger and Mitchell, 1995)
Pteropus rufus E. Geoffroy St.-Hilaire, 1803

Common Name	Taxon Name
Madagaskar-Freischwanzfledermaus (German)	<i>Paremballonura atrata</i> (Peters, 1874)
Madagaskar-Haftscheibenfledermäuse (German)	MYZOPODIDAE Thomas, 1904
Madagaskar-Haftscheibenfledermäuse (German)	<i>Myzopoda</i> Milne-Edwards and A. Grandidier, 1878
Madagaskar-Hausfledermaus (German)	<i>Scotophilus robustus</i> A. Milne-Edwards, 1881
Madagaskar-Höhlenflughund (German)	<i>Rousettus madagascariensis</i> G. Grandidier, 1929
Madagaskar-Langflügelfledermaus (German)	<i>Miniopterus sororculus</i> Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton, 2007
Madagaskar-Mausohr (German)	<i>Myotis goudoti</i> (A. Smith, 1834)
Madagaskar-Palmenflughund (German)	<i>Eidolon dupreanum</i> (Schegel, 1867)
Madagaskar-Rundblattnase (German)	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Madagaskar-Rundblattnase (German)	<i>Macronycteris gigas</i> (Wagner, 1845)
Madagaskar-Schiebeschwanzfledermaus (German)	<i>Coleura kibomalandy</i> Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012
Madagaskarse grootlosstertvlermuis (Afrikaans)	<i>Tadarida fulminans</i> (Thomas, 1903)
Madagaskar-Zwergfledermaus (German)	<i>Laephotis malagasyensis</i> (Peterson, Eger and Mitchell, 1995)
Madagassische Bulldoggfledermaus (German)	<i>Tadarida fulminans</i> (Thomas, 1903)
Madagassische Haftscheibenfledermaus (German)	MYZOPODIDAE Thomas, 1904
Madagassische hechtschijfvlermuizen (Dutch)	MYZOPODIDAE Thomas, 1904
Madagassische Riesen-Bulldoggfledermaus (German)	<i>Otomops madagascariensis</i> Dorst, 1953
Madeira dwerchflearmûs (Frisian)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeira dwergvlermuis (Dutch)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeira nahkhiir (Estonian)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeira pipistrelle (English)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeiradwergvlermuis (Dutch)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeiraflagermus (Danish)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeiraflaggermus (Norwegian)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeira-Fledermaus (German)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeirai törpedenevér (Hungarian)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeiran Pipistrelle Bat (English)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeiranpiippa (Finnish)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeirapipistrell (Swedish)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeira-Pipistrelle (German)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeiras sikspārnis (Latvian)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeira-Zwergfledermaus (German)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeira-Zwergfledermaus (Luxembourgish)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeirinis š ikš niukas (Lithuanian)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeirski šišmiš (Bosnian)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeirski mali šišmiš (Croatian)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeirski mali netopir (Slovenian)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Maderia Cüce Yarasaı (Turkish)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Maderiya və ya Kanar şəbpərəsi (Azerbaijani)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Maendeleo Horseshoe Bat (English)	<i>Rhinolophus maendeleo</i> Kock, Csorba and Howell, 2000
Maghreb mûsearflearmûs (Frisian)	<i>Myotis punicus</i> Felten, 1977
Maghrebian bent-wing bat (English)	<i>Miniopterus maghrebensis</i> Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014
Maghrebian Mouse-eared Bat (English)	<i>Myotis punicus</i> Felten, 1977
Maghrebijns langvleugelvleermuis (Dutch)	<i>Miniopterus maghrebensis</i> Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014
Maghreb-Langflügelfledermaus (German)	<i>Miniopterus maghrebensis</i> Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014
Maghreb-Mausohr (German)	<i>Myotis punicus</i> Felten, 1977
Maghreb-Mausouer (Luxembourgish)	<i>Myotis punicus</i> Felten, 1977
Magrebi denevér (Hungarian)	<i>Myotis punicus</i> Felten, 1977
Mahafaly-Langflügelfledermaus (German)	<i>Miniopterus mahafaliensis</i> Goodman, Bradman, Christides and Appleton, 2009
Mahgreb museøre (Danish)	<i>Myotis punicus</i> Felten, 1977
Mahgrebmusør re (Norwegian)	<i>Myotis punicus</i> Felten, 1977
Mahrep Farekulaklı Yarasaı (Turkish)	<i>Myotis punicus</i> Felten, 1977
Major Long-fingered Bat (English)	<i>Miniopterus majori</i> Thomas, 1906
Majors Langflügelfledermaus (German)	<i>Miniopterus majori</i> Thomas, 1906
Major's Long-fingered Bat (English)	<i>Miniopterus majori</i> Thomas, 1906
malý vrápenec podkovní (Czech)	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Malagasy Free-tailed Bat (English)	<i>Tadarida fulminans</i> (Thomas, 1903)
Malagasy Mouse-eared Bat (English)	<i>Myotis goudoti</i> (A. Smith, 1834)
Malagasy Serotine (English)	<i>Laephotis matroka</i> (Thomas and Schwann, 1905)
Malagasy Straw-colored Fruit Bat (English)	<i>Eidolon dupreanum</i> (Schegel, 1867)
Malagasy White-bellied Free-tailed Bat (English)	<i>Mops (Mops) leucostigma</i> (G.M. Allen, 1918)
Mali šišmiš (Bosnian)	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Mali šišmiš (Croatian)	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Mali brkati šišmiš (Bosnian)	<i>Myotis mystacinus</i> (Kuhl, 1817)

Common Name

Mali brkati šišmiš (Croatian)
 malí cíponosi (Czech)
 Mali Lajslerov šišmiš (Bosnian)
 malí morousi (Czech)
 Mali netopir (Slovenian)
 Mali noční šišmiš (Bosnian)
 mali podkovnjak (Slovenian)
 Mali podkovnjak (Slovenian)
 Mali potkovasti šišmiš (Bosnian)
 Mali potkovasti šišmiš (Croatian)
 Mali potkovičar (Montenegrin)
 Mali potkovnjak (Croatian)
 Mali slijepi mišić (Montenegrin)
 Mali večernjak (Croatian)
 Manavi Long-fingered Bat (English)
 Manavil Long-fingered Bat (English)
 Manavi-Langflügelfledermaus (German)
 Marisa's Leaf-nosed Bat (English)
 Marisas Rundblattnase (German)
 Marokkanische Rundblattnase (German)
 Marovaza House Bat (English)

 Marovaza-Hausfledermaus (German)

 Martiensen's Free-tailed Bat (English)
 Martienssen bat (English)
 Martienssen's Big-eared Bulldog Bat (English)
 Marungu-Rundblattnase (German)

 Marungu-Rundblattnase (German)
 Mascarene Flying-fox (English)
 Mastiff Bats (English)
 Mastiff Bats (English)
 Mastiff Bats (English)
 Mastino-Fledermäuse (German)
 Matroka Bat (English)
 Matroka-Pipistrelle (German)
 Matroka-Zwergfledermaus (German)
 Mauritian Flying Fox (English)
 Mauritian Tomb Bat (English)
 Mauritius Mastino-Fledermaus (German)
 Mauritius-Grabfledermaus (German)
 Maurizische Doggengrämmler (German)
 Mäuschen-Pipistrelle (German)
 Mäuschen-Zwergfledermaus (German)
 Mausohr (German)
 Mausohren (German)
 Mausohren (German)
 Mausschwanzfledermäuse (German)
 Mausschwanz-Fledermäuse (German)
 Mausschwanz-Fledermäuse (German)
 Mazais pakavdegūnis (Latvian)
 Mazais vakarsīkspārnis (Latvian)
 Méckefliedermaus (Luxembourgish)
 Medelhavshästskonäsa (Swedish)
 Medelhavsmusöra (Swedish)
 Mediteranski potkovasti šišmiš (Bosnian)
 Mediteranski potkovasti šišmiš (Croatian)
 Mediterranean Horseshoe Bat (English)
 Mediterranean Horseshoe Bat (English)
 Mediterranean Lesser pipistrelle (English)
 Mediterranean Long-eared Bat (English)
 Medje Free-tailed Bat (English)
 Medje Greater Free-tailed Bat (English)
 Medje Mops Bat (English)
 Medje-Bulldoggfledermaus (German)
 megaderma žlutokřídla (Czech)
 megaderma africká (Czech)
 Mégadermatidés (French)
 Megadermatidi (Italian)
 Megaderme à ailes orangées (French)
 Megaderme à ailes orangées (French)
 Mégadermes à ailes orangées (French)
 megadermovití (Czech)

Taxon Name

Myotis mystacinus (Kuhl, 1817)
Paratriaenops Benda and Vallo, 2009
Nyctalus leisleri (Kuhl, 1817)
Mops (Chaerephon) Dobson, 1874
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Nyctalus leisleri (Kuhl, 1817)
Miniopterus manavi Thomas, 1906
Miniopterus manavi Thomas, 1906
Miniopterus manavi Thomas, 1906
Hipposideros marisae Aellen, 1954
Hipposideros marisae Aellen, 1954
Hipposideros tephros (Cabrera, 1906)
Scotophilus marovaza Goodman, Ratrimomanarivo and Randrianandrianina, 2006
Scotophilus marovaza Goodman, Ratrimomanarivo and Randrianandrianina, 2006
Otomops martiensseni (Matschie, 1897)
Otomops martiensseni (Matschie, 1897)
Otomops martiensseni (Matschie, 1897)
Macronycteris commersoni (E. Geoffroy St.-Hilaire, 1813)
Macronycteris gigas (Wagner, 1845)
Pteropus niger (Kerr, 1792)
 MOLOSSIDAE Gervais, 1856
 Molossinae Gervais, 1856
Otomops Thomas, 1913
Mormopterus Peters, 1865
Laephotis matroka (Thomas and Schwann, 1905)
Laephotis matroka (Thomas and Schwann, 1905)
Laephotis matroka (Thomas and Schwann, 1905)
Pteropus niger (Kerr, 1792)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Mormopterus acetabulosus (Hermann, 1804)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Mormopterus acetabulosus (Hermann, 1804)
Hypsugo musciculus (Thomas, 1913)
Hypsugo musciculus (Thomas, 1913)
Myotis Kaup, 1829
Cistugo Thomas, 1912
Myotis Kaup, 1829
 RHINOPOMATIDAE Dobson, 1872
 RHINOPOMATIDAE Dobson, 1872
Rhinopoma E. Geoffroy St.-Hilaire, 1818
Rhinolophus hipposideros (Bechstein, 1800)
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus euryale Blasius, 1853
Myotis puniceus Felten, 1977
Rhinolophus euryale Blasius, 1853
Rhinolophus euryale Blasius, 1853
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Plecotus kolombatovici Dulic, 1980
Mops (Mops) congicus J.A. Allen, 1917
Mops (Mops) congicus J.A. Allen, 1917
Mops (Mops) congicus J.A. Allen, 1917
Mops (Mops) congicus J.A. Allen, 1917
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Cardioderma cor (Peters, 1872)
 MEGADERMATIDAE H. Allen, 1864
 MEGADERMATIDAE H. Allen, 1864
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Lavia Gray, 1838
 MEGADERMATIDAE H. Allen, 1864

Common Name

Taxon Name

Mégaloglosse de Woermann (French)	<i>Megaloglossus woermanni</i> Pagenstecher, 1885
Mégaloglosses (French)	<i>Megaloglossus</i> Pagenstecher, 1885
Mehēla pakavdegūnis (Latvian)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Méhely-Huffeisennues (Luxembourgish)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Meheli sagarnina (Estonian)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Mehelijev potkovičar (Montenegrin)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Mehelio pasagnosis (Lithuanian)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Meheljev potkovasti šišmiš (Bosnian)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Meheljev potkovnjak (Croatian)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Mehely Nalburunlu Yarasası (Turkish)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Mehely-hoefijzerneus (Dutch)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Mehely-Hufeisennase (German)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Mehelyi ferra-saguzar (Basque)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Mehelyjev podkovnjak (Slovenian)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Méhely-patkósdenevér (Hungarian)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Mehelys hestekonæse (Danish)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Mehely's hoefijzerneus (Dutch)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Mehely's hoefizernoas (Frisian)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Mehely's Horseshoe Bat (English)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Meridional serotine (English)	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
Metsälepakko (Finnish)	<i>Nyctalus leisleri</i> (Kuhl, 1817)
Mëttelmier-Huffeisennues (Luxembourgish)	<i>Rhinolophus euryale</i> Blasius, 1853
Microptère de Hayman (French)	<i>Epomophorus intermedius</i> (Hayman, 1963)
Midas Bat (English)	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Midas Free-tailed Bat (English)	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Midas Groove-cheeked Bat (English)	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Midas Mops Bat (English)	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Midas se losstertvlermuis (Afrikaans)	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Midas-Bulldoggfledermaus (German)	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Midas-losstertvlermuis (Afrikaans)	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Middelhavehestekonæse (Danish)	<i>Rhinolophus euryale</i> Blasius, 1853
Middelhavsfrihale (Norwegian)	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Middelhavshesteskonese (Norwegian)	<i>Rhinolophus euryale</i> Blasius, 1853
Middelhavshesteskonese (Norwegian)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Middelhavslangvinge (Norwegian)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Midge Bat (English)	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Mimétille (French)	<i>Mimetillus</i> Thomas, 1904
Mimétille de Moloney (French)	<i>Mimetillus moloneyi</i> (Thomas, 1891)
Mimétille de Thomas (French)	<i>Mimetillus thomasi</i> Hinton, 1920
Mimetilli (Italian)	<i>Mimetillus</i> Thomas, 1904
Mimetillo di Moloney (Italian)	<i>Mimetillus moloneyi</i> (Thomas, 1891)
Mimic Bats (English)	<i>Mimetillus</i> Thomas, 1904
Mindre brunfladdermis (Swedish)	<i>Nyctalus leisleri</i> (Kuhl, 1817)
Miniopter da l'ala lunga (Rhaeto-Romance)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Miniopter Schreibers (Breton)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Minioptère (French)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Minioptère à couronne (French)	<i>Miniopterus inflatus</i> Thomas, 1903
Minioptère à longues ailes (French)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Minioptère d'Afrique australe (French)	<i>Miniopterus fraterculus</i> Thomas and Schwann, 1906
Minioptère d'Afrique orientale (French)	<i>Miniopterus africanus</i> Sanborn, 1936
Minioptère de Montagne d'Ambre (French)	<i>Miniopterus ambohitrensis</i> Goodman, Ramasindrazana, Naughton and Appleton, 2015
Minioptère de Peterson (French)	<i>Miniopterus petersoni</i> Goodman, Bradman, Maminirina, Ryan, Christidis & Appleton, 2008
Minioptère de São Tomé (French)	<i>Miniopterus newtoni</i> Bocage, 1889
Minioptère de Schreibers (French)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Minioptère d'Eger (French)	<i>Miniopterus egeri</i> Goodman, Ramasindrazana, Maminirina, Schoeman and Appleton, 2011
Minioptère des Comores (French)	<i>Miniopterus griveaudi</i> Harrison, 1959
Minioptère du Maghreb (French)	<i>Miniopterus maghrebensis</i> Puechmaile, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014
Minioptère du Natal (French)	<i>Miniopterus natalensis</i> (A. Smith, 1833)
Minioptère malgache (French)	<i>Miniopterus gleni</i> Peterson, Eger and Mitchell, 1995
Minioptère minóre (Italian)	<i>Miniopterus minor</i> Peters, 1867
Minioptère minuscule (French)	<i>Miniopterus minor</i> Peters, 1867
Minioptères (French)	<i>Miniopterus Bonaparte</i> , 1837
Miniopteri i Schreibers-it (Albanian)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Minioptéridés (French)	<i>MINIOPTERIDAE</i> Dobson, 1875
Minioptéridi (Italian)	<i>MINIOPTERIDAE</i> Dobson, 1875
Minioptero di Schreïbers (Italian)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Minioptero maggióre di Thòmas (Italian)	<i>Miniopterus inflatus</i> Thomas, 1903
Miniotèridi (Italian)	<i>MINIOPTERIDAE</i> Dobson, 1875
Miniottero (Italian)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Miniottero di Schreïbers (Italian)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)

Common Name

Miniòttero maggióre di Thòmas (Italian)
 Miniòttero minóre (Italian)
 Minopter ad ala lunga (Rhaeto-Romance)
 Minor Epauletted Fruit Bat (English)
 Miombo pipistrelle (English)
 Misir meyvə yarasası (Azerbaijani)
 Mittelmeerhufeisennase (German)
 Mittelmeer-Hufeisennase (German)
 Mleme (Nyanja (Malawi))
 mmpān (plural) (Twi (Ghana))
 mmpāne (plural) (see Ohemeng *et al.*, 2017: 184)
 (Tanoboase (Ghana))
 Močvarni patuljasti šišmiš (Croatian)
 Mokombe (Kinyarwanda)
 Moloneyovi netopýři (Czech)
 Moloneys Flachkopf-Fledermaus (German)
 Moloney's Flat-headed Bat (English)
 Moloney's Flat-headed Vesper Bat (English)
 Moloney's Mimic Bat (English)
 Moloneys Schmalflügel-Fledermaus (German)
 Molos Cestoni (Breton)
 Molos europejski (Polish)
 Moloss bulldoc (Rhaeto-Romance)
 Molosse à crinière (French)
 Molosse à glandes caudales (French)
 Molosse à grandes oreilles (French)
 Molosse à grandes oreilles de Madagascar (French)
 Molosse d'Afrique du Sud (French)
 Molosse d'Angola (French)
 Molosse d'Ansorge (French)
 Molosse de Cestoni (French)
 Molosse de Madagascar (French)
 Molosse de Midas (French)
 Molosse d'Egypte (French)
 Molosse d'Europe (French)
 Molosse du Duc des Abruzzes (French)
 Molosse du Nigéria (French)
 Molosse du Zaïre (French)
 Molosse géant (French)
 Molosse Midas (French)
 Molosse pâle (French)
 Molosse tacheté (French)
 Molosses (French)
 Molosses (French)
 Molosses à grandes oreilles (French)
 Molossidés (French)
 Molòssidi (Italian)
 Molosso del Cestoni (Italian)
 Mongalla Free-tailed Bat (English)
 Mongalla Mops Bat (English)
 Mongalla-Bulldoggfledermaus (German)
 Mongallan Mops Bat (English)
 Montagne d'Ambre Long-fingered Bat (English)
 Mopek (Polish)
 Mopek zachodni (Polish)
 Mops Bats (English)
 Mops Free-tailed Bats (English)
 Mopsfledermaus (German)
 Mopsfledermäuse (German)
 Mopsilepakko (Finnish)
 Mopsvleermuis (Dutch)
 Morcego amarelo pequeno (Portuguese)
 Morcego anano (Galician (Spain))
 Morcego anao de Kuhl (Portuguese)
 Morcego bigotudo (Galician (Spain))
 Morcego borboleta (Portuguese)
 Morcego caseiro amarelo (Portuguese)
 Morcego caseiro de Rendall (Portuguese)
 Morcego caseiro do cabo (Portuguese)
 Morcego caseiro Hottentot (Portuguese)
 Morcego común (Galician (Spain))
 Morcego da Madeira (Portuguese)
 Morcego das bananeiras (Portuguese)

Taxon Name

Miniopterus inflatus Thomas, 1903
Miniopterus minor Peters, 1867
Miniopterus schreibersii (Kuhl, 1817)
Epomophorus minor Dobson, 1880
Neoromicia anchietae (Seabra, 1900)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rhinolophus euryale Blasius, 1853
Rhinolophus euryale Blasius, 1853
Epomophorus crypturus Peters, 1852
 CHIROPTERA Blumenbach, 1779
 CHIROPTERA Blumenbach, 1779
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Mimetillus Thomas, 1904
Mimetillus moloneyi (Thomas, 1891)
Mimetillus moloneyi (Thomas, 1891)
Mimetillus moloneyi (Thomas, 1891)
Mimetillus moloneyi (Thomas, 1891)
Mimetillus moloneyi (Thomas, 1891)
Tadarida teniotis (Rafinesque, 1814)
Tadarida teniotis (Rafinesque, 1814)
Tadarida teniotis (Rafinesque, 1814)
Mops (Chaerephon) chapini (J.A. Allen, 1917)
Mops (Chaerephon) bemmeleni (Jentink, 1879)
Tadarida lobata (Thomas, 1891)
Otomops madagascariensis Dorst, 1953
Mops (Chaerephon) ansorgei (Thomas, 1913)
Mops (Mops) condylurus (A. Smith, 1833)
Mops (Chaerephon) ansorgei (Thomas, 1913)
Tadarida teniotis (Rafinesque, 1814)
Tadarida fulminans (Thomas, 1903)
Mops (Mops) midas (Sundevall, 1843)
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Tadarida teniotis (Rafinesque, 1814)
Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
Mops (Chaerephon) nigeriae (Thomas, 1913)
Mops (Chaerephon) gallagheri (Harrison, 1975)
Tadarida ventralis (Heuglin, 1861)
Mops (Mops) midas (Sundevall, 1843)
Mops (Chaerephon) chapini (J.A. Allen, 1917)
Mops (Chaerephon) bivittatus (Heuglin, 1861)
 MOLOSSIDAE Gervais, 1856
 Molossinae Gervais, 1856
Otomops Thomas, 1913
 MOLOSSIDAE Gervais, 1856
 MOLOSSIDAE Gervais, 1856
Tadarida teniotis (Rafinesque, 1814)
Mops (Mops) demonstrator (Thomas, 1903)
Mops (Mops) demonstrator (Thomas, 1903)
Mops (Mops) demonstrator (Thomas, 1903)
Mops (Mops) demonstrator (Thomas, 1903)
Miniopterus ambohitrensis Goodman, Ramasindrazana, Naughton and Appleton, 2015
Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Mops Lesson, 1842
Mops Lesson, 1842
Barbastella barbastellus (Schreber, 1774)
Barbastella Gray, 1821
Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Glauconycteris variegata (Tomes, 1861)
Scotophilus dinganii (A. Smith, 1833)
Pseudoromicia rendalli (Thomas, 1889)
Laephotis capensis (A. Smith, 1829)
Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Afronycteris nana (Peters, 1852)

Common Name

Morcego das covas (Galician (Spain))
 Morcego das fragas (Galician (Spain))
 Morcego das hortas de Botta (Galician (Spain))
 Morcego das hortas meridional (Galician (Spain))
 Morcego das sepulturas das Maurícias (Portuguese)
 Morcego das sepulturas do egipto (Portuguese)
 Morcego de beira branca (Galician (Spain))
 Morcego de bigotes (Galician (Spain))
 Morcego de borde claro (Galician (Spain))
 Morcego de cabeça curta (Portuguese)
 Morcego de Capaccini (Portuguese)
 Morcego de cauda (Portuguese)
 Morcego de cauda livre (Portuguese)
 Morcego de cauda livre de Ansoorge (Portuguese)
 Morcego de cauda livre de ventre branco (Portuguese)
 Morcego de cauda livre do egipto (Portuguese)
 Morcego de cauda livre malhado (Portuguese)
 Morcego de ferradura de Blasius (Galician (Spain))
 Morcego de ferradura grande (Galician (Spain))
 Morcego de ferradura mediano (Galician (Spain))
 Morcego de ferradura mediterrâneo (Galician (Spain))
 Morcego de ferradura pequeno (Galician (Spain))
 Morcego de Hanak (Galician (Spain))
 Morcego de Hanak (Portuguese)
 Morcego de Hemprich (Portuguese)
 Morcego de Kuhl (Portuguese)
 Morcego de Leisler (Galician (Spain))
 Morcego de Madeira (Galician (Spain))
 Morcego de nariz enfolhado da cafraria (Portuguese)
 Morcego de nariz enfolhado de commerson (Portuguese)
 Morcego de nariz enfolhado de commerson (Portuguese)
 Morcego de orelhas fendidas (Galician (Spain))
 Morcego de pés grandes (Galician (Spain))
 Morcego de railer (Portuguese)
 Morcego de Savii (Portuguese)
 Morcego de Schlieffens (Portuguese)
 Morcego de schreiber (Portuguese)
 Morcego de tumba de cú espiado (Galician (Spain))
 Morcego do monte (Galician (Spain))
 Morcego dos Açores (Portuguese)
 Morcego exipcio (Galician (Spain))
 Morcego ferradura da Damarlandia (Portuguese)
 Morcego ferradura das savanas (Portuguese)
 Morcego ferradura de cabo (Portuguese)
 Morcego ferradura de Darling (Portuguese)
 Morcego ferradura de Dent (Portuguese)
 Morcego ferradura de Hildebrandt (Portuguese)
 Morcego ferradura de Lander (Portuguese)
 Morcego ferradura de nariz de sela (Portuguese)
 Morcego ferradura de Swinny (Portuguese)
 Morcego ferradura gigante (Portuguese)
 Morcego Frugívoro de Bocage (Portuguese)
 Morcego frugívoro de Peters (Portuguese)
 Morcego frugívoro de Wahlberg (Portuguese)
 Morcego frugívoro do Egipto (Portuguese)
 Morcego frugívoro gigante (Portuguese)
 Morcego gigante amarelo das casas (Portuguese)
 Morcego gigante de cauda livre (Portuguese)
 Morcego grande de dedos compridos (Portuguese)
 Morcego grande orelhudo (Portuguese)
 Morcego lamoso Harrison (Portuguese)
 Morcego lanoso da Damaralandia (Portuguese)
 Morcego lanudo de Bocage (Portuguese)
 Morcego lanudo de weltsch (Portuguese)
 Morcego mediterrâneo de ferradura (Galician (Spain))
 Morcego montañeiro (Galician (Spain))
 Morcego orelhudo do Egipto (Portuguese)
 Morcego orelhudo piloso (Portuguese)
 Morcego pequeno das casas (Portuguese)
 Morcego pequeno de cauda livre (Portuguese)
 Morcego pequeno de dedos compridos (Portuguese)
 Morcego pequeno de ferradura (Galician (Spain))
 Morcego rabudo (Galician (Spain))

Taxon Name

Miniopterus schreibersii (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Sauromys petrophilus (Roberts, 1917)
Myotis capaccinii (Bonaparte, 1837)
Coleura afra (Peters, 1852)
Mops (Mops) condylurus (A. Smith, 1833)
Mops (Chaerephon) ansorgei (Thomas, 1913)
Mops (Mops) niveiventer Cabrera and Ruxton, 1926
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Mops (Chaerephon) bivittatus (Heuglin, 1861)
Rhinolophus blasii Peters, 1867
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Otonycteris hemprichii Peters, 1859
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Hipposideros caffer (Sundevall, 1846)
Macronycteris commersoni (E. Geoffroy St.-Hilaire, 1813)
Macronycteris gigas (Wagner, 1845)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis capaccinii (Bonaparte, 1837)
Mops (Xiphonycteris) thersites (Thomas, 1903)
Hypsugo savii (Bonaparte, 1837)
Nycticeinops schlieffenii (Peters, 1859)
Miniopterus schreibersii (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Hypsugo savii (Bonaparte, 1837)
Nyctalus azoreum (Thomas, 1901)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus simulator K. Andersen, 1904
Rhinolophus capensis Lichtenstein, 1823
Rhinolophus darlingi K. Andersen, 1905
Rhinolophus denti Thomas, 1904
Rhinolophus hildebrandtii Peters, 1878
Rhinolophus landeri Martin, 1838
Rhinolophus blasii Peters, 1867
Rhinolophus swinnyi Gough, 1908
Rhinolophus clivosus Cretzschmar, 1828
Myonycteris angolensis (Bocage, 1898)
Epomophorus crypturus Peters, 1852
Epomophorus wahlbergi (Sundevall, 1846)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Eidolon helvum (Kerr, 1792)
Scotoophilus nigrita (Schreber, 1774)
Tadarida ventralis (Heuglin, 1861)
Miniopterus inflatus Thomas, 1903
Nycteris grandis Peters, 1865
Kerivoula lanosa (A. Smith, 1847)
Kerivoula argentata Tomes, 1861
Myotis bocagii (Peters, 1870)
Myotis welwitschii (Gray, 1866)
Rhinolophus euryale Blasius, 1853
Hypsugo savii (Bonaparte, 1837)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris hispida (Schreber, 1774)
Scotoecus albobfuscus (Thomas, 1890)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Miniopterus fraterculus Thomas and Schwann, 1906
Rhinolophus hipposideros (Bechstein, 1800)
Tadarida teniotis (Rafinesque, 1814)

Common Name

Morcego rateiro africano (Galician (Spain))
 Morcego tridentado (Portuguese)
 Morcego-anão (Portuguese)
 Morcego-arboricola-gigante (Portuguese)
 Morcego-arboricola-pequeno (Portuguese)
 Morcego-das-bananas (Portuguese)
 Morcego-de-bigodes (Portuguese)
 Morcego-de-ferradura de Blasius (Portuguese)
 Morcego-de-ferradura-grande (Portuguese)
 Morcego-de-ferradura-mediterrânico (Portuguese)
 Morcego-de-ferradura-mourisco (Portuguese)
 Morcego-de-ferradura-pequeno (Portuguese)
 Morcego-de-peluche (Portuguese)
 Morcego-de-ventre-nú (Portuguese)
 Morcego-frugívoro-egípcio (Portuguese)
 Morcego-hortelão de Botta (Portuguese)
 Morcego-hortelão-claro (Portuguese)
 Morcego-lanudo (Portuguese)
 Morcego-negro (Portuguese)
 Morcego-orelhudo-canarino (Portuguese)
 Morcego-pigmeu (Portuguese)
 Morcego-rabudo (Portuguese)
 Morcego-rato do Magrebe (Portuguese)
 Morcegos (Portuguese)
 Mòr-ialtag oidhcheach (Scottish Gaelic)
 morous širokouchý (Czech)
 morous africký (Czech)
 morous angolský (Czech)
 morous Ansoergeiv (Czech)
 morous bělobřichý (Czech)
 morous běloskrvný (Czech)
 morous chocholatý (Czech)
 morous egyptský (Czech)
 Morous evropský (Czech)
 morous guinejský (Czech)
 morous ještěří (Czech)
 morous jižní (Czech)
 morous kamerunský (Czech)
 morous kivuský (Czech)
 morous konžský (Czech)
 morous krátkokřídlý (Czech)
 morous krysový (Czech)
 morous liberijský (Czech)
 morous madagaskarský (Czech)
 morous malý (Czech)
 morous malgašský (Czech)

 morous Martienssenův (Czech)
 morous mauricijský (Czech)
 morous mongallanský (Czech)
 morous niangaranský (Czech)
 morous nigerijský (Czech)
 morous nubijský (Czech)
 morous oslí (Czech)
 morous Petersův (Czech)
 morous plochý (Czech)
 morous réunionský (Czech)

 morous sahelský (Czech)
 morous seychelský (Czech)
 morous skvrnitý (Czech)
 morous sněhobílý (Czech)
 morous spílavý (Czech)
 morous Stanleyův (Czech)
 morous tomášský (Czech)
 morous Trevorův (Czech)
 morous trpasličí (Czech)
 morous východoafrický (Czech)
 morous vévodský (Czech)
 morous západoafrický (Czech)
 morousovití (Czech)
 Morris' Mausohr (German)
 Morris's Bat (English)
 Morris's Mouse-eared Bat (English)

Taxon Name

Myotis punicus Felten, 1977
Cloeotis percivali Thomas, 1901
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Afronycteris nana (Peters, 1852)
Myotis mystacinus (Kuhl, 1817)
Rhinolophus blasii Peters, 1867
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Rhinolophus hipposideros (Bechstein, 1800)
Miniopterus schreibersii (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Barbastella barbastellus (Schreber, 1774)
Plecotus teneriffae Barrett-Hamilton, 1907
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Tadarida teniotis (Rafinesque, 1814)
Myotis punicus Felten, 1977
 CHIROPTERA Blumenbach, 1779
Nyctalus lasiopterus (Schreber, 1780)
Tadarida lobata (Thomas, 1891)
Tadarida ventralis (Heuglin, 1861)
Mops (Mops) condylurus (A. Smith, 1833)
Mops (Chaerephon) ansorgei (Thomas, 1913)
Mops (Chaerephon) leucogaster (A. Grandidier, 1869)
Mops (Mops) leucostigma (G.M. Allen, 1918)
Mops (Chaerephon) chapini (J.A. Allen, 1917)
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Tadarida teniotis (Rafinesque, 1814)
Mops (Chaerephon) russatus (J.A. Allen, 1917)
Sauromys petrophilus (Roberts, 1917)
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Mops (Xiphonycteris) petersoni (El Rayah, 1981)
Mops (Chaerephon) gallagheri (Harrison, 1975)
Mops (Mops) congicus J.A. Allen, 1917
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Myopterus daubentonii Desmarest, 1820
Mops (Chaerephon) bemmeleni (Jentink, 1879)
Otomops madagascariensis Dorst, 1953
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Mops (Chaerephon) jobimena (Goodman and Cardiff, 2004)
Otomops martiensseni (Matschie, 1897)
Mormopterus acetabulosus (Hermann, 1804)
Mops (Mops) demonstrator (Thomas, 1903)
Mops (Mops) niangarae J.A. Allen, 1917
Mops (Chaerephon) nigeriae (Thomas, 1913)
Mops (Chaerephon) major (Trouessart, 1897)
Mops (Mops) midas (Sundevall, 1843)
Mormopterus jugularis (Peters, 1865)
Platymops setiger (Peters, 1878)
Mormopterus francoismoutoui Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst, Bowie, 2008
Mops (Xiphonycteris) spurrelli (Dollman, 1911)
Mops (Chaerephon) pusillus (Miller, 1902)
Mops (Chaerephon) bivittatus (Heuglin, 1861)
Mops (Mops) niveiventer Cabrera and Ruxton, 1926
Mops (Xiphonycteris) thersites (Thomas, 1903)
Mops (Xiphonycteris) bakarii Stanley, 2009
Mops (Chaerephon) tomensis (Juste and Ibáñez, 1993)
Mops (Mops) trevori J.A. Allen, 1917
Mops (Xiphonycteris) nanulus J.A. Allen, 1917
Tadarida fulminans (Thomas, 1903)
Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
Myopterus whitleyi (Scharff, 1900)
 MOLOSSIDAE Gervais, 1856
Myotis morrisi Hill, 1971
Myotis morrisi Hill, 1971
Myotis morrisi Hill, 1971

Common Name

Taxon Name

Morris's Myotis (English)
 Mossambique-Hufeisennase (German)
 Mossambique-Langflügelfledermaus (German)
 motýlí netopýři (Czech)
 Mount Gargues Pipistrelle (English)
 Mount Mabu Horseshoe Bat (English)
 Mount Nimba Leaf-nosed Bat (English)
 Mount Nimba-Rundblattnase (German)
 Mountain Pipistrelle Bats (English)
 Mountain Rousette (English)
 Mouse-eared bat (English)
 Mouse-eared Bats (English)
 Mouselike Pipistrelle (English)
 Mouse-like Pipistrelle Bat (English)
 Mouse-tailed bats (English)
 Mouse-tailed bats (English)
 Mozambian Slit-faced Bat (English)
 Mozambican Horseshoe Bat (English)
 Mozambique Dämmerungsfledermaus (German)
 Mozambique long-fingered bat (English)
 Mozambique Sheath-tailed Bat (English)
 mračník bělobřichý (Czech)
 mračník Goodmanův (Czech)
 mračník guinejský (Czech)
 mračník madagaskarský (Czech)
 mračník ořechový (Czech)
 mračník olbřímí (Czech)
 mračník réunionský (Czech)
 mračník statný (Czech)
 mračník zelený (Czech)
 mračníkové (Czech)
 mračníkové (Czech)
 mrakolibové (Czech)
 Mroczek Boty (Polish)
 Mroczek pustynny (Polish)
 mroczkowate (Polish)
 Mt Gargues Pipistrelle (English)
 Mt Gargues Pipistrelle Bat (English)
 Mt Mabu Hufeisennase (German)
 Mückenfledermaus (German)
 Muisstaartveermuizen (Dutch)
 Mulima (Kinande (DRC))
 Mulima (Kiluba (DRC))
 Mulima (Kiluba (DRC))
 Mulima (Kiluba (DRC))
 Mulima (Kinande (DRC))
 Mulima (Kinande (DRC))
 Mulima (Kinande (DRC))
 Mulima (Kinande (DRC))
 Mulima (Kinande (DRC))
 Mulima (Kinande (DRC))
 Murciélago (Castilian (Spain))
 Murciélago Amarillo (Castilian (Spain))
 Murciélago bigotudo (Castilian (Spain))
 Murciélago común (Castilian (Spain))
 Murciélago dalmata de herradura (Castilian (Spain))
 Murciélago de Ala Larga (Castilian (Spain))
 Murciélago de borde claro (Castilian (Spain))
 Murciélago de bosque (Castilian (Spain))
 Murciélago de cabeza Plana (Castilian (Spain))
 Murciélago de Cabrera (Castilian (Spain))
 Murciélago de Cara Hueca (Castilian (Spain))
 Murciélago de cola libre y vientre desnudo (Castilian (Spain))
 Murciélago de cueva (Castilian (Spain))
 Murciélago de Geoffroy (Castilian (Spain))

Myotis morrisi Hill, 1971
Rhinolophus mossambicus Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Miniopterus mossambicus Monadjem, Goodman, Stanley and Appleton, 2013
Glauconycteris Dobson, 1875
Pipistrellus (Pipistrellus) aero Heller, 1912
Rhinolophus mabuensis Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Hipposideros lamottei Brosset, 1985
Hipposideros lamottei Brosset, 1985
Hypsugo Kolenati, 1856
Stenonycteris lanosus (Thomas, 1906)
Myotis mystacinus (Kuhl, 1817)
Myotis Kaup, 1829
Hypsugo musciculus (Thomas, 1913)
Hypsugo musciculus (Thomas, 1913)
 RHINOPOMATIDAE Dobson, 1872
Rhinopoma E. Geoffroy St.-Hilaire, 1818
Nycteris vinsoni Dalquest, 1965
Rhinolophus mossambicus Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Afronycteris nana (Peters, 1852)
Miniopterus mossambicus Monadjem, Goodman, Stanley and Appleton, 2013
Coleura afra (Peters, 1852)
Scotophilus leucogaster (Cretzschmar, 1826)
Scotophilus marovaza Goodman, Ratrimomanarivo and Randrianandrianina, 2006
Scotophilus nucella Robbins, 1983
Scotophilus tandrefana Goodman, Jenkins and Ratrimomanarivo, 2005
Scotophilus nux Thomas, 1904
Scotophilus nigrita (Schreber, 1774)
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Scotophilus robustus A. Milne-Edwards, 1881
Scotophilus viridis (Peters, 1852)
Scotophilinae Van Cakenberghe and Seamark, 2008
Scotophilus Leach, 1821
Scotophilus Leach, 1821
Eptesicus (Cnephæus) bottae (Peters, 1869)
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
 VESPERTILIONIDAE Gray, 1821
Pipistrellus (Pipistrellus) aero Heller, 1912
Pipistrellus (Pipistrellus) aero Heller, 1912
Rhinolophus mabuensis Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
 RHINOPOMATIDAE Dobson, 1872
Eidolon helvum (Kerr, 1792)
Epomophorus Bennett, 1836
Epomophorus pusillus Peters, 1868
Myonycteris angolensis (Bocage, 1898)
Myonycteris angolensis (Bocage, 1898)
Plerotes anchietae (Seabra, 1900)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Macronycteris gigas (Wagner, 1845)
Otomops martiensseni (Matschie, 1897)
Myotis welwitschii (Gray, 1866)
Hypsignathus monstrosus H. Allen, 1862
Pseudoromicia tenuipinnis (Peters, 1872)
Scotophilus nigrita (Schreber, 1774)
Myotis mystacinus (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rhinolophus blasii Peters, 1867
Miniopterus inflatus Thomas, 1903
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Mimetillus moloneyi (Thomas, 1891)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Nycteris arge Thomas, 1903
Taphozous nudiventris Cretzschmar, 1830
Miniopterus schreibersii (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)

Common Name

Murciélago de Hanak (Castilian (Spain))
 Murciélago de Herradura (Castilian (Spain))
 Murciélago de Herradura (Castilian (Spain))
 Murciélago de herradura de Blasius (Castilian (Spain))
 Murciélago de herradura mediterráneo (Castilian (Spain))
 Murciélago de la fruta egípcio (Castilian (Spain))
 Murciélago de las Tumbas (Castilian (Spain))
 Murciélago de Lengua Larga (Castilian (Spain))
 Murciélago de Madeira (Castilian (Spain))
 Murciélago de montaña (Castilian (Spain))
 Murciélago de Nariz de Hoja (Castilian (Spain))
 Murciélago de Orejas de Ratón (Castilian (Spain))
 Murciélago enan (Castilian (Spain))
 Murciélago Enano (Castilian (Spain))
 Murciélago enano (Castilian (Spain))
 Murciélago grande de herradura (Castilian (Spain))
 Murciélago Guanero (Castilian (Spain))
 Murciélago hortelano de Botta (Castilian (Spain))
 Murciélago hortelano mediterráneo (Castilian (Spain))
 Murciélago hortelano meridional (Castilian (Spain))
 Murciélago hortelano pardo (Castilian (Spain))
 Murciélago Mariposo (Castilian (Spain))
 Murciélago mediano de herradura (Castilian (Spain))
 Murciélago mediterráneo de herradura (Castilian (Spain))
 Murciélago montañero (Castilian (Spain))
 Murciélago nocturno (Castilian (Spain))
 Murciélago orejiroto (Castilian (Spain))
 Murciélago orejudo canario (Castilian (Spain))
 Murciélago patudo (Castilian (Spain))
 Murciélago pequeño de herradura (Castilian (Spain))
 Murciélago pigmeo (Castilian (Spain))
 Murciélago rabudo (Castilian (Spain))
 Murciélago ratonero bigotudo (Castilian (Spain))
 Murciélago ratonero moruno (Castilian (Spain))
 Murciélago ratonero pardo (Castilian (Spain))
 Murciélago ratonero patudo (Castilian (Spain))
 Murciélago ratonero Zenate (Castilian (Spain))

 Murcielagos (Castilian (Spain))
 Murcielagos chicos (Castilian (Spain))
 Murcielagos cola de raton o de cola libre (Castilian (Spain))
 Murciélagos de cola envainada (Castilian (Spain))
 Murciélagos de cola envainada (Castilian (Spain))
 Murin à bajoues argenté (French)
 Murin à bajoues d'Abo (French)
 Murin à bajoues d'Allen (French)
 Murin à bajoues de Béatrice (French)
 Murin à bajoues de Bibundi (French)
 Murin à bajoues de Glen (French)
 Murin à bajoues du Kenya (French)
 Murin à bajoues papillon (French)
 Murin à bajoues pie (French)
 Murin à moustaches (French)
 Murin à oreilles échancrées (French)
 Murin d'Angola (French)
 Murin d'Angola (French)
 Murin d'Anjouan (French)
 Murin de Capaccini (French)
 Murin de Du Bocage (French)
 Murin de Kock (French)
 Murin de Lesueur (French)
 Murin de Madagascar (French)
 Murin de Scott (French)
 Murin de Welwitsch (French)
 Murin d'Ethiopie (French)
 Murin du Maghreb (French)
 Murin roux (French)
 Murin tricolore (French)
 Murin velu du Cap (French)
 Murin Zenati (French)

 Murins (French)
 Murins à bajoues (French)

Taxon Name

Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Rhinolophus alcyone Temminck, 1853
Rhinolophus landeri Martin, 1838
Rhinolophus blasii Peters, 1867
Rhinolophus euryale Blasius, 1853
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Megaloglossus woermanni Pagenstecher, 1885
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Hypsugo savii (Bonaparte, 1837)
Hipposideros beatus (K. Andersen, 1906)
Myotis bocagii (Peters, 1870)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Epomophorus pusillus Peters, 1868
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Glauconycteris beatrix Thomas, 1901
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Hypsugo savii (Bonaparte, 1837)
Nyctalus leisleri (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Plecotus teneriffae Barrett-Hamilton, 1907
Myotis capaccinii (Bonaparte, 1837)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Tadarida teniotis (Rafinesque, 1814)
Myotis mystacinus (Kuhl, 1817)
Myotis punicus Felten, 1977
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis capaccinii (Bonaparte, 1837)
Myotis zenatius Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019
CHIROPTERA Blumenbach, 1779
VESPERTILIONIDAE Gray, 1821
MOLOSSIDAE Gervais, 1856
EMBALLONURIDAE Gervais, 1855
VESPERTILIONIDAE Gray, 1821
Glauconycteris argentata (Dobson, 1875)
Glauconycteris poensis (Gray, 1842)
Glauconycteris alboguttata J.A. Allen, 1917
Glauconycteris beatrix Thomas, 1901
Glauconycteris egeria Thomas, 1913
Glauconycteris gleni Peterson and Smith, 1973
Glauconycteris kenyacola Peterson, 1982
Glauconycteris variegata (Tomes, 1861)
Glauconycteris superba Hayman, 1939
Myotis mystacinus (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Cistugo seabrae Thomas, 1912
Myotis tricolor (Temminck, 1832)
Myotis anjouanensis Dorst, 1960
Myotis capaccinii (Bonaparte, 1837)
Myotis bocagii (Peters, 1870)
Myotis dieteri M. Happold, 2005
Cistugo lesueuri Roberts, 1919
Myotis goudoti (A. Smith, 1834)
Myotis scotti Thomas, 1927
Myotis welwitschii (Gray, 1866)
Myotis morrisi Hill, 1971
Myotis punicus Felten, 1977
Myotis bocagii (Peters, 1870)
Myotis tricolor (Temminck, 1832)
Myotis tricolor (Temminck, 1832)
Myotis zenatius Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019
Myotis Kaup, 1829
Glauconycteris Dobson, 1875

Common Name

Taxon Name

Murrcielago de cueva magrebí (Castilian (Spain))

Muskusbulfflearmûs (Frisian)
 Mustaschfladdermus (Swedish)
 Myonyctère d'Afrique orientale (French)
 Myonyctère de São Tomé (French)
 Myonyctères (French)
 Myoptère de Bini (French)
 Myoptère de Daubenton (French)
 Myoptère de Whitley (French)
 Myoptère du Sénégal (French)
 Myoptères (French)
 Myotis de Welwitsch (French)
 Myzopoda oreillard (French)

myzopoda ušatá (Czech)

Myzopodidés (French)
 myzopodovití (Czech)
 myzopody (Czech)
 Nøgenrumpet gravflagermus (Danish)
 Nəhəng axşam yarasası (Azerbaijani)
 Nacktbauch-Grabfledermaus (German)
 Nacktbäuchige Tempelfledermaus (German)
 Nagy patkósdenevér nagy patkósorrú denevér (Hungarian)
 Naked-bellied Taphozous (English)
 Naked-bellied Tomb Bat (English)
 Naked-rumped bat (English)
 Naked-rumped Tomb Bat (English)
 Nakengumpflaggermus (Norwegian)
 Nakengumpsfladdermuis (Swedish)
 Namib Long-eared Bat (English)
 Namibia-Langohrfledermaus (German)
 Namibian Long-eared Bat (English)
 Namib-langoorvlermuis (Afrikaans)
 Namoroka-Langflügelfledermaus (German)

Nanonyctère (French)
 Narrow-winged Bats (English)
 Nas fier-chaval grond (Rhaeto-Romance)
 Nas fier-chaval pitschen (Rhaeto-Romance)
 Natal Clinging Bat (English)
 Natal Flat-headed Bat (English)
 Natal Free-tailed Bat (English)
 Natal Long-fingered Bat (English)
 Natal Wrinkle-lipped Bat (English)
 Natalischen Schwirrfledermaus (German)
 Natal-Langflügelfledermaus (German)
 Natalse losstertvlermuis (Afrikaans)
 ndema (Comorian)
 Ndemia (Chewa (Malawi))
 Ndim (Azande (DRC))
 Ndim (Azande (DRC))
 ndrema (Comorian)
 Nectar Bat (English)
 nedopír širokouhý (Czech)
 Nedopír celouchý (Czech)
 Nedopír chlupatopěstný (Czech)
 Nedopír hvízdavý (Czech)
 Nedopír kašelný (Czech)
 nedopír walausatý (Czech)
 Nedopír wraubkowaný (Czech)
 Neoromicia Pipistrelle Bats (English)
 Neoromicie (Italian)
 netopýři (Czech)

netopýři širokouší (Czech)
 netopýři bradožlázi (Czech)
 netopýři buldočí (Czech)
 netopýři dlouhorucí (Czech)
 netopýři dlouhorucí (Czech)
 netopýři hladkonosí (Czech)
 netopýři holonosí (Czech)
 netopýři morousovití (Czech)

Miniopterus maghrebensis Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014
Tadarida teniotis (Rafinesque, 1814)
Myotis mystacinus (Kuhl, 1817)
Myonycteris relicta Bergmans, 1980
Myonycteris brachycephala (Bocage, 1889)
Myonycteris Matschie, 1899
Myopterus whiteleyi (Scharff, 1900)
Myopterus daubentonii Desmarest, 1820
Myopterus whiteleyi (Scharff, 1900)
Myopterus daubentonii Desmarest, 1820
Myopterus E. Geoffroy St.-Hilaire, 1818
Myotis welwitschii (Gray, 1866)
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
 MYZOPODIDAE Thomas, 1904
 MYZOPODIDAE Thomas, 1904
Myzopoda Milne-Edwards and A. Grandidier, 1878
Taphozous nudiventris Cretzschmar, 1830
Nyctalus lasiopterus (Schreber, 1780)
Taphozous nudiventris Cretzschmar, 1830
Taphozous nudiventris Cretzschmar, 1830
Rhinolophus ferrumequinum (Schreber, 1774)
Taphozous nudiventris Cretzschmar, 1830
Taphozous nudiventris Cretzschmar, 1830
Taphozous nudiventris Cretzschmar, 1830
Taphozous nudiventris Cretzschmar, 1830
Taphozous nudiventris Cretzschmar, 1830
Taphozous nudiventris Cretzschmar, 1830
Laephotis namibensis Setzer, 1971
Laephotis namibensis Setzer, 1971
Laephotis namibensis Setzer, 1971
Laephotis namibensis Setzer, 1971
Miniopterus brachytragos Goodman, Maminirina, Bradman, Christidis and Appleton, 2009
Nanonycteris veldkampii (Jentink, 1888)
Mimetillus Thomas, 1904
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Miniopterus natalensis (A. Smith, 1833)
Mormopterus acetabulosus (Hermann, 1804)
Mormopterus acetabulosus (Hermann, 1804)
Miniopterus natalensis (A. Smith, 1833)
Mormopterus acetabulosus (Hermann, 1804)
Scotophilus dinganii (A. Smith, 1833)
Miniopterus natalensis (A. Smith, 1833)
Mormopterus acetabulosus (Hermann, 1804)
Pteropus seychellensis A. Milne-Edwards, 1877
Afronycteris nana (Peters, 1852)
Epomophorus labiatus (Temminck, 1837)
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Pteropus seychellensis A. Milne-Edwards, 1877
Megaloglossus woermanni Pagenstecher, 1885
Miniopterus schreibersii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Myotis mystacinus (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Neoromicia Roberts, 1926
Neoromicia Roberts, 1926
 VESPERTILIONIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
Barbastella Gray, 1821
Taphozous E. Geoffroy St.-Hilaire, 1818
 MOLOSSIDAE Gervais, 1856
 MINIOPTERIDAE Dobson, 1875
Miniopterus Bonaparte, 1837
 EMBALLONURIDAE Gervais, 1855
 VESPERTILIONIDAE Gray, 1821
Tadarida Rafinesque, 1814

Common Name

netopýři myší (Czech)
 Netopýr černý (Czech)
 netopýr černý (Czech)
 Netopýr červenohlavý (Czech)
 netopýr šplhavý (Czech)
 netopýr abonský (Czech)
 Netopýr alpský (Czech)
 netopýr anžuánský (Czech)
 netopýr Anchietův (Czech)
 netopýr angolský (Czech)
 Netopýr azorský (Czech)
 netopýr bělobřichý (Czech)
 netopýr běloblanný (Czech)
 netopýr bělohrdlý (Czech)
 netopýr banánový (Czech)
 netopýr bečuánský (Czech)
 netopýr bibundinský (Czech)
 netopýr Bocageův (Czech)
 Netopýr Bottův (Czech)
 Netopýr brvitý (Czech)
 netopýr buldočí (Czech)
 Netopýr chlupatopestrý (Czech)
 netopýr damarský (Czech)
 netopýr damarský (Czech)
 Netopýr dlouhonožý (Czech)
 netopýr dvoubarvý (Czech)
 netopýr efulenský (Czech)
 netopýr Flowerův (Czech)
 netopýr galangánský (Czech)
 netopýr gambiánský (Czech)
 netopýr gargueský (Czech)
 netopýr Glenův (Czech)
 netopýr guinejský (Czech)
 netopýr habešský (Czech)
 Netopýr Hanákův (Czech)
 netopýr Hindeův (Czech)
 netopýr hnědý (Czech)
 netopýr hotentotský (Czech)
 netopýr hrobkový (Czech)
 Netopýr hvízdavý (Czech)
 netopýr iturijský (Czech)
 netopýr jižní (Czech)
 Netopýr jižní (Czech)
 netopýr kalabarský (Czech)
 netopýr kamerunský (Czech)
 Netopýr kanárský (Czech)
 netopýr kapský (Czech)
 netopýr keňský (Czech)
 netopýr Kockův (Czech)
 netopýr konžský (Czech)
 netopýr koptový (Czech)
 netopýr krátkouchý (Czech)
 Netopýr kyrénský (Czech)
 Netopýr Leislerův (Czech)
 Netopýr lysobřichý (Czech)
 netopýr Machadův (Czech)
 netopýr madagaskarský (Czech)
 Netopýr madeirský (Czech)
 Netopýr makaronéský (Czech)
 Netopýr malý (Czech)
 netopýr malgašský (Czech)
 netopýr mauricijský (Czech)
 netopýr mechový (Czech)
 netopýr medžeský (Czech)
 netopýr Moloneyův (Czech)
 Netopýr mopsličí (Czech)
 netopýr Morrisův (Czech)
 netopýr motýlí (Czech)
 netopýr namibský (Czech)
 Netopýr nejmenší (Czech)
 netopýr nigerijský (Czech)
 Netopýr obrovský (Czech)
 netopýr obtloustlý (Czech)
 Netopýr Ogněvův (Czech)

Taxon Name

Myotis Kaup, 1829
Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Afronycteris nana (Peters, 1852)
Glauconycteris poensis (Gray, 1842)
Hypsugo savii (Bonaparte, 1837)
Myotis anjouanensis Dorst, 1960
Neoromicia anchietae (Seabra, 1900)
Laephotis angolensis Monard, 1935
Nyctalus azureum (Thomas, 1901)
Scotophilus leucogaster (Cretzschmar, 1826)
Pseudoromicia tenuipinnis (Peters, 1872)
Scotoecus albigula Thomas, 1909
Afronycteris nana (Peters, 1852)
Laephotis botswanae Setzer, 1971
Glauconycteris egeria Thomas, 1913
Myotis bocagii (Peters, 1870)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Coleura afra (Peters, 1852)
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Laephotis capensis (A. Smith, 1829)
Myotis capaccinii (Bonaparte, 1837)
Vansonia rueppellii (J.B. Fischer, 1829)
Pipistrellus (Pipistrellus) nanulus Thomas, 1904
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Nycticeinops grandidieri (Dobson, 1876)
Scotoecus albofusus (Thomas, 1890)
Pipistrellus (Pipistrellus) aere Heller, 1912
Glauconycteris gleni Peterson and Smith, 1973
Nycticeinops eisentrauti (Hill, 1968)
Myotis scotti Thomas, 1927
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Scotoecus hindei Thomas, 1901
Laephotis capensis (A. Smith, 1829)
Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Glauconycteris superba Hayman, 1939
Pipistrellus (Pipistrellus) hesperidus (Temminck, 1840)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pseudoromicia brunnea (Thomas, 1880)
Pipistrellus (Pipistrellus) inexpectatus Aellen, 1959
Plecotus teneriffae Barrett-Hamilton, 1907
Laephotis capensis (A. Smith, 1829)
Glauconycteris kenyacola Peterson, 1982
Myotis dieteri M. Happold, 2005
Glauconycteris curryae Eger and Schlitter, 2001
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycticeinops schlieffenii (Peters, 1859)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Nyctalus leisleri (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Glauconycteris machadoi Hayman, 1963
Myotis goudoti (A. Smith, 1834)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Laephotis matroka (Thomas and Schwann, 1905)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Hypsugo musciculus (Thomas, 1913)
Glauconycteris humeralis J.A. Allen, 1917
Mimetillus moloneyi (Thomas, 1891)
Barbastella barbastellus (Schreber, 1774)
Myotis morrisi Hill, 1971
Glauconycteris variegata (Tomes, 1861)
Laephotis namibensis Setzer, 1971
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Eptesicus (Cnephæus) platyops (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Nycticeinops crassulus (Thomas, 1904)
Eptesicus (Cnephæus) bottae (Peters, 1869)

Common Name	Taxon Name
Netopýr pisklavý (Czech)	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
netopýr pouštní (Czech)	<i>Hypsugo ariel</i> (Thomas, 1904)
netopýr pruhovaný (Czech)	<i>Glauconycteris alboguttata</i> J.A. Allen, 1917
Netopýr punský (Czech)	<i>Myotis punicus</i> Felten, 1977
Netopýr pustinný (Czech)	<i>Otonycteris hemprichii</i> Peters, 1859
netopýr Rendallův (Czech)	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
netopýr rovníkový (Czech)	<i>Glauconycteris beatrix</i> Thomas, 1901
netopýr Rüppellův (Czech)	<i>Vansonia rueppellii</i> (J.B. Fischer, 1829)
netopýr savanový (Czech)	<i>Neoromicia guineensis</i> (Bocage, 1889)
Netopýr Saviův (Czech)	<i>Hypsugo savii</i> (Bonaparte, 1837)
netopýr Schlieffenův (Czech)	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
netopýr seychelský (Czech)	<i>Coleura seychellensis</i> Peters, 1868
netopýr slujový (Czech)	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
netopýr slunečný (Czech)	<i>Afronycteris helios</i> (Heller, 1912)
netopýr somálský (Czech)	<i>Neoromicia somalica</i> (Thomas, 1901)
netopýr stříbrný (Czech)	<i>Glauconycteris argentata</i> (Dobson, 1875)
Netopýr stromový (Czech)	<i>Nyctalus leisleri</i> (Kuhl, 1817)
netopýr tanganjický (Czech)	<i>Pipistrellus (Pipistrellus) permixtus</i> Aellen, 1957
Netopýr tripolitánský (Czech)	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
netopýr trojbarvý (Czech)	<i>Myotis tricolor</i> (Temminck, 1832)
netopýr vlašťovčí (Czech)	<i>Scotoecus hirundo</i> (de Winton, 1899)
netopýr vlnatý (Czech)	<i>Kerivoula africana</i> Dobson, 1878
Netopýr vousatý (Czech)	<i>Myotis mystacinus</i> (Kuhl, 1817)
Netopýr vrásofyskatý (Czech)	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Netopýr vroubený (Czech)	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
netopýr Welwitschův (Czech)	<i>Myotis welwitschii</i> (Gray, 1866)
netopýr Welwitschiův (Czech)	<i>Myotis welwitschii</i> (Gray, 1866)
netopýr Wintonův (Czech)	<i>Laephotis wintoni</i> Thomas, 1901
netopýr zelený (Czech)	<i>Scotophilus borbonicus</i> (E. Geoffroy St.-Hilaire, 1803)
netopýr zuluský (Czech)	<i>Neoromicia zuluensis</i> (Roberts, 1924)
Netopýra brvitého (Czech)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
netopýrcové (Czech)	<i>Myotinae</i> Tate, 1942
netopýrcové (Czech)	<i>Myotis</i> Kaup, 1829
Netopýrec brvitý (Czech)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
netopýrec vousatý (Czech)	<i>Myotis mystacinus</i> (Kuhl, 1817)
netopýrek ušatý (Czech)	<i>Myzopoda aurita</i> Milne-Edwards and A. Grandidier, 1878
netopýrkové ušatí (Czech)	<i>MYZOPODIDAE</i> Thomas, 1904
netopýrotvaří (Czech)	<i>VESPERTILIONIFORMACEI</i> Van Cakenberghe, Kearney and Seamark, 2007
netopýrov ci (Czech)	<i>VESPERTILIONOIDEA</i> Gray, 1821
netopýrovití (Czech)	<i>VESPERTILIONIDAE</i> Gray, 1821
Netopier brvitý (Slovak)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Netopier dlhonožý (Slovak)	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Netopier fúzatý (Slovak)	<i>Myotis mystacinus</i> (Kuhl, 1817)
Netopier magrebský (Slovak)	<i>Myotis punicus</i> Felten, 1977
Netopier stromový (Slovak)	<i>Nyctalus leisleri</i> (Kuhl, 1817)
Netzhäutigen Dämmerungsfledermaus (German)	<i>Pipistrellus (Pipistrellus) hesperidus</i> (Temminck, 1840)
Niangara Free-tailed Bat (English)	<i>Mops (Mops) niangarae</i> J.A. Allen, 1917
Niangara Mops Bat (English)	<i>Mops (Mops) niangarae</i> J.A. Allen, 1917
Niangara-Bulldoggfledermaus (German)	<i>Mops (Mops) niangarae</i> J.A. Allen, 1917
Nianganan Mops Bat (English)	<i>Mops (Mops) niangarae</i> J.A. Allen, 1917
Nicteride di Dobsòn (Italian)	<i>Nycteris macrotis</i> Dobson, 1876
Nicteride di Parisi (Italian)	<i>Nycteris parisii</i> (de Beaux, 1924)
Nicteride di Tèbe (Italian)	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
Nicteride irsùta (Italian)	<i>Nycteris hispida</i> (Schreber, 1774)
Nicteride ispida (Italian)	<i>Nycteris hispida</i> (Schreber, 1774)
Nicteride orecchiùta (Italian)	<i>Nycteris aurita</i> (K. Andersen, 1912)
Nicteride tebàna (Italian)	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
Nicteridi (Italian)	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795
Nicticèinope di Schlieffen (Italian)	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
Nicticèinopi (Italian)	<i>Nycticeinops</i> Hill & Harrison, 1987
Nigeria-Bulldoggfledermaus (German)	<i>Mops (Chaerephon) nigeriae</i> (Thomas, 1913)
Nigerian Free-tailed Bat (English)	<i>Mops (Chaerephon) nigeriae</i> (Thomas, 1913)
Nigerian Wrinkle-lipped Bat (English)	<i>Mops (Chaerephon) nigeriae</i> (Thomas, 1913)
Nigeriese losstertvlermuis (Afrikaans)	<i>Mops (Chaerephon) nigeriae</i> (Thomas, 1913)
Nijlrousette (Dutch)	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Nijlrousettus (Dutch)	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Niifluchhond (Luxembourgish)	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Niiflughund (Ägyptischer Höhlenflughund) (German)	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Niiflyghund (Swedish)	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Nilusi repülőkutya (Hungarian)	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)

Common Name

Taxon Name

Nimba long-fingered bat (English)

Miniopterus nimbae Monadjem, Shapiro, Richards, Karabulut, Crawley, Nielsen, Hansen, Bohmann and Mourier, 2020

Nimble Slit-faced Bat (English)

Nycteris arge Thomas, 1903

Nishpon Gadot (Hebrew)

Myotis capaccinii (Bonaparte, 1837)

Nishpon Pgoom Ozen (Hebrew)

Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)

Nitteride di Dòbson (Italian)

Nycteris macrotis Dobson, 1876

Nitteride di Parisi (Italian)

Nycteris parisii (de Beaux, 1924)

Nitteride di Tèbe (Italian)

Nycteris thebaica E. Geoffroy St.-Hilaire, 1818

Nitteride irsùta (Italian)

Nycteris hispida (Schreber, 1774)

Nitteride ispida (Italian)

Nycteris hispida (Schreber, 1774)

Nitteride orecchiùta (Italian)

Nycteris aurita (K. Andersen, 1912)

Nitteride tebàna (Italian)

Nycteris thebaica E. Geoffroy St.-Hilaire, 1818

Nitteridi (Italian)

Nycteris G. Cuvier and E. Geoffroy, 1795

Nitticèinope di Schlieffen (Italian)

Nycticeinops schlieffenii (Peters, 1859)

Nitticèinopi (Italian)

Nycticeinops Hill & Harrison, 1987

Nkulo (Azande (DRC))

Hipposideros caffer (Sundevall, 1846)

noční kaloni (Czech)

Rousettus Gray, 1821

Noack's Leaf-nosed Bat (English)

Hipposideros ruber (Noack, 1893)

Noack's Roundleaf Bat (English)

Hipposideros ruber (Noack, 1893)

Noacks Rundblattnase (German)

Hipposideros ruber (Noack, 1893)

Nocek długopalcy (Polish)

Myotis capaccinii (Bonaparte, 1837)

Nocek orzęsiony (Polish)

Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)

Nocek punicki (Polish)

Myotis punicus Felten, 1977

Nocek wasatek (Polish)

Myotis mystacinus (Kuhl, 1817)

nocki (Polish)

Myotis Kaup, 1829

Noctophile d'Angola (French)

Laephotis angolensis Monard, 1935

Noctophile de De Winton (French)

Laephotis wintoni Thomas, 1901

Noctophile du Botswana (French)

Laephotis botswanae Setzer, 1971

Noctophile du Namib (French)

Laephotis namibensis Setzer, 1971

Noctophiles (French)

Laephotis Thomas, 1901

Nòtutl de les Azores (Catalan (Spain))

Nyctalus azoreum (Thomas, 1901)

Nòtutl gegant (Catalan (Spain))

Nyctalus lasiopterus (Schreber, 1780)

Nòtutl petit (Catalan (Spain))

Nyctalus leisleri (Kuhl, 1817)

Noctule Bats (English)

Nyctalus Bowdich, 1825

Noctule de Leisler (French)

Nyctalus leisleri (Kuhl, 1817)

Noctule des Açores (French)

Nyctalus azoreum (Thomas, 1901)

Noctule géante (French)

Nyctalus lasiopterus (Schreber, 1780)

Noctules (English)

Nyctalus Bowdich, 1825

Noctules (French)

Nyctalus Bowdich, 1825

Nóctulo das Azores (Galician (Spain))

Nyctalus azoreum (Thomas, 1901)

Nóctulo de Azores (Castilian (Spain))

Nyctalus azoreum (Thomas, 1901)

Noctulo de Leisler (Castilian (Spain))

Nyctalus leisleri (Kuhl, 1817)

Nóctulo gigante (Castilian (Spain))

Nyctalus lasiopterus (Schreber, 1780)

Nóctulo grande (Castilian (Spain))

Nyctalus lasiopterus (Schreber, 1780)

Nóctulo menor (Castilian (Spain))

Nyctalus leisleri (Kuhl, 1817)

Nóctulo pequeno (Galician (Spain))

Nyctalus leisleri (Kuhl, 1817)

Nóctulo pequeño (Castilian (Spain))

Nyctalus leisleri (Kuhl, 1817)

Nóctulo xigante (Galician (Spain))

Nyctalus lasiopterus (Schreber, 1780)

Noisy Fruit-bats (English)

Epomops Gray, 1866

Noktula Żgħir (Maltese)

Nyctalus leisleri (Kuhl, 1817)

Noktula Kbir (Maltese)

Nyctalus lasiopterus (Schreber, 1780)

Noktula ta' l-Azores (Maltese)

Nyctalus azoreum (Thomas, 1901)

Noktulë e Azoreve (Albanian)

Nyctalus azoreum (Thomas, 1901)

Noktulë e Leisler-it (Albanian)

Nyctalus leisleri (Kuhl, 1817)

Noktulë gjigante (Albanian)

Nyctalus lasiopterus (Schreber, 1780)

nosalec malolupenný (Czech)

Rhinopoma microphyllum (Brünnich, 1782)

nosalecové (Czech)

Rhinopoma E. Geoffroy St.-Hilaire, 1818

Notch-eared Bat (English)

Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)

Nottola delle Azzorre (Italian)

Nyctalus azoreum (Thomas, 1901)

Nottola di Leisler (Italian)

Nyctalus leisleri (Kuhl, 1817)

Nottola gigante (Italian)

Nyctalus lasiopterus (Schreber, 1780)

Nozigell an Azorez (Breton)

Nyctalus azoreum (Thomas, 1901)

Nozigell Leisler (Breton)

Nyctalus leisleri (Kuhl, 1817)

Nozigell vras (Breton)

Nyctalus lasiopterus (Schreber, 1780)

Nucella House Bat (English)

Scotophilus nucella Robbins, 1983

Nußfarbene Hausfledermaus (German)

Scotophilus nux Thomas, 1904

Nut-colored Yellow Bat (English)

Scotophilus nux Thomas, 1904

Nut-coloured House Bat (English)

Scotophilus nux Thomas, 1904

Nux Yellow House Bat (English)

Scotophilus nux Thomas, 1904

Nyanza serotine (English)

Pseudoromicia nyanza Monadjem, Patterson, Webala and Demos, 2020

Nyctère à grandes oreilles (French)

Nycteris macrotis Dobson, 1876

Nyctère à longues oreilles (French)

Nycteris aurita (K. Andersen, 1912)

Common Name

Nyctère d'Aellen (French)
 Nyctère d'Andersen (French)
 Nyctère de Bates (French)
 Nyctère de Dobson (French)
 Nyctère de Gambie (French)
 Nyctère de Ja (French)
 Nyctère de la Thébaïde (French)
 Nyctère de Parisi (French)
 Nyctère de Vinson (French)
 Nyctère de Wood (French)
 Nyctère du Mozambique (French)
 Nyctère égyptienne (French)
 Nyctère hérissée (French)
 Nyctère hirsute (French)
 Nyctère moyen (French)
 Nyctère naine (French)
 Nyctères (French)
 Nyctères (French)
 Nyctéridés (French)
 Nycteris i alpios (Hungarian)
 Nycteris i blepharoti (Hungarian)
 Nycteris i kini (Hungarian)
 Nycteris i leukogyros (Hungarian)
 Nycteris i macropterys (Hungarian)
 Nycteris i macropus (Hungarian)
 Nycteris i mikromolossos (Hungarian)
 Nycteris i mystakophoros (Hungarian)
 Nycteris i urophoros (Hungarian)
 Nycteris pterygisti i megali (Hungarian)
 Nycteris pterygisti i mikra (Hungarian)
 nycteris velká (Czech)
 Nycticeinops (French)
 Nycticeinops de Schlieffen (French)
 nykteridoviti (Czech)
 nykteridy (Czech)
 nykteris egyptská (Czech)
 nykteris velkouchá (Czech)
 Nyrlûsette (Frisian)
 Nyugati pisedenevér (Hungarian)
 obecny (Czech)
 Okenlångöra (Swedish)
 Old World Big-eared Bats (English)
 Old World Fruit bats (English)
 Old World Long-eared Bats (English)
 Old World Sucker-footed Bat (English)
 Old-World leaf-nosed bat (English)
 Olivenbraune Schwirrfledermaus (German)
 Olivengrüne Schwirrfledermaus (German)
 Ollialtóg nochtúil (Irish Gaelic)
 Onechte vampieren (Dutch)
 Orecchione di Gaisler (Italian)
 Orecchione di Hemprich (Italian)
 Orecchione di Kolombatovic (Italian)
 Orecchione di Tenerife (Italian)
 Oreillard d'Angola (French)
 Oreillard de De Winton (French)
 Oreillard de Namibie (French)
 Oreillard de Ténérife (French)
 Oreillard d'Egypte (French)
 Oreillard Des Balkans (French)
 Oreillard d'Ethiopie (French)
 Oreillard d'Hemprich (French)
 Oreillard du Botswana (French)
 Oreillard du Gaisler (French)
 Oreillard du Namib (French)
 Oreillards (French)
 Oreillards d'Afrique (French)
 Oreillards du désert (French)
 Orejudo canario (Castilian (Spain))
 Orejudo de Hemprich (Castilian (Spain))
 Orelludo canario (Galician (Spain))
 Orelludo de Hemprich (Galician (Spain))
 Orellut canari (Catalan (Spain))

Taxon Name

Nycteris intermedia Aellen, 1959
Nycteris aurita (K. Andersen, 1912)
Nycteris arge Thomas, 1903
Nycteris macrotis Dobson, 1876
Nycteris gambiensis (K. Andersen, 1912)
Nycteris major (K. Andersen, 1912)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris parisii (de Beaux, 1924)
Nycteris vinsoni Dalquest, 1965
Nycteris woodi K. Andersen, 1914
Nycteris vinsoni Dalquest, 1965
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris hispida (Schreber, 1774)
Nycteris hispida (Schreber, 1774)
Nycteris intermedia Aellen, 1959
Nycteris nana (K. Andersen, 1912)
 NYCTERIDAE Van der Hoeven, 1855
Nycteris G. Cuvier and E. Geoffroy, 1795
 NYCTERIDAE Van der Hoeven, 1855
Hypsugo savii (Bonaparte, 1837)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Barbastella barbastellus (Schreber, 1774)
Myotis mystacinus (Kuhl, 1817)
Tadarida teniotis (Rafinesque, 1814)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Nycteris grandis Peters, 1865
Nycticeinops Hill & Harrison, 1987
Nycticeinops schlieffenii (Peters, 1859)
 NYCTERIDAE Van der Hoeven, 1855
Nycteris G. Cuvier and E. Geoffroy, 1795
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris macrotis Dobson, 1876
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Barbastella barbastellus (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Otonycteris hemprichii Peters, 1859
Plecotus E. Geoffroy St.-Hilaire, 1818
 PTEROPODIDAE Gray, 1821
Plecotus E. Geoffroy St.-Hilaire, 1818
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
 HIPPOSIDERIDAE Lydekker, 1891
Scotophilus leucogaster (Cretzschmar, 1826)
Scotophilus viridis (Peters, 1852)
Nyctalus lasiopterus (Schreber, 1780)
 MEGADERMATIDAE H. Allen, 1864
Plecotus gaisleri Benda, Kiefer, Hanak & Veith, 2004
Otonycteris hemprichii Peters, 1859
Plecotus kolombatovici Dulic, 1980
Plecotus teneriffae Barrett-Hamilton, 1907
Laephotis angolensis Monard, 1935
Laephotis wintoni Thomas, 1901
Laephotis namibensis Setzer, 1971
Plecotus teneriffae Barrett-Hamilton, 1907
Plecotus christii Gray, 1838
Plecotus kolombatovici Dulic, 1980
Plecotus balensis Kruskop & Lavrenchenko, 2000
Otonycteris hemprichii Peters, 1859
Laephotis botswanae Setzer, 1971
Plecotus gaisleri Benda, Kiefer, Hanak & Veith, 2004
Laephotis namibensis Setzer, 1971
Plecotus E. Geoffroy St.-Hilaire, 1818
Laephotis Thomas, 1901
Otonycteris Peters, 1859
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859
Plecotus teneriffae Barrett-Hamilton, 1907

Common Name

Orellut d'Hemprich (Catalan (Spain))
 Óriás koraidenevér (Hungarian)
 Östliche Haftscheibenfledermaus (German)

Östliche Mopsfledermaus (German)
 Otocarlino (Italian)
 Otocarlino di Martienssèn (Italian)
 ouška myši (Czech)
 Oznan (Hebrew)
 příšerec egipecký (Czech)
 příšerecové (Czech)
 přísavkovci (Czech)
 přísavkovcovití (Czech)
 přísavkovec bělobřichý (Czech)

přísavkovec ušatý (Czech)

Pūnijas naktssikspārnis (Latvian)
 Paarse hoefijzerneus (Dutch)
 Pale Free-tailed Bat (English)
 Pale Xantharpy (English)
 Paljaskõht-kääpanahkhiir (Estonian)
 Palmenflughund (German)
 Palmenflughunde (German)
 Pålsvingad brunfladdermus (Swedish)
 Panda Bat (English)
 Papo ("Congolese")
 Papo ("Congolese")
 Parisis Schlitznasen-Fledermaus (German)
 Parisi's Slit-faced Bat (English)
 Parissi's Slit-faced Bat (English)
 Parkpipistrell (Swedish)
 Parsaf Bahir (Hebrew)
 parsaf gadol (Hebrew)
 Parsaf Gamadi (Hebrew)
 Parsaf HaNegev (Hebrew)
 Parsaf Hiver (Hebrew)
 Parsaf Matsui (Hebrew)
 parsaf matzuy (Hebrew)
 Parsafon (Hebrew)
 pasafon (Hebrew)
 Pashaf Matsui (Hebrew)
 Patrizis Dreizackblattnase (German)
 Patrizi's Lesser Trident Bat (English)
 Patrizi's Trident Leaf-nosed Bat (English)
 Patuljasti šišmiš (Croatian)
 Patuljasti šišmiš (Bosnian)
 Patuljasti slijepi mišić (Bosnian)
 Patuljasti slijepi mišić (Montenegrin)
 Paulian's Trident Bat (English)
 pavrápencovití (Czech)
 pavrápenec abánský (Czech)
 pavrápenec Aellenův (Czech)
 pavrápenec africký (Czech)
 pavrápenec Commersonův (Czech)

pavrápenec eritrejský (Czech)
 pavrápenec guinejský (Czech)
 pavrápenec Jonesův (Czech)
 pavrápenec kamerunský (Czech)
 pavrápenec krátkoocasý (Czech)
 pavrápenec Kyklop (Czech)
 pavrápenec natalský (Czech)
 pavrápenec nimbánský (Czech)
 pavrápenec obrovský (Czech)
 pavrápenec pruhovaný (Czech)
 pavrápenec sahelský (Czech)
 pavrápenec sazový (Czech)
 pavrápenec trojzubcový (Czech)
 pavrápenec ušatý (Czech)
 Peak-saddle Horseshoe Bat (English)
 Pearse hoefizernoas (Frisian)
 Pel's Pouched Bat (English)
 Pels Taschenfledermaus (German)

Taxon Name

Otonycteris hemprichii Peters, 1859
Nyctalus lasiopterus (Schreber, 1780)
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Barbastella leucomelas (Cretzschmar, 1826)
Otomops Thomas, 1913
Otomops martiensseni (Matschie, 1897)
Myotis Kaup, 1829
Plecotus christii Gray, 1838
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Mops (Chaerephon) Dobson, 1874
Myzopoda Milne-Edwards and A. Grandidier, 1878
 MYZOPODIDAE Thomas, 1904
Myzopoda schliemanni Goodman, Rakotondraparany and Kofoky, 2007
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Myotis punicus Felten, 1977
Rhinolophus euryale Blasius, 1853
Mops (Chaerephon) chapini (J.A. Allen, 1917)
Eidolon helvum (Kerr, 1792)
Taphozous nudiventris Cretzschmar, 1830
Eidolon helvum (Kerr, 1792)
Eidolon Rafinesque, 1815
Nyctalus lasiopterus (Schreber, 1780)
Glauconycteris superba Hayman, 1939
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Taphozous nudiventris Cretzschmar, 1830
Nycteris parisii (de Beaux, 1924)
Nycteris parisii (de Beaux, 1924)
Nycteris parisii (de Beaux, 1924)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Rhinolophus euryale Blasius, 1853
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus mehelyi Matschie, 1901
Rhinolophus blasii Peters, 1867
Rhinolophus blasii Peters, 1867
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Tadarida teniotis (Rafinesque, 1814)
Asellia patrizii de Beaux, 1931
Asellia patrizii de Beaux, 1931
Asellia patrizii de Beaux, 1931
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Paratriaenops pauliani (Goodman and Ranivo, 2008)
 HIPPOSIDERIDAE Lydekker, 1891
Hipposideros abae J.A. Allen, 1917
Hipposideros marisae Aellen, 1954
Hipposideros caffer (Sundevall, 1846)
Macronycteris commersoni (E. Geoffroy St.-Hilaire, 1813)
Asellia patrizii de Beaux, 1931
Hipposideros beatus (K. Andersen, 1906)
Hipposideros jonesi Hayman, 1947
Doryrhina camerunensis (Eisentraut, 1956)
Hipposideros curtus G.M. Allen, 1921
Doryrhina cyclops (Temminck, 1853)
Hipposideros caffer (Sundevall, 1846)
Hipposideros lamottei Brosset, 1985
Macronycteris gigas (Wagner, 1845)
Macronycteris vittatus (Peters, 1852)
Hipposideros tephros (Cabrera, 1906)
Hipposideros fuliginosus (Temminck, 1853)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Hipposideros megalotis (Heuglin, 1861)
Rhinolophus blasii Peters, 1867
Rhinolophus euryale Blasius, 1853
Saccolaimus peli (Temminck, 1853)
Saccolaimus peli (Temminck, 1853)

Common Name

Taxon Name

Pemba Flying Fox (English)	<i>Pteropus voeltzkowi</i> Matschie, 1909
Pemba Flying-fox (English)	<i>Pteropus voeltzkowi</i> Matschie, 1909
Pemba vliegende hond (Dutch)	<i>Pteropus voeltzkowi</i> Matschie, 1909
Pemba-Bulldoggfledermaus (German)	<i>Mops (Xiphonycteris) bakarii</i> Stanley, 2009
Pemba-Flugfuchs (German)	<i>Pteropus voeltzkowi</i> Matschie, 1909
Percivals Kleinohr-Dreizackblattnase (German)	<i>Cloeotis percivali</i> Thomas, 1901
Percival's Short-eared Trident Bat (English)	<i>Cloeotis percivali</i> Thomas, 1901
Percival's Trident Bat (English)	<i>Cloeotis percivali</i> Thomas, 1901
Perforated Taphozous bat (English)	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
Pergamentflügel-Fledermäuse (German)	<i>Myotis</i> E. Geoffroy St.-Hilaire, 1818
Peters' Epaulettenflughund (German)	<i>Epomophorus crypturus</i> Peters, 1852
Peters' Flachkopf-Bulldoggfledermaus (German)	<i>Platymops setiger</i> (Peters, 1878)
Peters' Kleiner Epaulettenflughund (German)	<i>Epomophorus pusillus</i> Peters, 1868
Peters' Lesser Epauletted Fruit Bat (English)	<i>Epomophorus pusillus</i> Peters, 1868
Peters' Lesser Fruit-bat (English)	<i>Epomophorus pusillus</i> Peters, 1868
Peters se witkolvrugtevlermuis (Afrikaans)	<i>Epomophorus crypturus</i> Peters, 1852
Peter's Sheath-tailed Bat (English)	<i>Paremballonura atrata</i> (Peters, 1874)
Peter's Trident Bat (English)	<i>Trienops afer</i> Peters, 1877
Peters' Wrinkle-lipped Bat (English)	<i>Mormopterus jugularis</i> (Peters, 1865)
Peter's Zwergepaulettenflughund (German)	<i>Epomophorus pusillus</i> Peters, 1868
Petersons Bulldoggfledermaus (German)	<i>Mops (Xiphonycteris) petersoni</i> (El Rayah, 1981)
Peterson's Free-tailed Bat (English)	<i>Mops (Xiphonycteris) petersoni</i> (El Rayah, 1981)
Petersons Langflügel-fledermaus (German)	<i>Miniopterus petersoni</i> Goodman, Bradman, Maminirina, Ryan, Christidis & Appleton, 2008
Peterson's Long-fingered Bat (English)	<i>Miniopterus petersoni</i> Goodman, Bradman, Maminirina, Ryan, Christidis & Appleton, 2008
Peterson's Mops Bat (English)	<i>Mops (Xiphonycteris) petersoni</i> (El Rayah, 1981)
Petersovi morousi (Czech)	<i>Mormopterus</i> Peters, 1865
Peters's Dwarf Epauletted Fruit Bat (English)	<i>Epomophorus pusillus</i> Peters, 1868
Peters's Epauletted Fruit Bat (English)	<i>Epomophorus crypturus</i> Peters, 1852
Peters's Flat-headed Bat (English)	<i>Platymops setiger</i> (Peters, 1878)
Peters's flat-headed Bats (English)	<i>Platymops</i> Thomas, 1906
Peters's Goblin bat (English)	<i>Mormopterus jugularis</i> (Peters, 1865)
Peters's Horseshoe Bat (English)	<i>Rhinolophus blasii</i> Peters, 1867
Peters's Sheath-tailed Bat (English)	<i>Paremballonura atrata</i> (Peters, 1874)
Peters's sheath-tailed bats (English)	<i>Coleura</i> Peters, 1867
Peters's Wrinkle-lipped Bat (English)	<i>Mormopterus jugularis</i> (Peters, 1865)
Peters-witkolvrugtevlermuis (Afrikaans)	<i>Epomophorus crypturus</i> Peters, 1852
Petit Epomophore (French)	<i>Epomophorus minimus</i> Claessen & De Vree, 1991
Petit épomophore (French)	<i>Epomophorus labiatus</i> (Temminck, 1837)
Petit Epomophore frugivore d'Angola (French)	<i>Epomophorus grandis</i> (Sanborn, 1950)
Petit Fer-à-Cheval (French)	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Petit microptère (French)	<i>Epomophorus pusillus</i> Peters, 1868
Petit minioptère (French)	<i>Miniopterus fraterculus</i> Thomas and Schwann, 1906
Petit minioptère africain (French)	<i>Miniopterus minor</i> Peters, 1867
Petit minioptère malgache (French)	<i>Miniopterus manavi</i> Thomas, 1906
Petit molosse (French)	<i>Mormopterus francoismoutoui</i> Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst, Bowie, 2008
Petit molosse à glandes caudales (French)	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
Petit molosse de Port Louis (French)	<i>Mormopterus acetabulosus</i> (Hermann, 1804)
Petit murin brun (French)	<i>Myotis tricolor</i> (Temminck, 1832)
Petit Myonyctère (French)	<i>Myonycteris torquata</i> (Dobson, 1878)
Petit Renard volant de Mascareignes (French)	<i>Pteropus subniger</i> (Kerr, 1792)
Petit renard volant de Mascarene (French)	<i>Pteropus subniger</i> (Kerr, 1792)
Petit rhinolophe (French)	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Petit rhinolophe fer à cheval (French)	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Petit Rhinopome (French)	<i>Rhinopoma cystops</i> Thomas, 1903
Petit scotophile africain (French)	<i>Scotophilus viridis</i> (Peters, 1852)
Petite Chauve-souris frugivore à collier (French)	<i>Myonycteris torquata</i> (Dobson, 1878)
Petite Chauve-souris jaune des maisons (French)	<i>Scotophilus borbonicus</i> (E. Geoffroy St.-Hilaire, 1803)
Petite Chauve-souris laineuse (French)	<i>Kerivoula lanosa</i> (A. Smith, 1847)
Petite Noctule (French)	<i>Nyctalus leisleri</i> (Kuhl, 1817)
Petite tadaride (French)	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
Petite tadaride de Madagascar (French)	<i>Mops (Chaerephon) leucogaster</i> (A. Grandidier, 1869)
Petite vache-volante (French)	<i>Nanonycteris veldkampii</i> (Jentink, 1888)
Petra fruit bat (English)	<i>Myonycteris angolensis petraea</i> (Bergmans, 1997)
Phantom-winged Bats (English)	<i>Mormopterus</i> Peters, 1865
Phyllorhine à petites oreilles (French)	<i>Cloeotis percivali</i> Thomas, 1901
Phyllorhine d'Aba (French)	<i>Hipposideros abae</i> J.A. Allen, 1917
Phyllorhine de Benito (French)	<i>Hipposideros beatus</i> (K. Andersen, 1906)
Phyllorhine de Cafrerie (French)	<i>Hipposideros caffer</i> (Sundevall, 1846)
Phyllorhine de Percival (French)	<i>Cloeotis percivali</i> Thomas, 1901
Phyllorhine de Sundevall (French)	<i>Hipposideros caffer</i> (Sundevall, 1846)
Phyllorhine fuligineux (French)	<i>Hipposideros fuliginosus</i> (Temminck, 1853)

Common Name

Phyllorhines (French)
 Phyllorine (French)
 Phyllorine à grandes oreilles (French)
 Phyllorine à grandes oreilles d'Ethiopie (French)
 Phyllorine à queue courte (French)
 Phyllorine cendrée (French)
 Phyllorine cyclope (French)
 Phyllorine d'Aellen (French)
 Phyllorine d'Afrique occidentale (French)
 Phyllorine de Commerson (French)
 Phyllorine de Commerson (French)
 Phyllorine de Jones (French)
 Phyllorine de Lamotte (French)
 Phyllorine de Noack (French)
 Phyllorine d'Eisentraut (French)
 Phyllorine des Cyclopes (French)
 Phyllorine du Cameroun (French)
 Phyllorine du Mont Nimba (French)
 Phyllorine fuligineuse (French)
 Phyllorine naine (French)
 Phyllorine rouge (French)
 Phyllorine sombre (French)
 Pied Bat (English)
 Pied Butterfly Bat (English)
 Piesangvlermuis (Afrikaans)
 Pietinis pasagnosis (Lithuanian)
 Pīmejsikspārnis (Latvian)
 Pikkjalg-lendlane (Estonian)
 Pipistrel da fossa a venter niv (Rhaeto-Romance)
 Pipistrel da Hanak (Rhaeto-Romance)
 Pipistrel da Kuhl (Rhaeto-Romance)
 Pipistrel da las alps (Rhaeto-Romance)
 Pipistrel da Madeira (Rhaeto-Romance)
 Pipistrel da Savi (Rhaeto-Romance)
 Pipistrel i Hanak-ut (Albanian)
 Pipistrel i Kuhl-it (Albanian)
 Pipistrel i Savi-it (Albanian)
 Pipistrel i zakonshëm (Albanian)
 Pipistrel muschin (Rhaeto-Romance)
 Pipistrel nanin (Rhaeto-Romance)
 Pipistrel pigmeic (Rhaeto-Romance)
 Pipistrel xhuxh (Albanian)
 Pipistrel-la comuna (Catalan (Spain))
 Pipistrel-la de Hanak (Catalan (Spain))
 Pipistrel-la de Madeira (Catalan (Spain))
 Pipistrel-la de vores clares (Catalan (Spain))
 Pipistrel-la nana (Catalan (Spain))
 Pipistrellflagermus (Danish)
 Pipistrelli i Madeiras (Albanian)
 Pipistrelli i varrezave barkzhveshur (Albanian)
 Pipistrell (Maltese)
 Pipistrell (Swedish)
 Pipistrell Żgħir (Maltese)
 Pipistrell gorr (Breton)
 Pipistrell Komuni (Maltese)
 Pipistrell Kuhl (Breton)
 Pipistrell Libia (Breton)
 Pipistrell Madeira (Breton)
 Pipistrell ta' Hanak (Maltese)
 Pipistrell ta' Kuhl (Maltese)
 Pipistrell ta' Madejra (Maltese)
 Pipistrell ta' Savi (Maltese)
 Pipistrell voutin (Breton)
 Pipistrelle à ailes blanches (French)
 Pipistrelle à grosse tête (French)
 Pipistrelle ariel (French)
 Pipistrelle Bats (English)
 Pipistrelle commune (French)
 Pipistrelle d'Aellen (French)
 Pipistrelle d'Afrique centrale (French)
 Pipistrelle d'Afrique occidentale (French)
 Pipistrelle d'Anchieta (French)
 Pipistrelle de Grandidier (French)

Taxon Name

Hipposideros Gray, 1831
HIPPOSIDERIDAE Lydekker, 1891
Hipposideros megalotis (Heuglin, 1861)
Hipposideros megalotis (Heuglin, 1861)
Hipposideros curtus G.M. Allen, 1921
Hipposideros tephurus (Cabrera, 1906)
Doryrhina cyclops (Temminck, 1853)
Hipposideros marisae Aellen, 1954
Hipposideros jonesi Hayman, 1947
Macronycteris commersoni (E. Geoffroy St.-Hilaire, 1813)
Macronycteris gigas (Wagner, 1845)
Hipposideros jonesi Hayman, 1947
Hipposideros lamottei Brosset, 1985
Hipposideros ruber (Noack, 1893)
Doryrhina camerunensis (Eisentraut, 1956)
Doryrhina cyclops (Temminck, 1853)
Doryrhina camerunensis (Eisentraut, 1956)
Hipposideros lamottei Brosset, 1985
Hipposideros fuliginosus (Temminck, 1853)
Hipposideros beatus (K. Andersen, 1906)
Hipposideros ruber (Noack, 1893)
Hipposideros fuliginosus (Temminck, 1853)
Glauconycteris superba Hayman, 1939
Glauconycteris superba Hayman, 1939
Afronycteris nana (Peters, 1852)
Rhinolophus euryale Blasius, 1853
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Myotis capaccinii (Bonaparte, 1837)
Taphozous nudiventris Cretzschmar, 1830
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Taphozous nudiventris Cretzschmar, 1830
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pseudoromicia tenuipinnis (Peters, 1872)
Nycticeinops crassulus (Thomas, 1904)
Hypsugo ariel (Thomas, 1904)
Pipistrellus Kaup, 1829
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) inexpectatus Aellen, 1959
Pipistrellus (Pipistrellus) inexpectatus Aellen, 1959
Hypsugo musciculus (Thomas, 1913)
Neoromicia anchietae (Seabra, 1900)
Nycticeinops grandidieri (Dobson, 1876)

Common Name

Taxon Name

Pipistrelle de Guinée (French)	<i>Neoromicia guineensis</i> (Bocage, 1889)
Pipistrelle de Kuhl (French)	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Pipistrelle de Libye (French)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Pipistrelle de Madère (French)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Pipistrelle de Racey (French)	<i>Pipistrellus (Pipistrellus) raceyi</i> Bates, Ratrimomanarivo, Harrison and Goodman, 2006
Pipistrelle de Rendall (French)	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
Pipistrelle de Rueppel (French)	<i>Vansonia rueppellii</i> (J.B. Fischer, 1829)
Pipistrelle de Rüppell (French)	<i>Vansonia rueppellii</i> (J.B. Fischer, 1829)
Pipistrelle de Savi (French)	<i>Hypsugo savii</i> (Bonaparte, 1837)
Pipistrelle de Somalie (French)	<i>Neoromicia somalica</i> (Thomas, 1901)
Pipistrelle de Tanzanie (French)	<i>Pipistrellus (Pipistrellus) permixtus</i> Aellen, 1957
Pipistrelle d'Eisentraut (French)	<i>Nycticeinops eisentrauti</i> (Hill, 1968)
Pipistrelle des Melcks (French)	<i>Laephotis stanleyi</i> (Goodman, Kearney, Ratsimbazafy and Hassanin, 2017)
Pipistrelle du Kenya (French)	<i>Pipistrellus (Pipistrellus) aero</i> Heller, 1912
Pipistrelle du Miombo (French)	<i>Neoromicia anchietae</i> (Seabra, 1900)
Pipistrelle géante (French)	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Pipistrelle hespéride (French)	<i>Pipistrellus (Pipistrellus) hesperidus</i> (Temminck, 1840)
Pipistrelle jaune (French)	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
Pipistrelle minuscule (French)	<i>Pipistrellus (Pipistrellus) nanulus</i> Thomas, 1904
Pipistrelle murine (French)	<i>Hypsugo musciculus</i> (Thomas, 1913)
Pipistrelle naine (French)	<i>Afronycteris nana</i> (Peters, 1852)
Pipistrelle naine aux ailes brunes (French)	<i>Afronycteris nana</i> (Peters, 1852)
Pipistrelle pygmée (French)	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Pipistrelle rouille (French)	<i>Pipistrellus (Pipistrellus) rusticus</i> (Tomes, 1861)
Pipistrelle rustique (French)	<i>Pipistrellus (Pipistrellus) rusticus</i> (Tomes, 1861)
Pipistrelle soleil (French)	<i>Afronycteris helios</i> (Heller, 1912)
Pipistrelle soprane (French)	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Pipistrelle zouloue (French)	<i>Neoromicia zuluensis</i> (Roberts, 1924)
Pipistrellen (German)	<i>Pipistrellus</i> Kaup, 1829
Pipistrelles (English)	<i>Pipistrellus</i> Kaup, 1829
Pipistrelles (French)	<i>Pipistrellus</i> Kaup, 1829
Pipistrèlli (Italian)	<i>CHIROPTERA</i> Blumenbach, 1779
Pipistrèlli (Italian)	<i>Pipistrellus</i> Kaup, 1829
Pipistrèlli a coda di tòpo (Italian)	<i>RHINOPOMATIDAE</i> Dobson, 1872
Pipistrèlli délle tòmbe (Italian)	<i>Taphozous</i> E. Geoffroy St.-Hilaire, 1818
Pipistrello albolimbato (Italian)	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Pipistrèllo capènse (Italian)	<i>Laephotis capensis</i> (A. Smith, 1829)
Pipistrèllo carlino (Italian)	<i>Mops</i> Lesson, 1842
Pipistrello de Kühli (Italian)	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Pipistrello delle tombe a ventre nudo (Italian)	<i>Taphozous nudiventris</i> Cretzschmar, 1830
Pipistrèllo di Eisentraut (Italian)	<i>Nycticeinops eisentrauti</i> (Hill, 1968)
Pipistrello di Hanak (Italian)	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Pipistrèllo di Hèller (Italian)	<i>Afronycteris helios</i> (Heller, 1912)
Pipistrello di Madeira (Italian)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Pipistrèllo di Peters (Italian)	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
Pipistrèllo di Pèters (Italian)	<i>Afronycteris nana</i> (Peters, 1852)
Pipistrèllo di Rëndall (Italian)	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
Pipistrèllo di Rüppell (Italian)	<i>Vansonia rueppellii</i> (J.B. Fischer, 1829)
Pipistrello di Savi (Italian)	<i>Hypsugo savii</i> (Bonaparte, 1837)
Pipistrèllo giallàstro (Italian)	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
Pipistrèllo guineàno (Italian)	<i>Neoromicia guineensis</i> (Bocage, 1889)
Pipistrello nano (Italian)	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Pipistrèllo nàno déi banàni (Italian)	<i>Afronycteris nana</i> (Peters, 1852)
Pipistrello pigmeo (Italian)	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Pipistrèllo rugginóso (Italian)	<i>Pipistrellus (Pipistrellus) rusticus</i> (Tomes, 1861)
Pipistrèllo seràle (Italian)	<i>Pipistrellus (Pipistrellus) hesperidus</i> (Temminck, 1840)
Pipistrèllo sòmalo (Italian)	<i>Neoromicia somalica</i> (Thomas, 1901)
Pipistrello soprano (Italian)	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Pipistrello soprano (Italian)	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Pipistrello (Castilian (Spain))	<i>Pipistrellus (Pipistrellus) nanulus</i> Thomas, 1904
Pipistrello (Castilian (Spain))	<i>Nycticeinops crassulus</i> (Thomas, 1904)
Pipistrello arrunt (Basque)	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Pipistrello Enano (Castilian (Spain))	<i>Afronycteris nana</i> (Peters, 1852)
Pipistrello madeirar (Basque)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Pipistrello mediterraneo (Basque)	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Pisze denevér (Hungarian)	<i>Barbastella barbastellus</i> (Schreber, 1774)
Piszedenevér (Hungarian)	<i>Barbastella barbastellus</i> (Schreber, 1774)
Pita-ohiki-koumori (Japanese)	<i>Mormopterus jugularis</i> (Peters, 1865)
Pitkäsiipiyökkö (Finnish)	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Pitkäsormisiippa (Finnish)	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Plain-faced Bats (English)	<i>VESPERTILIONIDAE</i> Gray, 1821
Plakegbauchhe Griewerflatterer (Luxembourgish)	<i>Taphozous nudiventris</i> Cretzschmar, 1830

Common Name

Platicarlini (Italian)
 Platkop-losstertvlermuis (Afrikaans)
 plaví kaloni (Czech)
 Plérôte d'Anchieta (French)
 Plerote Fruit-bats (English)
 Plérôtes (French)
 Plerotes Fruit-bats (English)
 pletušánové (Czech)
 Plikapilvis maišiasparnis (Lithuanian)
 ploší morousi (Czech)
 pochvorep buldočí (Czech)
 pochvorep madagaskarský (Czech)

 pochvorep seychelský (Czech)
 pochvorepovití (Czech)
 Podkasaniec Schreibersa (Polish)
 podkováček (Czech)
 podkováčkové (Czech)
 Podkovár Blasiov (Slovak)
 Podkovár južný (Slovak)
 Podkovár malý (Slovak)
 Podkovár Mehélyho (Slovak)
 Podkovár veľký (Slovak)
 podkowáček (Czech)
 podkowcowate (Polish)
 Podkowiec średni (Polish)
 Podkowiec śródziemnomorski (Polish)
 Podkowiec Blasiusa (Polish)
 Podkowiec duży (Polish)
 Podkowiec mały (Polish)
 podvečerníkové (Czech)
 Poehle's Traenenflughund (German)
 Pohle's Fruit Bat (English)
 Pohle's Harlekin-Flughund (German)
 Pohle's Harlequin Fruit-bat (English)
 Pohle's Tear-drop Fruit-bat (English)
 Popo (Swahili)
 Popo (Swahili)
 Pouched Bats (English)
 Pouched Bats (English)
 praví hrobkovci (Czech)
 praví kaloni (Czech)
 praví morousi (Czech)
 praví netopýři (Czech)
 praví pavrápenci (Czech)
 praví ušani (Czech)
 praví vlnouškové (Czech)
 Prétopodidés (French)
 Primorski šišmiš (Croatian)
 Pseudo-vampire à nez en cœur (French)
 psi noční (Czech)
 Psohubec myšový (Czech)
 ptákossavci (Czech)
 Pteropi fantasma (Italian)
 Pteròpidi (Italian)
 Pteropo fantasma paglierino (Italian)
 Pteropòdidi (Italian)
 Pügme-nahkhiir (Estonian)
 Pundursikspārnis (Latvian)
 Punisches Mausohr (German)
 Punski šišmiš (Bosnian)
 Punski netopir (Slovenian)
 Punski več ernjak (Montenegrin)
 Pustinski škorpiojed (Montenegrin)
 pustinní ušani (Czech)
 Pygmy pipistrelle bat (English)
 Pygmy Pipistrellus (English)
 pyložraví kaloni (Czech)
 Pzymroczek Saviego (Polish)
 rýhonos štětinatý (Czech)
 rýhonos drobný (Czech)
 rýhonos egyptský (Czech)
 rýhonos guinejský (Czech)
 rýhonos jihoafrický (Czech)

Taxon Name

Platymops Thomas, 1906
Sauromys petrophilus (Roberts, 1917)
Eidolon Rafinesque, 1815
Plerotes anchietae (Seabra, 1900)
Plerotes K. Andersen, 1910
Plerotes K. Andersen, 1910
Plerotes K. Andersen, 1910
Plecotus E. Geoffroy St.-Hilaire, 1818
Taphozous nudiventris Cretzschmar, 1830
Platymops Thomas, 1906
Coleura afra (Peters, 1852)
Paremballonura tiavato (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)
Coleura seychellensis Peters, 1868
 EMBALLONURIDAE Gervais, 1855
Miniopterus schreibersii (Kuhl, 1817)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus Lacépède, 1799
Rhinolophus blasii Peters, 1867
Rhinolophus euryale Blasius, 1853
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus mehelyi Matschie, 1901
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
 RHINOLOPHIDAE Gray, 1825
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Rhinolophus blasii Peters, 1867
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Eptesicus Rafinesque, 1820
Casinycteris ophiodon (Pohle, 1943)
Casinycteris ophiodon (Pohle, 1943)
Casinycteris ophiodon (Pohle, 1943)
Casinycteris ophiodon (Pohle, 1943)
Casinycteris ophiodon (Pohle, 1943)
 CHIROPTERA Blumenbach, 1779
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Saccolaimus Temminck, 1838
Taphozous E. Geoffroy St.-Hilaire, 1818
Taphozous E. Geoffroy St.-Hilaire, 1818
Pteropus Brisson, 1762
Molossinae Gervais, 1856
Vespertilioninae Gray, 1821
Hipposideros Gray, 1831
Plecotus E. Geoffroy St.-Hilaire, 1818
Kerivoula Gray, 1842
 PTEROPODIDAE Gray, 1821
Hypsugo savii (Bonaparte, 1837)
Cardioderma cor (Peters, 1872)
Rousettus Gray, 1821
Tadarida teniotis (Rafinesque, 1814)
 CHIROPTERA Blumenbach, 1779
Eidolon Rafinesque, 1815
 PTEROPODIDAE Gray, 1821
Eidolon helvum (Kerr, 1792)
 PTEROPODIDAE Gray, 1821
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myotis punicus Felten, 1977
Myotis punicus Felten, 1977
Myotis punicus Felten, 1977
Myotis punicus Felten, 1977
Otonycteris hemprichii Peters, 1859
Otonycteris Peters, 1859
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Hypsugo ariel (Thomas, 1904)
Plerotes K. Andersen, 1910
Hypsugo savii (Bonaparte, 1837)
Nycteris hispida (Schreber, 1774)
Nycteris nana (K. Andersen, 1912)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris arge Thomas, 1903
Nycteris woodi K. Andersen, 1914

Common Name

rýhonos kamerunský (Czech)
 rýhonos mosambický (Czech)
 rýhonos prostřední (Czech)
 rýhonos somálský (Czech)
 rýhonos východoafrický (Czech)
 rýhonos velký (Czech)
 rýhonos velkouchý (Czech)
 rýhonos západoafrický (Czech)
 rýhonosi (Czech)
 rýhonosovití (Czech)
 Raceys Pipistrelle (German)

 Racey's Pipistrelle Bat (English)

 Raceys Zwergfledermaus (German)

 Railer Bat (English)
 Railer Mops Bat (English)
 Rainforest House Bat (English)
 Raniak azorský (Slovak)
 Raniak malý (Slovak)
 Raniak obrovský (Slovak)
 Rat penat comú (Catalan (Spain))
 Rat penat cuallarg (Catalan (Spain))
 Rat penat de bigotis (Catalan (Spain))
 Rat penat de bosc (Catalan (Spain))
 Rat penat de cova (Catalan (Spain))
 Rat penat de Madeira (Catalan (Spain))
 Rat penat de peus grans (Catalan (Spain))
 Rat penat de vores clares (Catalan (Spain))
 Rat penat gran de ferradura (Catalan (Spain))
 Rat penat mediterrani de ferradura (Catalan (Spain))
 Rat penat mitjà de ferradura (Catalan (Spain))
 Rat penat muntanyero (Catalan (Spain))
 Rat penat noctul gran (Catalan (Spain))
 Rat penat noctul petit (Catalan (Spain))
 Rat penat orellut canari (Catalan (Spain))
 Rat penat petit de ferradura (Catalan (Spain))
 Ratapinyada de cova (Catalan (Spain))
 Ratapinyada d'orella escapçada (Catalan (Spain))
 Ratapinyada gran de ferradura (Catalan (Spain))
 Ratapinyada mediterrània de ferradura (Catalan (Spain))
 Ratapinyada muntanyenca (Catalan (Spain))
 Ratapinyada petita de ferradura (Catalan (Spain))
 Ratpenat cuallarg (Catalan (Spain))
 Ratpenat de bigotis (Catalan (Spain))
 Ratpenat de bosc (Catalan (Spain))
 Ratpenat de cova (Catalan (Spain))
 Ratpenat de ferradura de Blasius (Catalan (Spain))
 Ratpenat de ferradura gran (Catalan (Spain))
 Ratpenat de ferradura mediterrani (Catalan (Spain))
 Ratpenat de ferradura mitjà (Catalan (Spain))
 Ratpenat de ferradura petit (Catalan (Spain))
 Ratpenat de peus grans (Catalan (Spain))
 Ratpenat de tomba de carpó nu (Catalan (Spain))
 Ratpenat dels graners de Botta (Catalan (Spain))
 Ratpenat dels graners meridional (Catalan (Spain))
 Ratpenat d'orelles dentades (Catalan (Spain))
 Ratpenat egipci (Catalan (Spain))
 Ratpenat muntanyenc (Catalan (Spain))
 Ratpenat rater africà (Catalan (Spain))
 Rat-tailed Bat (English)
 Rauhaarschlitznase (German)
 Red Flying Fox (English)
 Red Leaf-nosed Bat (English)
 Red Madagascan Trident Bat (English)
 Red Trident Bat (English)
 Relict Collared Fruit-bat (English)
 Reliktärer Halskrausen-Flughund (German)
 Renard volant d'Aldabra (French)
 Renard volant de l'île de Mascarene (French)
 Renard Volant de l'île de Rodriguez (French)
 Renard volant de Madagascar (French)
 Renard volant de Maurice (French)

Taxon Name

Nycteris major (K. Andersen, 1912)
Nycteris vinsoni Dalquest, 1965
Nycteris intermedia Aellen, 1959
Nycteris parisi (de Beaux, 1924)
Nycteris aurita (K. Andersen, 1912)
Nycteris grandis Peters, 1865
Nycteris macrotis Dobson, 1876
Nycteris gambiensis (K. Andersen, 1912)
Nycteris G. Cuvier and E. Geoffroy, 1795
 NYCTERIDAE Van der Hoeven, 1855
Pipistrellus (Pipistrellus) raceyi Bates, Ratrimomanarivo, Harrison and Goodman, 2006
Pipistrellus (Pipistrellus) raceyi Bates, Ratrimomanarivo, Harrison and Goodman, 2006
Pipistrellus (Pipistrellus) raceyi Bates, Ratrimomanarivo, Harrison and Goodman, 2006
Mops (Xiphonycteris) thersites (Thomas, 1903)
Mops (Xiphonycteris) thersites (Thomas, 1903)
Scotophilus nucella Robbins, 1983
Nyctalus azoreum (Thomas, 1901)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus lasiopterus (Schreber, 1780)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Tadarida teniotis (Rafinesque, 1814)
Myotis mystacinus (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Myotis capaccinii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Hypsugo savii (Bonaparte, 1837)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Plecotus teneriffae Barrett-Hamilton, 1907
Rhinolophus hipposideros (Bechstein, 1800)
Miniopterus schreibersii (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
Hypsugo savii (Bonaparte, 1837)
Rhinolophus hipposideros (Bechstein, 1800)
Tadarida teniotis (Rafinesque, 1814)
Myotis mystacinus (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)
Rhinolophus blasii Peters, 1867
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Rhinolophus hipposideros (Bechstein, 1800)
Myotis capaccinii (Bonaparte, 1837)
Taphozous nudiventris Cretzschmar, 1830
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Hypsugo savii (Bonaparte, 1837)
Myotis punicus Felten, 1977
Rhinopoma microphyllum (Brünnich, 1782)
Nycteris hispida (Schreber, 1774)
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Hipposideros ruber (Noack, 1893)
Triaenops menamena Goodman and Ranivo, 2009
Triaenops menamena Goodman and Ranivo, 2009
Myonycteris relicta Bergmans, 1980
Myonycteris relicta Bergmans, 1980
Pteropus aldabrensis True, 1893
Pteropus niger (Kerr, 1792)
Pteropus rodricensis Dobson, 1878
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Pteropus niger (Kerr, 1792)

Common Name

Taxon Name

Renard volant de Pemba (French)	<i>Pteropus voeltzkowi</i> Matschie, 1909
Renard volant de Rodrigues (French)	<i>Pteropus rodricensis</i> Dobson, 1878
Renard volant des Comores (French)	<i>Pteropus livingstonii</i> Gray, 1866
Renard volant des Mascaraïnes (French)	<i>Pteropus niger</i> (Kerr, 1792)
Renard volant des Seychelles (French)	<i>Pteropus seychellensis</i> A. Milne-Edwards, 1877
Renard volant malgache (French)	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
Renard volant noir (French)	<i>Pteropus livingstonii</i> Gray, 1866
Renard volant roux (French)	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
Renards volants (French)	<i>Pteropus</i> Brisson, 1762
Rendal-dakvlermuis (Afrikaans)	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
Rendall se dakvlermuis (Afrikaans)	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
Rendalls Pipistrelle (German)	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
Rendall's Pipistrelle (English)	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
Rendall's Pipistrelle Bat (English)	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
Rendall's Serotine (English)	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
Rendall's Serotine Bat (English)	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
Rendalls Zwergfledermaus (German)	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
Reunion Free-tailed bat (English)	<i>Mormopterus francoismoutoui</i> Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst, Bowie, 2008
Reunion Mastino-Fledermaus (German)	<i>Mormopterus francoismoutoui</i> Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst, Bowie, 2008
Réunion-Hausfledermaus (German)	<i>Scotophilus borbonicus</i> (E. Geoffroy St.-Hilaire, 1803)
Reuse losstertvlermuis (Afrikaans)	<i>Tadarida ventralis</i> (Heuglin, 1861)
Rhinolophe à selle pointue (French)	<i>Rhinolophus blasii</i> Peters, 1867
Rhinolophe Alcyon (French)	<i>Rhinolophus alcyone</i> Temminck, 1853
Rhinolophe d'Afrique du Sud (French)	<i>Rhinolophus capensis</i> Lichtenstein, 1823
Rhinolophe d'Afrique orientale (French)	<i>Rhinolophus deckenii</i> Peters, 1868
Rhinolophe de Blasius (French)	<i>Rhinolophus blasii</i> Peters, 1867
Rhinolophe de brousse (French)	<i>Rhinolophus simulator</i> K. Andersen, 1904
Rhinolophe de Caferie (French)	<i>Hipposideros caffer</i> (Sundevall, 1846)
Rhinolophe de Caferie (French)	<i>Hipposideros tephros</i> (Cabrera, 1906)
Rhinolophe de Cretzschmar (French)	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
Rhinolophe de Darling (French)	<i>Rhinolophus darlingi</i> K. Andersen, 1905
Rhinolophe de Decken (French)	<i>Rhinolophus deckenii</i> Peters, 1868
Rhinolophe de Dent (French)	<i>Rhinolophus denti</i> Thomas, 1904
Rhinolophe de Geoffroy (French)	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
Rhinolophe de Guinée (French)	<i>Rhinolophus guineensis</i> Eisentraut, 1960
Rhinolophe de Hill (French)	<i>Rhinolophus hilli</i> Aellen, 1973
Rhinolophe de Lander (French)	<i>Rhinolophus landeri</i> Martin, 1838
Rhinolophe de Lofa (French)	<i>Rhinolophus hillorum</i> Koopman, 1989
Rhinolophe de MacLaud (French)	<i>Rhinolophus macclaudi</i> Pousargues, 1898
Rhinolophe de Mehely (French)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Rhinolophe de Roumanie (French)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Rhinolophe de Rüppell (French)	<i>Rhinolophus fumigatus</i> Rüppell, 1842
Rhinolophe de Sakeji (French)	<i>Rhinolophus sakejiensis</i> Cotterill, 2002
Rhinolophe de Swinny (French)	<i>Rhinolophus swinnyi</i> Gough, 1908
Rhinolophe de Tanga (French)	<i>Rhinolophus maendeleo</i> Kock, Csorba and Howell, 2000
Rhinolophe des forêts (French)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Rhinolophe d'Halcyon (French)	<i>Rhinolophus alcyone</i> Temminck, 1853
Rhinolophe d'Hildebrandt (French)	<i>Rhinolophus hildebrandtii</i> Peters, 1878
Rhinolophe du Bushveld (French)	<i>Rhinolophus simulator</i> K. Andersen, 1904
Rhinolophe du Cap (French)	<i>Rhinolophus capensis</i> Lichtenstein, 1823
Rhinolophe du Congo (French)	<i>Rhinolophus adami</i> Aellen and Brosset, 1968
Rhinolophe du Ruwenzori (French)	<i>Rhinolophus ruwenzorii</i> J. Eric Hill, 1942
Rhinolophe du Sénégal (French)	<i>Rhinolophus guineensis</i> Eisentraut, 1960
Rhinolophe du Veld (French)	<i>Rhinolophus simulator</i> K. Andersen, 1904
Rhinolophe du Ziama (French)	<i>Rhinolophus ziama</i> Fahr, Vierhaus, Hutterer and Kock, 2002
Rhinolophe éloquent (French)	<i>Rhinolophus eloquens</i> K. Andersen, 1905
Rhinolophe euryale (French)	<i>Rhinolophus euryale</i> Blasius, 1853
Rhinolophe sylvestre (French)	<i>Rhinolophus silvestris</i> Aellen, 1959
Rhinolophe trident (French)	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
Rhinolophes (French)	<i>RHINOLOPHIDAE</i> Gray, 1825
Rhinolophes (French)	<i>Rhinolophus</i> Lacépède, 1799
Rhinolophidés (French)	<i>RHINOLOPHIDAE</i> Gray, 1825
Rhinolophos i megalis (Hungarian)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Rhinolophos i meheli (Hungarian)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Rhinolophos i mikra (Hungarian)	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Rhinopoma (Italian)	<i>Rhinopoma</i> E. Geoffroy St.-Hilaire, 1818
Rhinopoma afro-ovestasiatico (Italian)	<i>Rhinopoma cystops</i> Thomas, 1903
Rhinopoma moyen (French)	<i>Rhinopoma cystops</i> Thomas, 1903
Rhinopomatidés (French)	<i>RHINOPOMATIDAE</i> Dobson, 1872
Rhinopome de Hardwicke (French)	<i>Rhinopoma cystops</i> Thomas, 1903

Common Name

Taxon Name

Rhinopome de MacInnes (French)	<i>Rhinopoma macinnesi</i> Hayman, 1937
Rhinopome microphyllum (French)	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
Rhinopomes (French)	RHINOPOMATIDAE Dobson, 1872
Rhinopomes (French)	<i>Rhinopoma</i> E. Geoffroy St.-Hilaire, 1818
Ridi šišmiš (Croatian)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Ridi večernjak (Montenegrin)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Riesen Abendsegler (German)	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Riesenabendsegler (German)	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Riesen-Bulldoggfledermäuse (German)	<i>Otomops</i> Thomas, 1913
Riesen-Hausfledermaus (German)	<i>Scotophilus nigrita</i> (Schreber, 1774)
Riesen-Rundblattnase (German)	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Riesen-Rundblattnase (German)	<i>Macronycteris gigas</i> (Wagner, 1845)
Rinolof da Blasius (Rhaeto-Romance)	<i>Rhinolophus blasii</i> Peters, 1867
Rinolof da Méhely (Rhaeto-Romance)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Rinolof grond (Rhaeto-Romance)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Rinolof mediterranean (Rhaeto-Romance)	<i>Rhinolophus euryale</i> Blasius, 1853
Rinolof pitschen (Rhaeto-Romance)	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Rinòlofi (Italian)	<i>Rhinolophus</i> Lacépède, 1799
Rinolòfidi (Italian)	RHINOLOPHIDAE Gray, 1825
Rinòlofo affumicàto (Italian)	<i>Rhinolophus fumigatus</i> Rüppell, 1842
Rinolof de Blasius (Castilian (Spain))	<i>Rhinolophus blasii</i> Peters, 1867
Rinolof di Blàsius (Italian)	<i>Rhinolophus blasii</i> Peters, 1867
Rinòlofo di Cretzschmar (Italian)	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
Rinòlofo di Decken (Italian)	<i>Rhinolophus deckenii</i> Peters, 1868
Rinòlofo di Hildebrandt (Italian)	<i>Rhinolophus hildebrandtii</i> Peters, 1878
Rinòlofo di Länder (Italian)	<i>Rhinolophus landeri</i> Martin, 1838
Rinolof di Méhely (Italian)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Rinolof egipcio (Castilian (Spain))	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
Rinòlofo eloquente (Italian)	<i>Rhinolophus eloquens</i> K. Andersen, 1905
Rinolof Euriale (Italian)	<i>Rhinolophus euryale</i> Blasius, 1853
Rinolof grande de herradura (Castilian (Spain))	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Rinolof maggiore (Italian)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Rinolof mediano de herradura (Castilian (Spain))	<i>Rhinolophus mehelyi</i> Matschie, 1901
Rinolof mediterraneo de herradura (Castilian (Spain))	<i>Rhinolophus euryale</i> Blasius, 1853
Rinolof mediterraneo (Italian)	<i>Rhinolophus euryale</i> Blasius, 1853
Rinolof minore (Italian)	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Rinolof pequeño de herradura (Castilian (Spain))	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Rinòlofo simulatòre (Italian)	<i>Rhinolophus simulator</i> K. Andersen, 1904
Rinolofu Żgħir (Maltese)	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Rinolofu Kbir (Maltese)	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Rinolofu ta' Blasius (Maltese)	<i>Rhinolophus blasii</i> Peters, 1867
Rinolofu ta' Mehely (Maltese)	<i>Rhinolophus mehelyi</i> Matschie, 1901
Rinolofu tal-Mediterran (Maltese)	<i>Rhinolophus euryale</i> Blasius, 1853
Rinolophos i mesogiaki (Hungarian)	<i>Rhinolophus euryale</i> Blasius, 1853
Rinopoma di MacInnes (Italian)	<i>Rhinopoma macinnesi</i> Hayman, 1937
Rinopoma microfillo (Italian)	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
Rinopormatidi (Italian)	RHINOPOMATIDAE Dobson, 1872
Ripslendlane (Estonian)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Riseflaggermus (Norwegian)	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Riseg Bëschfliedermaus (Luxembourgish)	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Robbins Hausfledermaus (German)	<i>Scotophilus nucella</i> Robbins, 1983
Robbin's House Bat (English)	<i>Scotophilus nucella</i> Robbins, 1983
Robbins's House Bat (English)	<i>Scotophilus nucella</i> Robbins, 1983
Robbins's Yellow Bat (English)	<i>Scotophilus nucella</i> Robbins, 1983
Roberts' Flachkopf-Bulldoggfledermaus (German)	<i>Sauromys petrophilus</i> (Roberts, 1917)
Roberts' Flat-headed Bat (English)	<i>Sauromys petrophilus</i> (Roberts, 1917)
Roberts Zwergfledermaus (German)	<i>Laephotis robertsi</i> (Goodman, Taylor, Ratrimomanarivo and Hooper, 2012)
Roberts's Flat-headed Bat (English)	<i>Sauromys petrophilus</i> (Roberts, 1917)
Robust Yellow Bat (English)	<i>Scotophilus robustus</i> A. Milne-Edwards, 1881
Rock-dwelling Flat-headed Bat (English)	<i>Sauromys petrophilus</i> (Roberts, 1917)
Rock-loving Flat-headed Bat (English)	<i>Sauromys petrophilus</i> (Roberts, 1917)
Rodrigues Flying Fox (English)	<i>Pteropus rodricensis</i> Dobson, 1878
Rodrigues-Flugfuchs (German)	<i>Pteropus rodricensis</i> Dobson, 1878
Rodriguez Flying Fox (English)	<i>Pteropus rodricensis</i> Dobson, 1878
Rodriguez vliegende hond (Dutch)	<i>Pteropus rodricensis</i> Dobson, 1878
Rodriguez-Flughund (German)	<i>Pteropus rodricensis</i> Dobson, 1878
Roeskleurvermuis (Afrikaans)	<i>Pipistrellus (Pipistrellus) rusticus</i> (Tomes, 1861)
Ronkko (Finnish)	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Rooi langhaarvermuis (Afrikaans)	<i>Myotis bocagii</i> (Peters, 1870)
Rosevears Zwergfledermaus (German)	<i>Pseudoromicia roseveari</i> (Monadjem, Richards, Taylor and Stoffberg, 2013)
Rossetta (Italian)	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)

Common Name

Rossétti (Italian)
 Rossétto del Nilo (Italian)
 Rossétto egiziano (Italian)
 Rostfarbene Bulldoggfledermaus (German)
 Rostfarbene Pipistrelle (German)
 Rostfarbene Zwergfledermaus (German)
 rothbraune Doggenrämmler (German)
 Rothen Kamnase (German)
 Rötliche Dreizahn-Blattnase (German)
 Rougette (French)
 Roundleaf Bats (English)
 Roussette Fruit-bats (English)
 Roussettes (English)
 Roussette à collet rouge (French)
 Roussette à épaulettes d'Angola (French)
 Roussette à épaulettes de Peters (French)
 Roussette à épaulettes de Wahlberg (French)
 Roussette à long poils du Ruwenzori (French)
 Roussette à longs poils (French)
 Roussette à longs poils (French)
 Roussette à petit palais (French)
 Roussette d'Angola (French)
 Roussette de Bocage (French)
 Roussette de Buettikofer (Ewe (Ghana))
 Roussette de Büttikofer (Ewe (Ghana))
 Roussette de Dobson (French)
 Roussette de Franquet (Ewe (Ghana))
 Roussette de Madagascar (French)
 Roussette de Pohle (French)
 Roussette de Zenker (French)
 Roussette d'Égypte (French)
 Roussette des Comores (French)
 Roussette des palmiers africaine (Ewe (Ghana))
 Roussette des palmiers malgache (French)
 Roussette des Seychelles (French)
 Roussette du Kenya (French)
 Roussette du Ruwenzori (French)
 Roussette égyptienne (French)
 Roussette foncée (French)
 Roussette jaune de Madagascar (French)
 Roussette labiale (French)
 Roussette labiée (French)
 Roussette malgache couleur paille (French)
 Roussette naine de Hayman (French)
 Roussette naine de Peters (French)
 Roussette naine de Veldkamp (French)
 Roussette noire (French)
 Roussette paillée (Ewe (Ghana))
 Roussette rougeâtre (French)
 Roussettes (French)
 Roussettes à collier (French)
 Roussettes des palmiers (French)
 Roussettes géantes (French)
 Roussettes naines (French)
 Rudawka nilowa (Polish)
 Rufous Hairy Bat (English)
 Rufous Mouse-eared bat (English)
 Rufous Mouse-eared Myotis (English)
 Rufous Trident Bat (English)
 Rundbladneser (Norwegian)
 Rundblattnasen (German)
 Rundlappigen Kammnase (German)
 Rüppell se saalneusvlermuis (Afrikaans)
 Rüppell se vlermuis (Afrikaans)
 Rüppell's Bat (English)
 Rüppells dwergvlermuis (Dutch)
 Rüppell's Horseshoe Bat (English)
 Rüppells Hufeisennase (German)
 Rüppells Hufeisennase (German)
 Rüppell's Pipistrelle (English)
 Rüppells Pipistrelle (German)
 Rüppells Zwergfledermaus (German)
 Rüppell-saalneusvlermuis (Afrikaans)
 Rüppell-vlermuis (Afrikaans)

Taxon Name

Rousettus Gray, 1821
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Mops (Chaerephon) russatus (J.A. Allen, 1917)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Tadarida ventralis (Heuglin, 1861)
Rhinolophus alcyone Temminck, 1853
Triaenops menamena Goodman and Ranivo, 2009
Pteropus subniger (Kerr, 1792)
Hipposideros Gray, 1831
Rousettus Gray, 1821
Rousettus Gray, 1821
Pteropus subniger (Kerr, 1792)
Epomophorus angolensis Gray, 1870
Epomophorus crypturus Peters, 1852
Epomophorus wahlbergi (Sundevall, 1846)
Myonycteris angolensis ruwenzorii (Eisentraut, 1965)
Myonycteris angolensis (Bocage, 1898)
Stenonycteris lanosus (Thomas, 1906)
Casinycteris argynnis Thomas, 1910
Myonycteris angolensis (Bocage, 1898)
Myonycteris angolensis (Bocage, 1898)
Epomops buettikoferi (Matschie, 1899)
Epomops buettikoferi (Matschie, 1899)
Epomophorus dobsonii Bocage, 1889
Epomops franqueti (Tomes, 1860)
Rousettus madagascariensis G. Grandidier, 1929
Casinycteris ophiodon (Pohle, 1943)
Scotonycteris zenkeri Matschie, 1894
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus obliviosus Kock, 1978
Eidolon helvum (Kerr, 1792)
Eidolon dupreanum (Schegel, 1867)
Pteropus seychellensis A. Milne-Edwards, 1877
Myonycteris relicta Bergmans, 1980
Stenonycteris lanosus (Thomas, 1906)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pteropus subniger (Kerr, 1792)
Eidolon dupreanum (Schegel, 1867)
Epomophorus labiatus (Temminck, 1837)
Epomophorus labiatus (Temminck, 1837)
Eidolon dupreanum (Schegel, 1867)
Epomophorus intermedius (Hayman, 1963)
Epomophorus pusillus Peters, 1868
Nanonycteris veldkampii (Jentink, 1888)
Pteropus niger (Kerr, 1792)
Eidolon helvum (Kerr, 1792)
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Rousettus Gray, 1821
Myonycteris Matschie, 1899
Eidolon Rafinesque, 1815
Pteropus Brisson, 1762
Nanonycteris Matschie, 1899
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Myotis bocagii (Peters, 1870)
Myotis bocagii (Peters, 1870)
Myotis bocagii (Peters, 1870)
Triaenops menamena Goodman and Ranivo, 2009
HIPPOSIDERIDAE Lydekker, 1891
HIPPOSIDERIDAE Lydekker, 1891
Rhinolophus landeri Martin, 1838
Rhinolophus fumigatus Rüppell, 1842
Vansonia rueppellii (J.B. Fischer, 1829)
Vansonia rueppellii (J.B. Fischer, 1829)
Vansonia rueppellii (J.B. Fischer, 1829)
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus fumigatus Rüppell, 1842
Vansonia rueppellii (J.B. Fischer, 1829)
Vansonia rueppellii (J.B. Fischer, 1829)
Vansonia rueppellii (J.B. Fischer, 1829)
Rhinolophus fumigatus Rüppell, 1842
Vansonia rueppellii (J.B. Fischer, 1829)

Common Name

Rüppel's Pipistrelle (English)
 Rüppel's Pipistrelle Bat (English)
 Ruseta de Angola (Castilian (Spain))
 Ruseta Egipicia (Castilian (Spain))
 Ruskosiippa (Finnish)
 Rusoisolepakko (Finnish)
 Russbraunen Kamnase (German)
 Russet Free-tailed Bat (English)
 Russet Wrinkle-lipped bat (English)
 Rußfarbene Rundblattnase (German)
 Rusty Bat (English)
 Rusty Pipistrelle (English)
 Rusty Pipistrelle Bat (English)
 Ruwenzori Horseshoe bat (English)
 Ruwenzori Long-haired rousette (English)
 Ruwenzori Long-haired Rousette (English)
 Ruwenzori-Hufeisennase (German)
 Ruwenzori-Langhaarflughund (German)
 Sāvi sikspārnis (Latvian)
 Səhra gönlüçəsi (Azerbaijani)
 Saccolaime (French)
 Saccolaime de Pel (French)
 Sackfledermaus (German)
 Sackflügel-Fledermäuse (German)
 Sac-winged Bats (English)
 Saguzar (Basque)
 Saguzar arratoi-belarri mairutar (Basque)
 Saguzar biboteduna (Basque)
 Saguzar buztanluze europar (Basque)
 Saguzar buztanluzea (Basque)
 Saguzar hankahandia (Basque)
 Saguzar hatz-luze (Basque)
 Sakeji Horseshoe bat (English)
 Sakeji-Hufeisennase (German)
 Samburu Pipistrelle Bat (English)
 Samburu-Pipistrelle (German)
 Samburu-Zwergfledermaus (German)
 Samtfell-Flughunde (German)
 Sanborn's Epauletted Fruit-bat (English)
 Sanborns Epaulettenflughund (German)
 Sandy long-fingered bat (English)
 Sao Thome Langflügel-Fledermaus (German)
 Sao Thome Long-fingered bat (English)
 Sao Tomé Collared Fruit Bat (English)
 São Tomé Free-tailed Bat (English)
 São Tomé-Bulldoggfledermaus (German)
 saukušan (Czech)
 saukušanové (Czech)
 Sauromys (French)
 Sauromys à jugulaire (French)
 Sauromys à tête plate (French)
 Sauromys de Peters (French)
 Sauromys de Roberts (French)
 Sauromys du Natal (French)
 Savi şəbpərə si (Azerbaijani)
 Savi saguzar (Basque)
 Savi saguzarra (Basque)
 Savijev šišmiš (Bosnian)
 Savijev netopir (Slovenian)
 Savijev slijepi mišić (Montenegrin)
 Savi'nin Cüce Yarasası (Turkish)
 Savio šikšniukas (Lithuanian)
 Savir Chamchika (Bengali)
 Savi's dwergvleermuis (Dutch)
 Savis flagermus (Danish)
 Savi's Pipistrelle (English)
 Savi's Pipistrelle Bat (English)
 Schedestaartvleermuizen (Dutch)
 Schiebeschwanz-Flledermäuse (German)
 Schlangenzähniger Harlekin-Flughund (German)
 Schlieffen se vlermuis (Afrikaans)
 Schlieffenovi netopýři (Czech)
 Schlieffens Abendfledermaus (German)
 Schlieffen's Bat (English)

Taxon Name

Vansonia rueppellii (J.B. Fischer, 1829)
Vansonia rueppellii (J.B. Fischer, 1829)
Myonycteris angolensis (Bocage, 1898)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus capensis Lichtenstein, 1823
Mops (Chaerephon) russatus (J.A. Allen, 1917)
Mops (Chaerephon) russatus (J.A. Allen, 1917)
Hipposideros fuliginosus (Temminck, 1853)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Rhinolophus ruwenzorii J. Eric Hill, 1942
Myonycteris angolensis ruwenzorii (Eisentraut, 1965)
Stenonycteris lanosus (Thomas, 1906)
Rhinolophus ruwenzorii J. Eric Hill, 1942
Myonycteris angolensis ruwenzorii (Eisentraut, 1965)
Hypsugo savii (Bonaparte, 1837)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Saccolaimus Temminck, 1838
Saccolaimus peli (Temminck, 1853)
Miniopterus Bonaparte, 1837
EMBALLONURIDAE Gervais, 1855
EMBALLONURIDAE Gervais, 1855
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myotis punicus Felten, 1977
Myotis mystacinus (Kuhl, 1817)
Tadarida teniotis (Rafinesque, 1814)
Tadarida teniotis (Rafinesque, 1814)
Myotis capaccinii (Bonaparte, 1837)
Myotis capaccinii (Bonaparte, 1837)
Rhinolophus sakejiensis Cotterill, 2002
Rhinolophus sakejiensis Cotterill, 2002
Afronycteris helios (Heller, 1912)
Afronycteris helios (Heller, 1912)
Afronycteris helios (Heller, 1912)
Myonycteris Matschie, 1899
Epomophorus grandis (Sanborn, 1950)
Epomophorus grandis (Sanborn, 1950)
Miniopterus arenarius Heller, 1912
Miniopterus newtoni Bocage, 1889
Miniopterus newtoni Bocage, 1889
Myonycteris brachycephala (Bocage, 1889)
Mops (Chaerephon) tomensis (Juste and Ibáñez, 1993)
Mops (Chaerephon) tomensis (Juste and Ibáñez, 1993)
Barbastella barbastellus (Schreber, 1774)
Barbastella Gray, 1821
Sauromys Roberts, 1917
Mormopterus jugularis (Peters, 1865)
Platymops setiger (Peters, 1878)
Mormopterus jugularis (Peters, 1865)
Sauromys petrophilus (Roberts, 1917)
Mormopterus acetabulosus (Hermann, 1804)
Hypsugo savii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
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Hypsugo savii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
EMBALLONURIDAE Gervais, 1855
Coleura Peters, 1867
Casinycteris ophiodon (Pohle, 1943)
Nycticeinops schlieffenii (Peters, 1859)
Nycticeinops Hill & Harrison, 1987
Nycticeinops schlieffenii (Peters, 1859)
Nycticeinops schlieffenii (Peters, 1859)

Common Name

Schlieffen's Twilight Bat (English)
 Schlieffens vleermuis (Dutch)
 Schlieffen-vleermuis (Afrikaans)
 Schliemanns Haftscheibenfledermaus (German)
 Schliemann's Sucker-footed Bat (English)
 Schlitznasen (German)
 Schlitznasen (German)
 Schlitznasenfledermäuse (German)
 Schlitznasen-Fledermäuse (German)
 Schlitznasen-Fledermäuse (German)
 Schmalflügel-Fledermäuse (German)
 Schmetterlingsfledermaus (German)
 Schnauzbärtige Fledermaus (German)
 Schoudervlekkalong (Dutch)
 Schreber's Brown Bat (English)
 Schreber's Yellow Bat (English)
 Schreiber saguzarra (Basque)
 Schreiber-grotvleermuis (Afrikaans)
 Schreiber's Bat (English)
 Schreiber's Bent-winged Bat (English)
 Schreibers Fledermaus (German)
 Schreibers' langfingerflagermus (Danish)
 Schreibers Langflügelfledermaus (German)
 Schreibers saguzar (Basque)
 Schreibers se grotvleermuis (Afrikaans)
 Schreibers' vleermuis (Dutch)
 Schreibersge dik-angeli wawula (Sinhalese)
 Schreibers'langwjukflearmūs (Frisian)
 Schreibers's Long-fingered Bat (English)
 Schwanzdrüsen-Bulldoggfledermaus (German)
 Schwarzflügel-Hausfledermaus (German)
 Schwarz-Weiße Schmetterlingsfledermaus (German)
 Scotonyctère de Zenker (French)
 Scotoèchi (Italian)
 Scotoèco di Hinde (Italian)
 Scotoèco golabiànca (Italian)
 Scotoèco røndine (Italian)
 Scotoèque à ailes pâles (French)
 Scotoèque à gorge blanche (French)
 Scotoèque de Hinde (French)
 Scotoèque hirundine (French)
 Scotoèques (French)
 Scotòfili (Italian)
 Scotòfili di Dingan (Italian)
 Scotòfili panciabiànca (Italian)
 Scotòfili scùro (Italian)
 Scotòfili vérdi (Italian)
 Scotonyctère à dents de serpent (French)
 Scotonyctère de Bergmans (French)
 Scotonyctère de Hayman (French)
 Scotonyctères (French)
 Scotophile à ailes noires (French)
 Scotophile à ventre blanc (French)
 Scotophile à ventre jaune (French)
 Scotophile africain (French)
 Scotophile d'Andadoany (French)
 Scotophile de Bourbon (French)
 Scotophile de Madagascar (French)
 Scotophile de Madagascar (French)
 Scotophile de Marovaza (French)
 Scotophile de Robbins (French)
 Scotophile des hauteurs (French)
 Scotophile du crépuscule (French)
 Scotophile noisette (French)
 Scotophile nux (French)
 Scotophile robuste (French)
 Scotophile verdâtre (French)
 Scotophiles (French)

Taxon Name

Nycticeinops schlieffenii (Peters, 1859)
Nycticeinops schlieffenii (Peters, 1859)
Nycticeinops schlieffenii (Peters, 1859)
Myzopoda schliemanni Goodman, Rakotondraparany and Kofoky, 2007
Myzopoda schliemanni Goodman, Rakotondraparany and Kofoky, 2007
NYCTERIDAE Van der Hoeven, 1855
Nycteris G. Cuvier and E. Geoffroy, 1795
NYCTERIDAE Van der Hoeven, 1855
NYCTERIDAE Van der Hoeven, 1855
Nycteris G. Cuvier and E. Geoffroy, 1795
Mimetillus Thomas, 1904
Glauconycteris Dobson, 1875
Myotis mystacinus (Kuhl, 1817)
Pteropus livingstonii Gray, 1866
Scotophilus nigrita (Schreber, 1774)
Scotophilus nigrita (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Mops (Chaerephon) bemmeleni (Jentink, 1879)
Scotoecus hirundo (de Winton, 1899)
Glauconycteris superba Hayman, 1939
Scotonycteris zenkeri Matschie, 1894
Scotoecus Thomas, 1901
Scotoecus hindei Thomas, 1901
Scotoecus albigula Thomas, 1909
Scotoecus hirundo (de Winton, 1899)
Scotoecus albofuscus (Thomas, 1890)
Scotoecus albigula Thomas, 1909
Scotoecus hindei Thomas, 1901
Scotoecus hirundo (de Winton, 1899)
Scotoecus Thomas, 1901
Scotophilus Leach, 1821
Scotophilus dinganii (A. Smith, 1833)
Scotophilus leucogaster (Cretzschmar, 1826)
Scotophilus nigrita (Schreber, 1774)
Scotophilus viridis (Peters, 1852)
Casinycteris ophiodon (Pohle, 1943)
Scotonycteris bergmansi Hassanin, Khoudier, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015
Scotonycteris occidentalis Hayman, 1947
Scotonycteris Matschie, 1894
Scotoecus hirundo (de Winton, 1899)
Scotophilus leucogaster (Cretzschmar, 1826)
Scotophilus dinganii (A. Smith, 1833)
Scotophilus dinganii (A. Smith, 1833)
Scotophilus tandrefana Goodman, Jenkins and Ratrimomanarivo, 2005
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Scotophilus robustus A. Milne-Edwards, 1881
Scotophilus marovaza Goodman, Ratrimomanarivo and Randrianandrianina, 2006
Scotophilus nucella Robbins, 1983
Scotophilus nucella Robbins, 1983
Scotoecus albofuscus (Thomas, 1890)
Scotophilus nux Thomas, 1904
Scotophilus nux Thomas, 1904
Scotophilus robustus A. Milne-Edwards, 1881
Scotophilus viridis (Peters, 1852)
Scotophilus Leach, 1821

Common Name

Scotts Mausohr (German)
 Scott's Mouse-eared Bat (English)
 Scott's Mouse-eared Myotis (English)
 Seabra's Wing-gland Bat (English)
 Sena-Doggengrämler (German)
 Senegal Bat (English)
 Senegal Horseshoe Bat (English)
 Senegal Mastiff Bat (English)
 Sennaarische Doggengrämler (German)
 Serotin Botta (Breton)
 Serotin da Botta (Rhaeto-Romance)
 Serotin isabellin (Rhaeto-Romance)
 Serotin melenwenn (Breton)
 Serotin ta' Botta (Maltese)
 Serotin tas-Sider Pallidu (Maltese)
 Sérotine à ailes blanches (French)
 Sérotine à grands yeux (French)
 Serotine Bats (English)
 Sérotine brune d'Afrique (French)
 Sérotine brun-noire (French)
 Sérotine de Botta (French)
 Sérotine de Flower (French)
 Sérotine de Grandidier (French)
 Sérotine de Guinée (French)
 Sérotine de Lagos (French)
 Sérotine de Madagascar (French)
 Serotine de Rendall (French)
 Sérotine de Sakahara (French)
 Sérotine de Somalie (French)
 Sérotine des aloes (French)
 Sérotine des maisons à longue queue (French)
 Sérotine du Cap (French)
 Sérotine du Soudan (French)
 Serotinë e Botta's (Albanian)
 Sérotine hottentote (French)
 Sérotine hottentote (French)
 Serotinë Isabelline (Albanian)
 Sérotine isabelle (French)
 Sérotine isabelline (French)
 Sérotine jaune (French)
 Sérotine méditerranéen (French)
 Sérotine minuscule (French)
 Serotines (English)
 Sérotines (French)
 Serotino di Botta (Italian)
 Serotino isabellino (Italian)
 Seychellen Bulldoggfledermaus (German)
 Seychellen Schiebeschwanz-Fledermaus (German)
 Seychellen-Flugfuchs (German)
 Seychelles Flying Fox (English)
 Seychelles Free-tailed bat (English)
 Seychelles Fruit Bat (English)
 Seychelles Sheath-tailed Bat (English)
 Sgolanotg da las Azoras (Rhaeto-Romance)
 Sgolanotg gigant (Rhaeto-Romance)
 Sgolanotg pitschen (Rhaeto-Romance)
 Sheath-tailed Bat (English)
 Sheath-tailed Bats (English)
 Sheath-tailed bats (English)
 Shoa Kamnase (German)
 Short-eared Bats (English)
 Short-eared Trident Bat (English)
 Short-eared Trident Bats (English)
 Short-palate Fruit-bats (English)
 Short-palated Fruit Bat (English)
 Short-palated Fruit-bats (English)
 Short-tailed Leaf-nosed Bat (English)
 Short-tailed Roundleaf Bat (English)
 Short-winged Mops Bat (English)
 Siçanqlaq gecə şəbpərəsi (Azerbaijani)
 Sierra Leone Free-tailed Bat (English)
 Sierra Leone Mops Bat (English)
 Sierra-Leone Halskrausen-Flughund (German)

Taxon Name

Myotis scotti Thomas, 1927
Myotis scotti Thomas, 1927
Myotis scotti Thomas, 1927
Cistugo seabrae Thomas, 1912
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Scotophilus nigrita (Schreber, 1774)
Rhinolophus guineensis Eisentraut, 1960
Myopterus daubentonii Desmarest, 1820
Mops (Mops) midas (Sundevall, 1843)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Pseudoromicia tenuipinnis (Peters, 1872)
Eptesicus (Cnephaeus) platyops (Thomas, 1901)
Eptesicus Rafinesque, 1820
Pseudoromicia brunnea (Thomas, 1880)
Pseudoromicia brunnea (Thomas, 1880)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Nycticeinops grandidieri (Dobson, 1876)
Neoromicia guineensis (Bocage, 1889)
Eptesicus (Cnephaeus) platyops (Thomas, 1901)
Laephotis matroka (Thomas and Schwann, 1905)
Pseudoromicia rendalli (Thomas, 1889)
Laephotis malagasyensis (Peterson, Eger and Mitchell, 1995)
Neoromicia somalica (Thomas, 1901)
Neoromicia zuluensis (Roberts, 1924)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Laephotis capensis (A. Smith, 1829)
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Nycticeinops grandidieri (Dobson, 1876)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Neoromicia guineensis (Bocage, 1889)
Eptesicus Rafinesque, 1820
Eptesicus Rafinesque, 1820
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Mops (Chaerephon) pusillus (Miller, 1902)
Coleura seychellensis Peters, 1868
Pteropus seychellensis A. Milne-Edwards, 1877
Pteropus seychellensis A. Milne-Edwards, 1877
Mops (Chaerephon) pusillus (Miller, 1902)
Pteropus seychellensis A. Milne-Edwards, 1877
Coleura seychellensis Peters, 1868
Nyctalus azureus (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Coleura afra (Peters, 1852)
EMBALLONURIDAE Gervais, 1855
Coleura Peters, 1867
Rhinolophus fumigatus Rüppell, 1842
Cloeotis Thomas, 1901
Cloeotis percivali Thomas, 1901
Cloeotis Thomas, 1901
Casinycteris Thomas, 1910
Casinycteris argynnis Thomas, 1910
Casinycteris Thomas, 1910
Hipposideros curtus G.M. Allen, 1921
Hipposideros curtus G.M. Allen, 1921
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Myotis punicus Felten, 1977
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Myonycteris leptodon K. Andersen, 1908

Common Name

Taxon Name

Siipat (Finnish)	VESPERTILIONIDAE Gray, 1821
Silberne Schmetterlingsfledermaus (German)	<i>Glauconycteris argentata</i> (Dobson, 1875)
Silver Woolly Bat (English)	<i>Kerivoula argentata</i> Tomes, 1861
Silvered Bat (English)	<i>Glauconycteris argentata</i> (Dobson, 1875)
Silvered Bats (English)	<i>Glauconycteris</i> Dobson, 1875
Silvered Butterfly Bat (English)	<i>Glauconycteris argentata</i> (Dobson, 1875)
Silvered Woolly Bat (English)	<i>Kerivoula argentata</i> Tomes, 1861
Silvery Butterfly Bat (English)	<i>Glauconycteris argentata</i> (Dobson, 1875)
Silvery Woolly Bat (English)	<i>Kerivoula argentata</i> Tomes, 1861
Simandou serotine (English)	<i>Pipistrellus (Pipistrellus) simandouensis</i> Monadjem, Richards, Decher and Hutterer, 2020
Simple-nosed Bats (English)	VESPERTILIONIDAE Gray, 1821
Sinai Barbestelle (English)	<i>Barbastella leucomelas</i> (Cretzschmar, 1826)
Singing Fruit Bat (English)	<i>Epomophorus dobsonii</i> Bocage, 1889
Singing Fruit Bat (English)	<i>Epomops franqueti</i> (Tomes, 1860)
Singing Fruit-bats (English)	<i>Epomops</i> Gray, 1866
Sivatagi hosszúfülű-denevér (Hungarian)	<i>Otonycteris hemprichii</i> Peters, 1859
Sjevernoafrički oštrouhi šišmiš (Croatian)	<i>Myotis punicus</i> Felten, 1977
Skægflagermus (Danish)	<i>Myotis mystacinus</i> (Kuhl, 1817)
skalní netopýři (Czech)	<i>Hypsugo</i> Kolenati, 1856
Skedestertvlermuis (Afrikaans)	<i>Coleura afra</i> (Peters, 1852)
Skjeggflaggermus (Norwegian)	<i>Myotis mystacinus</i> (Kuhl, 1817)
Skouarnegell Hemprich (Breton)	<i>Otonycteris hemprichii</i> Peters, 1859
Skouarnegell Tenerife (Breton)	<i>Plecotus teneriffae</i> Barrett-Hamilton, 1907
Skropstainais naktsšķīpānis (Latvian)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Skupina brkatič netopirjev (Slovenian)	<i>Myotis mystacinus</i> (Kuhl, 1817)
Slender-winged Pipistrelle Bat (English)	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
Slijepi mišić sa Madeire (Montenegrin)	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Slit-faced bats (English)	NYCTERIDAE Van der Hoeven, 1855
Slit-faced bats (English)	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795
Small house-bat (English)	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Small mouse-tailed bat (English)	<i>Rhinopoma cystops</i> Thomas, 1903
Smaller Epauletted fruit bat (English)	<i>Epomophorus crypturus</i> Peters, 1852
Smaller Horseshoe Bat (English)	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Smithers Hufeisennase (German)	<i>Rhinolophus smithersi</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Smithers's Horseshoe Bat (English)	<i>Rhinolophus smithersi</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Smith's fruit bat (English)	<i>Myonycteris angolensis smithii</i> (Thomas, 1908)
Smiths Samtfell-Flughund (German)	<i>Myonycteris angolensis smithii</i> (Thomas, 1908)
Smith's Wollfledermaus (German)	<i>Kerivoula smithii</i> Thomas, 1880
Smith's Woolly Bat (English)	<i>Kerivoula smithii</i> Thomas, 1880
Smoky Horseshoe Bat (English)	<i>Rhinolophus fumigatus</i> Rüppell, 1842
Smooth-haired Fruit-bats (English)	<i>Myonycteris</i> Matschie, 1899
Snake-toothed Harlequin Fruit-bat (English)	<i>Casinycteris ophiodon</i> (Pohle, 1943)
Snake-toothed Tear-drop Fruit-bat (English)	<i>Casinycteris ophiodon</i> (Pohle, 1943)
Sod silli (see Lanza <i>et al.</i> , 2015: 13) (Somalian)	CHIROPTERA Blumenbach, 1779
Sodan silli (Somalian)	CHIROPTERA Blumenbach, 1779
Soft-furred Fruit-bats (English)	<i>Myonycteris</i> Matschie, 1899
Somali Pipistrelle Bat (English)	<i>Neoromicia somalica</i> (Thomas, 1901)
Somali Serotine (English)	<i>Neoromicia somalica</i> (Thomas, 1901)
Somali Serotine Bat (English)	<i>Neoromicia somalica</i> (Thomas, 1901)
Somali Trident Leaf-nosed Bat (English)	<i>Asellia italosomalica</i> de Beaux, 1931
Somalian Pipistrelle Bat (English)	<i>Neoromicia somalica</i> (Thomas, 1901)
Somalia-Pipistrelle (German)	<i>Neoromicia somalica</i> (Thomas, 1901)
Somalia-Zwergfledermaus (German)	<i>Neoromicia somalica</i> (Thomas, 1901)
Somali-dakvlermuis (Afrikaans)	<i>Neoromicia somalica</i> (Thomas, 1901)
Somalische Dreizackblattnase (German)	<i>Asellia italosomalica</i> de Beaux, 1931
Sooty Leaf-nosed Bat (English)	<i>Hipposideros fuliginosus</i> (Temminck, 1853)
Sooty Roundleaf Bat (English)	<i>Hipposideros fuliginosus</i> (Temminck, 1853)
Soprano Pipistrelle (English)	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Sororcula Long-fingered Bat (English)	<i>Miniopterus sororculus</i> Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton, 2007
Sororculus Longfingered Bat (English)	<i>Miniopterus sororculus</i> Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton, 2007
Sousouri banann (Creole)	<i>Coleura seychellensis</i> Peters, 1868
South African Lesser Leaf-nosed Bat (English)	<i>Hipposideros caffer</i> (Sundevall, 1846)
Southern Africa horseshoe bat (English)	<i>Rhinolophus capensis</i> Lichtenstein, 1823
Southern Sheath-tailed Bat (English)	<i>Coleura afra</i> (Peters, 1852)
spaka kryšowá (Czech)	<i>Myopterus daubentonii</i> Desmarest, 1820
spaky (Czech)	<i>Myopterus</i> E. Geoffroy St.-Hilaire, 1818
Spitssaalneusvlermuis (Afrikaans)	<i>Rhinolophus blasii</i> Peters, 1867
Spleetneusvlermuizen (Dutch)	NYCTERIDAE Van der Hoeven, 1855
Spotted Free-tailed Bat (English)	<i>Mops (Chaerephon) bivittatus</i> (Heuglin, 1861)

Common Name

Spotted Gland-tailed Bat (English)
 Spotted Wrinkle-lipped Bat (English)
 Spurrell trumpet-eared bat (English)
 Spurrells Bulldoggfledermaus (German)
 Spurrell's Free-tailed Bat (English)
 Spurrell's Mops Bat (English)
 Spurrells Wollfledermaus (German)
 Spurrell's Woolly Bat (English)
 srdcoví lyronosi (Czech)
 Sredozemni potkovičar (Montenegrin)
 Sredozemni slobodnorepac (Croatian)
 sreso ampane [= "original or normal bat"] (Tanoboase (Ghana))
 Srostlouch (Czech)
 srostlouchové (Czech)
 Stanleys Serotine bat (German)

Stejnoušan zejkatý (Czech)
 Stirngrubenfledermaus (German)
 Stirnlappen-Bulldoggfledermaus (German)
 Stor brunflagermus (Danish)
 Stor hestekonæse (Danish)
 Stor hesteskone (Norwegian)
 Storøret flagermus (Danish)
 Storfotsfladdermus (Swedish)
 Storhesteskone (Norwegian)
 Storøreflaggermus (Norwegian)
 Större hästskonäsa (Swedish)
 Strėliaausis lygianosis (Lithuanian)
 Straw-colored Fruit Bat (English)
 Straw-coloured Flying Fox (English)
 Straw-coloured Fruit-bats (English)
 Streifen-Rundblattnase (German)
 striflaggermus (Norwegian)
 Striped Butterfly Bat (English)
 Striped Leaf-nosed Bat (English)
 Striped or Banded Guano Bat (English)
 strostlouchové (Czech)
 Sucker-footed Bat (English)

Sucker-footed Bats (English)
 Sucker-footed Bats (English)
 Südafrikanischen Sackfledermaus (German)
 Sudan Horn-skinned Bat (English)
 Sudan Serotine (English)
 Sugeskålflaggermus (Norwegian)
 Sundevall se bladneusvlermuis (Afrikaans)
 Sundevall-bladneusvlermuis (Afrikaans)
 Sundevall's African Leaf-nosed Bat (English)
 Sundevall's Blattnase (German)
 Sundevall's Free-tailed Bat (English)
 Sundevall's Leaf-nosed Bat (English)
 Sundevall's Roundleaf Bat (English)
 Sundevalls Rundblattnase (German)
 Sundevallse blaarneusvlermuis (Afrikaans)
 Superb Butterfly Bat (English)
 Suur-sagarnina (Estonian)
 Suur-teravkõrv (Estonian)
 Svetlotrebušni poznik (Slovenian)
 Svijetlotrebušasti slijepi miš (Montenegrin)
 Swallow Lesser House Bat (English)
 Swinny se saalneusvlermuis (Afrikaans)
 Swinny's Horseshoe Bat (English)
 Swinnys Hufeisennase (German)
 Swinny-saalneusvlermuis (Afrikaans)
 Swinnyse Vlermuis (Afrikaans)
 Sydapistrell (Swedish)
 Szöröskarú denevér (Hungarian)
 Szöröskarú koraidenevér (Hungarian)
 Szoprán törpedenevér (Hungarian)
 Tadarida (Maltese)
 tadarida africká (Czech)
 Tadarida buldogovitá (Slovak)
 Tadarida de Midas (French)

Taxon Name

Mops (Chaerephon) bivittatus (Heuglin, 1861)
Mops (Chaerephon) bivittatus (Heuglin, 1861)
Kerivoula phalaena Thomas, 1912
Mops (Xiphonycteris) spurrelli (Dollman, 1911)
Mops (Xiphonycteris) spurrelli (Dollman, 1911)
Mops (Xiphonycteris) spurrelli (Dollman, 1911)
Kerivoula phalaena Thomas, 1912
Kerivoula phalaena Thomas, 1912
Cardioderma Peters, 1873
Rhinolophus blasii Peters, 1867
Tadarida teniotis (Rafinesque, 1814)
Epomophorus gambianus (Ogilby, 1835)

Barbastella barbastellus (Schreber, 1774)
Plecotus E. Geoffroy St.-Hilaire, 1818
Laephotis stanleyi (Goodman, Kearney, Ratsimbazafy and Hassanin, 2017)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Mops (Chaerephon) major (Trouessart, 1897)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Otonycteris hemprichii Peters, 1859
Myotis capaccinii (Bonaparte, 1837)
Rhinolophus ferrumequinum (Schreber, 1774)
 MEGADERMATIDAE H. Allen, 1864
Rhinolophus ferrumequinum (Schreber, 1774)
Otonycteris hemprichii Peters, 1859
Eidolon helvum (Kerr, 1792)
Eidolon helvum (Kerr, 1792)
Eidolon Rafinesque, 1815
Macronycteris vittatus (Peters, 1852)
 NYCTERIDAE Van der Hoeven, 1855
Glauconycteris alboguttata J.A. Allen, 1917
Macronycteris vittatus (Peters, 1852)
Tadarida teniotis (Rafinesque, 1814)
Barbastella Gray, 1821
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
 MYZOPODIDAE Thomas, 1904
Myzopoda Milne-Edwards and A. Grandidier, 1878
Miniopterus natalensis (A. Smith, 1833)
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
 MYZOPODIDAE Thomas, 1904
Hipposideros caffer (Sundevall, 1846)
Hipposideros caffer (Sundevall, 1846)
Hipposideros caffer (Sundevall, 1846)
Hipposideros caffer (Sundevall, 1846)
Mops (Mops) midas (Sundevall, 1843)
Hipposideros caffer (Sundevall, 1846)
Hipposideros caffer (Sundevall, 1846)
Hipposideros caffer (Sundevall, 1846)
Hipposideros caffer (Sundevall, 1846)
Hipposideros caffer (Sundevall, 1846)
Glauconycteris superba Hayman, 1939
Rhinolophus ferrumequinum (Schreber, 1774)
Otonycteris hemprichii Peters, 1859
Eptesicus (Cnephæus) bottae (Peters, 1869)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Scotoecus hirundo (de Winton, 1899)
Rhinolophus swinnyi Gough, 1908
Rhinolophus swinnyi Gough, 1908
Rhinolophus swinnyi Gough, 1908
Rhinolophus swinnyi Gough, 1908
Rhinolophus swinnyi Gough, 1908
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Tadarida teniotis (Rafinesque, 1814)
Tadarida ventralis (Heuglin, 1861)
Tadarida teniotis (Rafinesque, 1814)
Mops (Mops) midas (Sundevall, 1843)

Common Name

Tadarida egiziana (Italian)
 Tadarida evropská (Czech)
 tadarida jižní (Czech)
 tadarida malá (Czech)
 Tadarida ventrale (Italian)
 Tadaride (Italian)
 Tadaride à doigts blancs (French)
 Tadaride à glande caudale (French)
 Tadaride à lèvres ridées (French)
 Tadaride à oreillettes (French)
 Tadaride à queue libre de Madagascar (French)
 Tadaride à tête plate de Peters (French)
 Tadaride à ventre blanc (French)
 Tadaride à ventre blanc (French)
 Tadaride bornée (French)
 Tadaride d'Allen (French)
 Tadaride d'Angola (French)
 Tadaride d'Ansorge (French)
 Tadaride de Cestoni (French)
 Tadaride de Chapin (French)
 Tadaride de Gallagher (French)
 Tadaride de La Réunion (French)

 Tadaride de l'Afrique orientale (French)
 Tadaride de Madagascar (French)
 Tadaride de Madagascar à ventre blanc (French)
 Tadaride de Medje (French)
 Tadaride de Mongalla (French)
 Tadaride de Peters (French)
 Tadaride de Peterson (French)
 Tadaride de Railer (French)
 Tadaride de São Tomé (French)
 Tadaride de Spurell (French)
 Tadaride de Thomas (French)
 Tadaride de Trevor (French)
 Tadaride d'Egypte (French)
 Tadaride des Seychelles (French)
 Tadaride d'Europe (French)
 Tadaride du Duc des Abruzzes (French)
 Tadaride du Kenya (French)
 Tadaride du Niangara (French)
 Tadaride du Nigeria (French)
 Tadaride géante d'Afrique (French)
 Tadaride Midas (French)
 Tadaride naine (French)
 Tadaride rouge ou noir (French)

 Tadaride rousse (French)
 Tadaride tachetée (French)
 Tadarides (French)
 Tadarides (French)
 Tadarides (French)
 Tadarides (French)
 Tadarides à tête plate (French)
 tadaridovití (Czech)
 Tadarine Bats (English)
 Tafozoi (Italian)
 Tafozoi di Hamilton (Italian)
 Tafozoi di Hildegarde (Italian)
 Tafozoi mauriziano (Italian)
 Tafozoi nudiventre (Italian)
 Tafozoi perforato (Italian)
 Tanzania Pipistrelle (English)
 Tanzania-Hufeisennase (German)

 Tanzanian Woolly Bat (English)
 Tanzania-Wollfledermaus (German)
 Taphien à croupe nue (French)
 Taphien à ventre nu (French)
 Taphien de Hamilton (French)
 Taphien de Hildegarde (French)
 Taphien de l'île de France (French)
 Taphien de l'île Maurice (French)
 Taphien de Maurice (French)

Taxon Name

Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
 Tadarida teniotis (Rafinesque, 1814)
 Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
 Mops (Chaerephon) pumilus (Cretzschmar, 1826)
 Tadarida ventralis (Heuglin, 1861)
 Tadarida Rafinesque, 1814
 Mops (Xiphonycteris) brachypterus (Peters, 1852)
 Mops (Chaerephon) bemmeleni (Jentink, 1879)
 Mormopterus acetabulosus (Hermann, 1804)
 Mops (Chaerephon) major (Trouessart, 1897)
 Mops (Mops) leucostigma (G.M. Allen, 1918)
 Platymops setiger (Peters, 1878)
 Mops (Chaerephon) leucogaster (A. Grandidier, 1869)
 Mops (Mops) niveiventer Cabrera and Ruxton, 1926
 Mops (Chaerephon) pumilus (Cretzschmar, 1826)
 Mops (Mops) trevori J.A. Allen, 1917
 Mops (Mops) condylurus (A. Smith, 1833)
 Mops (Chaerephon) ansorgei (Thomas, 1913)
 Tadarida teniotis (Rafinesque, 1814)
 Mops (Chaerephon) chapini (J.A. Allen, 1917)
 Mops (Chaerephon) gallagheri (Harrison, 1975)
 Mormopterus francoismoutoui Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst, Bowie, 2008
 Tadarida lobata (Thomas, 1891)
 Mormopterus jugularis (Peters, 1865)
 Mops (Chaerephon) leucogaster (A. Grandidier, 1869)
 Mops (Mops) conigicus J.A. Allen, 1917
 Mops (Mops) demonstrator (Thomas, 1903)
 Mops (Xiphonycteris) brachypterus (Peters, 1852)
 Mops (Xiphonycteris) petersoni (El Rayah, 1981)
 Mops (Xiphonycteris) thersites (Thomas, 1903)
 Mops (Chaerephon) tomensis (Juste and Ibáñez, 1993)
 Mops (Xiphonycteris) spurrelli (Dollman, 1911)
 Tadarida fulminans (Thomas, 1903)
 Mops (Mops) trevori J.A. Allen, 1917
 Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
 Mops (Chaerephon) pusillus (Miller, 1902)
 Tadarida teniotis (Rafinesque, 1814)
 Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
 Tadarida lobata (Thomas, 1891)
 Mops (Mops) niangarae J.A. Allen, 1917
 Mops (Chaerephon) nigeriae (Thomas, 1913)
 Tadarida ventralis (Heuglin, 1861)
 Mops (Mops) midas (Sundevall, 1843)
 Mops (Xiphonycteris) nanulus J.A. Allen, 1917
 Mops (Chaerephon) jobimena (Goodman and Cardiff, 2004)
 Mops (Chaerephon) russatus (J.A. Allen, 1917)
 Mops (Chaerephon) bivittatus (Heuglin, 1861)
 Mops Lesson, 1842
 Mops (Chaerephon) Dobson, 1874
 Mormopterus Peters, 1865
 Tadarida Rafinesque, 1814
 Platymops Thomas, 1906
 MOLOSSIDAE Gervais, 1856
 Tadarida Rafinesque, 1814
 Taphozous E. Geoffroy St.-Hilaire, 1818
 Taphozous hamiltoni Thomas, 1920
 Taphozous hildegardae Thomas, 1909
 Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
 Taphozous nudiventris Cretzschmar, 1830
 Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
 Pipistrellus (Pipistrellus) permixtus Aellen, 1957
 Rhinolophus maendeleo Kock, Csorba and Howell, 2000
 Kerivoula africana Dobson, 1878
 Kerivoula africana Dobson, 1878
 Taphozous nudiventris Cretzschmar, 1830
 Taphozous nudiventris Cretzschmar, 1830
 Taphozous hamiltoni Thomas, 1920
 Taphozous hildegardae Thomas, 1909
 Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
 Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
 Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818

Common Name

Taphien des tombeaux (French)
 Taphien perforé (French)
 Taphiens (French)
 Taphiens (French)
 Taphozous bat of Mauritius (English)
 Täpläherkko (Finnish)
 Taschenfledermäuse (German)
 Tear-drop Fruit-bats (English)
 Temminck se langhaarvlermuis (Afrikaans)
 Temminck-langhaarvlermuis (Afrikaans)
 Temmincks Epaulettenflughund (German)
 Temminck's Hairy Bat (English)
 Temmincks Hufeisennase (German)
 Temminck's Mouse-eared Bat (English)
 Temmincks Rundblattnase (German)
 Tenerifes garausis (Latvian)
 Thebaanse spleetneusvlermuis (Dutch)
 Thersites' Bulldoggfledermaus (German)
 Thomas' House Bat (English)
 Thomas' Schmalflügel-Fledermaus (German)
 Thomas se vlermuis (Afrikaans)
 Thomas's Flat-headed Bat (English)
 Thomas's Lesser House Bat (English)
 Thomas's Short-palate Fruit-bat (English)
 Thomas-vlermuis (Afrikaans)
 Three-coloured bat (English)
 Thulo Ghodnale Chamero (Nepali)
 Tiny Pipistrelle (English)
 Tiny Pipistrelle Bat (English)
 Tiny Serotine (English)
 Tomb Bat (English)
 Tomb Bat (English)
 Tomb Bats (English)
 Tomb Bats (English)
 Tomosu Biasa (Indonesian)
 Törpe denevér (Hungarian)
 Törpe denevér (Hungarian)
 Transvaal Free-tailed Bat (English)
 Transvaalse losstertvlermuis (Afrikaans)
 Trevor's Bat (English)
 Trevors Bulldoggfledermaus (German)
 Trevor's Free-tailed Bat (English)
 Trevor's Mops Bat (English)
 Triaenops (French)
 Triaenops à grandes oreilles (French)
 Triaenops de Madagascar (French)
 Triaenops de Paulian (French)
 Triaenops de Trouessart (French)
 Triaenops malgache (French)
 Triaenops roux (French)
 Tricoloured Mouse-eared Bat (English)
 Trident (French)
 Trident bat (English)
 Trident bats (English)
 Trident Bats (English)
 Trident bats (English)
 Trident Bats (English)
 Trident de Patrizi (French)
 Trident du désert (French)
 Trident Leaf-nosed Bat (English)
 Trident Leaf-nosed Bats (English)
 Trident Leaf-nosed Bats (English)
 Trident Rhinolophe (English)
 Trident-nosed Bats (English)
 Tridents (French)
 Triënopoë afro (Italian)
 Triënopi (Italian)
 Triple nose-leaf bats (English)
 Trispalvis peléausis (Lithuanian)
 Trobojni šišmiš (Bosnian)
 Trobojni šišmiš (Croatian)
 trojzubcoví pavrápenci (Czech)
 Tropical Long-eared Bats (English)
 Trouessarts Dreizahn-Blattnase (German)

Taxon Name

Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
EMBALLONURIDAE Gervais, 1855
Taphozous E. Geoffroy St.-Hilaire, 1818
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Rhinolophus mehelyi Matschie, 1901
Saccolaimus Temminck, 1838
Scotonycteris Matschie, 1894
Myotis tricolor (Temminck, 1832)
Myotis tricolor (Temminck, 1832)
Epomophorus labiatus (Temminck, 1837)
Myotis tricolor (Temminck, 1832)
Rhinolophus alcyone Temminck, 1853
Myotis tricolor (Temminck, 1832)
Hipposideros fuliginosus (Temminck, 1853)
Plecotus teneriffae Barrett-Hamilton, 1907
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Mops (Xiphonycteris) thersites (Thomas, 1903)
Scotoecus albofuscus (Thomas, 1890)
Mimetillus thomasi Hinton, 1920
Scotoecus albofuscus (Thomas, 1890)
Mimetillus thomasi Hinton, 1920
Scotoecus albofuscus (Thomas, 1890)
Casinycteris argynnis Thomas, 1910
Scotoecus albofuscus (Thomas, 1890)
Myotis tricolor (Temminck, 1832)
Rhinolophus ferrumequinum (Schreber, 1774)
Pipistrellus (Pipistrellus) nanulus Thomas, 1904
Pipistrellus (Pipistrellus) nanulus Thomas, 1904
Neoromicia guineensis (Bocage, 1889)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
EMBALLONURIDAE Gervais, 1855
Taphozous E. Geoffroy St.-Hilaire, 1818
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Tadarida ventralis (Heuglin, 1861)
Tadarida ventralis (Heuglin, 1861)
Mops (Mops) trevori J.A. Allen, 1917
Mops (Mops) trevori J.A. Allen, 1917
Mops (Mops) trevori J.A. Allen, 1917
Mops (Mops) trevori J.A. Allen, 1917
Triaenops Dobson, 1871
Paratriaenops auritus (G. Grandidier, 1912)
Paratriaenops furculus (Trouessart, 1907)
Paratriaenops pauliani (Goodman and Ranivo, 2008)
Paratriaenops furculus (Trouessart, 1907)
Paratriaenops furculus (Trouessart, 1907)
Triaenops menamena Goodman and Ranivo, 2009
Myotis tricolor (Temminck, 1832)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
HIPPOSIDERIDAE Lydekker, 1891
Asellia Gray, 1838
RHINONYCTERIDAE J. E. Gray, 1866
Triaenops Dobson, 1871
Asellia patrizii de Beaux, 1931
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Asellia Gray, 1838
Triaenops Dobson, 1871
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Cloeotis Thomas, 1901
Asellia Gray, 1838
Triaenops afer Peters, 1877
Triaenops Dobson, 1871
Triaenops Dobson, 1871
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Asellia Gray, 1838
Laephotis Thomas, 1901
Paratriaenops furculus (Trouessart, 1907)

Common Name

Trouessart's Triden Bat (English)
 Trouessart's Trident Bat (English)
 Trujillo's House Bat (English)
 Trujillos Hausfledermaus (German)
 Trumpet-eared Bats (English)
 Tsingy-Freischwanzfledermaus (German)

Tuŋa (Vai (Liberia))
 Tunisiansiippa (Finnish)
 Turkinserotiini (Finnish)
 Tusseflaggermus (Norwegian)
 Twilight bats (English)
 Ušan širokouchý (Czech)
 ušan balkánský (Czech)
 ušan berberský (Czech)
 ušan egyptský (Czech)
 ušan habešský (Czech)
 Ušan kanárský (Czech)
 ušan pustinný (Czech)
 ušanové (Czech)
 Ucháč kanársky (Slovak)
 Uchaňa čierna (Slovak)
 Üçrəng gecə şəbpərəsi (Azerbaijani)
 Udnan (Hebrew)
 Unerwartete Pipistrelle (German)
 Unerwartete Zwergfledermaus (German)
 Uolinis šikšnys (Lithuanian)
 upír černolící (Czech)
 upír čerwenokrky (Czech)
 upír bledý (Czech)
 Upír egypcekyý (Czech)
 upír obecný (Czech)
 upírové (Czech)
 upírové (Czech)
 upírové (Czech)
 Upland Horseshoe bat (English)
 Ureglia gronda da Teneriffa (Rhaeto-Romance)
 Ureglia lunga da Hemprich (Rhaeto-Romance)
 Uszorek pustynny (Polish)
 Uzunayaklı Yarasa (Turkish)
 Uzunbarmaq gecə şəbpərəsi (Azerbaijani)
 Uzunkanatlı Yarasa (Turkish)
 Uzunkulaklı Çöl Yarasaı (Turkish)
 výložkoví kaloni (Czech)
 Vaaleaetelänlepakko (Finnish)
 Vaaleaherkko (Finnish)
 Vahemere lendlane (Estonian)
 Vahemere nahkhiir (Estonian)
 Vahemere sagarnina (Estonian)
 Väike-sagarnina (Estonian)
 Väikevidevlane (Estonian)
 Vaivaislepakko (Finnish)
 vakoví hrobkovci (Czech)
 Valevampyyrit (Finnish)
 Väliimerenherkko (Finnish)
 Valkopäälepakot (Finnish)
 vampýrovci (Czech)
 Van Bemmelen's Wrinkle-lipped Bat (English)
 Variable Bulldoggfledermaus (German)

Variegated Butterfly Bat (English)
 večerní netopýři (Czech)
 Večernica šípouchá (Slovak)
 Večernica Bottova (Slovak)
 večernica Hanáková (Slovak)
 Večernica hvízdavá (Slovak)
 Večernica južná (Slovak)
 Večernica Leachova (Slovak)
 Večernica madeirská (Slovak)
 Večernica malá (Slovak)
 Večernica Saviho (Slovak)
 večernica tripolitánska (Slovak)
 Veckläppad fladdermus (Swedish)
 Vejicati netopir (Slovenian)

Taxon Name

Paratriaenops furculus (Trouessart, 1907)
Paratriaenops furculus (Trouessart, 1907)
Scotophilus trujilloi Brooks and Bickham, 2014
Scotophilus trujilloi Brooks and Bickham, 2014
Kerivoula Gray, 1842
Paremballonura tiavato (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)
Hypsignathus monstrosus H. Allen, 1862
Myotis punicus Felten, 1977
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Nycticeinops Hill & Harrison, 1987
Barbastella barbastellus (Schreber, 1774)
Plecotus kolombatovici Dulic, 1980
Plecotus gaisleri Benda, Kiefer, Hanak & Veith, 2004
Plecotus christii Gray, 1838
Plecotus balensis Kruskop & Lavrenchenko, 2000
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859
Plecotus E. Geoffroy St.-Hilaire, 1818
Plecotus teneriffae Barrett-Hamilton, 1907
Barbastella barbastellus (Schreber, 1774)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Otonycteris hemprichii Peters, 1859
Pipistrellus (Pipistrellus) inexpectatus Aellen, 1959
Pipistrellus (Pipistrellus) inexpectatus Aellen, 1959
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Pteropus subniger (Kerr, 1792)
Eidolon helvum (Kerr, 1792)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pteropus subniger (Kerr, 1792)
 CHIROPTERA Blumenbach, 1779
 PTEROPODIDAE Gray, 1821
Pteropus Brisson, 1762
Rhinolophus hillorum Koopman, 1989
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859
Otonycteris hemprichii Peters, 1859
Myotis capaccinii (Bonaparte, 1837)
Myotis capaccinii (Bonaparte, 1837)
Miniopterus schreibersii (Kuhl, 1817)
Otonycteris hemprichii Peters, 1859
Epomophorus Bennett, 1836
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Rhinolophus blasii Peters, 1867
Myotis punicus Felten, 1977
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Rhinolophus euryale Blasius, 1853
Rhinolophus hipposideros (Bechstein, 1800)
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Saccolaimus Temminck, 1838
 MEGADERMATIDAE H. Allen, 1864
Rhinolophus euryale Blasius, 1853
 NYCTERIDAE Van der Hoeven, 1855
 NOCTILIONOIDEA Gray, 1821
Mops (Chaerephon) bemmeleni (Jentink, 1879)
Mops (Chaerephon) jobimena (Goodman and Cardiff, 2004)
Glauconycteris variegata (Tomes, 1861)
Eptesicus Rafinesque, 1820
Otonycteris hemprichii Peters, 1859
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Hypsugo savii (Bonaparte, 1837)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Tadarida teniotis (Rafinesque, 1814)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)

Common Name

Taxon Name

velcí cíponosi (Czech)
 velcí morousi (Czech)
 Veldkampovi kaloni (Czech)
 Veldkamp's Bat (English)
 Veldkamp's Dwarf Fruit-bat (English)
 Veldkamps Zwergflughund (German)
 veliký vrápenec podkovní (Czech)
 Veliki mračnik (Slovenian)
 Veliki nočnišišmiš (Bosnian)
 Veliki noćnik (Montenegrin)
 Veliki podkovnjak (Slovenian)
 Veliki potkovasti šišmiš (Bosnian)
 Veliki potkovasti šišmiš (Croatian)
 Veliki potkovičar (Montenegrin)
 Veliki potkovnjak (Croatian)
 Veliki večernjak (Croatian)
 velkouší morousi (Czech)
 Verwechselte Pipistrelle (German)
 Verwechselte Zwergfledermaus (German)
 Vesper Bats (English)
 Vespère à grosse tête (French)
 Vespère ariel (French)
 Vespère d'Anchieta (French)
 Vespère de Savi (French)
 Vespère d'Eisentraut (French)
 Vespère murin (French)
 Vespères (French)
 Vespertil cun tscheglias (Rhaeto-Romance)
 Vespertil cun tschegls (Rhaeto-Romance)
 Vespertil da Capaccini (Rhaeto-Romance)
 Vespertil dal barbis (Rhaeto-Romance)
 Vespertil pitschen dal barbis (Rhaeto-Romance)
 Vespertil ureglia-mieur punic (Rhaeto-Romance)
 Vespertilio di Capaccini (Italian)
 Vespertilio maghrebino (Italian)
 Vespertilio mustacchino (Italian)
 Vespertilio smarginato (Italian)
 Vespertilio smarginato (Italian)
 Vespertilion à moustaches (French)
 Vespertilion à oreilles échanrées (French)
 Vespertilion d'Angola (French)
 Vespertilion de Capaccini (French)
 Vespertilion de l'île d'Anjouan (French)
 Vespertilion de Madagascar (French)
 Vespertilion de Morris (French)
 Vespertilion de Scott (French)
 Vespertilion de Welwitsch (French)
 Vespertilion des montagnes (French)
 Vespertilion Doré (French)
 Vespertilion échanré (French)
 Vespertilion isabelle (French)
 Vespertilionid bats (English)
 Vespertilionidés (French)
 Vespertilionidi (Italian)
 Vespertilions dorés (French)
 Vestlig bredøret flagermus (Danish)
 Vetteflaggermus (Norwegian)
 Vidusjuras pakavdegunis (Latvian)
 Viiksisippa (Finnish)
 víkonos (Czech)
 víkonos africký (Czech)
 víkonos asijský (Czech)
 víkonos egyptský (Czech)
 víkonos východoafrický (Czech)
 víkonos velký (Czech)
 víkonosi (Czech)
 víkonosovití (Czech)
 Vinkeløreflaggermus (Norwegian)
 Vinson se spleetneusvlermuis (Afrikaans)
 Vinsons Schlitznasen-Fledermaus (German)
 Vinson's Slit-faced Bat (English)
 vlašťovčí netopýři (Czech)
 vlastní vrápenci (Czech)

Triaenops Dobson, 1871
Mops Lesson, 1842
Nanonycteris Matschie, 1899
Nanonycteris veldkampii (Jentink, 1888)
Nanonycteris veldkampii (Jentink, 1888)
Nanonycteris veldkampii (Jentink, 1888)
Rhinolophus ferrumequinum (Schreber, 1774)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Nyctalus lasiopterus (Schreber, 1780)
Otomops Thomas, 1913
Pipistrellus (Pipistrellus) permixtus Aellen, 1957
Pipistrellus (Pipistrellus) permixtus Aellen, 1957
 VESPERTILIONIDAE Gray, 1821
Nycticeinops crassulus (Thomas, 1904)
Hypsugo ariel (Thomas, 1904)
Neoromicia anchietae (Seabra, 1900)
Hypsugo savii (Bonaparte, 1837)
Nycticeinops eisentrauti (Hill, 1968)
Hypsugo musciculus (Thomas, 1913)
Hypsugo Kolenati, 1856
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis capaccinii (Bonaparte, 1837)
Myotis mystacinus (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Myotis punicus Felten, 1977
Myotis capaccinii (Bonaparte, 1837)
Myotis punicus Felten, 1977
Myotis mystacinus (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis mystacinus (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis tricolor (Temminck, 1832)
Myotis capaccinii (Bonaparte, 1837)
Myotis anjouanensis Dorst, 1960
Myotis goudoti (A. Smith, 1834)
Myotis morrisi Hill, 1971
Myotis scotti Thomas, 1927
Myotis welwitschii (Gray, 1866)
Myotis capaccinii (Bonaparte, 1837)
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
 VESPERTILIONIDAE Gray, 1821
 VESPERTILIONIDAE Gray, 1821
 VESPERTILIONIDAE Gray, 1821
 MYZOPODIDAE Thomas, 1904
Barbastella barbastellus (Schreber, 1774)
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus euryale Blasius, 1853
Myotis mystacinus (Kuhl, 1817)
Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma cystops Thomas, 1903
Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma macinnesi Hayman, 1937
Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma E. Geoffroy St.-Hilaire, 1818
 RHINOPOMATIDAE Dobson, 1872
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Nycteris vinsoni Dalquest, 1965
Nycteris vinsoni Dalquest, 1965
Nycteris vinsoni Dalquest, 1965
Scotoecus Thomas, 1901
 RHINOLOPHIDAE Gray, 1825

Common Name

Vleermuizen (Dutch)
 Vliegende honden (Dutch)
 Vlindervlermuis (Afrikaans)
 vlnoušek bronzový (Czech)
 vlnoušek damarský (Czech)
 vlnoušek guinejský (Czech)
 vlnoušek habešský (Czech)
 vlnoušek malý (Czech)
 vlnoušek východoafrický (Czech)
 vlnoušek západoafrický (Czech)
 vlnouškové (Czech)
 vrápenčí (Czech)
 vrápenčí (Czech)
 vrápenčovi (Czech)
 vrápenčovití (Czech)
 vrápenec (Czech)
 vrápenec Adamův (Czech)
 vrápenec Alcýone (Czech)
 Vrápenec Blasiův (Czech)
 vrápenec cíponosý (Czech)
 vrápenec Darlingův (Czech)
 vrápenec Dentův (Czech)
 vrápenec guinejský (Czech)
 vrápenec Hildebrandtův (Czech)
 vrápenec Hillův (Czech)
 Vrápenec jižní (Czech)
 vrápenec jihoafrický (Czech)
 vrápenec kouřový (Czech)
 vrápenec Landerův (Czech)
 vrápenec liberijský (Czech)

 vrápenec Maclaudův (Czech)
 Vrápenec malý (Czech)
 Vrápenec Mehelyův (Czech)
 vrápenec menší (Czech)
 vrápenec moravský (Czech)
 vrápenec mosambický (Czech)
 vrápenec pahrbeční (Czech)
 vrápenec pahrbeční (Czech)
 vrápenec podkovní (Czech)
 vrápenec podkovníček (Czech)
 vrápenec podkrovní (Czech)
 vrápenec pouštní (Czech)
 vrápenec ruwenzorijský (Czech)
 vrápenec sakežianský (Czech)
 Vrápenec středozemský (Czech)
 vrápenec tanganjický (Czech)

 vrápenec ugandský (Czech)
 vrápenec východoafrický (Czech)
 vrápenec veliký (Czech)
 Vrápenec velký (Czech)
 vrápenec západoafrický (Czech)
 vrápeník malý (Czech)
 vrápeník podkovní (Czech)
 vrápeníkové (Czech)
 vrápeníkové (Czech)
 vrápeníkové (Czech)
 Vuoristopii (Finnish)
 Wahlberg se vrugtevlermuis (Afrikaans)
 Wahlberg's Epauletted Fruit-bat (English)
 Wahlberg's Epauletted Fruit-bat (English)
 Wahlbergs Epaulettenflughund (German)
 Wahlberg-witkolvrugtevlermuis (Afrikaans)
 Wäissbordfliedermaus (Luxembourgish)
 Wald-Hufeisennase (German)
 Warzen-Breitflügelfledermaus (German)
 Wat-Wat (Arabian)
 wečerník prowtaný (Czech)
 wečerník senegalský (Czech)
 wečerníkové (Czech)
 Weißbauch-Bulldoggfledermaus (German)
 Weißbäuchige Hausfledermaus (German)
 Weissbauchige Schwirrfledermaus (German)

Taxon Name

CHIROPTERA Blumenbach, 1779
 PTEROPODIDAE Gray, 1821
Glauconycteris variegata (Tomes, 1861)
Kerivoula cuprosa Thomas, 1912
Kerivoula argentata Tomes, 1861
Kerivoula smithii Thomas, 1880
Kerivoula eriophora (Heuglin, 1877)
Kerivoula lanosa (A. Smith, 1847)
Kerivoula africana Dobson, 1878
Kerivoula phalaena Thomas, 1912
Kerivoulinae Miller, 1907
Rhinolophus Lacépède, 1799
 RHINOLOPHOIDEA J. E. Gray, 1825
 RHINOLOPHOIDEA J. E. Gray, 1825
 RHINOLOPHIDAE Gray, 1825
Rhinolophus Lacépède, 1799
Rhinolophus adami Aellen and Brosset, 1968
Rhinolophus alcýone Temminck, 1853
Rhinolophus blasii Peters, 1867
Triaenops afer Peters, 1877
Rhinolophus darlingi K. Andersen, 1905
Rhinolophus denti Thomas, 1904
Rhinolophus guineensis Eisentraut, 1960
Rhinolophus hildebrandtii Peters, 1878
Rhinolophus hilli Aellen, 1973
Rhinolophus euryale Blasius, 1853
Rhinolophus capensis Lichtenstein, 1823
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus landeri Martin, 1838
Rhinolophus ziamia Fahr, Vierhaus, Hutterer and Kock, 2002
Rhinolophus maclaudi Pousargues, 1898
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus mehelyi Matschie, 1901
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus swinnyi Gough, 1908
Rhinolophus blasii Peters, 1867
Rhinolophus clivus Cretzschmar, 1828
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus clivus Cretzschmar, 1828
Rhinolophus ruwenzorii J. Eric Hill, 1942
Rhinolophus sakejiensis Cotterill, 2002
Rhinolophus euryale Blasius, 1853
Rhinolophus maendeleo Kock, Csorba and Howell, 2000
Rhinolophus eloquens K. Andersen, 1905
Rhinolophus deckenii Peters, 1868
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hillorum Koopman, 1989
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus Lacépède, 1799
Rhinolophus Lacépède, 1799
Plecotus E. Geoffroy St.-Hilaire, 1818
Hypsugo savii (Bonaparte, 1837)
Epomophorus wahlbergi (Sundevall, 1846)
Epomophorus wahlbergi (Sundevall, 1846)
Epomophorus wahlbergi (Sundevall, 1846)
Epomophorus wahlbergi (Sundevall, 1846)
Epomophorus wahlbergi (Sundevall, 1846)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Rhinolophus silvestris Aellen, 1959
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Roussettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Taphozous E. Geoffroy St.-Hilaire, 1818
Mops (Mops) niveiventer Cabrera and Ruxton, 1926
Scotophilus leucogaster (Cretzschmar, 1826)
Scotophilus leucogaster (Cretzschmar, 1826)

Common Name

Taxon Name

Weissfluegel-Hausfledermaus (German)	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
Weißflügel-Hausfledermaus (German)	<i>Scotoecus albofuscus</i> (Thomas, 1890)
Weisskehl-Hausfledermaus (German)	<i>Scotoecus albigula</i> Thomas, 1909
Weißrandfledermaus (German)	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Weißrand-Pipistrelle (German)	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Weißflügel Pipistrelle (German)	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
Weißflügel-Zwergfledermaus (German)	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
weloblánec lupenatý (Czech)	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
Welwitch's Bat (English)	<i>Myotis welwitschii</i> (Gray, 1866)
Welwitsch se langhaarvlermuis (Afrikaans)	<i>Myotis welwitschii</i> (Gray, 1866)
Welwitsch-langhaarvlermuis (Afrikaans)	<i>Myotis welwitschii</i> (Gray, 1866)
Welwitsch's Hairy Bat (English)	<i>Myotis welwitschii</i> (Gray, 1866)
Welwitschs Mausohr (German)	<i>Myotis welwitschii</i> (Gray, 1866)
Welwitsch's Mouse-eared Bat (English)	<i>Myotis welwitschii</i> (Gray, 1866)
Welwitsch's Myotis (English)	<i>Myotis welwitschii</i> (Gray, 1866)
West African fruit bats (English)	<i>Scotonycteris</i> Matschie, 1894
West African Rousette (English)	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Westafrikanische Nachtfledermaus (German)	<i>Glauconycteris poensis</i> (Gray, 1842)
Westelijke mopsvlermuis (Dutch)	<i>Barbastella barbastellus</i> (Schreber, 1774)
Western Africa leaf-nosed bat (English)	<i>Hipposideros jonesi</i> Hayman, 1947
Western Africa Pipistrelle (English)	<i>Hypsugo musciculus</i> (Thomas, 1913)
Western Barbastelle (English)	<i>Barbastella barbastellus</i> (Schreber, 1774)
Western Barbastelle Bat (English)	<i>Barbastella barbastellus</i> (Schreber, 1774)
Western Sheath-tailed Bat (English)	<i>Paremballonura tiavato</i> (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)
Western Yellow Bat (English)	<i>Scotophilus tandrefana</i> Goodman, Jenkins and Ratrimomanarivo, 2005
Westliche Hausfledermaus (German)	<i>Scotophilus tandrefana</i> Goodman, Jenkins and Ratrimomanarivo, 2005
Westlicher Woermanns Langzungen-Flughund (German)	<i>Megaloglossus azagnyi</i> Nesi, Kadjo and Hassanin, 2012
Westlike breedearflearnûs (Frisian)	<i>Barbastella barbastellus</i> (Schreber, 1774)
Whiskered Bat (English)	<i>Myotis mystacinus</i> (Kuhl, 1817)
Whiskered Mouse-eared Bat (English)	<i>Myotis mystacinus</i> (Kuhl, 1817)
Whiskered Myotis (English)	<i>Myotis mystacinus</i> (Kuhl, 1817)
White stripe bat (English)	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
White-bellied Free-tailed Bat (English)	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
White-bellied Free-tailed Bat (English)	<i>Mops (Mops) niveiventer</i> Cabrera and Ruxton, 1926
White-bellied House Bat (English)	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
White-bellied Lesser House Bat (English)	<i>Scotoecus albigula</i> Thomas, 1909
White-bellied Mops Bat (English)	<i>Mops (Mops) niveiventer</i> Cabrera and Ruxton, 1926
White-bellied Yellow Bat (English)	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
White-fingered Free-tailed Bat (English)	<i>Mops (Xiphonycteris) brachypterus</i> (Peters, 1852)
White-shouldered bat (English)	<i>Mops (Mops) leucostigma</i> (G.M. Allen, 1918)
White-spotted Butterfly Bat (English)	<i>Glauconycteris alboguttata</i> J.A. Allen, 1917
White-throated Lesser House Bat (English)	<i>Scotoecus albigula</i> Thomas, 1909
White-winged Bat (English)	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
White-winged Pipistrelle Bat (English)	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
White-winged Serotine (English)	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
Whitleys Pergamentflügel-Fledermaus (German)	<i>Myotis whitleyi</i> (Scharff, 1900)
Whitley's Winged-mouse Bat (English)	<i>Myotis whitleyi</i> (Scharff, 1900)
Willards horseshoe bat (English)	<i>Rhinolophus willardi</i> Kerbis Peterhans and Fahr, 2013
Willards Hufeisennase (German)	<i>Rhinolophus willardi</i> Kerbis Peterhans and Fahr, 2013
Wilson's Long-fingered Bat (English)	<i>Miniopterus wilsoni</i> Monadjem, Guyton, Naskrecki, Richards, Kropff and Dalton, 2020
Wimperefledermaus (Luxembourgish)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Wimperfledermaus (German)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Winged-mouse Bats (English)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Winged-rat Free-tailed Bats (English)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Wing-gland bat (English)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Wing-gland Bats (English)	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Winton se langoorvlermuis (Afrikaans)	<i>Cistugo lesueuri</i> Roberts, 1919
Winton-langoorvlermuis (Afrikaans)	<i>Cistugo</i> Thomas, 1912
Winton's Long-eared Bat (English)	<i>Laephotis wintoni</i> Thomas, 1901
Winziger Epaulettenflughund (German)	<i>Laephotis wintoni</i> Thomas, 1901
Witlyfvlermuis (Afrikaans)	<i>Laephotis wintoni</i> Thomas, 1901
Woastynggrutearflearnûs (Frisian)	<i>Epomophorus minimus</i> Claessen & De Vree, 1991
Woermann's Bat (English)	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
Woermann's Fruit bat (English)	<i>Otonycteris hemprichii</i> Peters, 1859
Woermann's Langzungen-Flughund (German)	<i>Megaloglossus woermanni</i> Pagenstecher, 1885
Woermann's Long-tongued Fruit-bat (English)	<i>Megaloglossus woermanni</i> Pagenstecher, 1885
Woestijndwergvlermuis (Dutch)	<i>Megaloglossus woermanni</i> Pagenstecher, 1885
Woestijngrootoorvlermuis (Dutch)	<i>Megaloglossus woermanni</i> Pagenstecher, 1885
Wolf-dog Bats (English)	<i>Hypsugo ariel</i> (Thomas, 1904)
	<i>Otonycteris hemprichii</i> Peters, 1859
	<i>MOLOSSIDAE</i> Gervais, 1856

Common Name

Wolf-dog Bats (English)
 Wollfledermäuse (German)
 Wood se spleetneusvlermuis (Afrikaans)
 Wood's Long-eared Bat (English)
 Woods Schlitznasen-Fledermaus (German)
 Wood's Slit-faced Bat (English)
 Woodse Langoorvlermuis (Afrikaans)
 Wood-spleetneusvlermuis (Afrikaans)
 Woolly bats (English)
 Woolly Bats (English)
 Woolly Rousette (English)
 wrápenec menší (Czech)
 wrápenec pahrbečný (Czech)
 wrápenec podkowní (Czech)
 wrápenec trojzeykový (Czech)
 wrápenecové (Czech)
 Wrinkled-lipped Bats (English)
 Wrinkled-lipped Bats (English)
 Wrinkle-lipped Bat (English)
 Wrinkle-lipped Bats (English)
 Wüstegroussouer (Luxembourgish)
 Wüsten-Großohr (German)
 Wüsten-Großohren (German)
 Wytrâne dwerchflearmûs (Frisian)
 Yaznuy Katan (Hebrew)
 Yellow bats (English)
 Yellow House Bat (English)
 Yellow Pipistrelle Bat (English)
 Yellow Serotine (English)
 Yellow-bellied House Bat (English)
 Yellow-haired fruit bat (English)
 Yellow-winged Bat (English)
 Yellow-winged Bats (English)
 Yellow-winged Short-palate Fruit-bat (English)
 Ynsniene flearmûs (Frisian)
 Ystlum adain-gam Schreiber (Welsh)
 Ystlum adain-lydan Botta (Welsh)
 Ystlum adain-lydan llwydfelyn (Welsh)
 Ystlum barfog (Welsh)
 Ystlum beddrod tinfoel (Welsh)
 Ystlum clustlydan Maghreb (Welsh)
 Ystlum cynffonog (Welsh)
 Ystlum du (Welsh)
 Ystlum Geoffroy (Welsh)
 Ystlum hirfys (Welsh)
 Ystlum hirglust Hemprich (Welsh)
 Ystlum hirglust yr Ynysoedd Dedwydd (Welsh)
 Ystlum Leisler (Welsh)
 Ystlum lleiaf Hanak (Welsh)
 Ystlum lleiaf Kuhl (Welsh)
 Ystlum lleiaf Madeira (Welsh)
 Ystlum lleiaf Savi (Welsh)
 Ystlum mawr yr Asores (Welsh)
 Ystlum mwyaf (Welsh)
 Ystlum pedol Blasius (Welsh)
 Ystlum pedol lleiaf (Welsh)
 Ystlum pedol Mehely (Welsh)
 Ystlum pedol Môr y Canoldir (Welsh)
 Ystlum pedol mwyaf (Welsh)
 Ystlum-ffrwythau yr Aifft (Welsh)
 Ystlum-lleiaf cyffredin (Welsh)
 Ystlum-lleiaf soprano (Welsh)
 Zaire Gland-tailed Bat (English)
 zakrslí netopýři (Czech)
 Zenati Mausohr (German)

Zenati myotis (English)

Zenker's Fruit Bat (English)
 Zenkers Harlekin-Flughund (German)
 Zenker's Harlequin Fruit-bat (English)
 Zenker's Tear-drop Fruit-bat (English)
 Zenker's Traenenflughund (German)

Taxon Name

Molossinae Gervais, 1856
Kerivoula Gray, 1842
Nycteris woodi K. Andersen, 1914
Nycteris woodi K. Andersen, 1914
Nycteris woodi K. Andersen, 1914
Nycteris woodi K. Andersen, 1914
Nycteris woodi K. Andersen, 1914
Nycteris woodi K. Andersen, 1914
Kerivoulinae Miller, 1907
Kerivoula Gray, 1842
Stenonycteris lanosus (Thomas, 1906)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus ferrumequinum (Schreber, 1774)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Rhinolophus Lacépède, 1799
MOLOSSIDAE Gervais, 1856
Molossinae Gervais, 1856
Tadarida teniotis (Rafinesque, 1814)
Mops (Chaerephon) Dobson, 1874
Otonycteris hemprichii Peters, 1859
Otonycteris hemprichii Peters, 1859
Otonycteris Peters, 1859
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Rhinopoma cystops Thomas, 1903
Scotophilus Leach, 1821
Scotophilus dinganii (A. Smith, 1833)
Nycticeinops grandidieri (Dobson, 1876)
Nycticeinops grandidieri (Dobson, 1876)
Scotophilus dinganii (A. Smith, 1833)
Eidolon helvum (Kerr, 1792)
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Lavia Gray, 1838
Casinycteris argynnis Thomas, 1910
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Miniopterus schreibersii (Kuhl, 1817)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Myotis mystacinus (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Myotis punicus Felten, 1977
Tadarida teniotis (Rafinesque, 1814)
Barbastella barbastellus (Schreber, 1774)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis capaccinii (Bonaparte, 1837)
Otonycteris hemprichii Peters, 1859
Plecotus teneriffae Barrett-Hamilton, 1907
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Hypsugo savii (Bonaparte, 1837)
Nyctalus azoreum (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus blasius, 1867
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Rhinolophus ferrumequinum (Schreber, 1774)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Mops (Chaerephon) gallagheri (Harrison, 1975)
Pipistrellus Kaup, 1829
Myotis zenatius Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019
Myotis zenatius Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019
Scotonycteris zenkeri Matschie, 1894
Scotonycteris zenkeri Matschie, 1894
Scotonycteris zenkeri Matschie, 1894
Scotonycteris zenkeri Matschie, 1894
Scotonycteris zenkeri Matschie, 1894

Common Name

Ziama Horseshoe Bat (first use Simmons, 2005: 365)
(English)
Ziama-Hufeisennase (German)

Zorra Voladora (Castilian (Spain))
Zorra Voladora Amarilla (Castilian (Spain))
Zorra Voladora de Collar (Castilian (Spain))
Zorra Voladora de Zenker (Castilian (Spain))
Zorro volador de Aldabra (Castilian (Spain))
Zorro volador de la isla Rodrigues (Castilian (Spain))
Zorro volador de las Seychelles (Castilian (Spain))
Zorro Volador De Livingston (Castilian (Spain))
Zorro volador de Madagascar (Castilian (Spain))
Zorro volador de Voeltzkow (Castilian (Spain))
Zorro volador negro de Mauricio (Castilian (Spain))
Zorro volador oscuro de Mauricio (Castilian (Spain))
Zuideuropese bulvleermuis (Dutch)
Zulu Pipistrelle Bat (English)
Zulu Serotine Bat (English)
Zulu-Pipistrelle (German)
Zulu-Zwergfledermaus (German)
Zwarte Komorenkalong (Dutch)
zweistreifige Doggengrämmler (German)
Zwerg-Bulldoggfledermaus (German)
Zwerg-Doggengrämmler (German)
Zwerg-Epaulettenflughund (German)
Zwergfledermaus (German)
Zwergfledermäuse (German)
Zwergfledermaus (Luxembourgish)
Zwergflughunde (German)
Zwerg-Hausfledermaus (German)
Zwerg-Langflügelfledermaus (German)
Zwerg-Schlitznasenfledermaus (German)
Zyklopen-Rundblattnase (German)
Zyklossen-Blattnase (German)

Taxon Name

Rhinolophus ziama Fahr, Vierhaus, Hutterer and Kock, 2002
Rhinolophus ziama Fahr, Vierhaus, Hutterer and Kock, 2002
Epomophorus wahlbergi (Sundevall, 1846)
Eidolon helvum (Kerr, 1792)
Myonycteris torquata (Dobson, 1878)
Scotonycteris zenkeri Matschie, 1894
Pteropus aldabrensis True, 1893
Pteropus rodricensis Dobson, 1878
Pteropus seychellensis A. Milne-Edwards, 1877
Pteropus livingstonii Gray, 1866
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Pteropus voeltzkowi Matschie, 1909
Pteropus niger (Kerr, 1792)
Pteropus subniger (Kerr, 1792)
Tadarida teniotis (Rafinesque, 1814)
Neoromicia zuluensis (Roberts, 1924)
Neoromicia zuluensis (Roberts, 1924)
Neoromicia zuluensis (Roberts, 1924)
Neoromicia zuluensis (Roberts, 1924)
Pteropus livingstonii Gray, 1866
Mops (Chaerephon) bivittatus (Heuglin, 1861)
Mops (Xiphonycteris) nanulus J.A. Allen, 1917
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Epomophorus minor Dobson, 1880
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus Kaup, 1829
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Nanonycteris Matschie, 1899
Afronycteris nana (Peters, 1852)
Miniopterus minor Peters, 1867
Nycteris nana (K. Andersen, 1912)
Doryrhina cyclops (Temminck, 1853)
Doryrhina cyclops (Temminck, 1853)

Appendix 3f: Common Names by Language

"Congolese"

Kubukubo	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Kubukubo	<i>Taphozous nudiventris</i> Cretzschmar, 1830
Papo	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Papo	<i>Taphozous nudiventris</i> Cretzschmar, 1830

Afrikaans

Aalwyndakvlermuis; Aalwynvlermuis	<i>Neoromicia zuluensis</i> (Roberts, 1924)
Anchieta-vlermuis	<i>Neoromicia anchietae</i> (Seabra, 1900)
Angola-langhaarvlermuis	<i>Cistugo seabrae</i> Thomas, 1912
Angola-losstertvlermuis	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
Angolase Losstertvlermuis	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
Angolase Vlerkkliervlermuis	<i>Cistugo seabrae</i> Thomas, 1912
Angola-witkolvrugtevlermuis	<i>Epomophorus angolensis</i> Gray, 1870
Ansorge se losstertvlermuis	<i>Mops (Chaerephon) ansorgei</i> (Thomas, 1913)
Ansorge-losstertvlermuis	<i>Mops (Chaerephon) ansorgei</i> (Thomas, 1913)
Bakoor-losstertvlermuis	<i>Otomops martiensseni</i> (Matschie, 1897)
Bleek losstertvlermuis	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
Bocage se vrugtevlermuis	<i>Myonycteris angolensis</i> (Bocage, 1898)
Bocage se Losstertvlermuis	<i>Tadarida aegyptiaca bocagei</i> (Seabra, 1900)
Bocage-vrugtevlermuis	<i>Myonycteris angolensis</i> (Bocage, 1898)
Bosveldsaalneusvlermuis	<i>Rhinolophus simulator</i> K. Andersen, 1904
Bosveldvlermuis	<i>Rhinolophus simulator</i> K. Andersen, 1904
Botswana-langoorvlermuis	<i>Laephotis botswanae</i> Setzer, 1971
Commerson se bladneusvlermuis	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Commerson se bladneusvlermuis	<i>Macronycteris gigas</i> (Wagner, 1845)
Commerson-bladneusvlermuis	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Commerson-bladneusvlermuis	<i>Macronycteris gigas</i> (Wagner, 1845)
Commersonse Blaarneusvlermuis	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Commersonse Blaarneusvlermuis	<i>Macronycteris gigas</i> (Wagner, 1845)
Damaralandse Wolhaarvlermuis	<i>Kerivoula argentata</i> Tomes, 1861
Damara-wolhaarvlermuis	<i>Kerivoula argentata</i> Tomes, 1861
Darling se saalneusvlermuis	<i>Rhinolophus darlingi</i> K. Andersen, 1905
Darling-saalneusvlermuis	<i>Rhinolophus darlingi</i> K. Andersen, 1905
Dent se saalneusvlermuis	<i>Rhinolophus denti</i> Thomas, 1904
Dent-saalneusvlermuis	<i>Rhinolophus denti</i> Thomas, 1904
Dentse Vlermuis	<i>Rhinolophus denti</i> Thomas, 1904
Dobson se vrugtevlermuis	<i>Epomophorus dobsonii</i> Bocage, 1889
Dobson-vrugtevlermuis	<i>Epomophorus dobsonii</i> Bocage, 1889
Drietand-blaarneusvlermuis	<i>Cloeotis percivali</i> Thomas, 1901
Drietand-bladneusvlermuis	<i>Cloeotis percivali</i> Thomas, 1901
Egiptiese losstertvlermuis	<i>Tadarida aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
Egiptiese spleetneusvlermuis	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
Egiptiese vrugtevlermuis	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Egiptiese witlyfvlermuis	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
Gambiaanse witkolvrugtevlermuis	<i>Epomophorus gambianus</i> (Ogilby, 1835)
Geel Dakvlermuis	<i>Scotophilus dinganii</i> (A. Smith, 1833)
Geel vrugtevlermuis	<i>Eidolon helvum</i> (Kerr, 1792)
Geoffroy se saalneusvlermuis	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
Geoffroy-saalneusvlermuis	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
Geoffroyse vlermuis	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
Gevlekte losstertvlermuis	<i>Mops (Chaerephon) bivittatus</i> (Heuglin, 1861)
Gewone-spleetneusvlermuis	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
Groot geel dakvlermuis	<i>Scotophilus nigrita</i> (Schreber, 1774)
Groot grotvlermuis	<i>Miniopterus inflatus</i> Thomas, 1903
Groot spleetneusvlermuis	<i>Nycteris grandis</i> Peters, 1865
Grootoor-losstertvlermuis	<i>Tadarida lobata</i> (Thomas, 1891)
Groter spleetneusvlermuis	<i>Nycteris macrotis</i> Dobson, 1876
Haagnerse Platkopvlermuis	<i>Sauromys petrophilus haagneri</i> (Roberts, 1917)
Harige Langoorvlermuis	<i>Nycteris hispida</i> (Schreber, 1774)
Harige spleetneusvlermuis	<i>Nycteris hispida</i> (Schreber, 1774)
Hildebrandt se saalneusvlermuis	<i>Rhinolophus hildebrandtii</i> Peters, 1878
Hildebrandt-saalneusvlermuis	<i>Rhinolophus hildebrandtii</i> Peters, 1878
Hildebrandtse Vlermuis	<i>Rhinolophus hildebrandtii</i> Peters, 1878
Kaapse Blaarneusvlermuis	<i>Hipposideros caffer</i> (Sundevall, 1846)

Kaapse dakvlermuis	<i>Laephotis capensis</i> (A. Smith, 1829)
Kaapse Langhaarvlermuis	<i>Myotis tricolor</i> (Temminck, 1832)
Kaapse Langoorvlermuis	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
Kaapse saalneusvlermuis	<i>Rhinolophus capensis</i> Lichtenstein, 1823
Kaapse Vlermuis	<i>Rhinolophus capensis</i> Lichtenstein, 1823
Klein dakvlermuis	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
Klein geel dakvlermuis	<i>Scotophilus viridis</i> (Peters, 1852)
Klein grotvlermuis	<i>Miniopterus fraterculus</i> Thomas and Schwann, 1906
Klein losstertvlermuis	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
Klein Vrugtevlermuis	<i>Epomophorus crypturus</i> Peters, 1852
Klein wolhaarvlermuis	<i>Kerivoula lanosa</i> (A. Smith, 1847)
Kuhl se vlermuis	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Kuhl-vlermuis	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Lander se saalneusvlermuis	<i>Rhinolophus landeri</i> Martin, 1838
Lander-saalneusvlermuis	<i>Rhinolophus landeri</i> Martin, 1838
Langoor-vlermuise	<i>NYCTERIDAE</i> Van der Hoeven, 1855
Langstert-dakvlermuis	<i>Eptesicus (Cnephæus) hottentotus</i> (A. Smith, 1833)
Lesueur se langhaarvlermuis	<i>Cistugo lesueuri</i> Roberts, 1919
Lesueur-langhaarvlermuis	<i>Cistugo lesueuri</i> Roberts, 1919
Lesueurse Vlerkkliervlermuis	<i>Cistugo lesueuri</i> Roberts, 1919
Madagaskarse grootlosstertvlermuis	<i>Tadarida fulminans</i> (Thomas, 1903)
Midas se losstertvlermuis	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Midas-losstertvlermuis	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Namib-langoorvlermuis	<i>Laephotis namibensis</i> Setzer, 1971
Natalese losstertvlermuis	<i>Mormopterus acetabulosus</i> (Hermann, 1804)
Nigeriese losstertvlermuis	<i>Mops (Chaerephon) nigeriae</i> (Thomas, 1913)
Peters se witkolvrugtevlermuis	<i>Epomophorus crypturus</i> Peters, 1852
Peters-witkolvrugtevlermuis	<i>Epomophorus crypturus</i> Peters, 1852
Piesangvlermuis	<i>Afronycteris nana</i> (Peters, 1852)
Platkop-losstertvlermuis	<i>Sauromys petrophilus</i> (Roberts, 1917)
Rendal-dakvlermuis	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
Rendall se dakvlermuis	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
Reuse losstertvlermuis	<i>Tadarida ventralis</i> (Heuglin, 1861)
Roeskleurvlermuis	<i>Pipistrellus (Pipistrellus) rusticus</i> (Tomes, 1861)
Rooi langhaarvlermuis	<i>Myotis bocagii</i> (Peters, 1870)
Rüppell se saalneusvlermuis	<i>Rhinolophus fumigatus</i> Rüppell, 1842
Rüppell se vlermuis	<i>Vansonia rueppellii</i> (J.B. Fischer, 1829)
Rüppell-saalneusvlermuis	<i>Rhinolophus fumigatus</i> Rüppell, 1842
Rüppell-vlermuis	<i>Vansonia rueppellii</i> (J.B. Fischer, 1829)
Schlieffen se vlermuis	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
Schlieffen-vlermuis	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
Schreiber-grotvlermuis	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Schreibers se grotvlermuis	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Skedestertvlermuis	<i>Coleura afra</i> (Peters, 1852)
Somali-dakvlermuis	<i>Neoromicia somalica</i> (Thomas, 1901)
Spitssaalneusvlermuis	<i>Rhinolophus blasii</i> Peters, 1867
Sundevall se bladneusvlermuis	<i>Hipposideros caffer</i> (Sundevall, 1846)
Sundevall-bladneusvlermuis	<i>Hipposideros caffer</i> (Sundevall, 1846)
Sundevallse blaarneusvlermuis	<i>Hipposideros caffer</i> (Sundevall, 1846)
Swinny se saalneusvlermuis	<i>Rhinolophus swinnyi</i> Gough, 1908
Swinny-saalneusvlermuis	<i>Rhinolophus swinnyi</i> Gough, 1908
Swinnyse Vlermuis	<i>Rhinolophus swinnyi</i> Gough, 1908
Temminck se langhaarvlermuis	<i>Myotis tricolor</i> (Temminck, 1832)
Temminck-langhaarvlermuis	<i>Myotis tricolor</i> (Temminck, 1832)
Thomas se vlermuis	<i>Scotoecus albofuscus</i> (Thomas, 1890)
Thomas-vlermuis	<i>Scotoecus albofuscus</i> (Thomas, 1890)
Transvaalse losstertvlermuis	<i>Tadarida ventralis</i> (Heuglin, 1861)
Vinson se spleetneusvlermuis	<i>Nycteris vinsoni</i> Dalquest, 1965
Vlindervlermuis	<i>Glauconycteris variegata</i> (Tomes, 1861)
Wahlberg se vrugtevlermuis	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
Wahlberg-witkolvrugtevlermuis	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
Welwitsch se langhaarvlermuis	<i>Myotis welwitschii</i> (Gray, 1866)
Welwitsch-langhaarvlermuis	<i>Myotis welwitschii</i> (Gray, 1866)
Winton se langoorvlermuis	<i>Laephotis wintoni</i> Thomas, 1901
Winton-langoorvlermuis	<i>Laephotis wintoni</i> Thomas, 1901
Witlyfvlermuis	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
Wood se spleetneusvlermuis	<i>Nycteris woodi</i> K. Andersen, 1914
Woodse Langoorvlermuis	<i>Nycteris woodi</i> K. Andersen, 1914
Wood-spleetneusvlermuis	<i>Nycteris woodi</i> K. Andersen, 1914

Albanian

Barbastela e zakonshme	<i>Barbastella barbastellus</i> (Schreber, 1774)
Lakuriq nate bisht-lire	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Lakuriq nate bishtlirë	<i>Tadarida teniotis</i> (Rafinesque, 1814)

Անապատային մաշկեղ
 Բեղավոր գիշերաչղջիկ
 Բլազիի կամ
 Գաջան փոքրաչղջիկ
 Եգիպտական մեծաչղջիկ
 Եռագույն գիշերաչղջիկ
 Եվրոպական լայնական չղջիկ
 Թզուկ փոքրաչղջիկ
 Իզաբելլայի մաշկեղ
 Լայնական ծալքաշուրթ չղջիկ
 Կանարական կամ Տենեռիֆյան ականջեղ
 Հանակի փոքրաչղջիկ
 Հարավային պայտաքիթ
 Հսկա իրիկնաչղջիկ
 Մադեյրական փոքրաչղջիկ
 Մաշեղանման փոքրաչղջիկ
 Մեծ պայտաքիթ
 Մեհելիի պայտաքիթ
 Մերկիոր պարկաթ
 Միջերկրածովային գիշերաչղջիկ
 Միջերկրածովային փոքրաչղջիկ
 Սովորական երկարաթև չղջիկ
 Սպիտակափոր սրականջ
 Փյունիկյան գիշերաչղջիկ
 Փոքր իրիկնաչղջիկ
 Փոքր պայտաքիթ
 Միջերկրածովային պայտաքիթ

Eptesicus (Cnephaeus) bottae (Peters, 1869)
Myotis mystacinus (Kuhl, 1817)
Rhinolophus blasii Peters, 1867
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Barbastella barbastellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Tadarida teniotis (Rafinesque, 1814)
Plecotus teneriffae Barrett-Hamilton, 1907
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Rhinolophus euryale Blasius, 1853
Nyctalus lasiopterus (Schreber, 1780)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Hypsugo savii (Bonaparte, 1837)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus mehelyi Matschie, 1901
Taphozous nudiventris Cretzschmar, 1830
Myotis capaccinii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Otonycteris hemprichii Peters, 1859
Myotis punicus Felten, 1977
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus blasii Peters, 1867

Azande (DRC)

Fulo
 Fulo
 Fulo
 Ndim
 Ndim
 Nkulo

Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Mops (Chaerephon) nigeriae (Thomas, 1913)
Afronycteris nana (Peters, 1852)
Epomophorus labiatus (Temminck, 1837)
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Hipposideros caffer (Sundevall, 1846)

Azerbaijani

İzabel gönlüce
 Ağqarın oxqulaq yarasa
 Adi uzunqanad yarasa
 Avropa enliqulağı
 Azor axşam yarasası
 Bıqlı gecə şəbpərəsi
 Blazius nalburunu
 Böyük nalburun
 Bükükdodaq enliqulaq
 Cırtan şəbpərə
 Cənub nalburunu
 Eynəkli nalburun
 Hanaki cırtan şəbpərəsi
 İpaqqarın məzar yarasası
 Kanar palazqulağı
 Kiçik şəbpərə
 Kicik axşam yarasası
 Kiçik nalburun
 Küli şəbpərəsi
 Maderiya və ya Kanar şəbpərəsi
 Misir meyvə yarasası
 Nəhəng axşam yarasası
 Səhra gönlücəsi
 Savi şəbpərəsi
 Siçanqulaq gecə şəbpərəsi
 Üçrəng gecə şəbpərəsi
 Uzunbarmaq gecə şəbpərəsi

Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Otonycteris hemprichii Peters, 1859
Miniopterus schreibersii (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Nyctalus azoreum (Thomas, 1901)
Myotis mystacinus (Kuhl, 1817)
Rhinolophus blasii Peters, 1867
Rhinolophus ferrumequinum (Schreber, 1774)
Tadarida teniotis (Rafinesque, 1814)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Taphozous nudiventris Cretzschmar, 1830
Plecotus teneriffae Barrett-Hamilton, 1907
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Nyctalus lasiopterus (Schreber, 1780)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Hypsugo savii (Bonaparte, 1837)
Myotis punicus Felten, 1977
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis capaccinii (Bonaparte, 1837)

Basque

Baratze-saguzar isabeldar
 Baso-saguzar
 Baso-saguzar arrunt
 Belarrihandi kanariar
 Blasius ferra-saguzar

Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Plecotus teneriffae Barrett-Hamilton, 1907
Rhinolophus blasii Peters, 1867

Botta serotino
 Ferra-saguzar handi
 Ferra-saguzar mediterraneo
 Ferra-saguzar txikia
 Fruitu-saguzar egiptoar
 Gau-saguzar azoretar
 Gau-saguzar handi
 Geoffroy saguzar
 Hanak pipistrello
 Hemprich belarrihandi
 Hilobi-saguzar gibelsol
 Kuhl pipistrello
 Kuhl saguzar
 Leisler gau-saguzar
 Mehelyi ferra-saguzar
 Pipistrello arrunt
 Pipistrello madeirar
 Pipistrello mediterraneo
 Saguzar
 Saguzar arratoi-belarri mairutar
 Saguzar biboteduna
 Saguzar buztanluze europar
 Saguzar buztanluzea
 Saguzar hankahandia
 Saguzar hatz-luze
 Savi saguzar
 Savi saguzarra
 Schreiber saguzarra
 Schreibers saguzar

Eptesicus (Cnephaeus) bottae (Peters, 1869)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
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Rhinolophus mehelyi Matschie, 1901
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myotis punicus Felten, 1977
Myotis mystacinus (Kuhl, 1817)
Tadarida teniotis (Rafinesque, 1814)
Tadarida teniotis (Rafinesque, 1814)
Myotis capaccinii (Bonaparte, 1837)
Myotis capaccinii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)

Belarusian

Вушан канарскі
 Вячэрніца азорская
 Вячэрніца гіганцкая
 Вячэрніца малая
 Даўгакрыл звычайны
 Кажан Бота
 Кажан ізабэлавы
 Крылан егіпецкі
 Марільнік галабрухі
 Начніца вусатая
 Начніца даўгапалая
 Начніца магрыбская
 Начніца трохкаляровая
 Начніца фінікійская
 Нетапыр Ганака
 Нетапыр Куля
 Нетапыр Саві
 Нетапыр мадэрскі
 Нетапыр міжземнаморскі
 Нетапыр-карлік
 Нетапыр-пігмей
 Падкаванос Блазіўса
 Падкаванос Мегеля
 Падкаванос вялікі
 Падкаванос малы
 Падкаванос міжземнаморскі
 Стрэлавух Хемрыха
 Тадарыда еўрапейская
 Шыракавух еўрапейскі

Plecotus teneriffae Barrett-Hamilton, 1907
Nyctalus azoreum (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Taphozous nudiventris Cretzschmar, 1830
Myotis mystacinus (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Myotis punicus Felten, 1977
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis punicus Felten, 1977
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus blasii Peters, 1867
Rhinolophus mehelyi Matschie, 1901
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus euryale Blasius, 1853
Otonycteris hemprichii Peters, 1859
Tadarida teniotis (Rafinesque, 1814)
Barbastella barbastellus (Schreber, 1774)

Bengali

Indur-lenji Chamchika
 Lomba-leji Chamchika
 Savir Chamchika

Rhinopoma microphyllum (Brünnich, 1782)
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Hypsugo savii (Bonaparte, 1837)

Bosnian

Širokouhi šišmiš
 Šrajberov šišmiš
 Azorski noćni šišmiš
 Blazijev potkovasti šišmiš

Barbastella barbastellus (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)
Nyctalus azoreum (Thomas, 1901)
Rhinolophus blasii Peters, 1867

Botin šišmiš
 Dugokrili slijepi miš
 Dugoprsti šišmiš
 Egipatski letipas
 Golorepi šišmiš
 Golotrbusni grobni šišmiš
 Hanakov šišmiš
 Hemprihov dugouhi šišmiš
 Izabelin šišmiš
 Kanarski dugouhi šišmiš
 Kulijev šišmiš
 Madeirski šišmiš
 Mali šišmiš
 Mali brkati šišmiš
 Mali Lajslerov šišmiš
 Mali nočni šišmiš
 Mali potkovasti šišmiš
 Mediteranski potkovasti šišmiš
 Meheljev potkovasti šišmiš
 Patuljasti šišmiš
 Patuljasti slijepi mišić
 Punski šišmiš
 Savijev šišmiš
 Trobojni šišmiš
 Veliki nočni šišmiš
 Veliki potkovasti šišmiš

Eptesicus (Cnephaeus) bottae (Peters, 1869)
Miniopterus schreibersii (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Tadarida teniotis (Rafinesque, 1814)
Taphozous nudiventris Cretzschmar, 1830
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Otonycteris hemprichii Peters, 1859
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Plecotus teneriffae Barrett-Hamilton, 1907
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myotis mystacinus (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myotis punicus Felten, 1977
Hypsugo savii (Bonaparte, 1837)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)

Breton

Barbastell Europa
 Frigibell Blasius
 Frigibell greizdouarel
 Frigibell Méhely
 Frigibell vihan
 Frigibell vras
 Gousper Savi
 Gousperell ar Magreb
 Gousperell Capaccini
 Gousperell Geoffroy
 Gousperell varvek
 Lgodenn-dall rous Egipt
 Logodenn-dall kof noazh ar beziou
 Miniopler Schreibers
 Molos Cestoni
 Nozigell an Azorez
 Nozigell Leisler
 Nozigell vras
 Pipistrell gorr
 Pipistrell Kuhl
 Pipistrell Libia
 Pipistrell Madeira
 Pipistrell voutin
 Serotin Botta
 Serotin melenwenn
 Skouarnegell Hemprich
 Skouarnegell Tenerife

Barbastella barbastellus (Schreber, 1774)
Rhinolophus blasii Peters, 1867
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus ferrumequinum (Schreber, 1774)
Hypsugo savii (Bonaparte, 1837)
Myotis punicus Felten, 1977
Myotis capaccinii (Bonaparte, 1837)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis mystacinus (Kuhl, 1817)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Taphozous nudiventris Cretzschmar, 1830
Miniopterus schreibersii (Kuhl, 1817)
Tadarida teniotis (Rafinesque, 1814)
Nyctalus azoreum (Thomas, 1901)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus lasiopterus (Schreber, 1780)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Otonycteris hemprichii Peters, 1859
Plecotus teneriffae Barrett-Hamilton, 1907

Bulgarian

Азорски вечерник
 Булдогов прилеп
 Голокореман прилеп
 Голям вечерник
 Голям подковонос
 Дългопръст нощник
 Египетски плодояден прилеп
 Изабелинов прилеп
 Канарски дългоух прилеп
 Кафяво прилепче
 Лайслеров (малък) вечерник
 Магребски нощник
 Мадейрско прилепче
 Малко кафяво прилепче
 Малък подковонос
 Мустакат нощник

Nyctalus azoreum (Thomas, 1901)
Tadarida teniotis (Rafinesque, 1814)
Taphozous nudiventris Cretzschmar, 1830
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)
Myotis capaccinii (Bonaparte, 1837)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Plecotus teneriffae Barrett-Hamilton, 1907
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Nyctalus leisleri (Kuhl, 1817)
Myotis punicus Felten, 1977
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus hipposideros (Bechstein, 1800)
Myotis mystacinus (Kuhl, 1817)

Пещерен дългокрил
Подковонос на Мехели
Полунощен прилеп на Бот
Прилепче на Ханак
Пустинен дългоух прилеп
Савиево прилепче
Средиземноморски подковонос
Средиземноморско прилепче
Трицветен нощник
Широкоух прилеп
Южен подковонос

Miniopterus schreibersii (Kuhl, 1817)
Rhinolophus mehelyi Matschie, 1901
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Otonycteris hemprichii Peters, 1859
Hypsugo savii (Bonaparte, 1837)
Rhinolophus blasii Peters, 1867
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Barbastella barbastellus (Schreber, 1774)
Rhinolophus euryale Blasius, 1853

Castilian (Spain)

Barbastella
Cabeza Martillo
Murciélago
Murciélago Amarillo
Murciélago bigotudo
Murciélago común
Murciélago dalmata de herradura
Murciélago de Ala Larga
Murciélago de borde claro
Murciélago de bosque
Murciélago de cabeza Plana
Murciélago de Cabrera
Murciélago de Cara Hueca
Murciélago de cola libre y vientre desnudo
Murciélago de cueva
Murciélago de Geoffroy
Murciélago de Hanak
Murciélago de Herradura
Murciélago de Herradura
Murciélago de herradura de Blasius
Murciélago de herradura mediterráneo
Murciélago de la fruta egipcio
Murciélago de las Tumbas
Murciélago de Lengua Larga
Murciélago de Madeira
Murciélago de montaña
Murciélago de Nariz de Hoja
Murciélago de Orejas de Ratón
Murciélago enan
Murciélago Enano
Murciélago enano
Murciélago grande de herradura
Murciélago Guanero
Murciélago hortelano de Botta
Murciélago hortelano mediterráneo
Murciélago hortelano meridional
Murciélago hortelano pardo
Murciélago Mariposa
Murciélago mediano de herradura
Murciélago mediterráneo de herradura
Murciélago montaño
Murciélago nocturno
Murciélago orejirroto
Murciélago orejudo canario
Murciélago patudo
Murciélago pequeño de herradura
Murciélago pigmeo
Murciélago rabudo
Murciélago ratonero bigotudo
Murciélago ratonero moruno
Murciélago ratonero pardo
Murciélago ratonero patudo
Murciélago ratonero Zenate

Murciélagos
Murciélagos chicos
Murciélagos cola de raton o de cola libre
Murciélagos de cola envainada
Murciélagos de cola envainada
Murciélago de cueva magrebí

Barbastella barbastellus (Schreber, 1774)
Hypsignathus monstrosus H. Allen, 1862
Pseudoromicia tenuipinnis (Peters, 1872)
Scotophilus nigrita (Schreber, 1774)
Myotis mystacinus (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rhinolophus blasii Peters, 1867
Miniopterus inflatus Thomas, 1903
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Mimetillus moloneyi (Thomas, 1891)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Nycteris arge Thomas, 1903
Taphozous nudiventris Cretzschmar, 1830
Miniopterus schreibersii (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Rhinolophus alcyone Temminck, 1853
Rhinolophus landeri Martin, 1838
Rhinolophus blasii Peters, 1867
Rhinolophus euryale Blasius, 1853
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Megaloglossus woermanni Pagenstecher, 1885
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Hypsugo savii (Bonaparte, 1837)
Hipposideros beatus (K. Andersen, 1906)
Myotis bocagii (Peters, 1870)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Epomophorus pusillus Peters, 1868
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Glauconycteris beatrix Thomas, 1901
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Hypsugo savii (Bonaparte, 1837)
Nyctalus leisleri (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Plecotus teneriffae Barrett-Hamilton, 1907
Myotis capaccinii (Bonaparte, 1837)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Tadarida teniotis (Rafinesque, 1814)
Myotis mystacinus (Kuhl, 1817)
Myotis punicus Felten, 1977
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis capaccinii (Bonaparte, 1837)
Myotis zenatus Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019
CHIROPTERA Blumenbach, 1779
VESPERTILIONIDAE Gray, 1821
MOLOSSIDAE Gervais, 1856
EMBALLONURIDAE Gervais, 1855
VESPERTILIONIDAE Gray, 1821
Miniopterus maghrebiensis Puechmaille, Allegrini, Benda, Bilgin, Ibáñez and Juste, 2014

Nóctulo de Azores
 Noctulo de Leisler
 Nóctulo gigante
 Nóctulo grande
 Nóctulo menor
 Nóctulo pequeno
 Orejudo canario
 Orejudo de Hemprich
 Pipistrello
 Pipistrello
 Pipistrello Enano
 Rinolofo de Blasius
 Rinolofo egipcio
 Rinolofo grande de herradura
 Rinolofo mediano de herradura
 Rinolofo mediterraneo de herradura
 Rinolofo pequeno de herradura
 Ruseta de Angola
 Ruseta Egipcia
 Zorra Voladora
 Zorra Voladora Amarilla
 Zorra Voladora de Collar
 Zorra Voladora de Zenker
 Zorro volador de Aldabra
 Zorro volador de la isla Rodrigues
 Zorro volador de las Seychelles
 Zorro Volador De Livingston
 Zorro volador de Madagascar
 Zorro volador de Voeltzkow
 Zorro volador negro de Mauricio
 Zorro volador oscuro de Mauricio

Nyctalus azoreum (Thomas, 1901)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859
Pipistrellus (Pipistrellus) nanulus Thomas, 1904
Nycticeinops crassulus (Thomas, 1904)
Afronycteris nana (Peters, 1852)
Rhinolophus blasii Peters, 1867
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Rhinolophus hipposideros (Bechstein, 1800)
Myonycteris angolensis (Bocage, 1898)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Epomophorus wahlbergi (Sundevall, 1846)
Eidolon helvum (Kerr, 1792)
Myonycteris torquata (Dobson, 1878)
Scotonycteris zenkeri Matschie, 1894
Pteropus aldabrensis True, 1893
Pteropus rodricensis Dobson, 1878
Pteropus seychellensis A. Milne-Edwards, 1877
Pteropus livingstonii Gray, 1866
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Pteropus voeltzkowi Matschie, 1909
Pteropus niger (Kerr, 1792)
Pteropus subniger (Kerr, 1792)

Catalan (Spain)

Nòctul de les Azores
 Nòctul gegant
 Nòctul petit
 Orellut canari
 Orellut d'Hemprich
 Pipistrel·la comuna
 Pipistrel·la de Hanak
 Pipistrel·la de Madeira
 Pipistrel·la de vores clares
 Pipistrel·la nana
 Rat penat comú
 Rat penat cuallarg
 Rat penat de bigotis
 Rat penat de bosc
 Rat penat de cova
 Rat penat de Madeira
 Rat penat de peus grans
 Rat penat de vores clares
 Rat penat gran de ferradura
 Rat penat mediterrani de ferradura
 Rat penat mitjà de ferradura
 Rat penat muntanyero
 Rat penat nòctul gran
 Rat penat nòctul petit
 Rat penat orellut canari
 Rat penat petit de ferradura
 Ratapinyada de cova
 Ratapinyada d'orella escapçada
 Ratapinyada gran de ferradura
 Ratapinyada mediterrània de ferradura
 Ratapinyada muntanyera
 Ratapinyada petita de ferradura
 Ratpenat cuallarg
 Ratpenat de bigotis
 Ratpenat de bosc
 Ratpenat de cova
 Ratpenat de ferradura de Blasius
 Ratpenat de ferradura gran
 Ratpenat de ferradura mediterrani
 Ratpenat de ferradura mitjà
 Ratpenat de ferradura petit

Nyctalus azoreum (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Tadarida teniotis (Rafinesque, 1814)
Myotis mystacinus (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Myotis capaccinii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Hypsugo savii (Bonaparte, 1837)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Plecotus teneriffae Barrett-Hamilton, 1907
Rhinolophus hipposideros (Bechstein, 1800)
Miniopterus schreibersii (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
Hypsugo savii (Bonaparte, 1837)
Rhinolophus hipposideros (Bechstein, 1800)
Tadarida teniotis (Rafinesque, 1814)
Myotis mystacinus (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)
Rhinolophus blasii Peters, 1867
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Rhinolophus hipposideros (Bechstein, 1800)

Ratpenat de peus grans
 Ratpenat de tomba de carpó nu
 Ratpenat dels graners de Botta
 Ratpenat dels graners meridional
 Ratpenat d'orelles dentades
 Ratpenat egipci
 Ratpenat muntanyenc
 Ratpenat rater africà

Myotis capaccinii (Bonaparte, 1837)
Taphozous nudiventris Cretzschmar, 1830
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Hypsugo savii (Bonaparte, 1837)
Myotis punicus Felten, 1977

Chewa (Malawi)

Karuru
 Ndemia

Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Afronycteris nana (Peters, 1852)

Chinese

东非伏翼
 中非小伏翼
 串耳蹄蝠
 乌干达菊头蝠
 任氏伏翼
 佐氏菊头蝠
 侏凹脸蝠
 侏犬吻蝠
 侏长翼蝠
 侏颈囊果蝠
 兰德菊头蝠
 利文斯顿黄蝠
 北非果蝠
 北非犬吻
 南非伏翼
 南非墓蝠
 南非鼠耳蝠
 史密斯彩蝠
 吕氏伏翼
 哈氏巨犬吻蝠
 喀麦隆蹄蝠
 喀麦隆黄蝠
 垂耳犬吻蝠
 埃及墓蝠
 大白纹蹄蝠
 大眼蹄蝠
 大耳凹脸蝠
 大耳犬吻蝠
 大耳犬吻蝠
 大长翼蝠
 孤颌果蝠
 安哥拉犬吻蝠
 安哥拉颌果蝠
 安氏凹脸蝠
 安氏犬吻蝠
 安氏黄蝠
 宽首伏翼
 小彩蝠
 小犬吻蝠
 小长翼蝠
 小颈囊果蝠
 岛犬吻蝠
 巨蹄蝠
 布什维尔德菊头蝠
 希氏菊头蝠
 彩蝶蝠
 彼德犬吻蝠
 德氏菊头蝠
 斑犬吻蝠
 施氏墓蝠
 无畏犬吻蝠

Pipistrellus (Pipistrellus) aero Heller, 1912
Pipistrellus (Pipistrellus) nanulus Thomas, 1904
Hipposideros megalotis (Heuglin, 1861)
Rhinolophus eloquens K. Andersen, 1905
Pseudoromicia rendalli (Thomas, 1889)
Rhinolophus clivosus Cretzschmar, 1828
Nycteris nana (K. Andersen, 1912)
Mops (Xiphonycteris) nanulus J.A. Allen, 1917
Miniopterus minor Peters, 1867
Epomophorus minimus Claessen & De Vree, 1991
Rhinolophus landeri Martin, 1838
Scotophilus livingstonii Brooks and Bickham, 2014
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Laephotis capensis (A. Smith, 1829)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Myotis tricolor (Temminck, 1832)
Kerivoula smithii Thomas, 1880
Vansonia rueppellii (J.B. Fischer, 1829)
Otomops harrisoni Ralph, Richards, Taylor, Napier and Lamb, 2015
Doryrhina camerunensis (Eisentraut, 1956)
Scotophilus nux Thomas, 1904
Mops (Chaerephon) major (Trouessart, 1897)
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Macronycteris vittatus (Peters, 1852)
Doryrhina cyclops (Temminck, 1853)
Nycteris macrotis Dobson, 1876
Otomops martiensseni (Matschie, 1897)
Tadarida lobata (Thomas, 1891)
Miniopterus inflatus Thomas, 1903
Myonycteris relicta Bergmans, 1980
Mops (Mops) condylurus (A. Smith, 1833)
Myonycteris angolensis (Bocage, 1898)
Nycteris aurita (K. Andersen, 1912)
Mops (Chaerephon) ansorgei (Thomas, 1913)
Scotophilus andrewreborii Brooks and Bickham, 2014
Nycticeinops crassulus (Thomas, 1904)
Kerivoula lanosa (A. Smith, 1847)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Miniopterus fraterculus Thomas and Schwann, 1906
Epomophorus labiatus (Temminck, 1837)
Tadarida fulminans (Thomas, 1903)
Macronycteris gigas (Wagner, 1845)
Rhinolophus simulator K. Andersen, 1904
Rhinolophus hildebrandtii Peters, 1878
Glauconycteris variegata (Tomes, 1861)
Platymops setiger (Peters, 1878)
Rhinolophus deckenii Peters, 1868
Mops (Chaerephon) bivittatus (Heuglin, 1861)
Nycticeinops schlieffenii (Peters, 1859)
Mops (Xiphonycteris) thersites (Thomas, 1903)

暗翼宽吻蝠
 暗黑伏翼
 松氏蹄蝠
 查平犬吻蝠
 桑布鲁伏
 棕红鼠耳蝠
 淡翼宽吻
 淡色凹脸蝠
 温氏长耳蝠
 特鲁黄蝠
 狭齿果蝠
 珀氏三叉蝠
 白翼伏翼
 白腹黄♀
 短翼犬吻蝠
 祖鲁伏翼
 米达犬吻蝠
 粗毛凹脸蝠
 索马里伏翼
 纳塔尔长翼蝠
 肯尼亚墓蝠
 肯尼亚蝶蝠
 腺尾犬吻蝠
 花蝶蝠
 苏丹墓蝠
 莫桑比克长翼蝠

裸腹墓蝠
 诺氏蹄蝠
 贝尔墓蝠
 赤犬吻蝠
 达马拉菊头蝠
 银彩蝠
 银蝶蝠
 锈色伏翼
 锤头果蝠
 长尾棕蝠
 非洲三叉蝠
 非洲假吸血蝠
 非洲凹脸蝠
 非洲大犬吻蝠
 非洲大黄蝠
 非洲小狐蝠
 非洲扁颅蝠
 非洲长翼蝠
 非洲鞘尾蝠
 韦氏颈囊果蝠
 香蕉伏翼
 魁凹脸蝠
 魏氏鼠耳蝠
 麦氏鼠尾蝠
 黄伏翼
 黄毛果蝠
 黄翼蝠

Scotoecus hirundo (de Winton, 1899)
Pipistrellus (Pipistrellus) hesperidus (Temminck, 1840)
Hipposideros caffer (Sundevall, 1846)
Mops (Chaerephon) chapini (J.A. Allen, 1917)
Afronycteris helios (Heller, 1912)
Myotis bocagii (Peters, 1870)
Scotoecus albofuscus (Thomas, 1890)
Nycteris arge Thomas, 1903
Laephotis wintoni Thomas, 1901
Scotophilus trujilloi Brooks and Bickham, 2014
Stenonycteris lanosus (Thomas, 1906)
Cloeotis percivali Thomas, 1901
Pseudoromicia tenuipinnis (Peters, 1872)
Scotophilus leucogaster (Cretzschmar, 1826)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Neoromicia zuluensis (Roberts, 1924)
Mops (Mops) midas (Sundevall, 1843)
Nycteris hispida (Schreber, 1774)
Neoromicia somalica (Thomas, 1901)
Miniopterus natalensis (A. Smith, 1833)
Taphozous hildegardeae Thomas, 1909
Glauconycteris kenyacola Peterson, 1982
Mops (Chaerephon) bemmeleni (Jentink, 1879)
Glauconycteris humeralis J.A. Allen, 1917
Taphozous hamiltoni Thomas, 1920
Miniopterus mossambicus Monadjem, Goodman, Stanley and Appleton, 2013
Taphozous nudiventris Cretzschmar, 1830
Hipposideros ruber (Noack, 1893)
Saccolaimus peli (Temminck, 1853)
Mops (Chaerephon) russatus (J.A. Allen, 1917)
Rhinolophus fumigatus Rüppell, 1842
Kerivoula argentata Tomes, 1861
Glauconycteris argentata (Dobson, 1875)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Hypsignathus monstrosus H. Allen, 1862
Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
Triaenops afer Peters, 1877
Cardioderma cor (Peters, 1872)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Tadarida ventralis (Heuglin, 1861)
Scotophilus nigrita (Schreber, 1774)
Epomophorus pusillus Peters, 1868
Mimetillus moloneyi (Thomas, 1891)
Miniopterus africanus Sanborn, 1936
Coleura afra (Peters, 1852)
Epomophorus wahlbergi (Sundevall, 1846)
Afronycteris nana (Peters, 1852)
Nycteris grandis Peters, 1865
Myotis welwitschii (Gray, 1866)
Rhinopoma macinnesi Hayman, 1937
Nycticeinops grandidieri (Dobson, 1876)
Eidolon helvum (Kerr, 1792)
Lavia frons (E. Geoffroy St.-Hilaire, 1810)

Comorian

ndema
 ndrema

Pteropus seychellensis A. Milne-Edwards, 1877
Pteropus seychellensis A. Milne-Edwards, 1877

Creole

Sousouri banann

Coleura seychellensis Peters, 1868

Croatian

Širokouhi šišmiš\0353\
 Širokouhi mračnjak

Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)

Šrajberov šišmiš	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Azorský večernjak	<i>Nyctalus azoreum</i> (Thomas, 1901)
Bjelorubi šišmiš	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Blazijev potkovnjak	<i>Rhinolophus blasii</i> Peters, 1867
Bottin nočník	<i>Eptesicus (Cnephæus) bottae</i> (Peters, 1869)
Brkati šišmiš	<i>Myotis mystacinus</i> (Kuhl, 1817)
Dugokřilý pršňák	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Dugonogi šišmiš	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Dugoprstí šišmiš	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Egipatský plodojed šišmiš	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Golotrbi grobni šišmiš	<i>Taphozous nudiventris</i> Cretzschmar, 1830
Hanakov patuljasti šišmiš	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hemprichov dugouhi šišmiš	<i>Otonycteris hemprichii</i> Peters, 1859
Izabelin nočník	<i>Eptesicus (Cnephæus) isabellinus</i> (Temminck, 1840)
Južní potkovnjak	<i>Rhinolophus euryale</i> Blasius, 1853
Kanarský dugoušan	<i>Plecotus teneriffae</i> Barrett-Hamilton, 1907
Kolombatovicev dugoušan	<i>Plecotus kolombatovici</i> Dulic, 1980
Madeirský malý šišmiš	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Mali šišmiš	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Mali brkati šišmiš	<i>Myotis mystacinus</i> (Kuhl, 1817)
Mali potkovasti šišmiš	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Mali potkovnjak	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Mali večernjak	<i>Nyctalus leisleri</i> (Kuhl, 1817)
Mediteranský potkovasti šišmiš	<i>Rhinolophus euryale</i> Blasius, 1853
Meheljev potkovnjak	<i>Rhinolophus mehelyi</i> Matschie, 1901
Močvarní patuljasti šišmiš	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Patuljasti šišmiš	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Primorský šišmiš	<i>Hypsugo savii</i> (Bonaparte, 1837)
Ridi šišmiš	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Sjevernoafrický oštrouhi šišmiš	<i>Myotis punicus</i> Felten, 1977
Sredozemní slobodnorepac	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Trobojní šišmiš	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Veliký potkovasti šišmiš	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Veliký potkovnjak	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Veliký večernjak	<i>Nyctalus lasiopterus</i> (Schreber, 1780)

Czech

šerowec senegalský	<i>Nycteris hispida</i> (Schreber, 1774)
šerowec tébský	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
šerowecové	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795
širokouší morousi	<i>Tadarida</i> Rafinesque, 1814
širokouší ušanové	<i>Barbastella</i> Gray, 1821
žlázokřídlec Lesueurův	<i>Cistugo lesueurii</i> Roberts, 1919
žlázokřídlec Seabrův	<i>Cistugo seabrae</i> Thomas, 1912
žlázokřídlecové	<i>Cistugo</i> Thomas, 1912
žlázokřídlecovití	<i>CISTUGONIDAE</i> Lack, Roehrs, Stanley, Ruedi and Van den Bussche, 2010
žlutokřídlní lyronosi	<i>Lavia</i> Gray, 1838
banánovní netopýři	<i>Neoromicia</i> Roberts, 1926
bejložraví netopýři	<i>PTEROPODIDAE</i> Gray, 1821
buldočfí pochvovrepi	<i>Coleura</i> Peters, 1867
cíponos africký	<i>Triaenops afer</i> Peters, 1877
cíponos aldabranský	<i>Paratriaenops pauliani</i> (Goodman and Ranivo, 2008)
cíponos jihoafrický	<i>Cloeotis percivali</i> Thomas, 1901
cíponos madagaskarský	<i>Triaenops menamena</i> Goodman and Ranivo, 2009
cíponos malý	<i>Paratriaenops furculus</i> (Trouessart, 1907)
cíponos zlatý	<i>Paratriaenops auritus</i> (G. Grandidier, 1912)
dasíci	<i>Barbastella</i> Gray, 1821
dasík arabský (netopýr skalní)	<i>Barbastella leucomelas</i> (Cretzschmar, 1826)
de Wintonovi netopýři	<i>Laephotis</i> Thomas, 1901
dlouhohazyční kaloni	<i>Megaloglossus</i> Pagenstecher, 1885
dutonosec	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
dutonosecové	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795
egyptští kaloni	<i>Rousettus</i> Gray, 1821
embalonurovití	<i>EMBALLONURIDAE</i> Gervais, 1855
Grayovi kaloni	<i>Epomops</i> Gray, 1866
hadozubí kaloni	<i>Scotonycteris</i> Matschie, 1894
hrobkovec egyptský	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
hrobkovec guinejský	<i>Saccolaimus peli</i> (Temminck, 1853)
Hrobkovec lysobřichý	<i>Taphozous nudiventris</i> Cretzschmar, 1830
hrobkovec mauricijský	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
hrobkovec východoafrický	<i>Taphozous hamiltoni</i> Thomas, 1920
hrobkovec východoafrický	<i>Taphozous hildegardeae</i> Thomas, 1909
Hroznohled pyskatý	<i>Tadarida teniotis</i> (Rafinesque, 1814)

- hroznohledové
 ještěří morousi
 jihoafričtí cíponosi
 kaloň štítohlavý
 kaloň aldabránský
 kaloň angolský
 kaloň Ansellův
 kaloň Büttikoferův
 kaloň dlouhojazyčný
 kaloň Dobsonův
 Kaloň egyptský
 kaloň Franquetův
 kaloň Grandidierův
 kaloň habešský
 kaloň hadozubý
 kaloň Haymannův
 kaloň kladivohlavý
 kaloň Kockův
 kaloň komorský
 kaloň krátkonebý
 kaloň límcový
 kaloň měkkosrstý
 kaloň madagaskarský
 kaloň malgašský
 kaloň maskarénský
 kaloň mauricijský
 kaloň menší
 kaloň mosambický
 kaloň nigerijský
 Kaloň nilský
 kaloň obojkový
 kaloň palmový
 kaloň pembánský
 kaloň pemsbský
 kaloň plavý
 kaloň plavý
 kaloň pyložravý
 kaloň rodrigueský
 kaloň Sanbornův
 kaloň seychelský
 kaloň sudánský
 kaloň tomášský
 kaloň úzkokřídý
 kaloň východoafrický
 kaloň výložkový
 kaloň větší
 kaloň Veldkampův
 kaloň vlnatý
 kaloň Wahlbergův
 kaloň zanzibarský
 kaloň Zenkerův
 kaloň zlatý
 kaloňotvaří

 kaloňovci
 kaloňovití
 kaloni
 kerivula africká
 kerivuly
 kladivohlaví kaloni
 klaponos egyptský
 klaponosové
 kožnatcovití
 krátkonebí kaloni
 krysoví morousi
 lesní netopýři (lesní létači)
 létací lišky
 létací psi
 letaunové
 létavci
 létavcovití
 létavec Aellenův

 létavec Glenův
 létavec jihoafrický
 létavec komorský

Tadarida Rafinesque, 1814
Sauromys Roberts, 1917
Cloeotis Thomas, 1901
Hypsignathus monstrosus H. Allen, 1862
Pteropus aldabrensis True, 1893
Epomophorus angolensis Gray, 1870
Epomophorus anelli Bergmans and Van Strien, 2004
Epomops buettikoferi (Matschie, 1899)
Megaloglossus woermanni Pagenstecher, 1885
Epomophorus dobsonii Bocage, 1889
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Epomops franqueti (Tomes, 1860)
Rousettus madagascariensis G. Grandidier, 1929
Epomophorus minimus Claessen & De Vree, 1991
Casinycteris ophiodon (Pohle, 1943)
Epomophorus intermedius (Hayman, 1963)
Hypsignathus monstrosus H. Allen, 1862
Rousettus obliviosus Kock, 1978
Pteropus livingstonii Gray, 1866
Casinycteris argynnis Thomas, 1910
Myonycteris torquata (Dobson, 1878)
Myonycteris angolensis (Bocage, 1898)
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Eidolon dupreanum (Schegel, 1867)
Pteropus subniger (Kerr, 1792)
Pteropus niger (Kerr, 1792)
Epomophorus minor Dobson, 1880
Epomophorus crypturus Peters, 1852
Epomophorus pusillus Peters, 1868
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pteropus subniger (Kerr, 1792)
Eidolon helvum (Kerr, 1792)
Pteropus voeltzkowi Matschie, 1909
Pteropus voeltzkowi Matschie, 1909
Eidolon helvum (Kerr, 1792)
Epomops franqueti (Tomes, 1860)
Plerotes anchietae (Seabra, 1900)
Pteropus rodricensis Dobson, 1878
Epomophorus grandis (Sanborn, 1950)
Pteropus seychellensis A. Milne-Edwards, 1877
Epomophorus labiatus (Temminck, 1837)
Myonycteris brachycephala (Bocage, 1889)
Eidolon helvum (Kerr, 1792)
Myonycteris relicta Bergmans, 1980
Epomophorus gambianus (Ogilby, 1835)
Epomophorus gambianus (Ogilby, 1835)
Nanonycteris veldkampii (Jentink, 1888)
Stenonycteris lanosus (Thomas, 1906)
Epomophorus wahlbergi (Sundevall, 1846)
Epomophorus minor Dobson, 1880
Scotonycteris zenkeri Matschie, 1894
Pteropus rodricensis Dobson, 1878
 PTEROPODIFORMI Van Cakenberghe, Kearney and Seamark, 2007
 PTEROPODOIDEA Gray, 1821
 PTEROPODIDAE Gray, 1821
 PTEROPODOIDEA Gray, 1821
Kerivoula africana Dobson, 1878
Kerivoulinae Miller, 1907
Hypsignathus H. Allen, 1862
Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma E. Geoffroy St.-Hilaire, 1818
 MEGADERMATIDAE H. Allen, 1864
Casinycteris Thomas, 1910
Myopterus E. Geoffroy St.-Hilaire, 1818
Nyctalus Bowdich, 1825
 PTEROPODIDAE Gray, 1821
 PTEROPODIDAE Gray, 1821
 CHIROPTERA Blumenbach, 1779
Miniopterus Bonaparte, 1837
 MINIOPTERIDAE Dobson, 1875
Miniopterus aelleni Goodman, Maminirina, Weyeneth, Bradman, Christidis, Ruedi and Appleton, 2009
Miniopterus gleni Peterson, Eger and Mitchell, 1995
Miniopterus fraterculus Thomas and Schwann, 1906
Miniopterus griveaudi Harrison, 1959

létavec maghrebský

létavec manavijský

létavec natalský

létavec nejmenší

létavec Petersonův

létavec pískový

létavec sesterský

Létavec stěhovavý

létavec tomášský

létavec východoafrický

létavec východomadagaskarský

létavec větší

Letouň lesní

Letouň zakrpený

letouni či netopýři

letouni lysoblaní

límcoví kaloni

listonos africký

listonos trojzubcový

listonosovití

lyronos žlutokřídý

lyronos srdcový

lyronosovití

měkkosrstí kaloni

malý vrápenec podkovní

malí cíponosi

malí morousi

megaderma žlutokřídá

megaderma africká

megadermovití

Moloneyovi netopýři

morous širokouchý

morous africký

morous angolský

morous Ansorgeův

morous bělobřichý

morous běloskrvný

morous chocholatý

morous egyptský

Morous evropský

morous guinejský

morous ještěří

morous jižní

morous kamerunský

morous kivuský

morous konžský

morous krátkokřídý

morous krysový

morous liberijský

morous madagaskarský

morous malý

morous malgašský

morous Martienssenův

morous mauricijský

morous mongallanský

morous niangaranský

morous nigerijský

morous nubijský

morous oslí

morous Petersův

morous plochý

morous réunijský

morous sahelský

morous seychelský

morous skvrnitý

morous sněhobílý

morous spílavý

morous Stanleyův

morous tomášský

morous Trevorův

morous trpasličí

Miniopterus maghrebensis Puechmaille, Allegrini,

Benda, Bilgin, Ibañez and Juste, 2014

Miniopterus manavi Thomas, 1906

Miniopterus natalensis (A. Smith, 1833)

Miniopterus minor Peters, 1867

Miniopterus petersoni Goodman, Bradman, Maminirina,

Ryan, Christidis & Appleton, 2008

Miniopterus arenarius Heller, 1912

Miniopterus sororculus Goodman, Ryan, Maminirina,

Fahr, Christidis and Appleton, 2007

Miniopterus schreibersii (Kuhl, 1817)

Miniopterus newtoni Bocage, 1889

Miniopterus africanus Sanborn, 1936

Miniopterus majori Thomas, 1906

Miniopterus inflatus Thomas, 1903

Nyctalus leisleri (Kuhl, 1817)

Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)

CHIROPTERA Blumenbach, 1779

CHIROPTERA Blumenbach, 1779

Myonycteris Matschie, 1899

Hipposideros caffer (Sundevall, 1846)

Asellia tridens (E. Geoffroy St.-Hilaire, 1813)

HIPPOSIDERIDAE Lydekker, 1891

Lavia frons (E. Geoffroy St.-Hilaire, 1810)

Cardioderma cor (Peters, 1872)

MEGADERMATIDAE H. Allen, 1864

Myonycteris Matschie, 1899

Rhinolophus hipposideros (Bechstein, 1800)

Paratriaenops Benda and Vallo, 2009

Mops (Chaerephon) Dobson, 1874

Lavia frons (E. Geoffroy St.-Hilaire, 1810)

Cardioderma cor (Peters, 1872)

MEGADERMATIDAE H. Allen, 1864

Mimetillus Thomas, 1904

Tadarida lobata (Thomas, 1891)

Tadarida ventralis (Heuglin, 1861)

Mops (Mops) condylurus (A. Smith, 1833)

Mops (Chaerephon) ansorgei (Thomas, 1913)

Mops (Chaerephon) leucogaster (A. Grandidier, 1869)

Mops (Mops) leucostigma (G.M. Allen, 1918)

Mops (Chaerephon) chapini (J.A. Allen, 1917)

Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)

Tadarida teniotis (Rafinesque, 1814)

Mops (Chaerephon) russatus (J.A. Allen, 1917)

Sauromys petrophilus (Roberts, 1917)

Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)

Mops (Xiphonycteris) petersoni (El Rayah, 1981)

Mops (Chaerephon) gallagheri (Harrison, 1975)

Mops (Mops) conicus J.A. Allen, 1917

Mops (Xiphonycteris) brachypterus (Peters, 1852)

Myopterus daubentonii Desmarest, 1820

Mops (Chaerephon) bemmeleni (Jentink, 1879)

Otomops madagascariensis Dorst, 1953

Mops (Chaerephon) pumilus (Cretzschmar, 1826)

Mops (Chaerephon) jobimena (Goodman and Cardiff,

2004)

Otomops martiensseni (Matschie, 1897)

Mormopterus acetabulosus (Hermann, 1804)

Mops (Mops) demonstrator (Thomas, 1903)

Mops (Mops) niangarae J.A. Allen, 1917

Mops (Chaerephon) nigeriae (Thomas, 1913)

Mops (Chaerephon) major (Trouessart, 1897)

Mops (Mops) midas (Sundevall, 1843)

Mormopterus jugularis (Peters, 1865)

Platymops setiger (Peters, 1878)

Mormopterus francoismoutoui Goodman, Jansen Van

Vuuren, Ratrimomanarivo, Probst, Bowie, 2008

Mops (Xiphonycteris) spurrelli (Dollman, 1911)

Mops (Chaerephon) pusillus (Miller, 1902)

Mops (Chaerephon) bivittatus (Heuglin, 1861)

Mops (Mops) niveiventer Cabrera and Ruxton, 1926

Mops (Xiphonycteris) thersites (Thomas, 1903)

Mops (Xiphonycteris) bakarii Stanley, 2009

Mops (Chaerephon) tomensis (Juste and Ibañez, 1993)

Mops (Mops) trevori J.A. Allen, 1917

Mops (Xiphonycteris) nanulus J.A. Allen, 1917

morous východoafrický
 morous vévodský
 morous západoafrický
 morousovití
 motýlí netopýři
 mračník bělobřichý
 mračník Goodmanův

mračník guinejský
 mračník madagaskarský

mračník ořechový
 mračník olbřímí
 mračník réunionský
 mračník statný
 mračník zelený
 mračníkové
 mračníkové
 mrakolibové
 myzopoda ušatá

myzopodovití
 myzopody
 nedopír širokouchý
 Nedopír celouchý
 Nedopír chlupatopěstný
 Nedopír hvízdavý
 Nedopír kašelný
 nedopír walausatý
 Nedopír wraubkowaný
 netopýři

netopýři širokouší
 netopýři bradožlází
 netopýři buldočí
 netopýři dlouhorucí
 netopýři dlouhorucí
 netopýři hladkonosí
 netopýři holonosí
 netopýři morousovití
 netopýři myší
 Netopýr černý
 netopýr černý
 Netopýr červenohlavý
 netopýr šplhavý
 netopýr abonský
 Netopýr alpský
 netopýr anžuánský
 netopýr Anchietův
 netopýr angolský
 Netopýr azorský
 netopýr bělobřichý
 netopýr běloblanný
 netopýr bělohrdlý
 netopýr banánový
 netopýr bečuánský
 netopýr bibundinský
 netopýr Bocageův
 Netopýr Bottův
 Netopýr brvitý
 netopýr buldočí
 Netopýr chlupatopestrý
 netopýr damarský
 netopýr damarský
 Netopýr dlouhonožý
 netopýr dvoubarvý
 netopýr efulenský
 netopýr Flowerův
 netopýr galangánský
 netopýr gambiánský
 netopýr gargueský
 netopýr Glenův
 netopýr guinejský
 netopýr habešský
 Netopýr Hanákův
 netopýr Hindeův

Tadarida fulminans (Thomas, 1903)
Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
Myotis whiteyi (Scharff, 1900)
 MOLOSSIDAE Gervais, 1856
Glauconycteris Dobson, 1875
Scotophilus leucogaster (Cretzschmar, 1826)
Scotophilus marovaza Goodman, Ratrimomanarivo and Randrianandrianina, 2006
Scotophilus nucella Robbins, 1983
Scotophilus tandrefana Goodman, Jenkins and Ratrimomanarivo, 2005
Scotophilus nux Thomas, 1904
Scotophilus nigrita (Schreber, 1774)
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Scotophilus robustus A. Milne-Edwards, 1881
Scotophilus viridis (Peters, 1852)
Scotophilinae Van Cakenberghe and Seamark, 2008
Scotophilus Leach, 1821
Scotophilus Leach, 1821
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
 MYZOPODIDAE Thomas, 1904
Myzopoda Milne-Edwards and A. Grandidier, 1878
Miniopterus schreibersii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Myotis mystacinus (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
 VESPERTILIONIFORMACEI Van Cakenberghe, Kearney and Seamark, 2007
Barbastella Gray, 1821
Taphozous E. Geoffroy St.-Hilaire, 1818
 MOLOSSIDAE Gervais, 1856
 MINIOPTERIDAE Dobson, 1875
Miniopterus Bonaparte, 1837
 EMBALLONURIDAE Gervais, 1855
 VESPERTILIONIDAE Gray, 1821
Tadarida Rafinesque, 1814
Myotis Kaup, 1829
Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Afronycteris nana (Peters, 1852)
Glauconycteris poensis (Gray, 1842)
Hypsugo savii (Bonaparte, 1837)
Myotis anjouanensis Dorst, 1960
Neoromicia anchietae (Seabra, 1900)
Laephotis angolensis Monard, 1935
Nyctalus azoreum (Thomas, 1901)
Scotophilus leucogaster (Cretzschmar, 1826)
Pseudoromicia tenuipinnis (Peters, 1872)
Scotoecus albigula Thomas, 1909
Afronycteris nana (Peters, 1852)
Laephotis botswanae Setzer, 1971
Glauconycteris egeria Thomas, 1913
Myotis bocagii (Peters, 1870)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Coleura afra (Peters, 1852)
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Laephotis capensis (A. Smith, 1829)
Myotis capaccinii (Bonaparte, 1837)
Vansonia rueppellii (J.B. Fischer, 1829)
Pipistrellus (Pipistrellus) nanulus Thomas, 1904
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Nycticeinops grandidieri (Dobson, 1876)
Scotoecus albifuscus (Thomas, 1890)
Pipistrellus (Pipistrellus) aero Heller, 1912
Glauconycteris gleni Peterson and Smith, 1973
Nycticeinops eisentrauti (Hill, 1968)
Myotis scotti Thomas, 1927
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Scotoecus hindei Thomas, 1901

netopýr hnědý
 netopýr hotentotský
 netopýr hrobový
 Netopýr hvízdavý
 netopýr iturijský
 netopýr jižní
 Netopýr jižní
 netopýr kalabarský
 netopýr kamerunský
 Netopýr kanárský
 netopýr kapský
 netopýr keňský
 netopýr Kockův
 netopýr konžský
 netopýr koptový
 netopýr krátkouchý
 Netopýr kyrénský
 Netopýr Leislerův
 Netopýr lysobřichý
 netopýr Machadův
 netopýr madagaskarský
 Netopýr madeirský
 Netopýr makaronésý
 Netopýr malý
 netopýr malgašský
 netopýr mauricijský
 netopýr mechový
 netopýr medžeský
 netopýr Moloneyův
 Netopýr mopsličí
 netopýr Morrisův
 netopýr motýlí
 netopýr namibský
 Netopýr nejmenší
 netopýr nigerijský
 Netopýr obrovský
 netopýr obtloustlý
 Netopýr Ogněvův
 Netopýr pisklavý
 netopýr pouštní
 netopýr pruhovaný
 Netopýr punský
 Netopýr pustinný
 netopýr Rendallův
 netopýr rovníkový
 netopýr Rüppellův
 netopýr savanový
 Netopýr Saviův
 netopýr Schlieffenův
 netopýr seychelský
 netopýr slujový
 netopýr slunečný
 netopýr somálský
 netopýr stříbrný
 Netopýr stromový
 netopýr tanganjický
 Netopýr tripolitánský
 netopýr trojbarvý
 netopýr vlašťovčí
 netopýr vlnatý
 Netopýr vousatý
 Netopýr vrásopyskatý
 Netopýr vroubený
 netopýr Welwitschův
 netopýr Welwitschiův
 netopýr Wintonův
 netopýr zelený
 netopýr zuluský
 Netopýra brvitého
 netopýrcové
 netopýrcové
 Netopýrec brvitý
 netopýrec vousatý
 netopýrek ušatý
 netopýrkové ušatí

Laephotis capensis (A. Smith, 1829)
Eptesicus (Cnephæus) hottentotus (A. Smith, 1833)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Glauconycteris superba Hayman, 1939
Pipistrellus (Pipistrellus) hesperidus (Temminck, 1840)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pseudoromicia brunnea (Thomas, 1880)
Pipistrellus (Pipistrellus) inexpectatus Aellen, 1959
Plecotus teneriffae Barrett-Hamilton, 1907
Laephotis capensis (A. Smith, 1829)
Glauconycteris kenyacola Peterson, 1982
Myotis dieteri M. Happold, 2005
Glauconycteris curryae Eger and Schlitter, 2001
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycticeinops schlieffenii (Peters, 1859)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Nyctalus leisleri (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Glauconycteris machadoi Hayman, 1963
Myotis goudoti (A. Smith, 1834)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Laephotis matroka (Thomas and Schwann, 1905)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Hypsugo musciculus (Thomas, 1913)
Glauconycteris humeralis J.A. Allen, 1917
Mimetillus moloneyi (Thomas, 1891)
Barbastella barbastellus (Schreber, 1774)
Myotis morrisi Hill, 1971
Glauconycteris variegata (Tomes, 1861)
Laephotis namibensis Setzer, 1971
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Eptesicus (Cnephæus) platyops (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Nycticeinops crassulus (Thomas, 1904)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Hypsugo ariel (Thomas, 1904)
Glauconycteris alboguttata J.A. Allen, 1917
Myotis punicus Felten, 1977
Otonycteris hemprichii Peters, 1859
Pseudoromicia rendalli (Thomas, 1889)
Glauconycteris beatrix Thomas, 1901
Vansonia rueppellii (J.B. Fischer, 1829)
Neoromicia guineensis (Bocage, 1889)
Hypsugo savii (Bonaparte, 1837)
Nycticeinops schlieffenii (Peters, 1859)
Coleura seychellensis Peters, 1868
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Afronycteris helios (Heller, 1912)
Neoromicia somalica (Thomas, 1901)
Glauconycteris argentata (Dobson, 1875)
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) permixtus Aellen, 1957
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Myotis tricolor (Temminck, 1832)
Scotoecus hirundo (de Winton, 1899)
Kerivoula africana Dobson, 1878
Myotis mystacinus (Kuhl, 1817)
Tadarida teniotis (Rafinesque, 1814)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Myotis welwitschii (Gray, 1866)
Myotis welwitschii (Gray, 1866)
Laephotis wintoni Thomas, 1901
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Neoromicia zuluensis (Roberts, 1924)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotinae Tate, 1942
Myotis Kaup, 1829
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis mystacinus (Kuhl, 1817)
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
 MYZOPODIDAE Thomas, 1904

netopýrotvaří

netopýrov ci
 netopýrovití
 noční kaloni
 nosalec malolupenný
 nosalecové
 nycteris velká
 nykteridovití
 nykteridy
 nykteris egyptská
 nykteris velkouchá
 obecný
 ouška myší
 příšerec egipecký
 příšerecové
 přísavkovci
 přísavkovcovití
 přísavkovec bělobřichý

 přísavkovec ušatý

 pavrápenec abánský
 pavrápenec Aellenův
 pavrápenec africký
 pavrápenec Commersonův

pavrápenec eritrejský
 pavrápenec guinejský
 pavrápenec Jonesův
 pavrápenec kamerunský
 pavrápenec krátkoocasý
 pavrápenec Kyklop
 pavrápenec natalský
 pavrápenec nimbánský
 pavrápenec obrovský
 pavrápenec pruhovaný
 pavrápenec sahelský
 pavrápenec sazový
 pavrápenec trojzubcový
 pavrápenec ušatý
 Petersovi morousi
 plaví kaloni
 pletušánové
 ploší morousi
 pochvorep buldočí
 pochvorep madagaskarský

pochvorep seychelský
 pochvorepovití
 podkováček
 podkováčkové
 podkowáček
 podvečerníkové
 praví hrobkovci
 praví kaloni
 praví morousi
 praví netopýři
 praví pavrápenci
 praví ušani
 praví vlnouškové
 psi noční
 Psohubec myšový
 ptákossavci
 pustinní ušani
 pyložraví kaloni
 rýhonos štětinatý
 rýhonos drobný
 rýhonos egyptský
 rýhonos guinejský
 rýhonos jihoafrický
 rýhonos kamerunský
 rýhonos mosambický
 rýhonos prostřední
 rýhonos somálský

VESPERTILIONIFORMACEI Van Cakenberghe,
 Kearney and Seamark, 2007
 VESPERTILIONOIDEA Gray, 1821
 VESPERTILIONIDAE Gray, 1821
 Rousettus Gray, 1821
 Rhinopoma microphyllum (Brünnich, 1782)
 Rhinopoma E. Geoffroy St.-Hilaire, 1818
 Nycteris grandis Peters, 1865
 NYCTERIDAE Van der Hoeven, 1855
 Nycteris G. Cuvier and E. Geoffroy, 1795
 Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
 Nycteris macrotis Dobson, 1876
 Rhinolophus ferrumequinum (Schreber, 1774)
 Myotis Kaup, 1829
 Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
 Mops (Chaerephon) Dobson, 1874
 Myzopoda Milne-Edwards and A. Grandidier, 1878
 MYZOPODIDAE Thomas, 1904
 Myzopoda schliemanni Goodman, Rakotondraparany
 and Kofoky, 2007
 Myzopoda aurita Milne-Edwards and A. Grandidier,
 1878
 HIPPOSIDERIDAE Lydekker, 1891
 Hipposideros abae J.A. Allen, 1917
 Hipposideros marisae Aellen, 1954
 Hipposideros caffer (Sundevall, 1846)
 Macronycteris commersoni (E. Geoffroy St.-Hilaire,
 1813)
 Asellia patrizii de Beaux, 1931
 Hipposideros beatus (K. Andersen, 1906)
 Hipposideros jonesi Hayman, 1947
 Doryrhina camerunensis (Eisentraut, 1956)
 Hipposideros curtus G.M. Allen, 1921
 Doryrhina cyclops (Temminck, 1853)
 Hipposideros caffer (Sundevall, 1846)
 Hipposideros lamottei Brosset, 1985
 Macronycteris gigas (Wagner, 1845)
 Macronycteris vittatus (Peters, 1852)
 Hipposideros tephurus (Cabrera, 1906)
 Hipposideros fuliginosus (Temminck, 1853)
 Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
 Hipposideros megalotis (Heuglin, 1861)
 Mormopterus Peters, 1865
 Eidolon Rafinesque, 1815
 Plecotus E. Geoffroy St.-Hilaire, 1818
 Platymops Thomas, 1906
 Coleura afra (Peters, 1852)
 Paremballonura tiavato (Goodman, Cardiff, Ranivo,
 Russell and Yoder, 2006)
 Coleura seychellensis Peters, 1868
 EMBALLONURIDAE Gervais, 1855
 Rhinolophus ferrumequinum (Schreber, 1774)
 Rhinolophus Lacépède, 1799
 Rhinolophus ferrumequinum (Schreber, 1774)
 Eptesicus Rafinesque, 1820
 Taphozous E. Geoffroy St.-Hilaire, 1818
 Pteropus Brisson, 1762
 Molossinae Gervais, 1856
 Vespertilioninae Gray, 1821
 Hipposideros Gray, 1831
 Plecotus E. Geoffroy St.-Hilaire, 1818
 Kerivoula Gray, 1842
 Rousettus Gray, 1821
 Tadarida teniotis (Rafinesque, 1814)
 CHIROPTERA Blumenbach, 1779
 Otonycteris Peters, 1859
 Plerotes K. Andersen, 1910
 Nycteris hispida (Schreber, 1774)
 Nycteris nana (K. Andersen, 1912)
 Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
 Nycteris arge Thomas, 1903
 Nycteris woodi K. Andersen, 1914
 Nycteris major (K. Andersen, 1912)
 Nycteris vinsoni Dalquest, 1965
 Nycteris intermedia Aellen, 1959
 Nycteris parisii (de Beaux, 1924)

- rýhonos východoafrický
 rýhonos velký
 rýhonos velkouchý
 rýhonos západoafrický
 rýhonosi
 rýhonosovití
 saukušan
 saukušanové
 Schlieffenovi netopýři
 skalní netopýři
 spaka krysowá
 spaky
 srdcoví lyronosi
 Srostlouch
 srostlouchové
 Stejnoušan zejkatý
 strostlouchové
 tadarida africká
 Tadarida evropská
 tadarida jižní
 tadarida malá
 tadaridovití
 trojzubcoví pavrápenci
 Ušan širokouchý
 ušan balkánský
 ušan berberský
 ušan egyptský
 ušan habešský
 Ušan kanárský
 ušan pustinný
 ušanové
 upír černolící
 upír čerwenokrký
 upír bledý
 Upír egypcecký
 upír obecný
 upírové
 upírové
 upírové
 výložkoví kaloni
 vakoví hrobkovci
 vampýrovci
 večerní netopýři
 velcí cíponosi
 velcí morousi
 Veldkampovi kaloni
 veliký vrápenec podkovní
 velkouší morousi
 víkonos
 víkonos africký
 víkonos asijský
 víkonos egyptský
 víkonos východoafrický
 víkonos velký
 víkonosi
 víkonosovití
 vlaštovčí netopýři
 vlastní vrápenci
 vlnoušek bronzový
 vlnoušek damarský
 vlnoušek guinejský
 vlnoušek habešský
 vlnoušek malý
 vlnoušek východoafrický
 vlnoušek západoafrický
 vlnouškové
 vrápenci
 vrápenci
 vrápencovci
 vrápencovití
 vrápenec
 vrápenec Adamův
 vrápenec Alcyone
 Vrápenec Blasiův
 vrápenec cíponosý
 vrápenec Darlingův
Nycteris aurita (K. Andersen, 1912)
Nycteris grandis Peters, 1865
Nycteris macrotis Dobson, 1876
Nycteris gambiensis (K. Andersen, 1912)
Nycteris G. Cuvier and E. Geoffroy, 1795
 NYCTERIDAE Van der Hoeven, 1855
Barbastella barbastellus (Schreber, 1774)
Barbastella Gray, 1821
Nycticeinops Hill & Harrison, 1987
Hypsugo Kolenati, 1856
Myopterus daubentonii Desmarest, 1820
Myopterus E. Geoffroy St.-Hilaire, 1818
Cardioderma Peters, 1873
Barbastella barbastellus (Schreber, 1774)
Plecotus E. Geoffroy St.-Hilaire, 1818
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Barbastella Gray, 1821
Tadarida ventralis (Heuglin, 1861)
Tadarida teniotis (Rafinesque, 1814)
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
 MOLOSSIDAE Gervais, 1856
Asellia Gray, 1838
Barbastella barbastellus (Schreber, 1774)
Plecotus kolombatovici Dulic, 1980
Plecotus gaisleri Benda, Kiefer, Hanak & Veith, 2004
Plecotus christii Gray, 1838
Plecotus balensis Kruskop & Lavrenchenko, 2000
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859
Plecotus E. Geoffroy St.-Hilaire, 1818
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Pteropus subniger (Kerr, 1792)
Eidolon helvum (Kerr, 1792)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pteropus subniger (Kerr, 1792)
 CHIROPTERA Blumenbach, 1779
 PTEROPODIDAE Gray, 1821
Pteropus Brisson, 1762
Epomophorus Bennett, 1836
Saccolaimus Temminck, 1838
 NOCTILIONOIDEA Gray, 1821
Eptesicus Rafinesque, 1820
Triaenops Dobson, 1871
Mops Lesson, 1842
Nanonycteris Matschie, 1899
Rhinolophus ferrumequinum (Schreber, 1774)
Otomops Thomas, 1913
Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma cystops Thomas, 1903
Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma macinnesi Hayman, 1937
Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma E. Geoffroy St.-Hilaire, 1818
 RHINOPOMATIDAE Dobson, 1872
Scotoecus Thomas, 1901
 RHINOLOPHIDAE Gray, 1825
Kerivoula cuprosa Thomas, 1912
Kerivoula argentata Toms, 1861
Kerivoula smithii Thomas, 1880
Kerivoula eriophora (Heuglin, 1877)
Kerivoula lanosa (A. Smith, 1847)
Kerivoula africana Dobson, 1878
Kerivoula phalaena Thomas, 1912
Kerivoulinae Miller, 1907
Rhinolophus Lacépède, 1799
 RHINOLOPHOIDEA J. E. Gray, 1825
 RHINOLOPHOIDEA J. E. Gray, 1825
 RHINOLOPHIDAE Gray, 1825
Rhinolophus Lacépède, 1799
Rhinolophus adami Aellen and Brosset, 1968
Rhinolophus alcyone Temminck, 1853
Rhinolophus blasii Peters, 1867
Triaenops afer Peters, 1877
Rhinolophus darlingi K. Andersen, 1905

vrápenec Dentův
vrápenec guinejský
vrápenec Hildebrandtův
vrápenec Hillův
Vrápenec jižní
vrápenec jihoafrický
vrápenec kouřový
vrápenec Landerův
vrápenec liberijský

vrápenec Macclaudův
Vrápenec malý
Vrápenec Mehelyův
vrápenec menší
vrápenec moravský
vrápenec mosambický
vrápenec pahrbeční
vrápenec pahrbeční
vrápenec podkovní
vrápenec podkovníček
vrápenec podkrovní
vrápenec pouštní
vrápenec ruwenzorijský
vrápenec sakežienský
Vrápenec středozejský
vrápenec tanganjický

vrápenec ugandský
vrápenec východoafrický
vrápenec veliký
Vrápenec velký
vrápenec západoafrický
vrápeník malý
vrápeník podkovní
vrápeníkové
vrápeníkové
vrápeníkové
večerník prowrtaný
večerník senegalský
večerníkové
weloblánek lupenatý
vrápenec menší
vrápenec pahrbeční
vrápenec podkovní
vrápenec trojzýkový
vrápenecové
zakrslí netopýři

Rhinolophus denti Thomas, 1904
Rhinolophus guineensis Eisentraut, 1960
Rhinolophus hildebrandtii Peters, 1878
Rhinolophus hilli Aellen, 1973
Rhinolophus euryale Blasius, 1853
Rhinolophus capensis Lichtenstein, 1823
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus landeri Martin, 1838
Rhinolophus ziama Fahr, Vierhaus, Hutterer and Kock, 2002
Rhinolophus macclaudi Pousargues, 1898
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus mehelyi Matschie, 1901
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus swinnyi Gough, 1908
Rhinolophus blasii Peters, 1867
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus ruwenzorii J. Eric Hill, 1942
Rhinolophus sakejiensis Cotterill, 2002
Rhinolophus euryale Blasius, 1853
Rhinolophus maendeleo Kock, Csorba and Howell, 2000
Rhinolophus eloquens K. Andersen, 1905
Rhinolophus deckenii Peters, 1868
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hillorum Koopman, 1989
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus Lacépède, 1799
Rhinolophus Lacépède, 1799
Plecotus E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Taphozous E. Geoffroy St.-Hilaire, 1818
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus ferrumequinum (Schreber, 1774)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Rhinolophus Lacépède, 1799
Pipistrellus Kaup, 1829

Danish

Azorflagermus
Blasius' hestekonæse
Bottas sydflagermus
Bredøret flagermus
Capaccinis flagermus
Dværgflagermus
Dværgflagermus
Dværghesteskonaes
Egyptisk flyvehund
Europæisk bulldogflagermus
Geoffroys flagermus
Hanaks flagermus
Isabellasydflagermus
Kanarielangøre
Kuhls flagermus
Leislers flagermus
Leisler's flagermus
Lille hestekonæse
Madeiraflagermus
Mahgreb museøre
Mehelys hestekonæse
Middelhavehestekonæse
Nøgenrumpet gravflagermus
Pipistrellflagermus
Savis flagermus

Nyctalus azoreum (Thomas, 1901)
Rhinolophus blasii Peters, 1867
Eptesicus (Cnephæus) bottae (Peters, 1869)
Barbastella barbastellus (Schreber, 1774)
Myotis capaccinii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus hipposideros (Bechstein, 1800)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Tadarida teniotis (Rafinesque, 1814)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Plecotus teneriffae Barrett-Hamilton, 1907
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Myotis punicus Felten, 1977
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Taphozous nudiventris Cretzschmar, 1830
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Hypsugo savii (Bonaparte, 1837)

Schreibers' langfingerflagermus
Skægflagermus
Stor brunflagermus
Stor hestekonæse
Storøret flagermus
Vestlig bredøret flagermus

Miniopterus schreibersii (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)
Otonycteris hemprichii Peters, 1859
Barbastella barbastellus (Schreber, 1774)

Dutch

Alpendwergvleermuis
Alpenvleermuis
Arabische mopsvleermuis
Azoren rosse vleermuis
Azorenvleermuis
Baardvleermuis
Balkangrootoorvleermuis
Bladneusvleermuis
Blasius' hoefijzerneus
Blasius-hoefijzerneus
Bosvleermuis
Botta's vleermuis
Buldogvleermuis
Bulveermuis
Cabrera's rondbladneus
Canarische grootoorvleermuis
Capaccini's vleermuis
Drietandbladneusvleermuis
Dwarsoorvleermuis
Egyptische grafvleermuis
Egyptische grootoorvleermuis
Egyptische bulvleermuis
Europese bulvleermuis
Fenicische vale vleermuis
Gaislers grootoorvleermuis
Geoffroy's hoefijzerneus
Gewone dwergvleermuis
Gewone pisang blad vleermuis
Gladneusvleermuis
Grote fruitvleermuis
Grote hoefijzerneus
Grote klapneusvleermuis
Grote rosse vleermuis
Grote Schreiber's vleermuis
Hanaks dwergvleermuis
Hanak's dwergvleermuis
Hartneusvleermuis
Hoefijzerneusvleermuis
Hoefijzerneuzen
Hoefijzerneuzen
Horaceks hoefijzerneus
Ingekorven vleermuis
Isabelvleermuis
Kaalbuikgrafvleermuis
Kalongs
Kleine dwergvleermuis
Kleine hoefijzerneus
Kleine klapneusvleermuis
Knight's Egyptische vleermuis

Kuhls dwergvleermuis
Kuhl's dwergvleermuis
Langstaartvleermuis
Langvleugelvleermuis
Langvleugelvleermuis
Madagassische hechtschijfvleermuis
Madeira dwergvleermuis
Madeiradwergvleermuis
Maghrebijs langvleugelvleermuis

Mehely-hoefijzerneus
Mehely's hoefijzerneus
Mopsvleermuis
Muisstaartvleermuis
Nijlrousette
Nijlrousetus

Hypsugo savii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
Barbastella leucomelas (Cretzschmar, 1826)
Nyctalus azoreum (Thomas, 1901)
Nyctalus azoreum (Thomas, 1901)
Myotis mystacinus (Kuhl, 1817)
Plecotus kolombatovici Dulic, 1980
HIPPOSIDERIDAE Lydekker, 1891
Rhinolophus blasii Peters, 1867
Rhinolophus blasii Peters, 1867
Nyctalus leisleri (Kuhl, 1817)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
MOLOSSIDAE Gervais, 1856
Tadarida teniotis (Rafinesque, 1814)
Hipposideros tephros (Cabrera, 1906)
Plecotus teneriffae Barrett-Hamilton, 1907
Myotis capaccinii (Bonaparte, 1837)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Barbastella barbastellus (Schreber, 1774)
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Plecotus christii Gray, 1838
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Tadarida teniotis (Rafinesque, 1814)
Myotis punicus Felten, 1977
Plecotus gaisleri Benda, Kiefer, Hanak & Veith, 2004
Rhinolophus clivosus Cretzschmar, 1828
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myotis mystacinus (Kuhl, 1817)
VESPERTILIONIDAE Gray, 1821
PTEROPODIDAE Gray, 1821
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinopoma microphyllum (Brünnich, 1782)
Nyctalus lasiopterus (Schreber, 1780)
Miniopterus schreibersii (Kuhl, 1817)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Cardioderma cor (Peters, 1872)
RHINOLOPHIDAE Gray, 1825
RHINOLOPHIDAE Gray, 1825
Rhinolophus Lacépède, 1799
Rhinolophus horaceki Benda and Vallo, 2012
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Taphozous nudiventris Cretzschmar, 1830
Pteropus Brisson, 1762
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinopoma cystops Thomas, 1903
Aegyptonycteris knightae Simmons, Seiffert and Gunnell, 2016
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
RHINOPOMATIDAE Dobson, 1872
Miniopterus schreibersii (Kuhl, 1817)
MINIOPTERIDAE Dobson, 1875
MYZOPODIDAE Thomas, 1904
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Miniopterus maghrebiensis Puechmaile, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014
Rhinolophus mehelyi Matschie, 1901
Rhinolophus mehelyi Matschie, 1901
Barbastella barbastellus (Schreber, 1774)
RHINOPOMATIDAE Dobson, 1872
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)

Onechte vampieren
 Paarse hoefijzerneus
 Pemba vliegende hond
 Rodriguez vliegende hond
 Rüppells dwergvleermuis
 Savi's dwergvleermuis
 Schedestaartvleermuis
 Schlieffens vleermuis
 Schoudervlekkalong
 Schreibers' vleermuis
 Spleetneusvleermuis
 Thebaanse spleetneusvleermuis
 Vleermuisen
 Vliegende honden
 Westelijke mopsvleermuis
 Woestijndwergvleermuis
 Woestijngrootoorvleermuis
 Zuideuropese bulvleermuis
 Zwarte Komorenkalong

MEGADERMATIDAE H. Allen, 1864
Rhinolophus euryale Blasius, 1853
Pteropus voeltzkowi Matschie, 1909
Pteropus rodricensis Dobson, 1878
Vansonia rueppellii (J.B. Fischer, 1829)
Hypsugo savii (Bonaparte, 1837)
 EMBALLONURIDAE Gervais, 1855
Nycticeinops schlieffenii (Peters, 1859)
Pteropus livingstonii Gray, 1866
Miniopterus schreibersii (Kuhl, 1817)
 NYCTERIDAE Van der Hoeven, 1855
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
 CHIROPTERA Blumenbach, 1779
 PTEROPODIDAE Gray, 1821
Barbastella barbastellus (Schreber, 1774)
Hypsugo ariel (Thomas, 1904)
Otonycteris hemprichii Peters, 1859
Tadarida teniotis (Rafinesque, 1814)
Pteropus livingstonii Gray, 1866

English

45 kHz pipistrelle
 55 kHz pipistrelle
 Aba Leaf-nosed Bat
 Aba Roundleaf Bat
 Abo Bat
 Abo Butterfly Bat
 Abruzzi's Wrinkle-lipped Bat
 Abyssinian Horseshoe Bat
 Adam's Horseshoe Bat
 Aellen's Leaf-nosed Bat
 Aellen's Long-fingered Bat
 Aellen's Pipistrelle
 Aellen's Pipistrelle Bat
 Aellen's Roundleaf Bat
 Aequatorial Broad Winged Bat
 Aerial Pipistrelle Bat
 African Epauletted bats
 African False Vampire Bat
 African False Vampire Bats
 African Free-tailed Bats
 African Giant Free-tailed Bat
 African Heart-nosed Bat
 African Long-eared Bats
 African Long-fingered Bat
 African Long-tongued Bat
 African Long-tongued Fruit-bats
 African Nectar Fruit-bats
 African pipistrelle
 African Sheath-tailed Bat
 African Straw-coloured Fruit-bat
 African Taphozous
 African Trident Bat
 African Trident Bats
 African Trident-nosed Bats
 African Woolly Bat
 African Yellow Bat
 African Yellow House Bat
 African Yellow-winged Bat
 Aldabra Flying Fox
 Aldabra Flying-fox
 Allen's Butterfly Bat
 Allen's Spotted Bat
 Allen's Striped Bat
 Aloe Bat
 Aloe Serotine
 Aloe Serotine Bat
 Anchieta's Bat
 Anchieta's Fruit Bat
 Anchieta's Pipistrelle
 Anchieta's Plerote Fruit-bat
 Andersen's Slit-faced Bat

Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Hipposideros abae J.A. Allen, 1917
Hipposideros abae J.A. Allen, 1917
Glauconycteris poensis (Gray, 1842)
Glauconycteris poensis (Gray, 1842)
Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus adami Aellen and Brosset, 1968
Hipposideros marisae Aellen, 1954
Miniopterus aelleni Goodman, Maminirina, Weyeneth, Bradman, Christidis, Ruedi and Appleton, 2009
Pipistrellus (Pipistrellus) inexpectatus Aellen, 1959
Pipistrellus (Pipistrellus) inexpectatus Aellen, 1959
Hipposideros marisae Aellen, 1954
Miniopterus arenarius Heller, 1912
Pipistrellus (Pipistrellus) aero Heller, 1912
Epomops Gray, 1866
Cardioderma cor (Peters, 1872)
Cardioderma Peters, 1873
Myotis E. Geoffroy St.-Hilaire, 1818
Tadarida ventralis (Heuglin, 1861)
Cardioderma cor (Peters, 1872)
Laephotis Thomas, 1901
Miniopterus africanus Sanborn, 1936
Megaloglossus woermanni Pagenstecher, 1885
Megaloglossus Pagenstecher, 1885
Megaloglossus Pagenstecher, 1885
Pipistrellus (Pipistrellus) hesperidus (Temminck, 1840)
Coleura afra (Peters, 1852)
Eidolon helvum (Kerr, 1792)
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Cloeotis percivali Thomas, 1901
Triaenops afer Peters, 1877
Cloeotis Thomas, 1901
Cloeotis Thomas, 1901
Kerivoula africana Dobson, 1878
Scotophilus dinganii (A. Smith, 1833)
Scotophilus dinganii (A. Smith, 1833)
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Pteropus aldabrensis True, 1893
Pteropus aldabrensis True, 1893
Glauconycteris alboguttata J.A. Allen, 1917
Glauconycteris humeralis J.A. Allen, 1917
Glauconycteris alboguttata J.A. Allen, 1917
Neoromicia zuluensis (Roberts, 1924)
Neoromicia zuluensis (Roberts, 1924)
Neoromicia zuluensis (Roberts, 1924)
Neoromicia anchietae (Seabra, 1900)
Plerotes anchietae (Seabra, 1900)
Neoromicia anchietae (Seabra, 1900)
Plerotes anchietae (Seabra, 1900)
Nycteris aurita (K. Andersen, 1912)

- Andrew Rebori's House Bat
 Angolan Epauletted Fruit Bat
 Angolan Epauletted Fruit-bat
 Angolan Free-tailed Bat
 Angolan Fruit Bat
 Angolan Hairy Bat
 Angolan Long-eared Bat
 Angolan Long-haired Fruit-bat
 Angolan Mops Bat
 Angolan Rousette
 Angolan Soft-furred Fruit-bat
 Angolan Wing-gland Bat
 Anjouan Mouse-eared Bat
 Anjouan Myotis
 Ansoerge's Free-tailed Bat
 Ansoerge's Wrinkle-lipped Bat
 Arabian barbastelle
 Arabian Horseshoe Bat
 Ariel Pipistrelle
 Asian Barbastelle
 Asiatic Barbastelle
 Azorean Noctule Bat
 Azores noctule
 Badger bat
 Bale Big-eared Bat
 Bale Long-eared Bat
 Balkan long-eared bat
 Balli Slit-faced Bat
 Banana Bat
 Banana Pipistrelle
 Banana Pipistrelle Bat
 Barbastelle
 Barbastelle bat
 Barbastelle Bats
 Barbastelles
 Bates's Slit-faced Bat
 Bats
 Beatrix Bufferfly Bat
 Beatrix's Bat
 Benguela Fruit-bat
 Benito Leaf-nosed Bat
 Benito Roundleaf Bat
 Bent-winged bat
 Bent-winged Bats
 Bergmans's Collared Fruit Bat
 Bergmans's fruit bat
 Bergmans's Small Fruit Bat
 Bibundi Bat
 Bibundi Bufferfly Bat
 Big-brown bats
 Big-eared Bats
 Big-eared Bulldog Bats
 Big-eared Free-tailed Bat
 Big-eared Free-tailed Bats
 Big-eared Guano Bat
 Big-eared Kenya Free-tailed Bat
 Big-eared Kenyan Guano Bat
 Big-eared leaf-nosed bat
 Big-eared slit-faced bat
 Bini Free-tailed Bat
 Bini Winged-mouse Bat
 Black and Red Free-tailed Bat
 Black Clinging Bat
 Black Flying- fox
 Blackish Butterfly Bat
 Blasius's Horseshoe Bat
 Blue-grey Bats
 Bocage Banana Bat
 Bocage's Free-tailed Bat
 Bocage's Fruit-bat
 Bocage's Hairy Bat
Scotophilus andrewreborii Brooks and Bickham, 2014
Epomophorus angolensis Gray, 1870
Epomophorus angolensis Gray, 1870
Mops (Mops) condylurus (A. Smith, 1833)
Myonycteris angolensis (Bocage, 1898)
Cistugo seabrae Thomas, 1912
Laephotis angolensis Monard, 1935
Myonycteris angolensis (Bocage, 1898)
Mops (Mops) condylurus (A. Smith, 1833)
Myonycteris angolensis (Bocage, 1898)
Myonycteris angolensis (Bocage, 1898)
Cistugo seabrae Thomas, 1912
Myotis anjouanensis Dorst, 1960
Myotis anjouanensis Dorst, 1960
Mops (Chaerephon) ansorgei (Thomas, 1913)
Mops (Chaerephon) ansorgei (Thomas, 1913)
Barbastella leucomelas (Cretzschmar, 1826)
Rhinolophus clivosus Cretzschmar, 1828
Hypsugo ariel (Thomas, 1904)
Barbastella leucomelas (Cretzschmar, 1826)
Barbastella leucomelas (Cretzschmar, 1826)
Nyctalus azoreum (Thomas, 1901)
Nyctalus azoreum (Thomas, 1901)
Glauconycteris superba Hayman, 1939
Plecotus balensis Kruskop & Lavrenchenko, 2000
Plecotus balensis Kruskop & Lavrenchenko, 2000
Plecotus kolombatovici Dulic, 1980
Nycteris parisii (de Beaux, 1924)
Afronycteris nana (Peters, 1852)
Afronycteris nana (Peters, 1852)
Afronycteris nana (Peters, 1852)
Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Barbastella Gray, 1821
Barbastella Gray, 1821
Nycteris arge Thomas, 1903
CHIROPTERA Blumenbach, 1779
Glauconycteris beatrix Thomas, 1901
Glauconycteris beatrix Thomas, 1901
Plerotes anchietae (Seabra, 1900)
Hipposideros beatus (K. Andersen, 1906)
Hipposideros beatus (K. Andersen, 1906)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus Bonaparte, 1837
Myonycteris relicta Bergmans, 1980
Scotonycteris bergmansi Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015
Myonycteris relicta Bergmans, 1980
Glauconycteris egeria Thomas, 1913
Glauconycteris egeria Thomas, 1913
Eptesicus Rafinesque, 1820
Plecotus E. Geoffroy St.-Hilaire, 1818
Otomops Thomas, 1913
Tadarida lobata (Thomas, 1891)
Otomops Thomas, 1913
Tadarida lobata (Thomas, 1891)
Tadarida lobata (Thomas, 1891)
Tadarida lobata (Thomas, 1891)
Hipposideros megalotis (Heuglin, 1861)
Nycteris macrotis Dobson, 1876
Myopterus whitleyi (Scharff, 1900)
Myopterus whitleyi (Scharff, 1900)
Mops (Chaerephon) jobimena (Goodman and Cardiff, 2004)
Miniopterus fraterculus Thomas and Schwann, 1906
Pteropus livingstonii Gray, 1866
Glauconycteris atra Hassanin, Colombo, Gembu, Merle, Tu, Görföl, Musaba Akawa, Csorba, Kearney, Monadjem and Ing, 2017
Rhinolophus blasii Peters, 1867
Glauconycteris Dobson, 1875
Myotis bocagii (Peters, 1870)
Tadarida aegyptiaca bocagei (Seabra, 1900)
Myonycteris angolensis (Bocage, 1898)
Myotis bocagii (Peters, 1870)

Bocage's Mouse-eared Bat	<i>Myotis bocagii</i> (Peters, 1870)
Bonneted Bats	MOLOSSIDAE Gervais, 1856
Bonneted Bats	Molossinae Gervais, 1856
Botswana Long-eared Bat	<i>Laephotis botswanae</i> Setzer, 1971
Botswanan Long-eared Bat	<i>Laephotis botswanae</i> Setzer, 1971
Botta's Serotine	<i>Eptesicus (Cnephaeus) bottae</i> (Peters, 1869)
Botta's Serotine Bat	<i>Eptesicus (Cnephaeus) bottae</i> (Peters, 1869)
Bourbon Bat	<i>Scotophilus borbonicus</i> (E. Geoffroy St.-Hilaire, 1803)
Broad-headed Pipistrelle	<i>Nycticeinops crassulus</i> (Thomas, 1904)
Broad-headed Pipistrelle Bat	<i>Nycticeinops crassulus</i> (Thomas, 1904)
Brown bats	<i>Scotophilus</i> Leach, 1821
Brown House Bat	<i>Scotophilus nigrita</i> (Schreber, 1774)
Brown Pipistrelle Bat	<i>Pseudoromicia brunnea</i> (Thomas, 1880)
Brünnich's Mouse-tailed Bat	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
Buettikofer's Clamorous Fruit-bat	<i>Epomops buettikoferi</i> (Matschie, 1899)
Buettikofer's Epauletted Bat	<i>Epomops buettikoferi</i> (Matschie, 1899)
Buettikofer's Epauletted Fruit Bat	<i>Epomops buettikoferi</i> (Matschie, 1899)
Bulldog Bats	MOLOSSIDAE Gervais, 1856
Bulldog Bats	Molossinae Gervais, 1856
Bushveld Horseshoe Bat	<i>Rhinolophus simulator</i> K. Andersen, 1904
Butterfly Bat	<i>Glauconycteris variegata</i> (Tomes, 1861)
Butterfly Bats	<i>Glauconycteris</i> Dobson, 1875
Büttikofer's Fruit Bat	<i>Epomops buettikoferi</i> (Matschie, 1899)
Cameroon Leaf-nosed Bat	<i>Doryrhina camerunensis</i> (Eisentraut, 1956)
Campo-Ma'an fruit bat	<i>Casinycteris campomaanensis</i> Hassanin, 2014
Canary big-eared bat	<i>Plecotus teneriffae</i> Barrett-Hamilton, 1907
Canary Long-eared Bat	<i>Plecotus teneriffae</i> Barrett-Hamilton, 1907
Canon Smith's Rousette	<i>Myonycteris angolensis</i> (Bocage, 1898)
Capaccinii's Mouse-eared Bat	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Cape bat	<i>Laephotis capensis</i> (A. Smith, 1829)
Cape Hairy Bat	<i>Myotis tricolor</i> (Temminck, 1832)
Cape Hairy Myotis	<i>Myotis tricolor</i> (Temminck, 1832)
Cape Horseshoe Bat	<i>Rhinolophus capensis</i> Lichtenstein, 1823
Cape House Bat	<i>Laephotis capensis</i> (A. Smith, 1829)
Cape Leaf-nosed bat	<i>Hipposideros caffer</i> (Sundevall, 1846)
Cape Long-eared Bat	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
Cape Myotis	<i>Myotis tricolor</i> (Temminck, 1832)
Cape Pipistrelle Bat	<i>Laephotis capensis</i> (A. Smith, 1829)
Cape Serotine	<i>Laephotis capensis</i> (A. Smith, 1829)
Cape Serotine Bat	<i>Laephotis capensis</i> (A. Smith, 1829)
Central Africa Pipistrelle	<i>Pipistrellus (Pipistrellus) inexpectatus</i> Aellen, 1959
Chapin's Free-tailed Bat	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
Chapin's Wrinkle-lipped Bat	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
Christie's Long-eared Bat	<i>Plecotus christii</i> Gray, 1838
Clamorous Fruit-bats	<i>Epomops</i> Gray, 1866
Cohen's Horseshoe Bat	<i>Rhinolophus cohenae</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Collared Fruit-bats	<i>Myonycteris</i> Matschie, 1899
Commerson's Leaf-nosed Bat	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Commerson's Leaf-nosed Bat	<i>Macronycteris gigas</i> (Wagner, 1845)
Commerson's Rhinoloph	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Commerson's Rhinoloph	<i>Macronycteris gigas</i> (Wagner, 1845)
Commerson's roundleaf bat	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Commerson's roundleaf bat	<i>Macronycteris gigas</i> (Wagner, 1845)
Common African Leaf-nosed Bat	<i>Hipposideros caffer</i> (Sundevall, 1846)
Common Barbastelle	<i>Barbastella barbastellus</i> (Schreber, 1774)
Common Bent-winged Bat	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Common Butterfly Bat	<i>Glauconycteris argentata</i> (Dobson, 1875)
Common Long-fingered Bat	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Common Pipistrelle	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Common Pipistrelle Bat	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Common Slit-faced Bat	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
Comores Flying Fox	<i>Pteropus livingstonii</i> Gray, 1866
Comoro Black Flying Fox	<i>Pteropus livingstonii</i> Gray, 1866
Comoro Flying Fox	<i>Pteropus livingstonii</i> Gray, 1866
Comoro Rousette	<i>Rousettus obliviosus</i> Kock, 1978
Congo Horseshoe Bat	<i>Rhinolophus adami</i> Aellen and Brosset, 1968
Copper Woolly Bat	<i>Kerivoula cuprosa</i> Thomas, 1912
Crested Free-tail Bat	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
Crested Wrinkle-lipped Bat	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
Cretzschmar's Brown Bat	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
Cretzschmar's Horseshoe bat	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
Curry's Bat	<i>Glauconycteris curryae</i> Eger and Schlitter, 2001

- Curry's Butterfly Bat
 Cyclops Bat
 Cyclops Leaf-nosed Bat
 Cyclops Roundleaf Bat
 Damara Woolly Bat
 D'Anchieta's Fruit Bat
 Dar-es-salaam Pipistrelle
 Dar-es-Salaam Pipistrelle Bat
 Dark Flying Fox
 Dark House Bat
 Dark-brown Pipistrelle Bat
 Dark-brown Serotine
 Dark-winged Lesser House Bat
 Darling's Horseshoe Bat
 Daubenton's Free-tailed Bat
 Daubenton's Winged-mouse Bat
 De Beaux's Trident Bat
 De Winton's Long-eared Bat
 Decken's Horseshoe Bat
 Dent's Horseshoe Bat
 Desert Bats
 Desert Big-eared Bats
 Desert Long-eared bat
 Desert Long-eared Bats
 Desert Pipistrelle
 Dingaan's House Bat
 Dingaan's Yellow Bat
 Dingan's Bat
 Dja Slit-faced Bat; Large Slit-faced Bat
 Dobson's Bats
 Dobson's Epauletted Bat
 Dobson's Epauletted Fruit Bat
 Dobson's Fruit Bat
 Duke of Abruzzi's Free-tailed Bat
 Duke of Abruzzi's Wrinkle-lipped Bat
 Dusky leaf-nosed bat
 dusky pipistrelle
 Dwarf Epauletted Bat
 Dwarf Epauletted Fruit Bat
 Dwarf Free-tailed Bat
 Dwarf Fruit-bats
 Dwarf Leaf-nosed Bat
 Dwarf Mops Bat
 Dwarf Pipistrelle Bat
 Dwarf Slit-faced Bat
 East African Collared Fruit-bat
 East African Epauletted Fruit Bat
 East African Epauletted Fruit-bat
 East African Flat-headed Bat
 East African Flat-headed Bats
 East African Little Collared Fruit Bat
 East African Serotine
 East African Trident Bat
 Eastern Africa Free-tailed Bat
 Eastern Africa horseshoe bat
 Eastern Barbastelle
 Eastern Barbastelle Bat
 Eastern Sucker-footed Bat
 Eger's long-fingered bat
 Egyptian Desert Pipistrelle
 Egyptian Free-tailed Bat
 Egyptian Fruit Bat
 Egyptian Guano Bat
 Egyptian Mouse-tailed bat
 Egyptian Nyctinome
 Egyptian Rousette
 Egyptian Rousette Bat
 Egyptian sheath-tailed bat
 Egyptian Slit-faced Bat
 Egyptian Tomb Bat
 Egyptian Xantharpy
 Eidolon fruit bats
 Eisentraut's Leaf-nosed Bat
Glauconycteris curryae Eger and Schlitter, 2001
Doryrhina cyclops (Temminck, 1853)
Doryrhina cyclops (Temminck, 1853)
Doryrhina cyclops (Temminck, 1853)
Kerivoula argentata Tomes, 1861
Plerotes anchietae (Seabra, 1900)
Pipistrellus (Pipistrellus) permixtus Aellen, 1957
Pipistrellus (Pipistrellus) permixtus Aellen, 1957
Pteropus subniger (Kerr, 1792)
Scotophilus nux Thomas, 1904
Pseudoromicia brunnea (Thomas, 1880)
Pseudoromicia brunnea (Thomas, 1880)
Scotoecus hirundo (de Winton, 1899)
Rhinolophus darlingi K. Andersen, 1905
Myopterus daubentonii Desmarest, 1820
Myopterus daubentonii Desmarest, 1820
Asellia italosomalica de Beaux, 1931
Laephotis wintoni Thomas, 1901
Rhinolophus deckenii Peters, 1868
Rhinolophus denti Thomas, 1904
Otonycteris Peters, 1859
Otonycteris Peters, 1859
Otonycteris hemprichii Peters, 1859
Otonycteris Peters, 1859
Hypsugo ariel (Thomas, 1904)
Scotophilus dinganii (A. Smith, 1833)
Scotophilus dinganii (A. Smith, 1833)
Scotophilus dinganii (A. Smith, 1833)
Nycteris major (K. Andersen, 1912)
Glauconycteris Dobson, 1875
Epomophorus dobsonii Bocage, 1889
Epomophorus dobsonii Bocage, 1889
Epomophorus dobsonii Bocage, 1889
Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
Hipposideros fuliginosus (Temminck, 1853)
Pipistrellus (Pipistrellus) hesperidus (Temminck, 1840)
Myonycteris Matschie, 1899
Epomophorus pusillus Peters, 1868
Mops (Xiphonycteris) nanulus J.A. Allen, 1917
Nanonycteris Matschie, 1899
Hipposideros beatus (K. Andersen, 1906)
Mops (Xiphonycteris) nanulus J.A. Allen, 1917
Pipistrellus (Pipistrellus) nanulus Thomas, 1904
Nycteris nana (K. Andersen, 1912)
Myonycteris relicta Bergmans, 1980
Epomophorus minimus Claessen & De Vree, 1991
Epomophorus minimus Claessen & De Vree, 1991
Platymops setiger (Peters, 1878)
Platymops Thomas, 1906
Myonycteris relicta Bergmans, 1980
Laephotis kirinyaga Monadjem, Patterson, Webala and Demos, 2020
Cloeotis percivali Thomas, 1901
Tadarida lobata (Thomas, 1891)
Rhinolophus deckenii Peters, 1868
Barbastella leucomelas (Cretzschmar, 1826)
Barbastella leucomelas (Cretzschmar, 1826)
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Miniopterus egeri Goodman, Ramasindrazana, Maminirina, Schoeman and Appleton, 2011
Hypsugo ariel (Thomas, 1904)
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Rhinopoma cystops Thomas, 1903
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Taphozous nudiventris Cretzschmar, 1830
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Eidolon Rafinesque, 1815
Doryrhina camerunensis (Eisentraut, 1956)

Eisentraut's pipistrelle	<i>Nycticeinops eisentrauti</i> (Hill, 1968)
Eisentraut's Pipistrelle Bat	<i>Nycticeinops eisentrauti</i> (Hill, 1968)
Ejeta's House Bat	<i>Scotophilus ejetai</i> Brooks and Bickham, 2014
Eloquent Horseshoe Bat	<i>Rhinolophus eloquens</i> K. Andersen, 1905
Epaulet bats	<i>Epomops</i> Gray, 1866
Epauletted bats	<i>Epomops</i> Gray, 1866
Epauletted Fruit-bats	<i>Epomophorus</i> Bennett, 1836
Ethiopian Big-eared Bat	<i>Plecotus balensis</i> Kruskop & Lavrenchenko, 2000
Ethiopian Epauletted Fruit Bat	<i>Epomophorus labiatus</i> (Temminck, 1837)
Ethiopian Epauletted Fruit-bat	<i>Epomophorus labiatus</i> (Temminck, 1837)
Ethiopian Large-eared Leaf-nosed Bat	<i>Hipposideros megalotis</i> (Heuglin, 1861)
Ethiopian Large-eared Roundleaf Bat	<i>Hipposideros megalotis</i> (Heuglin, 1861)
Ethiopian Long-eared Bat	<i>Plecotus balensis</i> Kruskop & Lavrenchenko, 2000
Ethiopian Woolly Bat	<i>Kerivoula eriophora</i> (Heuglin, 1877)
Eurasian Free-tailed Bat	<i>Tadarida teniotis</i> (Rafinesque, 1814)
European Free-tailed Bat	<i>Tadarida teniotis</i> (Rafinesque, 1814)
European Guano Bat	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Evening bats	<i>VESPERTILIONIDAE</i> Gray, 1821
Fairy Pipistrelle	<i>Hypsugo ariel</i> (Thomas, 1904)
False Vampire Bats	<i>MEGADERMATIDAE</i> H. Allen, 1864
Felten's Myotis	<i>Myotis punicus</i> Felten, 1977
Flat-faced Serotine Bat	<i>Eptesicus (Cnephæus) platyops</i> (Thomas, 1901)
Flat-headed Bats	<i>Mormopterus</i> Peters, 1865
Flat-headed Bats	<i>Mimetillus</i> Thomas, 1904
Flat-headed Bulldog Bats	<i>Mormopterus</i> Peters, 1865
Flat-headed free-tailed bat	<i>Platymops setiger</i> (Peters, 1878)
Flat-headed Free-tailed Bat	<i>Sauromys petrophilus</i> (Roberts, 1917)
Flat-headed Vesper Bats	<i>Mimetillus</i> Thomas, 1904
Flitter Mouse	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Flower's Pipistrelle Bat	<i>Eptesicus (Rhinopterus) floweri</i> (de Winton, 1901)
Flying Calf	<i>Nanonycteris veldkampii</i> (Jentink, 1888)
Flying Foxes	<i>Pteropus</i> Brisson, 1762
Flying-foxes	<i>Pteropus</i> Brisson, 1762
Forest Bats	<i>Kerivoulinæ</i> Miller, 1907
Forest Horseshoe Bat	<i>Rhinolophus silvestris</i> Aellen, 1959
Franquet's Clamorous Fruit-bat	<i>Epomops franqueti</i> (Tomes, 1860)
Franquet's Epauletted Fruit Bat	<i>Epomops franqueti</i> (Tomes, 1860)
Franquet's Fruit Bat	<i>Epomops franqueti</i> (Tomes, 1860)
Free-tailed Bats	<i>MOLOSSIDAE</i> Gervais, 1856
Free-tailed Bats	<i>Molossinae</i> Gervais, 1856
Free-tailed Bats	<i>Tadarida</i> Rafinesque, 1814
Gaisler's Long-eared Bat	<i>Plecotus gaisleri</i> Benda, Kiefer, Hanak & Veith, 2004
Gallagher's Free-tailed Bat	<i>Mops (Chaerephon) gallagheri</i> (Harrison, 1975)
Gallagher's Wrinkle-lipped Bat	<i>Mops (Chaerephon) gallagheri</i> (Harrison, 1975)
Gambian Epauletted Bat	<i>Epomophorus gambianus</i> (Ogilby, 1835)
Gambian Epauletted Fruit bat	<i>Epomophorus gambianus</i> (Ogilby, 1835)
Gambian Slit-faced Bat	<i>Nycteris gambiensis</i> (K. Andersen, 1912)
Geat Brown Bat	<i>Scotophilus nigrita</i> (Schreber, 1774)
Geoffroy's Bat	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Geoffroy's Horseshoe Bat	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
Geoffroy's Lesser Tident Bat	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
Geoffroy's Myotis	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Geoffroy's Nycteris	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
Geoffroy's Tomb Bat	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
Geoffroy's Trident Leaf-nosed Bat	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
Giant African Free-tailed Bat	<i>Tadarida ventralis</i> (Heuglin, 1861)
Giant African Guano Bat	<i>Tadarida ventralis</i> (Heuglin, 1861)
Giant Brown Bat	<i>Scotophilus nigrita</i> (Schreber, 1774)
Giant Brown House Bat	<i>Scotophilus nigrita</i> (Schreber, 1774)
Giant Free-tailed Bat	<i>Tadarida ventralis</i> (Heuglin, 1861)
Giant gland-tailed bat	<i>Mops (Chaerephon) major</i> (Trouessart, 1897)
Giant Guano Bat	<i>Tadarida ventralis</i> (Heuglin, 1861)
Giant House Bat	<i>Scotophilus nigrita</i> (Schreber, 1774)
Giant Leaf-nosed Bat	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Giant Leaf-nosed Bat	<i>Macronycteris gigas</i> (Wagner, 1845)
Giant Mastiff Bat	<i>Otomops martiensseni</i> (Matschie, 1897)
Giant noctule	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Giant Noctule Bat	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Giant Yellow House Bat	<i>Scotophilus nigrita</i> (Schreber, 1774)
Gland-tailed Free-tailed Bat	<i>Mops (Chaerephon) bemmeleni</i> (Jentink, 1879)
Gland-tailed Wrinkle-lipped Bat	<i>Mops (Chaerephon) bemmeleni</i> (Jentink, 1879)
Glauconycteris	<i>Glauconycteris</i> Dobson, 1875
Glen's Butterfly Bat	<i>Glauconycteris gleni</i> Peterson and Smith, 1973
Glen's Long-fingered Bat	<i>Miniopterus gleni</i> Peterson, Eger and Mitchell, 1995
Glen's Wattled Bat	<i>Glauconycteris gleni</i> Peterson and Smith, 1973

- Goblin bats
Golden Bat
- Golden Short-palate Fruit-bat
Golden Short-palated Fruit Bat
Goodman's Trident Bat
Grandidier's Pipistrelle
Grandidier's Free-tailed Bat
Grandidier's trident bat
Great Brown House Bat
Great pipistrelle
Greater Bent-winged Bat
Greater Cyclops
Greater Free-tailed Bats
Greater Horseshoe Bat
Greater Long-fingered Bat
Greater Mascarene Flying Fox
Greater Mouse-tailed Bat
Greater noctule
Greater Noctule Bat
Greater Rat-tailed Bat
Greater Roundleaf Bat
Greater Serotine Bats
Greater Slit-faced Bat
Greater Trident Bats
Green House Bat
Greenish Yellow Bat
Griveaud's Long-fingered bat
Guano Bats
Guinea Serotine
Guinean Horseshoe Bat (Csorba *et al.*, 2003: 59 - 61; Simmons, 2005)
Guinean Pipistrelle Bat
Haagner's Flat-headed Bat
Hairy Bats
Hairy Long-eared Bat
Hairy Slit-faced Bat
Hairy-armed bat
Halcyon Horseshoe Bat
Hamilton's Naked-bellied Tomb Bat
Hamilton's Naked-rumped Tomb Bat
Hamilton's Tomb Bat
Hammer-headed bat
Hammer-headed bats
Hammer-headed Fruit Bat
Hanak's Pipistrelle
Happolds' Pipistrelle
- Hardwicke's Mouse-tailed Bat
Harlequin Fruit-bats
Harrison's large-eared giant mastiff bat
- Harrison's fruit bat
Hayman's tear-drop fruit bat
Hayman's Dwarf Epauletted Fruit Bat
Hayman's Epauletted Fruit Bat
Hayman's Lesser Fruit Bat
Hayman's Lesser Fruit-bat
Heart-nosed Bat
Heart-nosed Bats
Heart-nosed Big-eared Bat
Helios Pipistrelle Bat
Heller's Pipistrelle
Heller's Pipistrelle Bat
Hemprich's arrow-eared bat
Hemprich's Arrow-headed Bat
Hemprich's Big-eared Bat
Hemprich's Desert Bat
Hemprich's Desert Long-eared Bat
Hemprich's Long-eared Bat
Hemprich's Long-eared Desert Bat
High-crowned bat
Hildebrandt's Horseshoe Bat
Hildegard's Tomb Bat
Hill's Horseshoe Bat
Hill's Horseshoe bat
- Mormopterus* Peters, 1865
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Casinycteris argynnis Thomas, 1910
Casinycteris argynnis Thomas, 1910
Triaenops goodmani Samonds, 2007
Nycticeinops grandidieri (Dobson, 1876)
Mops (Chaerephon) leucogaster (A. Grandidier, 1869)
Paratriaenops auritus (G. Grandidier, 1912)
Scotophilus nigrita (Schreber, 1774)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Doryrhina camerunensis (Eisentraut, 1956)
Mops Lesson, 1842
Rhinolophus ferrumequinum (Schreber, 1774)
Miniopterus inflatus Thomas, 1903
Pteropus niger (Kerr, 1792)
Rhinopoma microphyllum (Brünnich, 1782)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus lasiopterus (Schreber, 1780)
Rhinopoma microphyllum (Brünnich, 1782)
Doryrhina camerunensis (Eisentraut, 1956)
Eptesicus Rafinesque, 1820
Nycteris macrotis Dobson, 1876
Triaenops Dobson, 1871
Scotophilus viridis (Peters, 1852)
Scotophilus viridis (Peters, 1852)
Miniopterus griveaudi Harrison, 1959
Tadarida Rafinesque, 1814
Neoromicia guineensis (Bocage, 1889)
Rhinolophus guineensis Eisentraut, 1960
- Neoromicia guineensis* (Bocage, 1889)
Sauromys petrophilus haagneri (Roberts, 1917)
Myotis Kaup, 1829
Nycteris hispida (Schreber, 1774)
Nycteris hispida (Schreber, 1774)
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus alcyone Temminck, 1853
Taphozous hamiltoni Thomas, 1920
Taphozous hamiltoni Thomas, 1920
Taphozous hamiltoni Thomas, 1920
Hypsignathus monstrosus H. Allen, 1862
Hypsignathus H. Allen, 1862
Hypsignathus monstrosus H. Allen, 1862
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Nycticeinops happoldorum (Hutterer, Decher, Monadjem and Astrin, 2019)
Rhinopoma cystops Thomas, 1903
Scotonycteris Matschie, 1894
Otomops harrisoni Ralph, Richards, Taylor, Napier and Lamb, 2015
Myonycteris angolensis goliath (Bergmans, 1997)
Scotonycteris occidentalis Hayman, 1947
Epomophorus intermedius (Hayman, 1963)
Epomophorus intermedius (Hayman, 1963)
Epomophorus intermedius (Hayman, 1963)
Epomophorus intermedius (Hayman, 1963)
Cardioderma cor (Peters, 1872)
Cardioderma Peters, 1873
Cardioderma cor (Peters, 1872)
Afronycteris helios (Heller, 1912)
Afronycteris helios (Heller, 1912)
Afronycteris helios (Heller, 1912)
Otonycteris hemprichii Peters, 1859
Otonycteris hemprichii Peters, 1859
Otonycteris hemprichii Peters, 1859
Otonycteris hemprichii Peters, 1859
Otonycteris hemprichii Peters, 1859
Otonycteris hemprichii Peters, 1859
Otonycteris hemprichii Peters, 1859
Miniopterus inflatus Thomas, 1903
Rhinolophus hildebrandtii Peters, 1878
Taphozous hildegardeae Thomas, 1909
Rhinolophus hilli Aellen, 1973
Rhinolophus hillorum Koopman, 1989

Hinde's Lesser House Bat	<i>Scotoecus hindei</i> Thomas, 1901
Hispid bats	NYCTERIDAE Van der Hoeven, 1855
Hollow-faced bats	NYCTERIDAE Van der Hoeven, 1855
Hollow-faced bats	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795
Horn-skinned Bat	<i>Eptesicus (Rhinopterus) floweri</i> (de Winton, 1901)
Horn-skinned Pipistrelle Bat	<i>Eptesicus (Rhinopterus) floweri</i> (de Winton, 1901)
Horn-skinned Serotine	<i>Eptesicus (Rhinopterus) floweri</i> (de Winton, 1901)
Horseshoe bats	RHINOLOPHIDAE Gray, 1825
Horseshoe bats	<i>Rhinolophus</i> Lacépède, 1799
Hottentot bat	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
Hottentot Serotine Bat	<i>Eptesicus (Cnephaeus) hottentotus</i> (A. Smith, 1833)
House bats	<i>Scotophilus</i> Leach, 1821
House Bats	<i>Scotoecus</i> Thomas, 1901
Intermediate Slit-faced Bat	<i>Nycteris intermedia</i> Aellen, 1959
Isabelline Serotine Bat	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
Isabelline White-winged Serotine	<i>Pseudoromicia isabella</i> (Decher, Hutterer and Monadjem, 2016)
Isalo Serotine	<i>Laephotis malagasyensis</i> (Peterson, Eger and Mitchell, 1995)
Ja Slit-faced Bat	<i>Nycteris major</i> (K. Andersen, 1912)
Jones' Leaf-nosed Bat	<i>Hipposideros jonesi</i> Hayman, 1947
Jones' Roundleaf Bat	<i>Hipposideros jonesi</i> Hayman, 1947
Jones's Roundleaf Bat	<i>Hipposideros jonesi</i> Hayman, 1947
Kahuzi horseshoe bat	<i>Rhinolophus kahuzi</i> Fahr and Kerbis Peterhans, 2013
Kenya Barbastelle	<i>Glauconycteris kenyacola</i> Peterson, 1982
Kenya Pipistrelle	<i>Pipistrellus (Pipistrellus) aero</i> Heller, 1912
Kenyan Big-eared Free-tailed Bat	<i>Tadarida lobata</i> (Thomas, 1891)
Kenyan Butterfly Bat	<i>Glauconycteris kenyacola</i> Peterson, 1982
Kenyan Wattle Bat	<i>Glauconycteris kenyacola</i> Peterson, 1982
Kityo's serotine	<i>Pseudoromicia kityoi</i> Monadjem, Kerbis Peterhans, Nalikka, Waswa, Demos and Patterson, 2020
Knight's Egyptian bat	<i>Aegyptonycteris knightae</i> Simmons, Seiffert and Gunnell, 2016
Knob-tailed Mops Bat	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
Knob-tailed Nyctinome	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
Kolombatovic's Big-eared Bat	<i>Plecotus kolombatovici</i> Dulic, 1980
Kolombatovic's Long-eared Bat	<i>Plecotus kolombatovici</i> Dulic, 1980
Kordofan Sheath-tail-bat	<i>Taphozous hamiltoni</i> Thomas, 1920
Kruger serotine bat (Taylor, 2005: 305 - first use)	<i>Laephotis stanleyi</i> (Goodman, Kearney, Ratsimbazafy and Hassanin, 2017)
Kuhl's Bat	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Kuhl's pipistrelle	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Kuhl's Pipistrelle Bat	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Lagos Serotine	<i>Eptesicus (Cnephaeus) platyops</i> (Thomas, 1901)
Lagos Serotine Bat	<i>Eptesicus (Cnephaeus) platyops</i> (Thomas, 1901)
Lamotte's Leaf-nosed Bat	<i>Hipposideros lamottei</i> Brosset, 1985
Lamotte's Roundleaf Bat	<i>Hipposideros lamottei</i> Brosset, 1985
Lander's Horseshoe Bat	<i>Rhinolophus landeri</i> Martin, 1838
Lappet-eared Free-tailed Bat	<i>Mops (Chaerephon) major</i> (Trouessart, 1897)
Lappet-eared Wrinkle-lipped Bat	<i>Mops (Chaerephon) major</i> (Trouessart, 1897)
Large Bentwing Bat	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
large bent-winged bat	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Large Free-tailed Bat	<i>Tadarida fulminans</i> (Thomas, 1903)
Large Guano Bat	<i>Tadarida fulminans</i> (Thomas, 1903)
Large Headed Pipistrelle	<i>Nycticeinops macrocephalus</i> (Hutterer and Kerbis Peterhans, 2020)
Large Slit-faced Bat	<i>Nycteris grandis</i> Peters, 1865
Large Wrinkle-lipped Bat	<i>Mops (Chaerephon) major</i> (Trouessart, 1897)
Large-eared Free-tailed Bat	<i>Otomops martiensseni</i> (Matschie, 1897)
Large-eared Giant Mastiff Bat	<i>Otomops martiensseni</i> (Matschie, 1897)
Large-eared Leaf-nosed Bat	<i>Hipposideros megalotis</i> (Heuglin, 1861)
Large-eared Slit-faced Bat	<i>Nycteris macrotis</i> Dobson, 1876
Large-footed Bats	<i>Myotis</i> Kaup, 1829
Larger Horseshoe Bat	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Larger Mouse-tailed Bat	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
Larger Rat-tailed Bat	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
Large-winged Bats	MEGADERMATIDAE H. Allen, 1864
Leafnosed Bats	<i>Hipposideros</i> Gray, 1831
Leaf-nosed bats	HIPPOSIDERIDAE Lydekker, 1891
Leaf-nosed Bats	<i>Hipposideros</i> Gray, 1831
Leaf-winged bat	<i>Glauconycteris variegata</i> (Tomes, 1861)
Least Epauletted Fruit-bat	<i>Epomophorus minimus</i> Claessen & De Vree, 1991
Least horseshoe bat	<i>Rhinolophus gorongosae</i> Taylor, MacDonald, Goodman, Kearney, Cotterill, Stoffberg, Monadjem, Schoeman, Guyton, Naskrecki and Richards, 2018
Least Long-fingered Bat	<i>Miniopterus minor</i> Peters, 1867

- Least Pipistrelle Bat
 Leisler's Bat
 Leisler's noctule
 Lesser (Sundevall's) Leaf-nosed Bat
 Lesser Angolan Epauletted Fruit Bat
 Lesser bent-winged bat
 Lesser Epauletted Bat
 Lesser Free-tailed Bat
 Lesser Free-tailed Bats
 Lesser Horseshoe Bat
 Lesser House Bats
 Lesser Leaf-nosed bat
 Lesser leaf-nosed bat
 Lesser Long-fingered Bat
 Lesser Mascarene Flying Fox
 Lesser Mascarene Flying-fox
 Lesser Mastiff Bats
 Lesser Mouse-tailed Bat
 Lesser noctule
 Lesser Noctule Bat
 Lesser Nut-coloured House Bat
 Lesser Rat-tailed Bat
 Lesser Sudan Horn-skinned Bat
 Lesser Trident Bats
 Lesser Woolly Bat
 Lesser Yellow Bat
 Lesser Yellow House Bat
 Lesser Yellow House Bat
 Lesser Yellow House Bat
 Lesueur's Hairy Bat
 Lesueur's Myotis
 Lesueur's Wing-gland Bat
 Lightning Guano Bat
 Light-winged Lesser House Bat
 Little Brown Bats
 Little Bulldog Bats
 Little Collared Fruit Bat
 Little Collared Fruit-bat
 Little Collared Fruit-bats
 Little Epauletted Fruit-bat
 Little Flying Cow
 Little Flying Cow
 Little Free-tailed Bat
 Little Goblin Bats
 Little Mastiff Bats
 Little Nut-coloured House Bat
 Little Wrinkle-lipped Bat
 Livingstone's House Bat
 Livingstone's Flying Fox
 Lobe-lipped Bats
 Long-crested Free-tailed Bat
 Long-crested Gland-tailed Bat
 Long-crested Wrinkle-lipped Bat
 Long-eared bats
 Long-eared bats
 Long-eared Bats
 Long-eared Desert Bat
 Long-eared Desert Bats
 Long-fingered Bat
 Long-fingered Bats
 Long-fingered Bats
 Long-fingered Myotis
 Long-haired Rousette
 Long-haired Rousette
 Long-tailed Bat
 Long-tailed Greater Serotine Bat
 Long-tailed House Bat
 Long-tailed Serotine Bat
 Long-wing Bat
 Machado's Butterfly Bat
 MacInnes's Mouse-tailed Bat
 MacLaud's Horseshoe Bat
 Madagascan Flying Fox
 Madagascan Flying-fox
 Madagascan Free-tailed Bat
 Madagascan Fruit Bat
Hypsugo musciculus (Thomas, 1913)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Hipposideros tephros (Cabrera, 1906)
Epomophorus grandis (Sanborn, 1950)
Miniopterus fraterculus Thomas and Schwann, 1906
Myonycteris Matschie, 1899
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Mops (Chaerephon) Dobson, 1874
Rhinolophus hipposideros (Bechstein, 1800)
Scotoecus Thomas, 1901
Hipposideros caffer (Sundevall, 1846)
Hipposideros tephros (Cabrera, 1906)
Miniopterus fraterculus Thomas and Schwann, 1906
Pteropus subniger (Kerr, 1792)
Pteropus subniger (Kerr, 1792)
Mops (Chaerephon) Dobson, 1874
Rhinopoma cystops Thomas, 1903
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Scotophilus nucella Robbins, 1983
Rhinopoma cystops Thomas, 1903
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Asellia Gray, 1838
Kerivoula lanosa (A. Smith, 1847)
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Scotophilus leucogaster (Cretzschmar, 1826)
Scotophilus viridis (Peters, 1852)
Cistugo lesueuri Roberts, 1919
Cistugo lesueuri Roberts, 1919
Cistugo lesueuri Roberts, 1919
Tadarida fulminans (Thomas, 1903)
Scotoecus albofuscus (Thomas, 1890)
Myotis Kaup, 1829
Mormopterus Peters, 1865
Myonycteris torquata (Dobson, 1878)
Myonycteris torquata (Dobson, 1878)
Myonycteris Matschie, 1899
Epomophorus labiatus (Temminck, 1837)
Nanonycteris Matschie, 1899
Nanonycteris veldkampii (Jentink, 1888)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Mormopterus Peters, 1865
Mormopterus Peters, 1865
Scotophilus nucella Robbins, 1983
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Scotophilus livingstonii Brooks and Bickham, 2014
Pteropus livingstonii Gray, 1866
Glauconycteris Dobson, 1875
Mops (Chaerephon) chapini (J.A. Allen, 1917)
Mops (Chaerephon) chapini (J.A. Allen, 1917)
Mops (Chaerephon) chapini (J.A. Allen, 1917)
 NYCTERIDAE Van der Hoeven, 1855
Plecotus E. Geoffroy St.-Hilaire, 1818
Laephotis Thomas, 1901
Otonycteris hemprichii Peters, 1859
Otonycteris Peters, 1859
Myotis capaccinii (Bonaparte, 1837)
 MINIOPTERIDAE Dobson, 1875
Miniopterus Bonaparte, 1837
Myotis capaccinii (Bonaparte, 1837)
Myonycteris angolensis (Bocage, 1898)
Stenonycteris lanosus (Thomas, 1906)
Rhinopoma microphyllum (Brünnich, 1782)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Miniopterus schreibersii (Kuhl, 1817)
Glauconycteris machadoi Hayman, 1963
Rhinopoma macinnesi Hayman, 1937
Rhinolophus macclaudi Pousargues, 1898
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Tadarida fulminans (Thomas, 1903)
Eidolon dupreanum (Schegel, 1867)

Madagascar Large Free-tailed Bat	<i>Tadarida fulminans</i> (Thomas, 1903)
Madagascar Large Guano Bat	<i>Tadarida fulminans</i> (Thomas, 1903)
Madagascar Roussette	<i>Rousettus madagascariensis</i> G. Grandidier, 1929
Madagascar Straw-coloured Fruit-bat	<i>Eidolon dupreanum</i> (Schegel, 1867)
Madagascar white-bellied Free-tailed Bat	<i>Mops (Chaerephon) leucogaster</i> (A. Grandidier, 1869)
Madagascar cryptic leaf-nosed bat	<i>Macronycteris cryptovalorona</i> (Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016)
Madagascar Flying Fox	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
Madagascar Free-tailed Bat	<i>Otomops madagascariensis</i> Dorst, 1953
Madagascar Fruit Bat	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
Madagascar Large Free-tailed Bat	<i>Tadarida fulminans</i> (Thomas, 1903)
Madagascar Leaf-nosed Bat	<i>Paratriaenops furculus</i> (Trouessart, 1907)
Madagascar Mastiff Bat	<i>Otomops madagascariensis</i> Dorst, 1953
Madagascar Roussette	<i>Rousettus madagascariensis</i> G. Grandidier, 1929
Madagascar serotine	<i>Laephotis matroka</i> (Thomas and Schwann, 1905)
Madagascar Sheath-tailed Bat	<i>Coleura kibomalandy</i> Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012
Madagascar sheath-tailed bat	<i>Paremballonura atrata</i> (Peters, 1874)
Madagascar Sucker-footed Bat	<i>Myzopoda aurita</i> Milne-Edwards and A. Grandidier, 1878
Madagascar White-bellied Free-tailed Bat	<i>Mops (Mops) leucostigma</i> (G.M. Allen, 1918)
Madagascar Yellow House Bat	<i>Scotophilus borbonicus</i> (E. Geoffroy St.-Hilaire, 1803)
Madeira pipistrelle	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeiran Pipistrelle Bat	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Maendeleo Horseshoe Bat	<i>Rhinolophus maendeleo</i> Kock, Csorba and Howell, 2000
Maghrebian bent-wing bat	<i>Miniopterus maghrebensis</i> Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014
Maghrebian Mouse-eared Bat	<i>Myotis punicus</i> Felten, 1977
Major Long-fingered Bat	<i>Miniopterus majori</i> Thomas, 1906
Major's Long-fingered Bat	<i>Miniopterus majori</i> Thomas, 1906
Malagasy Free-tailed Bat	<i>Tadarida fulminans</i> (Thomas, 1903)
Malagasy Mouse-eared Bat	<i>Myotis goudoti</i> (A. Smith, 1834)
Malagasy Serotine	<i>Laephotis matroka</i> (Thomas and Schwann, 1905)
Malagasy Straw-colored Fruit Bat	<i>Eidolon dupreanum</i> (Schegel, 1867)
Malagasy White-bellied Free-tailed Bat	<i>Mops (Mops) leucostigma</i> (G.M. Allen, 1918)
Manavi Long-fingered Bat	<i>Miniopterus manavi</i> Thomas, 1906
Manavil Long-fingered Bat	<i>Miniopterus manavi</i> Thomas, 1906
Marisa's Leaf-nosed Bat	<i>Hipposideros marisae</i> Aellen, 1954
Marovaza House Bat	<i>Scotophilus marovaza</i> Goodman, Ratrimomanarivo and Randrianandrianina, 2006
Martensen's Free-tailed Bat	<i>Otomops martiensseni</i> (Matschie, 1897)
Martienssen bat	<i>Otomops martiensseni</i> (Matschie, 1897)
Martienssen's Big-eared Bulldog Bat	<i>Otomops martiensseni</i> (Matschie, 1897)
Mascarene Flying-fox	<i>Pteropus niger</i> (Kerr, 1792)
Mastiff Bats	<i>MOLOSSIDAE</i> Gervais, 1856
Mastiff Bats	<i>Molossinae</i> Gervais, 1856
Mastiff Bats	<i>Otomops</i> Thomas, 1913
Matroka Bat	<i>Laephotis matroka</i> (Thomas and Schwann, 1905)
Mauritian Flying Fox	<i>Pteropus niger</i> (Kerr, 1792)
Mauritian Tomb Bat	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
Mediterranean Horseshoe Bat	<i>Rhinolophus euryale</i> Blasius, 1853
Mediterranean Horseshoe Bat	<i>Rhinolophus mehelyi</i> Matschie, 1901
Mediterranean Lesser pipistrelle	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Mediterranean Long-eared Bat	<i>Plecotus kolombatovici</i> Dulic, 1980
Medje Free-tailed Bat	<i>Mops (Mops) congicus</i> J.A. Allen, 1917
Medje Greater Free-tailed Bat	<i>Mops (Mops) congicus</i> J.A. Allen, 1917
Medje Mops Bat	<i>Mops (Mops) congicus</i> J.A. Allen, 1917
Mehely's Horseshoe Bat	<i>Rhinolophus mehelyi</i> Matschie, 1901
Meridional serotine	<i>Eptesicus (Cnephæus) isabellinus</i> (Temminck, 1840)
Midas Bat	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Midas Free-tailed Bat	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Midas Groove-cheeked Bat	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Midas Mops Bat	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Midge Bat	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Mimic Bats	<i>Mimetillus</i> Thomas, 1904
Minor Epauletted Fruit Bat	<i>Epomophorus minor</i> Dobson, 1880
Miombo pipistrelle	<i>Neoromicia anchietae</i> (Seabra, 1900)
Moloney's Flat-headed Bat	<i>Mimetillus moloneyi</i> (Thomas, 1891)
Moloney's Flat-headed Vesper Bat	<i>Mimetillus moloneyi</i> (Thomas, 1891)
Moloney's Mimic Bat	<i>Mimetillus moloneyi</i> (Thomas, 1891)
Mongalla Free-tailed Bat	<i>Mops (Mops) demonstrator</i> (Thomas, 1903)
Mongalla Mops Bat	<i>Mops (Mops) demonstrator</i> (Thomas, 1903)
Mongallan Mops Bat	<i>Mops (Mops) demonstrator</i> (Thomas, 1903)

Montagne d'Ambre Long-fingered Bat

Mops Bats

Mops Free-tailed Bats

Morris's Bat

Morris's Mouse-eared Bat

Morris's Myotis

Mount Gargues Pipistrelle

Mount Mabu Horseshoe Bat

Mount Nimba Leaf-nosed Bat

Mountain Pipistrelle Bats

Mountain Rousette

Mouse-eared bat

Mouse-eared Bats

Mouselike Pipistrelle

Mouse-like Pipistrelle Bat

Mouse-tailed bats

Mouse-tailed bats

Mozambian Slit-faced Bat

Mozambican Horseshoe Bat

Mozambique long-fingered bat

Mozambique Sheath-tailed Bat

Mt Gargues Pipistrelle

Mt Gargues Pipistrelle Bat

Naked-bellied Taphozous

Naked-bellied Tomb Bat

Naked-rumped bat

Naked-rumped Tomb Bat

Namib Long-eared Bat

Namibian Long-eared Bat

Narrow-winged Bats

Natal Clinging Bat

Natal Flat-headed Bat

Natal Free-tailed Bat

Natal Long-fingered Bat

Natal Wrinkle-lipped Bat

Nectar Bat

Neoromicia Pipistrelle Bats

Niangara Free-tailed Bat

Niangara Mops Bat

Niangaran Mops Bat

Nigerian Free-tailed Bat

Nigerian Wrinkle-lipped Bat

Nimba long-fingered bat

Nimble Slit-faced Bat

Noack's Leaf-nosed Bat

Noack's Roundleaf Bat

Noctule Bats

Noctules

Noisy Fruit-bats

Notch-eared Bat

Nucella House Bat

Nut-colored Yellow Bat

Nut-coloured House Bat

Nux Yellow House Bat

Nyanza serotine

Old World Big-eared Bats

Old World Fruit bats

Old World Long-eared Bats

Old World Sucker-footed Bat

Old-World leaf-nosed bat

Pale Free-tailed Bat

Pale Xantharpy

Panda Bat

Parisi's Slit-faced Bat

Parissi's Slit-faced Bat

Patrizi's Lesser Trident Bat

Patrizi's Trident Leaf-nosed Bat

Paulian's Trident Bat

Miniopterus ambohitrensis Goodman, Ramasindrazana, Naughton and Appleton, 2015

Mops Lesson, 1842

Mops Lesson, 1842

Myotis morrisoni Hill, 1971

Myotis morrisoni Hill, 1971

Myotis morrisoni Hill, 1971

Pipistrellus (Pipistrellus) aero Heller, 1912

Rhinolophus mabuensis Taylor, Stoffberg, Monadjem,

Schoeman, Bayliss and Cotterill, 2012

Hipposideros lamottei Brosset, 1985

Hypsugo kolenati, 1856

Stenonycteris lanosus (Thomas, 1906)

Myotis mystacinus (Kuhl, 1817)

Myotis Kaup, 1829

Hypsugo musciculus (Thomas, 1913)

Hypsugo musciculus (Thomas, 1913)

RHINOPOMATIDAE Dobson, 1872

Rhinopoma E. Geoffroy St.-Hilaire, 1818

Nycteris vinsoni Dalquest, 1965

Rhinolophus mossambicus Taylor, Stoffberg,

Monadjem, Schoeman, Bayliss and Cotterill, 2012

Miniopterus mossambicus Monadjem, Goodman,

Stanley and Appleton, 2013

Coleura afra (Peters, 1852)

Pipistrellus (Pipistrellus) aero Heller, 1912

Pipistrellus (Pipistrellus) aero Heller, 1912

Taphozous nudiventris Cretzschmar, 1830

Taphozous nudiventris Cretzschmar, 1830

Taphozous nudiventris Cretzschmar, 1830

Taphozous nudiventris Cretzschmar, 1830

Laephotis namibensis Setzer, 1971

Laephotis namibensis Setzer, 1971

Mimetillus Thomas, 1904

Miniopterus natalensis (A. Smith, 1833)

Mormopterus acetabulosus (Hermann, 1804)

Mormopterus acetabulosus (Hermann, 1804)

Miniopterus natalensis (A. Smith, 1833)

Mormopterus acetabulosus (Hermann, 1804)

Megaloglossus woermanni Pagenstecher, 1885

Neoromicia Roberts, 1926

Mops (Mops) niangarae J.A. Allen, 1917

Mops (Mops) niangarae J.A. Allen, 1917

Mops (Mops) niangarae J.A. Allen, 1917

Mops (Chaerephon) nigeriae (Thomas, 1913)

Mops (Chaerephon) nigeriae (Thomas, 1913)

Miniopterus nimbae Monadjem, Shapiro, Richards,

Karabulut, Crawley, Nielsen, Hansen, Bohmann and

Mourier, 2020

Nycteris arge Thomas, 1903

Hipposideros ruber (Noack, 1893)

Hipposideros ruber (Noack, 1893)

Nyctalus Bowdich, 1825

Nyctalus Bowdich, 1825

Epomops Gray, 1866

Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)

Scotophilus nucella Robbins, 1983

Scotophilus nux Thomas, 1904

Scotophilus nux Thomas, 1904

Scotophilus nux Thomas, 1904

Pseudoromicia nyanza Monadjem, Patterson, Webala

and Demos, 2020

Plecotus E. Geoffroy St.-Hilaire, 1818

PTEROPODIDAE Gray, 1821

Plecotus E. Geoffroy St.-Hilaire, 1818

Myzopoda aurita Milne-Edwards and A. Grandidier,

1878

HIPPOSIDERIDAE Lydekker, 1891

Mops (Chaerephon) chapini (J.A. Allen, 1917)

Eidolon helvum (Kerr, 1792)

Glauconycteris superba Hayman, 1939

Nycteris parisii (de Beaux, 1924)

Nycteris parisii (de Beaux, 1924)

Asellia patrizii de Beaux, 1931

Asellia patrizii de Beaux, 1931

Paratriaenops pauliani (Goodman and Ranivo, 2008)

- Peak-saddle Horseshoe Bat
 Pel's Pouched Bat
 Pemba Flying Fox
 Pemba Flying-fox
 Percival's Short-eared Trident Bat
 Percival's Trident Bat
 Perforated Taphozous bat
 Peters' Lesser Epauletted Fruit Bat
 Peters' Lesser Fruit-bat
 Peter's Sheath-tailed Bat
 Peter's Trident Bat
 Peters' Wrinkle-lipped Bat
 Peterson's Free-tailed Bat
 Peterson's Long-fingered Bat

 Peterson's Mops Bat
 Peters's Dwarf Epauletted Fruit Bat
 Peters's Epauletted Fruit Bat
 Peters's Flat-headed Bat
 Peters's flat-headed Bats
 Peters's Goblin bat
 Peters's Horseshoe Bat
 Peters's Sheath-tailed Bat
 Peters's sheath-tailed bats
 Peters's Wrinkle-lipped Bat
 Petra fruit bat
 Phantom-winged Bats
 Pied Bat
 Pied Butterfly Bat
 Pipistrelle Bats
 Pipistrelles
 Plain-faced Bats
 Plerote Fruit-bats
 Plerotes Fruit-bats
 Pohle's Fruit Bat
 Pohle's Harlequin Fruit-bat
 Pohle's Tear-drop Fruit-bat
 Pouched Bats
 Pouched Bats
 Pygmy pipistrelle bat
 Pygmy Pipistrellus
 Racey's Pipistrelle Bat

 Railer Bat
 Railer Mops Bat
 Rainforest House Bat
 Rat-tailed Bat
 Red Flying Fox
 Red Leaf-nosed Bat
 Red Madagascan Trident Bat
 Red Trident Bat
 Relict Collared Fruit-bat
 Rendall's Pipistrelle
 Rendall's Pipistrelle Bat
 Rendall's Serotine
 Rendall's Serotine Bat
 Reunion Free-tailed bat

 Robbin's House Bat
 Robbins's House Bat
 Robbins's Yellow Bat
 Roberts' Flat-headed Bat
 Roberts's Flat-headed Bat
 Robust Yellow Bat
 Rock-dwelling Flat-headed Bat
 Rock-loving Flat-headed Bat
 Rodrigues Flying Fox
 Rodriguez Flying Fox
 Roundleaf Bats
 Rousette Fruit-bats
 Rousettes
 Rufous Hairy Bat
 Rufous Mouse-eared bat
 Rufous Mouse-eared Myotis
 Rufous Trident Bat
 Rüppell's Bat

Rhinolophus blasii Peters, 1867
Saccolaimus peli (Temminck, 1853)
Pteropus voeltzkowi Matschie, 1909
Pteropus voeltzkowi Matschie, 1909
Cloeotis percivali Thomas, 1901
Cloeotis percivali Thomas, 1901
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Epomophorus pusillus Peters, 1868
Epomophorus pusillus Peters, 1868
Paremballonura atrata (Peters, 1874)
Trienops afer Peters, 1877
Mormopterus jugularis (Peters, 1865)
Mops (Xiphonycteris) petersoni (El Rayah, 1981)
Minioporus petersoni Goodman, Bradman, Maminirina, Ryan, Christidis & Appleton, 2008
Mops (Xiphonycteris) petersoni (El Rayah, 1981)
Epomophorus pusillus Peters, 1868
Epomophorus crypturus Peters, 1852
Platymops setiger (Peters, 1878)
Platymops Thomas, 1906
Mormopterus jugularis (Peters, 1865)
Rhinolophus blasii Peters, 1867
Paremballonura atrata (Peters, 1874)
Coleura Peters, 1867
Mormopterus jugularis (Peters, 1865)
Myonycteris angolensis petraea (Bergmans, 1997)
Mormopterus Peters, 1865
Glauconycteris superba Hayman, 1939
Glauconycteris superba Hayman, 1939
Pipistrellus Kaup, 1829
Pipistrellus Kaup, 1829
 VESPERTILIONIDAE Gray, 1821
Plerotes K. Andersen, 1910
Plerotes K. Andersen, 1910
Casinycteris ophiodon (Pohle, 1943)
Casinycteris ophiodon (Pohle, 1943)
Casinycteris ophiodon (Pohle, 1943)
Saccolaimus Temminck, 1838
Taphozous E. Geoffroy St.-Hilaire, 1818
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Hypsugo ariel (Thomas, 1904)
Pipistrellus (Pipistrellus) raceyi Bates, Ratrimomanarivo, Harrison and Goodman, 2006
Mops (Xiphonycteris) thersites (Thomas, 1903)
Mops (Xiphonycteris) thersites (Thomas, 1903)
Scotophilus nucella Robbins, 1983
Rhinopoma microphyllum (Brünnich, 1782)
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Hipposideros ruber (Noack, 1893)
Trienops menamena Goodman and Ranivo, 2009
Trienops menamena Goodman and Ranivo, 2009
Myonycteris relicta Bergmans, 1980
Pseudoromicia rendalli (Thomas, 1889)
Pseudoromicia rendalli (Thomas, 1889)
Pseudoromicia rendalli (Thomas, 1889)
Pseudoromicia rendalli (Thomas, 1889)
Mormopterus francoismoutoui Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst, Bowie, 2008
Scotophilus nucella Robbins, 1983
Scotophilus nucella Robbins, 1983
Scotophilus nucella Robbins, 1983
Sauromys petrophilus (Roberts, 1917)
Sauromys petrophilus (Roberts, 1917)
Scotophilus robustus A. Milne-Edwards, 1881
Sauromys petrophilus (Roberts, 1917)
Sauromys petrophilus (Roberts, 1917)
Pteropus rodricensis Dobson, 1878
Pteropus rodricensis Dobson, 1878
Hipposideros Gray, 1831
Rousettus Gray, 1821
Rousettus Gray, 1821
Myotis bocagii (Peters, 1870)
Myotis bocagii (Peters, 1870)
Myotis bocagii (Peters, 1870)
Trienops menamena Goodman and Ranivo, 2009
Vansonia rueppellii (J.B. Fischer, 1829)

- Rüppell's Horseshoe Bat
 Ruppell's Pipistrelle
 Rüppel's Pipistrelle
 Rüppel's Pipistrelle Bat
 Russet Free-tailed Bat
 Russet Wrinkle-lipped bat
 Rusty Bat
 Rusty Pipistrelle
 Rusty Pipistrelle Bat
 Ruwenzori Horseshoe bat
 Ruwenzori Long-haired rousette
 Ruwenzori Long-haired Rousette
 Sac-winged Bats
 Sakeji Horseshoe bat
 Samburu Pipistrelle Bat
 Sanborn's Epauletted Fruit-bat
 Sandy long-fingered bat
 Sao Thome Long-fingered bat
 Sao Tomé Collared Fruit Bat
 São Tomé Free-tailed Bat
 Savi's Pipistrelle
 Savi's Pipistrelle Bat
 Schlieffen's Bat
 Schlieffen's Twilight Bat
 Schliemann's Sucker-footed Bat
- Schreber's Brown Bat
 Schreber's Yellow Bat
 Schreiber's Bat
 Schreiber's Bent-winged Bat
 Schreibers's Long-fingered Bat
 Scott's Mouse-eared Bat
 Scott's Mouse-eared Myotis
 Seabra's Wing-gland Bat
 Senegal Bat
 Senegal Horseshoe Bat
 Senegal Mastiff Bat
 Serotine Bats
 Serotines
 Seychelles Flying Fox
 Seychelles Free-tailed bat
 Seychelles Fruit Bat
 Seychelles Sheath-tailed Bat
 Sheath-tailed Bat
 Sheath-tailed Bats
 Sheath-tailed bats
 Short-eared Bats
 Short-eared Trident Bat
 Short-eared Trident Bats
 Short-palate Fruit-bats
 Short-palated Fruit Bat
 Short-palated Fruit-bats
 Short-tailed Leaf-nosed Bat
 Short-tailed Roundleaf Bat
 Short-winged Mops Bat
 Sierra Leone Free-tailed Bat
 Sierra Leone Mops Bat
 Silver Woolly Bat
 Silvered Bat
 Silvered Bats
 Silvered Butterfly Bat
 Silvered Woolly Bat
 Silvery Butterfly Bat
 Silvery Woolly Bat
 Simandou serotine
- Simple-nosed Bats
 Sinai Barbestelle
 Singing Fruit Bat
 Singing Fruit Bat
 Singing Fruit-bats
 Slender-winged Pipistrelle Bat
 Slit-faced bats
 Slit-faced bats
 Small house-bat
 Small mouse-tailed bat
- Rhinolophus fumigatus* Rüppell, 1842
Vansonia rueppellii (J.B. Fischer, 1829)
Vansonia rueppellii (J.B. Fischer, 1829)
Vansonia rueppellii (J.B. Fischer, 1829)
Mops (Chaerephon) russatus (J.A. Allen, 1917)
Mops (Chaerephon) russatus (J.A. Allen, 1917)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Rhinolophus ruwenzorii J. Eric Hill, 1942
Myonycteris angolensis ruwenzorii (Eisentraut, 1965)
Stenonycteris lanosus (Thomas, 1906)
 EMBALLONURIDAE Gervais, 1855
Rhinolophus sakejiensis Cotterill, 2002
Afronycteris helios (Heller, 1912)
Epomophorus grandis (Sanborn, 1950)
Miniopterus arenarius Heller, 1912
Miniopterus newtoni Bocage, 1889
Myonycteris brachycephala (Bocage, 1889)
Mops (Chaerephon) tomensis (Juste and Ibáñez, 1993)
Hypsugo savii (Bonaparte, 1837)
Hypsugo savii (Bonaparte, 1837)
Nycticeinops schlieffenii (Peters, 1859)
Nycticeinops schlieffenii (Peters, 1859)
Myzopoda schliemannii Goodman, Rakotondraparany and Kofoky, 2007
Scotophilus nigrita (Schreber, 1774)
Scotophilus nigrita (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Myotis scotti Thomas, 1927
Myotis scotti Thomas, 1927
Cistugo seabrae Thomas, 1912
Scotophilus nigrita (Schreber, 1774)
Rhinolophus guineensis Eisentraut, 1960
Myotis daubentonii Desmarest, 1820
Eptesicus Rafinesque, 1820
Eptesicus Rafinesque, 1820
Pteropus seychellensis A. Milne-Edwards, 1877
Mops (Chaerephon) pusillus (Miller, 1902)
Pteropus seychellensis A. Milne-Edwards, 1877
Coleura seychellensis Peters, 1868
Coleura afra (Peters, 1852)
 EMBALLONURIDAE Gervais, 1855
Coleura Peters, 1867
Cloeotis Thomas, 1901
Cloeotis percivali Thomas, 1901
Cloeotis Thomas, 1901
Casinycteris Thomas, 1910
Casinycteris argynnis Thomas, 1910
Casinycteris Thomas, 1910
Hipposideros curtus G.M. Allen, 1921
Hipposideros curtus G.M. Allen, 1921
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Kerivoula argentata Tomes, 1861
Glauconycteris argentata (Dobson, 1875)
Glauconycteris Dobson, 1875
Glauconycteris argentata (Dobson, 1875)
Kerivoula argentata Tomes, 1861
Glauconycteris argentata (Dobson, 1875)
Kerivoula argentata Tomes, 1861
Pipistrellus (Pipistrellus) simandouensis Monadjem, Richards, Decher and Hutterer, 2020
 VESPERTILIONIDAE Gray, 1821
Barbastella leucomelas (Cretzschmar, 1826)
Epomophorus dobsonii Bocage, 1889
Epomops franqueti (Tomes, 1860)
Epomops Gray, 1866
Pseudoromicia tenuipinnis (Peters, 1872)
 NYCTERIDAE Van der Hoeven, 1855
Nycteris G. Cuvier and E. Geoffroy, 1795
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rhinopoma cystops Thomas, 1903

Smaller Epauletted fruit bat	<i>Epomophorus crypturus</i> Peters, 1852
Smaller Horseshoe Bat	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Smithers's Horseshoe Bat	<i>Rhinolophus smithersi</i> Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Smith's fruit bat	<i>Myonycteris angolensis smithii</i> (Thomas, 1908)
Smith's Woolly Bat	<i>Kerivoula smithii</i> Thomas, 1880
Smoky Horseshoe Bat	<i>Rhinolophus fumigatus</i> Rüppell, 1842
Smooth-haired Fruit-bats	<i>Myonycteris</i> Matschie, 1899
Snake-toothed Harlequin Fruit-bat	<i>Casinycteris ophiodon</i> (Pohle, 1943)
Snake-toothed Tear-drop Fruit-bat	<i>Casinycteris ophiodon</i> (Pohle, 1943)
Soft-furred Fruit-bats	<i>Myonycteris</i> Matschie, 1899
Somali Pipistrelle Bat	<i>Neoromicia somalica</i> (Thomas, 1901)
Somali Serotine	<i>Neoromicia somalica</i> (Thomas, 1901)
Somali Serotine Bat	<i>Neoromicia somalica</i> (Thomas, 1901)
Somali Trident Leaf-nosed Bat	<i>Asellia italosomalica</i> de Beaux, 1931
Somalian Pipistrelle Bat	<i>Neoromicia somalica</i> (Thomas, 1901)
Sooty Leaf-nosed Bat	<i>Hipposideros fuliginosus</i> (Temminck, 1853)
Sooty Roundleaf Bat	<i>Hipposideros fuliginosus</i> (Temminck, 1853)
Soprano Pipistrelle	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Sororcula Long-fingered Bat	<i>Miniopterus sororculus</i> Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton, 2007
Sororculus Longfingered Bat	<i>Miniopterus sororculus</i> Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton, 2007
South African Lesser Leaf-nosed Bat	<i>Hipposideros caffer</i> (Sundevall, 1846)
Southern Africa horseshoe bat	<i>Rhinolophus capensis</i> Lichtenstein, 1823
Southern Sheath-tailed Bat	<i>Coleura afra</i> (Peters, 1852)
Spotted Free-tailed Bat	<i>Mops (Chaerephon) bivittatus</i> (Heuglin, 1861)
Spotted Gland-tailed Bat	<i>Mops (Chaerephon) bivittatus</i> (Heuglin, 1861)
Spotted Wrinkle-lipped Bat	<i>Mops (Chaerephon) bivittatus</i> (Heuglin, 1861)
Spurrell trumpet-eared bat	<i>Kerivoula phalaena</i> Thomas, 1912
Spurrell's Free-tailed Bat	<i>Mops (Xiphonycteris) spurrelli</i> (Dollman, 1911)
Spurrell's Mops Bat	<i>Mops (Xiphonycteris) spurrelli</i> (Dollman, 1911)
Spurrell's Woolly Bat	<i>Kerivoula phalaena</i> Thomas, 1912
Straw-colored Fruit Bat	<i>Eidolon helvum</i> (Kerr, 1792)
Straw-coloured Flying Fox	<i>Eidolon helvum</i> (Kerr, 1792)
Straw-coloured Fruit-bats	<i>Eidolon</i> Rafinesque, 1815
Striped Butterfly Bat	<i>Glauconycteris alboguttata</i> J.A. Allen, 1917
Striped Leaf-nosed Bat	<i>Macronycteris vittatus</i> (Peters, 1852)
Striped or Banded Guano Bat	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Sucker-footed Bat	<i>Myzopoda aurita</i> Milne-Edwards and A. Grandidier, 1878
Sucker-footed Bats	<i>MYZOPODIDAE</i> Thomas, 1904
Sucker-footed Bats	<i>Myzopoda</i> Milne-Edwards and A. Grandidier, 1878
Sudan Horn-skinned Bat	<i>Eptesicus (Rhinopterus) floweri</i> (de Winton, 1901)
Sudan Serotine	<i>Eptesicus (Rhinopterus) floweri</i> (de Winton, 1901)
Sundevall's African Leaf-nosed Bat	<i>Hipposideros caffer</i> (Sundevall, 1846)
Sundevall's Free-tailed Bat	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Sundevall's Leaf-nosed Bat	<i>Hipposideros caffer</i> (Sundevall, 1846)
Sundevall's Roundleaf Bat	<i>Hipposideros caffer</i> (Sundevall, 1846)
Superb Butterfly Bat	<i>Glauconycteris superba</i> Hayman, 1939
Swallow Lesser House Bat	<i>Scotoecus hirundo</i> (de Winton, 1899)
Swinny's Horseshoe Bat	<i>Rhinolophus swinnyi</i> Gough, 1908
Tadarine Bats	<i>Tadarida</i> Rafinesque, 1814
Tanzania Pipistrelle	<i>Pipistrellus (Pipistrellus) permixtus</i> Aellen, 1957
Tanzanian Woolly Bat	<i>Kerivoula africana</i> Dobson, 1878
Taphozous bat of Mauritius	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
Tear-drop Fruit-bats	<i>Scotonycteris</i> Matschie, 1894
Temminck's Hairy Bat	<i>Myotis tricolor</i> (Temminck, 1832)
Temminck's Mouse-eared Bat	<i>Myotis tricolor</i> (Temminck, 1832)
Thomas' House Bat	<i>Scotoecus albofuscus</i> (Thomas, 1890)
Thomas's Flat-headed Bat	<i>Mimetillus thomasi</i> Hinton, 1920
Thomas's Lesser House Bat	<i>Scotoecus albofuscus</i> (Thomas, 1890)
Thomas's Short-palate Fruit-bat	<i>Casinycteris argynnis</i> Thomas, 1910
Three-coloured bat	<i>Myotis tricolor</i> (Temminck, 1832)
Tiny Pipistrelle	<i>Pipistrellus (Pipistrellus) nanulus</i> Thomas, 1904
Tiny Pipistrelle Bat	<i>Pipistrellus (Pipistrellus) nanulus</i> Thomas, 1904
Tiny Serotine	<i>Neoromicia guineensis</i> (Bocage, 1889)
Tomb Bat	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
Tomb Bat	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
Tomb Bats	<i>EMBALLONURIDAE</i> Gervais, 1855
Tomb Bats	<i>Taphozous</i> E. Geoffroy St.-Hilaire, 1818
Transvaal Free-tailed Bat	<i>Tadarida ventralis</i> (Heuglin, 1861)
Trevor's Bat	<i>Mops (Mops) trevori</i> J.A. Allen, 1917
Trevor's Free-tailed Bat	<i>Mops (Mops) trevori</i> J.A. Allen, 1917
Trevor's Mops Bat	<i>Mops (Mops) trevori</i> J.A. Allen, 1917
Tricoloured Mouse-eared Bat	<i>Myotis tricolor</i> (Temminck, 1832)

- Trident bat
 Trident bats
 Trident Bats
 Trident bats
 Trident Bats
 Trident Leaf-nosed Bat
 Trident Leaf-nosed Bats
 Trident Leaf-nosed Bats
 Trident Rhinolophe
 Trident-nosed Bats
 Triple nose-leaf bats
 Tropical Long-eared Bats
 Trouessart's Triden Bat
 Trouessart's Trident Bat
 Trujillo's House Bat
 Trumpet-eared Bats
 Twilight bats
 Upland Horseshoe bat
 Van Bemmelen's Wrinkle-lipped Bat
 Variegated Butterfly Bat
 Veldkamp's Bat
 Veldkamp's Dwarf Fruit-bat
 Vesper Bats
 Vespertilionid bats
 Vinson's Slit-faced Bat
 Wahlberg's Epauletted Fruit-bat
 Wahlberg's Epauletted Fruit-bat
 Welwitch's Bat
 Welwitsch's Hairy Bat
 Welwitsch's Mouse-eared Bat
 Welwitsch's Myotis
 West African fruit bats
 West African Rousette
 Western Africa leaf-nosed bat
 Western Africa Pipistrelle
 Western Barbastelle
 Western Barbastelle Bat
 Western Sheath-tailed Bat

 Western Yellow Bat

 Whiskered Bat
 Whiskered Mouse-eared Bat
 Whiskered Myotis
 White stripe bat
 White-bellied Free-tailed Bat
 White-bellied Free-tailed Bat
 White-bellied House Bat
 White-bellied Lesser House Bat
 White-bellied Mops Bat
 White-bellied Yellow Bat
 White-fingered Free-tailed Bat
 White-shouldered bat
 White-spotted Butterfly Bat
 White-throated Lesser House Bat
 White-winged Bat
 White-winged Pipistrelle Bat
 White-winged Serotine
 Whitley's Winged-mouse Bat
 Willards horseshoe bat
 Wilson's Long-fingered Bat

 Winged-mouse Bats
 Winged-rat Free-tailed Bats
 Wing-gland bat
 Wing-gland Bats
 Winton's Long-eared Bat
 Woermann's Bat
 Woermann's Fruit bat
 Woermann's Long-tongued Fruit-bat
 Wolf-dog Bats
 Wolf-dog Bats
 Wood's Long-eared Bat
 Wood's Slit-faced Bat
 Woolly bats
 Woolly Bats
- Asellia tridens* (E. Geoffroy St.-Hilaire, 1813)
 HIPPOSIDERIDAE Lydekker, 1891
Asellia Gray, 1838
 RHINONYCTERIDAE J. E. Gray, 1866
Triaenops Dobson, 1871
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Asellia Gray, 1838
Triaenops Dobson, 1871
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Cloeotis Thomas, 1901
Triaenops Dobson, 1871
Laephotis Thomas, 1901
Paratriaenops furculus (Trouessart, 1907)
Paratriaenops furculus (Trouessart, 1907)
Scotophilus trujilloi Brooks and Bickham, 2014
Kerivoula Gray, 1842
Nycticeinops Hill & Harrison, 1987
Rhinolophus hillorum Koopman, 1989
Mops (Chaerephon) bemmeleni (Jentink, 1879)
Glauconycteris variegata (Tomes, 1861)
Nanonycteris veldkampii (Jentink, 1888)
Nanonycteris veldkampii (Jentink, 1888)
 VESPERTILIONIDAE Gray, 1821
 VESPERTILIONIDAE Gray, 1821
Nycteris vinsoni Dalquest, 1965
Epomophorus wahlbergi (Sundevall, 1846)
Epomophorus wahlbergi (Sundevall, 1846)
Myotis welwitschii (Gray, 1866)
Myotis welwitschii (Gray, 1866)
Myotis welwitschii (Gray, 1866)
Myotis welwitschii (Gray, 1866)
Scotonycteris Matschie, 1894
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Hipposideros jonesi Hayman, 1947
Hypsugo musciculus (Thomas, 1913)
Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Paremballonura tiavato (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)
Scotophilus tandrefana Goodman, Jenkins and Ratrimomanarivo, 2005
Myotis mystacinus (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Mops (Mops) niveiventer Cabrera and Ruxton, 1926
Scotophilus leucogaster (Cretzschmar, 1826)
Scotoecus albigula Thomas, 1909
Mops (Mops) niveiventer Cabrera and Ruxton, 1926
Scotophilus leucogaster (Cretzschmar, 1826)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Mops (Mops) leucostigma (G.M. Allen, 1918)
Glauconycteris alboguttata J.A. Allen, 1917
Scotoecus albigula Thomas, 1909
Pseudoromicia tenuipinnis (Peters, 1872)
Pseudoromicia tenuipinnis (Peters, 1872)
Pseudoromicia tenuipinnis (Peters, 1872)
Myopterus whitleyi (Scharff, 1900)
Rhinolophus willardi Kerbis Peterhans and Fahr, 2013
Miniopterus wilsoni Monadjem, Guyton, Naskrecki, Richards, Kropff and Dalton, 2020
Myopterus E. Geoffroy St.-Hilaire, 1818
Myopterus E. Geoffroy St.-Hilaire, 1818
Cistugo lesueuri Roberts, 1919
Cistugo Thomas, 1912
Laephotis wintoni Thomas, 1901
Megaloglossus woermanni Pagenstecher, 1885
Megaloglossus woermanni Pagenstecher, 1885
Megaloglossus woermanni Pagenstecher, 1885
 MOLOSSIDAE Gervais, 1856
Molossinae Gervais, 1856
Nycteris woodi K. Andersen, 1914
Nycteris woodi K. Andersen, 1914
Kerivoulinae Miller, 1907
Kerivoula Gray, 1842

Woolly Rousette	<i>Stenonycteris lanosus</i> (Thomas, 1906)
Wrinkled-lipped Bats	<i>MOLOSSIDAE</i> Gervais, 1856
Wrinkled-lipped Bats	<i>Molossinae</i> Gervais, 1856
Wrinkle-lipped Bat	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Wrinkle-lipped Bats	<i>Mops (Chaerephon)</i> Dobson, 1874
Yellow bats	<i>Scotophilus</i> Leach, 1821
Yellow House Bat	<i>Scotophilus dinganii</i> (A. Smith, 1833)
Yellow Pipistrelle Bat	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
Yellow Serotine	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
Yellow-bellied House Bat	<i>Scotophilus dinganii</i> (A. Smith, 1833)
Yellow-haired fruit bat	<i>Eidolon helvum</i> (Kerr, 1792)
Yellow-winged Bat	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
Yellow-winged Bats	<i>Lavia</i> Gray, 1838
Yellow-winged Short-palate Fruit-bat	<i>Casinycteris argynnis</i> Thomas, 1910
Zaire Gland-tailed Bat	<i>Mops (Chaerephon) gallagheri</i> (Harrison, 1975)
Zenati myotis	<i>Myotis zenatus</i> Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019
Zenker's Fruit Bat	<i>Scotonycteris zenkeri</i> Matschie, 1894
Zenker's Harlequin Fruit-bat	<i>Scotonycteris zenkeri</i> Matschie, 1894
Zenker's Tear-drop Fruit-bat	<i>Scotonycteris zenkeri</i> Matschie, 1894
Ziama Horseshoe Bat (first use Simmons, 2005: 365)	<i>Rhinolophus ziama</i> Fahr, Vierhaus, Hutterer and Kock, 2002
Zulu Pipistrelle Bat	<i>Neoromicia zuluensis</i> (Roberts, 1924)
Zulu Serotine Bat	<i>Neoromicia zuluensis</i> (Roberts, 1924)

Estonian

Alpi nahkhiir	<i>Hypsugo savii</i> (Bonaparte, 1837)
Assoori videvlane	<i>Nyctalus azoreum</i> (Thomas, 1901)
Egiptuse öötiibur	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Euroopa kurdmokk	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Euroopa laikõrv	<i>Barbastella barbastellus</i> (Schreber, 1774)
Euroopa pikktiib	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Habelendlane	<i>Myotis mystacinus</i> (Kuhl, 1817)
Hele-sagarnina	<i>Rhinolophus blasii</i> Peters, 1867
Hiidvidevlane	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Ibeeria nahkhiir	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
Kõrbe-nahkhiir	<i>Eptesicus (Cnephaeus) bottae</i> (Peters, 1869)
Kääbus-nahkhiir	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Kanaari suurkõrv	<i>Plecotus teneriffae</i> Barrett-Hamilton, 1907
Liibüa nahkhiir	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Madeira nahkhiir	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Meheli sagarnina	<i>Rhinolophus mehelyi</i> Matschie, 1901
Paljaskõht-kääpanahkhiir	<i>Taphozous nudiventris</i> Cretzschmar, 1830
Pikkjalg-lendlane	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Pügme-nahkhiir	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Ripslendlane	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Suur-sagarnina	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Suur-teravkõrv	<i>Otonycteris hemprichii</i> Peters, 1859
Vahemere lendlane	<i>Myotis punicus</i> Felten, 1977
Vahemere nahkhiir	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Vahemere sagarnina	<i>Rhinolophus euryale</i> Blasius, 1853
Väike-sagarnina	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Väikevidevlane	<i>Nyctalus leisleri</i> (Kuhl, 1817)

Ewe (Ghana)

Aguto	<i>Eidolon helvum</i> (Kerr, 1792)
Aguto	<i>Epomophorus gambianus</i> (Ogilby, 1835)
Aguto	<i>Epomops buettikoferi</i> (Matschie, 1899)
Aguto	<i>Epomops franqueti</i> (Tomes, 1860)
Chauve-souris à épauettes de Gambie	<i>Epomophorus gambianus</i> (Ogilby, 1835)
chauve-souris paillée	<i>Eidolon helvum</i> (Kerr, 1792)
Chien volant à épauettes du Congo	<i>Epomops franqueti</i> (Tomes, 1860)
Epomophore de Buettikofer	<i>Epomops buettikoferi</i> (Matschie, 1899)
French:: Epomophore de Büttikofer	<i>Epomops buettikoferi</i> (Matschie, 1899)
French:: Epomophore de Franquet	<i>Epomops franqueti</i> (Tomes, 1860)
French:: Epomophore de Gambie	<i>Epomophorus gambianus</i> (Ogilby, 1835)
French:: Roussette jaune	<i>Eidolon helvum</i> (Kerr, 1792)
Roussette de Buettikofer	<i>Epomops buettikoferi</i> (Matschie, 1899)
Roussette de Büttikofer	<i>Epomops buettikoferi</i> (Matschie, 1899)
Roussette de Franquet	<i>Epomops franqueti</i> (Tomes, 1860)
Roussette des palmiers africaine	<i>Eidolon helvum</i> (Kerr, 1792)
Roussette paillée	<i>Eidolon helvum</i> (Kerr, 1792)

Fang

angon

Doryrhina cyclops (Temminck, 1853)

Finnish

Aavikkolepakko
 Arohautalepakko
 Azorienisolepakko
 Doggilepakko
 Doggilepakot
 Euroopanisoherkko
 Euroopanpikkuherkko
 Hautalepakot
 Hedelmälepakot
 Hevosenenkäyököt
 Kääpiölepakko
 Kanariankorvayökkö
 Kapearäpyläpiippa
 Kaviokuonoyököt
 Keltapaimulepakko
 Libyanlepakko
 Madeiranpiippa
 Metsälepakko
 Mopsilepakko
 Pitkäsiipiyökkö
 Pitkäsormisiippa
 Ronkko
 Ruskosiippa
 Rusoisolepakko
 Siipat
 Täpläherkko
 Tunisiansiippa
 Turkinserotiini
 Vaaleaetälänlepakko
 Vaaleaherkko
 Vaivaislepakko
 Valevampyyrit
 Välimerenherkko
 Valkopäälepakot
 Viikisiippa
 Vuoristopiippa

Otonycteris hemprichii Peters, 1859
Taphozous nudiventris Cretzschmar, 1830
Nyctalus azoreum (Thomas, 1901)
Tadarida teniotis (Rafinesque, 1814)
 MOLOSSIDAE Gervais, 1856
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
 EMBALLONURIDAE Gervais, 1855
 PTEROPODIDAE Gray, 1821
 RHINOLOPHIDAE Gray, 1825
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Plecotus teneriffae Barrett-Hamilton, 1907
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
 RHINOLOPHIDAE Gray, 1825
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Nyctalus leisleri (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Nyctalus lasiopterus (Schreber, 1780)
 VESPERTILIONIDAE Gray, 1821
Rhinolophus mehelyi Matschie, 1901
Myotis punicus Felten, 1977
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Rhinolophus blasii Peters, 1867
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
 MEGADERMATIDAE H. Allen, 1864
Rhinolophus euryale Blasius, 1853
 NYCTERIDAE Van der Hoeven, 1855
Myotis mystacinus (Kuhl, 1817)
Hypsugo savii (Bonaparte, 1837)

French

Asellia
 Asellia à trois endentures
 Asellia d'Afrique orientale
 Asellia de Patrizi
 Barbastelle
 Barbastelle argentée
 Barbastelle bariolée
 Barbastelle commune
 Barbastelle d'Abo
 Barbastelle d'Allen
 Barbastelle de Béatrix
 Barbastelle de Glen
 Barbastelle d'Europe
 Barbastelle du Bibundi
 Barbastelle du Kenya
 Barbastelle occidentale
 Barbastelle orientale
 Barbastelles
 Cardioderme à nez en cœur
 Cardiodermes
 Casinyctère
 Casinyctère
 Casinyctère de Campo-Ma'an
 Casinyctère dorée
 Chauve-souris à 3 endentures
 Chauve-souris à face creuse
 Chauve-souris à fer à cheval
 Chauve-souris à longues oreilles d'Angola

Asellia Gray, 1838
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Cloeotis percivali Thomas, 1901
Asellia patrizii de Beaux, 1931
Barbastella barbastellus (Schreber, 1774)
Glauconycteris argentata (Dobson, 1875)
Glauconycteris superba Hayman, 1939
Barbastella barbastellus (Schreber, 1774)
Glauconycteris poensis (Gray, 1842)
Glauconycteris alboguttata J.A. Allen, 1917
Glauconycteris beatrix Thomas, 1901
Glauconycteris gleni Peterson and Smith, 1973
Barbastella barbastellus (Schreber, 1774)
Glauconycteris egeria Thomas, 1913
Glauconycteris kenyacola Peterson, 1982
Barbastella barbastellus (Schreber, 1774)
Barbastella leucomelas (Cretzschmar, 1826)
Barbastella Gray, 1821
Cardioderma cor (Peters, 1872)
Cardioderma Peters, 1873
Casinycteris Thomas, 1910
Casinycteris argynnis Thomas, 1910
Casinycteris campomaanensis Hassanin, 2014
Casinycteris argynnis Thomas, 1910
Asellia Gray, 1838
 NYCTERIDAE Van der Hoeven, 1855
 RHINOLOPHIDAE Gray, 1825
Laephotis angolensis Monard, 1935

- Chauve-souris à longues oreilles de De Winton
 Chauve-souris à longues oreilles de Namibie
 Chauve-souris à longues oreilles du Botswana
 Chauve-souris à oreilles en trompette de Spurell
 Chauve-souris à oreilles en trompette de Spurell
 Chauve-souris à oreilles pointues de Hemprich
 Chauve-souris à pieds à ventouses de Madagascar

 Chauve-souris à queue de souris
 Chauve-souris à queue en fourreau d'Afrique
 Chauve-souris à queue en fourreau des Seychelles
 Chauve-souris à queue gainée d'Afrique
 Chauve-souris à queue gainée de Pel
 Chauve-souris à queue gainée des Seychelles
 chauve-souris à tête de marteau
 Chauve-souris à tête plate
 Chauve-souris à tête plate de Moloney
 Chauve-souris à ventouses de Schliemann

 Chauve-souris bouledogue géante
 Chauve-souris de Commerson à nez feuillu

 Chauve-souris de Commerson à nez feuillu
 Chauve-souris de Schlieffen
 Chauve-souris de Schreiber
 Chauve-souris de Thomas
 Chauve-souris de Veldkamp
 chauve-souris des Hauts
 Chauve-souris des Tombes
 Chauve-souris des tombes
 Chauve-souris des tombes égyptiennes
 Chauve-souris domestique malgache
 Chauve-souris du crépuscule
 Chauve-souris frugivore à collier d'Afrique orientale
 Chauve-souris frugivore à collier de São Tomé
 Chauve-souris frugivore à palais court
 Chauve-souris frugivore d'Anchieta
 Chauve-souris jaune des maisons
 Chauve-souris laineuse
 Chauve-souris laineuse cuivrée
 Chauve-souris laineuse d'Ethiopie
 Chauve-souris laineuse du Damara
 Chauve-souris malgache à oreille de souris
 Chauve-souris malgache à pieds à ventouses

 Chauve-souris malgache à queue en fourreau
 Chauve-souris mauritienne des tombeaux
 Chauve-souris papillon
 Chauve-souris peinte argentée
 Chauve-souris peinte cuivrée
 Chauve-souris peinte de Smith
 Chauve-souris peinte de Tanzanie
 Chauve-souris peinte d'Ethiopie
 Chauve-souris peinte phalène
 Chauve-souris réticulée
 Chauve-souris trident
 Chauve-souris vesper
 Chauves-souris
 Chauves-souris à épauettes
 Chauves-souris à grandes oreilles
 Chauves-souris à longues oreilles
 Chauves-souris à nez large
 Chauves-souris à queue de souris
 Chauves-souris à queue gainée
 Chauves-souris à queue libre
 Chauves-souris à ventouses
 Chauves-souris des maisons
 Chauves-souris frugivore de l'ancien monde
 Chauves-souris peintes
 Chien volant à tête en marteau
 Cléothé de Percival
 Cléothés
 Emballonure d'Afrique
 Emballonure de Madagascar

Laephotis wintoni Thomas, 1901
Laephotis namibensis Setzer, 1971
Laephotis botswanae Setzer, 1971
Kerivoula phalaena Thomas, 1912
Kerivoula phalaena Thomas, 1912
Otonycteris hemprichii Peters, 1859
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Rhinopoma cystops Thomas, 1903
Coleura afra (Peters, 1852)
Coleura seychellensis Peters, 1868
Coleura afra (Peters, 1852)
Saccolaimus peli (Temminck, 1853)
Coleura seychellensis Peters, 1868
Hypsignathus monstrosus H. Allen, 1862
Mimetillus Thomas, 1904
Mimetillus moloneyi (Thomas, 1891)
Myzopoda schliemanni Goodman, Rakotondraparany and Kofoky, 2007
Otomops martiensseni (Matschie, 1897)
Macronycteris commersoni (E. Geoffroy St.-Hilaire, 1813)
Macronycteris gigas (Wagner, 1845)
Nycticeinops schlieffenii (Peters, 1859)
Miniopterus schreibersii (Kuhl, 1817)
Scotoecus albofuscus (Thomas, 1890)
Nanonycteris veldkampii (Jentink, 1888)
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Taphozous E. Geoffroy St.-Hilaire, 1818
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Scotophilus robustus A. Milne-Edwards, 1881
Nycticeinops schlieffenii (Peters, 1859)
Myonycteris relicta Bergmans, 1980
Myonycteris brachycephala (Bocage, 1889)
Casinycteris argynnis Thomas, 1910
Plerotes anchietae (Seabra, 1900)
Scotophilus dinganii (A. Smith, 1833)
Kerivoula lanosa (A. Smith, 1847)
Kerivoula cuprosa Thomas, 1912
Kerivoula eriophora (Heuglin, 1877)
Kerivoula argentata Tomes, 1861
Myotis goudoti (A. Smith, 1834)
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Paremballonura atrata (Peters, 1874)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Glauconycteris variegata (Tomes, 1861)
Kerivoula argentata Tomes, 1861
Kerivoula cuprosa Thomas, 1912
Kerivoula smithii Thomas, 1880
Kerivoula africana Dobson, 1878
Kerivoula eriophora (Heuglin, 1877)
Kerivoula phalaena Thomas, 1912
Glauconycteris variegata (Tomes, 1861)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
 VESPERTILIONIDAE Gray, 1821
 CHIROPTERA Blumenbach, 1779
Epomophorus Bennett, 1836
Plecotus E. Geoffroy St.-Hilaire, 1818
Laephotis Thomas, 1901
Nyctalus Bowdich, 1825
 RHINOPOMATIDAE Dobson, 1872
Coleura Peters, 1867
 MOLOSSIDAE Gervais, 1856
Myzopoda Milne-Edwards and A. Grandidier, 1878
Scotoecus Thomas, 1901
 PTEROPODIDAE Gray, 1821
Kerivoula Gray, 1842
Hypsignathus monstrosus H. Allen, 1862
Cloeotis percivali Thomas, 1901
Cloeotis Thomas, 1901
Coleura afra (Peters, 1852)
Coleura kibomalandy Goodman, Puechmairie, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012

- Emballonure de Madagascar
Emballonure des rochers
- Emballonure des Seychelles
Emballonuridés
Epomophore d'Angola
Epomophore d'Ansell
Epomophore de Dobson
Epomophore de Peters
Epomophore de Sanborn
Epomophore d'Ethiopie
Epomophore frugivore de Wahlberg
Epomophore nain
Epomophores
Epomophores de l'ouest
Faux vampire de l'ancien monde
Fer à cheval d'Hildebrandt
Fer à cheval du Kenya
Fer-à-Cheval d'Afrique orientale
Fer-à-Cheval de Darling
Fers-à-cheval
Fers-à-cheval
Feuille de Daubenton
Glaucoryctère à gorge blanche
Glaucoryctère argentée
Glaucoryctère d'Abo
Glaucoryctère de Béatrix
Glaucoryctère de Bibundi
Glaucoryctère de Curry
Glaucoryctère de Glen
Glaucoryctère de Machado
Glaucoryctère du Kenya
Glaucoryctère papillon
Glaucoryctère pie
Glaucoryctère sombre
- Glaucoryctère tacheté
Glaucoryctères
Grand Fer-à-Cheval
Grand minioptère
Grand minioptère africain
Grand minioptère malgache
Grand molosse à glandes caudales
Grand molosse à grandes oreilles
Grand rhinolophe
Grand Rhinopome
Grand scotophile africain
Grande noctule
Grande nyctère
grande Roussette des Mascareignes
Grande tadaride africaine
Grande Tadaride de Madagascar
Hipposidérédés
Hypsignates
Hypsignathe monstrueux
Kérivoule de Tanzanie
la petite roussette des Comores
La Phyllorhine cryptique de Madagascar
- la roussette de Livingstone
Lavie à ailes jaunes
le grand rhinolophe fer à cheval
Le Megaderme feuille
le vespertilion de Kuhl
le vespertilion de Leisler
le vespertilion pygmée
Lissonyctère d'Angola
Lissonyctère de Smith
Lissonyctère d'Ethiopie
Lissonyctère du Ruwenzori
Lissonyctère goliath
Lissonyctères
Macroglosse de Woermann
Mégadermatidés
Megaderme à ailes orangées
- Paremballonura atrata* (Peters, 1874)
Paremballonura tiavato (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)
Coleura seychellensis Peters, 1868
EMBALLONURIDAE Gervais, 1855
Epomophorus angolensis Gray, 1870
Epomophorus anselli Bergmans and Van Strien, 2004
Epomophorus dobsonii Bocage, 1889
Epomophorus crypturus Peters, 1852
Epomophorus grandis (Sanborn, 1950)
Epomophorus labiatus (Temminck, 1837)
Epomophorus wahlbergi (Sundevall, 1846)
Epomophorus minimus Claessen & De Vree, 1991
Epomophorus Bennett, 1836
Epomops Gray, 1866
MEGADERMATIDAE H. Allen, 1864
Rhinolophus hildebrandtii Peters, 1878
Rhinolophus hildebrandtii Peters, 1878
Rhinolophus deckenii Peters, 1868
Rhinolophus darlingi K. Andersen, 1905
RHINOLOPHIDAE Gray, 1825
Rhinolophus Lacépède, 1799
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Glaucorycteris alboguttata J.A. Allen, 1917
Glaucorycteris argentata (Dobson, 1875)
Glaucorycteris poensis (Gray, 1842)
Glaucorycteris beatrix Thomas, 1901
Glaucorycteris egeria Thomas, 1913
Glaucorycteris curryae Eger and Schlitter, 2001
Glaucorycteris gleni Peterson and Smith, 1973
Glaucorycteris machadoi Hayman, 1963
Glaucorycteris kenyacola Peterson, 1982
Glaucorycteris variegata (Tomes, 1861)
Glaucorycteris superba Hayman, 1939
Glaucorycteris atra Hassanin, Colombo, Gembu, Merle, Tu, Görföl, Musaba Akawa, Csorba, Kearney, Monadjem and Ing, 2017
Glaucorycteris humeralis J.A. Allen, 1917
Glaucorycteris Dobson, 1875
Rhinolophus ferrumequinum (Schreber, 1774)
Miniopterus inflatus Thomas, 1903
Miniopterus inflatus Thomas, 1903
Miniopterus majori Thomas, 1906
Mops (Chaerephon) major (Trouessart, 1897)
Otomops martiensseni (Matschie, 1897)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinopoma microphyllum (Brünnich, 1782)
Scotophilus nigrita (Schreber, 1774)
Nyctalus lasiopterus (Schreber, 1780)
Nycteris grandis Peters, 1865
Pteropus niger (Kerr, 1792)
Tadarida ventralis (Heuglin, 1861)
Tadarida fulminans (Thomas, 1903)
HIPPOSIDERIDAE Lydekker, 1891
Hypsignathus H. Allen, 1862
Hypsignathus monstrosus H. Allen, 1862
Kerivoula africana Dobson, 1878
Rousettus obliviosus Kock, 1978
Macronycteris cryptovalorona (Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016)
Pteropus livingstonii Gray, 1866
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Rhinolophus ferrumequinum (Schreber, 1774)
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Myonycteris angolensis (Bocage, 1898)
Myonycteris angolensis smithii (Thomas, 1908)
Myonycteris angolensis petraea (Bergmans, 1997)
Myonycteris angolensis ruwenzorii (Eisentraut, 1965)
Myonycteris angolensis goliath (Bergmans, 1997)
Myonycteris Matschie, 1899
Megaloglossus woermanni Pagenstecher, 1885
MEGADERMATIDAE H. Allen, 1864
Lavia frons (E. Geoffroy St.-Hilaire, 1810)

Megaderme à ailes orangées	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
Mégadermes à ailes orangées	<i>Lavia</i> Gray, 1838
Mégaloglosse de Woermann	<i>Megaloglossus woermanni</i> Pagenstecher, 1885
Mégaloglosses	<i>Megaloglossus</i> Pagenstecher, 1885
Microptère de Hayman	<i>Epomophorus intermedius</i> (Hayman, 1963)
Mimétille	<i>Mimetillus</i> Thomas, 1904
Mimétille de Moloney	<i>Mimetillus moloneyi</i> (Thomas, 1891)
Mimétille de Thomas	<i>Mimetillus thomasi</i> Hinton, 1920
Minioptère	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Minioptère à couronne	<i>Miniopterus inflatus</i> Thomas, 1903
Minioptère à longues ailes	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Minioptère d'Afrique australe	<i>Miniopterus fraterculus</i> Thomas and Schwann, 1906
Minioptère d'Afrique orientale	<i>Miniopterus africanus</i> Sanborn, 1936
Minioptère de Montagne d'Ambre	<i>Miniopterus ambohitrensis</i> Goodman, Ramasindrazana, Naughton and Appleton, 2015
Minioptère de Peterson	<i>Miniopterus petersoni</i> Goodman, Bradman, Maminirina, Ryan, Christidis & Appleton, 2008
Minioptère de São Tomé	<i>Miniopterus newtoni</i> Bocage, 1889
Minioptère de Schreibers	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Minioptère d'Eger	<i>Miniopterus egeri</i> Goodman, Ramasindrazana, Maminirina, Schoeman and Appleton, 2011
Minioptère des Comores	<i>Miniopterus griveaudi</i> Harrison, 1959
Minioptère du Maghreb	<i>Miniopterus maghrebensis</i> Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014
Minioptère du Natal	<i>Miniopterus natalensis</i> (A. Smith, 1833)
Minioptère malgache	<i>Miniopterus gleni</i> Peterson, Eger and Mitchell, 1995
Minioptère minuscule	<i>Miniopterus minor</i> Peters, 1867
Minioptères	<i>Miniopterus</i> Bonaparte, 1837
Minioptéridés	MINIOPTERIDAE Dobson, 1875
Molosse à crinière	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
Molosse à glandes caudales	<i>Mops (Chaerephon) bemmeleni</i> (Jentink, 1879)
Molosse à grandes oreilles	<i>Tadarida lobata</i> (Thomas, 1891)
Molosse à grandes oreilles de Madagascar	<i>Otomops madagascariensis</i> Dorst, 1953
Molosse d'Afrique du Sud	<i>Mops (Chaerephon) ansorgei</i> (Thomas, 1913)
Molosse d'Angola	<i>Mops (Mops) condylurus</i> (A. Smith, 1833)
Molosse d'Ansorge	<i>Mops (Chaerephon) ansorgei</i> (Thomas, 1913)
Molosse de Cestoni	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Molosse de Madagascar	<i>Tadarida fulminans</i> (Thomas, 1903)
Molosse de Midas	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Molosse d'Egypte	<i>Tadarida aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
Molosse d'Europe	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Molosse du Duc des Abruzzes	<i>Mops (Chaerephon) aloysiisabaudiae</i> (Festa, 1907)
Molosse du Nigéria	<i>Mops (Chaerephon) nigeriae</i> (Thomas, 1913)
Molosse du Zaïre	<i>Mops (Chaerephon) gallagheri</i> (Harrison, 1975)
Molosse géant	<i>Tadarida ventralis</i> (Heuglin, 1861)
Molosse Midas	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Molosse pâle	<i>Mops (Chaerephon) chapini</i> (J.A. Allen, 1917)
Molosse tacheté	<i>Mops (Chaerephon) bivittatus</i> (Heuglin, 1861)
Molosses	MOLOSSIDAE Gervais, 1856
Molosses	<i>Molossinae</i> Gervais, 1856
Molosses à grandes oreilles	<i>Otomops</i> Thomas, 1913
Molossidés	MOLOSSIDAE Gervais, 1856
Murin à bajoues argenté	<i>Glauconycteris argentata</i> (Dobson, 1875)
Murin à bajoues d'Abo	<i>Glauconycteris poensis</i> (Gray, 1842)
Murin à bajoues d'Allen	<i>Glauconycteris alboguttata</i> J.A. Allen, 1917
Murin à bajoues de Béatrice	<i>Glauconycteris beatrix</i> Thomas, 1901
Murin à bajoues de Bibundi	<i>Glauconycteris egeria</i> Thomas, 1913
Murin à bajoues de Glen	<i>Glauconycteris gleni</i> Peterson and Smith, 1973
Murin à bajoues du Kenya	<i>Glauconycteris kenyacola</i> Peterson, 1982
Murin à bajoues papillon	<i>Glauconycteris variegata</i> (Tomes, 1861)
Murin à bajoues pie	<i>Glauconycteris superba</i> Hayman, 1939
Murin à moustaches	<i>Myotis mystacinus</i> (Kuhl, 1817)
Murin à oreilles échancrées	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Murin d'Angola	<i>Cistugo seabrae</i> Thomas, 1912
Murin d'Angola	<i>Myotis tricolor</i> (Temminck, 1832)
Murin d'Anjouan	<i>Myotis anjouanensis</i> Dorst, 1960
Murin de Capaccini	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Murin de Du Bocage	<i>Myotis bocagii</i> (Peters, 1870)
Murin de Kock	<i>Myotis dieteri</i> M. Happold, 2005
Murin de Lesueur	<i>Cistugo lesueuri</i> Roberts, 1919
Murin de Madagascar	<i>Myotis goudoti</i> (A. Smith, 1834)
Murin de Scott	<i>Myotis scotti</i> Thomas, 1927
Murin de Welwitsch	<i>Myotis welwitschii</i> (Gray, 1866)
Murin d'Ethiopie	<i>Myotis morrisi</i> Hill, 1971
Murin du Maghreb	<i>Myotis punicus</i> Felten, 1977
Murin roux	<i>Myotis bocagii</i> (Peters, 1870)

- Murin tricolore
 Murin velu du Cap
 Murin Zenati

 Murins
 Murins à bajoues
 Myonyctère d'Afrique orientale
 Myonyctère de São Tomé
 Myonyctères
 Myoptère de Bini
 Myoptère de Daubenton
 Myoptère de Whitley
 Myoptère du Sénégal
 Myoptères
 Myotis de Welwitsch
 Myzopoda oreillard

 Myzopodidés
 Nanonyctère
 Noctophile d'Angola
 Noctophile de De Winton
 Noctophile du Botswana
 Noctophile du Namib
 Noctophiles
 Noctule de Leisler
 Noctule des Açores
 Noctule géante
 Noctules
 Nyctère à grandes oreilles
 Nyctère à longues oreilles
 Nyctère d'Aellen
 Nyctère d'Andersen
 Nyctère de Bates
 Nyctère de Dobson
 Nyctère de Gambie
 Nyctère de Ja
 Nyctère de la Thébaïde
 Nyctère de Parisi
 Nyctère de Vinson
 Nyctère de Wood
 Nyctère du Mozambique
 Nyctère égyptienne
 Nyctère hérissée
 Nyctère hirsute
 Nyctère moyen
 Nyctère naine
 Nyctères
 Nyctères
 Nyctéridés
 Nycticeinops
 Nycticeinops de Schlieffen
 Oreillard d'Angola
 Oreillard de De Winton
 Oreillard de Namibie
 Oreillard de Ténérife
 Oreillard d'Égypte
 Oreillard Des Balkans
 Oreillard d'Éthiopie
 Oreillard d'Hemprich
 Oreillard du Botswana
 Oreillard du Gaisler
 Oreillard du Namib
 Oreillards
 Oreillards d'Afrique
 Oreillards du désert
 Petit Epomophore
 Petit épomophore
 Petit Epomophore frugivore d'Angola
 Petit Fer-à-Cheval
 Petit microptère
 Petit minioptère
 Petit minioptère africain
 Petit minioptère malgache
 Petit molosse

 Petit molosse à glandes caudales

Myotis tricolor (Temminck, 1832)
Myotis tricolor (Temminck, 1832)
Myotis zenatus Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019
Myotis Kaup, 1829
Glauconycteris Dobson, 1875
Myonycteris relicta Bergmans, 1980
Myonycteris brachycephala (Bocage, 1889)
Myonycteris Matschie, 1899
Myopterus whitleyi (Scharff, 1900)
Myopterus daubentonii Desmarest, 1820
Myopterus whitleyi (Scharff, 1900)
Myopterus daubentonii Desmarest, 1820
Myopterus E. Geoffroy St.-Hilaire, 1818
Myotis welwitschii (Gray, 1866)
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
 MYZOPODIDAE Thomas, 1904
Nanonycteris veldkampii (Jentink, 1888)
Laephotis angolensis Monard, 1935
Laephotis wintoni Thomas, 1901
Laephotis botswanae Setzer, 1971
Laephotis namibensis Setzer, 1971
Laephotis Thomas, 1901
Nyctalus leisleri (Kuhl, 1817)
Nyctalus azoreum (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus Bowdich, 1825
Nycteris macrotis Dobson, 1876
Nycteris aurita (K. Andersen, 1912)
Nycteris intermedia Aellen, 1959
Nycteris aurita (K. Andersen, 1912)
Nycteris arge Thomas, 1903
Nycteris macrotis Dobson, 1876
Nycteris gambiensis (K. Andersen, 1912)
Nycteris major (K. Andersen, 1912)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris parisii (de Beaux, 1924)
Nycteris vinsoni Dalquest, 1965
Nycteris woodi K. Andersen, 1914
Nycteris vinsoni Dalquest, 1965
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris hispida (Schreber, 1774)
Nycteris hispida (Schreber, 1774)
Nycteris intermedia Aellen, 1959
Nycteris nana (K. Andersen, 1912)
 NYCTERIDAE Van der Hoeven, 1855
Nycteris G. Cuvier and E. Geoffroy, 1795
 NYCTERIDAE Van der Hoeven, 1855
Nycticeinops Hill & Harrison, 1987
Nycticeinops schlieffenii (Peters, 1859)
Laephotis angolensis Monard, 1935
Laephotis wintoni Thomas, 1901
Laephotis namibensis Setzer, 1971
Plecotus teneriffae Barrett-Hamilton, 1907
Plecotus christii Gray, 1838
Plecotus kolombatovici Dulic, 1980
Plecotus balensis Kruskop & Lavrenchenko, 2000
Otonycteris hemprichii Peters, 1859
Laephotis botswanae Setzer, 1971
Plecotus gaisleri Benda, Kiefer, Hanak & Veith, 2004
Laephotis namibensis Setzer, 1971
Plecotus E. Geoffroy St.-Hilaire, 1818
Laephotis Thomas, 1901
Otonycteris Peters, 1859
Epomophorus minimus Claessen & De Vree, 1991
Epomophorus labiatus (Temminck, 1837)
Epomophorus grandis (Sanborn, 1950)
Rhinolophus hipposideros (Bechstein, 1800)
Epomophorus pusillus Peters, 1868
Miniopterus fraterculus Thomas and Schwann, 1906
Miniopterus minor Peters, 1867
Miniopterus manavi Thomas, 1906
Mormopterus francoismoutoui Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst, Bowie, 2008
Mops (Chaerephon) pumilus (Cretzschmar, 1826)

Petit molosse de Port Louis	<i>Mormopterus acetabulosus</i> (Hermann, 1804)
Petit murin brun	<i>Myotis tricolor</i> (Temminck, 1832)
Petit Myonyctère	<i>Myonycteris torquata</i> (Dobson, 1878)
Petit Renard volant de Mascareignes	<i>Pteropus subniger</i> (Kerr, 1792)
Petit renard volant de Mascarene	<i>Pteropus subniger</i> (Kerr, 1792)
Petit rhinolophe	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Petit rhinolophe fer à cheval	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Petit Rhinopome	<i>Rhinopoma cystops</i> Thomas, 1903
Petit scotophile africain	<i>Scotophilus viridis</i> (Peters, 1852)
Petite Chauve-souris frugivore à collier	<i>Myonycteris torquata</i> (Dobson, 1878)
Petite Chauve-souris jaune des maisons	<i>Scotophilus borbonicus</i> (E. Geoffroy St.-Hilaire, 1803)
Petite Chauve-souris laineuse	<i>Kerivoula lanosa</i> (A. Smith, 1847)
Petite Noctule	<i>Nyctalus leisleri</i> (Kuhl, 1817)
Petite tadaride	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
Petite tadaride de Madagascar	<i>Mops (Chaerephon) leucogaster</i> (A. Grandidier, 1869)
Petite vache-volante	<i>Nanonycteris veldkampii</i> (Jentink, 1888)
Phyllorhine à petites oreilles	<i>Cloeotis percivali</i> Thomas, 1901
Phyllorhine d'Aba	<i>Hipposideros abae</i> J.A. Allen, 1917
Phyllorhine de Benito	<i>Hipposideros beatus</i> (K. Andersen, 1906)
Phyllorhine de Caferie	<i>Hipposideros caffer</i> (Sundevall, 1846)
Phyllorhine de Percival	<i>Cloeotis percivali</i> Thomas, 1901
Phyllorhine de Sundevall	<i>Hipposideros caffer</i> (Sundevall, 1846)
Phyllorhine fuligineux	<i>Hipposideros fuliginosus</i> (Temminck, 1853)
Phyllorhines	<i>Hipposideros</i> Gray, 1831
Phyllorine	<i>HIPPOSIDERIDAE</i> Lydekker, 1891
Phyllorine à grandes oreilles	<i>Hipposideros megalotis</i> (Heuglin, 1861)
Phyllorine à grandes oreilles d'Ethiopie	<i>Hipposideros megalotis</i> (Heuglin, 1861)
Phyllorine à queue courte	<i>Hipposideros curtus</i> G.M. Allen, 1921
Phyllorine cendrée	<i>Hipposideros tephros</i> (Cabrera, 1906)
Phyllorine cyclope	<i>Doryrhina cyclops</i> (Temminck, 1853)
Phyllorine d'Aellen	<i>Hipposideros marisae</i> Aellen, 1954
Phyllorine d'Afrique occidentale	<i>Hipposideros jonesi</i> Hayman, 1947
Phyllorine de Commerson	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Phyllorine de Commerson	<i>Macronycteris gigas</i> (Wagner, 1845)
Phyllorine de Jones	<i>Hipposideros jonesi</i> Hayman, 1947
Phyllorine de Lamotte	<i>Hipposideros lamottei</i> Brosset, 1985
Phyllorine de Noack	<i>Hipposideros ruber</i> (Noack, 1893)
Phyllorine d'Eisentraut	<i>Doryrhina camerunensis</i> (Eisentraut, 1956)
Phyllorine des Cyclopes	<i>Doryrhina cyclops</i> (Temminck, 1853)
Phyllorine du Cameroun	<i>Doryrhina camerunensis</i> (Eisentraut, 1956)
Phyllorine du Mont Nimba	<i>Hipposideros lamottei</i> Brosset, 1985
Phyllorine fuligineuse	<i>Hipposideros fuliginosus</i> (Temminck, 1853)
Phyllorine naine	<i>Hipposideros beatus</i> (K. Andersen, 1906)
Phyllorine rouge	<i>Hipposideros ruber</i> (Noack, 1893)
Phyllorine sombre	<i>Hipposideros fuliginosus</i> (Temminck, 1853)
Pipistrelle à ailes blanches	<i>Pseudoromicia tenuipinnis</i> (Peters, 1872)
Pipistrelle à grosse tête	<i>Nycticeinops crassulus</i> (Thomas, 1904)
Pipistrelle ariel	<i>Hypsugo ariel</i> (Thomas, 1904)
Pipistrelle commune	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Pipistrelle d'Aellen	<i>Pipistrellus (Pipistrellus) inexpectatus</i> Aellen, 1959
Pipistrelle d'Afrique centrale	<i>Pipistrellus (Pipistrellus) inexpectatus</i> Aellen, 1959
Pipistrelle d'Afrique occidentale	<i>Hypsugo musciculus</i> (Thomas, 1913)
Pipistrelle d'Anchieta	<i>Neoromicia anchietae</i> (Seabra, 1900)
Pipistrelle de Grandidier	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
Pipistrelle de Guinée	<i>Neoromicia guineensis</i> (Bocage, 1889)
Pipistrelle de Kuhl	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Pipistrelle de Hulbe	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Pipistrelle de Madère	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Pipistrelle de Racey	<i>Pipistrellus (Pipistrellus) raceyi</i> Bates, Ratrimomanarivo, Harrison and Goodman, 2006
Pipistrelle de Rendall	<i>Pseudoromicia rendalli</i> (Thomas, 1889)
Pipistrelle de Rueppel	<i>Vansonia rueppellii</i> (J.B. Fischer, 1829)
Pipistrelle de Rüppell	<i>Vansonia rueppellii</i> (J.B. Fischer, 1829)
Pipistrelle de Savi	<i>Hypsugo savii</i> (Bonaparte, 1837)
Pipistrelle de Somalie	<i>Neoromicia somalica</i> (Thomas, 1901)
Pipistrelle de Tanzanie	<i>Pipistrellus (Pipistrellus) permixtus</i> Aellen, 1957
Pipistrelle d'Eisentraut	<i>Nycticeinops eisentrauti</i> (Hill, 1968)
Pipistrelle des Melcks	<i>Laephotis stanleyi</i> (Goodman, Kearney, Ratsimbazafy and Hassanin, 2017)
Pipistrelle du Kenya	<i>Pipistrellus (Pipistrellus) aero</i> Heller, 1912
Pipistrelle du Miombo	<i>Neoromicia anchietae</i> (Seabra, 1900)
Pipistrelle géante	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Pipistrelle hespéride	<i>Pipistrellus (Pipistrellus) hesperidus</i> (Temminck, 1840)
Pipistrelle jaune	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
Pipistrelle minuscule	<i>Pipistrellus (Pipistrellus) nanulus</i> Thomas, 1904

- Pipistrelle murine
 Pipistrelle naine
 Pipistrelle naine aux ailes brunes
 Pipistrelle pygmée
 Pipistrelle rouille
 Pipistrelle rustique
 Pipistrelle soleil
 Pipistrelle soprane
 Pipistrelle zouloue
 Pipistrelles
 Plérote d'Anchieta
 Plérotés
 Prétopodidés
 Pseudo-vampire à nez en cœur
 Renard volant d'Aldabra
 Renard volant de l'île de Mascarene
 Renard Volant de l'île de Rodriguez
 Renard volant de Madagascar
 Renard volant de Maurice
 Renard volant de Pemba
 Renard volant de Rodrigues
 Renard volant des Comores
 Renard volant des Mascaraïnes
 Renard volant des Seychelles
 Renard volant malgache
 Renard volant noir
 Renard volant roux
 Renards volants
 Rhinolophe à selle pointue
 Rhinolophe Alcyon
 Rhinolophe d'Afrique du Sud
 Rhinolophe d'Afrique orientale
 Rhinolophe de Blasius
 Rhinolophe de brousse
 Rhinolophe de Cafrerie
 Rhinolophe de Cafrerie
 Rhinolophe de Cretzschmar
 Rhinolophe de Darling
 Rhinolophe de Decken
 Rhinolophe de Dent
 Rhinolophe de Geoffroy
 Rhinolophe de Guinée
 Rhinolophe de Hill
 Rhinolophe de Lander
 Rhinolophe de Lofa
 Rhinolophe de MacLaud
 Rhinolophe de Mehely
 Rhinolophe de Roumanie
 Rhinolophe de Rüppell
 Rhinolophe de Sakeji
 Rhinolophe de Swinny
 Rhinolophe de Tanga

 Rhinolophe des forêts
 Rhinolophe d'Halcyon
 Rhinolophe d'Hildebrandt
 Rhinolophe du Bushveld
 Rhinolophe du Cap
 Rhinolophe du Congo
 Rhinolophe du Ruwenzori
 Rhinolophe du Sénégal
 Rhinolophe du Veld
 Rhinolophe du Ziama

 Rhinolophe éloquent
 Rhinolophe euryale
 Rhinolophe sylvestre
 Rhinolophe trident
 Rhinolophes
 Rhinolophes
 Rhinolophidés
 Rhinopoma moyen
 Rhinopomatidés
 Rhinopome de Hardwicke
 Rhinopome de MacInnes
 Rhinopome microphyllé
- Hypsugo musciculus* (Thomas, 1913)
Afronycteris nana (Peters, 1852)
Afronycteris nana (Peters, 1852)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Afronycteris helios (Heller, 1912)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Neoromicia zuluensis (Roberts, 1924)
Pipistrellus Kaup, 1829
Plerotes anchietae (Seabra, 1900)
Plerotes K. Andersen, 1910
 PTEROPODIDAE Gray, 1821
Cardioderma cor (Peters, 1872)
Pteropus aldabrensis True, 1893
Pteropus niger (Kerr, 1792)
Pteropus rodricensis Dobson, 1878
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Pteropus niger (Kerr, 1792)
Pteropus voeltzkowi Matschie, 1909
Pteropus rodricensis Dobson, 1878
Pteropus livingstonii Gray, 1866
Pteropus niger (Kerr, 1792)
Pteropus seychellensis A. Milne-Edwards, 1877
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Pteropus livingstonii Gray, 1866
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Pteropus Brisson, 1762
Rhinolophus blasii Peters, 1867
Rhinolophus alcyone Temminck, 1853
Rhinolophus capensis Lichtenstein, 1823
Rhinolophus deckenii Peters, 1868
Rhinolophus blasii Peters, 1867
Rhinolophus simulator K. Andersen, 1904
Hipposideros caffer (Sundevall, 1846)
Hipposideros tephros (Cabrera, 1906)
Rhinolophus clivus Cretzschmar, 1828
Rhinolophus darlingi K. Andersen, 1905
Rhinolophus deckenii Peters, 1868
Rhinolophus denti Thomas, 1904
Rhinolophus clivus Cretzschmar, 1828
Rhinolophus guineensis Eisentraut, 1960
Rhinolophus hilli Aellen, 1973
Rhinolophus landeri Martin, 1838
Rhinolophus hillorum Koopman, 1989
Rhinolophus macclaudi Pousargues, 1898
Rhinolophus mehelyi Matschie, 1901
Rhinolophus mehelyi Matschie, 1901
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus sakejiensis Cotterill, 2002
Rhinolophus swinnyi Gough, 1908
Rhinolophus maendeleo Kock, Csorba and Howell, 2000
Rhinolophus mehelyi Matschie, 1901
Rhinolophus alcyone Temminck, 1853
Rhinolophus hildebrandtii Peters, 1878
Rhinolophus simulator K. Andersen, 1904
Rhinolophus capensis Lichtenstein, 1823
Rhinolophus adami Aellen and Brosset, 1968
Rhinolophus ruwenzorii J. Eric Hill, 1942
Rhinolophus guineensis Eisentraut, 1960
Rhinolophus simulator K. Andersen, 1904
Rhinolophus ziama Fahr, Vierhaus, Hutterer and Kock, 2002
Rhinolophus eloquens K. Andersen, 1905
Rhinolophus euryale Blasius, 1853
Rhinolophus silvestris Aellen, 1959
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
 RHINOLOPHIDAE Gray, 1825
Rhinolophus Lacépède, 1799
 RHINOLOPHIDAE Gray, 1825
Rhinopoma cystops Thomas, 1903
 RHINOPOMATIDAE Dobson, 1872
Rhinopoma cystops Thomas, 1903
Rhinopoma macinnesi Hayman, 1937
Rhinopoma microphyllum (Brünnich, 1782)

Rhinopomes	<i>RHINOPOMATIDAE</i> Dobson, 1872
Rhinopomes	<i>Rhinopoma</i> E. Geoffroy St.-Hilaire, 1818
Rougette	<i>Pteropus subniger</i> (Kerr, 1792)
Roussette à collet rouge	<i>Pteropus subniger</i> (Kerr, 1792)
Roussette à épaulettes d'Angola	<i>Epomophorus angolensis</i> Gray, 1870
Roussette à épaulettes de Peters	<i>Epomophorus crypturus</i> Peters, 1852
Roussette à épaulettes de Wahlberg	<i>Epomophorus wahlbergi</i> (Sundevall, 1846)
Roussette à long poils du Ruwenzori	<i>Myonycteris angolensis ruwenzorii</i> (Eisentraut, 1965)
Roussette à longs poils	<i>Myonycteris angolensis</i> (Bocage, 1898)
Roussette à longs poils	<i>Stenonycteris lanosus</i> (Thomas, 1906)
Roussette à petit palais	<i>Casinycteris argynnis</i> Thomas, 1910
Roussette d'Angola	<i>Myonycteris angolensis</i> (Bocage, 1898)
Roussette de Bocage	<i>Myonycteris angolensis</i> (Bocage, 1898)
Roussette de Dobson	<i>Epomophorus dobsonii</i> Bocage, 1889
Roussette de Madagascar	<i>Rousettus madagascariensis</i> G. Grandidier, 1929
Roussette de Pohle	<i>Casinycteris ophiodon</i> (Pohle, 1943)
Roussette de Zenker	<i>Scotonycteris zenkeri</i> Matschie, 1894
Roussette d'Égypte	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Roussette des Comores	<i>Rousettus obliviosus</i> Kock, 1978
Roussette des palmiers malgache	<i>Eidolon dupreanum</i> (Schegel, 1867)
Roussette des Seychelles	<i>Pteropus seychellensis</i> A. Milne-Edwards, 1877
Roussette du Kenya	<i>Myonycteris relicta</i> Bergmans, 1980
Roussette du Ruwenzori	<i>Stenonycteris lanosus</i> (Thomas, 1906)
Roussette égyptienne	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Roussette foncée	<i>Pteropus subniger</i> (Kerr, 1792)
Roussette jaune de Madagascar	<i>Eidolon dupreanum</i> (Schegel, 1867)
Roussette labiale	<i>Epomophorus labiatus</i> (Temminck, 1837)
Roussette labiée	<i>Epomophorus labiatus</i> (Temminck, 1837)
Roussette malgache couleur paille	<i>Eidolon dupreanum</i> (Schegel, 1867)
Roussette naine de Hayman	<i>Epomophorus intermedius</i> (Hayman, 1963)
Roussette naine de Peters	<i>Epomophorus pusillus</i> Peters, 1868
Roussette naine de Veldkamp	<i>Nanonycteris veldkampii</i> (Jentink, 1888)
Roussette noire	<i>Pteropus niger</i> (Kerr, 1792)
Roussette rougeâtre	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
Roussettes	<i>Rousettus</i> Gray, 1821
Roussettes à collier	<i>Myonycteris</i> Matschie, 1899
Roussettes des palmiers	<i>Eidolon Rafinesque</i> , 1815
Roussettes géantes	<i>Pteropus</i> Brisson, 1762
Roussettes naines	<i>Nanonycteris</i> Matschie, 1899
Saccolaime	<i>Saccolaimus</i> Temminck, 1838
Saccolaime de Pel	<i>Saccolaimus peli</i> (Temminck, 1853)
Sauromys	<i>Sauromys</i> Roberts, 1917
Sauromys à jugulaire	<i>Mormopterus jugularis</i> (Peters, 1865)
Sauromys à tête plate	<i>Platymops setiger</i> (Peters, 1878)
Sauromys de Peters	<i>Mormopterus jugularis</i> (Peters, 1865)
Sauromys de Roberts	<i>Sauromys petrophilus</i> (Roberts, 1917)
Sauromys du Natal	<i>Mormopterus acetabulosus</i> (Hermann, 1804)
Scotonyctère de Zenker	<i>Scotonycteris zenkeri</i> Matschie, 1894
Scotoèque à ailes pâles	<i>Scotoecus albofuscus</i> (Thomas, 1890)
Scotoèque à gorge blanche	<i>Scotoecus albigula</i> Thomas, 1909
Scotoèque de Hinde	<i>Scotoecus hindei</i> Thomas, 1901
Scotoèque hirundine	<i>Scotoecus hirundo</i> (de Winton, 1899)
Scotoèques	<i>Scotoecus</i> Thomas, 1901
Scotonyctère à dents de serpent	<i>Casinycteris ophiodon</i> (Pohle, 1943)
Scotonyctère de Bergmans	<i>Scotonycteris bergmansi</i> Hassanin, Khouider, Gembu, Goodman, Kado, Nesi, Pourrut, Nakouné and Bonillo, 2015
Scotonyctère de Hayman	<i>Scotonycteris occidentalis</i> Hayman, 1947
Scotonyctères	<i>Scotonycteris</i> Matschie, 1894
Scotophile à ailes noires	<i>Scotoecus hirundo</i> (de Winton, 1899)
Scotophile à ventre blanc	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
Scotophile à ventre jaune	<i>Scotophilus dinganii</i> (A. Smith, 1833)
Scotophile africain	<i>Scotophilus dinganii</i> (A. Smith, 1833)
Scotophile d'Andadoany	<i>Scotophilus tandrefana</i> Goodman, Jenkins and Ratrimomanarivo, 2005
Scotophile de Bourbon	<i>Scotophilus borbonicus</i> (E. Geoffroy St.-Hilaire, 1803)
Scotophile de Madagascar	<i>Scotophilus borbonicus</i> (E. Geoffroy St.-Hilaire, 1803)
Scotophile de Madagascar	<i>Scotophilus robustus</i> A. Milne-Edwards, 1881
Scotophile de Marovaza	<i>Scotophilus marovaza</i> Goodman, Ratrimomanarivo and Randrianandrianina, 2006
Scotophile de Robbins	<i>Scotophilus nucella</i> Robbins, 1983
Scotophile des hauteurs	<i>Scotophilus nucella</i> Robbins, 1983
Scotophile du crépuscule	<i>Scotoecus albofuscus</i> (Thomas, 1890)
Scotophile noisette	<i>Scotophilus nux</i> Thomas, 1904
Scotophile nux	<i>Scotophilus nux</i> Thomas, 1904
Scotophile robuste	<i>Scotophilus robustus</i> A. Milne-Edwards, 1881

- Scotophile verdâtre
 Scotophiles
 Sérotine à ailes blanches
 Sérotine à grands yeux
 Sérotine brune d'Afrique
 Sérotine brun-noire
 Sérotine de Botta
 Sérotine de Flower
 Sérotine de Grandidier
 Sérotine de Guinée
 Sérotine de Lagos
 Sérotine de Madagascar
 Sérotine de Rendall
 Sérotine de Sakahara

 Sérotine de Somalie
 Sérotine des aloes
 Sérotine des maisons à longue queue
 Sérotine du Cap
 Sérotine du Soudan
 Sérotine hottentote
 Sérotine hottentote
 Sérotine isabelle
 Sérotine isabelline
 Sérotine jaune
 Sérotine méditerranéen
 Sérotine minuscule
 Sérotines
 Tadarida de Midas
 Tadaride à doigts blancs
 Tadaride à glande caudale
 Tadaride à lèvres ridées
 Tadaride à oreillettes
 Tadaride à queue libre de Madagascar
 Tadaride à tête plate de Peters
 Tadaride à ventre blanc
 Tadaride à ventre blanc
 Tadaride bornée
 Tadaride d'Allen
 Tadaride d'Angola
 Tadaride d'Ansorge
 Tadaride de Cestoni
 Tadaride de Chapin
 Tadaride de Gallagher
 Tadaride de La Réunion

 Tadaride de l'Afrique orientale
 Tadaride de Madagascar
 Tadaride de Madagascar à ventre blanc
 Tadaride de Medje
 Tadaride de Mongalla
 Tadaride de Peters
 Tadaride de Peterson
 Tadaride de Railer
 Tadaride de São Tomé
 Tadaride de Spurrell
 Tadaride de Thomas
 Tadaride de Trevor
 Tadaride d'Egypte
 Tadaride des Seychelles
 Tadaride d'Europe
 Tadaride du Duc des Abruzzes
 Tadaride du Kenya
 Tadaride du Niangara
 Tadaride du Nigeria
 Tadaride géante d'Afrique
 Tadaride Midas
 Tadaride naine
 Tadaride rouge ou noir

 Tadaride rousse
 Tadaride tachetée
 Tadarides
 Tadarides
 Tadarides
 Tadarides
 Tadarides

Scotophilus viridis (Peters, 1852)
Scotophilus Leach, 1821
Pseudoromicia tenuipinnis (Peters, 1872)
Eptesicus (Cnephaeus) platyops (Thomas, 1901)
Pseudoromicia brunnea (Thomas, 1880)
Pseudoromicia brunnea (Thomas, 1880)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Nycticeinops grandidieri (Dobson, 1876)
Neoromicia guineensis (Bocage, 1889)
Eptesicus (Cnephaeus) platyops (Thomas, 1901)
Laephotis matroka (Thomas and Schwann, 1905)
Pseudoromicia rendalli (Thomas, 1889)
Laephotis malagasyensis (Peterson, Eger and Mitchell, 1995)
Neoromicia somalica (Thomas, 1901)
Neoromicia zuluensis (Roberts, 1924)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Laephotis capensis (A. Smith, 1829)
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Nycticeinops grandidieri (Dobson, 1876)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Neoromicia guineensis (Bocage, 1889)
Eptesicus Rafinesque, 1820
Mops (Mops) midas (Sundevall, 1843)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Mops (Chaerephon) bemmeleni (Jentink, 1879)
Mormopterus acetabulosus (Hermann, 1804)
Mops (Chaerephon) major (Trouessart, 1897)
Mops (Mops) leucostigma (G.M. Allen, 1918)
Platymops setiger (Peters, 1878)
Mops (Chaerephon) leucogaster (A. Grandidier, 1869)
Mops (Mops) niveiventer Cabrera and Ruxton, 1926
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Mops (Mops) trevori J.A. Allen, 1917
Mops (Mops) condylurus (A. Smith, 1833)
Mops (Chaerephon) ansorgei (Thomas, 1913)
Tadarida teniotis (Rafinesque, 1814)
Mops (Chaerephon) chapini (J.A. Allen, 1917)
Mops (Chaerephon) gallagheri (Harrison, 1975)
Mormopterus francoismoutoui Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst, Bowie, 2008
Tadarida lobata (Thomas, 1891)
Mormopterus jugularis (Peters, 1865)
Mops (Chaerephon) leucogaster (A. Grandidier, 1869)
Mops (Mops) conigicus J.A. Allen, 1917
Mops (Mops) demonstrator (Thomas, 1903)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Mops (Xiphonycteris) petersoni (El Rayah, 1981)
Mops (Xiphonycteris) thersites (Thomas, 1903)
Mops (Chaerephon) tomensis (Juste and Ibáñez, 1993)
Mops (Xiphonycteris) spurrelli (Dollman, 1911)
Tadarida fulminans (Thomas, 1903)
Mops (Mops) trevori J.A. Allen, 1917
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Mops (Chaerephon) pusillus (Miller, 1902)
Tadarida teniotis (Rafinesque, 1814)
Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
Tadarida lobata (Thomas, 1891)
Mops (Mops) niangarae J.A. Allen, 1917
Mops (Chaerephon) nigeriae (Thomas, 1913)
Tadarida ventralis (Heuglin, 1861)
Mops (Mops) midas (Sundevall, 1843)
Mops (Xiphonycteris) nanulus J.A. Allen, 1917
Mops (Chaerephon) jobimena (Goodman and Cardiff, 2004)
Mops (Chaerephon) russatus (J.A. Allen, 1917)
Mops (Chaerephon) bivittatus (Heuglin, 1861)
Mops Lesson, 1842
Mops (Chaerephon) Dobson, 1874
Mormopterus Peters, 1865
Tadarida Rafinesque, 1814

Tadarides à tête plate	<i>Platymops</i> Thomas, 1906
Taphien à croupe nue	<i>Taphozous nudiventris</i> Cretzschmar, 1830
Taphien à ventre nu	<i>Taphozous nudiventris</i> Cretzschmar, 1830
Taphien de Hamilton	<i>Taphozous hamiltoni</i> Thomas, 1920
Taphien de Hildegarde	<i>Taphozous hildegardeae</i> Thomas, 1909
Taphien de l'île de France	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
Taphien de l'île Maurice	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
Taphien de Maurice	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
Taphien des tombeaux	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
Taphien perforé	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
Taphiens	EMBALLONURIDAE Gervais, 1855
Taphiens	<i>Taphozous</i> E. Geoffroy St.-Hilaire, 1818
Triaenops	<i>Triaenops</i> Dobson, 1871
Triaenops à grandes oreilles	<i>Paratriaenops auritus</i> (G. Grandidier, 1912)
Triaenops de Madagascar	<i>Paratriaenops furculus</i> (Trouessart, 1907)
Triaenops de Paulian	<i>Paratriaenops pauliani</i> (Goodman and Ranivo, 2008)
Triaenops de Trouessart	<i>Paratriaenops furculus</i> (Trouessart, 1907)
Triaenops malgache	<i>Paratriaenops furculus</i> (Trouessart, 1907)
Triaenops roux	<i>Triaenops menamena</i> Goodman and Ranivo, 2009
Trident	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
Trident de Patrizi	<i>Asellia patrizii</i> de Beaux, 1931
Trident du désert	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
Tridents	<i>Asellia</i> Gray, 1838
Vespère à grosse tête	<i>Nycticeinops crassulus</i> (Thomas, 1904)
Vespère ariel	<i>Hypsugo ariel</i> (Thomas, 1904)
Vespère d'Anchieta	<i>Neoromicia anchietae</i> (Seabra, 1900)
Vespère de Savi	<i>Hypsugo savii</i> (Bonaparte, 1837)
Vespère d'Eisentraut	<i>Nycticeinops eisentrauti</i> (Hill, 1968)
Vespère murin	<i>Hypsugo musciculus</i> (Thomas, 1913)
Vespères	<i>Hypsugo</i> Kolenati, 1856
Vespertilion à moustaches	<i>Myotis mystacinus</i> (Kuhl, 1817)
Vespertilion à oreilles échancrées	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Vespertilion d'Angola	<i>Myotis tricolor</i> (Temminck, 1832)
Vespertilion de Capaccini	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Vespertilion de l'île d'Anjouan	<i>Myotis anjouanensis</i> Dorst, 1960
Vespertilion de Madagascar	<i>Myotis goudoti</i> (A. Smith, 1834)
Vespertilion de Morris	<i>Myotis morrisi</i> Hill, 1971
Vespertilion de Scott	<i>Myotis scotti</i> Thomas, 1927
Vespertilion de Welwitsch	<i>Myotis welwitschii</i> (Gray, 1866)
Vespertilion des montagnes	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Vespertilion Doré	<i>Myzopoda aurita</i> Milne-Edwards and A. Grandidier, 1878
Vespertilion échancré	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Vespertilion isabelle	<i>Eptesicus (Cnephæus) isabellinus</i> (Temminck, 1840)
Vespertilionidés	VESPERTILIONIDAE Gray, 1821
Vespertillons dorés	MYZOPODIDAE Thomas, 1904

Frisian

Azoaren reade flearmûs	<i>Nyctalus azoreum</i> (Thomas, 1901)
Blasius' hoefizernoas	<i>Rhinolophus blasii</i> Peters, 1867
Boskflearmûs	<i>Nyctalus leisleri</i> (Kuhl, 1817)
Botta's lette flearmûs	<i>Eptesicus (Cnephæus) bottae</i> (Peters, 1869)
Burdflearmûs	<i>Myotis mystacinus</i> (Kuhl, 1817)
Gewoane dwerchflearmûs	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Grutfuottige flearmûs	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Grutte hoefizernoas	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Grutte reade flearmûs	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Hanaks dwerchflearmûs	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Isabelkleurde lette flearmûs	<i>Eptesicus (Cnephæus) isabellinus</i> (Temminck, 1840)
Kanaryske grutearflearmûs	<i>Plecotus teneriffae</i> Barrett-Hamilton, 1907
Kealbûktimpelflearmûs	<i>Taphozous nudiventris</i> Cretzschmar, 1830
Lytse Alpenflearmûs	<i>Hypsugo savii</i> (Bonaparte, 1837)
Lytse dwerchflearmûs	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Lytse hoefizernoas	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Madeira dwerchflearmûs	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Maghreb mûsearflearmûs	<i>Myotis punicus</i> Felten, 1977
Mehely's hoefizernoas	<i>Rhinolophus mehelyi</i> Matschie, 1901
Muskusbulflearmûs	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Nylrûsette	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Pearse hoefizernoas	<i>Rhinolophus euryale</i> Blasius, 1853
Schreibers'langwjukflearmûs	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Westlike breedearflearmûs	<i>Barbastella barbastellus</i> (Schreber, 1774)
Woastynggrutearflearmûs	<i>Otonycteris hemprichii</i> Peters, 1859
Wytrâne dwerchflearmûs	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)

Ynsniene flearmús

Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)

Galician (Spain)

Morcego anano
 Morcego bigotudo
 Morcego común
 Morcego das covas
 Morcego das fragas
 Morcego das hortas de Botta
 Morcego das hortas meridional
 Morcego de beira branca
 Morcego de bigotes
 Morcego de borde claro
 Morcego de ferradura de Blasius
 Morcego de ferradura grande
 Morcego de ferradura mediano
 Morcego de ferradura mediterráneo
 Morcego de ferradura pequeno
 Morcego de Hanak
 Morcego de Leisler
 Morcego de Madeira
 Morcego de orelas fendidas
 Morcego de pés grandes
 Morcego de tumba de cú espido
 Morcego do monte
 Morcego exipcio
 Morcego mediterráneo de ferradura
 Morcego montañoiro
 Morcego pequeno de ferradura
 Morcego rabudo
 Morcego rateiro africano
 Nóctulo das Azores
 Nóctulo pequeno
 Nóctulo xigante
 Orelludo canario
 Orelludo de Hemprich

Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Myotis mystacinus (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Rhinolophus blasii Peters, 1867
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis capaccinii (Bonaparte, 1837)
Taphozous nudiventris Cretzschmar, 1830
Hypsugo savii (Bonaparte, 1837)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rhinolophus euryale Blasius, 1853
Hypsugo savii (Bonaparte, 1837)
Rhinolophus hipposideros (Bechstein, 1800)
Tadarida teniotis (Rafinesque, 1814)
Myotis punicus Felten, 1977
Nyctalus azoreum (Thomas, 1901)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus lasiopterus (Schreber, 1780)
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859

Georgian

აზორული მეღამურა
 ბლაზის ცხვირნალა
 გიგანტური მეღამურა
 დიდი ცხვირნალა
 ეგვიპტური მფრინავი ძაღლი
 ევროპული მაჩუბთელა
 ევროპული ტადარიდა
 კანარიული ყურა
 კაპაჩინის მღამიობი
 კულის
 მადეირული ღამორი
 მალრიბული მღამიობი
 მეპელის ცხვირნალა
 მცირე მეღამურა
 მცირე ცხვირნალა
 პაჩია ღამორი
 სავის ღამორი
 სამფერი მღამიობი
 სამხრეთული ცხვირნალა
 ტყის მეგვიანე
 უდაბნოს მეგვიანე
 ულვამა მღამიობი
 მიშველმუცელა ქარქაშკუდა
 ჩვეულებრივი ფრთაგრძელი
 ხმელთაშუაზღვიური ღამორი
 ჯუჯა ღამორი
 პანაკის ღამორი
 ჰემპრიხის ისარყურა

Nyctalus azoreum (Thomas, 1901)
Rhinolophus blasii Peters, 1867
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Barbastella barbastellus (Schreber, 1774)
Tadarida teniotis (Rafinesque, 1814)
Plecotus teneriffae Barrett-Hamilton, 1907
Myotis capaccinii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Myotis punicus Felten, 1977
Rhinolophus mehelyi Matschie, 1901
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Hypsugo savii (Bonaparte, 1837)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Rhinolophus euryale Blasius, 1853
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Myotis mystacinus (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Miniopterus schreibersii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Otonycteris hemprichii Peters, 1859

German

Aba-Rundblattnase	<i>Hipposideros abae</i> J.A. Allen, 1917
Abendfledermäuse	<i>Nycticeinops</i> Hill & Harrison, 1987
Abendsegler	<i>Nyctalus</i> Bowdich, 1825
Abendstern-Pipistrelle	<i>Pipistrellus</i> (<i>Pipistrellus</i>) <i>hesperidus</i> (Temminck, 1840)
Abendstern-Zwergfledermaus	<i>Pipistrellus</i> (<i>Pipistrellus</i>) <i>hesperidus</i> (Temminck, 1840)
Abo-Schmetterlingsfledermaus	<i>Glauconycteris poensis</i> (Gray, 1842)
Adams Hufeisennase	<i>Rhinolophus adami</i> Aellen and Brosset, 1968
Aellens Langflügelfledermaus	<i>Miniopterus aelleni</i> Goodman, Maminirina, Weyeneth, Bradman, Christidis, Ruedi and Appleton, 2009
Afrikanische Dreizahn-Blattnase	<i>Triaenops afer</i> Peters, 1877
Afrikanische Langflügelfledermaus	<i>Miniopterus africanus</i> Sanborn, 1936
Afrikanische Langohrfledermäuse	<i>Laephotis</i> Thomas, 1901
Afrikanische Langzungen-Flughunde	<i>Megaloglossus</i> Pagenstecher, 1885
Afrikanische Schiebeschwanz-Fledermaus	<i>Coleura afra</i> (Peters, 1852)
Afrikanischer Langzungen-Flughund	<i>Megaloglossus woermanni</i> Pagenstecher, 1885
Afrikanischer Palmenflughund	<i>Eidolon helvum</i> (Kerr, 1792)
Afrikanischer strohfarbener Flughund	<i>Eidolon helvum</i> (Kerr, 1792)
Ägyptische Bulldoggfledermaus	<i>Tadarida aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
Ägyptische Grabfledermaus	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
Ägyptische Mausschwanz-Fledermaus	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
Ägyptische Schlitznasen-Fledermaus	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
Aldabra- Dreizahnblattnase	<i>Paratriaenops pauliani</i> (Goodman and Ranivo, 2008)
Aldabra-Flugfuchs	<i>Pteropus aldabrensis</i> True, 1893
Alpenfledermaus	<i>Hypsugo savii</i> (Bonaparte, 1837)
Alpen-Pipistrelle	<i>Hypsugo savii</i> (Bonaparte, 1837)
Alpenzwergfledermaus	<i>Hypsugo savii</i> (Bonaparte, 1837)
Altwelt-Rundblattnasen	<i>Hipposideros</i> Gray, 1831
Ambre-Langflügelfledermaus	<i>Miniopterus ambohitrensis</i> Goodman, Ramasindrazana, Naughton and Appleton, 2015
Anchietas Breitgesicht-Flughund	<i>Plerotes anchietae</i> (Seabra, 1900)
Anchietas Pipistrelle	<i>Neoromicia anchietae</i> (Seabra, 1900)
Anchietas Zwergfledermaus	<i>Neoromicia anchietae</i> (Seabra, 1900)
Andersens Hufeisennase	<i>Rhinolophus eloquens</i> K. Andersen, 1905
Andersens Schlitznasen-Fledermaus	<i>Nycteris aurita</i> (K. Andersen, 1912)
Andrews Hausfledermaus	<i>Scotophilus andrewreborii</i> Brooks and Bickham, 2014
Angola-Bulldoggfledermaus	<i>Mops</i> (<i>Mops</i>) <i>condylurus</i> (A. Smith, 1833)
Angola-Epaulettenflughund	<i>Epomophorus angolensis</i> Gray, 1870
Angola-Langohrfledermaus	<i>Laephotis angolensis</i> Monard, 1935
Angola-Mausohr	<i>Cistugo seabrae</i> Thomas, 1912
Angolanischer Samtfell-Flughund	<i>Myonycteris angolensis</i> (Bocage, 1898)
Angola-Weichfellflughund	<i>Myonycteris angolensis</i> (Bocage, 1898)
Anjouan-Mausohr	<i>Myotis anjouanensis</i> Dorst, 1960
Ansells Epaulettenflughund	<i>Epomophorus anselli</i> Bergmans and Van Strien, 2004
Ansorges Bulldoggfledermaus	<i>Mops</i> (<i>Chaerephon</i>) <i>ansorgei</i> (Thomas, 1913)
Äthiopische Hausfledermaus	<i>Scotophilus ejetai</i> Brooks and Bickham, 2014
Äthiopischer Samtfell-Flughund	<i>Myonycteris angolensis petraea</i> (Bergmans, 1997)
Aufgeblasene Langflügelfledermaus	<i>Miniopterus inflatus</i> Thomas, 1903
Azoren-Abendsegler	<i>Nyctalus azureum</i> (Thomas, 1901)
Bakaris Bulldoggfledermaus	<i>Mops</i> (<i>Xiphonycteris</i>) <i>bakarii</i> Stanley, 2009
Bale-Langohr	<i>Plecotus balensis</i> Kruskop & Lavrenchenko, 2000
Balkan-Langohr	<i>Plecotus kolombatovici</i> Dulic, 1980
Bananen Zwergfledermaus	<i>Afronycteris nana</i> (Peters, 1852)
Bananen-Fledermaus	<i>Afronycteris nana</i> (Peters, 1852)
Bananen-Mausohr	<i>Myotis bocagii</i> (Peters, 1870)
Bananen-Pipistrelle	<i>Afronycteris nana</i> (Peters, 1852)
Bartfledermaus	<i>Myotis mystacinus</i> (Kuhl, 1817)
Bates' Schlitznasen-Fledermaus	<i>Nycteris arge</i> Thomas, 1903
Bemmelens Bulldoggfledermaus	<i>Mops</i> (<i>Chaerephon</i>) <i>bemmeleni</i> (Jentink, 1879)
Benito-Blattnase	<i>Hipposideros beatus</i> (K. Andersen, 1906)
Benito-Rundblattnase	<i>Hipposideros beatus</i> (K. Andersen, 1906)
Bergmans Harlekin-Flughund	<i>Scotonycteris bergmansi</i> Hassanin, Khouider, Gembu, Goodman, Kadjo, Nesi, Pourrut, Nakouné and Bonillo, 2015
Bibundi Schmetterlingsfledermaus	<i>Glauconycteris egeria</i> Thomas, 1913
Bini Pergamentflügel-Fledermaus	<i>Myopterus whiteyi</i> (Scharff, 1900)
Blasius-Hufeisennase	<i>Rhinolophus blasii</i> Peters, 1867
Botswana-Langohrfledermaus	<i>Laephotis botswanae</i> Setzer, 1971
Bottas Breitflügelfledermaus	<i>Eptesicus</i> (<i>Cnephaeus</i>) <i>bottae</i> (Peters, 1869)
Bottas Fledermaus	<i>Eptesicus</i> (<i>Cnephaeus</i>) <i>bottae</i> (Peters, 1869)
Bourbonische Schwirrfledermaus	<i>Scotophilus borbonicus</i> (E. Geoffroy St.-Hilaire, 1803)
Breitflügelfledermäuse	<i>Eptesicus Rafinesque</i> , 1820
Breitgesicht-Flughunde	<i>Plerotes</i> K. Andersen, 1910

- Breitköpfige Pipistrelle
 Breitköpfige Zwergfledermaus
 Bulldoggfledermaus
 Bulldoggfledermäuse
 Bulldogg-Fledermäuse
 Bunte Wollfledermaus
 Buschland-Rundblattnase
 Buschveld-Hufeisennase
 Büttikofers Epaulettenflughund
 Campomaan-Flughund
 Capische Abendfledermaus
 Capische Trugfledermaus
 Capischen Kamnase
 Zentralafrikanische Doggengrämler
 Chagrin Fledermaus
 Chapins Bulldoggfledermaus
 Christis Graues Langohr
 Cohens Hufeisennase
- Currys Schmetterlingsfledermaus
 Damara-Hufeisennase
 Darlings Hufeisennase
 Daubentons Pergamentflügel-Fledermaus
 De Wintons Langohrfledermaus
 Deckens Hufeisennase
 Dents Hufeisennase
 Dja Schlitznasen-Fledermaus
 Dja-Schlitznasenfledermaus
 Dobsons Epaulettenflughund
 Dongola-Dämmerungsfledermaus
 Dreifarb-Mausohr
 Dreizackblattnase
 Dreizackblattnasen
 Dreizahn-Blattnasen
 Dreyzahnige Kammnase
 Dunkelbraune Pipistrelle
 Dunkelbraune Zwergfledermaus
 Dunkle Schmetterlingsfledermaus
- Eger-Langflügelfledermaus
- Egyptischer fliegender Hund
 Eisentrauts Pipistrelle
 Eisentrauts Zwergfledermaus
 Elfen-Pipistrelle
 Elfen-Zwergfledermaus
 Epaulettenflughund
 Epaulettenflughunde
 Epaulettenflughunde
 Europäische Bulldoggfledermaus
 Europäische Sackfledermaus
 Falsche Vampire
 Flachkopf-Bulldoggfledermäuse
 Flachkopf-Bulldoggfledermäuse
 Flachköpfige Dämmerungsfledermaus
 Flachschnauzigen Schwirrfledermaus
 Flattnasenfreischwänze
 Flederhunde
 Fledermaus mit gerändelten Ohren
 Fledermäuse
 Flugfuchse
 Flughunde
 Franquet's Epaulettenflugfuchs
 Franquets Epaulettenflughund
 Freischeinige Stelfussfledermaus
 Fürst von Abruzzens Bulldoggfledermaus
 Gaislers Langohrfledermaus
 Gallaghers Bulldoggfledermaus
 Gambia Epaulettenflughund
 Gambia Schlitznasen-Fledermaus
 Gebirgs-Höhlenflughund
 Gefleckte Bulldoggfledermaus
 Gefleckte Bulldoggfledermaus
 Gefleckte Schmetterlingsfledermaus
 Gelbbäuchige Hausfledermaus
- Nycticeinops crassulus* (Thomas, 1904)
Nycticeinops crassulus (Thomas, 1904)
Tadarida teniotis (Rafinesque, 1814)
 MOLOSSIDAE Gervais, 1856
 MOLOSSIDAE Gervais, 1856
Kerivoula argentata Tomes, 1861
Rhinolophus simulator K. Andersen, 1904
Rhinolophus simulator K. Andersen, 1904
Epomops buettikoferi (Matschie, 1899)
Casinycteris campomaanensis Hassanin, 2014
Laephotis capensis (A. Smith, 1829)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Rhinolophus capensis Lichtenstein, 1823
Mops (Mops) condylurus (A. Smith, 1833)
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Mops (Chaerephon) chapini (J.A. Allen, 1917)
Plecotus christii Gray, 1838
Rhinolophus cohenae Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Glauconycteris curryae Eger and Schlitter, 2001
Rhinolophus damarensis Roberts, 1946
Rhinolophus darlingi K. Andersen, 1905
Myopterus daubentonii Desmarest, 1820
Laephotis wintoni Thomas, 1901
Rhinolophus deckenii Peters, 1868
Rhinolophus denti Thomas, 1904
Nycteris major (K. Andersen, 1912)
Nycteris major (K. Andersen, 1912)
Epomophorus dobsonii Bocage, 1889
Vansonia rueppellii (J.B. Fischer, 1829)
Myotis tricolor (Temminck, 1832)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Asellia Gray, 1838
Triaenops Dobson, 1871
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Pseudoromicia brunnea (Thomas, 1880)
Pseudoromicia brunnea (Thomas, 1880)
Glauconycteris atra Hassanin, Colombo, Gembu, Merle, Tu, Görföl, Musaba Akawa, Csorba, Kearney, Monadjem and Ing, 2017
Miniopterus egeri Goodman, Ramasindrazana, Maminirina, Schoeman and Appleton, 2011
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Nycticeinops eisentrauti (Hill, 1968)
Nycticeinops eisentrauti (Hill, 1968)
Hypsugo ariel (Thomas, 1904)
Hypsugo ariel (Thomas, 1904)
Epomophorus Bennett, 1836
Epomophorus Bennett, 1836
Epomops Gray, 1866
Tadarida teniotis (Rafinesque, 1814)
Miniopterus schreibersii (Kuhl, 1817)
 MEGADERMATIDAE H. Allen, 1864
Platymops Thomas, 1906
Sauromys Roberts, 1917
Pipistrellus (Pipistrellus) hesperidus (Temminck, 1840)
Scotophilus dinganii (A. Smith, 1833)
 EMBALLONURIDAE Gervais, 1855
 PTEROPODIDAE Gray, 1821
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
 CHIROPTERA Blumenbach, 1779
Pteropus Brisson, 1762
 PTEROPODIDAE Gray, 1821
Epomops franqueti (Tomes, 1860)
Epomops franqueti (Tomes, 1860)
Myotis capaccinii (Bonaparte, 1837)
Mops (Chaerephon) aloysiisabaudiae (Festa, 1907)
Plecotus gaisleri Benda, Kiefer, Hanak & Veith, 2004
Mops (Chaerephon) gallagheri (Harrison, 1975)
Epomophorus gambianus (Ogilby, 1835)
Nycteris gambiensis (K. Andersen, 1912)
Stenonycteris lanosus (Thomas, 1906)
Mops (Chaerephon) bivittatus (Heuglin, 1861)
Mops (Mops) leucostigma (G.M. Allen, 1918)
Glauconycteris humeralis J.A. Allen, 1917
Scotophilus dinganii (A. Smith, 1833)

Gelbbauchige Schwirrfledermaus	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
Gelbe Pipistrelle	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
Gelbe Zwergfledermaus	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
Gelbflügel-Fledermaus	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
Gelbflügel-Fledermäuse	<i>Lavia</i> Gray, 1838
Gelbflügelige Großblattnase	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
Gemeine Langflügelfledermaus	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Gemeine Schlitznasen-Fledermaus	<i>Nycteris hispida</i> (Schreber, 1774)
Genetzte Schmetterlingsfledermaus	<i>Glauconycteris variegata</i> (Tomes, 1861)
Geoffroys Dreizackblattnase	<i>Asellia tridens</i> (E. Geoffroy St.-Hilaire, 1813)
Geoffroys Hufeisennase	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
Gesäumte Doggenrämmler	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
Gestreifte Schmetterlingsfledermaus	<i>Glauconycteris alboguttata</i> J.A. Allen, 1917
Gewöhnliche Bulldoggfledermäuse	<i>Tadarida Rafinesque</i> , 1814
Gewöhnliche Rundblattnase	<i>Hipposideros caffer</i> (Sundevall, 1846)
Glattnasen	<i>VESPERTILIONIDAE</i> Gray, 1821
Glattnasen-Fledermäuse	<i>VESPERTILIONIDAE</i> Gray, 1821
Glattnasen-Freischwänze	<i>EMBALLONURIDAE</i> Gervais, 1855
Gleichsatteligen Kammnase	<i>Rhinolophus euryale</i> Blasius, 1853
Glens Langflügelfledermaus	<i>Miniopterus gleni</i> Peterson, Eger and Mitchell, 1995
Glens Schmetterlingsfledermaus	<i>Glauconycteris gleni</i> Peterson and Smith, 1973
Goldener Kurzgaumen-Flughund	<i>Casinycteris argynnis</i> Thomas, 1910
Gorongosa-Hufeisennase	<i>Rhinolophus gorongosae</i> Taylor, MacDonald, Goodman, Kearney, Cotterill, Stoffberg, Monadjem, Schoeman, Guyton, Naskrecki and Richards, 2018
Grabflatterer	<i>Taphozous</i> E. Geoffroy St.-Hilaire, 1818
Grabfledermäuse	<i>Taphozous</i> E. Geoffroy St.-Hilaire, 1818
Grandidiers Bulldoggfledermaus	<i>Mops (Chaerephon) leucogaster</i> (A. Grandidier, 1869)
Grandidiers Dreizahn-Blattnase	<i>Paratriaenops auritus</i> (G. Grandidier, 1912)
Grandidiers Zwergfledermaus	<i>Nycticeinops grandidieri</i> (Dobson, 1876)
Griffiths Langflügelfledermaus	<i>Miniopterus griffithsi</i> Goodman, Maminirina, Bradman, Christidis and Appleton, 2009
Großabendsegler	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Grossblattnasen	<i>MEGADERMATIDAE</i> H. Allen, 1864
Großblattnasen	<i>MEGADERMATIDAE</i> H. Allen, 1864
Große Bulldoggfledermaus	<i>Tadarida ventralis</i> (Heuglin, 1861)
Große Hufeisennase	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Große Schlitznase	<i>Nycteris grandis</i> Peters, 1865
Große Schlitznasen-Fledermaus	<i>Nycteris grandis</i> Peters, 1865
Große Schmetterlingsfledermaus	<i>Glauconycteris superba</i> Hayman, 1939
Grosser Maskarenen-Flugfuchs	<i>Pteropus niger</i> (Kerr, 1792)
Großhufeisennase	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Großohr-Bulldoggfledermaus	<i>Tadarida lobata</i> (Thomas, 1891)
Großohrhohnase	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
Großohrige Riesen-Bulldoggfledermaus	<i>Otomops martiensseni</i> (Matschie, 1897)
Großohrige Schlitznasen-Fledermaus	<i>Nycteris macrotis</i> Dobson, 1876
Großohr-Rundblattnase	<i>Hipposideros megalotis</i> (Heuglin, 1861)
Grünliche Hausfledermaus	<i>Scotophilus viridis</i> (Peters, 1852)
Guinea-Hufeisennase	<i>Rhinolophus guineensis</i> Eisentraut, 1960
Guinea-Pipistrelle	<i>Neoromicia guineensis</i> (Bocage, 1889)
Guinea-Zwergfledermaus	<i>Neoromicia guineensis</i> (Bocage, 1889)
Haarbindigen Dämmerungsfledermaus	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Haarige Schlitznasenfledermaus	<i>Nycteris hispida</i> (Schreber, 1774)
Halcyon Rundblattnase	<i>Rhinolophus alcyone</i> Temminck, 1853
Halskrausen-Flughunde	<i>Myonycteris</i> Matschie, 1899
Hamiltons Grabfledermaus	<i>Taphozous hamiltoni</i> Thomas, 1920
Hammerkopf	<i>Hypsignathus monstrosus</i> H. Allen, 1862
Hammerkopf-Flughund	<i>Hypsignathus monstrosus</i> H. Allen, 1862
Hammerkopf-Flughunde	<i>Hypsignathus</i> H. Allen, 1862
Hanaks Pipistrelle	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hanaks Zwergfledermaus	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hardwicks Mausschwanz-Fledermaus	<i>Rhinopoma cystops</i> Thomas, 1903
Harlekin-Flughunde	<i>Scotonycteris</i> Matschie, 1894
Harrison-Riesen-Bulldoggfledermaus	<i>Otomops harrisoni</i> Ralph, Richards, Taylor, Napier and Lamb, 2015
Harrisons Samtfell-Flughund	<i>Myonycteris angolensis goliath</i> (Bergmans, 1997)
Hausfledermäuse	<i>Scotophilus</i> Leach, 1821
Haymans Harlekin-Flughund	<i>Scotonycteris occidentalis</i> Hayman, 1947
Haymans Kleiner Epaulettenflughund	<i>Epomophorus intermedius</i> (Hayman, 1963)
Hemprichs Wüstengroßohr	<i>Otonycteris hemprichii</i> Peters, 1859
Herznasen-Fledermaus	<i>Cardioderma cor</i> (Peters, 1872)
Herznasen-Fledermäuse	<i>Cardioderma</i> Peters, 1873
herzohrige Abendfledermaus	<i>Hypsugo savii</i> (Bonaparte, 1837)
Heuglins Wollfledermaus	<i>Kerivoula eriophora</i> (Heuglin, 1877)
Hildebrandts Hufeisennase	<i>Rhinolophus hildebrandtii</i> Peters, 1878
Hildegardes Grabfledermaus	<i>Taphozous hildegardae</i> Thomas, 1909

- Hills Hufeisennase
Hindes Hausfledermaus
Hochland-Hufeisennase
Höhlenflughunde
Hottentotten-Breitflügelfledermaus
Hufeisennase
Hufeisennasen
Hufeisennasen
Hufeisennasen-Fledermäuse
Hufeisennasen-Fledermäuse
Hundkopf-Schwirrfledermaus
Intermediäre Schlitznasenfledermaus
Intermediäre Schlitznasen-Fledermaus
Isabell-Breitflügelfledermaus
isabellfarbenen Abendfledermaus
Isabellfledermaus
Isabell-Weissflügel Serotine
- Jones' Rundblattnase
Kaffern-Sackfledermaus
Kahlschienige Dämmerungsfledermaus
Kahuzi-Hufeisennase
Kamerun-Blattnase
Kamerun-Blattnase
Kamerun-Rundblattnase
Kanaren-Langohr
Kanaren-Langohrfledermaus
Kap-Hufeisennase
Kap-Pipistrelle
Kap-Zwergfledermaus
Kastanienrothe Kammnase
Kenya-Schmetterlingsfledermaus
Kerbohrige Ohrenfledermaus
Kleinabendsegler
Kleine Abendsegler
Kleine Bartfledermaus
Kleine Bulldoggfledermaus
Kleine Hausfledermäuse
Kleine Hufeisennase
Kleine Langflügelfledermaus
Kleine Mausschwanzfledermaus
Kleine Pipistrelle
Kleine Schmetterlingsfledermaus
Kleine Waldfledermaus
Kleine Wollfledermaus
Kleine Zwergfledermaus
Kleiner Abendsegler
Kleiner Halsbandflughund
Kleiner Halskrausen-Flughund
Kleiner Maskarenen-Flugfuchs
Kleinhufeisennase
Kleinohr-Dreizackblattnase
Kleinste Schlitznasen-Fledermaus
Kocks Mausohr
Komoren-Höhlenflughund
Komoren-Langflügelfledermaus
Kongo-Bulldoggfledermaus
Kryptische Madagaskar-Rundblattnase
- Kuhlsche Fledermaus
Kupferfarbene Wollfledermaus
Kupferfarbenes Mausohr
Kurzflügel-Bulldoggfledermaus
Kurzflügeligen Doggengrämler
Kurzgaumen-Flughunde
Kurzmaulige Fledermaus
Kurzschwanz-Rundblattnase
Lagos-Breitflügelfledermaus
Lamottes Rundblattnase
Landers Hufeisennase
Lander's Rundblattnase
Lanflügelfledermaus
Langflügelfledermaus
Langflügelfledermäuse
Langflügelfledermäuse
Langfußfledermaus
- Rhinolophus hilli* Aellen, 1973
Scotoecus hindei Thomas, 1901
Rhinolophus hillorum Koopman, 1989
Rousettus Gray, 1821
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Rhinolophus ferrumequinum (Schreber, 1774)
RHINOLOPHIDAE Gray, 1825
Rhinolophus Lacépède, 1799
RHINOLOPHIDAE Gray, 1825
Rhinolophus Lacépède, 1799
Scotophilus nigrita (Schreber, 1774)
Nycteris intermedia Aellen, 1959
Nycteris intermedia Aellen, 1959
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Pseudoromicia isabella (Decher, Hutterer and Monadjem, 2016)
Hipposideros jonesi Hayman, 1947
Miniopterus natalensis (A. Smith, 1833)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rhinolophus kahuzi Fahr and Kerbis Peterhans, 2013
Doryrhina camerunensis (Eisentraut, 1956)
Hipposideros ruber (Noack, 1893)
Doryrhina camerunensis (Eisentraut, 1956)
Plecotus teneriffae Barrett-Hamilton, 1907
Plecotus teneriffae Barrett-Hamilton, 1907
Rhinolophus capensis Lichtenstein, 1823
Laephotis capensis (A. Smith, 1829)
Laephotis capensis (A. Smith, 1829)
Rhinolophus landeri Martin, 1838
Glauconycteris kenyacola Peterson, 1982
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Scotoecus Thomas, 1901
Rhinolophus hipposideros (Bechstein, 1800)
Miniopterus fraterculus Thomas and Schwann, 1906
Rhinopoma cystops Thomas, 1903
Pipistrellus (Pipistrellus) nanulus Thomas, 1904
Glauconycteris beatrix Thomas, 1901
Nyctalus leisleri (Kuhl, 1817)
Kerivoula lanosa (A. Smith, 1847)
Pipistrellus (Pipistrellus) nanulus Thomas, 1904
Nyctalus leisleri (Kuhl, 1817)
Myonycteris torquata (Dobson, 1878)
Myonycteris torquata (Dobson, 1878)
Pteropus subniger (Kerr, 1792)
Rhinolophus hipposideros (Bechstein, 1800)
Cloeotis Thomas, 1901
Nycteris nana (K. Andersen, 1912)
Myotis dieteri M. Happold, 2005
Rousettus obliviosus Kock, 1978
Miniopterus griveaudi Harrison, 1959
Mops (Mops) congicus J.A. Allen, 1917
Macronycteris cryptovalorona (Goodman, Schoeman, Rakotoarivelo and Willows-Munro, 2016)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Kerivoula cuprosa Thomas, 1912
Myotis bocagii (Peters, 1870)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Casinycteris Thomas, 1910
Barbastella barbastellus (Schreber, 1774)
Hipposideros curtus G.M. Allen, 1921
Eptesicus (Cnephaeus) platyops (Thomas, 1901)
Hipposideros lamottei Brosset, 1985
Rhinolophus landeri Martin, 1838
Rhinolophus landeri Martin, 1838
Miniopterus villiersi Aellen, 1956
Miniopterus schreibersii (Kuhl, 1817)
MINIOPTERIDAE Dobson, 1875
Miniopterus Bonaparte, 1837
Myotis capaccinii (Bonaparte, 1837)

Langohren	<i>Plecotus</i> E. Geoffroy St.-Hilaire, 1818
Langohrfledermäuse	<i>Plecotus</i> E. Geoffroy St.-Hilaire, 1818
Leislersche Fledermaus	<i>Nyctalus leisleri</i> (Kuhl, 1817)
Leseuers Mausohr	<i>Cistugo lesueuri</i> Roberts, 1919
Livingstone-Hausfledermaus	<i>Scotophilus livingstonii</i> Brooks and Bickham, 2014
Livingstones Flugfuchs	<i>Pteropus livingstonii</i> Gray, 1866
Luftige Pipistrelle	<i>Pipistrellus (Pipistrellus) aero</i> Heller, 1912
Luftige Zwergfledermaus	<i>Pipistrellus (Pipistrellus) aero</i> Heller, 1912
Lybische Hufeisennase	<i>Rhinolophus horaceki</i> Benda and Vallo, 2012
Machados Schmetterlingsfledermaus	<i>Glauconycteris machadoi</i> Hayman, 1963
MacInnes' Mausschwanz-Fledermaus	<i>Rhinopoma macinnesi</i> Hayman, 1937
Maclauds Hufeisennase	<i>Rhinolophus maclaudi</i> Pousargues, 1898
Madagaskar- Dreizack-Blattnasen	<i>Paratriaenops</i> Benda and Vallo, 2009
Madagaskar Mastino-Fledermaus	<i>Mormopterus jugularis</i> (Peters, 1865)
Madagaskar Pipistrelle	<i>Laephotis malagasyensis</i> (Peterson, Eger and Mitchell, 1995)
Madagaskar-Flugfuchs	<i>Pteropus rufus</i> E. Geoffroy St.-Hilaire, 1803
Madagaskar-Freischwanzfledermaus	<i>Paremballonura atrata</i> (Peters, 1874)
Madagaskar-Haftscheibenfledermäuse	MYZOPODIDAE Thomas, 1904
Madagaskar-Haftscheibenfledermäuse	<i>Myzopoda</i> Milne-Edwards and A. Grandidier, 1878
Madagaskar-Hausfledermaus	<i>Scotophilus robustus</i> A. Milne-Edwards, 1881
Madagaskar-Höhlenflughund	<i>Rousettus madagascariensis</i> G. Grandidier, 1929
Madagaskar-Langflügelfledermaus	<i>Miniopterus sororculus</i> Goodman, Ryan, Maminirina, Fahr, Christidis and Appleton, 2007
Madagaskar-Mausohr	<i>Myotis goudoti</i> (A. Smith, 1834)
Madagaskar-Palmenflughund	<i>Eidolon dupreanum</i> (Schegel, 1867)
Madagaskar-Rundblattnase	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Madagaskar-Rundblattnase	<i>Macronycteris gigas</i> (Wagner, 1845)
Madagaskar-Schiebeschwanzfledermaus	<i>Coleura kibomalandy</i> Goodman, Puechmaille, Friedli-Weyeneth, Gerlach, Ruedi, Schoeman, Stanley and Teeling, 2012
Madagaskar-Zwergfledermaus	<i>Laephotis malagasyensis</i> (Peterson, Eger and Mitchell, 1995)
Madagassische Bulldoggfledermaus	<i>Tadarida fulminans</i> (Thomas, 1903)
Madagassische Haftscheibenfledermaus	MYZOPODIDAE Thomas, 1904
Madagassische Riesen-Bulldoggfledermaus	<i>Otomops madagascariensis</i> Dorst, 1953
Madeira-Fledermaus	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeira-Pipistrelle	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Madeira-Zwergfledermaus	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Maghreb-Langflügelfledermaus	<i>Miniopterus maghrebensis</i> Puechmaille, Allegrini, Benda, Bilgin, Ibañez and Juste, 2014
Maghreb-Mausohr	<i>Myotis punicus</i> Felten, 1977
Mahafaly-Langflügelfledermaus	<i>Miniopterus mahafaliensis</i> Goodman, Bradman, Christides and Appleton, 2009
Majors Langflügelfledermaus	<i>Miniopterus majori</i> Thomas, 1906
Manavi-Langflügelfledermaus	<i>Miniopterus manavi</i> Thomas, 1906
Marisas Rundblattnase	<i>Hipposideros marisae</i> Aellen, 1954
Marokkanische Rundblattnase	<i>Hipposideros tephurus</i> (Cabrera, 1906)
Marovaza-Hausfledermaus	<i>Scotophilus marovaza</i> Goodman, Ratrimomanarivo and Randrianandrianina, 2006
Marungu-Rundblattnase	<i>Macronycteris commersoni</i> (E. Geoffroy St.-Hilaire, 1813)
Marungu-Rundblattnase	<i>Macronycteris gigas</i> (Wagner, 1845)
Mastino-Fledermäuse	<i>Mormopterus</i> Peters, 1865
Matroka-Pipistrelle	<i>Laephotis matroka</i> (Thomas and Schwann, 1905)
Matroka-Zwergfledermaus	<i>Laephotis matroka</i> (Thomas and Schwann, 1905)
Mauritius Mastino-Fledermaus	<i>Mormopterus acetabulosus</i> (Hermann, 1804)
Mauritius-Grabfledermaus	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
Maurizische Doggengrämler	<i>Mormopterus acetabulosus</i> (Hermann, 1804)
Mäuschen-Pipistrelle	<i>Hypsugo musciculus</i> (Thomas, 1913)
Mäuschen-Zwergfledermaus	<i>Hypsugo musciculus</i> (Thomas, 1913)
Mausohr	<i>Myotis</i> Kaup, 1829
Mausohren	<i>Cistugo</i> Thomas, 1912
Mausohren	<i>Myotis</i> Kaup, 1829
Mausschwanzfledermäuse	RHINOPOMATIDAE Dobson, 1872
Mausschwanz-Fledermäuse	RHINOPOMATIDAE Dobson, 1872
Medje-Bulldoggfledermaus	<i>Rhinopoma</i> E. Geoffroy St.-Hilaire, 1818
Mehely-Hufeisennase	<i>Mops (Mops) congicus</i> J.A. Allen, 1917
Midas-Bulldoggfledermaus	<i>Rhinolophus mehelyi</i> Matschie, 1901
Mittelmeerhufeisennase	<i>Mops (Mops) midas</i> (Sundevall, 1843)
Mittelmeer-Hufeisennase	<i>Rhinolophus euryale</i> Blasius, 1853
Moloneys Flachkopf-Fledermaus	<i>Rhinolophus euryale</i> Blasius, 1853
Moloneys Schmalflügel-Fledermaus	<i>Mimetillus moloneyi</i> (Thomas, 1891)
Mongalla-Bulldoggfledermaus	<i>Mimetillus moloneyi</i> (Thomas, 1891)
	<i>Mops (Mops) demonstrator</i> (Thomas, 1903)

- Mopsfledermaus
Mopsfledermäuse
Morris' Mausohr
Mossambique-Hufeisennase
- Mossambique-Langflügelfledermaus
- Mount Nimba-Rundblattnase
Mozambique Dämmerungsfledermaus
Mt Mabu Hufeisennase
- Mückenfledermaus
Nacktbau-Grabfledermaus
Nacktbäuchige Tempelfledermaus
Namibia-Langohrfledermaus
Nemoroka-Langflügelfledermaus
- Natalischen Schwirrfledermaus
Natal-Langflügelfledermaus
Netzhäutigen Dämmerungsfledermaus
Niagara-Bulldoggfledermaus
Nigeria-Bulldoggfledermaus
Nilflughund (Ägyptischer Höhlenflughund)
Noacks Rundblattnase
Nußfarbene Hausfledermaus
Olivenbraune Schwirrfledermaus
Olivengrüne Schwirrfledermaus
Östliche Haftscheibenfledermaus
- Östliche Mopsfledermaus
Palmenflughund
Palmenflughunde
Parisis Schlitznasen-Fledermaus
Patrizis Dreizackblattnase
Pels Taschenfledermaus
Pemba-Bulldoggfledermaus
Pemba-Flugfuchs
Percivals Kleinohr-Dreizackblattnase
Pergamentflügel-Fledermäuse
Peters' Epaulettenflughund
Peters' Flachkopf-Bulldoggfledermaus
Peters' Kleiner Epaulettenflughund
Peter's Zwergepaulettenflughund
Petersons Bulldoggfledermaus
Petersons Langflügelfledermaus
- Pipistrellen
Poehle's Traenenflughund
Pohles Harlekin-Flughund
Punisches Mausohr
Raceys Pipistrelle
- Raceys Zwergfledermaus
- Rauhaarschlitznase
Reliktärer Halskrausen-Flughund
Rendalls Pipistrelle
Rendalls Zwergfledermaus
Reunion Mastino-Fledermaus
- Réunion-Hausfledermaus
Riesen Abendsegler
Riesenabendsegler
Riesen-Bulldoggfledermäuse
Riesen-Hausfledermaus
Riesen-Rundblattnase
- Riesen-Rundblattnase
Robbins Hausfledermaus
Roberts' Flachkopf-Bulldoggfledermaus
Roberts Zwergfledermaus
- Rodrigues-Flugfuchs
Rodriguez-Flughund
Rosevears Zwergfledermaus
- Barbastella barbastellus* (Schreber, 1774)
Barbastella Gray, 1821
Myotis morrisi Hill, 1971
Rhinolophus mossambicus Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Miniopterus mossambicus Monadjem, Goodman, Stanley and Appleton, 2013
Hipposideros lamottei Brosset, 1985
Afronycteris nana (Peters, 1852)
Rhinolophus mabuensis Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Taphozous nudiventris Cretzschmar, 1830
Taphozous nudiventris Cretzschmar, 1830
Laephotis namibensis Setzer, 1971
Miniopterus brachytragos Goodman, Maminirina, Bradman, Christidis and Appleton, 2009
Scotophilus dinganii (A. Smith, 1833)
Miniopterus natalensis (A. Smith, 1833)
Pipistrellus (Pipistrellus) hesperidus (Temminck, 1840)
Mops (Mops) niangarae J.A. Allen, 1917
Mops (Chaerephon) nigeriae (Thomas, 1913)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Hipposideros ruber (Noack, 1893)
Scotophilus nux Thomas, 1904
Scotophilus leucogaster (Cretzschmar, 1826)
Scotophilus viridis (Peters, 1852)
Myzopoda aurita Milne-Edwards and A. Grandidier, 1878
Barbastella leucomelas (Cretzschmar, 1826)
Eidolon helvum (Kerr, 1792)
Eidolon Rafinesque, 1815
Nycteris parisi (de Beaux, 1924)
Asellia patrizii de Beaux, 1931
Saccolaimus peli (Temminck, 1853)
Mops (Xiphonycteris) bakarii Stanley, 2009
Pteropus voeltzkowi Matschie, 1909
Cloeotis percivali Thomas, 1901
Myopterus E. Geoffroy St.-Hilaire, 1818
Epomophorus crypturus Peters, 1852
Platymops setiger (Peters, 1878)
Epomophorus pusillus Peters, 1868
Epomophorus pusillus Peters, 1868
Mops (Xiphonycteris) petersoni (El Rayah, 1981)
Miniopterus petersoni Goodman, Bradman, Maminirina, Ryan, Christidis & Appleton, 2008
Pipistrellus Kaup, 1829
Casinycteris ophiodon (Pohle, 1943)
Casinycteris ophiodon (Pohle, 1943)
Myotis punicus Felten, 1977
Pipistrellus (Pipistrellus) raceyi Bates, Ratrimomanarivo, Harrison and Goodman, 2006
Pipistrellus (Pipistrellus) raceyi Bates, Ratrimomanarivo, Harrison and Goodman, 2006
Nycteris hispida (Schreber, 1774)
Myonycteris relicta Bergmans, 1980
Pseudoromicia rendalli (Thomas, 1889)
Pseudoromicia rendalli (Thomas, 1889)
Mormopterus francoismoutoui Goodman, Jansen Van Vuuren, Ratrimomanarivo, Probst, Bowie, 2008
Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus lasiopterus (Schreber, 1780)
Otomops Thomas, 1913
Scotophilus nigrita (Schreber, 1774)
Macronycteris commersoni (E. Geoffroy St.-Hilaire, 1813)
Macronycteris gigas (Wagner, 1845)
Scotophilus nucella Robbins, 1983
Sauromys petrophilus (Roberts, 1917)
Laephotis robertsi (Goodman, Taylor, Ratrimomanarivo and Hofer, 2012)
Pteropus rodricensis Dobson, 1878
Pteropus rodricensis Dobson, 1878
Pseudoromicia roseveari (Monadjem, Richards, Taylor and Stoffberg, 2013)

- Rostfarbene Bulldoggfledermaus
 Rostfarbene Pipistrelle
 Rostfarbene Zwergfledermaus
 rothbraune Doggengrämler
 Rothen Kamnase
 Rötliche Dreizahn-Blattnase
 Rundblattnasen
 Rundlappigen Kammnase
 Rüppells Hufeisennase
 Rüppells Hufeisennase
 Rüppells Pipistrelle
 Rüppells Zwergfledermaus
 Russbraunen Kamnase
 Rußfarbene Rundblattnase
 Ruwenzori-Hufeisennase
 Ruwenzori-Langhaarflughund
 Sackfledermaus
 Sackflügel-Fledermäuse
 Sakeji-Hufeisennase
 Samburu-Pipistrelle
 Samburu-Zwergfledermaus
 Samtfell-Flughunde
 Sanborns Epaulettenflughund
 Sao Thome Langflügelfledermaus
 São Tomé-Bulldoggfledermaus
 Schiebeschwanz-Fledermäuse
 Schlangenzähniger Harlekin-Flughund
 Schlieffens Abendfledermaus
 Schliemanns Haftscheibenfledermaus

 Schlitznasen
 Schlitznasen
 Schlitznasenfledermäuse
 Schlitznasen-Fledermäuse
 Schlitznasen-Fledermäuse
 Schmalflügel-Fledermäuse
 Schmetterlingsfledermäuse
 Schnauzbärtige Fledermaus
 Schreibers Fledermaus
 Schreibers Langflügelfledermaus
 Schwanzdrüsen-Bulldoggfledermaus
 Schwarzflügel-Hausfledermaus
 Schwarz-Weiße Schmetterlingsfledermaus
 Scotts Mausohr
 Sena-Doggengrämler
 Sennaarische Doggengrämler
 Seychellen Bulldoggfledermaus
 Seychellen Schiebeschwanz-Fledermaus
 Seychellen-Flugfuchs
 Shoa Kamnase
 Sierra-Leone Halskrausen-Flughund
 Silberne Schmetterlingsfledermaus
 Smithers Hufeisennase

 Smiths Samtfell-Flughund
 Smith's Wollfledermaus
 Somalia-Pipistrelle
 Somalia-Zwergfledermaus
 Somalische Dreizackblattnase
 Spurrells Bulldoggfledermaus
 Spurrells Wollfledermaus
 Stanleys Serotine bat

 Stirngrubenfledermaus
 Stirnlappen-Bulldoggfledermaus
 Streifen-Rundblattnase
 Südafrikanischen Sackfledermaus
 Sundevall's Blattnase
 Sundevalls Rundblattnase
 Swinnys Hufeisennase
 Tanzania-Hufeisennase

 Tanzania-Wollfledermaus
 Taschenfledermäuse
 Temmincks Epaulettenflughund
 Temmincks Hufeisennase

Mops (Chaerephon) russatus (J.A. Allen, 1917)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)
Tadarida ventralis (Heuglin, 1861)
Rhinolophus alcyone Temminck, 1853
Triaenops menamena Goodman and Ranivo, 2009
 HIPPOSIDERIDAE Lydekker, 1891
Rhinolophus landeri Martin, 1838
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus fumigatus Rüppell, 1842
Vansonia rueppellii (J.B. Fischer, 1829)
Vansonia rueppellii (J.B. Fischer, 1829)
Rhinolophus capensis Lichtenstein, 1823
Hipposideros fuliginosus (Temminck, 1853)
Rhinolophus ruwenzorii J. Eric Hill, 1942
Myonycteris angolensis ruwenzorii (Eisentraut, 1965)
Miniopterus Bonaparte, 1837
 EMBALLONURIDAE Gervais, 1855
Rhinolophus sakejiensis Cotterill, 2002
Afronycteris helios (Heller, 1912)
Afronycteris helios (Heller, 1912)
Myonycteris Matschie, 1899
Epomophorus grandis (Sanborn, 1950)
Miniopterus newtoni Bocage, 1889
Mops (Chaerephon) tomensis (Juste and Ibáñez, 1993)
Coleura Peters, 1867
Casinycteris ophiodon (Pohle, 1943)
Nycticeinops schlieffenii (Peters, 1859)
Myzopoda schliemanni Goodman, Rakotondraparany and Kofoky, 2007
 NYCTERIDAE Van der Hoeven, 1855
Nycteris G. Cuvier and E. Geoffroy, 1795
 NYCTERIDAE Van der Hoeven, 1855
 NYCTERIDAE Van der Hoeven, 1855
Nycteris G. Cuvier and E. Geoffroy, 1795
Mimetillus Thomas, 1904
Glauconycteris Dobson, 1875
Myotis mystacinus (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Mops (Chaerephon) bemmeleni (Jentink, 1879)
Scotoecus hirundo (de Winton, 1899)
Glauconycteris superba Hayman, 1939
Myotis scotti Thomas, 1927
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Mops (Mops) midas (Sundevall, 1843)
Mops (Chaerephon) pusillus (Miller, 1902)
Coleura seychellensis Peters, 1868
Pteropus seychellensis A. Milne-Edwards, 1877
Rhinolophus fumigatus Rüppell, 1842
Myonycteris leptodon K. Andersen, 1908
Glauconycteris argentata (Dobson, 1875)
Rhinolophus smithersi Taylor, Stoffberg, Monadjem, Schoeman, Bayliss and Cotterill, 2012
Myonycteris angolensis smithii (Thomas, 1908)
Kerivoula smithii Thomas, 1880
Neoromicia somalica (Thomas, 1901)
Neoromicia somalica (Thomas, 1901)
Asellia italosomalica de Beaux, 1931
Mops (Xiphonycteris) spurrelli (Dollman, 1911)
Kerivoula phalaena Thomas, 1912
Laephotis stanleyi (Goodman, Kearney, Ratsimbazafy and Hassanin, 2017)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Mops (Chaerephon) major (Trouessart, 1897)
Macronycteris vittatus (Peters, 1852)
Miniopterus natalensis (A. Smith, 1833)
Hipposideros caffer (Sundevall, 1846)
Hipposideros caffer (Sundevall, 1846)
Rhinolophus swinnyi Gough, 1908
Rhinolophus maendeleo Kock, Csorba and Howell, 2000
Kerivoula africana Dobson, 1878
Saccolaimus Temminck, 1838
Epomophorus labiatus (Temminck, 1837)
Rhinolophus alcyone Temminck, 1853

- Temmincks Rundblattnase
 Thersites' Bulldoggfledermaus
 Thomas' Schmalflügel-Fledermaus
 Trevors Bulldoggfledermaus
 Trouessarts Dreizahn-Blattnase
 Trujillos Hausfledermaus
 Tsingy-Freischwanzfledermaus

 Unerwartete Pipistrelle
 Unerwartete Zwergfledermaus
 Variable Bulldoggfledermaus

 Veldkamps Zwergflughund
 Verwechselte Pipistrelle
 Verwechselte Zwergfledermaus
 Vinsons Schlitznasen-Fledermaus
 Wahlbergs Epaulettenflughund
 Wald-Hufeisennase
 Warzen-Breitflügel-Fledermaus
 Weißbauch-Bulldoggfledermaus
 Weißbäuchige Hausfledermaus
 Weissbauchige Schwirrfledermaus
 Weissflügel-Hausfledermaus
 Weißflügel-Hausfledermaus
 Weisskehl-Hausfledermaus
 Weißrandfledermaus
 Weißrand-Pipistrelle
 Weißflügel Pipistrelle
 Weißflügel-Zwergfledermaus
 Welwitschs Mausohr
 Westafrikanische Nachtfledermaus
 Westliche Hausfledermaus

 Westlicher Woermanns Langzungen-Flughund

 Whitleys Pergamentflügel-Fledermaus
 Willards Hufeisennase
 Wimperfledermaus
 Winziger Epaulettenflughund
 Woermann's Langzungen-Flughund
 Wollfledermäuse
 Woods Schlitznasen-Fledermaus
 Wüsten-Großohr
 Wüsten-Großohren
 Zenati Mausohr

 Zenkers Harlekin-Flughund
 Zenker's Traenenflughund
 Ziamo-Hufeisennase

 Zulu-Pipistrelle
 Zulu-Zwergfledermaus
 zweistreifige Doggengrämler
 Zwerg-Bulldoggfledermaus
 Zwerg-Doggengrämler
 Zwerg-Epaulettenflughund
 Zwergfledermaus
 Zwergfledermäuse
 Zwergflughunde
 Zwerg-Hausfledermaus
 Zwerg-Langflügel-Fledermaus
 Zwerg-Schlitznasenfledermaus
 Zyklopen-Rundblattnase
 Zyklossen-Blattnase

Hipposideros fuliginosus (Temminck, 1853)
Mops (Xiphonycteris) thersites (Thomas, 1903)
Mimetillus thomasi Hinton, 1920
Mops (Mops) trevori J.A. Allen, 1917
Paratriaenops furculus (Trouessart, 1907)
Scotophilus trujilloi Brooks and Bickham, 2014
Paremballonura tiavato (Goodman, Cardiff, Ranivo, Russell and Yoder, 2006)
Pipistrellus (Pipistrellus) inexpectatus Aellen, 1959
Pipistrellus (Pipistrellus) inexpectatus Aellen, 1959
Mops (Chaerephon) jobimena (Goodman and Cardiff, 2004)
Nanonycteris veldkampii (Jentink, 1888)
Pipistrellus (Pipistrellus) permixtus Aellen, 1957
Pipistrellus (Pipistrellus) permixtus Aellen, 1957
Nycteris vinsoni Dalquest, 1965
Epomophorus wahlbergi (Sundevall, 1846)
Rhinolophus silvestris Aellen, 1959
Eptesicus (Rhinopterus) floweri (de Winton, 1901)
Mops (Mops) niveiventer Cabrera and Ruxton, 1926
Scotophilus leucogaster (Cretzschmar, 1826)
Scotophilus leucogaster (Cretzschmar, 1826)
Pseudoromicia tenuipinnis (Peters, 1872)
Scotoecus albofuscus (Thomas, 1890)
Scotoecus albigula Thomas, 1909
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pseudoromicia tenuipinnis (Peters, 1872)
Pseudoromicia tenuipinnis (Peters, 1872)
Myotis welwitschii (Gray, 1866)
Glauconycteris poensis (Gray, 1842)
Scotophilus tandrefana Goodman, Jenkins and Ratrimomanarivo, 2005
Megaloglossus azagnyi Nesi, Kadjo and Hassanin, 2012
Myopterus whitleyi (Scharff, 1900)
Rhinolophus willardi Kerbis Peterhans and Fahr, 2013
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Epomophorus minimus Claessen & De Vree, 1991
Megaloglossus woermanni Pagenstecher, 1885
Kerivoula Gray, 1842
Nycteris woodi K. Andersen, 1914
Otonycteris hemprichii Peters, 1859
Otonycteris Peters, 1859
Myotis zenatus Ibáñez, Juste, Salicini, Puechmaille and Ruedi, 2019
Scotonycteris zenkeri Matschie, 1894
Scotonycteris zenkeri Matschie, 1894
Rhinolophus ziamo Fahr, Vierhaus, Hutterer and Kock, 2002
Neoromicia zuluensis (Roberts, 1924)
Neoromicia zuluensis (Roberts, 1924)
Mops (Chaerephon) bivittatus (Heuglin, 1861)
Mops (Xiphonycteris) nanulus J.A. Allen, 1917
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Epomophorus minor Dobson, 1880
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus Kaup, 1829
Nanonycteris Matschie, 1899
Afronycteris nana (Peters, 1852)
Miniopterus minor Peters, 1867
Nycteris nana (K. Andersen, 1912)
Doryrhina cyclops (Temminck, 1853)
Doryrhina cyclops (Temminck, 1853)

Greek

- Ατρίχη ταφονυχτερίδα
 Αιγυπτιακή φρουτονυχτερίδα
 Ασιατική τρανονυχτερίδα
 Βουνονυχτερίδα
 Λευκονυχτερίδα
 Μεγάλος νυκτοβάτης
 Μεσορινόλοφος
 Μεσορινόλοφος

Taphozous nudiventris Cretzschmar, 1830
Roussettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus blasii Peters, 1867
Rhinolophus euryale Blasius, 1853

Μικρονυκτοβάτης
 Μικρονυχτερίδα
 Μικρορινόλοφος
 Μουστακονυχτερίδα
 Μπαρμπαστέλλος
 Νανονυχτερίδα
 Νανονυχτερίδα του Hanak
 Νυκτοβάτης των Αζόρων
 Νυχτερίδας της Μαδέρας
 Νυχτονόμος
 Ποδαρομυτίδα
 Πτερυγονυχτερίδα
 Πυρρομυτίδα
 Ρινόλοφος του Blasius
 Ρινόλοφος του Mehely
 Τρανομυτίδα της Καρχηδόνας
 Τρανορινόλοφος
 Ωτονυχτερίδα της Τενερίφης
 Ωτονυχτερίδα της ερήμου
 Ωχρή τρανονυχτερίδα

Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus hipposideros (Bechstein, 1800)
Myotis mystacinus (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Nyctalus azoreum (Thomas, 1901)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Tadarida teniotis (Rafinesque, 1814)
Myotis capaccinii (Bonaparte, 1837)
Miniopterus schreibersii (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Rhinolophus blasii Peters, 1867
Rhinolophus mehelyi Matschie, 1901
Myotis punicus Felten, 1977
Rhinolophus ferrumequinum (Schreber, 1774)
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)

Hebrew

אודן
 אודן
 אפלול מצוי
 בלומף שחור
 כנפן
 לילן
 נשפון גדול
 עטלפון אירופי
 עטלפון סאבי
 פרספ בהיר
 פרספ גדול
 פרספ גמדי
 פרספ הנגב
 פרספ חיוור
 פרספ מצוי
 פרספון
 Aflul Matsui
 aflul matzuy
 Ashman Gadol
 Ashman Katan
 Atalef Perot
 Atalef Yaznuy Gadol
 atalefon eropi
 atalefon savi
 Atalfon Aeropi
 Atalfon Ariel
 Atalfon Leven Shulayim
 Atalfon Rippel
 Atalfon Savi
 Blumaf Shahor
 Kanfan
 leilan
 Leylan
 Nishpon Gadot
 Nishpon Pgoom Ozen
 Oznan
 Parsaf Bahir
 parsaf gadol
 Parsaf Gamadi
 Parsaf HaNegev
 Parsaf Hiver
 Parsaf Matsui
 parsaf matzuy
 Parsafon
 pasafon
 Pashaf Matsui
 Udnan
 Yaznuy Katan

Otonycteris hemprichii Peters, 1859
Plecotus christii Gray, 1838
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Barbastella leucomelas (Cretzschmar, 1826)
Miniopterus schreibersii (Kuhl, 1817)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Myotis capaccinii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Hypsugo savii (Bonaparte, 1837)
Rhinolophus euryale Blasius, 1853
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus mehelyi Matschie, 1901
Rhinolophus blasii Peters, 1867
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Taphozous nudiventris Cretzschmar, 1830
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rhinopoma microphyllum (Brünnich, 1782)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Hypsugo ariel (Thomas, 1904)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Vansonia rueppellii (J.B. Fischer, 1829)
Hypsugo savii (Bonaparte, 1837)
Barbastella leucomelas (Cretzschmar, 1826)
Miniopterus schreibersii (Kuhl, 1817)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Myotis capaccinii (Bonaparte, 1837)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Plecotus christii Gray, 1838
Rhinolophus euryale Blasius, 1853
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus mehelyi Matschie, 1901
Rhinolophus blasii Peters, 1867
Rhinolophus blasii Peters, 1867
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Asellia tridens (E. Geoffroy St.-Hilaire, 1813)
Tadarida teniotis (Rafinesque, 1814)
Otonycteris hemprichii Peters, 1859
Rhinopoma cystops Thomas, 1903

Hungarian

Alpesi denevér

Hypsugo savii (Bonaparte, 1837)

Azori-szigeteki koraidenevér
 Bajuszos denevér
 Botta-késeidenevér
 Csonkafülű denevér
 Csúcsosnyergű patkósdenevér
 Csupaszhasú sárlakó-denevér
 Európai hosszúszárnyú-denevér
 Európai szabadfarkú-denevér
 Fehérszélű törpedenevér
 Hanák-törpedenevér
 Hosszúlábú denevér
 Hosszúszárnyú denevér
 Izabella-késeidenevér
 Kanári-szigeteki hosszúfülű-denevér
 Kereknyergű patkósdenevér
 Kereknyergű patkósorrú denevér
 Kis patkósdenevér
 Kis patkósorrú denevér
 Közönséges törpedenevér
 Madeirai törpedenevér
 Magrebi denevér
 Méhely-patkósdenevér
 Nagy patkósdenevér nagy patkósorrú denevér
 Nílusi repülőkutya
 Nycteris i alpius
 Nycteris i blepharodoti
 Nycteris i kini
 Nycteris i leukogyros
 Nycteris i macropterys
 Nycteris i macropus
 Nycteris i mikromolossos
 Nycteris i mystakophoros
 Nycteris i urophoros
 Nycteris pterygisti i megalis
 Nycteris pterygisti i mikra
 Nyugati piszedenevér
 Óriás koraidenevér
 Píse denevér
 Písedenevér
 Rhinolophos i megalis
 Rhinolophos i meheli
 Rhinolophos i mikra
 Rhinolophos i mesogiaki
 Sivatagi hosszúfülű-denevér
 Szőrös karú denevér
 Szőrös karú koraidenevér
 Szoprán törpedenevér
 Törpe denevér
 Törpe denevér

Nyctalus azoreum (Thomas, 1901)
Myotis mystacinus (Kuhl, 1817)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Rhinolophus blasii Peters, 1867
Taphozous nudiventris Cretzschmar, 1830
Miniopterus schreibersii (Kuhl, 1817)
Tadarida teniotis (Rafinesque, 1814)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Myotis capaccinii (Bonaparte, 1837)
Miniopterus schreibersii (Kuhl, 1817)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Plecotus teneriffae Barrett-Hamilton, 1907
Rhinolophus euryale Blasius, 1853
Rhinolophus euryale Blasius, 1853
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Myotis punicus Felten, 1977
Rhinolophus mehelyi Matschie, 1901
Rhinolophus ferrumequinum (Schreber, 1774)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Hypsugo savii (Bonaparte, 1837)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Barbastella barbastellus (Schreber, 1774)
Myotis mystacinus (Kuhl, 1817)
Tadarida teniotis (Rafinesque, 1814)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Nyctalus lasiopterus (Schreber, 1780)
Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus mehelyi Matschie, 1901
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus euryale Blasius, 1853
Otonycteris hemprichii Peters, 1859
Nyctalus leisleri (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Miniopterus schreibersii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)

Indonesian

Kelelawar ekor lembing
 Kelelawar Ekor-tikus
 Kelelawar mini biasa
 Kelelawar mini schreiber
 Kelelawar myotis pucuk
 Tomosu Biasa

Rhinopoma microphyllum (Brünnich, 1782)
Rhinopoma microphyllum (Brünnich, 1782)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)

Irish Gaelic

Crú-ialtóg bheag
 Crú-ialtóg Blasius
 Crú-ialtóg Mehely
 Crú-ialtóg Mheánmhuirí
 Crú-ialtóg mhór
 Ialtóg Asórach
 Ialtóg bharbastail Iartharach
 Ialtóg Botta
 Ialtóg earrshaor Eorpach
 Ialtóg fhadchluasach Chanárach
 Ialtóg fhadchluasach Hemprich
 Ialtóg fhadmhéarach
 Ialtóg fheascrach
 Ialtóg Geoffroy

Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus blasii Peters, 1867
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Rhinolophus ferrumequinum (Schreber, 1774)
Nyctalus azoreum (Thomas, 1901)
Barbastella barbastellus (Schreber, 1774)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Tadarida teniotis (Rafinesque, 1814)
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859
Myotis capaccinii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)

laltóg ghiobach
laltóg Hanak
laltóg isibéalach
laltóg Kuhl
laltóg Leisler
laltóg luch-chluasach Mhaigribeach
laltóg meas Éigipteach
laltóg Mhaidéarach
laltóg mhéarfhada Schreiber
laltóg nochtphrampach thuama
laltóg Savi
laltóg shopránach
Ollialtóg nochtúil

Myotis mystacinus (Kuhl, 1817)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Myotis punicus Felten, 1977
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Miniopterus schreibersii (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Nyctalus lasiopterus (Schreber, 1780)

isiXhosa

idludaka
ilulwane

CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779

Italian

Asèllia di Patrizi
Asèllia italosòmala
Asèllie
Barbastello
Cardiodèrma
Cardiodèrma nàso a cuòre
Carlino brachittero
Carlino condilùro
Carlino Mida
Carlino setolòso
Cherefónte bivittàto
Cherefónte di Ansòrge
Cherefónte nigeriàno
Cherefónte pigmèò
Cherefónti
Cherivoula di Smith
Cherivoule
Chiròtreri
Cloeòtide di Pèrcival
Cloeòtidi
Còleura africàna
Èidoli
Èidolo paglierìno
Emballonùridi
Epomòfori
Epomòforo di Wählberg
Epomòforo labiàto
Epomòforo mìnimò
Fàlsi vampìri
Fàlsi vampìri dàlle àli gialle
Fàlso vampìre dàlle ali gialle
Fàlso vampìro dàl nàso a cuòre
Fèrri di cavàllo
Fèrro di cavàllo affumicàto
Fèrro di cavàllo di Blàsìus
Fèrro di cavàllo di Cretzschmar
Fèrro di cavàllo di Dècken
Fèrro di cavàllo di Hildebrandt
Fèrro di cavàllo di Méhely
Fèrro di cavàllo eloquènte
Fèrro di cavàllo Euriale
Fèrro di Cavàllo Maggiore
ferro di cavàllo mediterraneò
Fèrro di Cavàllo Minore
Fèrro di cavàllo simulatòre
Glaucônictèride argentàta
Glaucônictèride keniòta
Glaucônictèride variegàta
Glaucônitteride argentàta
Glaucônitteride keniòta
Glaucônitteride variegàta
Glaucônitterìdi
Ipposidèri
Ipposidèridi

Asellia patrizii de Beaux, 1931
Asellia italosomalica de Beaux, 1931
Asellia Gray, 1838
Barbastella barbastellus (Schreber, 1774)
Cardioderma Peters, 1873
Cardioderma cor (Peters, 1872)
Mops (Xiphonycteris) brachypterus (Peters, 1852)
Mops (Mops) condylurus (A. Smith, 1833)
Mops (Mops) midas (Sundevall, 1843)
Platymops setiger (Peters, 1878)
Mops (Chaerephon) bivittatus (Heuglin, 1861)
Mops (Chaerephon) ansorgei (Thomas, 1913)
Mops (Chaerephon) nigeriae (Thomas, 1913)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Mops (Chaerephon) Dobson, 1874
Kerivoula smithii Thomas, 1880
Kerivoula Gray, 1842
CHIROPTERA Blumenbach, 1779
Cloeotis percivali Thomas, 1901
Cloeotis Thomas, 1901
Coleura afra (Peters, 1852)
Eidolon Rafinesque, 1815
Eidolon helvum (Kerr, 1792)
EMBALLONURIDAE Gervais, 1855
Epomophorus Bennett, 1836
Epomophorus wahlbergi (Sundevall, 1846)
Epomophorus labiatus (Temminck, 1837)
Epomophorus minimus Claessen & De Vree, 1991
MEGADERMATIDAE H. Allen, 1864
Lavia Gray, 1838
Lavia frons (E. Geoffroy St.-Hilaire, 1810)
Cardioderma cor (Peters, 1872)
Rhinolophus Lacépède, 1799
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus blasii Peters, 1867
Rhinolophus clivosus Cretzschmar, 1828
Rhinolophus deckenii Peters, 1868
Rhinolophus hildebrandtii Peters, 1878
Rhinolophus mehelyi Matschie, 1901
Rhinolophus eloquens K. Andersen, 1905
Rhinolophus euryale Blasius, 1853
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus simulator K. Andersen, 1904
Glauconycteris argentata (Dobson, 1875)
Glauconycteris kenyacola Peterson, 1982
Glauconycteris variegata (Tomes, 1861)
Glauconycteris argentata (Dobson, 1875)
Glauconycteris kenyacola Peterson, 1982
Glauconycteris variegata (Tomes, 1861)
Glauconycteris Dobson, 1875
Hipposideros Gray, 1831
HIPPOSIDERIDAE Lydekker, 1891

- Ipposidero càfro
 Ipposidero orecchiùto
 Ipposidero vittàto
 Ipsùghi
 Lefòte di Winton
 Lefòti
 Lissonictèride angolàna
 Lissonittèride angolàna
 Megadermatìdi
 Mimetilli
 Mimetillo di Moloney
 Miniòptere minóre
 Miniòptèridi
 Miniòptero di Schrèibers
 Miniòptero maggióre di Thòmas
 Miniottèridi
 Miniottero
 Miniottero di Schrèibers
 Miniottero maggióre di Thòmas
 Miniottero minóre
 Molòssidi
 Molosso del Cestoni
 Neoromicie
 Nictèride di Dobsòn
 Nictèride di Parìsi
 Nictèride di Tèbe
 Nictèride irsùta
 Nictèride ìspida
 Nictèride orecchiùta
 Nictèride tebàna
 Nictèridi
 Nicticèinope di Schlieffen
 Nicticèinopi
 Nitteride di Dòbson
 Nitteride di Parìsi
 Nitteride di Tèbe
 Nitteride irsùta
 Nitteride ìspida
 Nitteride orecchiùta
 Nitteride tebàna
 Nitteridi
 Nitticèinope di Schlieffen
 Nitticèinopi
 Nottola delle Azzorre
 Nottola di Leisler
 Nottola gigante
 Orecchione di Gaisler
 Orecchione di Hemprich
 Orecchione di Kolombatovic
 Orecchione di Tenerife
 Otocarlino
 Otocarlino di Martienssèn
 Pipistrèlli
 Pipistrèlli
 Pipistrèlli a còda di tòpo
 Pipistrèlli dèlle tòmbe
 Pipistrello albolimbato
 Pipistrèllo capènse
 Pipistrèllo carlino
 Pipistrello de Kühli
 Pipistrello delle tombe a ventre nudo
 Pipistrèllo di Eisentràut
 Pipistrello di Hanak
 Pipistrèllo di Hèller
 Pipistrello di Madeira
 Pipistrèllo di Peters
 Pipistrèllo di Pèters
 Pipistrèllo di Rëndall
 Pipistrèllo di Rùppell
 Pipistrello di Savi
 Pipistrèllo giallàstro
 Pipistrèllo guineàno
 Pipistrello nano
 Pipistrèllo nàno déi banàni
 Pipistrello pigmeo
 Pipistrèllo rugginóso
Hipposideros caffer (Sundevall, 1846)
Hipposideros megalotis (Heuglin, 1861)
Macronycteris vittatus (Peters, 1852)
Hypsugo Kolenati, 1856
Laephotis wintoni Thomas, 1901
Laephotis Thomas, 1901
Myonycteris angolensis (Bocage, 1898)
Myonycteris angolensis (Bocage, 1898)
 MEGADERMATIDAE H. Allen, 1864
Mimetillus Thomas, 1904
Mimetillus moloneyi (Thomas, 1891)
Miniopterus minor Peters, 1867
 MINIOPTERIDAE Dobson, 1875
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus inflatus Thomas, 1903
 MINIOPTERIDAE Dobson, 1875
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus inflatus Thomas, 1903
Miniopterus minor Peters, 1867
 MOLOSSIDAE Gervais, 1856
Tadarida teniotis (Rafinesque, 1814)
Neoromicia Roberts, 1926
Nycteris macrotis Dobson, 1876
Nycteris parisii (de Beaux, 1924)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris hispida (Schreber, 1774)
Nycteris hispida (Schreber, 1774)
Nycteris aurita (K. Andersen, 1912)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris G. Cuvier and E. Geoffroy, 1795
Nycticeinops schlieffenii (Peters, 1859)
Nycticeinops Hill & Harrison, 1987
Nycteris macrotis Dobson, 1876
Nycteris parisii (de Beaux, 1924)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris hispida (Schreber, 1774)
Nycteris hispida (Schreber, 1774)
Nycteris aurita (K. Andersen, 1912)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris G. Cuvier and E. Geoffroy, 1795
Nycticeinops schlieffenii (Peters, 1859)
Nycticeinops Hill & Harrison, 1987
Nyctalus azoreum (Thomas, 1901)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus lasiopterus (Schreber, 1780)
Plecotus gaisleri Benda, Kiefer, Hanak & Veith, 2004
Otonycteris hemprichii Peters, 1859
Plecotus kolombatovici Dulic, 1980
Plecotus teneriffae Barrett-Hamilton, 1907
Otomops Thomas, 1913
Otomops martiensseni (Matschie, 1897)
 CHIROPTERA Blumenbach, 1779
Pipistrellus Kaup, 1829
 RHINOPOMATIDAE Dobson, 1872
Taphozous E. Geoffroy St.-Hilaire, 1818
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Laephotis capensis (A. Smith, 1829)
Mops Lesson, 1842
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Nycticeinops eisentrauti (Hill, 1968)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Afronycteris helios (Heller, 1912)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pseudoromicia tenuipinnis (Peters, 1872)
Afronycteris nana (Peters, 1852)
Pseudoromicia rendalli (Thomas, 1889)
Vansonia rueppellii (J.B. Fischer, 1829)
Hypsugo savii (Bonaparte, 1837)
Nycticeinops grandidieri (Dobson, 1876)
Neoromicia guineensis (Bocage, 1889)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Afronycteris nana (Peters, 1852)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) rusticus (Tomes, 1861)

Pipistrèllo seràle	<i>Pipistrellus (Pipistrellus) hesperidus</i> (Temminck, 1840)
Pipistrèllo sòmalo	<i>Neoromicia somalica</i> (Thomas, 1901)
Pipistrello soprano	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Pipistrello soprano	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Platicarlini	<i>Platymops</i> Thomas, 1906
Ptèropi fantasma	<i>Eidolon</i> Rafinesque, 1815
Pteròpidi	<i>PTEROPODIDAE</i> Gray, 1821
Ptèropo fantasma paglierino	<i>Eidolon helvum</i> (Kerr, 1792)
Pteropòdidi	<i>PTEROPODIDAE</i> Gray, 1821
Rhinopoma	<i>Rhinopoma</i> E. Geoffroy St.-Hilaire, 1818
Rhinopoma afro-ovestasiatico	<i>Rhinopoma cystops</i> Thomas, 1903
Rinòlofi	<i>Rhinolophus</i> Lacépède, 1799
Rinolòfidi	<i>RHINOLOPHIDAE</i> Gray, 1825
Rinòlofo affumicàto	<i>Rhinolophus fumigatus</i> Rüppell, 1842
Rinòlofo di Blàsius	<i>Rhinolophus blasii</i> Peters, 1867
Rinòlofo di Cretzschmar	<i>Rhinolophus clivosus</i> Cretzschmar, 1828
Rinòlofo di Dècken	<i>Rhinolophus deckenii</i> Peters, 1868
Rinòlofo di Hildebrandt	<i>Rhinolophus hildebrandtii</i> Peters, 1878
Rinòlofo di Lànder	<i>Rhinolophus landeri</i> Martin, 1838
Rinòlofo di Méhely	<i>Rhinolophus mehelyi</i> Matschie, 1901
Rinòlofo eloquente	<i>Rhinolophus eloquens</i> K. Andersen, 1905
Rinòlofo Euriale	<i>Rhinolophus euryale</i> Blasius, 1853
Rinòlofo maggiore	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Rinòlofo mediterraneo	<i>Rhinolophus euryale</i> Blasius, 1853
Rinòlofo minore	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Rinòlofo simulatòre	<i>Rhinolophus simulator</i> K. Andersen, 1904
Rinopoma di MacInnes	<i>Rhinopoma macinnesi</i> Hayman, 1937
Rinopoma microfillo	<i>Rhinopoma microphyllum</i> (Brünnich, 1782)
Rinopormatidi	<i>RHINOPOMATIDAE</i> Dobson, 1872
Rossetta	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Rosséti	<i>Rousettus</i> Gray, 1821
Rossétto del Nilo	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Rossétto egiziano	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Scotoèchi	<i>Scotoecus</i> Thomas, 1901
Scotoeco di Hinde	<i>Scotoecus hindei</i> Thomas, 1901
Scotoeco golabianca	<i>Scotoecus albigula</i> Thomas, 1909
Scotoeco røndine	<i>Scotoecus hirundo</i> (de Winton, 1899)
Scotòfili	<i>Scotophilus</i> Leach, 1821
Scotòfilo di Dìngan	<i>Scotophilus dinganii</i> (A. Smith, 1833)
Scotòfilo panciabianca	<i>Scotophilus leucogaster</i> (Cretzschmar, 1826)
Scotòfilo scùro	<i>Scotophilus nigrita</i> (Schreber, 1774)
Scotòfilo vèrde	<i>Scotophilus viridis</i> (Peters, 1852)
Serotino di Botta	<i>Eptesicus (Cnephaeus) bottae</i> (Peters, 1869)
Serotino isabellino	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
Tadarida egiziana	<i>Tadarida aegyptiaca</i> (E. Geoffroy St.-Hilaire, 1818)
Tadarida ventrale	<i>Tadarida ventralis</i> (Heuglin, 1861)
Tadaride	<i>Tadarida</i> Rafinesque, 1814
Tafozò	<i>Taphozous</i> E. Geoffroy St.-Hilaire, 1818
Tafozò di Håmilton	<i>Taphozous hamiltoni</i> Thomas, 1920
Tafozò di Hildegarde	<i>Taphozous hildegardeae</i> Thomas, 1909
Tafozò mauriziano	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
Tafozò nudiventre	<i>Taphozous nudiventris</i> Cretzschmar, 1830
Tafozò perforato	<i>Taphozous perforatus</i> E. Geoffroy St.-Hilaire, 1818
Triènope afro	<i>Triaenops afer</i> Peters, 1877
Triènopi	<i>Triaenops</i> Dobson, 1871
Vespertilio di Capaccini	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Vespertilio maghrebino	<i>Myotis punicus</i> Felten, 1977
Vespertilio mustacchino	<i>Myotis mystacinus</i> (Kuhl, 1817)
Vespertilio smarginato	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Vespertilio smarginato	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Vespertiliònidi	<i>VESPERTILIONIDAE</i> Gray, 1821

Japanese

Pita-ohiki-koumori	<i>Mormopterus jugularis</i> (Peters, 1865)

Kabyle (Algeria)

Imtchaghyeye	<i>CHIROPTERA</i> Blumenbach, 1779

Kiande (DRC)

Kakorokombe	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795

Kiluba (DRC)

Kasusu	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
Kasusu	<i>Rhinolophus</i> Lacépède, 1799
Kasusu	<i>Hipposideros caffer</i> (Sundevall, 1846)
Kasusu	<i>Rhinolophus hildebrandtii</i> Peters, 1878
Kasusu	<i>Rhinolophus landeri</i> Martin, 1838
Kasusu	<i>Cloeotis percivali</i> Thomas, 1901
Kasusu	<i>Nycteris</i> G. Cuvier and E. Geoffroy, 1795
Kasusu	<i>Nycteris thebaica</i> E. Geoffroy St.-Hilaire, 1818
Kasusu	<i>Mops (Chaerephon) pumilus</i> (Cretzschmar, 1826)
Kasusu	<i>Kerivoula argentata</i> Tomes, 1861
Kasusu	<i>Afronycteris nana</i> (Peters, 1852)
Kasusu	<i>Laephotis capensis</i> (A. Smith, 1829)
Kasusu	<i>Nycticeinops schlieffenii</i> (Peters, 1859)
Katutu	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
Katutu	<i>Nycteris arge</i> Thomas, 1903
Katutu	<i>Nycteris hispida</i> (Schreber, 1774)
Mulima	<i>Epomophorus</i> Bennett, 1836
Mulima	<i>Epomophorus pusillus</i> Peters, 1868
Mulima	<i>Myonycteris angolensis</i> (Bocage, 1898)
Mulima	<i>Plerotes anchietae</i> (Seabra, 1900)
Mulima	<i>Myotis welwitschii</i> (Gray, 1866)

Kinande (DRC)

Apopo	<i>Epomophorus labiatus</i> (Temminck, 1837)
Kahazo-komule	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
Kakoro kombe	<i>Lavia frons</i> (E. Geoffroy St.-Hilaire, 1810)
Kakorokombe	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818
Karakonube	<i>Stenonycteris lanosus</i> (Thomas, 1906)
Mulima	<i>Eidolon helvum</i> (Kerr, 1792)
Mulima	<i>Myonycteris angolensis</i> (Bocage, 1898)
Mulima	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Mulima	<i>Macronycteris gigas</i> (Wagner, 1845)
Mulima	<i>Otomops martiensseni</i> (Matschie, 1897)
Mulina	<i>Hypsignathus monstrosus</i> H. Allen, 1862

Kinyarwanda

Mokombe	<i>Taphozous mauritanus</i> E. Geoffroy St.-Hilaire, 1818

Latvian

Ēģiptes augšsīkspārnis	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Šreibersa sīkspārnis	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Azoru vakarsīkspārnis	<i>Nyctalus azoreum</i> (Thomas, 1901)
Bārdainais naktssīkspārnis	<i>Myotis mystacinus</i> (Kuhl, 1817)
Blāzija pakavdegunis	<i>Rhinolophus blasii</i> Peters, 1867
Botas sīkspārnis	<i>Eptesicus (Cnephaeus) bottae</i> (Peters, 1869)
Brivastes sīkspārnis	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Eiropas platausis	<i>Barbastella barbastellus</i> (Schreber, 1774)
Garpirkstu naktssīkspārnis	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Hanaka sīkspārnis	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Hempriha sīkspārnis	<i>Otonycteris hemprichii</i> Peters, 1859
Izabellas sīkspārnis	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
Kūla sīkspārnis	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Kailvēdera sīkspārnis	<i>Taphozous nudiventris</i> Cretzschmar, 1830
Lielais pakavdegunis	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Lielais vakarsīkspārnis	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Madeiras sīkspārnis	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Mazais pakavdegunis	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Mazais vakarsīkspārnis	<i>Nyctalus leisleri</i> (Kuhl, 1817)
Meheļa pakavdegunis	<i>Rhinolophus mehelyi</i> Matschie, 1901
Pūnijas naktssīkspārnis	<i>Myotis punicus</i> Felten, 1977
Pigmejsīkspārnis	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Pundursīkspārnis	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Sāvi sīkspārnis	<i>Hypsugo savii</i> (Bonaparte, 1837)
Skropstainais naktssīkspārnis	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Tenerifes garausis	<i>Plecotus teneriffae</i> Barrett-Hamilton, 1907
Vidusjuras pakavdegunis	<i>Rhinolophus euryale</i> Blasius, 1853

Lithuanian

Šikšniukas mažylis	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Šikšniukas nykštukas	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Šreiberio ilgapištis	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Ūsuotasis pelėausis	<i>Myotis mystacinus</i> (Kuhl, 1817)
Azorinis nakviša	<i>Nyctalus azoreum</i> (Thomas, 1901)
Blazijaus pasagnosis	<i>Rhinolophus blasii</i> Peters, 1867
Didysis nakviša	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Didysis pasagnosis	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Egiptinis vaisėdis	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Europinis plačiaausis	<i>Barbastella barbastellus</i> (Schreber, 1774)
Europinis raukšlėtūpis	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Finikinis pelėausis	<i>Myotis punicus</i> Felten, 1977
Hanako šikšniukas	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Ilgapištis pelėausis	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Izabelos šikšnys	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
Kanarinis ausylis	<i>Plecotus teneriffae</i> Barrett-Hamilton, 1907
Kulio šikšniukas	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Mažasis nakviša	<i>Nyctalus leisleri</i> (Kuhl, 1817)
Mažasis pasagnosis	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Madeirinis šikšniukas	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Mehelio pasagnosis	<i>Rhinolophus mehelyi</i> Matschie, 1901
Pietinis pasagnosis	<i>Rhinolophus euryale</i> Blasius, 1853
Plikapilvis maišiarnaris	<i>Taphozous nudiventris</i> Cretzschmar, 1830
Savio šikšniukas	<i>Hypsugo savii</i> (Bonaparte, 1837)
Strėliausis lygianosis	<i>Otonycteris hemprichii</i> Peters, 1859
Trispalvis pelėausis	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Uolinis šikšnys	<i>Eptesicus (Cnephaeus) bottae</i> (Peters, 1869)

Luxembourgish

Alpen-Fledermaus	<i>Hypsugo savii</i> (Bonaparte, 1837)
Azoren-Bëschrfliedermaus	<i>Nyctalus azoreum</i> (Thomas, 1901)
Blasius-Huffeisennues	<i>Rhinolophus blasii</i> Peters, 1867
Botta-Fledermaus	<i>Eptesicus (Cnephaeus) bottae</i> (Peters, 1869)
Europäesch Bulldogfledermaus	<i>Tadarida teniotis</i> (Rafinesque, 1814)
Europäesch Mopsfledermaus	<i>Barbastella barbastellus</i> (Schreber, 1774)
Grouss Huffeisennues	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Isabel-Fledermaus	<i>Eptesicus (Cnephaeus) isabellinus</i> (Temminck, 1840)
Kanaren-Laangouer	<i>Plecotus teneriffae</i> Barrett-Hamilton, 1907
Kleng Baartfledermaus	<i>Myotis mystacinus</i> (Kuhl, 1817)
Kleng Bëschrfliedermaus	<i>Nyctalus leisleri</i> (Kuhl, 1817)
Kleng Fledermaus	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)
Kleng Huffeisennues	<i>Rhinolophus hipposideros</i> (Bechstein, 1800)
Laangflillekefledermaus	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Laangfoussfledermaus	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Libesch Zwergfledermaus	<i>Pipistrellus (Pipistrellus) hanaki</i> Hulva and Benda, 2004
Madeira-Zwergfledermaus	<i>Pipistrellus (Pipistrellus) maderensis</i> (Dobson, 1878)
Maghreb-Mausouer	<i>Myotis punicus</i> Felten, 1977
Méckefledermaus	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Méhely-Huffeisennues	<i>Rhinolophus mehelyi</i> Matschie, 1901
Mëttelmier-Huffeisennues	<i>Rhinolophus euryale</i> Blasius, 1853
Nilfluchhond	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)
Plakegbauchhege Griewerflatterer	<i>Taphozous nudiventris</i> Cretzschmar, 1830
Riseg Bëschrfliedermaus	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Wäissbordfledermaus	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Wimperefledermaus	<i>Myotis emarginatus</i> (E. Geoffroy St.-Hilaire, 1806)
Wüstegroussouer	<i>Otonycteris hemprichii</i> Peters, 1859
Zwergfledermaus	<i>Pipistrellus (Pipistrellus) pipistrellus</i> (Schreber, 1774)

Macedonian

Џуцест лилјак	<i>Pipistrellus (Pipistrellus) pygmaeus</i> (Leach, 1825)
Азорски вечерник	<i>Nyctalus azoreum</i> (Thomas, 1901)
Белорабен лилјак [= Beloraben Liljak]	<i>Pipistrellus (Pipistrellus) kuhlii</i> (Kuhl, 1817)
Бласиев потковичар [= Blasiev Potkovichar]	<i>Rhinolophus blasii</i> Peters, 1867
Ботев лилјак	<i>Eptesicus (Cnephaeus) bottae</i> (Peters, 1869)
Голем вечерник	<i>Nyctalus lasiopterus</i> (Schreber, 1780)
Голем потковичар [= Golem Potkovichar]	<i>Rhinolophus ferrumequinum</i> (Schreber, 1774)
Долгокрилест лилјак	<i>Miniopterus schreibersii</i> (Kuhl, 1817)
Долгопрст ноћник [= Dolgoprst Nokjnik]	<i>Myotis capaccinii</i> (Bonaparte, 1837)
Египетски овошен лилјак	<i>Rousettus aegyptiacus</i> (E. Geoffroy St.-Hilaire, 1810)

Изабелов лилјак
 Магребски ноќник
 Мал лилјак
 Мал потковичар [= Mal Potkovichar]
 Медеирски лилјак
 Медитерански потковичар
 Мехелов потковичар
 Мустаќест ноќник [= Mustakjest Nokjnik]
 Опашест лилјак [= Opashest Liljak]
 Савиев лилјак [= Saviev Liljak]
 Тенерифов ушест лилјак
 Торбокрилест лилјак
 Тробоен ноќник [= Troboen Nokjnik]
 Ханаков лилјак
 Хемпричиев долгоушест лилјак
 Широкоушест лилјак
 Шумски вечерник [= Shumski Vechernik]
 Dolgokrilest Liljak
 Dzudzest Liljak
 Juzhen Potkovichar
 Kafeav Dzudzest Liljak

Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Myotis punicus Felten, 1977
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Myotis mystacinus (Kuhl, 1817)
Tadarida teniotis (Rafinesque, 1814)
Hypsugo savii (Bonaparte, 1837)
Plecotus teneriffae Barrett-Hamilton, 1907
Taphozous nudiventris Cretzschmar, 1830
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Otonycteris hemprichii Peters, 1859
Barbastella barbastellus (Schreber, 1774)
Nyctalus leisleri (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)

Malagasy

Angavo
 Fanihy
 kitrotroke

Eidolon dupreanum (Schegel, 1867)
Pteropus rufus E. Geoffroy St.-Hilaire, 1803
Miniopterus mahafaliensis Goodman, Bradman, Christides and Appleton, 2009

Maltese

Barbastell
 Farfett il-Lejl Daqni
 Farfett il-Lejl Ghieri
 Farfett il-Lejl ta' Widnejh Imnaqqxa
 Farfett il-Lejl ta' Xrajber
 Farfett il-Lejl tal-Frott ta' L-Egittu
 Farfett il-Lejl tan-Nala
 Farfett il-Lejl tan-Nala kbir
 Farfett il-Lejl tas-Swaba Twal
 Farfett il-Lejl Widnejh Twal tad-Dežert
 Farfett il-Lejl Widnejh Twal tal-Gžejjer Kanarji
 Farfett il-Lejl Widnet il-gurdien ta' Maghreb
 Farfett il-Lejl Widnet il-Gurdien
 Noktula Žghir
 Noktula Kbir
 Noktula ta' l-Azores
 Pipistrell
 Pipistrell Žghir
 Pipistrell Komuni
 Pipistrell ta' Hanak
 Pipistrell ta' Kuhl
 Pipistrell ta' Madejra
 Pipistrell ta' Savi
 Rinolofu Žghir
 Rinolofu Kbir
 Rinolofu ta' Blasius
 Rinolofu ta' Mehely
 Rinolofu tal-Mediterran
 Serotin ta' Botta
 Serotin tas-Sider Pallidu
 Tadarida

Barbastella barbastellus (Schreber, 1774)
Myotis mystacinus (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Miniopterus schreibersii (Kuhl, 1817)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus ferrumequinum (Schreber, 1774)
Myotis capaccinii (Bonaparte, 1837)
Otonycteris hemprichii Peters, 1859
Plecotus teneriffae Barrett-Hamilton, 1907
Myotis punicus Felten, 1977
Myotis punicus Felten, 1977
Nyctalus leisleri (Kuhl, 1817)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus azoreum (Thomas, 1901)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Hypsugo savii (Bonaparte, 1837)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus blasii Peters, 1867
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Tadarida teniotis (Rafinesque, 1814)

Manganja (Malawi)

Chuchu

Afronycteris nana (Peters, 1852)

Misuku (Malawi)

Kashusha

Afronycteris nana (Peters, 1852)

Montenegrin

Šumski noćnik

Nyctalus leisleri (Kuhl, 1817)

širokouhi slijepi miš
 Azorski noćnik
 Bijeloruski slijepi mišić
 Brkati večernjak
 Dugoprsti večernjak
 Dugorepi slijepi miš
 Dugorili prstenjak
 Egipatski letipas
 Golotrbušni grobni slijepi miš
 Južni potkovičar
 Kanarski slijepi miš
 Kraljičin slijepi miš
 Libijski patuljasti slijepi mišić
 Mali potkovičar
 Mali slijepi mišić
 Mehelijev potkovičar
 Patuljasti slijepi mišić
 Punski večernjak
 Pustinski škorpiojed
 Ridi večernjak
 Savijev slijepi mišić
 Slijepi mišić sa Madeire
 Sredozemni potkovičar
 Svijetlotrbušasti slijepi miš
 Veliki noćnik
 Veliki potkovičar

Barbastella barbastellus (Schreber, 1774)
Nyctalus azoreum (Thomas, 1901)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Tadarida teniotis (Rafinesque, 1814)
Miniopterus schreibersii (Kuhl, 1817)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Taphozous nudiventris Cretzschmar, 1830
Rhinolophus euryale Blasius, 1853
Plecotus teneriffae Barrett-Hamilton, 1907
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rhinolophus mehelyi Matschie, 1901
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Myotis punicus Felten, 1977
Otonycteris hemprichii Peters, 1859
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Rhinolophus blasii Peters, 1867
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)

Nepali

Bange Chamero
 Thulo Ghodnale Chamero

Miniopterus schreibersii (Kuhl, 1817)
Rhinolophus ferrumequinum (Schreber, 1774)

Norwegian

Ørkenlangøre
 Afrikagrotteflygehund
 Asorflaggermus
 Balkanhesteskonese
 Blekflaggermus
 Bredøre
 Buldoggflaggermus
 Dvergflaggermus
 Dvergflaggermus
 Dverghesteskonese
 Ekte hesteskonese
 falske hestekoneser
 falske vampyrer
 Flyvende hunder
 foldeleppede flaggermus
 Frihalete flaggermus
 Geoffroyflaggermus
 Glattneseflaggermus
 Glattneser
 gravflaggermus
 Hestekoneser
 Hullneser
 Hvitrandflaggermus
 Isabellaflaggermus
 kalong
 Kanarilangøre
 klaffneser
 Klippeflaggermus
 Kyrenaikaflaggermus
 Langfotflaggermus
 Langhalete flaggermus
 Liten hestekoneser
 Madeiraflaggermus
 Mahgrebmusø re
 Middelhavsfrihale
 Middelhavshestekoneser
 Middelhavshestekoneser
 Middelhavslangvinge
 Nakengumpflaggermus
 Riseflaggermus
 Rundbladneser

Otonycteris hemprichii Peters, 1859
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Nyctalus azoreum (Thomas, 1901)
Rhinolophus blasii Peters, 1867
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Barbastella barbastellus (Schreber, 1774)
 MOLOSSIDAE Gervais, 1856
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus hipposideros (Bechstein, 1800)
 RHINOLOPHIDAE Gray, 1825
 HIPPOSIDERIDAE Lydekker, 1891
 MEGADERMATIDAE H. Allen, 1864
 PTEROPODIDAE Gray, 1821
 MOLOSSIDAE Gervais, 1856
 EMBALLONURIDAE Gervais, 1855
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
 VESPERTILIONIDAE Gray, 1821
 VESPERTILIONIDAE Gray, 1821
 EMBALLONURIDAE Gervais, 1855
 RHINOLOPHIDAE Gray, 1825
 NYCTERIDAE Van der Hoeven, 1855
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
 PTEROPODIDAE Gray, 1821
Plecotus teneriffae Barrett-Hamilton, 1907
 RHINOPOMATIDAE Dobson, 1872
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Myotis capaccinii (Bonaparte, 1837)
 RHINOPOMATIDAE Dobson, 1872
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Myotis punicus Felten, 1977
Tadarida teniotis (Rafinesque, 1814)
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Miniopterus schreibersii (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Nyctalus lasiopterus (Schreber, 1780)
 HIPPOSIDERIDAE Lydekker, 1891

Skjeggflaggermus
 Stor hesteskonese
 Storhesteskonese
 Storøreflaggermus
 striflaggermus
 Sugeskålflaggermus
 Tusseflaggermus
 Vetteflaggermus
 Vinkeløreflaggermus

Myotis mystacinus (Kuhl, 1817)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
 MEGADERMATIDAE H. Allen, 1864
 NYCTERIDAE Van der Hoeven, 1855
 MYZOPODIDAE Thomas, 1904
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Nyctalus leisleri (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)

Nyanja (Malawi)

Mleme

Epomophorus crypturus Peters, 1852

Nyungwe (Malawi)

Kalemawalema (not specific)
 Kalemawalema (not specific)

Hipposideros caffer (Sundevall, 1846)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)

Polish

Borowiaczek
 Borowiec azorski
 Borowiec olbrzymi
 Gacek kanaryjski
 Grobownik gołobrzuchy
 Karlik średni
 Karlik cyrenejski
 Karlik drobny
 Karlik Kuhla
 Karlik maderski
 Karlik malutki
 Molos europejski
 Mopek
 Mopek zachodni
 Mroczek Botty
 Mroczek pustynny
 mroczkowate
 Nocek długopalcy
 Nocek orzęsiony
 Nocek punicki
 Nocek wasatek
 nocki
 Podkasaniec Schreibersa
 podkowcowate
 Podkowiec średni
 Podkowiec śródziemnomorski
 Podkowiec Blasiusa
 Podkowiec duży
 Podkowiec mały
 Pzymroczek Saviego
 Rudawka nilowa
 Uszorek pustynny

Nyctalus leisleri (Kuhl, 1817)
Nyctalus azoreum (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Plecotus teneriffae Barrett-Hamilton, 1907
Taphozous nudiventris Cretzschmar, 1830
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Tadarida teniotis (Rafinesque, 1814)
Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
 VESPERTILIONIDAE Gray, 1821
Myotis capaccinii (Bonaparte, 1837)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis punicus Felten, 1977
Myotis mystacinus (Kuhl, 1817)
Myotis Kaup, 1829
Miniopterus schreibersii (Kuhl, 1817)
 RHINOLOPHIDAE Gray, 1825
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Rhinolophus blasii Peters, 1867
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Hypsugo savii (Bonaparte, 1837)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Otonycteris hemprichii Peters, 1859

Portuguese

Morcego amarelo pequeno
 Morcego anao de Kuhl
 Morcego borboleta
 Morcego caseiro amarelo
 Morcego caseiro de Rendall
 Morcego caseiro do cabo
 Morcego caseiro Hottentot
 Morcego da Madeira
 Morcego das bananeiras
 Morcego das sepulturas das Mauricias
 Morcego das sepulturas do egipto
 Morcego de cabeça curta
 Morcego de Capaccini
 Morcego de cauda
 Morcego de cauda livre
 Morcego de cauda livre de Ansoorge
 Morcego de cauda livre de ventre branco
 Morcego de cauda livre do egipto

Scotophilus borbonicus (E. Geoffroy St.-Hilaire, 1803)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Glauconycteris variegata (Tomes, 1861)
Scotophilus dinganii (A. Smith, 1833)
Pseudoromicia rendalli (Thomas, 1889)
Laephotis capensis (A. Smith, 1829)
Eptesicus (Cnephaeus) hottentotus (A. Smith, 1833)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Afronycteris nana (Peters, 1852)
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818
Taphozous perforatus E. Geoffroy St.-Hilaire, 1818
Sauromys petrophilus (Roberts, 1917)
Myotis capaccinii (Bonaparte, 1837)
Coleura afra (Peters, 1852)
Mops (Mops) condylurus (A. Smith, 1833)
Mops (Chaerephon) ansorgei (Thomas, 1913)
Mops (Mops) niveiventer Cabrera and Ruxton, 1926
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)

Morcego de cauda livre malhado
 Morcego de Hanak
 Morcego de Hemprich
 Morcego de Kuhl
 Morcego de nariz enfolhado da cafraria
 Morcego de nariz enfolhado de commerson

Morcego de nariz enfolhado de commerson
 Morcego de railer
 Morcego de Savii
 Morcego de Schlieffens
 Morcego de schreiber
 Morcego dos Açores
 Morcego ferradura da Damarlandia
 Morcego ferradura das savanas
 Morcego ferradura de cabo
 Morcego ferradura de Darling
 Morcego ferradura de Dent
 Morcego ferradura de Hildebrandt
 Morcego ferradura de Lander
 Morcego ferradura de nariz de sela
 Morcego ferradura de Swinny
 Morcego ferradura gigante
 Morcego Frugivoro de Bocage
 Morcego frugivoro de Peters
 Morcego frugivoro de Wahlberg
 Morcego frugivoro do Egipto
 Morcego frugivoro gigante
 Morcego gigante amarelo das casas
 Morcego gigante de cauda livre
 Morcego grande de dedos compridos
 Morcego grande orelhudo
 Morcego lamoso Harrison
 Morcego lanoso da Damaralandia
 Morcego lanudo de Bocage
 Morcego lanudo de weltsch
 Morcego orelhudo do Egipto
 Morcego orelhudo piloso
 Morcego pequeno das casas
 Morcego pequeno de cauda livre
 Morcego pequeno de dedos compridos
 Morcego tridentado
 Morcego-anão
 Morcego-arboricola-gigante
 Morcego-arboricola-pequeno
 Morcego-das-bananas
 Morcego-de-bigodes
 Morcego-de-ferradura de Blasius
 Morcego-de-ferradura-grande
 Morcego-de-ferradura-mediterrânico
 Morcego-de-ferradura-mourisco
 Morcego-de-ferradura-pequeno
 Morcego-de-peluche
 Morcego-de-ventre-nú
 Morcego-frugivoro-egípcio
 Morcego-hortelão de Botta
 Morcego-hortelão-claro
 Morcego-lanudo
 Morcego-negro
 Morcego-orelhudo-canarino
 Morcego-pigmeu
 Morcego-rabudo
 Morcego-rato do Magrebe
 Morcegos

Mops (Chaerephon) bivittatus (Heuglin, 1861)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Otonycteris hemprichii Peters, 1859
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Hipposideros caffer (Sundevall, 1846)
Macronycteris commersoni (E. Geoffroy St.-Hilaire, 1813)
Macronycteris gigas (Wagner, 1845)
Mops (Xiphonycteris) thersites (Thomas, 1903)
Hypsugo savii (Bonaparte, 1837)
Nycticeinops schlieffenii (Peters, 1859)
Miniopterus schreibersii (Kuhl, 1817)
Nyctalus azoreum (Thomas, 1901)
Rhinolophus fumigatus Rüppell, 1842
Rhinolophus simulator K. Andersen, 1904
Rhinolophus capensis Lichtenstein, 1823
Rhinolophus darlingi K. Andersen, 1905
Rhinolophus denti Thomas, 1904
Rhinolophus hildebrandtii Peters, 1878
Rhinolophus landeri Martin, 1838
Rhinolophus blasii Peters, 1867
Rhinolophus swinnyi Gough, 1908
Rhinolophus clivosus Cretzschmar, 1828
Myonycteris angolensis (Bocage, 1898)
Epomophorus crypturus Peters, 1852
Epomophorus wahlbergi (Sundevall, 1846)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Eidolon helvum (Kerr, 1792)
Scotophilus nigrita (Schreber, 1774)
Tadarida ventralis (Heuglin, 1861)
Miniopterus inflatus Thomas, 1903
Nycteris grandis Peters, 1865
Kerivoula lanosa (A. Smith, 1847)
Kerivoula argentata Tomes, 1861
Myotis bocagii (Peters, 1870)
Myotis welwitschii (Gray, 1866)
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Nycteris hispida (Schreber, 1774)
Scotoecus albofuscus (Thomas, 1890)
Mops (Chaerephon) pumilus (Cretzschmar, 1826)
Miniopterus fraterculus Thomas and Schwann, 1906
Cloeotis percivali Thomas, 1901
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Afronycteris nana (Peters, 1852)
Myotis mystacinus (Kuhl, 1817)
Rhinolophus blasii Peters, 1867
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Rhinolophus hipposideros (Bechstein, 1800)
Miniopterus schreibersii (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Barbastella barbastellus (Schreber, 1774)
Plecotus teneriffae Barrett-Hamilton, 1907
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Tadarida teniotis (Rafinesque, 1814)
Myotis punicus Felten, 1977
 CHIROPTERA Blumenbach, 1779

Rhaeto-Romance

Barbastel
 Barbastel ureglia lada
 Chaun sgulant da l'Egipta
 Miniopter da l'ala lunga
 Miniopter ad ala lunga
 Moloss buldoc
 Nas fier-chaval grond
 Nas fier-chaval pitschen

Barbastella barbastellus (Schreber, 1774)
Barbastella barbastellus (Schreber, 1774)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Tadarida teniotis (Rafinesque, 1814)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)

Pipistrel da fossa a venter niv
 Pipistrel da Hanak
 Pipistrel da Kuhl
 Pipistrel da las alps
 Pipistrel da Madeira
 Pipistrel da Savi
 Pipistrel muschin
 Pipistrel nanin
 Pipistrel pigmeic
 Rinolof da Blasius
 Rinolof da Méhely
 Rinolof grond
 Rinolof mediterranean
 Rinolof pitschen
 Serotin da Botta
 Serotin isabellin
 Sgolanotg da las Azoras
 Sgolanotg gigant
 Sgolanotg pitschen
 Ureglia gronda da Teneriffa
 Ureglia lunga da Hemprich
 Vespertil cun tscheglias
 Vespertil cun tschegls
 Vespertil da Capaccini
 Vespertil dal barbis
 Vespertil pitschen dal barbis
 Vespertil ureglia-mieur punic

Taphozous nudiventris Cretzschmar, 1830
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus blasii Peters, 1867
Rhinolophus mehelyi Matschie, 1901
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
Rhinolophus hipposideros (Bechstein, 1800)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Nyctalus azoreum (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis capaccinii (Bonaparte, 1837)
Myotis mystacinus (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Myotis punicus Felten, 1977

Romanian

Lilacul cu coadă liberă europeană
 Liliac-cârn
 Liliac-cu-aripei-lungi
 Liliac-cu-gene-lungi
 Liliac-cu-mustăți
 Liliac-cu-picioare-lungi
 Liliac-mare-cupotcoavă
 Liliac-mic-cupotcoavă
 Liliac-românesc
 Liliac-sudic
 Liliacul cărămiziu
 Liliacul cârn
 Liliacul cu aripi late mediteranean
 Liliacul cu aripi lungi
 Liliacul cu burtă golașă
 Liliacul cu dungă albă
 Liliacul cu picioare lungi
 Liliacul cu potcoavă a lui Blasius
 Liliacul cu potcoavă a lui Mehely
 Liliacul de amurg din Azore
 Liliacul din Maghreb
 Liliacul fructivor egiptean
 Liliacul lui Botta
 Liliacul lui Savi
 Liliacul mare cu potcoavă
 Liliacul mare de amurg
 Liliacul mediteranean cu potcoavă
 Liliacul mic cu potcoavă
 Liliacul mic de amurg
 Liliacul mustăcio
 Liliacul pigmeu
 Liliacul pitic
 Liliacul pitic a lui Hanak
 Liliacul pitic al lui Kuhl
 Liliacul pitic din Madeira
 Liliacul urecheat de desert
 Liliacul urecheat din Tenerife
 Liliacul-lui-Blasius
 Liliaci cu pliuri nazale
 Liliaci cu tragus

Tadarida teniotis (Rafinesque, 1814)
Barbastella barbastellus (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis mystacinus (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Barbastella barbastellus (Schreber, 1774)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Miniopterus schreibersii (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Rhinolophus blasii Peters, 1867
Rhinolophus mehelyi Matschie, 1901
Nyctalus azoreum (Thomas, 1901)
Myotis punicus Felten, 1977
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Hypsugo savii (Bonaparte, 1837)
Rhinolophus ferrumequinum (Schreber, 1774)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus euryale Blasius, 1853
Rhinolophus hipposideros (Bechstein, 1800)
Nyctalus leisleri (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Otonycteris hemprichii Peters, 1859
Plecotus teneriffae Barrett-Hamilton, 1907
Rhinolophus blasii Peters, 1867
 RHINOLOPHIDAE Gray, 1825
 VESPERTILIONIDAE Gray, 1821

Russian

Азиатская широкоушка [= Aeiatskaya shirokouchka]
 Большой подковонос [= Bolshoj podkovonos]

Barbastella leucomelas (Cretzschmar, 1826)
Rhinolophus ferrumequinum (Schreber, 1774)

Бульдоговые
 Вечерница азорская
 Вечерница гигантская
 Вечерница малая
 Гигантская вечерница [= Gigantskaya vechernicha]
 Гладконосые
 Длиннокрыл обыкновенный
 Длиннокрыловые
 Кожан изабелловый
 Кожан пустынный
 Кожановидный нетопырь [= Kozhanovidny netopyr]
 Крылан египетский
 Крылановые
 Лжевампиры
 Листоносы
 Ложные подковоносы
 Малая вечерница [= Mapaya vechernicha]
 Малый нетопырь [= Malyj Neopyr]
 Мешкокрыл голобрюхий
 Нетопырь Ганака
 Нетопырь Куля
 Нетопырь кожановидный
 Нетопырь мадейский
 Нетопырь малый (пигмей)
 Нетопырь средиземноморский
 Нетопырь-карлик [= Netopyr karlik]
 Ночница средиземная (итальянская)
 Ночница трёхцветная
 Ночница усатая
 Ночница финикийская
 Обыкновенный длиннокрыл [= Obyknovennyj dlinnokryl]
 Подковогубы
 Подковонос Мегели (очковый)
 Подковонос большой
 Подковонос малый [= Podkovonos malyj]
 Подковонос средиземноморский
 Подковонос южный
 Подковоносые
 Свободнохвостые
 Складчатогуб широкоухий
 Средиземноморский нетопырь [= Sredizemnomorskij netopyr]
 Стрелоух белобрюхий
 Трёхцветная нуночница [= Tryokhtsvetnaya nochnitsa]
 Усатая ночница [= Usataya nochnitsa]
 Ушан канарский
 Футлярохвостые
 Широкоушка европейская
 Южный подковонос [= Yuzhnyj podkovonos]

MOLOSSIDAE Gervais, 1856
Nyctalus azoreum (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus lasiopterus (Schreber, 1780)
 VESPERTILIONIDAE Gray, 1821
Miniopterus schreibersii (Kuhl, 1817)
 MINIOPTERIDAE Dobson, 1875
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Hypsugo savii (Bonaparte, 1837)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
 PTEROPODIDAE Gray, 1821
 MEGADERMATIDAE H. Allen, 1864
 HIPPOSIDERIDAE Lydekker, 1891
 HIPPOSIDERIDAE Lydekker, 1891
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Taphozous nudiventris Cretzschmar, 1830
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myotis capaccinii (Bonaparte, 1837)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis mystacinus (Kuhl, 1817)
Myotis punicus Felten, 1977
Miniopterus schreibersii (Kuhl, 1817)
 HIPPOSIDERIDAE Lydekker, 1891
Rhinolophus mehelyi Matschie, 1901
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus blasii Peters, 1867
Rhinolophus euryale Blasius, 1853
 RHINOLOPHIDAE Gray, 1825
 MOLOSSIDAE Gervais, 1856
Tadarida teniotis (Rafinesque, 1814)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)

Otonycteris hemprichii Peters, 1859
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis mystacinus (Kuhl, 1817)
Plecotus teneriffae Barrett-Hamilton, 1907
 EMBALLONURIDAE Gervais, 1855
Barbastella barbastellus (Schreber, 1774)
Rhinolophus euryale Blasius, 1853

Scottish Gaelic

Crudh-ialtag bheag
 Crudh-ialtag Blasii
 Crudh-ialtag Meadhan-thireach
 Crudh-ialtag Mehelyi
 Crudh-ialtag mhòr
 Ialtag Asorach
 Ialtag bharbastail iarach
 Ialtag Botta
 Ialtag eàrr-shaor Eòrpach
 Ialtag fhad-chluasach Chanaireach
 Ialtag fhad-chluasach Hemprich
 Ialtag fhad-mheurach
 Ialtag fheasgarach
 Ialtag Geoffroy
 Ialtag ghibeach
 Ialtag Hanak
 Ialtag isibealach
 Ialtag Kuhl
 Ialtag Leisler
 Ialtag luch-chluasach Mhaigribeach
 Ialtag meas Èipheiteach
 Ialtag Mhaideurach
 Ialtag mheur-fhada Schreiber

Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus blasii Peters, 1867
Rhinolophus euryale Blasius, 1853
Rhinolophus mehelyi Matschie, 1901
Rhinolophus ferrumequinum (Schreber, 1774)
Nyctalus azoreum (Thomas, 1901)
Barbastella barbastellus (Schreber, 1774)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Tadarida teniotis (Rafinesque, 1814)
Plecotus teneriffae Barrett-Hamilton, 1907
Otonycteris hemprichii Peters, 1859
Myotis capaccinii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis mystacinus (Kuhl, 1817)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Nyctalus leisleri (Kuhl, 1817)
Myotis punicus Felten, 1977
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Miniopterus schreibersii (Kuhl, 1817)

laltag nochd-thònach thuaim
laltag Savi
laltag shoprànach
Mòr-laltag oidhcheach

Taphozous nudiventris Cretzschmar, 1830
Hypsugo savii (Bonaparte, 1837)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Nyctalus lasiopterus (Schreber, 1780)

Serbian

Обични пећински летипас [= Obični pećinski letipas]
Јужни потковичар [= Južni potkovičar]
Азорски ноћник [= Azorski noćnik]
Атлантски сл[и]јепи мишић [= Atlantski sl[ij]epi mišić]
Б[и]јелоруби слепи мишић [= Bj[el]orubi slepi mišić]
Блискоисточни поноћњак [= Bliskoistočni ponoćnjak]
Велики ноћник [= Veliki noćnik]
Велики потковичар [= Veliki potkovičar]
Голотрби гробнички м[и]јешокрилаш [= Golotربي grobnički m[ij]ješokrilaš]
Дугодлаки сл[и]јепи мишић [= Dugodlaki sl[ij]epi mišić]
Дугопрсти вечерњак [= Dugoprsti večernjak]
Европска широкоушка [= Evropejskaya širokouška]
Европски дугокрилаш
Европски широкоушан [= Evropski širokoušan]
Канарски сиви дугоушан
Либијски сл[и]јепи мишић [= Libijski sl[ij]epi mišić]
Магребски велики вечерњак [= Magrebski veliki večernjak]
Магребски поноћњак [= Magrebski ponoć njak]
Мали ноћник [= Mali noćnik]
Мали потковичар [= Mali potkovičar]
Обични сл[и]јепи мишић [= Obični sl[ij]epi mišić]
Патуљаст сл[и]јепи мишић [= Patuljasti sl[ij]epi mišić]
Пустински дугоушан [= Pustinjski dugoušan]
Риђи вечерњак [= Riđi večernjak]
Средоземни потковичар [= Sredozemni potkovičar]
Средоземни репаш [= Sredozemni repaš]
Тамнолики бркати вечерњак [= Tamnoliki brkati večernjak]
Тамнооки потковичар [= Tamnooki potkovičar]
Широкоухиј складчатогуб

Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Rhinolophus blasii Peters, 1867
Nyctalus azureum (Thomas, 1901)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)
Taphozous nudiventris Cretzschmar, 1830
Hypsugo savii (Bonaparte, 1837)
Myotis capaccinii (Bonaparte, 1837)
Barbastella barbastellus (Schreber, 1774)
Miniopterus schreibersii (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Plecotus teneriffae Barrett-Hamilton, 1907
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Myotis punicus Felten, 1977
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Otonycteris hemprichii Peters, 1859
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Rhinolophus euryale Blasius, 1853
Tadarida teniotis (Rafinesque, 1814)
Myotis mystacinus (Kuhl, 1817)
Rhinolophus mehelyi Matschie, 1901
Tadarida teniotis (Rafinesque, 1814)

Serbian Latin

Evropski dugokrilaš
Kanarski sivi dugoušan

Miniopterus schreibersii (Kuhl, 1817)
Plecotus teneriffae Barrett-Hamilton, 1907

Sinhalese

Schreibersge dik-angeli wawula

Miniopterus schreibersii (Kuhl, 1817)

SiSwati

Lilulwane
Lilulwane
Lilulwane

Cloeotis percivali Thomas, 1901
Miniopterus fraterculus Thomas and Schwann, 1906
Miniopterus natalensis (A. Smith, 1833)

Slovak

Hladkonos holobručný
Kaloň egyptský
Lietavec stahovavý
Netopier brvitý
Netopier dlhonohý
Netopier fúzatý
Netopier magrebský
Netopier stromový
Podkovár Blasiov
Podkovár južný
Podkovár malý
Podkovár Mehélyho
Podkovár veľký
Raniak azorský
Raniak malý
Raniak obrovský
Tadarida buldogovitá
Ucháč kanársky
Uchaňa čierna

Taphozous nudiventris Cretzschmar, 1830
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Miniopterus schreibersii (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis capaccinii (Bonaparte, 1837)
Myotis mystacinus (Kuhl, 1817)
Myotis punicus Felten, 1977
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus blasii Peters, 1867
Rhinolophus euryale Blasius, 1853
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus mehelyi Matschie, 1901
Rhinolophus ferrumequinum (Schreber, 1774)
Nyctalus azureum (Thomas, 1901)
Nyctalus leisleri (Kuhl, 1817)
Nyctalus lasiopterus (Schreber, 1780)
Tadarida teniotis (Rafinesque, 1814)
Plecotus teneriffae Barrett-Hamilton, 1907
Barbastella barbastellus (Schreber, 1774)

Večernica šípouchá
 Večernica Bottova
 večernica Hanákova
 Večernica hvízdavá
 Večernica južná
 Večernica Leachova
 Večernica madeirská
 Večernica malá
 Večernica Saviho
 večernica tripolitánska

Otonycteris hemprichii Peters, 1859
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Hypsugo savii (Bonaparte, 1837)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)

Slovenian

Širokouhi netopir
 Škorpjonjedi puščavež
 Azorski mračnik
 Belorobi netopir
 Blasijev podkovnjak
 Brkati netopir
 Dolgokrili netopir
 Dolgonogi netopir
 Dolgorepi netopir
 Drobni netopir
 Egipčanski leteči pes
 Golotrebušni mavzolejnik
 Gozdni mračnik
 Južni podkovnjak
 Kanarski uhatež
 Kraljičin poznik
 Libijski drobni netopir
 Liliacul-pitič mediteranean
 Madeirski mali netopir
 Mali netopir
 mali podkovnjak
 Mali podkovnjak
 Mehelyjev podkovnjak
 Punski netopir
 Savijev netopir
 Skupina brkati netopirjev
 Svetlotrebušni poznik
 Vejicati netopir
 Veliki mračnik
 Veliki podkovnjak

Barbastella barbastellus (Schreber, 1774)
Otonycteris hemprichii Peters, 1859
Nyctalus azoreum (Thomas, 1901)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Rhinolophus blasii Peters, 1867
Myotis mystacinus (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Tadarida teniotis (Rafinesque, 1814)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Taphozous nudiventris Cretzschmar, 1830
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus euryale Blasius, 1853
Plecotus teneriffae Barrett-Hamilton, 1907
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
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Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus mehelyi Matschie, 1901
Myotis punicus Felten, 1977
Hypsugo savii (Bonaparte, 1837)
Myotis mystacinus (Kuhl, 1817)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)

Somalian

Auo-rnèr
 Fid Mer
 Fidmair
 Fidmer
 Kibil
 Kibille
 Kibillei
 Sod silli (see Lanza *et al.*, 2015: 13)
 Sodan silli

CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779

Swahili

Popo
 Popo

CHIROPTERA Blumenbach, 1779
Taphozous mauritanus E. Geoffroy St.-Hilaire, 1818

Swedish

Alpfladdermus
 Azorfladdermus
 Balkanhästskonäsa
 Barbastell
 Blek sydflddermus
 Dvärgfladdermus
 Dvärghästskonäsa
 Dvärgpipistrell
 Europeisk vikvinge
 Flikörad fladdermus
 Isabellfladdermus

Hypsugo savii (Bonaparte, 1837)
Nyctalus azoreum (Thomas, 1901)
Rhinolophus blasii Peters, 1867
Barbastella barbastellus (Schreber, 1774)
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus hipposideros (Bechstein, 1800)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Miniopterus schreibersii (Kuhl, 1817)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)

Kanarielångöra
Kretapipistrell
Kuhls fladdermus
Kusthästskonäsa
Leislers fladdermus
Madeirapipistrell
Medelhavshästskonäsa
Medelhavsmusöra
Mindre brunfladdermus
Mustaschfladdermus
Nakengumpsfladdermus
Nilflyghund
Okenlångöra
Pälsvingad brunfladdermus
Parkpipistrell
Pipistrell
Storfotsfladdermus
Större hästskonäsa
Sydpipistrell
Veckläppad fladdermus

Plecotus teneriffae Barrett-Hamilton, 1907
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Rhinolophus mehelyi Matschie, 1901
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Rhinolophus euryale Blasius, 1853
Myotis punicus Felten, 1977
Nyctalus leisleri (Kuhl, 1817)
Myotis mystacinus (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Otonycteris hemprichii Peters, 1859
Nyctalus lasiopterus (Schreber, 1780)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Myotis capaccinii (Bonaparte, 1837)
Rhinolophus ferrumequinum (Schreber, 1774)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Tadarida teniotis (Rafinesque, 1814)

Tamil

எகிப்தி ய வாலற்ற வெளவால்
திறந்த ரம்படு கல்லறை வெளவால்
ஸ்ரைப ர்ஸ்-ன் நீண்ட விரலுள்ள வெளவால்
Sraipars-ṇ Nīṇṭa Viralulla Veļavāl
Ekiptiya Vāṇṇra Veļavā
Iṛanta Rampṭu Kallaṛai Veļāl

Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Taphozous nudiventris Cretzschmar, 1830
Miniopterus schreibersii (Kuhl, 1817)
Miniopterus schreibersii (Kuhl, 1817)
Tadarida aegyptiaca (E. Geoffroy St.-Hilaire, 1818)
Taphozous nudiventris Cretzschmar, 1830

Tanoboase (Ghana)

afrifraa
ahwenekron
ampane (singular)
ampane ankasa
ampane kronfo [= "thief or criminal bat"] (see Ohemeng *et al.*, 2017: 184)
frede frede
mmpane (plural) (see Ohemeng *et al.*, 2017: 184)
sreso ampane [= "original or normal bat"]

MOLOSSIDAE Gervais, 1856
Hypsignathus monstrosus H. Allen, 1862
CHIROPTERA Blumenbach, 1779
Eidolon helvum (Kerr, 1792)
Epomophorus gambianus (Ogilby, 1835)
MOLOSSIDAE Gervais, 1856
CHIROPTERA Blumenbach, 1779
Epomophorus gambianus (Ogilby, 1835)

Turkish

Akdeniz Cüce Yarasaı
Akdeniz Nalburunlu Yarasaı
Avrupa Serbest Kuyruklu Yarasaı
Azorlar Akşamcı Yarasaı
Bıyıklı Siyah Yarasa
Basıkburunlu Yarasa
Beyazseritli Yarasa
Blasius Nalburunlu Yarasa
Bottae'nin Genişkanatlı Yarasaı
Büyük Akşamcı Yarasa
Büyük Nalburunlu Yarasa
Çıplakkarınlı Mezar Yarasaı
Cüce Yarasa
Hanak'ın Cüce Yarasaı
İzabel Genişkanatlı Yarasaı
Kanaryalar Uzunkulaklı Yarasaı
Kırpıklı Yarasa
Küçük Ağaç Yarasaı
Küçük Nalburunlu Yarasa
Mısır Meyve Yarasaı
Maderia Cüce Yarasaı
Mahrep Farekulaklı Yarasaı
Mehely Nalburunlu Yarasaı
Savi'nin Cüce Yarasaı
Uzunayaklı Yarasa
Uzunkanatlı Yarasa
Uzunkulaklı Çöl Yarasaı

Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Rhinolophus euryale Blasius, 1853
Tadarida teniotis (Rafinesque, 1814)
Nyctalus azoreum (Thomas, 1901)
Myotis mystacinus (Kuhl, 1817)
Barbastella barbastellus (Schreber, 1774)
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Rhinolophus blasii Peters, 1867
Eptesicus (Cnephaeus) bottae (Peters, 1869)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus ferrumequinum (Schreber, 1774)
Taphozous nudiventris Cretzschmar, 1830
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Eptesicus (Cnephaeus) isabellinus (Temminck, 1840)
Plecotus teneriffae Barrett-Hamilton, 1907
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Nyctalus leisleri (Kuhl, 1817)
Rhinolophus hipposideros (Bechstein, 1800)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Myotis punicus Felten, 1977
Rhinolophus mehelyi Matschie, 1901
Hypsugo savii (Bonaparte, 1837)
Myotis capaccinii (Bonaparte, 1837)
Miniopterus schreibersii (Kuhl, 1817)
Otonycteris hemprichii Peters, 1859

Twi (Ghana)

apan (singular)
mmpan (plural)

CHIROPTERA Blumenbach, 1779
CHIROPTERA Blumenbach, 1779

Ukrainian

Вечірниця азорська
Вечірниця гігантська
Вечірниця мала
Вухань канарський
Довгокрил звичайний
Довгокрили [= Dovyhokryly]
Звичайні листоноси [= Zvychayni lystonosy]
ілики (Пергач) пустельний
Крилан єгипетський
Летючі лисиці
Лилик ізабеліновий
Лилики [= Lylyky]
Лиликові [= Lylykovy]
Листоніс Коммерсона [= Lystonis Kommersona]

Листоніс південноафриканський [= Lystonis
pivdennoafrykans'kyu]
Малий еполетовий крилан [= Malyy epoletovyy krylan]
Мишкокрил голобрюхий
Неороміція [= Neoromitsiya]
Нетопир Ганака
Нетопир білосмугий
Нетопир карлик
Нетопир мадейський
Нетопир малий [= Netopyr malyy]
Нетопир пігмей
Нетопир скельний (Саві)
Нічниця вусата
Нічниця довгопала
Нічниця магрибська
Нічниця триколірна
Нічниці [= Nichnytsi]
Підковик (Підковоніс) Блазіуса
Підковик (Підковоніс) Мегеля
Підковик (Підковоніс) великий [= Pidkovyk velyky]
Підковик (Підковоніс) південний
Підковик (Підковоніс) малий [= Pidkovyk malyy]
Підковикові [= Pidkovykovy]
Стріловух білочеревий
Тадарида європейська
Трилистоніс африканський [= Trylystonis afrykans'kyu]
Широковух європейський [= Shyrovukh zvychaynyy]

Nyctalus azoreum (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Nyctalus leisleri (Kuhl, 1817)
Plecotus teneriffae Barrett-Hamilton, 1907
Miniopterus schreibersii (Kuhl, 1817)
MINIOPTERIDAE Dobson, 1875
HIPPOSIDERIDAE Lydekker, 1891
Eptesicus (Cnephæus) bottae (Peters, 1869)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
PTEROPODIDAE Gray, 1821
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Vespertilioninae Gray, 1821
VESPERTILIONIDAE Gray, 1821
Macronycteris commersoni (E. Geoffroy St.-Hilaire, 1813)
Hipposideros caffer (Sundevall, 1846)

Epomophorus labiatus (Temminck, 1837)
Taphozous nudiventris Cretzschmar, 1830
Neoromicia Roberts, 1926
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)
Hypsugo savii (Bonaparte, 1837)
Myotis mystacinus (Kuhl, 1817)
Myotis capaccinii (Bonaparte, 1837)
Myotis punicus Felten, 1977
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotinae Tate, 1942
Rhinolophus blasii Peters, 1867
Rhinolophus mehelyi Matschie, 1901
Rhinolophus ferrumequinum (Schreber, 1774)
Rhinolophus euryale Blasius, 1853
Rhinolophus hipposideros (Bechstein, 1800)
RHINOLOPHIDAE Gray, 1825
Otonycteris hemprichii Peters, 1859
Tadarida teniotis (Rafinesque, 1814)
Triaenops afer Peters, 1877
Barbastella barbastellus (Schreber, 1774)

Vai (Liberia)

Tuŋa

Hypsignathus monstrosus H. Allen, 1862

Vietnamese

Họ dơi cánh
Họ dơi muỗi
Họ dơi quạ
Họ dơi thò đuôi
Họ dơi bao
Họ dơi lá
Họ dơi ma
Họ dơi mũi

MINIOPTERIDAE Dobson, 1875
VESPERTILIONIDAE Gray, 1821
PTEROPODIDAE Gray, 1821
MOLOSSIDAE Gervais, 1856
EMBALLONURIDAE Gervais, 1855
RHINOLOPHIDAE Gray, 1825
MEGADERMATIDAE H. Allen, 1864
HIPPOSIDERIDAE Lydekker, 1891

Welsh

Ystlum adain-gam Schreiber
Ystlum adain-lydan Botta
Ystlum adain-lydan llwydfelyn
Ystlum barfog
Ystlum beddrod tinfoel
Ystlum clustlydan Maghreb

Miniopterus schreibersii (Kuhl, 1817)
Eptesicus (Cnephæus) bottae (Peters, 1869)
Eptesicus (Cnephæus) isabellinus (Temminck, 1840)
Myotis mystacinus (Kuhl, 1817)
Taphozous nudiventris Cretzschmar, 1830
Myotis punicus Felten, 1977

Ystlum cynffonog
 Ystlum du
 Ystlum Geoffroy
 Ystlum hirfys
 Ystlum hirglust Hemprich
 Ystlum hirglust yr Ynysoedd Dedwydd
 Ystlum Leisler
 Ystlum lleiaf Hanak
 Ystlum lleiaf Kuhl
 Ystlum lleiaf Madeira
 Ystlum lleiaf Savi
 Ystlum mawr yr Asores
 Ystlum mwyaf
 Ystlum pedol Blasius
 Ystlum pedol lleiaf
 Ystlum pedol Mehely
 Ystlum pedol Môr y Canoldir
 Ystlum pedol mwyaf
 Ystlum-ffrwythau yr Aifft
 Ystlum-lleiaf cyffredin
 Ystlum-lleiaf soprano

Tadarida teniotis (Rafinesque, 1814)
Barbastella barbastellus (Schreber, 1774)
Myotis emarginatus (E. Geoffroy St.-Hilaire, 1806)
Myotis capaccinii (Bonaparte, 1837)
Otonycteris hemprichii Peters, 1859
Plecotus teneriffae Barrett-Hamilton, 1907
Nyctalus leisleri (Kuhl, 1817)
Pipistrellus (Pipistrellus) hanaki Hulva and Benda, 2004
Pipistrellus (Pipistrellus) kuhlii (Kuhl, 1817)
Pipistrellus (Pipistrellus) maderensis (Dobson, 1878)
Hypsugo savii (Bonaparte, 1837)
Nyctalus azoreum (Thomas, 1901)
Nyctalus lasiopterus (Schreber, 1780)
Rhinolophus blasii Peters, 1867
Rhinolophus hipposideros (Bechstein, 1800)
Rhinolophus mehelyi Matschie, 1901
Rhinolophus euryale Blasius, 1853
Rhinolophus ferrumequinum (Schreber, 1774)
Rousettus aegyptiacus (E. Geoffroy St.-Hilaire, 1810)
Pipistrellus (Pipistrellus) pipistrellus (Schreber, 1774)
Pipistrellus (Pipistrellus) pygmaeus (Leach, 1825)

Yao (Malawi)

Lichinji (applied to all large bats)
 Lichinji (applied to all largish bats)
 Liputiputi (applied to all small bats)
 Liputiputi (applied to all small bats)

Epomophorus minor Dobson, 1880
Rhinolophus hildebrandtii Peters, 1878
Nycteris thebaica E. Geoffroy St.-Hilaire, 1818
Afronycteris nana (Peters, 1852)

Appendix 3g: Original Descriptions

[*\[Epomophorus \(Nanonycteris\)\]*](#) Matschie, 1899

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Subgen. *Nanonycteris* Mtsch, subgen. nov,

Die Entfernung zwischen dem vorderen Augenwinkel und der Nasenspitze ist viel grösser als die Breite des Gesichtes an den Mundwinkeln. Auf dem Gaumen stehen parallele querfalten. Die Lippe ist nicht gefurcht. Der Schwanz ist noch kürzer als bei *Micropteropus*. Auf dem Plagiopatagium durchschneiden weniger als 32 Fascien dem vom Ellenbogen zur Spitze des fünften Fingers verlaufenden Strang. Die Reihe der Molaren ist nur so lang wie die Entfernung zwischen den Spitzen der oberen Caninen; der letzte obere Molar reicht nur bis zur Höhe des Vorderrandes des Foramen infraorbitale. Der knöcherne Gaumen ist wie bei *Epomops* gebildet. Der Unterarm ist kürzer als 60 mm.

Typus: *Ep. veldkampii* Jent.

Hab. Ober- und Mittel-Guinea.

[*\[Epomophorus macrocephalus\] var. angolensis*](#) Gray, 1870

p. 125

Pale brown; head with a dark streak from the nostrils to the front of the lower edge of the eye, and a second rather above and parallel with the upper lip; chest and beneath whitish; epaulette pale brown; forearm 3 1/4 inches. Female. B. M.

Hab. Angola, *Montiero* (in spirit).

[*\[Epomophorus macrocephalus\] var. unicolor*](#) Gray, 1870

p. 125

Brown above and below; epaulet white, rather small. B. M.

Hab. Banks of the Zambesi at Shupanga. Eating figs. *Kirk*.

[*\[Nyctinomus \(Nyctinomus\) pumilus\] Var. Major*](#) Trouessart, 1897

p. 146

[Nyctinomus (Nyctinomus) pumilus] Var. major, Dobs., l.c., p. 428 (vel nov.sp.).

Nilus super

[*\[Pipistrellus Kühli\] latastei*](#) Laurent, 1937

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Forme saharienne blanchâtre, non séparée de *P. Kühli* par LATASTE, qui le rapprochait de la sous espèce italienne *P. K. albolimbatus* KUSTER, laquelle est d'ailleurs absolument identique au type (Ca.) (2); serait la Chauve-Souris la plus commune au Sahara (LAVAUDEN), très commune dans la Tunisie méridionale et à Djerba (Bl.)... *Pipistrellus Kühli albolimbatus* LAVAUDEN non KUSTER.

Footnote (2).

Selon Ca., il serait nécessaire de donner à cette forme un autre nom, et je proposerais *Latastei* en l'honneur du savant naturaliste qui fut le premier à en observer les principaux caractères.

[*\[Rhinolophus hipposideros\] escalerae*](#) K. Andersen, 1918

Female. B.M. no. 10.11.24.1. Ha-ha, Mogador, Morocco. Collected by M. de la Escalera. Presented by Oldfield Thomas.

Characters based on the *Rh. hipposideros* subspecies key on p. 377

Infraorbital bridge nearly always much broadened.

p_3 nearly always absent. Size as *minimus*. Forearm of type 37 mm. Skull length to front of canine 15.3, condyle to canine 15.3, condyle to canine 13.6, $c-m^3$ 5.5. (Morocco).

[\[*Vespertilio auritus*\] *β aegyptius*](#) J.B. Fischer, 1829

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β. Aegyptius; minor; vellere magis rufo; vertebra caudali ultima a patagio anali liberiore. Geoffr. et Desmar. l.c.

[\[*Vespertilio pipistrellus* var.\] *β Aegyptius*](#) J.B. Fischer, 1829

p. 105

β. Aegyptius; cineraceus: pilis apice cinereis. Vespertilion pipistrelle d'Egypte Geoffr. et Desmar. l. c. (An species distincta?)

Longit. tota capitis corporisque 1 poll. 2 lin., capitis 6 lin., auriculārum $3\frac{1}{2}$ lin. (quarum latit. 4 lin.), caudae 11 lin. ; volatus amplitudo 6 poll. 5 lin. Dentes mol. $5-5 / 5-5$. In Europa (Gallia, Germania, Italia, Anglia, Dania); *β* in cryptis Thebatum Aegypti; in Buchara, teste Eversmann.

[\[*Vesperugo \(Vesperus\) serotinus*\] Var. *Gabonensis*](#) Trouessart, 1897

p. 108

[*Vesperugo (Vesperus) serotinus*] Var. *gabonensis*, Dobson, Catal., 1878. p. 192

Africa Occid., Gabon.

[\[*Xantharpyia \(Myonycteris\)*\]](#) Matschie, 1899

p. 63

Subgen. *Myonycteris* Mtsch, subgen. nov.

Der erste untere Molar ist kürzer als der zweite und der letzte untere Molar zusammengekommen. Der letzte obere Molar ist ungefähr $\frac{2}{3}$ so lang wie der vorletzte obere Molar. Der erste obere Praemolar ist kaum grösser als ein oberer Incisivus. Der erste untere Praemolar ist sehr klein, nur etwas grösser als ein unterer Schneidezahn. Auf dem

p. 64

Gaumen stehen hinter drei ungetheilten Querfalten vier getheilte Querfalten, und zwei gezähnelte Linien bilten den Abschluss des Gaumens nach hinten. Der Fuss mit den Krallen ist höchstens so lang wie das Ohr. Auf dem Plagiopatagium durchschneiden höchstens 16 Facien den vom Ellenbogen zur Spitze des fünften Fingers verlaufenden Strang.

Typus: *Cynonycteris torquata* Dobson.

Hab. Pungo Andongo, Cahata, Quibula, sämmtlich in Angola (Bocage). Angola (Wellwich nach Gray), Bismarckburg in Togo (B.M.); Liberia (Jentink); Bukoba am Victoria-Nyansa (B.M.).

[\[*Allomops*](#) J.A. Allen, 1917

p. 470

Text Figs. 12-15.

Type, *Chærephon (Allomops) osborni* sp. nov.

Supraoccipital border of skull enormously developed in old males, the lambdoid crest high and broad, expanded upward and backward into an oblique overhang, equal in height to about one half the interorbital breadth of the skull, or considerably exceeding the width of the foramen magnum, the upper border nearly straight, equal in length; to about one half the width of the braincase, with an extension downward from the lateral upper corner on either side to the mastoid process; less developed

but strongly indicated in females and young males; sagittal crest weakly developed in the smaller species of the group, very heavily so in the larger forms.

General form of the skull as in *Nyctinomus*, the braincase broad and flattened, preorbital furrow or depression conspicuous, but preorbital processes absent; premaxillæ usually fully ossified on front border, but sometimes emarginate.

Dental formula: $i \ 1\frac{1}{2-2}$, $c \ 1\frac{1}{1-1}$, $p \ 2\frac{2}{2-2}$ (or $1\frac{1}{2-2}$), p^2 being deciduous or wholly absent in some species), $m \ 3\frac{3}{3-3} = 30$ (or 28). Upper incisors small, about one third the length of the canines, parallel, widely separated from each other and the canines;

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lower incisors very, small, bilobed, the outer smaller than the middle ones, postero-external to them in position, often one or both outer lower incisors deciduous. Upper canines very long, slender and sharp-pointed, with a broad, shallow longitudinal groove on the front face, and a narrow postero-internal cingulum; lower canines correspondingly slender, with a broad postero-internal cingulum. First upper premolar (p^2) minute, in the toothrow (usually) or external to the base of the canine, sometimes deciduous; p^4 about half the size of m^1 , larger than m^3 . First lower premolar (p_2) larger than p_4 in males, smaller than p_4 in females, the sexual difference in the size of p_2 well marked in all the (four) known species. Upper molars with m^3 small, about one third the size of m^2 . Molar-premolar series in both jaws heavy, low and broad, with, very broad mandibular rami.

Males larger than the females, the sexual difference in size especially evident in the skulls, where it is emphasized by the much larger size of p_2 in the males than in the females relatively to the size of p_4 . Ears of medium size, thick and heavy, united at the base by a broad membrane, which extends forward as a conspicuous protuberance in front of the ears. Tail relatively short, about two fifths of the length of head and body, rather more than the apical half free. Sides of back naked, the naked space continuous with the naked hind limbs and naked hinder portion of the body, which is bare of fur on both surfaces posterior to a line joining the hips. From the region of the hips arises a fringe of scattered blackish bristles, directed backward and reaching to or beyond the middle of the uropatagium, the number of these bristly hairs varying, even in different individuals of the same species, from a few to a fairly conspicuous fringe. Pelage of the upperparts exceptionally short, soft and velvety; on the underparts longer and more dense. Color pattern distinctive, - very dark

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above and along the sides of the body below, without white at the base of the patagium. Wing membranes blackish, thick and leathery.

The present group, whether considered as of generic or as only of subgeneric value, is a natural group, much more sharply defined from all others than is *Chærephon* from *Nyctinomus*, although these two groups have been long currently given recognition as genera. In the present connection it is given subgeneric rank. The four known species are here described as new, although one of them; may be the same as *Nyctinomus thersites* of Thomas, which apparently belongs to the *Allomops* group. They are small to medium sized species, one of them, *Allomops nanulus*, being the smallest known African molossid.

In working out this group, in connection with other African molossid material, I have been impressed with the small value of the character most commonly considered as an important index to the relation of species referred to *Nyctinomus* and *Chærephon*, namely, the emargination or non-emargination of the front border of the premaxillæ. In one instance in the study of a series of 23 specimens collected the same day at the same locality, and evidently beyond question conspecific, I took up the skulls first, with skulls of other species, to determine their generic affinities. Later on collating the skulls (marked only with their catalogue numbers) with the skins to which they belonged, I found I had referred part to *Nyctinomus* and part to *Chærephon*! Further study of the same series showed that in this case at least emargination and non-emargination of the front border of the premaxillæ was largely dependent on the age of the specimen, the vacuity behind the incisors becoming gradually closed by ossification with the increased age of the specimen.

In one of the species here referred to *Allomops*, represented by six adult specimens, two, on the basis of the condition of the premaxillæ, are typically *Nyctinomus* and the other four typically *Chærephon*. They were otherwise indistinguishable, either on the basis of external and cranial characters or by the age of the specimen. In another species here referred to *Allomops*, represented by seven specimens, all of the skulls had the front palatal border emarginate, but as all of the other characters, both external and cranial, were typical of the *Allomops* group, it seemed proper to give preference to the combined weight of the *Allomops* characters, despite the *Nyctinomus*-like condition of the premaxillæ.

Alobus Peters, 1868

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Vespertilio (Alobus) Temminckii Rüppell

Von dieser kleinen Art habe ich kürzlich ein Weingeistexemplar erhalten und dabei gefunden, dafs sie sich von allen andern der Gattung *Vesperugo* (mit ^{5/6} Backzähnen) durch den gänzlichen Mangel eines Spornlappens, durch eine der von *V. dasyncneme* ähnliche Ohrklappe, sowie im Schädelbau durch die besondere Kürze und Abplattung des Schnauzentheils auszeichnet, weshalb ich sie als eine besondere Untergattung, *Alobus*, trennen zu müssen glaube, welche ein Mittelglied zwischen *Vesperugo* und *Vespertilio* bildet.

Asellia tridens diluta K. Andersen, 1918

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Like *A. tridens tridens*, but averaging larger, and colour of fur conspicuously paler.

Forearm 52.2 mm.

Skull: length to foot of canine 18.7; cond.-can. 16.6; $c-m^3$ 7; $c-m_3$ 7.7.

Hab. (of type). El Golea, Algerian Sahara. Other specimens from Biskra.

Type. Female. B.M. no. 12.11.14.2. Original number 42. Collected 16th May, 1912, by Dr. E. Hartert. Presented by Lord Rothschild.

Asellia tridens pallida Laurent, 1937

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Le genre *Asellia* Gray, n'était jusqu'ici connu en Afrique du Nord que par une seule sous-espèce locale, *A. tridens diluta* Andersen, différente par sa taille plus grande et sa coloration plus pâle du type de l'espèce *A. tridens* Geoffroy; cette forme, dont Lataste avait déjà observé les caractères distinctifs lorsqu'il en vit à Gafsa, le premier représentant barbaresque, n'avait pas, jusqu'à maintenant, été signalée au delà de Figuig (Foley) et nulle part ailleurs au Maroc.

Or, dans une collection de chauves-souris confiées au Muséum d'Histoire Naturelle par l'Institut scientifique Chérifien, figurait une *Asellia tridens* recueilli en 1930 dans la région de l'Oued Tatta (collecteur, le Chaouch Ahmed), soit dans l'Anti-Atlas à 400 kilomètres environ en ligne droite de Figuig. Mais cet individu diffère suffisamment tant par sa coloration que par quelques détails morphologiques de *A. tridens diluta* Andersen, pour qu'il ne soit pas possible de le rapporter à la sous-espèce du Sahara Algérien; j'ai donc cru pouvoir en faire une forme nouvelle, sans doute à étroite localisation géographique et je la décris ici sous le nom de: *Asellia tridens pallida* subsp. nov.

DESCRIPTION. - *Asellia tridens* de teinte extrêmement pâle, plus pâle encore et plus petite que *A. t. diluta*, et présentant de plus quelques particularités morphologiques.

Ornementation nasale très semblable à celle de *A. t. diluta*; toutefois la légère convexité sagittale que porte en sa partie médiane le fer à cheval apparaît au bord antérieur libre sous l'aspect d'une pointe plus saillante. La concavité semi-lunaire de

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la base du trident est par contre moins marquée, les denticules du trident sont moins élevées, en particulier le denticule médian. Le premier repli périsasal s'insère nettement plus en avant (à peu près 0.5 mm.) sur la lèvre supérieure, tandis que le repli qui le suit extérieurement s'insère au même niveau mais en arrière, au lieu de prendre attache un peu plus bas comme dans la forme algérienne. Enfin il existe extérieurement un 3^e repli pérional ou jugal, sous la forme d'une petite languette cutanée à bord libre frangé, d'une longueur de 1.2 à 1.5 mm. de direction oblique d'avant en arrière et de haut en bas, dont l'intérêt réside en ce que cette languette jugale, parfaitement visible chez l'individu marocain, est à peine distincte chez les sujets algériens, sans toutefois manquer complètement: ce qui explique pourquoi elle est omise par la description d'Andersen, que rapporte Cabrera (... herradura; con dos hojuelas accesorias a cada lado), "Mammiferos de Maruccos"... p. 89).

La lèvre supérieure glabre en sa partie moyenne, porte un petit bourrelet médian plus visible chez *A. t. diluta* que dans la forme nouvelle, mais présent chez l'une et l'autre.

Les oreilles de forme tout à fait semblable, sont striées longitudinalement d'une vingtaine (ou au moins d'une douzaine, bien

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visibles) de sillons transversaux parallèles, profondément marqués et très apparents chez *A. t. diluta*, au contraire à peine marqués et peu visibles chez *A. t. pallida*.

Le reste de la face ne présente aucun caractère particulièrement différent; le corps, les membres et la formule dentaire sont tout à fait comparables à ceux la forme algérienne. Les ailes, naturellement de même disposition, portent un gaufrage, superficiel à peu près identique.

L'insertion de la membrane interfémorale se fait sur le tiers supérieur de l'avant-dernière vertèbre; mais ce caractère n'a pas, comme je l'avais cru tout d'abord, de valeur différentielle, car l'examen de plus nombreux exemplaires algériens m'a montré qu'il était inconstant, de même que le rapport entre la partie libre de la queue et sa longueur totale. Dans l'exemplaire marocain, la longueur de la queue qui est libre en dehors de la membrane interfémorale, atteint, comme le type égyptien, le tiers de la longueur totale, et chez les individus algériens que j'ai examinés, cette même dimension varie de 30 à 42 p. 100,

Enfb. l'aspect extérieur de *A.t. pallida* est trop peu différent de celui du type de l'espèce pour l'examen complet du squelette entraînant la destruction de l'unique échantillon, soit indispensable. Il n'y a pas de raison pour que la forme nouvelle pût présenter un quelconque caractère osseux différentiel.

MENSURATIONS. - Voici les mesures somatiques du type de la description, avec dans la colonne en regard, celles d'un exemplaire de l'espèce: *A. t. diluta* du Sud-Algérien ⁽¹⁾ ce sujet étant le plus grand que j'aie jusqu'ici mesuré ⁽²⁾.

F, M

Tête et corps: 46, 49

Queue: 25, 25

Queue (sa partie libre): 8, 9

Oreille: 16.5, 16.5

Oreille (sa plus grande largeur): 10.5, 13.5

Footnotes:

^(a) Cet exemplaire, don du D^r FOLEY, de l'Institut Pasteur d'Alger, capturé le 14 septembre 1936, à Hassi Fohra, Beni-Abbès, Sud-Oranais Saharien et qui a figuré sous le numéro 900 dans ma collection personnelle, fait maintenant partie de celle du Muséum d'Histoire Naturelle à Paris.

⁽¹⁾ Cette dimension est certainement de toutes la plus sujette à caution: elle peut varier selon les observateurs et à divers stades de la conservation d'un même sujet; aussi je me demande quelle valeur peut-on attribuer pour la classification systématique à ce caractère, lorsqu'il ne s'agit que d'une variation de quelques millimètres, alors que toutes les autres dimensions concordent.

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F, M

Largeur du fer à cheval ⁽²⁾: 6.7, 7.6

- de la bouche ⁽³⁾: 7.1, 7.1

Avant-bras: 51, 51

III^e doigt métacarpien ⁽⁴⁾: 38, 37

- 1^{re} phalange: 12.5 - 13, 15 - 17

- 2^e phalange: 15.5 - 16.0, 17

IV^e doigt métacarpien: 36, 35 - 37

- 1^{re} phalange: 12, 11.5 - 12

- 2^e phalange: 8, 8

V^e doigt métacarpien: 28.5 - 29, 28.5 - 31

- 1^{re} phalange: 13.5 - 14.5, 13.5

- 2^e phalange: 9.5, 10

Tibia: 18.5 - 19, 20 - 20.5

Pied (sans les ongles): 8, 8.5

Poids à sec: 8 gr. 5, 9 gr. 55

COLORATION. - Animal beaucoup plus pâle que *A. t. diluta* et même presque complètement blanc, les parties nues étant elles-mêmes de teinte beaucoup plus claire. Le poil est partout d'un blanc par à sa base, aussi bien sur le ventre que sur le dos, mais la pointe en est lavée d'une nuance bistre très clair sur les côtés du thorax, dans les aisselles, sur les épaules, et en quelques points symétriques du dos, des lombes et de la nuque. La face supérieure du crâne, les joues, le cou, la gorge, les cuisses et toutes les parties inférieures sont entièrement blanches; ce blanc étant beaucoup plus pur et plus soutenu que chez *A. t. diluta*, où il est plutôt terne et un peu jaunâtre. Le pelage couvre les joues, mais ne cache pas les yeux dont les paupières sont nues, et laisse nue la face autour et au-dessous de l'ornementation nasale, ainsi que le menton.

La lèvre supérieure, nue au milieu, porte de chaque côté une brosse de poils argentés plus régulièrement croissante extérieurement et plus fournie que chez les sujets algériens. Le bord interne de la face antérieure du pavillon porte de même une rangée de poils plus longs et d'un blanc beaucoup plus vif; par contre, les longs poils tactiles de la face présentent exactement même longueur et même disposition. Les organes génitaux sont nus dans les deux formes, mais celle du Maroc ne présente ni organes érectiles saillants

Footnotes:

⁽²⁾ D'un bord à l'autre de la convexité, laissant en dehors les replis périnasaux.

⁽³⁾ Mesuré sous le menton, d'une commissure à l'autre.

⁽⁴⁾ Lorsque deux chiffres sont donnés, l'un plus grand que l'autre, c'est que le segment de membre de l'un des deux côtés du corps n'a pas la même dimension que son homologue de l'autre côté: ce cas est très fréquent.

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ni mamelles apparentes (une paire abdominale, et une paire pectorale) ce qui me fait penser qu'il s'agit d'une femelle adulte mais jeune.

Les oreilles sont de teinte paille terne, les membranes alaires et interfémorales, sépia clair et non brun noirâtre comme chez *A. t. diluta*.

DIAGNOSE. - Cette forme se distingue de suite du type de l'espèce par sa teinte presque blanche, qui peut la distinguer aussi de la sous-espèce algérienne déjà claire. Du point de vue coloration, cette dernière forme est intermédiaire entre la Marocaine et l'Egyptienne typique. Les petits détails différentiels qui ont été énumérés au cours de la description, caractérisent bien *A. t. pallida*, mais leur observation nécessite l'examen comparatif des deux formes côte à côte. Je n'ai pu conserver comme caractère différentiel indubitable l'insertion de la membrane interfémorale sur l'une ou l'autre des dernières vertèbres caudales, ce caractère, qui m'avait paru constant, n'ayant pas résisté à l'examen d'un nombre plus important d'exemplaires. Je crois donc que la coloration du poil, blanche à la base partout, est le principal caractère différentiel entre les deux formes, avec, naturellement la teinte d'ensemble beaucoup plus claire. On peut ensuite accorder quelque valeur au développement plus ou moins net du 3^e repli périnasal, à la bordure frangée ou non du 1^{er} repli, à l'importance de la striation de l'oreille, etc...

Jusqu'à un certain point, la sous-espèce marocaine pourrait se rapprocher de la toute petite *Asellia patrizii* de Beaux, car son avant-bras est plutôt plus grand que la dimension métacarpien + 1^{re} phalange du 3^e doigt, caractère qui sert à De Beaux pour isoler cette forme naine de Dancalie, de toutes les *Asellia tridens* Arabes et Égyptiennes, où cette dimension dépasse de 1 à 3 millimètres celle de l'avant-bras; ce qui est aussi le cas dans toutes les *A. t. diluta* que j'ai jusqu'ici examinées (le dépassement va même jusqu'à 5 ou 6 millimètres). Mais *A. patrizii* si éloignée de par sa localisation géographique, se différencie suffisamment par ailleurs pour qu'on ne puisse accorder à ce caractère qu'une simple valeur de convergence.

Il me semble raisonnable de penser que c'est de *A. t. diluta* que la nouvelle forme se rapproche le plus, mais je n'ai actuellement pas la possibilité d'établir de comparaison avec les autres sous-espèces de *A. tridens* ce qui ne manquerait pas d'un vif intérêt.

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DISTRIBUTION GEOGRAPHIQUE. - Le Maroc montagneux et méridional est suffisamment inexploré et mal connu du point de vue scientifique, pour qu'on puisse espérer retrouver cette forme ailleurs, que dans le Bled Tatta d'où elle provient; notons toutefois qu'*Asellia tridens diluta* parvient jusqu'à Figuig et Beni Abbès (d'où viennent tous les individus que j'ai examinés grâce à la complaisance du Dr Foley), à peu près situées sous la même longitude et que Heim de Balsac la signale en divers points du Sahara; elle est évidemment beaucoup plus répandue qu'*Asellia tridens pallida*, que je ne crois pas être autre chose qu'une intéressante forme évolutive locale.

[Attalepharca](#) Menu, 1987

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Groupe Collectif: *Attalepharca* nov.

Origine du nom: de "attaleph", nom hébreu des chauves-souris dans la Bible (selon Maisonneuve, 1878), et "arca", l'arche. Il laisse sous-entendre un statut transitoire, une instance, le temps d'une réflexion.

Il est pour le principal formé par des espèces myotodontes, jusqu'alors classées dans le genre *Pipistrellus* compte tenu de leurs formules dentaires identiques à celle des pipistrelles. Il comprend aussi une espèce, *flavescens* (SEABRA, 1910), classée parmi les *Eptesicus* pour avoir perdu la P2I, alors que ses structures dentaires indiquent une origine différente, tant de ce dernier genre que de *Nycterikaupius*.

Du point de vue odontologique, les espèces rassemblées dans le Groupe Collectif *Attalepharca* sont proches entre elles, ce qui n'exclut pas certaines particularités dont l'éventuelle importance, spécifique ou générique, reste à définir. La même remarque peut être faite au regard de l'anatomie générale. Dans ces conditions, et avant une révision exhaustive, la proposition d'un Groupe Collectif s'impose pratiquement. Considérer d'emblée ces espèces comme étant congénériques, créer pour elles un taxon commun, serait user d'un faux-semblant qui compliquerait inutilement la taxonomie dans un proche avenir.

La possibilité de les rassembler à titre temporaire en un Groupe Collectif est conforme aux possibilités précisées au Chapitre I, art. 1 (d) du Code International de Nomenclature Zoologique. Notre proposition semble recevable, puisque la présente situation correspond exactement à celle définie par le Code (Edition de 1985, p. 279): "Ensemble d'espèces nominales qui ne peuvent pas être placées avec certitude dans les genres connus".

Pour faciliter les vérifications, la liste commentée des taxons placés dans le Groupe Collectif comportera les immatriculations des spécimens de référence. Ici aussi, la liste est restreinte aux espèces qu'il nous a été donné de soumettre à l'examen; elle n'a donc pas un caractère limitatif.

- Eléments spécifiques du Groupe Collectif *Attalepharca*:

anchietae (SEABRA, 1900). L'espèce présente certains caractères conservateurs: métaconules bien développés sur M1/ et M2/; maintien d'un paralophe et d'un métalophe sur les mêmes dents; présence d'une très petite P2/ et d'un léger espace entre la canine et P4/.

Références: BMNH 69.1248, 10.2632, 10.2634 - Zambie.

bodenheimeri (HARRISON, 1960). Cette espèce conserve une P2/ résiduelle. On remarque une grande ressemblance de la denture avec celle de *Hypsugo*, constatation n'ayant d'ailleurs pas échappé au descripteur qui précisait: "Its dentition shows that it belongs to the *P. savii* group of which it appears to be a small and very distinct Arabian representative". Nous la plaçons dans le Groupe Collectif à titre de précaution, mais la possibilité d'un rattachement à *Hypsugo*, après examen complémentaire, ne doit pas être exclue.

Références: BMNH 67.1255 - Ile Socotra.

crassulus (THOMAS, 1915). Cette petite espèce, à crâne relativement large, possède une denture proche de celle de *Hypsugo*. L'examen d'un matériel plus important,

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portant aussi sur les parties molles de l'animal, est nécessaire avant d'envisager son transfert.

Références: BMNH 63.1174 - Congo.

flavescens (SEABRA, 1900). La denture est très proche de celles de *Hypsugo savii* et de *Pipistrellus mordax*. Le fait que le matériel muséologique est, a priori, restreint ne facilite pas les comparaisons souhaitables. Les II/ et C. sup. du seul spécimen examiné en font un néoéptésiforme de statut générique incertain, ce qui nous a amené à le placer en instance dans le Groupe Collectif.

Références: MNHN 1900.536 - Angola.

joffrei (THOMAS, 1915). Cette espèce bien particulière a été décrite comme appartenant au genre *Nyctalus*, puis rapportée ensuite par Tate (1942) au genre *Pipistrellus*. Sa nette myotodontie ne permet pas son maintien dans l'un ou l'autre des deux genres précités. Elle est remarquable par sa C. sup. porteuse d'une cuspide secondaire, sa P2/ résiduelle, ses M1/-M2/ à protofossa largement ouvertes vers l'arrière et sa M3/ très peu réduite. Le crâne est, entre autres particularités, caractérisé par une forte apophyse anté-orbitaire. Sous réserves, le statut générique adéquat pourrait être indépendant.

Références: BMNH 16.3.26.2 - Chin Hills, SW Mackenzie; BMNH 16.3.26.84 - Birmanie.

kitcheneri (THOMAS, 1915). La P2/ est petite, et le contact est établi entre la C. sup. et la P4/. Les molaires supérieures ne présentent pas de paralophe, sauf, parfois, à l'état résiduel, et pas de métalophe; l'importance du métaconule est très variable. La proximité de parenté de cette espèce doit être recherchée avec *macrotis* et *vordermanni*.

Références: BMNH 10.4.5.49, 10.4.5.50, 10.4.5.53, 10.4.5.154 - tous de Bornéo.

macrotis (TEMMINCK, 1835). Primitivement attribuée à *Pipistrellus*, c'est une espèce qui doit trouver place parmi les formes à crâne globuleux susceptibles de constituer une des futures subdivisions du Groupe Collectif. Au Muséum de Leiden, quatre individus, ayant préalablement fait l'objet d'une étude par Koopman, sont regroupés. Ils sont accompagnés de l'annotation suivante: "These four specimens (cotypes from Sumatra and one other from Bornéo) are *P. macrotis*, not *imbricatus*". Ce sont les spécimens m, n, o et p du Catalogue de Jentink. Compte tenu de son ancienneté, ce matériel est en mauvais état et se prête très mal à une recherche odontologique. Il est difficile de distinguer, sur la seule base de la morphologie crânienne et de la denture, une différence marquée entre *macrotis* et *vordermanni* (souvent appelé *imbricatus*). Il semble toutefois que M/ soit un peu plus large sur le premier, et que la même dent tende à devenir subcarrée chez le second (voir ci-après *vordermanni*). Les *macrotis* conservent une P2/ bien cingulée qui ne s'oppose pas au contact entre les canine et P4/. Sur la rangée inférieure la P4 est petite, et légèrement déplacée vers l'intérieur.

Références: RNH spécimens m, n, o, p du Catalogue de Jentink - Sumatra et Bornéo ; BMNH 23.1.2.12 - Sumatra.

mordax (PETERS, 1866). Sur le plan dentaire, *Pipistrellus mordax* apparaît proche des *Hypsugo*. La P2/, bien cingulée, est relativement forte, son importance est du même ordre que celle observée chez *Hypsugo pulveratus*. Nous avons examiné le spécimen n° 156 de la collection étudiée par Sody (1937), provenant de Tjiboeni, Java. Ses mensurations de l'avant-bras et du crâne correspondent à celles données par Peters pour le type.

Références: RNH n° 156 Coll. Sody - Java; MNHN 1876.356 - Philippines; BMNH 79.11.21.203 - Kumaon, et 16.3.25.5 - Darjeeling. Les deux derniers présentent une morphologie dentaire conforme à celle de *mordax*, cependant une plus petite taille et des provenances géographiques éloignées de celles du type

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rendent souhaitable une vérification de leur identité spécifique.

musculus (THOMAS, 1913). La présence d'une petite P2/ a contribué au maintien de cette espèce dans le genre *Pipistrellus*. Il arrive cependant que cette dent manque. La structure de II/ et une nette myotodontie nous la font placer dans le Groupe Collectif. Sa morphologie dentaire reste très proche de celles des pipistrelles.

Références: MHNG 914.42, 914.43, 914.45 - NE Zaïre ; CB 008 - Gabon.

nilsoni (KEYSERLING et BLASIUS, 1839). La position générique de cette espèce reste à définir. Nous l'avons placée provisoirement dans le Groupe Collectif pour attirer l'attention sur ce cas particulier. La morphologie de II/ l'éloigne autant des *Eptesicus* que des *Nycterikaupius*; son crâne plat la sépare des espèces à crâne globuleux (*macrotis*, *vordermanni*); elle est autre que les *Hypsugo* par l'absence permanente de P2/, les structures de la canine supérieure, et le développement du flexus labial de P4/. Nous ne serions pas étonnés que des investigations multiformes mènent à une indépendance générique. Dans ce cas, le nom du taxon qui deviendrait prioritaire serait *Amblyotus* KOLENATI, 1858.

Références: BMNH 25.11.3.4 - Norvège; MHNG 890.42, 975.83, 1492.65, 1492.93 - Suisse, et 986.4 - Autriche.

vordermanni (JENTINK, 1890). La grande proximité entre cette espèce et *macrotis* semble être définitivement établie.

Comme le dernier, *vordermanni* devra prendre rang parmi les formes à crâne globuleux dont nous avons fait état précédemment. Un examen comparatif des deux espèces montre que la P4/ de *macrotis* présente un degré de réduction plus accusé que celle de *vordermanni*. Par ailleurs, les molaires supérieures de *vordermanni* possèdent un métalophe qui ferme la protofossa vers l'arrière, ce qui n'est pas le cas pour *macrotis*.

Références: RNH spécimen a O (type) - Billiton, juin 1888; MNHN 1959.221 - Bornéo; BMNH 10.4.6.29, 9.1.5.286, 9.1.5.312, 9.1.5.313, 9.1.5.315 - tous de Java.

C[oleura] g[allarum] nilosa Thomas, 1915

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Essential characters of the Somali *C. gallarum*, but the colour, instead of being more or less uniformly pale brown, is a darker brown - sepia - on the surface, the bases of the hairs being strongly contrasted white; the long hairs of the name are white for about 3.5 mm., the terminal 2 mm. being sepia. Below, the general colour is rather greyer, but the bases of the hairs are similarly contrasted whitish, the contrast showing well in the long hairs of the flanks, which are almost uniformly coloured in true *gallarum*.

Dimension (see table on p. 579).

Hab. Upper Nile. Type-series from near the mouth of the Bahr-el-Zeraf.

Type. Adult male. B.M. no. 15.3.6.76. Original number 77. Collected 3rd February, 1914, by Willoughby P. Lowe. Twelve specimens.

C[oleura] gallarum Thomas, 1915

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Size decidedly smaller than in *C. afra*, the forearm usually 47 - 48 mm. in length, the skull markedly smaller in all dimensions. General colour above pale brown - rather darker than "wood-brown," - the hairs slightly lighter towards their bases, but not conspicuously or abruptly bicolor. Under surface similar in general tone, but the bases of the hairs not lighter than the tips.

Skull as in *C. afra*, except for its smaller size and smaller teeth. Position and development of the small premolar variable.

Dimensions (see table on p. 579).

Hab. Suakin, Somaliland, and Aden. Type from Zeyla, Somaliland. Sea-level.

Type. Old female, B.M. no. 11.8.2.4. Collected 29th October, 1910, and presented by Dr. R. E. Drake-Brockman. Four skins and six skulls from the type-locality.

Other specimens from Suakin (*Anderson, Holled Smith*), Aden (*Yerbury*), Sheikh, Somaliland (*Drake-Brockman*), Lugh, Somali (*Bottego*).

C[oleura] silhouettæ Thomas, 1915

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Like *C. seychellensis*, but smaller.

General proportions as in *C. seychellensis*, the wings similarly longer than in the mainland species, the forearm about $3\frac{3}{4}$ times as long as the skull measured from the condyle to the front of the canines. Fur rather shorter than in *C. gallarum* and extending less upon the membranes; hairs of back about 3, of nape 4.5 mm. in length. Colour dark brown, rather darker than "bistre," the bases of the hairs scarcely lighter; under surface similar in general tone, but the tips of the hairs are slightly lighter than the bases.

Skull like that of *C. seychellensis*, but smaller throughout. A mesial septal ridge generally present in the basial pit.

Dimensions (see table on p. 579).

Hab. Silhouette Island, Seychelles.

Type. Adult male (skinned from spirit). B.M. no. 6.3.18.2. Collected and presented by Prof. J. Stanley Gardiner. Six specimens.

The Museum contains two authentic specimens, one of them a co-type, of Peters's *C. seychellensis*, besides a third from "Zanzibar." These differ so uniformly from the six examples obtained by Prof. Gardiner on Silhouette Island that it is evident that the Seychelles contain different forms of the genus, no doubt inhabiting different islands. While it is not definitely recorded on which island Dr. Percival Wright obtained the typical examples of *C. seychellensis*, for he mentions in connection with the species Mahé, Praslin, and Silhouette, it is presumably from the first of these islands that he got the specimens, as he says that "it was very common in the neighbourhood of the town of Port Victoria, Mahé," and it was there that he stayed for most of his time in the Seychelles, merely making casual visits to Praslin and Silhouette.

The example from "Zanzibar" referred to *C. seychellensis* by Dobson appears to me to be rightly so named, but I should think it possible that its asserted locality is erroneous. It was presented by a captain in the Navy, whose station would have included the Seychelles, where the bat may have either been caught or come on board, to be afterwards captured at Zanzibar.

As illustrating the greater length of the wing in the Seychelles species, it will be seen from the table of dimensions that *C. silhouettæ* has a smaller skull than *C. gallarum*, but yet has an absolutely longer forearm.

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[table removed - eds.]

Casinycteris Thomas, 1910

p. 111

External characters, including even coloration, precisely as in *Scotonycteris*. The ears larger than in *S. zenkeri*. Wings reticulated, said to be orange-coloured in the fresh state.

Skull at once distinguishable from that of *Scotonycteris* by its palate being markedly shortened posteriorly and the bony floor to the long mesopterygoid fossa completely obsolete, so that the palation is practically at the level of the back of the single molar. In *Scotonycteris* there is the usual elongated palate. Palatal edge forming a reversed W (M), the median angle in continuation with a long and high vomerine ridge. Rostrum shorter in proportion than in *Scotonycteris*, bony palate more arched mesially, zygomatics more abruptly expanded, postorbital processes and general cranial ridges more strongly developed. Rami of lower jaw thickened and expanded.

Dental formula as in *Scotonycteris*.

Teeth in a general way high, pointed, shorter in section antero-posteriorly, broader transversely than in *Scotonycteris*. Canines very long, curved, flattened. Cheek-teeth, both upper and lower, with the inner cusp well developed and separate, instead of being suppressed or joined to the outer, as in *Scotonycteris*; this is especially noticeable in p_3 , the inner cusp being distinct, two-thirds the height of the outer, while in *Scotonycteris* the outer and inner enamel-ridges of the tooth are smoothly continuous with each other.

Type. *Casinycteris argynnis*, sp. n.

This striking bat, which adds another to the long list of new genera discovered by Mr. Bates, is remarkable for possessing a palate quite unlike that of other fruit-eating bats, and more recalling that found in some of the Microchiroptera. The astonishing resemblance of the type species to *Scotonycteris zenkeri* is also noticeable. Probably both bats bear a protective resemblance to the leaves, fresh or dry, of some local tree.

Casinycteris argynnis Thomas, 1910

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General external appearance - at least in the dried state - precisely as in *Scotonycteris zenkeri*, except that the size is

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materially larger, and the ears are also larger. Fur soft and fine, the general mass of the hairs about 8 mm. in length on the back, but with other longer hairs, attaining 13 - 14 mm., intermixed with them. Colour above about as in *S. zenkeri*, the hairs brown at base, then greyish white, with pale russet-brown tips. Head with the brown and white markings more strongly defined than in that animal, the ground-colour a little darker, the white patches prominent, present between the eyes, at the posterior angle of the eyes, at the anterior bases of the ears, and along the upper lips. In all other respects, in colour, degree of hairiness, and the marked reticulation of the wings, *C. argynnis* is so like *S. zenkeri* as not to need detailed description.

At least this is the case so far as dried specimens are concerned, but Mr. Bates states that in the fresh state the wings, ears, eyelids, and muzzle were "bright orange colour," and some traces of yellow are still present on such parts of the type as have been protected from light. Possibly the same may be the case with *Scotonycteris*, but we have no evidence on the subject.

Dimensions of the type (the body and ear measurements taken in the flesh): -

Forearm 60 mm.

Head and body 95; ear 20; index finger (c. u.) 44.5; third finger, metacarpus 41, first phalanx 28, second phalanx 36; lower leg and foot (c. u.) 40.

Skull: greatest length 28.3; basal length 25; zygomatic breadth 20; interorbital breadth 5.2; tip to tip of postorbital processes 11.7; brain-case, breadth 13.6; palatal length 10.6; post-palatal length 14.4; front of canine to back of m^1 9.2.

Hab. Bitye, Ja River, S.E. Kameruns. Alt. 2000'.

Type. Adult female. Original number 502. Collected 19th November, 1909, by Mr. G. L. Bates.

Cephalotes teniotis Rafinesque, 1814

p. 12

Cephalotes teniotis. Entièrement gris-brun, oreilles plus longues de la tête, sans oreillons conchiformes et striées intérieurement en travers, une verrue entre les deux incisives. - Obs. Cette nouvelle espèce remarquable se trouve en Sicile: le *Vespertilio cephalotes* de Pallas ou *Pteropus cephalotes* des modernes appartient aussi à ce genre et lui donne son nom, je le nommerai *Cephalotes pallasii*.

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CORRECTION.

Page 12. - Je viens de m'apercevoir que Mr. Geoffroy St. Hilaire a établi en 1810, le *G. Cephalotes* avec le *Pteropus palasii* et une autre espèce *C. peronii*; mais nos caractères diffèrent en ce que dans mon *C. teniotis* il n'y a aucune incisive inférieure: s'il compose un G. particulier, il faudra le nommer TADARIDA TENIOTIS.

[the above (p. 55) is also the description of the genus *Tadarida*. This publication is conceived as a letter to Mr. Ch. H. Persoon M.D., Célèbre auteur des *Sinopsis plantarum et fungorum* in Paris, and was dated 3 June 1814.] - Victor Van Cakenberghe, December 2009.

Chærephon (Lophomops) J.A. Allen, 1917

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Text Figs. 9-11.

Type *Chærephon (Lophomops) chapini* sp. nov.

Skull and dentition as in the smaller species of *Chærephon*, but outer lower incisors often deciduous, giving an incisive formula of 1-1/1-1 instead of 1-1/2-2.

Ears united in front by a deep membrane, from the back of which arises a heavy crest of long straight hairs, occupying the whole posterior face of the membrane and rising above the tops of the ears as a broad frontal transverse crest, the hairs of which are dark brown for the basal half and lighter brown or (in one of the species, selected the type) white for the apical half. It is apparently a sexual character, but only males have thus far been examined except in the rather aberrant *C. (L.) abæ*. For

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decorative effect, especially when the crest hairs are parti-colored - rufous at base and pure white for the apical half, as in *C. (L.) chapini* - it vies with the spectacular head ornamentation occurring in many groups of birds.

The taxonomic value of *Lophomops* may be open to question. The peculiar character of the crest is not only its great size and striking effect, but the fact of its origin being from the whole posterior face of the deep membrane connecting the ears, instead of from the front of the head behind the ears. A number of species of both *Chærephon* and *Nyctinomus* have slightly lengthened tufts of glandular hairs behind the ears arising from the front part of the crown (not from the connecting membrane of the ears), consisting usually of lengthened crinkled hairs, a little longer and coarser than the adjoining fur, and usually of a darker color. These patches of specialized hairs frequently appear to be restricted to males, but in some species are present in both sexes, as is the case with the gular sac. They are, however, inconspicuous in comparison with the crests of the *Lophomops* group. Three species of this group are here designated as the type.

Dysopes limbatus Peters (now regarded as a *Chærephon*) is described by de Winton as having a "long crest of erect hairs behind the connecting membrane of the ears in the males," but he does not say that they arise from its posterior face. Peters does not describe nor does his figure represent any such crest as is seen in *Lophomops*. He simply says: "Die Haare auf dem Kopfe hinter der Vereinigung der Ohren sind länger und steifer als am übrigen Körper." "The type locality of *Dysopes limbatus* is Mozambique Island, and it is possible that the specimens de Winton had in hand were not the true *limbatus* of Peters, and are perhaps referable to some form of *Lophomops*, *C. limbatus* (Peters) is probably a near relative of *C. hindei* (Thomas).

Chærephon (Lophomops) abæ J.A. Allen, 1917

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Text Fig. 11.

Type, No. 48887, F ad.; skin and skull. Aba, northeastern Belgian Congo, Dec. 17, 1911; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 1823.

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Above hair-brown, with a slight grayish sheen; below a narrow median whitish band from breast to anal region, and a narrow white border at base of wing membranes from humerus to femur; rest of underparts like the back but without the grayish sheen; membranes lightish brown above and below. Male with a conspicuous cinnamon-brown, crest, lacking in the female.

Ears thin, dark brown, rising separately from the crown, the intervening space being occupied by an erectile membrane, convex above and about 5 mm. high, from the back of which arises, in the male, a thick mass of brown hairs, about 7 mm, in length and reaching 3 mm. above the crest of the convex membrane (Fig. 11); the basal three fifths of the hairs a darker shade of cinnamon-brown than the apical two fifths; in females (at least in dry skins), the membrane is directed backward, covering the crown) which is naked or covered only with a slight pubescence on the upper border and in front; crown beneath bare (as seen by softening and raising the disk or lappet). The size of the lappet varies in different specimens, being not fully developed in the younger examples.

Skull with the braincase broad and flat, rising only slightly above the rostral portion of the skull. Sagittal crest weakly developed or barely indicated; lambdoid crest moderate, preorbital processes strongly developed; premaxillæ well ossified in old skulls, but in young adults there is often a small U-shaped vacuity behind the incisors.

Collectors' measurements of type: Total length, 103 mm.; head and body, 69; tail, 34; foot, 16; ear, 17. Forearm (from skin), 43.

Collectors' measurements of type and 22 topotypes: Total length, 102 (97-107); head and body, 67.3 (62-73); tail, 33 (29-35); foot, 10.6 (10-11); ear, 18 (17-19).

Forearm, from skin (same specimens), 42 (40-44).

Skull (type), total length, 18.5; condylobasal length, 17.3; zygomatic breadth, 11.2; mastoid breadth, 10.7; interorbital breadth, 3.8; maxillar breadth, 8.1;

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breadth at base of canines, 5.4; breadth of braincase, 9; depth of braincase at pterygoids, 6.1; length of upper tooththrow (c-m³), 6.7; length of mandible, 12.3; angle to condyle, 3.5; depth at coronoid, 8.4; length of lower tooththrow, 7.6.

Skull, average of 10 specimens, total length, 18 (17.5-18.8); zygomatic breadth, 11.2 (11-11.4); breadth of braincase, 9.2 (8.7-9.8).

Represented here by a series of 25 specimens, all collected the same day at Aba. Four additional specimens from Faradje, collected Feb. 25, 1911, are so similar in every respect to the Aba series that they seem preferably referable to the same species. The external measurements are slightly less, but the skulls indicate greater maturity and are as large as those of the Aba series. The coloration, however, is less dark and more buffy. They probably represent a slightly differential local race of the Aba form.

Chærephon (Lophomops) abæ resembles, in size and coloration, *C. major* (Trouessart) and *C. emini* (de Winton) in size, but not in color. As neither of these species, so far as known, is crested in the males, nor possess the conspicuous and (probably) erectile lappet in the females, and ununited ears, they require no special consideration in the present connection. The uniform brown coloration of the membranes, and usually the size, will readily separate *abæ* from the other known forms of the *Lophomops* group.

Chærephon (Lophomops) chapini J.A. Allen, 1917

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Text Fig. 9

Type, No. 48841, M ad. (skin and skull; topotype, No. 49209, M ad., alcoholic), Faradje, northeastern Belgian Congo, Nov. 11, 1912 (topotype, Jan. 12, 1913); Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 1971.

Head in males conspicuously crested, the crest arising from the posterior face of the deep membrane connecting the ears; breadth of the crest at base, 8.5 mm.; length of crest, 13 mm. in type, 14 in topotype; basal half reddish chestnut, apical

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half white; breadth of crest across the top when, expanded (Figs. 9, A, B, C) about 20 mm. A broad tuft of soft whitish hairs covers the front of the crown behind the crest. Body above pale cinnamon-brown, the fur being whitish basally; below grayish brown, the middle of the belly whitish; a narrow band of white at proximal edge of wing membranes, between humerus and femur, continuous with a similar band at base of uropatagi; wing membranes white, becoming yellowish proximally; the portion adjoining the body, between humerus and femur, thickly punctated with minute blackish specks; interfemoral membrane dark brown above and below; limb bones dark brown above in strong contrast with the light colored membranes, below light brown like the membranes; tail blackish. No gular pouch.

Collectors' measurements of type: Total length, 84 mm.; head and body, 56; tail, 28; foot, 7; ear, 14.

Additional measurements from the dry skin: Forearm, 34; third metacarpal, 37; ear from crown, 8.5. Tragus minute, quadrate, about 1.5 square; antitragus small, about 5 broad at base and 3 high.

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Measurements of the alcoholic topotype: Total length, 80; head and body, 48; tail, 32; forearm, 37; third metacarpal, 30; third digit, 72; tibia, 11.8; foot, 7.7; ear from crown, 9.5, from notch behind antitragus, 12.8; expanse of both ears from tip to tip, 27.

Skull (measurements of topotype in parenthesis), total length, 15.5 (16); zygomatic breadth, - (9.5); interorbital breadth, 3.5 (3.5); mastoid breadth, - (9); breadth of braincase, 7.0 (7.7); maxillar breadth, 6.5 (6.8); upper tooththrow (c-m³), 5.4 (5.8); length of mandible, 10 (10.4); angle to condyle, 2.0 (2.0); depth at coronoid, 2.0 (2.0); lower tooththrow, 5.9 (6.4). Upper incisors parallel, slightly separated, a wide space between them and the canines; lower incisors, bifid, 1-1 (in both type and topotype), completely filling the space between the canines. First upper premolar minute, in the tooththrow, separating the canine and p⁴; first lower premolar smaller and much lower crown than the second. Premaxillæ fully ossified; palatal foramina coalesced,

forming a small U-shaped vacuity; presphenoid pits deep, slightly longer than wide; preorbital (lacrymal) processes strongly developed; median crest slightly indicated, lambdoid crest moderately strong. In general features the skull is as in other small species of *Chærephon*.

Chærephon (Lophomops) chapini is represented by only the type and a topotype, both old males from Faradje. It evidently has no close relationship to any known species. Its conspicuously heavy and long parti-colored crest distinguishes it at once from the other known species of the *Lophomops* group.

Chærephon (Lophomops) cristatus J.A. Allen, 1917

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Text Fig. 10, A, B.

Type, No 48844, M ad. (skin and skull), Boma, near mouth of Congo River, Belgian Congo, Jan. 26, 1915; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 2628.

Head crested, the crest arising from the posterior face of the deep membrane uniting the ears. Crest (Fig. 10, A, B) shorter than in *C. chapini*, the hairs reddish apically, the basal half darker, the transition between the two tones abrupt. Whole top of the head behind the crest naked, but covered by the crest hairs when the crest is laid back. Body above uniform russet, the basal portion of the fur but little lighter than the tips; underparts dark cinnamon-drab, the middle area with light tips to the hairs, in contrast with the sides; an indistinct median whitish band from the pectoral region, posteriorly; a narrow band of soft white fur along the inner edge of the wing membrane from the humerus to the femur and continued across the lower abdomen at base of the uropatugium. Face and gular region naked and blackish, bordered below by a naked flesh-colored band. Wing membranes as in *C. chapini* but whiter, the upper surface in strong contrast with the blackish limb bones; the dorsal side of the membrane along the sides of the body, between humerus and femur, thickly set with blackish dots, most of which support a minute bristly hair, visible only under a strong lens, and more obvious than in the specimens of *C. chapini*. Interfemoral membrane dark slaty brown above, somewhat lighter below. No gular pouch. Upper lips corrugated.

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Collectors' measurements of type: Total length, 87 mm.; head and body, 56; tail, 31; foot, 9; ear, 13.

Additional measurements from the dry skin: Forearm, 36; third metacarpal, 35; tibial, 10; foot; 8.7; thumb, 6; ear from crown, 9.2; length of crest hairs, 9.5.

Three male topotypes (Nos. 49216, 49237, 49359) in alcohol: Total length, 84, 83, 80; head and body, 51.5, 53, 48; tail, 32, 31, 32; forearm, 36, 36.2, 36.4; third metacarpal, 30, 34, 35; third digit, 66, 65, 66.5; ear from crown, 9, 8, 8; height from notch behind antitragus, 14, 13, 13.5; extent of both ears from tip to tip, 28, 29, 29; tibia, 11, 11.5, 11; foot, 7, 6.7, 8; length of crest, 9, 8, 9; width at base, about 6.5.

Skull, type (topotype No. 49216, in parenthesis), total length, 16.3 (16); zygomatic breadth, 9.7 (9.8); mastoid breadth, 9 (9.2); interorbital breadth, 3.6 (3.8); maxillar breadth, 7 (7.2); breadth of braincase, 8 (8); depth of skull at pterygoids, 6 (6.3); upper toothrow (c-m³), 5.5 (5.7); length of mandible, 10 (10.3); angle to condyle, 3 (3.2); depth at coronoid, 2.8 (2.8); lower toothrow, 6 (6.2).

Chærephon (Lophomops) cristatus is represented by the type and three topotypes, all collected at Boma, January 25-29 and June 15, 1915. The type lacks the middle pair and one of the outer lower incisors, which had been shed in life; one of the topotypes has only the middle pair, which occupies the whole space between the canines; the other has the middle pair and a minute outer incisor on the left side, showing that in the *Lophomops* group the lower incisors are doubtless normally 2-2. This species is slightly larger than *C. chapini* and differs from it strikingly in coloration, aside from the difference in the color and length of the crest.

Chærephon (Lophomops) shortridgei Thomas, 1926

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M. 1389. Ukualukasi, N.W. Ovamboland. 3400'. 7 Oct., 1924. B.M. No. 25 12.4.24. Type

A remarkable tufted species, alone related to *C.(L.) chapini* Allen, but differently coloured.

General characters as in the smaller species of *Chærephon*, but with a conspicuous bicolor tuft, as in the Bat figured by Allen. Size rather larger than in *chapini*. General colour of upper surface drab, the base of the hairs lighter and their extreme tips also whitish. Line of fur along the sides between humerus and femur brown, and that across the base of the interfemoral drabby brown, in neither case white as in *chapini*. Hairs of aural tuft about 13 mm. in length, their basal halves blackish brown, not reddish chestnut as in *chapini*, their terminal halves whitish. Wings pale brown throughout, quite without the contrasts of black and white and the punctuation of the membranes described in *chapini*, which are apparently very much as in *C. hindei*. Under surface pale drabby, the ends of the hairs whitish. Ears apparently as in *chapini*, their connecting membrane high, with a distinct vertical ridge on the centre of its front surface.

Skull smooth and rounded, as usual in *Chærephon*. First upper molar with an unusually prominent hypocone. Two lower incisors only in the single specimen.

Dimensions of the type:-

Forearm 39 mm

Head and body 56 mm.; tail 34; ear 16. Third finger, metacarpal 40, first phalanx 15; second phalanx 18; lower leg and foot 19.5.

Skull: greatest length 16.6 mm.; zygomatic breadth 10; mastoid breadth 9.1; front of canine to back of m3 6.1.

Hab. And Type as above.

This most remarkable tufted Bat is a very interesting and valuable accession to our collections, as we had nothing showing its really wonderful specialization. But I am in considerable doubt as to the taxonomic value of this specialization, striking as it is, for the species described by Dr. Allen as *C. cristatus* and *abae* have it far less marked, while *C. hindei* also has the connecting membrane of the ear tufted behind, the character chiefly relied on by Dr. Allen in forming the subgenus *Lophomops*. The reduction to two of the lower incisors, usual in *Lophomops* according to its describer, does not occur in *C. hindei*, which has four, as in other members of *Chaerephon*.

For the present, however, as an indication of the position of the species I provisionally use Dr. Allen's sub-generic name, while thinking it possible that the character of the aural tuft will merely prove another specific specialization in the plastic genus *Chaerephon*.

I have named this striking species in honour of Capt. Shortridge, to whose energy and enthusiasm the great success of the South West African collecting expeditions has been almost wholly due, and who was greatly struck by its remarkable appearance.

Chaerephon frater J.A. Allen, 1917

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Type, No. 49275, F ad. (alcoholic), Malela (near Boma), southwestern Belgian Congo, July 8, 1915; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 2613.

Similar to *Chaerephon hindei* in general coloration but smaller, skull less massive and the dentition weaker, with p^2 much smaller.

Coloration, including the wing membranes, practically the same as in *C. hindei*, so far as can be determined from alcoholic material, except that the proximal border

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of the wing membranes, between the humerus and femur, is without the white band of fur seen in *hindei*.

Entire body dark brown except a broad, usually well-defined pure white median area from opposite the shoulders to base of tail, covering about one-third of the ventral surface of this part of the body, varying somewhat in extent in different specimens. Wing membranes white above, except proximally from elbow to knee where they are brownish, the tone deepening toward the body; below wholly white. Interfemoral membrane brown both above and below. Ears joined by a deep membrane, fringed above with short stiff hairs about 0.5 to 1 mm, in length, and similarly covered in front and on the central part of its posterior face. Tragus minute, quadrate, about 1 mm. square. Antitragus rounded above, about 2.5 mm. in height and about 3 mm. wide at base. In 18 of the 22 well-preserved alcoholic specimens the front half of the crown behind the membrane joining the ears is naked; in the other four this area is slightly covered with fine fur. The ears are thinner and broader than in *C. hindei*.

External measurements of the type: Total length, 80 mm.; head and body, 50; tail, 30; forearm, 37; third metacarpal, 35.5; thumb to base of the prominent pad, 6; tibia, 10.5; foot, 7.2; ear from crown, 8.7; from the notch behind the antitragus, 12.

Forearm, average of 22 topotypes, 30 (35-37). Forearm in type of *hindei*, 40; in five specimens from near the type locality, 37.6 (37-39.4).

Skull, type (corresponding measurements of type of *hindei* in parenthesis): total length, 15.2 (17.6); zygomatic breadth, 9.4 (11.4); upper toothrow, 5.4 (6.7).

Seven skulls (removed from alcoholic specimens), 1 male and 6 females, measure as follows: Total length, 15.9 (15.4-16.4); condylobasal length (4 skulls), 14.3 (13.8-14.8); zygomatic breadth, 9.5 (9.0-9.9); mastoid breadth, 8.9 (8.7-9.2); breadth of braincase, 8.3 (7.9-8.4); interorbital breadth, 3.4 (3.3-3.6); breadth at base of canines, 4.2 (3.6-4.5); maxillary breadth, 6.8 (6.4-7.8); upper toothrow (c-m³), 5.4 (5.2-5.6); length of mandible, 10.1 (9.5-10.6); angle to condyle, 2.5 (2.3-2.9); depth at coronoid, 2.6 (2.2-2.9); lower toothrow (c-m₃), 5.9 (5.6-6.1).

Represented by 22 specimens in alcohol, all from Malela, the type locality.

Six skulls of *C. hindei*, two from the type locality (Fort Hall, B. E. Africa) and three from a nearby point (collection of United States National Museum) measure as follows (the corresponding measurements of six skulls of *C. frater* are given in brackets): Total length, 17.5 (17.3-18) [15.9 (15.4-16.4)]; zygomatic breadth, 10.6 (10.5-10.8) [9.5 (9.2-9.9)]; length of mandible, 11.4 (11.2-12) [10.1 (9.5-10.6)]. A single skull from Mombasa (collection of Museum of Comparative Zoology), exactly agrees in measurements with the average of the five from the Fort Hall region.

In general form the skull is a miniature of that of *C. hindei* but disproportionately weaker, especially in respect to the mandible and dentition. In four of the seven skulls examined, the premaxillae are fully ossified, as in typical *Chærephon*; in the others there is a U-shaped vacuity behind the incisors : and the incisors are not united by an osseous border. In this species; as in *C. abæ* and some other species of *Chærephon* of which I have been able to examine a series of skulls, a similar condition exists, the younger specimens often having the premaxillae imperfectly ossified at the incisive border. In one skull p^2 is in the toothrow on the left side, but on the right

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side is on the outer edge of the toothrow, as it is on both sides in all of the other five skulls. Usually p^2 separates the canine from p^4 , but in two instances p^2 is crowded so far out that the canine and p^4 are in contact. The first lower premolar is about one-third smaller than p_4 .

The nearest known form to *C. frater* is *C. hindei* (type locality, Fort Hall, Kenya district, British East Africa), from which *frater* differs in smaller size, especially in the smaller size of the skull and much weaker dentition. It also differs in color, particularly in having a larger area of white on the middle of the belly and in the absence of white on the proximal border of the wing membranes. As shown above, in a series of six skulls of each form, the extreme measurements do not overlap, the smallest *hindei* skull being considerably larger than the largest *frater* skull. The males of *hindei* have a low broad tuft of dark hair behind the membrane joining the ears, arising from the front part of the crown (not from the back of the frontal membrane, as in *Lophomops*), but in *frater* this part of the crown is usually bare in both sexes, and hence of course without the frontal tuft found in the males of *hindei*.

C. frater is about the size of *C. limbatus* (Peters) from Mozambique, with which it shares the character of white wing membranes, but *limbatus* evidently (judging from descriptions) belongs to a different section of the genus (*Lophomops*), the males being said to have a "long crest of erect hairs behind the connecting membrane of the ears" (de Winton). The absence of white on the lower parts and the character of p^2 would also exclude *limbatus* from further consideration in this connection.

Chærephon lancasteri Hayman, 1938

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Type. - B.M. no. 1937.12.8.25. Original number 724. M skin and skull, collected August 1936 at Lunzi River, Lundazi District, Loangwa Valley, N.E. Rhodesia, by D. Gordon Lancaster.

Description. - Closely similar in size to *C. (Lophomops) shortridgei* Thomas, slightly larger than *C. (L.) chapini* Allen. Crest long and bicoloured as in both these forms. Body above greyish drab, the hairs paler at the bases and slightly paler at the tips, thus producing a not very noticeable frosted effect. On the nape the hairs are notably paler throughout. The aural crest, rising from the back of the deep connecting membrane between the ears, is strongly bicoloured. The basal third is dark brown, the remainder pale buff, much as in *shortridgei*. Its length at the centre is 14.9 mm. A triangular patch on the crown, with its apex at the base of the centre of the tuft, is naked, and is entirely covered by the crest when laid flat. On the entire underside every hair is pure white from base to tip, including the narrow band along the edge of the membrane from humerus to groin. The wing-membranes are whitish, in strong contrast with the

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interfemoral membrane, which is dark greyish brown. The membrane connecting humerus and femur is heavily spotted, as in *C. limbatus* Peters and as described in *chapini*. The skull and teeth present no points of special interest. The skull in size and proportions is close to that of typical *Chærephon pumilus*. P^2 is in the toothrow, and there are four lower incisors, the inner pair slightly bifid.

Comparison. - *Chærephon lancasteri* differs from *chapini* Allen, from Faradje, N.E. Congo (the first member of the genus described with strongly developed crest), in the following respects: - *chapini* is smaller, with a forearm of 34 and 37 for type and topotype, against 39 in *lancasteri*; skull-length of 15.5 and 16, compared with 17 in the Rhodesian form; upper tooth-row 5.4 and 5.8, against 6.2. The body-colour of *chapini* is different, being described as pale cinnamon-brown above, fur whitish basally and below greyish brown, middle of belly whitish. The wing-membranes of *chapini* and *lancasteri* appear to be similar in colour.

C. (L.) cristatus Allen, from Boma, mouth of the Congo, has a short crest (9 mm.), is russet above, dark cinnamon-brown below, and has a forearm of 36.

C. (L.) abæ Allen, from Aba, N.E. Congo, has a short crest, an entirely different type of inter-aural membrane, is much larger (forearm 43), and is dark brown above and below.

C. (L.) shortridgei Thomas, from Ovamboland, S.W. Africa, is about the same size as *lancasteri*, but apart from the crest is quite distinct in colour. The dorsal surface is greyish drab rather heavily frosted by the pale hair-tips. There is a marked contrasting band of chestnut along the flanks above, between humerus and femur. Below it is pale drabby, with a pinkish tinge, only the tips of the hairs being a dirty whitish. All the membranes are of a uniform greyish hue without contrast.

C. (L.) nigri Hatt, from Bourem, Cercle de Timbouctoo, French West Africa, has a short crest (5-5 mm.), is smaller (forearm 36 mm.), has brown membranes, and is grey above and below.

C. (L.) langi Roberts, from N. Bechuanaland, in size is close to *chapini*, *shortridgei*, and the new form, having a forearm of 37.5 and upper tooth-row 6.1 mm. The

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colour above is dusky drab, below lighter, but only white along lower part of abdomen and along border of membrane below between humerus and femur. It has a unicoloured short crest.

Measurements of *lancasteri* (type). - Head and body 52 mm. ; tail 34.8 ; hind foot 6.3 ; ear 15 mm. height from meatus, width 12.5, connecting membrane between ears 6 mm. high; length of crest 14.9; forearm 39; third metacarpal 38.8, first phalanx 15.7, second phalanx 15.7; fourth metacarpal 37, first phalanx 12.5, second phalanx 8.5. Skull: total length 17; condylo-basal length 16.7; mastoid breadth 9.3; zygomatic breadth 9.7; breadth at m^3 7.4; upper tooth-row 6.2; greatest depth of skull 7.2.

Notes. - Allen's subgenus *Lophomops*, for *chapini*, *cristatus*, and *abæ*, was based on two characters: (1) the long aural crest arising from the back of the connecting membrane between the ears; (2) the reduction of the lower incisors to two. Dealing with the latter character first, it is only necessary to point out that Allen, on p. 464, in discussing *cristatus*, states that in the type and topotypes certain of the incisors had obviously been shed in life, and continues, "in the *Lophomops* group the lower incisors are doubtless normally 2-2." Allen does not mention the incisors of his third species of *Lophomops*, *abæ*. In *shortridgei* there are only two lower incisors, but they are obviously one inner and one outer. In other species of *Chærephon* some of the lower incisors are often found to have been shed.

The second character used (with some diffidence, it is true) by Allen for *Lophomops*, the erectile crest between the ears, has already been thoroughly dealt with in detail by Braestrup (1933). This author has shown that a crest, of greater or lesser length, may be developed at times of sexual activity by males of any species of *Chærephon*, and points out that Allen's subgenus must be given up, a course hinted at by Thomas in discussing *shortridgei*. While agreeing with Braestrup that the crest alone, being restricted to males at certain seasons, is itself not diagnostic in distinguishing forms, I think that when strongly developed crests such as are found in *chapini*, *shortridgei*, and *lancasteri* are associated with other differences in size and colour, these forms should be distinguished. Probably

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it will be found, when further collecting has shed more light on the distribution of these long-crested forms, that they may be linked as races of *chapini*. Until then, in view of the present wide gaps in their known distribution, it seems best to treat them as species. I have much pleasure in associating with this Rhodesian form the name of Mr. Gordon Lancaster, in appreciation of his frequent assistance to the Museum.

Chærephon nigeriæ Thomas, 1913

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Closely allied to *Ch. plicatus*, conspicuously larger than in any of the members of the *Ch. pumilus* group.

Size about as in *Ch. plicatus*. General colour dark brown throughout, except that the band of fur on the under side of the wing-membrane bordering the body between the humerus and femur is white, strongly contrasting with the uniformly dark body. Wing-membranes dull brown above and distally below, whitish proximally below. Ears and other soft parts apparently all as in *Ch. plicatus*.

Skull about the size of that of *Ch. plicatus*, but much longer and flatter, the upper profile almost straight instead of being strongly sinuate. Teeth as in the Indian species.

Dimensions of the type (measured on the spirit-specimen): -

Forearm 46 mm. (47 in second specimen).

Head and body 68; tail 37.5; ear 21; third finger, metacarpus 47.5, first phalanx 20, second phalanx 21; fifth finger, metacarpus 27; first phalanx 15; lower leg and hind foot (c.u.) 24.

Skull: condylo-basal length 18.6; mastoid breadth 11.5; front of canine to back of m^3 7.7.

Hab. Zaria Province, Northern Nigeria.

Type. Adult female. B.M. no. 11.3.22.1. Collected and presented by A. C. Francis, Esq. Two specimens.

This bat is no doubt most closely allied to the Abyssinian *Ch. bivittatus*, Heugl., a species we have not got, but which is said to be almost identical with the Indian *Ch. plicatus*. Heuglin's bat has, however, a pale under surface, which would distinguish it from *Ch. nigeriæ*, whether it proved to have the high brain-case of *Ch. plicatus* or the more flattened one of the present species.

Chaerephon pumilus naivashæ Hollister, 1916

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Type, from Naivasha Station, British East Africa. United States National Museum No. 166658, male, in alcohol, with skull removed. Collected August 7, 1909, by J. Alden Loring. Orig. No. 6955.

Description. - Like *Chaerephon pumilus pumilus* Cretzschmar, but larger, with longer forearm and larger skull; color averaging somewhat darker.

Measurements. - Type, compared with adult male of true *pumilus* from Saaita, Eritrea (number 143166), measurements of the latter in parentheses: Forearm, 42 (38) mm.; skull, condylobasal length, 16.0 (15.4); zygomatic breadth, 10.8 (10.4); interorbital constriction, 4.0 (3.6); mastoid breadth, 9.9 (9.2); mandible, 11.7 (11.8); maxillary tooth row, including canine, 6.3 (6.2); entire lower tooth row, 7.3 (7.0). Average of length of forearm in fifteen adults of *naivashæ*, 40.3; in eighteen adults of *pumilus* from Eritrea, Sudan, and Northern Uganda, 36.5.'

Chaerephon russatus J.A. Allen, 1917

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Text Fig. 25, p. 550.

Type, No. 48925, M ad., skin and skull, Medje, Belgian Congo, Sept., 8, 1910. Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 993.

Above uniform russet-brown, with a large median nearly hairless spot at shoulders; below cinnamon-brown, the extreme tips of the hairs on the median area faintly lighter in some specimens but as a rule not appreciably so; a band of cinnamon-brown bordering the underside of wing membrane between humerus and femur; wing membrane (in dry skins) dark brown proximally as far as the fourth digit, the outer portion slightly paler and semitranslucent; uropatagium dull brown, both above and below. Ears thickened on front border, blackish brown, united by a membrane at base, which projects as a rounded protuberance in front of the ears. Tragus minute, narrow, about 3.5 mm, high by about 1 mm. wide, blackish. Antitragus rather small, about 5 mm. wide at base and 3.5 in height, evenly rounded above. Tail about 34 mm, in length, the apical half free, the basal third below

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furred. Pad at the base of the thumb large. (For illustration of wing see Part II, p. 550, by Lang and Chapin.)

Collectors' measurements of type: Total length, 102 mm.; head and body, 68; tail, 31; foot, 10; ear, 21.

Additional measurements from the dry skin: Forearm, 45.7; third metacarpal, 45.4; tibia, 38; foot, 10.5; ear, length of outer border, 25.7; height from crown, 19.3.

Collectors' measurements, type and 23 adult topotypes. (4 males, 20 females): Total length, 100 (93-105); head and body, 67.8 (62-73); tail, 33.1 (29-36); foot, 11.2 (10-13); ear, 20.2 (19-22). Forearm, from the skins (same specimens), 44.6 (42-40).

Skull rather deep and narrow; median and lambdoid crests weakly developed, especially the former; preorbital ridge conspicuous; premaxillæ fully ossified, enclosing a pair of small palatal foramina. Upper incisors small, slightly separated at base with their tips nearly or actually in contact (in different specimens); lower incisors $2-2/2-2$ small, all bifid, the inner more deeply so than the outer with the inner lobe the larger; outer incisors partly posterior to the middle pair. Upper canines with a broad, deep longitudinal groove and small cingulum without cusps; lower canines without distinctive features. First upper premolar minute, closely crowded between the adjoining teeth and wholly separating them; second upper premolar large, more than half the size of m1; first lower premolar a little smaller than the second.

Skull (type), total length, 18.7; condylobasal length, 16.6; zygomatic breadth, 11; mastoid breadth, 10.1; interorbital breadth, 4.1; maxillary breadth, 7.9; breadth of braincase, 9.3; upper toothrow (with canine), 6.4; length of mandible, 12.2; angle to condyle, 2.5; depth at coronoid, 3; lower toothrow, 7.

Skull, type and 22 topotypes, total length, 18.5 (17.8-19.4); zygomatic breadth, 11 (10.5-11.4).

Represented by 27 specimens (24 skins with skulls, 3 alcoholic), all taken at Medje, September 8, 1910.

The series is exceedingly uniform in both coloration and measurements. The pelage of the dorsal surface is without light tips to the hairs, and the pelage of the ventral surface is nearly so, a few of the specimens, however, having the extreme tips of the hairs pale buffy over the median area. *Chaerephon russatus* closely approaches *C. demonstrator* (Thomas) in size and proportions, but the latter is somewhat larger, especially in cranial measurements, the zygomatic breadth being about 2 mm. greater. The upper canine and second upper premolar are said to be in contact in *C. demonstrator*, with the minute first premolar in the outer angle between them, while *russatus* the small premolar is in the middle of the toothrow, completely separating the canine from p⁴. The description of the coloration of *demonstrator* also does not agree well with that of *russatus*, nor does the indicated structure of the ears. The fleshy protuberance in front of the ears seen in *russatus* is found also in other species, which in other respects are widely different. It is said not to be present in *demonstrator*. In an alcoholic male of *russatus* there is no indication of the pair of scent glands

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at the base of the penis, mentioned as a distinctive feature of the unique type of *demonstrator*.

Chærophon (Lophomops) nigri Hatt, 1928

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Type: sexe M, adulte; localité: dans la case des passagers (située sur la berge du Niger) de Bourem, subdivision de Bourem, Cercle de Tombouctou, Soudan français; date: 6 janvier 1928; dans l'alcool; collecteur: TH. MONOD (Mission saharienne Augiérès-Draper); n° de récolte: M. S. A. D. Zoologie, n° 736 a; Mus. Paris.

Description: Similar to *C. (L.) cristatus* J. Allen. Pelage above and below, deep gray in contrast to the russet back and cinnamon drab underparts of *cristatus*. There is no band of soft

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white fur along the inner edge of the wing membrane, characteristic of *cristatus*. The fur hairs of the back extend on to the base of the wing, unlike the condition in *cristatus*. The wing membranes are a light smoke gray, as opposed to the ivory colored membranes of *cristatus*. There is little contrast between the wing membranes and the skin covering the wing bones. The wing membranes have a distinctly lighter posterior border, lacking in *cristatus*. The spotted area of the dorsal surface of the wings next to the body is more extensive than in *cristatus*. The body of this bat is smaller than that of *cristatus*.

Matériel examiné: trois spécimens, comme suit: 1 M adulte (type); 1 M [M. S. A. D. Z. n° 736 b] (de même taille que le M

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type, mais sans touffe de poils sur la couronne) et 1 F adulte [n° 736 c] (co-types), même localité, date, etc. que le type. Mus. Paris (F) et Amer. Mus. (M).

Les spécimens ont été comparés aux types et co-types de *C. (L.) cristatus* Allen, *C. (L.) Chapini* Allen et *C. (L.) abæ* Allen.

[table removed - eds.]

Les caractères crâniens ont été notés sur la F: longueur totale: 16; largeur mastoïde: 9; largeur interorbitaire: 4; largeur maxillaire: 7.5; largeur de la boîte crânienne: 8; longueur de la rangée supérieure de dents: 5; longueur de la rangée inférieure: 6.5; longueur de la mandibule: 10; mandibule, de l'angle au condyle: 4

Chaerophon pumilus elphicki Roberts, 1926

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Kershaw (*Ann. Durban Mus.* III, p. 29, 1921) has recently published an important note on the forms of *C. pumilus*, in which he places *C. limbatus* Peters as a subspecies, pointing out that the northern, typical *pumilus* has no white below as in *limbatus*, while specimens from south of the tropics again lose the white, and he therefore places them with *pumilus*. There are objections to the use of the same name for the animals occurring respectively in the northern and southern latitudes with intermediates between them, as there are sure to be differences in other respects besides those which are commonly accepted for guidance. There are a number of such cases in birds and mammals, and the present one is no exception, there being average cranial differences which require recognition. Hollister (*Bull. 99, U.S. Nat. Mus.* p. 96, 1918) has been good enough to publish dimensions of a number of specimens from E. Africa and northwards. He has apparently been guided to some extent by the length of forearm, a character dependent largely upon the distance the animals are called upon to fly from their shelters to their feeding grounds, and

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it seems evident that his Sudan and Uganda specimens should be separated, as there is a difference in the skull dimensions. Quoting his figures and adding those of specimens from Transvaal, the following results are shown:

[table removed - eds.]

Those from Zanzibar represent *pumilus*, those from Uganda and Sudan have also been allocated to the same form, but evidently require removal, those from Naivasha represent *naivashae*, those from Zanzibar *limbatus*, and those from Malelane elphicki. It is to be noted that there is an increase in size from the Equator southwards, *elphicki* representing the largest form. In colour these Transvaal specimens are very dark, glossy brown, almost black, above, and paler below on account of the paling of the tips of the hair, but still dark brown; white only appears distinctly as a narrow band between the limbs, along the flanks and inner border of the flying integument.

Type: T.M. No. 2488, adult male, Malelane Estate, E. Transvaal. Named after Capt. G. J. Elphick, to whom I am indebted for the specimens.

Chalinolobus argentatus Dobson, 1875

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The crown of the head is abruptly elevated above the face-line as in *Miniopterus*, but to a much less extent; muzzle short, very obtuse in front, broad and flattened above; nasal apertures wide apart in front; separated by a slightly concave space, opening sublaterally, bounded laterally by the front margins of the labial glandular prominences which are separated from the nostrils above by a sulcus on each side; as in the other species of this genus. Beneath, on each side of the chin, below the under lip, a smooth broad rounded elevation exists separated from its fellow of the opposite side by the

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small naked space on the lip in front of the lower incisors. These elevated sides of the chin are covered with short hairs and enclose a hollow space between. Ears very like those in *Miniopterus*, short and rather sloped backwards; the inner margin commences in a long lobule directed backwards; the margin of the ear is almost regularly convex all round from the commencement of the inner margin to a point in the outer margin opposite the base of the tragus, where it becomes slightly emarginated, and immediately beyond abruptly convex, forming a short erect lobe connected by a low band with a wart in front at the angle of the mouth, which is continuous with a horizontal lappet of thickened skin extending forwards along the lower lip almost as far as a point opposite the lower canine. With this lappet of the lower lip the outer margin of the ear is thus directly continuous. Tragus semilunate, the inner margin slightly concave, the outer regularly convex, at the base a rather large equilateral triangular lobule very acutely pointed.

First phalanx of longest finger short, intermediate in length between that of *Miniopterus* and that of *Vesperugo*, terminal phalanx very long, flexed forwards on the under surface of the first phalanx in repose; in this position it extends nearly to the middle of the metacarpal bone. Feet small; wings to the base of the toes; calcaneum long and straight, extending quite three fourths the distance between the ankle and the tip of the tail; no postcalcaneal lobule.

Tail as long as the head and body, wholly contained within the interfemoral membrane.

The ears and the face are nearly naked, a few short hairs only appearing on the glandular prominences between the eyes and nostrils; the fur of the body above and beneath extends upon the wing-membranes as far as a line drawn from the middle of the humerus to the middle of the femur; the interfemoral membrane above and beneath is quite naked, except where a small triangular patch of hair appears at the root of the tail.

The fur is tricoloured, the basal third of the hairs black, the middle third white, and the terminal third a beautiful dark silvery grey. This is the arrangement of the colours about the middle of the body; but the grey is more prevalent towards and on the head, while the dark shades prevail slightly over the grey towards the tail.

The integument of the ears and face pearly white; wing- and interfemoral membranes dusky white, translucent, traversed by well-defined reticulations and parallel lines.

The teeth are very peculiar. The inner upper incisors very long, remarkably slender and acute; the outer incisor on each side fills up the space between the inner incisor and the canine by its broad base; but its unicuspidate vertically directed summit is very short, and scarcely exceeds the cingulum of the inner incisor. The canines are extremely long and slender, and are directed almost vertically downwards and slightly outwards; the lower canines are also very slender, but scarcely more than half the length of the upper ones. The upper premolar is very acute and close to the canine;

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the posterior upper molar little more than half the size of the second molar. The lower incisors are distinctly trifid, some even appearing to have a fourth lobe; they are not crowded, and form a regular semi-circle across the wide space between the canines. The lower premolars are also, like the canines, very slender and acutely pointed, the first premolar about half the size of the second.

Length (of an adult F preserved in alcohol): head and body 2".0; tail 2".0; head 0".55; ear 0".5, tragus 0".22 x 0".1; forearm 1".7; thumb 0".25; first finger 1".7; second finger-metacarp. 1".7, 1st ph. 3".6, 2nd ph. 1".3; third finger-metacarp. 1".55, 1st ph. 0".45, 2nd ph. 0".5; fourth finger-metacarp. 1".35, 1st ph. 0".4, 2nd ph. 0".3; tibia 0".72; foot and claws 0".3.

Hab. Cameroon Mountains, western equatorial Africa.

This species has a general resemblance to *Miniopterus schreibersi* in the shape of the head and ears, in the shortness of the first phalanx and great length of the terminal phalanx of the longest finger, in the long and slender tail wholly contained within the interfemoral membrane; added to which the grey colour of the fur and corresponding size would cause specimens of this species to be readily confounded, on a superficial examination, with specimens of the European grey-coloured *M. schreibersi*.

Chalinolobus congicus Noack, 1889

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Taf. II, Fig. 1; Taf. V, Fig. 66 - 68.

Genus *Chalinolobus* bei DOBSON, Cat. Chiropt., p. 252.

Acht Spiritus-Exemplare, 4 M und 4 F. Netonna, April u. Mai. Coll. HESSE.

Die vorliegende Collection bietet nicht unerhebliche Schwierigkeiten, weil die Exemplare nach Geschlecht und mehr noch nach dem Alter in der Färbung ziemlich stark abweichen, während Schädel und Gebiss beweisen, dass sie zu einer Art gehören, welche ich geglaubt habe mit *Chalinolobus* vereinigen zu müssen, obwohl die inneren I zweispitzig wie bei *Vesperus* sind. Die Bildung des Ohres dagegen und des Mundwinkels ist durchaus die von *Chalinolobus*.

Diagnose. *Chalinolobus* mit zweispitzigen inneren I, halbmondförmigem Lappen am unteren Mundwinkel und Warze über dem Mundwinkel, rundem, nach hinten umgebogenem Lappen am unteren Ohrande, Tragus mit gerader innerer, gebogener äusserer Kaute mit spitzem Lappen am unteren Rande, weisslichgelber bis hell rötlichbrauner Flughaut, ganz von der Flughaut eingeschlossenem Schwanz und olivengelblich-silbergrauer Färbung und mehr oder weniger deutlichen weisslichen Streifen oben und unten an den Schultern.

Beschreibung. Kopf und Nase kurz, letztere breit abgerundet, die weit von einander entfernten Nasenlöcher durch eine seichte Furche getrennt. Oberlippe an den Seiten wulstig, die Lippenränder ganz vereinzelt kurz behaart. Mundwinkel mit einem grossen, halbmondförmig herunterhängenden Lappen, der nach unten mehr oder weniger scharf begrenzt ist. Auch vor dem Lappen ist die Unterlippe wulstig. Oberlippe im Mundwinkel warzig verdickt. Eine runde Warze steht über dem Mundwinkel nach dem Ohr hin. Auge sehr klein, näher dem Ohr als der Nasenspitze. Ohr gross, aber kürzer als der Kopf, breit abgerundet, innen nackt, aussen mit nackter Spitze. Der innere Hand unten zu einem runden Lappen umgebogen, der äussere verdickte Rand mit kleinem, rundem Lappen bis an den Mundwinkel vorgezogen. Tragus innen ausgehöhlt, an der Basis schmal, der äussere Rand springt über der schmalen Basis mit kleinem dreieckigem Lappen vor, mittlerer und oberer Theil des Tragus ziemlich gleich breit, die innere Kante gerade, die äussere rundlich gebogen. Zunge kurz und dick, am Rande mit einzelnen kleineren Wärzchen, sonst mit feinen Papillen. Von den 7 Gaumenfalten ist die erste und die

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letzte ungebrochen, bei den übrigen wie gewöhnlich die Innenseite der Bogen Bach hinten gezogen. Die Flughäute sind sehr zart und stark zugespitzt, zwischen Unterarm und fünftem Finger von starken, bräunlichen Adern durchzogen, sonst fein geädert, der Lappen der Schwanzflughaut unbedeutend, der Schwanz mit 7 Wirbeln ganz von der Flughaut eingeschlossen. Die Schwanzflughaut oben bis zum dritten Wirbel dünn behaart, der Humerus oben auf $\frac{1}{3}$ Länge, etwas stärker die Unterseite der Flughaut vom Ellbogengelenk bis zum Anfang des Unterschenkels. Der Daumen frei, an der Basis ohne stärkere Schwiele.

Färbung der Gesichtsseiten und des Ohrs weisslich, vor der Stirn olivenfarben, vom Auge zum Ohr ein dunkler Streifen, der nur bei einem M stärker hervortritt. Extremitäten unten weisslich, oben gelbbraun. Nägel tief schwarz. Färbung der Flughaut zwischen den Fingern weisslich, doch bei einigen M mehr rothbraun, an den Seiten und zwischen den Schenkeln hell rötlichgrau, Haar lang und fein, etwas flockig, Haarbasis etwas heller als das Haar. Färbung der Oberseite olivengrau mit silbergrauen Haarspitzen, nach hinten zu mehr olivengelb, Kehle etwas heller, sonst die Unterseite etwas dunkler als die Oberseite. Die Unterseite der Flughaut hell gelbbraun behaart. Zwei weissliche Streifen von der Schulter bis zu den Schenkeln treten nur bei einem M deutlich hervor. Die gelbliche Färbung des Hinterrückens ist bei einem Exemplar ebenfalls streitig, übrigens der Farbenton bei den alten F heller als bei den M, nur ein M sehr hell.

Bei *Chalinolobus argentatus* ist die Haarbasis schwarz, die Haarmitte weiss, die Spitze silbergrau, *Chal. variegatus* ist viel kleiner, bei *Chal. poensis* die Flughaut immer braun. Das Subgenus *Glauconycteris* ist grösser. Die breite Vorhaut ist zweilappig, an der Spitze kurz borstig behaart, das kleine Scrotum dicht hinter dem 4 mm langen Penis, dicht dahinter der After. Auch die Scheide dicht vor dem After, die Clitoris sehr klein, unten gefurcht und an der Spitze durchbohrt. Die beiden Mammæ nur bei einem F kahl mit angesogenen Zitzen.

Maasse. M, F adult.
Körper: 52 - 56, 50
Ohr: 9.5 - 10, 9.5
Mittlere Breite: 7, 7
Tragus: 4 - 4.5, 4.5
Schwanz: 42 - 45, 45
Humerus: 26 - 28, 25
Unterarm: 41 - 43, 40

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Daumen: 4.5, 4
II: 53 - 58, 54
III: 78 - 84, 81
IV: 57 - 61, 59
V: 50 - 52, 50
Femur: 17 - 18, 18
Unterschenkel: 17 - 18, 18
Fuss: 6 - 7, 7
Sporn: 12 - 14, 13

Schädel (Taf. V, Fig. 66) kurz und breit, der obere Theil des abgerundeten Occiput nach vorn gegen den Scheitel umgebogen, Stirnbeine etwas blasig aufgetrieben, Nasenbasis stark eingesenkt, der kurze und breite Oberkiefer nach oben gebogen, die Nase seicht gefurcht, die Seiten der Nasen beine mit rundlicher Leiste, die Bullae audit, mittelgross, aussen flach, innen kräftig entwickelt, der schlanke Jochbogen in der Mitte eckig in die Höhe gezogen.

Am Unterkiefer die Symphyse breit und kräftig, schräg nach vorn gerichtet und unten mit kleinem Zacken, der horizontale Ast gerade, der aufsteigende Ast niedrig und breit, nach hinten gerichtet, der niedrige Proc. coron. nach vorn und stark nach aussen gebogen, die obere Kante des aufsteigenden Astes flach eingebogen.

Maasse von M adult.: Scheitellänge 12, Basilarlänge 11.5, Schädelkapsel 8.5, Hinterhaupt über dem For. occipit. 5, For. occipit. 3 breit, 2.5 hoch, Scheitelbreite hinten 7.25, die Stirn hinter der Nase 5 lang und breit, Bullae aud. 3, Höhe der Schädelkapsel 6, Kieferlänge von I bis M III 5, Gaumenlänge 5, Breite 3.25. Die Schädel der F sind um ein Geringes kleiner. Unterkiefer bis zum Condylus 9, Höhe der Symphyse fast 2, des horizontalen Astes 1, des aufsteigenden Astes unter dem Proc. coron. 3.

Gebiss (Taf. V, Fig. 67 - 68) von *Chalinolobus*:

$I^{2+2}/6 \ C^{1+1}/_{1+1} \ P^{1+1}/_{1+1} \ M^{3+3}/_{3+3}$.

Oben I innen durch eine 1.25 breite Lücke getrennt, stark nach vorn und mit der Spitze etwas nach einwärts gerichtet, massig gebogen, stark zugespitzt mit kleinem hinteren Nebenzacken, der bei allen Exemplaren vorhanden ist. I aussen sehr klein, eng zwischen I innen und C, stumpf zugespitzt, wie I innen gerichtet. C schlank gebogen, etwas weniger nach vorn gerichtet als I innen, hinten flach cannelirt, Basalwulst wie bei I, P und M schwach, innen zackig vorspringend. P kaum $1/2$ C, nach innen gerichtet, an der Basis ziemlich breit, aussen cannelirt, mit schlanker

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Spitze, wenig gebogen. M I und II identisch, die Aussenzacken niedrig, der vordere stärker, erheblich nach vorn gerichtet. Der hintere innere W-Zacken stärker und höher als der vordere, der innere Nebenzacken hoch, mit scharfem Rande und starkem vorderen, nach innen gerichteten Zacken. M III schmal, der vordere Aussenzacken stark, nach vorn gerichtet, die niedrigen hinteren Zacken ebenfalls nach vorn gebogen.

Unten I klein, undeutlich dreilappig, in der Richtung der Kiefer-Symphyse schräg nach vorn gerichtet. C schlank, stark nach aussen und etwas nach hinten gerichtet, vorn innen mit Nebenzacken. P I sehr klein und niedrig, kaum $1/4$ C, breit dreieckig zugespitzt, Aussenseite etwas nach innen gerichtet. P II schlank, $= 1/2$ C. Die Aussenzacken von M breit dreieckig, der vordere um $1/3$ höher als der hintere, Die Hauptzacken der M sind etwas niedriger als C und nehmen nach hinten etwas an Grösse ab. Der innere Nebenzacken von M I ist stark nach vorn, der hintere von M II stark nach hinten gerichtet

[*Chamtwaria*](#) Butler, 1984

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CHAMTWARIA, nov. genus

Diagnosis: Resembling *Kerivoula* in having three upper premolars, the middle one (P3/) smaller than the anterior one (P2/) but less reduced than in *Myotis*; differing from *Kerivoula* in the longer, narrower face, nasals extending above canines, infraorbital foramen close to tooth row, the sharp infraorbital crest, and the less elevated paracone of P4/.

Type and only species: *Chamtwaria pickfordi* nov. spec.

[*Chamtwaria pickfordi*](#) Butler, 1984

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Chamtwaria pickfordi nov. spec.

Diagnosis: With the characters of the genus. P2/-M3/ 10.3 mm.

Holotype: KNM.CA 2237, the anterior part of a skull, from Chamtwara. This is the only known specimen.

The species is named after Dr. Martin Pickford.

Description (fig. 25 A, A'):

The skull is broken off posteriorly at the back of the olfactory chamber, and most of the alveolar portion of the maxilla is missing on the right side. On the left side the orbit and the root of the zygoma are preserved, and the three premolars are complete, but the canine and molars are represented only by their broken bases or roots.

Chamtwaria is large for a vespertilionid; in size of molars and width of palate it is comparable with *Scotophilus nigrita* and *S. heathi*, but the postcanine tooth-row is longer.

The face anterior to P4/ is narrower and less depressed than in *Myotis* and *Kerivoula*. Its width, behind the canines, is about equal to the total length of C - P4/, as in *Natalus*. The dorsal surface of the face is flattened, lacking the median concavity of *Kerivoula* and *Myotis* and thus resembling *Natalus*. The profile rises rapidly above the orbits. There are no sutures, except a trace of the internasal suture. Grooves show the position of the ascending processes of the premaxillae, which reached back to above the posterior end of the canines. The nasals are long, extending beyond the canines and reducing the size of the external narial opening. The large anterior emargination of the skull of other Vespertilionidae is absent, but there is more

resemblance to *Natalus*. The zygomatic arch arises above M2-3/. Its dorsal edge continues as a sharp crest below and anterior to the orbit, passing in front of the lachrymal foramen, as in *Myotis*; in *Kerivoula* this crest is weak and the lachrymal foramen opens on the face. The lachrymal foramen is above the anterior end of M1/ as in *Kerivoula* and *Myotis*; in *Natalus* it is above M2/. The infraorbital foramen is low on the face above the anterior end of P4/; in *Kerivoula* and *Myotis* it is higher, nearly level with the lachrymal foramen; in *Natalus* it is low but over P3/. The palate is proportionately longer and narrower than in *Kerivoula* and *Myotis*, and also more arched; it resembles *Natalus* in proportions. Its anterior border is not well preserved, but the emargination did not reach beyond mid-canine level. Posteriorly, the lateral part of the palate ends immediately behind M3/; the median part extended back under the nasopharynx, but its posterior edge is not preserved.

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To judge by its broken base, the canine seems to have been comparatively small. The first two premolars (P2,3/) are proportionately larger than in *Kerivoula*, so that the total length of the premolar series is 79 % of the length of the molar series, compared with 55 -65 % in *Kerivoula*. In *Myotis* both P2/ and P3/ are reduced, especially P3/ which in some species is minute and displaced lingually. P2/ is larger than P3/ in *Chamtwarina*, as in *Kerivoula* and *Myotis*, but in *Natalus* it is much smaller. Alveoli on the right side of the specimen show that P2/ has two roots. The crown is broadly oval in outline, and less compressed anteroposteriorly than in *Kerivoula*. There is a single cusp, blunted by wear, and buccal and lingual cingula. P3/ is proportionately narrower than P2/, and not compressed as in *Kerivoula*; it is separated from P4/ by a short space. In *Natalus* P3/ is triangular. P4/ of *Chamtwarina* approaches an equilateral triangle in outline; it is much shorter and less wide than M1/. The paracone is not elevated as in *Kerivoula*, but only about as high as on P2/. As far as can be seen from their broken bases, M1/ and M2/ resemble those teeth in *Kerivoula* and *Myotis*. On M1/ the metastylar apex projects farther laterally than the parastyle, and on M2/ parastylar and metastylar apices are

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equally prominent. The lingual region is more extended on M1/ than on M2/. The roots of M3/ indicate a short, transverse tooth. In *Natalus* M1/ and M2/ are more quadrate and M3/ is less reduced in length.

Relationships:

The fossil genus that is most like *Chamtwarina* is *Stehlinia*, from the U. Eocene-Oligocene of Europe (Revilliod 1922, Sigé 1975). This agrees in having a long face, with nasals extending above the canines, though the face is flatter and there is a median dorsal concavity; the lower edge of the orbit is sharp; the infraorbital foramen is low, near the teeth. Except the dorsal concavity, these characters are also shared with *Natalus*. The premolars of *Stehlinia* resemble *Natalus* and differ from *Chamtwarina*: P2/

is small and one-rooted; P3/ is a larger, triangular tooth which frequently develops a third root; P4/ is wider in comparison with M1/, and its paracone is elevated. Sigé (1975) considers *Stehlinia* to represent a primitive vespertilionoid stock of which *Natalus* and

Kerivoula are the least modified living descendants. *Kerivoula* and *Chamtwarina* have advanced beyond *Stehlinia* in the simplification of P3/ and enlargement of P2/. *Myotis* also has P3/ smaller than P2/, but both teeth have diminished in size as the face has shortened, leading to the disappearance of P3/ in other vespertilionids. Thus *Kerivoula* and *Chamtwarina* show in their premolars an early stage in the development of a derived vespertilionid character. *Chamtwarina* is more primitive than *Kerivoula* in skull characters in which it resembles *Stehlinia* and *Natalus*.

How to express these relationships taxonomically is mainly a matter of taste. Sigé (1975) removed *Kerivoula* from the Vespertilionidae and included it with *Stehlinia* in a family Kerivoulidae, but he retained the Natalidae as a separate family. Van Valen

(1979) combined Kerivoulidae and Natalidae with Thyropteridae, Furipteridae and Myzopodidae into a single paraphyletic family Natalidae, noting that they lack derived characters of Vespertilionidae, mostly characters of the humerus. Both these arrangements

have the disadvantage of obscuring the relationship of *Kerivoula* to *Myotis*. I would therefore retain *Kerivoula* in the Vespertilionidae, subfamily Kerivoulinae, but would exclude *Stehlinia*. *Chamtwarina*, unlike *Stehlinia* but like *Kerivoula*, has advanced in the vespertilionid direction in premolar organisation, and it may also be included in the Vespertilionidae. It is provisionally placed in the Kerivoulinae, though a case might be made for making it the type of new subfamily.

Choerephon (Lophomops) langi Roberts, 1932

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Choerephon (Lophomops) langi sp. nov.

In size coming near to *C. (L.) shortridgei* Thomas of Ovamboland, but differing in colour and its shorter and uni-colour crest. Colour above dusky drab, including the whole frontal crest; below lighter coloured, becoming pure white on the lower part of abdomen and along the border of the membrane in a fairly broad band to the base of the humeri.

Type: T.M. No. 6554, adult ♂, Tsotsoroga Pan, Northern Bechuanaland, 29th June, 1930, No. 870, Vernay-Lang Kalahari, Expedition. Length of head and body 53, tail 38, hind foot (c.u.) 8, ear 16. Crest 9, forearm 37.5, first phalanx of 3rd digit 36, 2nd 16.5, 3rd 23 mm. Skull: length from canines 17, zygomatic width 10.4, width of brain case 8.5, of palate across m² 7.6, width across canines 4.2, length of upper tooth-row (c-m³) 6.1 mm.

Chrysopteron Jentink, 1910

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Chrysopteron n. g.Dental formula: $i. 2-2/3-3$, $c. 1-1/1-1$, $pm. 3-3/3-3$, $m. 3-3/3-3 = 38$.

Braincase normal; second upper premolar very small and not to be seen from without; middle and second lower incisors with four cusps.

Type of the genus is *Weberi* Jentink.

Cistugo Thomas, 1912

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Allied to *Myotis*, both with differently proportioned teeth and with glands in the wings.

Skull essentially as in *Myotis*, but the brain-case not specially vaulted and the muzzle rather less pinched in laterally.

Dental formula as in *Myotis*.

Incisors of the same essential structure as in *Myotis*, but shorter. Canines similar. Small premolars subequal, minute, not half as large as the incisors, short, stumpy, quite without the similarity to a minute canine shown at least by the anterior one in *Myotis*, their tips barely rising to the level of the cingulum of the canine, the two closely pressed together and just filling the space between the canine and the large premolar. Large premolar with an unusually well-developed antero-internal cusp, as high as the large inner cusp of the molars. Lower incisors as in *Myotis*; canines proportionally short, barely rising as high as the posterior premolar; premolars all with their antero-posterior less than their transverse diameter, the two small ones closely crowded together between the canine and posterior premolar.

General external characters as in the smaller species of *Myotis*. Tragus of medium length, differing from that of most species of *Myotis* by being broader slightly above its base than at the base, its inner and outer edges both slightly convex.

Wings with peculiar thickened glands in them on the outer side of the forearms distally; three present on the left side and two on the right in the single specimen, but the situation of the third one is perceptible in the right wing, so that the normal number is probably three; the glands themselves about 3 - 3.5 mm. in length by 1 - 1.5 in breadth, more sharply outlined than the corresponding glands in *Pizonyx*; also situated closer to the forearm than in that genus, less in the centre of the wing.

Type: -

Cistugo lesueuri Roberts, 1919

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This genus was first described by Thomas (*ann. and Mag. N.H.* ser. 8, vol. x. pp; 204-206, 1907) on a single specimen taken in Angola. Apparently no more specimens have since been collected, and it is therefore pleasing to be able to record yet another specimen from so far south, and apparently resembling another species. The present one differs most markedly from *Cistugo seabrae* in size, the cranial and external characters seemingly differing but little; there appear to be no glands on the wing, however, and the colouration is somewhat different, both of which may eventually prove to be of little importance. *C. seabrae* is described as: "General colour dull drab, the bases of the hairs everywhere slaty, the tips above drab, below whitish. Membranes brown, with whitish edges, the light-coloured reticulations conspicuous." In the present species, the base of the hair is black, the upper parts as far as the crown terminally "honey-yellow" (Ridgway, 1912), the under parts as far as the chin much paler, yellowish white. The top of the head shows less of the yellowish colour and merges into the dark brown which characterises the face. Hair on the edge of the wing membrane merging from

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the yellow of the back to brown externally. Ears and nose almost naked, the former more thickly haired at the base posteriorly. The specimen being a dried skin cannot be accurately described as regards the ears; but apparently the tragus is almost straight on its inner margin, the outer margin convex. In the teeth, which are worn, the inner anterior cusp of P^4 is in line between the same cusps of P^3 and M^1 .

The following comparative measurements will serve to show the difference in size:

Cistugo seabrae; *Cistugo lesueuri*
 Head and body: 40; 47
 Tail: 40; 43
 Tibia and hind foot (c.u.): 18.2; 19.5
 Ear: 12; 13
 Skull: greatest length: 13.2; 14

Brain case: 6.6; 7
 Front of canine to back of M^3 : 4.6; 5.3
 Forearm: 32.5; 34.5
 Third metacarpal: 31.5; 33.8
 First phalanx: 10.7; 11
 Second phalanx: 9.7; 10.8

Type, old M, T. M. no. 2286, taken at Lormarins, Paarl District, 15th September, 1917, by J. S. Le Sueur, Esquire, by whom it was rescued from a cat.

Cistugo seabrae Thomas, 1912

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General appearance that of a *Pipistrellus*, say *P. kuhlii*, to which there is a considerable resemblance in size and colour. Ears of average size, their anterior margin convex at base, then nearly straight to the tip, which is narrowly

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rounded; outer edge angularly concave above, convex below. Tragus pointed, its inner and outer edges both slightly convex, a rounded lobe at its outer base. Wings from the base of the toes. No post-calcarea lobule. Tip of tail projecting. General colour dull drab, the bases of the hairs everywhere slaty, the tips above drab, below whitish. Membranes brown, with whitish edges, the light-coloured reticulations conspicuous.

Skull and teeth as described above.

Dimensions of the type (measured on the spirit-specimen): -

Forearm 32.5 mm.

Head and body 40 mm.; tail 40; ear 12; tragus on inner edge 5; third finger, metacarpal 31.5, first phalanx 10.7, second phalanx 9.7; lower leg and foot (c. u.) 18.2.

Skull: greatest length 13.2; basi-sinual length 10.1; breadth of brain-case 6.6; front of canine to back of m^3 4.6.

Hab. Mossamedes.

Type. Adult female. B.M. no. 6.1.3.3. Presented by the Lisbon Museum.

This interesting little bat, which I have named in honour of Senhor A. F. de Seabra, C.M.Z.S., of the Lisbon Museum, is distinguishable from *Myotis* by the presence of glands in its wings, by the reduced proportions of its anterior premolars, and the large antero-internal cusp on p^4 . Its general appearance is rather that of a *Pipistrellus* than a *Myotis*.

Clæotis Thomas, 1901

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Allied to *Hipposiderus*, *Asellia*, and *Triænops*.

Nose-leaf tridentate, its details apparently somewhat as in *Triænops*.

Ears short, rim-like, almost without tip, their outer and inner edges arising close together.

Thumbs extremely minute.

Tail scarcely projecting from membrane.

Anterior upper premolars present, but minute. Last molars nearly as large as the penultimate.

Nasal part of skull disproportionally small and feeble. Basal region broad between bullæ, without vacuities, strongly ridged laterally.

This bat does not appear to be assignable to any known genus. Its ear-structure, described in detail below, is quite unique, the shape of the ears being in this family very characteristic of the different genera. The complicated tridentate structure of the nose-leaf recalls *Triænops*, but the proportions of the skull are very different, nor is there any trace of the remarkable vertical expansion of the zygomata present in that bat. Nor do any of the species of *Asellia* show any approximation to *Clæotis*.

It is unfortunate that the two specimens were both sent as skins, so that, although one has been softened, the nose-leaf cannot yet be accurately described in detail, nor can any figure be given.

Clæotis Percivali Thomas, 1901

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Size very small, perhaps less than in any other member of the family. General build very light and delicate. Nose-

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leaf with many resemblances to that of *Triænops*, the posterior part similarly tridentate, the three points thinly hairy; below them in the middle line there is a small median projection, not so long as in *Triænops*; in front of that, again, there is something representing the anterior median plate in *Triænops*, but it is produced forwards into two small projecting points; horizontal horseshoe small, a narrow supplementary leaf apparently present. Outside and behind the tridentate leaf, between it and the eyes, there are two small projections covered with long hairs. Ears short, their outer and inner margins rising close together some way behind the eyes; the two margins are symmetrical with each other, rising vertically a short distance, then bending round abruptly and passing straight back to meet each other at the scarcely perceptible tip; the greatest height of the ear behind is only about one fifth higher than the point at which the two margins bend backwards in front. In fact the whole ear is very like a man's "stand-up" collar with the angles in front rounded off.

Wings very delicately made; thumbs quite minute, the terminal part free from the membrane only about 2 millim. long, including the claw. Wings attached to the distal end of the tibiae. Calcars short, reaching only about halfway up the tibiae. Tail apparently of only five joints, the last one scarcely projecting from the membrane, which is acutely pointed behind.

Fur fine, soft, and silky, about 6 millim. long on the back. Face bright buffy; crown greyish; back greyish brown, the hairs uniformly coloured. Hairs of underside slaty grey basally, yellowish white terminally.

Wing and interfemoral membranes uniformly smoky brown.

Skull with the part in front of the orbits much less broad and swollen than in other members of the family, the braincase being disproportionally large. Zygomata strongly convergent forwards. Palate ending opposite the middle of m^2 . Basal region of skull broad between the bullae, without vacuities, but with two strongly marked divergent ridges running from the base of the pterygoids to the antero-internal corners of the bullae; sphenoidal fissures large and widely open.

Upper incisors spatulate, indistinctly bicuspid. Canines with a small posterior secondary cusp at half their height. Anterior upper premolars minute, crushed in in the outer angle between the approximated canine and posterior premolar. Last molar about four fifths the size in cross-section of

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penultimate molar. Lower incisors tricuspid, overlapping. Anterior lower premolar well developed, two thirds the height of the next.

Dimensions of the type: -

Forearm 31 millim.

Head and body (in flesh) 35; tail 28; ear 8.

Skull: greatest length 13; basal length 9.5; zygomatic breadth 7; breadth above orbits 3.3; interorbital constriction 1.8; mastoid breadth 6.5; front of canine to back of m^3 3.8.

Dimensions of a specimen softened and put in spirit: -

Forearm 32.

Head and body (c.) 34; tail 26; height of ear behind 5; third finger, metacarpus 26, first phalanx 10.5, second phalanx 14; lower leg 13.5; hind foot (s. u.) 5.5; calcar 8.

Hab. Takaungu, N. of Mombasa, British East Africa.

Type. Male. B.M. no. 1.5.1.11. Collected 15th February, 1901, and presented by Mr. A. B. Percival.

Mr. Percival is to be congratulated on the discovery of this interesting little bat, which is widely different from anything hitherto known.

Clæotis percivali australis Roberts, 1917

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3 MM, 5 FF, Mooimeisjesfontein, Rustenburg (Powell).

This southern race differs from the typical one in being slightly larger, and in the lower anterior premolar not reaching to half the height of the posterior premolar. The following are maximum and minimum measurements taken from the series as compared with the typical race :-

C. p. australis; *C. p. percivali**

Head and body: 41-33; 35

Tail: 31-22; 28

Ear: 9-8; 8

Skull-

Extreme length: 13.6-13.2; 13

Greatest width: 7.6-7.2; 7

Mastoid width: 7.6-6.8; 6.5

Dental series: 4.3-4.1; 3.8

Forearm: 35-33; 31

Third finger, metacarpus: 29-27; 26

Third finger, first phalanx: 11.8-10.6; 10.5

*Cf. *Ann. and Mag. Nat. Hist.* (7), Vol. VIII (1901), p. 28.

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Colour.- Face "warm buffy," browner towards and round the lips; top of head and back buffish-brown, lighter on the posterior margin of the wings; under surface varying from light buff to golden orange, the base of the fur darker, in proportion to the colour of the tips.

Type. - T.M. No. M 1670, F.

Metatypes.- T.M. Nos. M 1669, 1671-1676.

Coleura kummeri Monard, 1939

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Collection: N^{os} 879 M, 880 F, 881 F, 882 F, 883 F, 884 M, 885 F, 886 M, 887 M, 888 F, 889 F, 890 M, 891 F, 3 crânes 892, Madina, Boé. Tous les exemplaires proviennent de la même station, une case indigène dans les environs de Madina Boé.

Le genre *Coleura* comprend de petites Chauves-souris appartenant à la famille des Emballonuridae. Les espèces de ce genre sont extrêmement rares dans les collections, peu nombreuses et répandues seulement dans l'Est du continent et dans les îles Seychelles. Le Catalogue de DOBSON (1878) n'en cite que deux espèces: la première, *seychellensis* de PETERS (1868), la seconde *afra*, du même auteur (1867); la première vient, comme son nom l'indique des îles Seychelles et de Zanzibar; la seconde est du continent. du Mozambique. Le Catalogue de TROUESSART et son Supplément n'ajoutent rien à ces renseignements. En 1915, OLDFIELD THOMAS décrit deux nouvelles espèces de ce genre, *gallarum* et *silhouettæ*, (avec deux sous-espèces de *gallarum*) et la distribution du genre gagne un peu vers le centre du continent: Suakim, Aden, Somaliland, région du Nil supérieur, du Congo au Zambèze. Les dernières citations de ce genre sont celles de ALLEN, LANG et CHAPIN (Bull. Am. Museum 37, 1917. p. 424) à Aba (Congo Belge) et de ALLEN et LOVERIDGE (1933) qui ont trouvé l'espèce dans les matériaux de l'expédition américaine au Tanganyika; la surface distributive est accrue de Voi, à la côte, jusque dans la région des grands lacs (Mwanza). Ainsi donc, le genre *Coleura* apparaît à la fois comme fort rare, oligomorphe, étroitement localisé dans les Seychelles et dans

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la partie Est du continent. Ce me fut donc une très grande surprise d'en trouver des exemplaires dans l'extrême Ouest africain, en Guinée portugaise, dans l'intéressante région du Boé. L'énorme hiatus qui sépare le Boé du lac Tanganyika est de l'ordre de 5000 km au moins.

Comme il fallait s'y attendre, les exemplaires de Guinée montrent avec *Coleura afra* des différences constantes dans les 13 exemplaires rapportés; ces différences sont suffisantes pour élever au rang d'espèce la forme guinéenne; elles sont du même ordre, pour le moins, que celles utilisées par THOMAS POUR LA CREATION DE SES ESPECES; EN OUTRE, CHOSE CURIEUSE, NOTRE NOUVELLE ESPECE OFFRE SOUVENT DES CARACTERES INTERMEDIAIRES ENTRE *SEYCHELLENSIS* ET *AFRA*.

NOUS NE POUVONS MIEUX FAIRE QUE DE DEDIER CETTE INTERESSANTE ESPECE A M. ALBERT KUMMER, agent de la Compagnie française, à Bissau, notre ami à qui nous avons dû un aimable foyer et une aide toujours active et bienveillante.

Description. Voisine de *afra* PETERS et en offrant les dimensions, à l'exception du tibia, toujours plus long que dans cette espèce et intermédiaire entre *afra* et *seychellensis*.

Museau allongé. conique, dépassant l'ouverture buccale de 3 mm environ; narines ouvertes à l'extrémité de deux courts tubes juxtaposés, séparés par un profond sillon longitudinal; le milieu de la lèvre supérieure montre un petit tubercule nu. Lèvre supérieure et commissure buccale garnies d'une frange de cils raides. Lèvre inférieure munie en son milieu d'un organe (glandulaire ?) nu, saillant, cordiforme, souvent partagé par un sillon médian, comme dans *seychellensis*, mais à la différence de *afra*.

Oreille large, arrondie, atteignant, rabattue en avant, l'origine des tubes nasaux; le lobe commissural est plus redressé que dans *afra*. Tragus de trois mm environ, à bords parallèles. à sommet arrondi, muni à sa base, du côté externe, d'un petit tubercule.

Membranes aliformes brunes transparentes; le pelage, à la différence de *afra* où il s'étend du milieu de l'humérus au tiers distal du fémur et jusqu'au point de sortie de la queue, ne dépasse pas le corps dans la région interfémorale et le débordé à peine sur les côtés du corps. La longueur de l'uropatagium, prise sur l'axe, est de deux fois celle de la queue. La membrane naît de la cheville ou légèrement en dessus.

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Le pelage est entièrement brun-roux, la base des poils blanchâtres, comme dans *gallarum nilosa* THOS.

Mâchoire supérieure: incisives obliques, convergentes, largement séparées, leurs pointes séparées par un intervalle égal à celui qui les sépare des canines. Celles-ci avec cingulum développé, formant en avant et en arrière des pointes accessoires, l'antérieure longue et aiguë, la postérieure plus courte. Première prémolaire toute petite, n'atteignant que le milieu de la pointe accessoire postérieure, formée d'une base cylindrique large d'où s'élèvent deux petits tubercules. Deuxième prémolaire presque aussi grande que la canine. Molaires bien conformées, la première avec parastyle et première commissure rudimentaires, la deuxième complète avec la première commissure recourbée à son origine en avant et en dedans, la troisième avec métastyle, quatrième commissure et hypocône absents ou rudimentaires. Protocônes diminuant de la première à la seconde molaires.

Mâchoire inférieure: les trois incisives trifides, juxtaposées, un peu séparées des canines. Cingulum de la canine bien développé, formant une pointe accessoire, bien visible en avant, cachée par la prémolaire en arrière. Première prémolaire bien développée, plus petite que la seconde. Molaires toutes à cinq tubercules bien développés, les protoconides les plus hauts.

Crâne: Rostre large et plat, à peine concave au milieu. Bord antéro-inférieur de l'orbite saillant, cachant la série dentaire en vue sagittale. Processus postorbitaire long, étroit, falciforme. Impression du basisphénoïde large, profonde, bordée en arrière par une crête saillante du basioccipital. Dimensions à peu près celles de *afra* les plus grandes du genre.

Les points principaux qui permettent de différencier cette espèce sont: longueur du tibia (18 mm), présence d'un sillon à la lèvre inférieure, membranes nues dès le corps. Ces caractères placent *C. kummeri* entre *seychellensis* et *afra*. La nudité des membranes, la présence d'un sillon labial sont de *seychellensis*; la forme du museau et des narines, les dimensions sont de *afra*; enfin la longueur du tibia est intermédiaire entre ces deux espèces (15 mm dans *afra* et 25 dans *seychellensis*). La grandeur est celle de *afra* et non de *gallarum*, plus petit.

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[table removed - eds.]

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Crâne, longueur totale: 17 mm
Largeur zygomatique: 9.6
" mastoïdienne: 8.8
" niveau M₂: 7.4
Série dentaire supérieure: 8

Mœurs. Les mœurs sont crépusculaires et nocturnes; l'espèce forme ordinairement des troupes assez nombreuses. Au repos, la première phalange du doigt II se replie sur le métacarpe et la seconde sur la première. Au troisième doigt, ce même mouvement n'est qu'ébauché par la première phalange, mais accompli nettement par la deuxième, à la manière de *Saccolaimus peli* TEM. (v. Bull. Am. Museum 37, 1917, p. 515).

Cynonycteris Peters, 1852

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Dentes primores $4/4$, canini $1^{1/1}$, molares $5^{5/6}$; index unguiculatus; cranium maxime ante processum zygomaticum ossis frontis coarctatum; mammae anteriores; qlans penis mollis; pollex patagio involutus; cauda brevis vel brevissima.

Die Arten sind zum Theil in Indien, zum Theil in Africa zu Hause

Cynonycteris Angolensis Bocage, 1898

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Cynonycteris Angolensis.

Syn. *Cynonycteris aegyptiaca*, Bocage (non Geoffroy). *Jorn. Ac. Sc. Lisboa.*, 2^a série, t. I, 1889, p. 15; *Cynonycteris* sp? Bocage, *Jorn. Ac. Sc. Lisboa*, 2^a série, t. II, 1892, p. 174, fig. 2.

Taille à peine inférieure à celle de la *C. collaris*. Tête de forme pyramidale, large en arrière, à museau étroit et obtus; les yeux plus rapprochés de l'oreille que de la narine; les oreilles beaucoup plus longues que la distance de la narine à l'œil, ovales, arrondies au bout. Membranes de l'aile et des oreilles noirâtres.

Pelage laineux et abondant, plus court sur la poitrine et l'abdomen; les poils de chaque côté de la tête, de la gorge et de la face intérieure du cou, sensiblement plus longs.

Les membres sont couverts en dessus de poils longs et serrés, les antérieurs jusqu'à un peu plus de la moitié de l'avant-bras, les postérieurs jusqu'à l'articulation du pied; en dessous, les poils couvrent les membres antérieurs jusqu'à la moitié de l'avant-bras, et sur les membres postérieurs arrivent à peine jusqu'au premier tiers de la jambe. Les ailes présentent sur leur face supérieure, de chaque côté de l'avant-bras, une étroite bande de poils et une bande plus large de chaque côté du corps, allant de la moitié de l'humerus à l'extrémité supérieure du tibia, et se prolongeant sur le côté externe de celui-ci jusqu'au pied. La membrane interfémorale est presque entièrement couverte en dessous de poils aussi longs et aussi fournis que ceux du dos.

La membrane pré-brachiale est garnie en dessous de petits poils laineux clairsemés; des poils de même nature forment au dessous du membre antérieur une bordure, limitée par une ligne courbe, qui finit sur le milieu de l'avant-bras. L'interfémorale, couverte de poils dans sa portion centrale, laisse de chaque côté un espace nu.

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Sur le dos et la face supérieure des ailes les poils sont bruns, couleur de tabac; le dessous de la tête est d'une teinte un peu plus claire, mais le museau est plus rembruni. Le cou et la gorge sont d'un brun pâle tirant au grisâtre; la couleur des parties inférieures se rapproche de celle du dos mais prend un ton plus clair, grisâtre, sur le milieu de la poitrine et du ventre.

Plis du palais au nombre de sept, dont trois simples suivis de quatre divisés; les deux derniers dentelés, tous les autres lisses. Les trois plis antérieurs, simples, touchent respectivement par leurs extrémités à la première, à la seconde et à la troisième prémolaire; des quatre plis divisés, les deux premiers sont en rapport avec les deux molaires et les deux derniers les suivent, gardant, entre eux et avec celui qui les précède, à peu près la même distance. Le fond du palais est limité en avant par deux lignes dentelées concentriques assez rapprochées entre elles.*

Dimensions:

M; F
Tête et corps: 120 mm.; 140 mm.
Queue: 13; 13
Tête: 45; 48
De l'œil à la narine: 15; 15
Oreille: 23; 22
Avant-bras: 80; 79
Pouce: 32; 33
3.^{me} doigt, métacarpe: 56; 60
" " 1.^{re} phalange: 40; 40
" " 2.^{me} " : 50; 53
3.^{me} doigt, métacarpe: 53; 56
" " 1.^{re} phalange: 26; 26
" " 2.^{me} " : 28; 29
Tibia: 32; 33
Pied: 20; 21

*: L'examen du palais de trois espèces d'Angola, dans nos collections, nous autorise à considérer les différences qu'elles présentent dans la forme et la disposition de ses plis, comme pouvant fournir des caractères différentiels de quelque valeur pour la détermination spécifique des Chiroptères du genre *Cynonycteris*. Le palais du *C. straminea* porte quatre plis simples et trois divisés, dont les deux derniers sont dentelés; et en arrière de ces plis, au fond du palais, se trouvent quatre lignes courbes concentriques également dentelées; chez le *C. collaris* il y a aussi quatre plis simples et trois divisés, mais le fond du palais est limité en avant par une seule ligne dentelée disposée en angle aigu; enfin le palais du *C. angolensis* nous présente trois plis simples et quatre divisés, et plus en arrière deux lignes dentelées. (V. *Jorn. Ac. Sc. de Lisboa*, II ser., t. 2.^{do} p.p. 174 a 177, figs. 1, 2, 3.)

Nous appelons sur ce sujet, qui nous semble assez intéressant, l'attention des zoologistes que seraient à même de pouvoir examiner le palais des autres espèces du genre *Cynonycteris*.

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Habitat: *Pungo-Andongo* au nord du *Quanza*, *Cahata* et *Quibula* dans l'intérieur de *Benguella*.

La *C. angolensis* est une espèce à ajouter aux trois espèces dont on avait déjà constaté l'existence en Angola: *C. straminea* dont nous possédons des exemplaires recueillis par Anchieta dans plusieurs localités; *C. collaris* très répandue dans l'Afrique équatoriale et australe et à laquelle nous croyons pouvoir rapporter des individus envoyés par notre regretté naturaliste de *Pungo Andongo* et de *Quidumbo*; enfin, *C. torquata*. Dobson, dont les exemplaires typiques, provenant d'Angola, font partie des collections du Muséum Britannique.

Il est impossible de confondre la nouvelle espèce, avec la *C. straminea* et la *C. collaris*. En effet, même sans tenir compte d'autres caractères différentiels, tels que l'infériorité de la taille, la diverse conformation de certaines parties, et la différente coloration du pelage, la sculpture de son palais nous permet de la séparer, nettement, de ses deux congénères, car, comme nous l'avons déjà observé, le palais de la *C. angolensis* présente trois plis simples et quatre plis divisés, tandis que les deux autres espèces ont quatre plis simples et trois plis divisés.

Quant à la *C. torquata*, la description publiée par Dobson, et surtout les dimensions signalées par cet auteur, d'après un individu adulte du Muséum Britannique nous semblaient contraires à toute idée d'assimilation; mais désirant arriver par comparaison directe à un résultat plus positif, nous avons adressé à notre ami Mr. Oldfield Thomas un de nos exemplaires de *Quibula* avec prière de vouloir bien le comparer aux exemplaires typiques du Muséum Britannique. Or le savant zoologiste du Muséum Britannique, vient de nous informer, que la *Cynonycteris* de *Quibula* ressemble à la *C. torquata* sous le rapport des plis du palais, trois simples et quatre divisés, mais qu'elle en diffère par la supériorité de sa taille, par sa tête plus longue, par ses oreilles presque doubles en longueur, par l'absence d'un collier etc, ce qui nous permet de conclure qu'il s'agit réellement d'une espèce inédite.

Cynonycteris torquata Dobson, 1878

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Cynopterus collaris, Gray (non Geoffroy), *Catal. Monkeys and Fruit-eating Bats*, p. 123 (1870).

Slightly more than half the size of *C. collaris*. The head also is comparatively broader and shorter, and the crown of the skull more elevated above the face-line.

The male has the under surface of the neck clothed with remarkably long, densely set yellow hairs, which extend outward upon the shoulders and backwards upon the breast, terminating abruptly, forming a broad well-defined ruff, very distinct from the short fur of the body (Plate V. fig. 2).

Above, half the forearm is covered with short hairs, and the wing-membrane as far as a line drawn from the middle of the humerus to the knee; nearly the whole of the interfemoral is thickly covered, but the lower third of the tibiae and the legs are quite naked. Beneath, a few fine hairs appear upon the antebrachial membrane and upon the wing-membrane between the humerus and the femur and behind the elbow, but no band of fur passes outwards behind the forearm to the carpus.

Above, reddish brown, head paler; beneath, pale brown except the ruff, which is bright reddish yellow.

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Upper incisors arranged in an angular series between the canines; last lower premolar placed on the commencement of the ascending ramus of the jaw, so that, although much smaller, its crown is higher than that of the antepenultimate molar.

In the following columns are given the measurements of an adult male (a dried specimen) and of a younger male (the type, preserved in alcohol) :

Length, head and body: -;	3.5
" Tail: -;	0.35
" head: -;	1.3
Distance of the eye from the nostril: -;	0.45
Length, ear: 0.6;	0.6
" forearm: 2.3;	2-.5
" thumb: 0.85;	0.85
" second finger: 1.7;	1.5
" third finger, metacarpal: 1.55;	1.35
" " 1st phalanx: 1.05;	1.0
" " 2nd " 1.55;	1.4
" fifth finger, metacarpal: 1.5;	1.35
" " 1st phalanx: 0.7;	0.65
" " 2nd " 0.85;	0.7
" tibia: 0.85;	08.8
" foot: 0.5;	0.5

Hab. Africa (Angola).

A. (type) M, al. (not full-grown). Angola. Dr. J. E. Gray.

B. ad. M sk. Africa. Sir J. Richardson.

Cynopterus collaris Gray, 1870

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Fur reddish brown, beneath rather duller; throat with a broad collar of rigid unctuous red hairs; hairy part of back rather narrow; shoulders and lower half of forearm hairy; forearm-bone $2\frac{1}{4}$ inches. Young.

Xanthopyia collaris, Gray, List. Mamm. B. M. 1843.

Hab. West Africa, *Currer* (from Haslar Museum); Angola, *Wellwich* (in spirits).

See also *Ptenochirus*, a subgenus of *Cynopterus*, Peters.

Doryrhina Peters, 1871

A redescription of the genus is provided by Foley *et al.* (2017: 12).

Dysopes brachypterus Peters, 1852

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Tafel XV. Fig. 1.

D. ferrugineofuscus, gastraeo medio griseo; auriculis multo latioribus quam altis, connatis; labris plicatis, crassis, in margine glabris; cauda ultra medium involuta; calcaribus brevioribus.

Longitudo tota 0,090; *capitis* 0,026; *caudae* 0,029; *antibrachii* 0,037; *volatus* 0,230.

Habitatio: Africa orientalis, Insula Mossambique, 15° Lat. Austr.

Diese Art ist der vorhergehenden sehr ähnlich. Die Ohren haben fast dieselbe Gestalt wie bei *Dysopes limbatus*, doch bildet der vordere Theil der Helix einen deutlichen Lappen, und der Tragus ist am Ende abgerundet. Die Lippen sind weniger faltig und am Rande ungekerbt, glatt. Der Kopf und Körper sind gröfser und stärker, die Flughäute dagegen kürzer, ebenso der Schwanz und die Spornen. Die Behaarung des Körpers ist sehr kurz, am Vorderhalse langer, und bekleidet den angrenzenden Theil der Seitenflughäute; die Schenkelflughaut ist aber weniger stark behaart, und umfaßt den Schwanz weit über die Hälfte bis zum viertletzten Gliede. Die Phalangen des dritten Fingers sind kürzer als das Mittelhandglied desselben.

Die Farbe des Rückens und der Seiten des Halses, der Brust und des Bauches ist dunkel rostbraun, die ganze Mitte der Bauchseite dagegen vom nackten Kinn bis zum After zeigt eine graue Färbung. Sammtliche Haare sind an der Basis und Endspitze blasser, weißlich, indem entweder das Braune, wie am Rücken und an den Seiten des Bauches, oder das Weiße, wie an der Mitte der Unterseite, überwiegend ist; einige Haare der Kehle sind ganz weiß. Die Flughäute und Ohren sind von derselben Farbe, aber dunkler; die Nägel bräunlich weiß.

Der Schädel ist viel gröfser als der von *D. limbatus*. Die Zahl der Zähne ist dieselbe, $3^2/_{3,2}1^1/_{1,4}1^1/_{1,2}3/_{2,3} = 30$; der letzte obere Backzahn ist aber nur halb so grofs wie der vorhergehende, so dafs seine Schmelzfalte V förmig ist, während dieselbe bei *D. limbatus* ein schief gestelltes N bildet. Ebenso ist der hinterste untere Backzahn weniger vollständig als bei *D. limbatus*. Die Form des Skelets, die Zahl der Wirbel und Rippen, so wie der Bau der Eingeweide ist Ganz wie bei der vorhergehenden Art. Den Blasenhals umfaßt eine grofse Prostata ebenso wie bei den Mannehen von *D. limbatus*; die Testikel, obwohl sehr entwickelt, liegen aber nicht aufserhalb sondern innerhalb der Bauchhöhle.

Ein einziges Exemplar dieser Art, ein Männchen, fand ich in einem Kamine in Mossambique.

Mafse in Millimetern.

Ganze Länge von der Schnauze bis zur Schwanzspitze: 90
Flugweite: 230

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Länge des Kopfes: 26

Höhe des vorderen Ohrandes: $7\frac{1}{2}$

Breite des ganzen Ohres: 15

Länge des Oberarms: 22

Länge des Vorderarms: 37

Länge des Daumens (Mittelh. 3. 1.Gl. 3. 2.Gl. $2\frac{1}{2}$): $8\frac{1}{2}$

Länge d. 2ten Fingers (Mittelh. 34): 34

Länge d. 3ten Fingers (Mittelh. $36\frac{1}{2}$. 1.Gl. 15. 2.Gl. 12. 3.Gl. $4\frac{1}{2}$): 70

Länge d. 4ten Fingers (Mittelh. 35. 1.Gl. 12. 2.Gl. $8\frac{1}{2}$. 3.Gl. $1\frac{1}{4}$): $56\frac{3}{4}$

Länge d. 5ten Fingers (Mittelh. 22. 1.Gl. $3\frac{1}{4}$. 2.Gl. $3\frac{1}{4}$. 3.Gl. $1\frac{1}{4}$): $35\frac{1}{2}$

Länge des Oberschenkels: 14

Länge des Unterschenkels: 12

Länge des Fusses mit den Krallen: $8\frac{1}{2}$

Länge des Schwanzes: 29

Länge des freien Theils desselben: $11\frac{1}{2}$

Länge des Sporns: 10

Länge des Schädels: $21\frac{1}{2}$

Länge der Wirbelsäule vom Atlas bis zum ersten Schwanzwirbel: 47

Obgleich diese Art mit der vorhergehenden große Ähnlichkeit hat, so ist sie doch durch die Grösse des Schädels, durch die grössere Länge der Schenkelflughaut, durch die Kürze der Spornen, durch die Form einiger Theile des Ohres und endlich durch die Färbung zu sehr von ihr verschieden, um mit derselben vereinigt werden zu können.

Dysopes dubius Peters, 1852

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Tafel XV. Fig. 2.

D. priori similis, sed multo major.

Habitatio: Africa orientalis, Sena, 17° Lat. Austr.

Obgleich ich von dieser Art nur ein einziges ganz junges Exemplar zur Vergleichung benutzen kann, so geht doch daraus hervor, dass sie sich nicht mit einer der bekannten vereinigen lasse. Die Ohren sind viel breiter als hoch, über der Stirn mit einander verwachsen, und die Lippen sind faltig, wie bei den vorhergehenden Arten. Durch die Form der Ohren unterscheidet sie sich auch sogleich von den übrigen africanischen Arten. Die Füße sind von bedeutender Grösse, 3 Mm. länger als die von *D. brachypterus*, woraus, ebenso wie aus den Verhältnissen des Schädels hervorgeht, dass das ausgewachsene Thier viel grösser ist, als *D. brachypterus* oder *D. Geoffroyi*.

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Am Schädel ist besonders zu bemerken, dass die Nasenbeine nur aus einem einzigen Knochenstücke bestehen, dass sich zwischen den grossen Scheitelbeinen und der Hinterhauptsschuppe zwei quadratische Ossa interparietalia befinden, und dass die Pars mastoidea des Schläfenbeins einen besondern Knochen bildet. Von den bleibenden Zähnen sind noch keine zum Durchbruch gekommen: jeder Zwischenkiefer trägt zwei feine conische, an der Spitze nach aufsen gekrümmte Schneidezähne, und in jedem Oberkiefer steht ein ganz ähnlich geformter Wechsel-Eckzahn. Im Unterkiefer stehen sechs feine zweilappige grade Schneidezähne und zwei Eckzähne, welche ihre hakenförmige Spitze nach aufsen wenden.

Dieses Thier wurde mir in Sena von einem Neger gebracht, welcher es im Walde gefunden haben wollte.

Dysopes hepaticus Heuglin, 1864

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Minor; supra cinereo-fuscus; gastraeo medio albido, reliquo lateribusque laete hepatico-brunneis, hypochondriis magis cinerascens; mento et gula nudiusculis, carnicoloribus; auriculis, facie et unguibus fuscis; cauda nigricante; patagiis pallide fuscis, pellucidis, subtus basin versus coerulescentibus.

Labris rugosis, hispidis; auriculis latis trapezoidiformibus, operculatis, margine superiore replicatis, fronte connatis, basi anteriore lobulo accessorio rotundato et trago minore arrecto, acutiusculo instructis; patagio anali plicato, tertiam partem caudae circumdante; cauda crassiuscula, dimidii corporis longitudine.

Long. corporis ad basin caudae 2" 10''' - caud. 1" 5''' - extens. patag. 13" - antibrach. 1" 9''' -

Obiges die Maasse eines alten F. Die MM scheinen constant kleiner, bei einigen ist die ganze Oberseite des Körpers fein weiss gescheckt, bei andern das Gesicht ganz schwärzlich.

Die Nasenlöcher stehen etwas seitlich und sind stumpf röhrenförmig aufgetrieben; die wulstige Oberlippe mit 7 - 8 Querfalten; die Behaarung des Gesichtes rauh. Das verhältnissmässig grosse Auge liegt in zwei tiefen Furchen nahe an der vordern Ohrbasis. An der hintern Basis des kurzen, breiten unregelmässig viereckigen Ohres ist ein grösserer häutiger Ansatz, der umgeschlagen den Gehörgang schliesst; vor der Basis des Vorderrandes des Ohres ein kleiner rundlicher, beweglicher Lappen, der sich an der Oberlippenwulst hinter dem Mundwinkel anschliesst.

Im Alter $\frac{1}{1}$, in der Jugend $\frac{1}{2}$ Schneidezähne. Die obern kräftig, mit der Spitze etwas convergirend, die untern mit unbewaffnetem Auge kaum sichtbar.

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Die obern Eckzähne vorne einfach gefurcht, die untern mit stark divergirenden Spitzen und einem spitzigen, nebensahn-artigen Absatz auf der vordern, innern Seite der Basis der Krone.

Im Gaumen 5 - 6 undeutliche Falten.

Auf dem äussersten Glied der ersten und fünften Zehe und am Daumen einige lange, rauhe, die Nägel weit überragende weissliche Borsten.

Die Flughäute fast ganz nackt.

Diese hübsch gefärbte Art lebt gesellschaftlich in dürrer, dichtem Laub auf unersteiglichen Doleb-palmen (*Borassus aethiopiensis*) im Lande der Req-Neger bis zum Djurfluss. Selten sieht man sie bei Tage fliegen, doch ist sie immer munter und sieht bei grellem Sonnenlicht.

Dysopes limbatus Peters, 1852

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[The poor quality of the copy might have resulted in some errors in the text below - Eds.]

Tafel XIV.

D. nigrofuscus, subtus pallidior, ventre medio lateribusque albis; auriculis multo tatoribus quam altis, connatis; labris crassiss plicatis; cauda ultra medium libera; patagio anali piloso.

Longitudo tota 0,095; capitis 0,022; caudae 0,034; antibrachii 0,037; volatus 0,260.

Habitatio: Africa orientalis, Insula Mossambique, Sena, a 15° ad 17° Lat. Aust.

Eine der kleinsten Arten dieser Gattung. Die Ohren ragen mit ihrem vordem Ende bis in die Mitte zwischen Auge und Schnauzenspitze, sind niedrig, breit, auf der Stirn durch eine breite Hautwulst vereinigt, am innern Rande der Helix und an der Basis dicht behaart der untere Theil der Helix ist schmal, und läßt keinen besondern Lappen erkennen; der Tragus ist sehr klein, eckig, am obern Rande fast grade abgeschnitten. Die Haare auf den Köpfe hinter der Vereinigung der Ohren sind länger und steifer als am übrigen Körper. Die Schnauze ist breit, stumpf, niedergedrückt, un vorn schräg von oben nach unter und hinter eingeschr..en. Die Oberlippe ist dick, .. am Rande ge... und mit kurzen steifen Haaren besetzt. Die Behaarung des Körpers ist sehr fein und weich, am Rücken und Bauch gleich lang, an der Vorderseite des Halses ein wenig länger, und geht zu beiden Seiten in einem breiten Saum auf die Flughäute über. Diese letzteren sind schmal

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zwischen dem Oberarm und Schenkel mit punctförmigen Rüschelein von Härchen besetzt. Nur das Mittelhandbied des Daumens wird von der Halsflughaut umfaßt. Vor dem Gelenke zwischen dem Mittelhand und dem ersten Gliede des Daumens befindet sich eine hornartige nackte Hautscheibe. Die Glieder des dritten Fingers zusammen sind so lang oder länger als die Mittelhand desselben. Die Flughäute gehen nicht ganz bis zum Ende des Unterschenkels herab. Der Schwanz ragt mit seiner größeren Endhölle aus der Schenkelflughaut hervor, welche der Quere nach gefaltet und sowohl oben wie unten zur Hälfte behaart ist.

Die Farbe des ausgewachsenen Männchen Fig 1., ist auf dem Rücken dunkel schwärzlich röstbraun, an der Bauchseite etwas blasser, die Mitte und die Seiten des Unterleibs so wie das ... weifs. Die Ohren sind schwarzbraun; die Flughäute neben den Körperseiten braun, die übrigen Theile derselben durchscheinend bräunlich. Die braune Theile viel heller rostfarbig. Die Rückenhaare haben eine blasser weifsliche Basis; die braunen Haare der Bauchseite zeigen ausserdem hellere Spitzen. Die Nägel sind brünnlich, am convexen Rande dunkler gefärbt.

Zähne im erwachsenen Zustande $3\frac{2}{3}2\frac{1}{1}1\frac{1}{1}1\frac{2}{2}3\frac{3}{2}3 = 30$. Die jungen Thiere zeigen vor ihren beiden oberen bleibenden Vorderzähnen die vier noch nicht ausgestoßenen Wechselzähne (Fig. 3a.). Die Wirbelsäule wird aus 41 Wirbelkörpern gebildet; darunter sind 7 Halswirbel, 13 Rückenwirbel, 6 Lendenwirbel, 5 Kreuzbeinwirbel, 10 Schwanzwirbel. Das Brustbein besteht aus drei Knochenstücken: der Kiel desselben ist sehr schwach; es verbinden sich sechs Paar Rippen mit demselben. Der zweite Finger hat eine Phalanx, der Mittelfinger drei, und eben so der vierte und fünfte Finger, bei denen aber das letzte Glied kurz umgebogen ist. Die Fibula ist vollständig entwickelt. Die Schleimhaut des Gaumens zeigt sechs Querfalten. Die Papillen der vordern Hälfte der Zunge sind platt und schuppenförmig; auf der hintern Hälfte sind sie grösser, zerstreuter, warzenförmig; nahe der Basis finden sich mehrere in ringförmigen Vertiefungen, von denen zwei durch ihre Grösse sich auszeichnen. Zu jeder Seite am Halse liegt eine grosse 7 Millim. im Durchmesser haltende Speicheldrüse. Der Oesophagus mündet nahe unter dem Zwerchfell in einen grossen sackförmigen Magen, welcher in einen nur anfangs etwas weiteren 130 Mm. langen Darm übergeht. Die Leber liegt mit ihrem grösseren Theil in der rechten Seite, hat einen mittleren schwachen, einen rechten tiefen Einschnitt; in letzterem liegt die Gallenblase. Die Milz ist 15 Mm. lang, am breitesten Theile 4 Mm. breit. Die Nieren sind einfach, bohnenförmig, 6 Mm. lang, 4 Mm. breit; die linke liegt viel tiefer als die rechte. Die Blase ist dünnwandig, und wird, bei dem Männchen, an ihrem Halse oben und an den Seiten von einer halbmondförmigen 6 Mm. im Durchmesser haltenden Prostata umgeben. Die Ruthe ist ganz weich, ohne innere Skelettheile; neben ihrer Wurzel liegen die 5 Mm. langen Hoden, und treiben die Haut so hervor, daß die äusseren Genitalien mit den menschlichen eine aufserordentliche Ähnlichkeit haben. Der Kehlkopf ist 5 Mm. lang. Die Luftröhre, welche keine Anschwellungen zeigt, enthält 24 knorpelige Halbringe. Die rechte

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Lunge zeigt zwei kleine Nebenlappen; die linke ist einlappig. Das Herz ist länglich, 10 Mm lang, 5 Mm. breit.

Diesen Grämmler erhielt ich auf der Insel Mossambique und in Sena aus dunklen Räumen der Wohnungen. Der Inhalt des Verdauungscanals verschiedener Individuen liefs Theile von Insecten erkennen.

Masse in Millimetern.

Mas.; Fem.

Ganze Länge von der Schnauze bis zur Schwanzspitze: 95; 80
 Flugweite: 260; 230
 Länge des Kopfes: 22; 20
 Höhe des Ohres am vordern Rande: 8; $7\frac{1}{2}$
 Breite des Ohres: 15; 13
 Länge des Oberarms: 21; 21
 Länge des Vorderarms: 37; 35
 Länge des Daumens (Mittelh. 3, 3. 1.Gl. 3, 3. 2.Gl. 2, 2): 8; 8
 Länge d. 2ten Fingers (Mittelh. 35, $32\frac{1}{2}$, 1.Gl. $1\frac{1}{2}$, $1\frac{1}{2}$): $36\frac{1}{2}$; 34
 Länge d. 3ten Fingers (Mittelh. $38\frac{1}{2}$, 35, 1.Gl. $16\frac{1}{4}$, 14, 2.Gl. $16\frac{1}{4}$, 13, 3.Gl. 8, 7): $79\frac{1}{2}$; 69
 Länge d. 4ten Fingers (Mittelh. $37\frac{1}{2}$, 34, 1.Gl. $14\frac{1}{2}$, 11, 2.Gl. $11\frac{1}{4}$, 9, 3.Gl. $1\frac{1}{2}$, $1\frac{1}{2}$): $64\frac{3}{4}$; $55\frac{1}{2}$
 Länge d. 5ten Fingers (Mittelh. 23, 22, 1.Gl. 11, 10, 2.Gl. 4, 3, 3.Gl. 1, 1): 39; 36
 Länge des Oberschenkels: $13\frac{1}{2}$; $12\frac{1}{2}$
 Länge des Unterschenkels: 12; $10\frac{1}{2}$
 Länge des ganzen Fusses mit den Krallen: 8; 8
 Länge des Schwanzes: 20; 15
 Länge des freien Endes desselben: 20; 15
 Länge des Sporns: 13; 12
 Länge des Schädels: $16\frac{1}{2}$; 16
 Länge der Wirbelsäule ohne die Schwanzwirbel: 43; 36

Nyctinomus aegyptiacus Geoffr. (*Dysopes Geoffroyi* Temm.) hat am meisten Ähnlichkeit mit dieser Art, ist aber viel größer (Vorderarm bei dem jungen Thier 50 Mm.), und auch durch die Form und Stellung der oberen Schneidezähne, welche kürzer sind und nahe beisammen stehen, verschieden. *Dysopus pumilus* Rüpp. unterscheidet sich von ihr durch größere Länge der Ohren und durch verschiedene Färbung der Bauchseite. *Dysopes midas*, Hedenborg, und *Dysopes Rüppellii* Temm. (*Dysopes Cestonii* Savi) sind durch ihre viel bedeutendere Größe, durch viel höhere Ohren, erstere auch durch die nackten Flughäute leicht von ihr zu unterscheiden. *Dysopes natalensis*, Smith, die einzige bisher bekannte südafrikanische Form dieser Gattung, ist oben und unten braun gefärbt, hat dreieckige zugespitzte Ohren, glatte Lippen, und eine viel breitere Schwanzflughaut, so daß sie in keiner Weise mit unserer Art zu verwechseln ist.

Dysopes pumilus Cretzschmar, 1826

p. 21
 Zwerg-Grämmler

Diagnos. *Dysopes corporis colore ex fusco nigrescente supra obscuriori infra dilutiori.*

Ausmessungen.

Fuß. Zoll. Lin.

Länge des Körpers von dem Scheitel bis zu dem Schwanz: - 1 9

Länge des Schwanzes: - - 10

Flügelweite: - 7 6

Höhe der Ohren: - - 6

Länge des Kopfes: - - 6

Beschreibung

Das Haar äußerst fine und wollig, nicht sehr lang aber dicht. Die Farbe auf den oberen Theilen des Körpers schwarzbraun, auf den unteren heller. Die Flügel- und Zwischenschenkel-Membranen schwarzbraun. Dieselbe Färbung an den Ohren. Diese mit starken wulstigen Knorpeln versehen, haben einen kleinen $\frac{1}{2}$ Linie langen Ohrdeckel (*operculum*), welcher bei dem nach dem Gesicht sehr vorliegenden Ohre nicht gesehen werden kann, ohne dass dasselbe nach hinten zurück gebogen worden, und welchen wir hier deswegen anführen, da wir ihn bei allen aus Afrika erhaltenen Grämlern vorfinden. Er ist von dem eigentlichen Ohrläppchen (*auricula*) bedeckt **). Im übrigen ist dieser Grämmler seinen Formen nach ein Miniaturbild von dem *Dysopes Rüppellii*, welchen Temminck in den *Monographies de mammalogie* pag. 224 beschrieben und planche XVIII abgebildet hat, weswegen wir in Hinsicht der Charakteristik dieses Geschlechtes als auch unserer hier bezeichneten Art im allgemeinen auf die vorzügliche Bearbeitung derselben hinweisen. Die kleinen schwarzblauen Augen sind von dem Ohre fast ganz bedeckt.

Vaterland. Massauah.

Aufenthalt. Alte verödete Wohnungen.

**) Geoffroy hat diesen Ohrdeckel nicht angegeben. Temminck dagegen desselben erwähnt, aber die *auricula* als eine doppelte, eine äußere und eine innere angeführt, während diese Theile ihrem Baue und ihrer Bedeutung gemäß, wie schon Kuhl angegeben, verschieden sind.

Eleutherura unicolor Gray, 1870

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Fur soft, uniform grey brown, rather paler below; hair one-coloured; ears large; wings pale, with white hairs near margin; forearm 3 inches 8 lines; tail very short; false grinders in each jaw well developed, lower largest.

Hab. West Africa: Gaboon, Verreaux.

See *Pteropus mollipilosus*, Allen, Proc. Acad. N. Sci. Philad. 1861, p. 159. Head small; ears large; tail very small, free; fur olive-brown, brighter on the back of the neck. *Hab.* Western Africa (Gaboon), *Du Chaillu*. "The intermaxillary bone of the skull is not thrown forwards and downwards as in other *Pteropi*, but is on the same level as the roof of the mouth, the first upper premolar larger than the incisors, the same tooth of lower of a good size and less tubercular than in other species." - Allen. Perhaps male of preceding.

Emballonura afra Peters, 1852

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Tafel XII. Tafel XIII. Fig. 18. 19.

E. ferrugineofusca, *gastraeo pallidior*, *auriculis triangularibus rotundatis*, *trago mediocri*; *rostro prominente*, *apice bifido*; *dentibus primoribus superioribus duobus*; *phalangibus primis digiti quarti et quinti duplo longioribus quam secundis*.

Longitudo tota 0,080; *caudae* 0,020; *antibrachii* 0,050; *volatus* 0,310.

Habitatio: Africa orientalis, Tette.

Von der Gröfse unserer *Vespertilio discolor*. Die Ohren stehen weit von einander getrennt, sind um ein Viertel kürzer als der Kopf, von dreieckig abgerundeter Gestalt, und nur an der Basis und am Längskiel behaart. Ihr vorderer grader Rand steht mitten über dem Auge, ihr hinterer convexer, unten schwach ausgeschnittener Rand setzt sich geschwungen bis nahe hinter und unter dem Mundwinkel fort. Es lassen sich an ihnen neun, oben gedrängter stehende, Querfalten unterscheiden. Der Tragus ist frei, doppelt so lang wie breit, an seiner Basis hinten mit einer kleinen nach vorn umgeschlagenen Spitze versehen, am Rande behaart. Die Augen sind gröfser als sie gewöhnlich bei den Thieren dieser Ordnung erscheinen, und haben ihre Lage in der Mitte zwischen dem vorderen Ohrande und dem Mundwinkel. Die Plica semilunaris in ihrem innern Winkel ist sehr entwickelt. Die Nase tritt oben doppelröhrig hervor, und bildet den am meisten vorspringenden Theil der Schnauze; sie ist zu jeder Seite schräg abgestutzt, so dafs die Nasenlöcher unter einem stumpfen Winkel

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von einander abgewandt sind. Das Maul ist bis unter die Augen gespalten. Die Oberlippe ragt mit ihrem breiten platten Rande über die Unterlippe hervor, ist mit steifen über der Mundrand gerichteten Haaren besetzt, jederseits an ihrer innern Oberfläche durch einen warzigen Vorsprung, und vor der Mitte durch eine platte Erhabenheit ausgezeichnet, welche in eine mittlere Vertiefung der Unterlippe eingreift, deren Schleimhaut sich hier mit einer dreieckigen breiten Platte nach aufsen umschlägt. Eine hintere Erweiterung der Backen in taschen förmige Vertiefungen ist nicht vorhanden. Die Gaumenhaut bildet sechs quere Falten; die erste derselben ist kurz und liegt mitten zwischen den Eckzähnen; die zweite ungetheilte liegt zwischen dem vordern Ende der beiden grofsen Lückenzähne; die dritte und vierte sind in der Mitte getrennt und nehmen den Raum zwischen dem ersten Paar der grofsen Backzähne ein; die fünfte ist ebenfalls in der Mitte getheilt, und bildet einen nach vorn stark convexen Bogen, so dafs ihre äufseren Enden mit der letzten graden Querfalte zwischen und an dem hintersten Theile des zweiten Paares der wahren Backzähne zusammenstofsen. Die Behaarung des Körpers ist dicht und weich, vor der Brust und dem Halse etwas länger als an den übrigen Theilen, wo man sie eher kurz nennen kann; sie geht in einem breiten Saum auf die Seitenflughäute über, an der Bauchseite etwas weiter aber lichter als an der Rückenseite; ebenso setzt sie sich auf der obern Seite der Schenkelflughaut bis zur Durchbohrungsstelle des Schwanzes fort. Die Flughäute sind lang und schmal, schliefsen von Daumen nur das Mittelhandglied ein, und gehen nicht über das untere Ende des Unterschenkels herab; die Rückenseite der Halsflughaut ist leicht von kurzem wolligen Haar bedeckt, und auf den erhabenen Linien zwischen Arm und Schenkel lassen sich Reihen kurzer Härchen wahrnehmen. Der Oberarm hat die Länge des Kopfes; der Unterarm ragt neben dem Körper gelegt weit über die Spitze der Schnauze hervor, und ist länger als die Entfernung der Schnauzenspitze vom After. Die Mittelhand des Daumens ist ebenso lang wie das erste Glied desselben; die Haut vor dem Gelenke zwischen diesen Gliedern bildet eine kleine nackte Scheibe. Der zweite Finger erreicht lange nicht das Ende der Mittelhand des dritten Fingers, und diese ragt doppelt so weit über das Mittelhandglied des vierten Fingers wie dieses über dasselbe Glied des fünften Fingers hinaus. Die erste Phalanx des vierten und fünften Fingers ist doppelt so lang wie die zweite. Die Beine sind im Verhältnifs zur obern Extremität nur kurz, indem sowohl Ober- wie Unterschenkel kaum ein Drittheil so lang wie der Vorderarm sind. Die Füfse dagegen sind ziemlich lang, länger als die Hälfte des Unterschenkels: die Zehen ragen alle fast gleich weit vor, doch ist die äufseren und innere ein wenig kürzer als die drei mittleren gleich langen. Die Krallen der Zehen sind etwas länger als die Daumenkrallen. Der Schwanz und die Spornen sind ungefähr so lang wie der Unterschenkel. Die Schenkelflughaut ist am Endrande schwach bogenförmig ausgeschnitten, wird ungefähr in ihrer Mitte oben vom Schwanzende, welches sich wie in eine Scheide zurückziehen kann, durchbohrt, und zeichnet sich durch zahlreiche Querfalten aus, welche an der Bauchseite mit weifsen Härchen bewimpert sind.

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Die Farbe des ganzen Thieres ist braun, rauchbraun, chocoladenbraun oder, wie in den meisten Fällen, dunkelrostbraun, an der Bauchseite etwas heller. Die einzelnen Haare sind auf der Rückenseite und dem Kopfe einfarbig braun bis auf den ganz

kurzen weiflichen Grundtheil, die Bauchhaare haben aufser dem hellen Grundtheile auch noch ganz kurze blasse Spitzen. Die Flügel und Ohren sind von derselben braunen Farbe, aber etwas dunkler. Die Nägel sind bräunlich weifs.

Der Schädel (Tafel XIII. Fig. 18. 19) zeigt ganz die der Gattung *Emballonura* eigenthümliche Form und die Entwicklung der Processus zygomatici des Stirnbeins, wie wir sie auch bei den *Taphozous* wiederfinden. Zähne: $\frac{3}{2}/\frac{1}{3} \frac{1}{1} \frac{1}{6} \frac{1}{1} \frac{2}{3}/\frac{2}{3} = 32$. Bei mehr als hundert Exemplaren fand ich stets nur zwei obere Schneidezähne, und nach dem Bau der Zwischenkiefer zu urtheilen, ist auch nur für einen einzigen Zahn in jedem derselben Platz. Diese beiden Zähne stehen weit auseinander, mit den inneren lang ausgezogenen Spitzen convergirend. Die unteren sechs Vorderzähne stehen in der Richtung des Kiefferrandes und sind dreilappig. Die oberen Eckzähne sind gekrümmt, sowohl vorn wie hinten mit einem deutlichen Absatz versehen. Gleich hinter ihnen liegt ein kleiner, rundlicher, einwurzeliger Lückenzahn, der nicht über den hinteren Absatz des Eckzahns herabragt, und an seiner Krone, genau betrachtet, dreispitzig ist. Hierauf folgt durch eine Lücke getrennt ein großer zweiwurzeliger Lückenzahn, welcher in seiner Form der ersten Hälfte des daran stoßenden ersten wahren Backzahns entspricht. Dieser letztere ist nicht ganz so groß wie der folgende vierte, welcher allein eine vollständig W förmige Schmelzfalte besitzt, und daher aufsen drei, innen zwei Spitzen, und oben an seiner Basis noch zwei durch die Entwicklung des Cingulums gebildete Höcker zeigt. Der fünfte und letzte obere Backzahn ist nur halb so groß wie der vierte. Die unteren Eckzähne sind von der Gestalt der oberen, aber beträchtlich kleiner. Der erste Lückenzahn des Unterkiefers ist einwurzelig, doppelt so groß wie der entsprechende obere, mit niedriger, unregelmäßig dreikantiger Spitze. Der zweite Lückenzahn ist zweiwurzelig, etwas länger und breiter, um das Doppelte höher, und von der Gestalt des Eckzahns. Die drei folgenden wahren Backzähne sind viel schmaler als die entsprechenden oberen, sämtlich fünfspitzig mit nach innen offenstehender W förmiger Schmelzfalte.

Die Wirbelsäule wird von 39 Wirbelkörpern zusammengesetzt; hiervon sind 7 Halswirbel, 14 Rückenwirbel, 5 Lendenwirbel, 7 Kreuzbeinwirbel und 6 Schwanzwirbel. Mit dem Brustbein, welches nur am Manubrium gekielt erscheint, verbinden sich 8 Paar Rippen, die übrigen 6 Paare sind falsche. Der Unterschenkel zeigt eine vollständig entwickelte Fibula.

Die Zunge ist mit feinen schuppenartigen Papillen bedeckt, welche nach der Mitte hin am Rande gestachelt erscheinen; an der Basis stehen größere knopförmige Papillen und zwei von einer ringförmigen Vertiefung umgebene Warzen. Der Oesophagus geht sogleich unter dem Zwerchfell in einen einfachen kleinen rundlichen Magen über, welcher durch eine schwache Pförtner-Einschnürung vom Darm abgesetzt ist. Dieser letztere ist anfangs ein wenig erweitert, und hat eine Länge von 150 Mm. Die Leber liegt mit ihrem größten Theile

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in der linken Seite, und hat zwei Einschnitte; unter dem rechten liegt eine ziemlich große Gallenblase. Am Fundus des Magens liegt die Milz, ein schmaler, platter Körper von 10 Mm Länge. Die Nieren sind ungelappt, bohnenförmig, 6 Mm. lang und 4 Mm, breit. Die Harnblase ist dick und musculös. Der Uterus geht in zwei lange Hörner aus. An den Seiten der Brust liegt ein einziges Paar von Zitzen; am Bauche ist keine Spur von zitzenähnlichen Vorsprüngen vorhanden. Die Ruthe des Männchens ist ganz weich, ohne Knorpel oder Knochen zu beiden Seiten vor den Schambeinen liegen die platten, rundlichen Hoden außerhalb der Bauchhöhle. Der Schildknorpel des Kehlkopfs bildet einen schmalen Halbring, den man leicht mit dem Zungenbein verwechseln könnte, von dessen hinteren Enden ein kurzer obere Fortsatz zur Verbindung mit dem großen Horn des Zungenbeins, ein sehr langer unterer zur Verbindung mit dem Ringknorpel abgeht. Der Ringknorpel ist dagegen sehr lang, an seinen hintern Fläche mit einer sehr hohen Leiste versehen, mit welcher sich oben die Giefsbeckenknorpel verbinden. Die Luftröhre besteht bis zur Theilungsstelle aus 17 knorpeligen Halbringen; ihr häutiger Theil bildet im ersten Drittel eine große spindelförmige Erweiterung. Die Lungen bestehen jederseits aus einem einzigen Lappen. Das Herz ist breit und abgerundet, 7 Mm. lang, 5 Mm. breit.

Ich fand diese erste africanische Art der Gattung *Emballonura* in dunkeln Kellerräumen in Tette. Sie lebt in großen Gesellschaften beisammen, in denen die Zahl der Männchen überwiegend zu sein scheint, indem ich unter hundert Exemplaren nur zwanzig Weibchen fund. Im Darm fand sich eine breiartige Masse mit Schalenrudimenten von Insecten vermischt.

Mafse in Millimetern.

Fem.; Mas.

Ganze Länge von der Schnauze bis zur Schwanzspitze: 80; 75

Flugweite: 310; 300

Länge des Kopfes: 22; 22

Entfernung des Auges von der Spitze der Schnauze: 8; 8

Länge des ganzen Ohres: 17; 17

Breite des Ohres: 12; 12

Länge des Tragus: 6; 6

Breite desselben: 2; 2

Länge des Oberarms: 27; $26\frac{1}{2}$

Länge des Unterarms: 50; 49

Länge des Daumens (Mittelh. $4\frac{1}{2}$, $4\frac{1}{2}$. 1.Gl. $4\frac{1}{2}$, $4\frac{1}{2}$. 2.Gl. $2\frac{1}{2}$, $2\frac{1}{2}$): $11\frac{1}{2}$; $11\frac{1}{2}$

Länge d. 2ten Fingers (Mittelh. 44, 41): 44; 41

Länge d. 3ten Fingers (Mittelh. 48, 41. 1.Gl. $16\frac{1}{2}$, 16. 2.Gl. $19\frac{1}{2}$, 19): 84, 79

Länge d. 4ten Fingers (Mittelh. 37, 35. 1.Gl. $12\frac{1}{2}$, $11\frac{3}{4}$. 2.Gl. $6\frac{1}{4}$, $5\frac{3}{4}$): $55\frac{3}{4}$; $52\frac{1}{2}$

Länge d. 5ten Fingers (Mittelh. $29\frac{1}{2}$, 29. 1.Gl. 13, $12\frac{1}{2}$. 2.Gl. 6, 5): $48\frac{1}{2}$; $46\frac{1}{2}$

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Länge des Oberschenkels: 16; 16

Länge des Unterschenkels: $15\frac{1}{2}$; $15\frac{1}{2}$

Länge des Fufses mit den Krallen: 11; 11

Länge des Schwanzes: 20; 17

Länge des Schenkelflughaut: 25; 25
 Länge des Sporns: 16; 16
 Länge des Schädels: 18; 18
 Länge der ganzen Wirbelsäule: 55; 55

Emballonura monticola, Temminck, mit welcher diese Art in der Färbung und Gestalt die meiste Verwandtschaft hat, unterscheidet sich von ihr durch geringere Gröfse, so wie, nach der Temminckschen Abbildung zu urtheilen, durch gröfsere Länge des fünften Mittelhandgliedes, und durch die ziemlich gleiche Länge der Phalangen des vierten und fünften Fingers.

Eomops Thomas, 1905

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In 1900 Dr. Scharff described a bat from Benin under the name of *Mormopterus Whitleyi* and was good enough to transfer the typical specimen to the British Museum.

Later on Dr. W. J. Ansorge obtained on the Lower Niger two examples of the same bat, and in examining these I find that a mistake has inadvertently been made in the dental formula given, and that, instead of being a *Mormopterus*, this

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bat belongs to quite a different group, hitherto unknown in the Old World.

For it proves to have the characteristic incisors of *Molossus*, the upper pair being large, nearly filling up the space between the canines, and touching each other in the middle line. The lower ones are only two in number, not four as originally stated, and while appearing from the front to be of the normal shape - i.e., narrow, broadening upward, each with a deep central notch dividing it into two cusps, of which the inner is the higher - they are curiously deep antero-posteriorly, deeper than broad, their hinder part running back between the canines.

The premaxillary region of the palate is very completely ossified, in great contrast to all the members of the *Nyctinomus* group, the only remnant of the premaxillary notch being two minute foramina on each side of the middle line, between and opposite the centre of the canines.

The skull is smooth and rounded, not broadened and flattened across the muzzle as it is in *Myopterus*, nor specially crested as in *Molossus*. The base of the skull has a narrow median ridge between two well-defined sphenoid pits.

Under these circumstances it is clear that this bat cannot be referred to *Nyctinomus* or *Mormopterus*, but is more nearly allied to *Molossus*, *Myopterus*, and their allies. The peculiar shape of its lower incisors, however, and the detailed structure of its skull prevent its being referred to any one of the existing American genera, and I would therefore propose to erect for its reception a special genus, which might bear the name of *Eomops*.

It is also to be noted that, owing to the bad condition of his specimens, Dr. Scharff described the underside of *Eomops Whitleyi* as "of a light reddish-yellow colour"; but Dr. Ansorge's fresh examples show that this is pure white, as are also the wing-membranes above and below, the digits, and the underside of the interfemoral; the upper surface of the forearms, tibiæ, and interfemoral are pale whitish brown.

Epomophorus (Epomophorus) büttikoferi Matschie, 1899

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1888. *Epomophorus gambianus* Jentink, Not. Leyd. Mus. X. p.50. - 1888. Jentink, Cat, Syst. p. 137c.

Hab. Schieffelinville, Junk River, Liberia.

Bei dem mir zu Gebote stehenden Material von *Epomophorus* variiert die Länge des Unterarms bei Exemplaren, deren letzter oberer Molar vollständig ausgebildet ist, nicht allzusehr. Die Männchen sind allerdings gewöhnlich grösser als die Weibchen, aber bei beiden Geschlechtern bleibt die Variation innerhalb von je 3 - 6 mm. Die alten Männchen von Angola erreichen eine Unterarm-Länge von 93 mm, die kleinsten ausgewachsenen Weibchen von dort haben einen Unterarm von 82 mm Länge. Die Angola-*Epomophorus* sind aber die grössten, welche ich aus der Gattung kenne. Der von Herrn Jentink erwähnte *Epomophorus* von Junk River in Liberia hat eine Unterarm-Länge von 101 mm, also 8 mm mehr als der grösste sonst bekannte *Epomophorus*.

Die Arten von *Epomophorus* s. str. bewohnen beschränkte Gebiete; es ist mir nicht gelungen nachzuweisen, dass zwei Formen dieser Gruppe neben einander in derselben Gegend leben. Wo man derartige Fälle in der Litteratur findet, da darf man stets an der Richtigkeit der Fundortsangabe oder der Bestimmung zweifeln.

Bei Bolama in Portugiesisch-Guinea lebt *Ep. macrocephalus*, an der Goldküste eine sehr ähnliche Form, die ich jetzt als *Ep. zechi* abtrenne. Von Liberia ist nur die von Jentink als *gambianus* bezeichnete Form bekannt. *Ep. gambianus* stammt aber aus den Sammlungen, welche Rendall vom Gambia heimbrachte und wird wahrscheinlich nördlich vom Gambia im Senegal-Gebiet erlangt worden sein. Es ist also von vorn herein sehr wahrscheinlich, dass in Sierra-Leone und Liberia eine neue Art von *Epomophorus* vorkommt. Jentink giebt die Unterarm-Länge für das eine der beiden im Leydener Museum befindlichen

Stücke auf 101 mm an. Diagnose von *Epomophorus büttikoferi* Mtsch. *Epomophorus*, gambiano aff. sed multo major, radii longitudine 93 mm superante.

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Als Typus ist das von Stampfli gesammte M des Leydener Museums von Schieffelinville am Junk River aufzufassen. Ich habe mir erlaubt, diese Art Herrn Direktor Dr. Büttikofer in Rotterdam zu widmen, durch welchen wir die Fauna von Liberia so gut kennen gelernt haben.

Das Original-Exemplar, ein sehr altes M mit schön entwickelten Schultertaschen hat nach Jentink folgende Maasse:

Unterarm: 101 mm

3. Finger, Metacarpale: 75.5

" " 1. Phalanx: 48

" " 2. " : 64.5

5. " Metacarpale: 73

" " 1. Phalanx: 35

" " 2. " : 35

Unterschenkel: 43

Auge bis zur Nasenspitze 30

Ep[omophorus (Epomophorus)] doriae Matschie, 1899

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- 1879. (?) *Ep. labiatus* Dobson, P.Z.S. p.716 - 717. - 1880. (?) Dobson, Rep. Brit. Ass. p. 2 - 3. - 1887. *Ep. labiatus* von Keren, Bogos, Monticelli, An. Mus. Civ. St. Nat. Genova, 2. ser, (V) p. 524.

Epomophorus doriae Mtsch, spec. nov, aff. *Ep. minor*, molaribus angustis, 1.5 mm latis, differt radii longitudine apud F 69.5 mm.

Ich verknüpfe mit diesem Flederhund den Namen des Herrn Marquis Doria, welchem unsere Sammlung so ausserordentlich grosse Förderung zu danken hat.

E. doriae unterscheidet sich von *Ep. labiatus* durch seine schmalen Backenzähne, welche nur 1.5 mm breit sind und durch die kurze Molarenreihe (9.2 mm); er unterscheidet sich von *minor* durch den längeren Unterarm (69.5 mm bei F) und durch die längere Schnauze (15.9 mm vom vordersten Punkte des Augenraudes neben dem Foramen infraorbitale bis zum Gnathion - gegen 14.1 bei einem *E. minor*-Weibchen).

Wie sich *Ep. doriae* zu *Ep. schoensis* verhält, vermag ich vorläufig nicht zu sagen. Von dem Kitimba-Flederhund, mit welchem er in der Länge der Zahnreihe und der Gestalt der Molaren übereinstimmt, unterscheidet er sich durch die grössere Länge des Unterarmes und dadurch, dass bei *doriae* der erste untere Molar viel länger ist als der letzte Praemolar, während bei dem Kirimba-Fledefhund beide Zähne ungefähr gleich lang sind.

Hab. Küste des Rothen Meeres in der Erythraea.

Ich möchte vermuthen, dass die *Ep. labiatus* aus der Robb'schen Sammlung hierher gehören, ebenso wie die Exemplare von Bogos (Beccari) im Museum von Genua.

[B.M.] F Bogos, Gerrard.

Ep[omophorus (Epomophorus)] neumanni Matschie, 1899

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- 1876. *Ep. crypturus* Ptrs. Monatsb. Akad. Berlin p. 913. - 1878. Peters l.c. p. 195. - 1879. *Ep. labiatus* Peters l.c. p. 831.

Epomophorus neumanni Mtsch. spec. nov. alis concoloribus neque ad corporis latera albido tinctis, fasciis in plagiopatagio 23 31; radii longitudo 73 - 78 mm (F), 77 - 80 mm (M).

Hab. Küstengebiet von Britisch Ost-Afrika (Mombasa, Takaungu, Malindi) und Moschi am Kilima-Ndjaru.

Ep. neumanni, den ich Herrn Oscar Neumann widme. unterscheidet sich von allen andern Flederhunden der Untergattung *Epomophorus* mit Ausnahme von *Ep. wahlbergi* und *stuhlmanni* dadurch, dass auf dem Plagiopatagium mindestens 23 Stränge den Hauptstrang durchschneiden; junge Thiere zeigen weniger Stränge, aber immerhin über 20, während junge Exemplare der übrigen Arten höchstens 16 aufweisen. Von *Ep. wahlbergi* und *stuhlmanni* unterscheidet sich *Ep. neumanni* durch die weit geringere Grösse, von *stuhlmanni* auch durch die gleichmässig braun gefarbten, an den Körperseiten nicht weisslichen Flughäute. Die Gaumeufalten sind ungefähr so, wie sie Dobson für *gambianus* abbildet. *Ep. neumanni* sieht

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Ep. zenkeri von Kamerun sehr ähnlich, ist aber kleiner und hat zahlreichere Stränge auf der Flughaut. '

[B.M.] 2 MM, 1 F Mombasa, Hildebrandt; M jun, Malindi, Fischer; F juv. Moschi am Kilima-Ndscharo, Oscar Neumann; M, F Takaungu, Thomas.

Epomophorus (Epomophorus) stuhlmanni Matschie, 1899

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1878. *Ep. gambianus* von Dar es Salaam. Dobson l.c. p. 11. - 1890. (?) *Ep. minor* von Bagamoyo, Thomas, P.Z.S. p. 446. - 1891. *Ep. gambianus* Noack, Jahrb. Hamb. Wiss. Anst. IX. p. 57 - 58. - 1891. *Ep. minor* Noack, l.c. p. 58 - 59. - 1895 *Ep. gambianus* Matschie, Säugethiere Deutsch-Ost-Afrikas, p. 16 Fig. 7 (Kopf). - 1897. Matschie, Arch. Naturg. I. p. 83.

Hab. Küstengebiet von Deutsch-Ost-Afrika und Zanzibar.

Diese Abart hat ebenso wie *wahlbergi* und wie die von Mombas bekannten Flederhunde auf dem Plagiopatagium mehr als 23 Stränge, welche den Hauptstrang durchschneiden. Von *wahlbergi* und dem Mombas-Flederhund lässt sie sich leicht durch ihre gegen die Körperseiten weisslich überflogenen Flughäute unterscheiden; sie ist grösser als der Mombas-Flederhund, aber kleiner als *wahlbergi*. Ihre Diagnose könnte man in dem Satze zusammen fassen:

Epomophorus stuhlmanni Mtsch. spec. nov., alis ad corporis latera albido tinctis, fasciis in plagiopatagio 23 - 30; radii longitudo 85 - 87 mm (M), 80 - 81 mm (F).

In der Gaumenbildung steht diese Abart der Kamerun-Form sehr nahe und dem "gambianus" auf Dobson's Tafel. Noack's *Ep. minor* von Zanzibar sind junge *Ep. stuhlmanni*; ich möchte auch vermuthen, dass die *Ep. minor*, welche Oldf. Thomas von Bagamoyo aufführt, junge *Ep. stuhlmanni* sind. Die grössten *Ep. minor* FF, welche ich kenne, haben einen Unterarm von 63 mm; bei den MM ist der Unterarm 64 - 68 mm lang; ganz junge *Ep. stuhlmanni* haben schon eine Unterarm-Länge von 68 - 69 mm.

Ich nenne diese Abart nach Herrn Regierungsrath Dr. Stuhlmann, dem ich den grösseren Theil der mir zur Verfügung stehenden Exemplare verdanke.

[B.M.] M pull., F ad, Ras Kisimkani auf Zanzibar, Stuhlmann; 2 MM Mojoni auf Zanzibar, O. Neumann; 2 FF pull., 3 FF, 1 M Dar es Salaam, Stuhlmann; M juv, Zanzibar, Wessel; 4 MM, 2 FF Vikindo in Usaramo, Stuhlmann; F Lindi, Füllehorn.

Epomophorus (Epomophorus) zechi Matschie, 1899

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1853. *Pachysoma whitei* partim Temminck, Esqu. Zool. Côte de Guinée p. 65 - 68 (Beschreibung des F p. 67 und des jungen M p. 68). - 1878. *Ep. macrocephalus* partim Dobson l. c. p. 8 - 10, Taf. II. Fig. 2 (Gaumen mit Gebiss). - 1887. *Ep. macrocephalus* Jentink l. c. p. 251. - 1888. Jentink l. c. p. 136. - 1893. Matschie, Sitzb. Ges. naturf. Fr. p. 256 - 257. - 1894. Matschie, Mitth. Geogr. Ges. Naturh. Mus. Lübeck, 2 ser. p. 133. 1897. partim Trouessart l. c. p. 88.

Hab. Accra, Goldküste (Jentink), Gross-Popo, Misahöhe in Togoland (B. M.), Lagos (B. M. und Stuttgarter Museum).

Epomophorus macrocephalo affinis, differt alis laete brunneis, radii longitudine 85 - 87 mm (F), 88 - 90 mm (M); dorsi colore laete brunneo griseo.

Der *Epomophorus* von der Goldküste, von Togo und von Lagos ist bisher immer als *Ep. macrocephalus* aufgefasst worden, trotzdem *Ep. macrocephalus* M nur einen Unterarm von 86 mm hat und trotzdem seine Flughaut schwärzlichbraun angegeben wird. Der *Epomophorus* on Mittel-Guinea hat einen Unterarm von 85 - 87 (F) resp. 88 - 90. (M) mm und seine Flughaut ist hellbraun. Der Hauptstrang des Plagiopatagiums wird von 18 - 22 Strängen durchschnitten. Die Entfernung der Aussenkanten der letzten oberen Molaren von einander ist beim M wie bei *macrocephalus* M etwas länger als die Länge der oberen Molaren-Reihe (cf. die Abbildungen bei Barboza und Tomes) mit der Abbildung auf Taf. 10. Die Zahnreihe ist länger als die Hälfte der Entfernung der ersten von der 7. Gaumenfalte. Das Bild auf Taf. II. Fig. 2 des Dobson'schen Catalogs entspricht dem Bilde, welches der Gaumen eines F bietet. - Ich widme diese Art dem durch seine zoologischen Forschungen im Togolande verdienten Herrn Graf Zech.

Abbildung des Schädels eines M von Lagos auf Taf. [10], Fig. 3 als *E. macrocephalus*. [B.M.] 3 MM Misahöhe und Kradji, Baumann; M pull. Gross-Popo, Bloess; 2 MM Lagos, Salmin; M, F, M juv., F juv. Accra, Reichnow, Unger und Pel.

Epomophorus (Epomophorus) zenkeri Matschie, 1899

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1865. *Ep. gambianus* Peters, P.Z.S. 1865 p. 400. - 1878. *Ep. gambianus* Dobson l.c. p. 11 vom Gabun. - 1885. *Ep. gambianus* von Niam-Niam. Jentink, Not. Leyd. Mus., p. 35 - 1887. specim. b., Jentink l.c. p. 251. - 1888. specim. b., Jentink l.c. p. 137. - 1889. *Ep. macro-*

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cephalus und *gambianus* von Porto da Lenha und Netonna, Noack, Zool. Jahrb. IV. p.200 - 205, Taf. V. Fig. 50 - 58 (Schädel von M ad. und juv., Gaumen von M ad. und juv.). - 1889. *Ep. gambianus* vom Rio Cuillo, Pungo-Andongo, Ambacca; Barboza du Bocage, Journ. Sc. Math. Phys. Nat. Lisboa, 2. ser. I. p. 14. - 1893. *Ep. gambianus* Matschie, Sitzb. Ges. naturf. Fr. Berlin p. 256 - 257. - 1896. E. de Pousargues, Ann. Sc. Nat. Zool. III. p. 251 - 253. - 1898. partim Barboza du Bocage, Journ. Sc. Math. Phys. Nat. Lisboa, 2. ser. V. p. 136.

Hab. Vom Kamerun-Fluss nach Süden bis zum Cuanza, nach Osten his Niam-Niam, also Süd-Kamerun, Gabun, Loango, Congo und Loanda; im besonderen bekannt vom Kamerun-Delta (Reichenow), vom Gabun (Büttner, Dobson), von Porto da Lenha (Noack), von Tschintschoscho (Falkenstein), von Netonna (Noack), von Malandje (von Mechow), von Pungo-Andongo, Ambacca, Quissango und vom Rio Cuillo (Barboza du Bocage), von Loanda (Peters), von Semmie in Niam-Niam (Jentink), von Franceville am oberen Ogowe (de Pousargues).

Dass diese *Epomophorus* von Nieder-Guinea nicht zu *Ep. gambianus* gehören, mit welchen sie in der Gestalt der Gaumenfalten grosse Aehnlichkeit haben, lässt sich aus folgenden Betrachtungen schliessen:

Bei *Ep. gambianus* Ogilb. variirt nach Tomes (P.Z.S. 1860 p. 531) die Länge des Unterarmes zwischen 76.2 und 85 mm, bei den Exemplaren von Nieder-Guinea zwischen 79 und 85 mm; es haben also die FF aus Nieder-Guinea einen ungefähr so langen Unterarm wie die MM von *gambianus* Ogilb. Nach der Ogilby'schen Original-Beschreibung von *gambianus* soll die Körperlänge $6\frac{3}{4}$ Zoll = 171.4 mm, die Kopflänge von der Nase zur Ohrwurzel $1\frac{3}{4}$ Zoll = 44.45 mm und die Spannweite der Flügel 1 Fuss 8 Zoll = 508 mm betragen. Die Abbildung des Schädels (P.Z.S. 1861 Taf. I. Fig. 2) beweist, dass wir es mit einem alten Exemplar zu thun haben, dessen Molaren schon stark abgekaut sind. Die entsprechenden Maasse für ein altes M von Tschintschoscho und ein altes F von Malandje betragen: 138 mm; 44 mm; ca. 555 mm resp. 127 mm, 43 mm, ca. 525 mm. Wir erkennen hieraus, dass *Ep. gambianus* von den *Epomophorus* aus Nieder-Guinea verschieden sein muss.

Von *Ep. gambianus* würde sich die hier zu betrachtende Art, welche ich nach meinem Freunde G. Zenker, dem ausgezeichneten Sammler in Kamerun, benenne, durch folgende Diagnose unterscheiden:

Epomophorus comptus H. Allen, 1862

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Hair soft ; thick above, thinner below. Color on the back a delicate reddish fawn, becoming darker on the loins and base of arms. Under surface with a large ovoid patch of dirty white, bounded laterally by longitudinal fawn colored stripes. Face covered with short brown hairs. Small yellow tufts at base of ears. Chin whitish. Epaulettes faintly marked. Lips moderate.

Dental formula, m $\frac{3}{5}$, c $\frac{1}{1}$, i $\frac{2}{4}$, c $\frac{1}{1}$, m $\frac{3}{5}$ = 26.

The fur of this bat differs from that of any other of the genus to which it belongs in not being unicolored. That upon the back of the neck and shoulders possesses three well-defined hues - the base is of a dark brown, the middle of a paler hue, while the tip forms a delicate reddish brown. This, however, can only be seen where the hair is thickest. Lower down upon the back and on the belly the entire length of each individual hair is of one color. Everywhere the hair covering is soft and yielding. All that space beginning between the eyes and terminating at the middle of the back is of thick, fine, beautiful hair. The sides are much thinner and darker; they form a continuous line with that of the ulna, where it extends down upon the wing membrane some little distance. The continuation of the dorsal fur forms on the ventral surface of the interfemoral membrane a sparsely furnished patch, which goes to make the lower portion of the lateral fawn-colored bands. These latter are narrow inferiorly, wider superiorly, where they terminate gradually in the lighter hue of the cervical region. From the external side a prolongation is sent up along the line of the humerus and ulna to within an inch of the carpal joint; both

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above and below this line the membrane has upon it several points of short lanuginous hairs of a whitish color.

The customary clumps of hair at the base of the ears are present. The anterior spot is larger than the posterior.

The characteristic shoulder tufts are by no means conspicuous; they are composed of thin long hairs of a light color. The centres are quite naked and placed more upon the neck than is usual.

The skull is smaller than that of *E. gambianus* Ogil. The post-orbital processes are small but well defined; temporal fossæ not approximate. The intermaxillaries slight, terminating in a point, not abruptly, as in *E. gambianus*, and supports but two incisors. The latter occupy the centre of the inter-canine space, and are slightly separated from one another.*

The other teeth differ in no particular from those of *E. gambianus* except in their mode of articulation, in which the inferior incisors close in front to the superior, as already seen in *Hypsignathus*, the superior teeth, however, are not entirely hidden.

This species holds a position between *E. gambianus* Ogil. and *E. shoensis* Rüpp. It differs from the former in the relative length of the nose, in the tricolored hair of back, and in the greater extent of the epigastric patch; from the latter in being much larger, and in the absence of the dark colored abdomen.

Dimensions.

Length of head and body: 6.0

" from snout to base of ears: 1.6
 " " eye to snout: 0.8
 " " anterior part of eye to ear: 0.8
 Height of ears: 0.9
 Breadth of ears: 0.6
 Length from nose to wing membrane: 2.8
 " of forearm: 3.0
 " longest finger: 6.0
 " fourth " : 4.8
 " thumb and claws: 1.2
 " basal joint: 0.3
 " second joint: 0.8
 " tibia: 1.6
 " foot and claws: 0.8
 Expanse of wing membrane: 18.0

Hab. Western Africa ; discovered by M. DuChaillu.

Epomophorus crypturus Peters, 1852

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Tafel V. Tafel XIII. Fig. 1 - 6.

E. colore corporis dilute umbrino, subtus pallidiori in cinereum migrante; patagiis umbrinis; ad basin auriculae utrinque macula alba.

Longitudo totalis 0,16, capitis 0,055, antibrachii 0,077; latitudo alar. expans. 0,5.

Habitatio: Africa orientalis, Tette, 17° Lat. Aust.

Der Kopf ist sehr grofs im Verhältnifs zum übrigen Körper, doppelt so lang wie breit, vor den Augen plötzlich verschmälert, so dafs die vorderen Augenwinkel nur halb so weit von einander entfernt stehen wie die hinteren. Der Abstand des Auges von der Schnauzenspitze ist viel gröfser als die Entfernung desselben vom Ohr. Die Pupille ist rund. Die Ohren sind elliptisch, an der Spitze abgerundet, ihr Rand ist ringsum saumartig verdickt, hinten schwach ausgeschnitten; an ihrer äufsern Fläche erheben sich neun bis zehn Querfalten, und der übrige Theil ist von netzförmig erhabenen Linien durchzogen; die innere Fläche ist sparsam behaart, die äufsere in ihrer obern Hälfte nackt, an der Basis aber, so wie an dem vordem und hintern Rande bis über die Hälfte hinauf ist die Behaarung lang und reich. Die Nase, welche über das Maul hinausragt, ist durch eine tiefe dreieckige Furche gespalten; die Nasenlöcher stehen seitlich und sind spiralförmig geschlitzt; die feingekörnte breite Nasenkuppe setzt sich, durch eine enge Längsspalte in der Mitte getrennt, bis zum Rande der Oberlippe fort; das vordere Ende der Unterlippe ist glatt und unbehaart. Die Entfernung der Nasen-

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löcher vom Rande der Oberlippe beträgt nur zwei Drittel von der Breite der zwischen ihnen Befindlichen Scheidewand. Die Maulöffnung reicht nicht ganz bis unter die Augen, und krümmt sich vorn in die Höhe. Der Körper, durch einen schmalen Hals vom Kopf abgesetzt, ist so kurz, dafs die Entfernung von dem obern Rande des Brustbeins bis zu dem After kaum die Länge des Kopfes erreicht; die Seitenflügel befestigen sich so an der Körperseite, dass sie in der Beckengegend nur halb so weit, 16 Millim., von einander entfernt stehen wie unter dem Oberarm. Das Schwanzrudiment liegt, in den langen Haaren versteckt, vor der Schenkelflughaut, an seiner Basis durch eine kleine Falle mit derselben verbunden. Tafel V. Fig. 3. Der Oberarm ist viel kürzer als der Kopf und etwas mehr als halb so lang wie der Vorderarm. Das erste Glied des Daumens, welches fast eben so lang ist, wie die drei Glieder des zweiten Fingers zusammen, wird beinahe zur Hälfte von der Flughaut eingehüllt. Der zweite Finger ragt mit seinem Nagel bis an das Ende vom Mittelhandknochen des dritten Fingers. Die erste Phalanx des dritten Fingers ist so lang wie das Mittelhandglied des zweiten Fingers; die zweite Phalanx desselben so lang wie sein eigenes Mittelhandglied. Die beiden Phalangen des vierten Fingers zusammen sind ein wenig länger, die des fünften Fingers ein wenig kürzer als ihre gleich langen Mittelhandglieder. Die Zehen sind an ihrer Basis bis zur Mitte ihres ersten Gliedes durch Häute mit einander verbunden. Die fünfte Zehe ist ein wenig kürzer als die drei mittleren gleich langen; die erste Zehe ragt mit ihrem ersten Gliede über das zweite Drittheil des zweiten Gliedes der folgenden hinaus. Die erste Zehe besteht aus zwei, die übrigen vier aus drei Gliedern; wol nur durch Unachtsamkeit sind in der Abbildung von *Epomophorus Whitei* (*Transact. Zool. Soc. II. pl. 7*) alle Zehen mit drei Gliedern gezeichnet. Die Körperflughaut setzt sich über den Fufsrücken bis zur Mitte des ersten Gliedes der zweiten Zehe fort. Die Schenkelflughaut ist schmal wie bei den schwanzlosen Pteropen, bogenförmig ausgeschnitten, an ihren Ecken durch einen schwachen Sporn sestützt, und endigt an der Basis des Mittelfufses. Die Krallen sind scharf und spitz, unten gefurcht; die Daumenkrallen ist um ebenso viel gröfser als die Zehenkrallen, wie die letzteren die Krallen des zweiten Fingers überragen. Die Behaarung ist dicht und wollig, an der Rückenseite des Körpers länger als an der Bauchseite desselben, am Vorderhalse kragenartig verlängert, über der Schulter bei Männchen und Weibchen einen Wirbel bildend; sie geht an den Oberextremitäten über das zweite Fünftel des Vorderarms hinaus, an der Rückseite der Unterextremität bis nahe zur Fufswurzel, an der vordem Seite derselben bis zur Mitte des Unterschenkels herab; sie dehnt sich an der Bauchseite weiter über die Flughaut aus, ist hier aber viel dünner als an der Rückseite derselben; die Schenkelflughaut wird bis auf ihre äufseren Winkel ganz durch die langen Haare verdeckt; die Rückseite der Flughaut ist ausserdem noch bis nahe zum fünften Finger hin mit zahlreichen Reihen kurzer Haarbüschel besetzt, welche nach der Fufsecke hin gedrängt und länger erscheinen. Die Zehen sind mit kurzen, steifen zerstreuten Haaren versehen. Aus den kurzen Gesichtshaaren springen zur Seite der Schnauze und über dem Auge schwache Schnurrhaare hervor.

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Die Farbe der Rückseite ist blafs umbrabraun, wie Milchkaffee, die Schultern, der Hals und die Bauchseite heller, die Mitte des Bauches ins Graue streifend; um die Augen herum und an den Seiten des Gesichts ist die Farbe gesättigt; vor und hinter dem Ohr befindet sich ein großer gelblichweißer Fleck. Die Iris ist bei jungen Thieren graubraun, bei den alten rothbraun. Die Nasenkuppe, Ohren und Flughäute sind ebenfalls umbrabraun aber etwas dunkler gefärbt. Die einzelnen Haare sind einfarbig, am Grunde etwas dunkler. Die Nägel sind von schwarzbrauner Farbe.

Der Schädel ist sehr niedrig, ohne Längsleiste; die Oberkiefer und Gaumenbeine sind sehr verlängert, so daß der sehr kurze Processus postorbitalis weit hinter der Mitte des Schädels zu liegen kommt; die Breite des Schädels hinter dem Processus postorbitalis ist viel größer als vor demselben; der Jochbogen verläuft horizontal; der Infraorbitalcanal ist ein wirklicher Canal wie bei *Macroglossus*. Der Unterkiefer ist sehr niedrig, so daß die größte Höhe fast drei Mal in der Länge desselben enthalten ist. An sieben Exemplaren verschiedene Alters besteht das Gebiß beständig aus $\frac{4}{4}$ Vorderzähnen, $\frac{1}{1}$ Eckzähnen, $\frac{3}{5}$ Backzähnen. Die oberen Vorderzähne sind conisch und nach hinten gekrümmt, die unteren dreilappig. Die oberen und unteren Eckzähne sind von gleicherhakenförmig gekrümmter Gestalt, und ihrer hintern Seite abgeflacht, die unteren um ein Drittel kleiner als die oberen. Die drei oberen Backzähne sind zweiwurzelig; die Krone des ersten ist eckzahnförmig, dreikantig, mit einer äußern convexen, einer vordern innern graden, und einer hintern innern concaven Fläche versehen; der zweite besteht aus einer einfachen Schmelzlamelle, welche eine biscuitförmige Grube umschließt, und aufsen einen starken, innen einen schwachen Zacken bildet; der dritte ist eben so lang wie der vorhergehende, aber viel niedriger, nur aufsen mit einem schwachen Zacken versehen. Im Unterkiefer folgt zunächst auf den Eckzahn ein kleiner einwurzeliger Lückenzahn mit kurzer conischer Spitze; darauf kommen drei zweiwurzelige Zähne, welche in ihrer Form den drei oberen Backzähnen entsprechen, aber etwas kleiner sind, und zuletzt ein kleiner zweiwurzeliger Höckerzahn, welcher nur halb so lang ist wie der vorhergehende. Die Wirbelsäule besteht aus 35 Wirbelkörpern, von denen 7 flache Halswirbel, 14 rippentragende Rückenwirbel und 4 Lendenwirbel sind, die folgenden 7 bis zur Vereinigung der Sitzbeine als Kreuzbeinwirbel und die letzten 3 als Schwanzwirbel betrachtet werden können. Das Brustbein besteht aus 5 Stücken und trägt 7 Rippenpaare. Das Schlüsselbein ist bogenförmig gekrümmt. Das Oberarmbein ist so lang wie der Schädel, an seinem untern Ende undurchbohrt, und am Condylus externus durch ein Gelenkstück vergrößert, welches den Streckmuskeln der Hand zum Ansatz dient. Die Ulna ist sehr dünn und mit dem Radius verwachsen; das Olecranon getrennt. Die Handwurzel besteht aus 7 Knochen, drei in der ersten, vier in der zweiten Reihe. Die Fibula ist unvollständig: die Zahl der Fußwurzelknochen ist 7; drei in der ersten, vier in zweiten Reihe; dann folgen fünf Mittelfußknochen und endlich Phalangen, von denen man zwei an der ersten Zehe, drei an den vier übrigen zählt. Die Schleimhaut des Gaumens bildet 7 wulstige Querfalten, von denen die beiden

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letzten getheilt sind; ganz hinten befinden sich noch zwei flache Quervorsprünge mit gezähnelten Rändern. Die Lippen sind sehr breit wie bei den *Macroglossus* und an ihrer innern Seite durch gezackte Vorsprünge der Schleimhaut ausgezeichnet, welche besonders stark um Mundwinkel entwickelt sind. Die Zunge ist von hornigen rückwärts gerichteten Papillen bedeckt, die an der abgerundeten Zungenspitze fein, lanzettförmig, auf der Mitte ihres ersten Drittheils sehr groß und dreispitzig, auf den übrigen Theilen platt und dachziegelförmig gestellt sind; an ihrer Basis stehen drei große warzenförmige flache Papillen. Der Oesophagus mündet in den 43 Millim. langen Magen, nur 7 Millim. vom Pförtner entfernt; der einfache Darm hat eine Länge von 960 Millim. Die Leber zerfällt durch zwei mehr oder weniger tiefgehende Einschnitte in drei Lappen; die längliche Gallenblase liegt unter dem rechten, oder zwischen diesem und dem mittlern Lappen. Die Milz ist 22 Millim. lang, 2 Millim. breit, und liegt hinter dem convexen Rande des Magens. Das Pancreas besteht in einer dünnen durchscheinenden Drüsenmasse, welche sich nur erkennen läßt, wenn man das Mesenterium gegen das Licht hält. Die Nieren sind zwei länglich abgerundete, 12 Millim. lange und 8 Millim. breite Körper, an deren oberen vorderen Enden die Nebennieren, kleine scheibenförmige Körper von 2 Millim. Durchmesser liegen. Die trächtigen Weibchen enthalten in jedem Horn des Uterus einen einzigen Fötus, der durch eine einfache Placenta ernährt wird. Die Brustwarzen sitzen am Rande des Brustmuskels auf, vor und unter der Achselgrube. Die Ruthe der Männchen besitzt weder knorpelige noch knöcherne Skelettheile; die Haut der Eichel ist in Längsfalten zusammengelegt. Das Zungenbein besteht aus einem sattelförmigen, unten breiteren Mittelstücke, mit dem jederseits unten das säbelförmige große Horn verbunden ist; oben setzen sich die kleinen Hörner an, welche aus zwei Gliedern bestehen, von denen das erste sehr klein, das zweite dagegen sehr groß, auffallend breit und von lanzettförmiger Gestalt mit einem langen feinen Fortsatz versehen ist. Der Kehledeckel ist so lang wie breit, von rhomboidaler Gestalt, mit abgerundeten Ecken. Der Schildknorpel ist unten viel breiter ab oben, woselbst der Rand sich in eine mittlere Spitze verlängert. Die Luftröhre enthält bis zu ihrer Theilungsstelle 21 - 22 Halbringe. Die linke Lunge ist einlappig und vierzipflig, die rechte durch zwei bis drei tiefe Einschnitte in drei bis vier Lappen abgetheilt. Das Herz hat eine eiförmige Gestalt, eine Länge von 13, und an der Basis eine Breite von 10 Millimetern; die Spitze der linken Herzkammer ragt weit über die linke hervor; die Vorkammern stehen durch eine weite Oeffnung mit einander in Verbindung; das rechte Herzohr ist breiter und abgerundeter als das linke; der kurze Stamm der aufsteigenden Aorta theilt sich in zwei Trunci anonymi, von denen der linke etwas früher als der rechte in seine beiden Äste, die Carotis und Subclavia, zerfällt.

Diese Thiere leben, wie es scheint, ausschließlic von Vegetabilien, da ich in ihrem Magen nie etwas andres als Ueberbleibsel und Kerne von Früchten vorfand.

Der einzige Ort, wo ich diese Art angetroffen habe, ist Tette, gegen 120 Meilen von der Küste und im 171° Südl. Br. gelegen. Die Eingebornen haben für diesen Flederhund zwei verschiedene Namen, *djæmea* und *demuanjundo*.

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Mafse in Millimetern.

Ausgewachsen Männch.; Trächt. Weibch.; Trächt. Weibch.; Junges Weibch.; Skelet Weibch

Ganze Länge von der Schnauze bis zur Mitte der Schenkelflughaut: 160; 155; 160; 120; -

Flugweite: 500; 530; 504; 480; -

Länge des Kopfes zwischen Schnauzenende und Hinterhaupt: 55; 50; 50; 44; 48

Breite des Kopfes an den Jochbogen: 23; 25; 26; 23; 24

Entfernung der hinteren Augenwinkel: 21; 20; 22; 20; -
 Entfernung der vorderen Augenwinkel: 12; 11; 12; 11; -
 Entfernung des Auges vom Ohr: 12; 11; 12; 10; -
 Entfernung des Auges von der Mitte des Nasenendes: 23; 22; 18; 17; -
 Länge des Ohres am vorderen Rande: 21; 21; 23; 17; -
 Breite des Ohres: 14; 14; 15; 12; -
 Länge des Oberarms: 50; 48; 50; 31; 48
 Länge des Vorderarms: 77; 79; 78; 70; 78
 Lg.1.F. (Mittelh.10,10,11,10. 1.Gl.18,16,18,16. 2.Gl. $6\frac{1}{2}$,6,6,6): 35; 32; 35; 32; 32
 Lg.2.F. (Mittelh.36,39,40,33. 1.Gl.10,7,8, $\frac{1}{2}$. 2.Gl. 6, $4\frac{1}{2}$,6, 5, 3.Gl. $4\frac{1}{2}$,5,5,5): 57; 56; 59; 52; 56
 Lg.3.F. (Mittelh.52,55,56,49. 1.Gl.37,37,38,32. 2.Gl.50,55,53,46): 142; 147; 148; 127; 147
 Lg.4.F. (Mittelh.51,53,55,46. 1.Gl.26,25,28 $\frac{1}{2}$,24. 2.Gl.29,30 $\frac{1}{2}$,30,26): 116; 110; 111; 98; 114
 Lg.5.F. (Mittelh.50,52,54,46. 1.Gl.25,24,26,23. 2.Gl.26,27,26 $\frac{1}{2}$,24): 103; 105; 106; 95; 106
 Länge des Oberschenkels: 19; 18; 18; 16; 19
 Länge des Unterschenkels: 31; 31; 30; 27; 31
 Länge des ganzen Fusses: 22; 22; 22; 21; 22 $\frac{1}{2}$
 Länge der mittlern Zehen mit dem Nagel: 16; 15; 15; 15; 15
 Länge des Sporns: 9 $\frac{1}{2}$; 8; 8; 6; -
 Schenkelflughaut an der Mitte des Unterschenkels: 10; 10; 12; 9; -
 Länge des Schwanzes: 5; 5; 5; 2; 5
 Länge der ganzen Wirbelsäule: -; -; -; -; 90
 Länge des Schlüsselbeins in grader Richtung: -; -; -; -; 20
 Länge des Schulterblatts: -; -; -; -; 25
 Breite des Schulterblatts: -; -; -; -; 8
 Länge des Brustbeins: -; -; -; -; 2?
 Breite des Brustbeins am Manubrium: -; -; -; -; 1?
 Höhe des Beckens: -; -; -; -; 1?
 Abstand der Schambeine: -; -; -; -; 1?

Diese Art steht dem *E. macrocephalus*, Ogilby (*Proceedings of the zool. society of London*. 1835. III. pag. 101) außerordentlich nahe; letztere unterscheidet sich aber von ?? nach Ogilbys Beschreibung durch die sehr kurzen Flügel (1 Fufs 3 Zoll Engl.) und die schwärz-

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liche Farbe derselben, *E. gambianus*, Ogilby, besitzt nach Gray (*Magazine of zoology and botany*. 1838. II. pag 504) keine weissen Haarbüschel an der Basis der Ohren, dagegen dergleichen über der Schulter. *E. Whitei*, Bennett (*Pteropus epomophorus*, Bennett; *Pteropus labiatus*, Temminck) ist durch die weissen Büschel über den Schultern ausgezeichnet. *Pteropus schoënsis*, Rüppell, ist viel kleiner. *Pteropus Wahlbergii*, Sundevall, scheint nach der gegebenen Diagnose einen längeren Schwanz zu haben und vielleicht auch anderes Gebiß. Alle diese Arten sind jedoch sehr verwandt, und die Zahl derselben wird sich durch Untersuchung großer Reihenfolgen wahrscheinlich noch vermindern. Das zoologische Museum hat ganz kürzlich ein ausgestopftes Exemplar von Herrn Temminck, aus Guinea durch Herrn Pel erhalten, welches keine weitere Unterscheidung von meiner Art aus Mossambique zuläßt, als dafs die Augen in der Mitte zwischen Ohr und Nasenende liegen

Epomophorus Dobsonii Bocage, 1889

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Ressemble Beaucoup à l'*E. gambianus* par son aspect, par sa taille et par ses proportions: mais il a une tête plus courte, des oreilles plus étroites vers l'extrémité, à bord extérieur légèrement concave dans leur tiers supérieur, et des couleurs différentes: d'un brun-roux uniforme sur la tête et en dessus, sans aucun mélange de teintes jaunâtres, plus pâles en dessous et tirant au grisâtre sur la partie inférieure du ventre; les membranes alaires sont de la couleur du dos.

Il porte, comme l'*E. gambianus*, une petite touffe de poils blanchâtres à l'épaule et quelques poils de la même couleur de chaque côté de la base de l'oreille.

Les dimensions du mâle adulte, représentant unique de l'espèce en notre possession, sont à peu-près celles d'un mâle de la même taille de l'*E. gambianus* à l'exception de la tête, qui est un peu plus longue chez celui-ci. Nous remarquons cependant une particularité que nous ne pouvons passer sous silence, en attendant qu'elle soit confirmée par l'observation d'autres exemplaires: chez tous nos individus de l'*E. gambianus* la première phalange du 4^e doigt a à peu-près la moitié de la longueur du métacarpe correspondant, tandis que chez la nouvelle espèce elle est beaucoup plus longue (V. tableau).

Les plis du palais, dont s'est servi si avantageusement M. Dobson pour bien caractériser les différentes espèces d'*Epomophorus*, nous fournissent des caractères différentiels d'une importance décisive, qui ne permettent pas de confondre cette espèce avec aucune de ses congénères. Sous ce rapport elle s'écarte absolument de toutes les espèces connues.

Il est difficile, ... , de faire bien saisir par une ... les description la disposition et ... Formes toutes spéciales ... de ces plis; mais la figure qui accompagne le texte ... à nous faire mieux comprendre.

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Le palais de notre individu présente 5 plis distincts, derrière lesquels se trouvent encore quelques lignes onduleuses, plus ou moins saillantes, finement denticulées. Il y a d'abord un premier pli du palais pli derrière les canines, qui ressemble au

premier pli du palais chez l'*E. gambianus*, mais il forme un angle plus saillant en arrière et porte au milieu une incision bien marquée. Après celui-ci, nous remarquons dans l'intervalle de la 1^e à la 2^e prémolaire un gros pli transversal droit, dont les extrémités se divisent en deux branches qui vont terminer respectivement sur chacune des deux dents pré-citées. Le troisième pli correspond, à la molaire et ressemble assez bien au cinquième pli de l'*E. gambianus*. Plus en arrière et après un intervalle plus grand, se trouvent deux plis, l'un derrière l'autre et plus rapprochés entre eux, qui n'ont pas d'équivalents, quant à la forme, chez les autres espèces du genre; ils sont aplatis, disposés horizontalement et à bords antérieurs libres; l'antérieur (4^e pli) a une petite incision au centre et chacune de ses moitiés représente un triangle dont le vertex porte en dessus une petite papille; le 5^e pli, placé à petite distance du 4^e et concentrique avec lui, a à peu-près la même forme, mais ses dimensions sont plus restreintes.

Les lignes finement dentelées qui complètent le dessin du palais se trouvent bien indiquées dans la figure 1.

Notre individu, un beau mâle adulte, nous a été envoyé récemment de *Quindumbo*, dans l'intérieur de Benguela, par notre zélé naturaliste M. d'Anchieta.

Le tableau ci-après montre les principales dimensions de notre exemplaire en regard de celles d'un individu du même sexe de l'*E. gambianus*, recueilli par M. d'Anchieta dans la même localité.

E. Dobsonii, M ad.; *E. gambianus*, M ad.

Tête et corps: 172 mm.; 176 mm.

Tête: 62; 65

De la narine à l'œil: 24; 30

De l'œil à l'oreille: 13; 13

Oreille: 25; 24

Avant-bras: 88; 91

Pollex: 35; 35

3^e doigt (met. 62, 1^e ph. 43, 2^e ph. 65) 170; (met. 65, 1^e ph. 41, 2^e ph. 59) 165

4^e doigt (met. 56, 1^e ph. 33, 2^e ph. 37) 126; (met. 62, 1^e ph. 30, 2^e ph. 34) 126

Jambe: 35; 36

Pied: 22; 23

Epomophorus franqueti Tomes, 1860

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(Pl. LXXV.)

If the species in the present monograph took rank according as they are more or less typical in form, the present one should appear as second, the *E. labiatus* probably as third, followed by *E. gambianus*, and the list should be completed by the smallest and least typical species - *E. schoënsis*. But the first and most typical species is succeeded by the one which was described at very nearly the same time, as being much better known than those which were to follow.

The present one is much the largest species, attaining an expanse of more than 2 feet, and has the same singular tufts of hair on the shoulders as are recorded of the first Species in the list - *E. macrocephalus*, but much more developed than in that species, and of a pale yellow colour. The only known specimen was forwarded to the French National Collection by Dr. Franquet of the French Imperial Navy, and from it I have, by the kind permission of M. Geoffrey St. Hilaire, taken the description which follows, and have had a carefully executed drawing made by M. Oudart, from which the illustrations accompanying the present paper have been copied. Its country is the same as that of the Gorilla.

The head is not nearly so long and narrow relatively as that of *E. macrocephalus*, but more nearly resembles that of *E. gambianus*. The ears, as in the other species of the genus, are of medium size, oval, and a little narrowed towards the tips; they are furnished with small tufts of fine white hair at the base of their inner and outer margins, like those of all the other species here described. The lips, as far as can be ascertained from the inspection of a mounted specimen, are large, although perhaps not quite equal to those of some of the other species. The interfemoral membrane is rather more ample than is usual in the genus.

The fur extends considerably on to the membranes, above and below, as in *E. macrocephalus*, and it is similarly unicolour, and possesses the same soft cottony texture. That of all the upper parts is of a cinnamon-brown colour, brighter and deeper than in the other species; the under parts similar, but the patch of whitish on the abdomen, which is faint as in the others, here takes the form of a clearly-defined oval space of pure white, as much as 2½ inches long.

The shoulder tufts are very much developed, and differ somewhat from those of *E. macrocephalus*. They occupy a space on the shoulder of as much as 1½ inch in length, in a descending direction; the lower half of this space consists of fur, which is of the same length and texture as that of the surrounding parts, but is of a buffy-yellow colour; whilst the upper part, constituting the real shoulder tuft, is composed of long yellow hairs, which spring outwards, and then curve downwards, partially hiding the short yellow hair already mentioned. All this yellow fur, both long and short, has a clear and well-defined outline. All the membranes are of a darkish cinnamon-brown.

The cranium is much less elongated than in either of the preceding species, and in its general proportions bear some resemblance to that

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of *Pachysome stramineum*, especially in the expansion of the zygomatic arches; but the teeth are of precisely similar number and relative proportion with each other, as in *E. macrocephalus*, although they generally are stouter than in that species. The lower jaw exhibits most unequivocally the peculiarities mentioned in detailing the generic characters. The dimensions of the cranium will be given with those of some of the other species, so as to afford a more direct means of comparison.

" "

Length of the head and body: 7 3

----- of the head: 2 8

----- of the ears: 0 10

----- of the fore-arm: 3 9

----- of the longest finger: 7 3

----- of the fourth finger: 5 6

----- of the tibia: 1 6

----- of the foot and claws: 0 11

Expanse of wings, about: 30 0

Hab. Gaboon.

Epomophorus guineensis Bocage, 1898

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Epomophorus guineensis, n. sp.

E. gambianus? Bocage, *Jorn. Ac. Sc. Lisb*, 2.° serie, t. I, pp. 2 et 3, fig. 2, (le palais).

Un mâle adulte de *Bolama* (Barahona).

Cet individu nous paraît distinct de l'*E. Gambianus*, par sa tête sensiblement plus allongée à museau plus étroit et surtout par le nombre et la disposition des plis du palais. Pour s'en assurer il suffit de comparer la figure ci contre à la figure de Dobson qui représente bien exactement les plis du palais de l'*E. gambianus*.

Epomophorus reii Aellen, 1950

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MATERIEL. - Le matériel faisant l'objet de cette note préliminaire a été récolté par la Mission scientifique suisse au Cameroun, en 1947. Il appartient au musée d'histoire naturelle de La Chaux-de-Fonds.

N° 1437, type: F adulte; Rei Bouba (région de Garoua, Cameroun), 13 septembre 1947.

1430, F jeune; même lieu, même date.

1438, M jeune (sur la femelle N° 1437).

DESCRIPTION. - *Epomophorus reii* est voisin de *E. gambianus* Ogilby et *E. angolensis* Gray. Les mesures externes correspondent aux mesures données pour *gambianus* par ANDERSEN et *angolensis* par MONARD, à part celles de quelques segments alaires.

Le crâne est moins allongé que dans *gambianus*: la largeur zygomatic mesure plus de la moitié de la longueur totale du crâne. La longueur du palais est 2 1/5 fois la largeur mesurée des bords externes des molaires supérieures (M1-M1); cette dernière mesure est plus grande que la longueur du palais postdentaire. Les diverses mesures sont comprises, pour la plupart, dans les limites que donne ANDERSEN pour *E. crypturus* Peters. Les plis du palais sont disposés nettement autrement que chez les autres espèces du genre: le 4° pli palatal est plus près du 3° que du 5°, mais la distance du 4° au 5° n'est pas le double de celle du 3° au 4°. Le 4° pli n'est pas entièrement devant une ligne allant des bords antérieurs de M1-M1: il est exactement entre les prémolaires postérieures (Pm4-Pm4) et M1-M1; son bord le plus avancé atteint le tiers de la longueur de Pm4. Le 5° pli est très en arrière de la rangée dentaire. Le 6° est derrière le milieu du palais post-dentaire.

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Le pelage n'a pas une coloration différente, dans ses traits essentiels, de celle des autres *Epomophorus*:

Faces supérieures: brun-roux, assez foncé vers l'arrière. Front plus clair. Touffes de poils blancs à la base des oreilles.

Côtés: joues et flancs comme le dessus, côté du cou nettement plus clair.

Faces inférieures: brun-jaunâtre clair. Poitrine comme les côtés du cou. Ventre blanc pur, tranchant nettement avec la couleur des flancs. Parties postgénétales de la couleur du dos.

Les poils de la gorge sont plus longs que le reste du pelage.

Les autres caractères sont ceux des espèces voisines.

COMPARAISON AVEC LES ESPECES VOISINES. *Epomophorus reii* diffère des autres espèces du genre par les caractères suivants (femelles):

de *crypturus* Peters.

Les mesures externes sont plus grandes, les mesures du crâne concordent à peu près. La distance du 3^e au 4^e pli palatal est plus petite que celle du 4^e au 5^e, alors que chez *crypturus* et *gambianus* elle est plus grande. Le 4^e pli est situé entre les dents Pm4-Pm4 et M1-M1 au lieu d'être à côté de M1-M1. La longueur du palais est 2 1/5 fois celle mesurée des bords externes de M1-M1 (2 fois chez *crypturus*). Le rapport C-M1 / M1-M1 est de 1.40 pour *reii* et 1.20 à 1.32

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pour *crypturus*. *Epomophorus crypturus* habite le Zambèze et la vallée du Limpopo.

De *gambianus* Ogilby.

Les mesures externes sont semblables. Les mesures du crâne sont plus fortes chez *gambianus*. Pour l'arrangement des plis du palais, voir les différences signalées avec *crypturus*. La largeur zygomatique mesure plus de la moitié de la longueur totale du crâne, alors que dans *gambianus* cette largeur est plus petite. Chez *gambianus*, la longueur du palais est 2 1/3 à 2 1/4 fois celle mesurée des bords externes des molaires M1; cette dernière mesure est plus petite que la longueur du palais postdentaire, alors qu'elle est plus grande chez *reii*. Le rapport rostr / C-M1 est de 1.05 chez *reii* et de 1.14-1.17 chez *gambianus*. L'indice digital (rapport du 3^e doigt au 5^e) est de 1.36 chez *reii* et de 1.31 à 1.32 chez *gambianus*. L'aire de répartition de *Epomophorus gambianus* comprend la zone s'étendant du Sénégal au sud de l'Abyssinie par la Sierra Leone, la Côte de l'Or, le Togo et la Nigeria.

De *angolensis* Gray.

Les mesures externes correspondent en gros. Les mesures du crâne ne sont pas comparables, car les auteurs ne donnent pas de séries complètes de mensurations de crânes de femelles: les quelques chiffres indiqués par ANDERSEN et MONARD conviennent à *reii*. Le 4^e pli est à côté des prémolaires Pm4, chez *angolensis*, au lieu d'être entre Pm4 et M2. Le 5^e pli touche à une ligne joignant les dents M1, alors qu'il est très en arrière de la rangée dentaire chez *reii*. Le 6^e pli est au milieu du palais postdentaire et non en arrière du milieu comme dans *reii* ou *gambianus*. *Epomophorus angolensis* habite l'Angola et le Damara.

De *pousarguesi* Trouessart.

Les mesures externes et les mesures du crâne sont plus fortes chez *pousarguesi*, connu seulement par le type: une femelle. L'arrangement des plis palataux ne serait pas différent de celui des plis de *angolensis*. Comme chez *gambianus*, la largeur zygomatique est plus petite que la moitié de la longueur du crâne (elle est plus grande

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[table removed - eds.]

chez *reii*). Le rapport C-M1 / M1-M1 est de 1.34 (1.40 chez *reii*) et le rapport rostre / C-M1 1.11 (1.05 chez *reii*). L'indice digital est de 1.30. *Epomophorus pousarguesi* a été trouvé une seule fois dans le cours supérieur du Chari.

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Par l'arrangement de ses plis palataux, par ses rapports de diverses mesures crâniennes, notre espèce se montre intermédiaire entre *gambianus* (et *crypturus*) et *angolensis*.

La nouvelle espèce est dédiée à la ville de Rei, sur la rivière du même nom (affluent de la Bénoué); dans la région de Garoua (Cameroun).

Les mœurs ne sont malheureusement pas connues, les chauves-souris nous ayant été apportées par des indigènes.

Epomophorus veldkampii Jentink, 1888

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One specimen, Buluma, Fisherman Lake (Büttikofer and Sala).

It is of the size of *Epomophorus pusillus* and is very difficult to distinguish externally from this species, but the palate-ridges are so different from the peculiar V-shaped groove in *Ep. pusillus*, that it can never be confounded with this species. First palate-ridge undivided, straight, between the canines; second and third, undivided, semicircular, between the first and second premolars; the following ridges, five in number, are divided in their centre, they form semicircular lines, the portion near the central groove is thickened; the remaining ridges are represented by toothed lines.

Ears with a thickened dark-colored margin; wing-membrane from the middle of the back of the first phalanx of the second toe; tail shorter than in *Ep. pusillus*.

Measurements of the type, an adult female, in alcohol (in mm):

Eye from tip of nostrils: 9.5

ear: 15

forearm: 50

thumb: 22

third finger, metacarpal: 38

" " , 1st phalanx: 22.5

" " , 2nd " : 33

fifth finger, metacarpal: 36

" " , 1st phalanx: 16

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fifth finger, 2nd phalanx: 16

tibia: 21.5

The color of the far is somewhat lighter tinged than in *Ep. pusillus*; distribution of fur like in that species.

I call this new species, *veldkampii*, as Biittikofer wishes to connect the name of one of his Liberian friends with this new acquisition. Mr. Veldkamp, at present Consul for the Netherlands; in Liberia, has helped our travellers as much as he could, has promoted their investigations in every way and finally assisted to Sala's funeral.

Epomops Gray, 1870

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Like *Epomophorus*, but head ovate. Skull ovate, broad. Face moderate, not nearly so long as the brain-case. Zygomatic arches convex. Cutting-teeth separate, far apart in the intercanine space. Grinders $\frac{4}{5}$, close together.

Epomophorus, sp., Tomes.

Eptesicus Rafinesque, 1820

p. 2

N.G EPTESICUS. Four acute fore-teeth to the upper jaw, in two equal pairs, separated by a great interval and a large flat wart, each pair has two unequal teeth, the outside tooth is much larger and unequally bifid, the outside one much larger, inside tooth small and entire. Six fore-teeth to the lower jaw, equal

p. 3

very small, close and truncate. Canine teeth very sharp, curved and long. Grinders unequally trifid. Snout plain, nose without appendages. Ears separated, auriculated. Tail mucronate. - This genus appears to differ from all those of Geoffroy and Cuvier, among the extensive tribe of Bats. The name means house-flyer.

Eptesicus ater J.A. Allen, 1917

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Type, No. 48988, M ad., skin and skull, Faradje, March 1, 1911; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No, 1466.

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Similar in general appearance to *E. tenuipinnis* but smaller, with black ears and foot, and wholly black above.

Pelage of upperparts brownish black to base of hairs, which are without light tips; below the hair of foreneck and upper chest brownish black tipped with whitish, rest of lower parts clear white, the hairs dusky at base; wing membranes white above and below; interfemoral membrane brownish white above, paler below; ears, feet, tail and limb bones dark brown, in strong contrast with the light-colored membranes. Size and general structure, including ears and tragus, as in *E. tenuipinnis*.

Collectors' measurements of type: Total length, 68 mm.; head and body, 39; tail, 29; foot, 6; ear, 12. Forearm (from skin), 27.5; in an adult male topotype, 27.9; in two subadult specimens from Niangara, M 27, F 25.

Skull, type, total length, 12.2; condylobasal length, 11.6; zygomatic breadth, 7.5; breadth of braincase, 6.6; interorbital breadth, 3.4; maxillar breadth, 5.2; breadth at basis of canines, 4; length of upper tooththrow (c-m³), 4; length of mandible, 8.4; angle to condyle, 1.5; depth at coronoid, 2.5.

Represented by 4 specimens: Faradje, 2 (type skin and skull and alcoholic topotype); Niangara, 2 (subadult. skins with skulls).

E. ater is nearly related to *E. tenuipinnis*, from which it differs in the much darker coloration of the pelage, the ears and feet blackish instead of pale yellow, and in slightly smaller size.

Eptesicus capensis angolensis J.Eric Hill, 1937

p. 1

TYPE. - No. 85535, Amer. Mus. Nat. Hist.; M adult, skin and skull; from Chitau, Angola. Secured by the Vernay Angola Expedition, Mr. H. Lang, collector, orig. no. 620, August 5, 1925.

The American Museum has a series of 18 from the type locality; there are two skulls, also topotypical, in the Carnegie Museum, Pittsburgh. Records of *E. capensis* in Angola, possibly referring to this race are Caconda and Huilla; Vila da Ponte. The more southern records may possibly concern *E. c. nkatiensis* Roberts.

GENERAL CHARACTERS. - A race of *Eptesicus capensis* with blackish wings and yellowish-brown body, the hairs above and below without dark bases.

COLORATION. - Upperparts darker than tawny olive, the bases of the hairs paler. Underparts slightly grayer than cinnamon-buff. Wing membranes and ears blackish, uropatagium slightly paler.

SKULL. - Braincase considerably higher than rostrum; the latter broad and short. Occipital height more than 65 per cent of mastoid breadth in adults.

DENTITION. - Medial upper incisor (I^2) deeply bifid; I^3 with small posterior cusp, only about one-third shorter than I^2 . I_{1-3} placed in line with the margin of lower jaw. Other teeth about as in *E. minutus*.

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MEASUREMENTS.-Head and body, 56; tail, 34; hind foot, 10; ear, 12.5; forearm, 37.0; digit III, 34.1 X 12.9 X 12.0 X 6.8. Skull: greatest length, 14.2; basilar length, 11.6; palatilar length, 5.9; zygomatic breadth, 10.4 (approximate); interorbital breadth, 4.2; mastoid breadth, 8.5; outside breadth across M^2 - M^2 , 6.9; height occiput, 5.8; maxillary alveoli, 5.4; crowns P^4 - M^3 , 4.3; width M^1 , 1.5.

Eptesicus capensis nkatiensis Roberts, 1932

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A small form of about the same size as *E. c. gracilior* Thomas and Schwann, but altogether lighter coloured above (near clay colour or rather lighter) and below whiter, the hairs on the legs and tail pure white to the base, only the extreme base of the hair about the abdomen dark, but the white decreasing and the dark base of the hair increasing forwards to the chest and throat, the chin and cheeks like the back.

Type: T.M. No. 6549, adult F, Nkate, Northern Bechuanaland, 18th August, 1930, No. 999, Vernay-Lang Kalahari Expedition. Length of head and body 44, tail 36, hind foot (c.u.) 8, ear 14. Skull: canines to occiput 13.3, zygomatic width 8.7, width of brain case 7, width of palate across outside of $msuper$ 2 5.7, length of upper tooth row (c- m^3) 4.8, of mandible to condyle 10 mm.

Eptesicus faradjius J.A. Allen, 1917

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Type, No. 49045, F ad., skin and skull, Faradje, northeastern Belgian Congo, Feb. 21; 1911; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 1914.

Upperparts tawny olive, the hairs dusky at extreme base; underparts similar but much paler, the tips of the hairs light tawny on throat, breast and middle of belly, more fulvous on side and pubic region; ears pale brown; limb bones and interfemoral membrane light brown above, paler below; wing membranes pale yellow on both surfaces.

Collectors' measurements of type: Total length, 93; head and body, 59; tail, 39; foot, 8; ear, 11. Forearm (from skin), 33.

Collectors' measurements of ten topotypes (5 males, 5 females): Total length, M 88 (83-95), F 92.6 (90-96); head and body, M 51 (46-56), F 54.2 (52-56); tail, M 37 (35-38), F 39 (35-43); foot, M 8.4 (8-9), F 8.4 (8-9); ear, M 11.4 (11-12), F 11.4 (11-12). Forearm (same specimens), M 33.5 (32.5-35.6), F 34.9 (32.5-36.2).

Skull (type), total length, 13.8; condylobasal length, 13; zygomatic breadth, 8.8; breadth of braincase, 7; mastoid breadth, 8.2; interorbital breadth, 4; maxillar breadth, 6.1; breadth at base of canines, 4.0; upper tooththrow (c- m^3), 4.0; length of mandible, 10.2; angle to condyles, 1.8; depth at coronoid, 3.2; lower tooththrow, 5.3.

Skull, 9 topotypes (4 males, 5 females): Total length, M 13.5 (13.2-13.0), F 13.7 (13.4-13.7).

Represented by 24 specimens: 14 (topotypes) from Faradje, collected February 18-21, March 1-30, and August 2, 1911, and 10 from Niangara, collected November 16-December 26, 1910.

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The two series are indistinguishable in every respect. Each series, however, shows a considerable range of variation in color, some of it due to age but mainly purely individual, the fawn color of the upper parts varying greatly in tone in different specimens, independently of sex and age.

There is apparently a slight sexual difference in size, the females having the forearm averaging about 2 mm. longer than the males, and there is a corresponding but slighter average difference in the length of the skull.

Eptesicus faradjus appears to be related to *E. flavescens* (Seabra), but it is smaller than the latter and should be readily distinguishable from it by the deep fawn color of the upperparts (varying in different specimens from light yellowish to olivaceous and even to slightly rufescent in different specimens), combined with a forearm length of about 34 mm. as against 37 in *flavescens*. It is very different in coloration from *E. phasma*, with a large series of which it has been compared, and with which it closely agrees in size.

Eptesicus garambæ J.A. Allen, 1917

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Type (and only specimen), No. 49340, M ad. in alcohol, Garamba, March 14, 1912; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 1888.

Upperparts very dark brown, the hairs without appreciably lighter tips; underparts similar but slightly paler, the tips of the hairs faintly grayish; membranes naked, blackish with a very narrow whitish edging; inner surface lighter and, slightly grayish; lower surface of limb bones whitish, in strong contrast with the dark membranes; wing membranes from base of outer toe; last caudal vertebra free for half its length; proximal vertebrae slightly hairy.

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Total length, 77.6 mm.; head and body, 44.6; tail, 33; forearm, 33; third metacarpal, 31.3; tibia, 12.6; foot, 9.1; ear from crown, 7.7; from notch, 12; length of tragus, 5.8.

Skull, total length, 13.7; condylobasal length, 13.1; zygomatic breadth, (?); mastoid breadth, 7.6; interorbital breadth, 4.7; breadth of braincase, 6.7; maxillar breadth, 5.4; breadth at canines, 4; upper toothrow (c-m³), 4.5; length of mandible, 9.4; angle to condyle, 1.7; depth at coronoid, 3.4; lower toothrow, 4.8.

Eptesicus garambæ, in size and external characters, resembles *E. ugandæ* Hollister, from Ledgus, Uganda, but exceeds it considerably in size, and differs from it in the form of the tragus and other characters, the last caudal vertebrae extending half its length beyond the membrane instead of wholly enclosed within it as in *ugandæ*. The skull proves, on comparison with topotype skulls of *ugandæ*, to not only exceed it greatly in size, but is much broader and flatter, and the rostral portion much broader, the skull as whole much more massive and depressed.

Eptesicus hottentotus bensoni Roberts, 1946

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Differs from *E. h. hottentotus* (A. Smith, *S. Afr. Quart. J. ser. 2*, no. 1, pt. 2, p. 581; 1833: Uitenhage and Albany districts), which I have pointed out in *Samab* (III, no. 8, p. 238, Dec. 1944) is an earlier name than *E. megalurus* (Temminck, *Monogr. de Mamm.* II, p. 206, 1835-41: 'Interior of South Africa'), in its slightly darker colour and in its smaller skull. Probably Shortridge's '*Scotophilus angusticeps*' (*Ann. S. Afr. Mus.* XXXII, 282; 1938: Hex River Estate, Citrusdal, Cape Province) is also a synonym of *E. h. hottentotus*, as it was placed in the wrong genus, as I found to be the case upon examination of the type; and also a specimen from the Cape Peninsula, in the South African Museum, does not differ materially from it and both practically conform to the descriptions of the colour by Andrew Smith and Temminck. In 1941 I collected a series of specimens on Eronga Mountain, Omaruru District, South-west Africa, which agree very well with the type of Shortridge's *E. megalurus pallidior* (*Ann. S. Afr. Mus.* XXXVI, 37; 1942: Goodhouse on the Orange River), and, although this form has somewhat longer wings than the typical southern one (wing-span in five measured from Eronga Mountain 340 - 365, as against about 12 in. = 304 according to Andrew Smith and 308 - 322 according to Temminck for *hottentotus*), the length of forearm, tibia and foot (c.u.) and skull proportions are much the same in all of them, whereas they are less in this new Nyasaland form. In colour *E. h. bensoni* is near 'tawny olive' for the terminal fourth, and blackish for the basal three-fourths of the hairs of all the upper parts, and below paler, the hairs being slightly tipped with whitish from chest to abdomen, with the pubic region and hairs along the margin of the wing membrane at the abdomen distinctly white. Both above and below a shade darker than the specimen from the Cape Peninsula. In the skull, which is unfortunately lacking the occipital part, the width of the muzzle is less, and the length of tooth rows and mandible shorter, as will be seen upon comparison of the figures given hereunder. The measurements taken by the collector of the specimen in the flesh do not indicate much difference between it and those of the typical form and *pallidior*, except the hind foot is shorter, which is confirmed by the measurements of the tibia and foot (c.u.)

taken from the dried skins. The forearm length is within the range of *hottentotus* and *pallidior*, but the digits conform more to those of the former.

Measurements of the type, with those of records and other measurements in brackets for comparison: Head and body 65 (of *hottentota* A. Smith 65, of *minuta* A. Smith 65, of *megalurus* Temminck, 'Pied du Roi' 2 in. 3 lines = 63 mm., of *angusticeps* Shortridge 77, 74, of *pallidior* Shortridge 74, of fourteen from Eronga Mountain 65 - 78); tail 57 (of *hottentota* A. Smith 50.6, of *minuta* A. Smith 50.6, of *megalurus* Temminck 56, of *angusticeps* Shortridge 58, 57, of *pallidior* Shortridge 48, of fourteen from Eronga Mountain 46 - 56); hind foot (c.u.) 8 (of *angusticeps* Shortridge 12, 11.5, of *pallidior* Shortridge 11, of fourteen from Eronga Mountain 9 - 11); ear 17 (of *megalurus*, according to Dobson, 15.5, no doubt shrunken in spirits, of *angusticeps* Shortridge 18, 17.5, of *pallidior* Shortridge 19, of fourteen

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from Eronga Mountain 15.5 - 20). Forearm 49.5 (of *minuta* A. Smith 1 in. 8 lines = 42 mm., obviously wrong when it had an expanse of wings of 12 in. (304 mm.), the same as he gave for *Scotophilus dinganii*, a bat of much the same size; of *megalurus* Temminck 1 in. 10 lines ('Pied du Roi') = 50 mm.; of *angusticeps* Shortridge 53 - I made it 51.5; of *pallidior* Shortridge 53.5; of fourteen from Eronga Mountain 48 - 52.5; the specimen from the Cape Peninsula 48). 3rd digit metacarpal 45 (type of *megalurus*, according to Dobson, 43, of the Cape Peninsula specimen 44.5, of the type of *angusticeps* 49, of the type of *pallidior* 52, of fourteen from Eronga Mountain 48 - 53); 4th digit metacarpal 44 (of the type of *megalurus* according to Dobson 39, of the type of *angusticeps* 47.5, of the type of *pallidior* 50.5, of fourteen from Eronga Mountain 46 - 53). Tibia and foot (c.u.) 27 (of *megalurus* according to Dobson 31.3, of *angusticeps* type 31, of *pallidior* type 29, of fourteen from Eronga Mountain 28 - 32). Skulls were not recorded by Andrew Smith, Temminck and Dobson; but I have measured the type skulls of *angusticeps* and *pallidior*, the specimen from Cape Peninsula and the fourteen from Eronga Mountain, and get the following figures: greatest length, *angusticeps* 21.3, Cape Peninsula 21.3, *pallidior* 20.7, Eronga Mountain 20 - 21.6; length from canines to condyles, *angusticeps* 19.2, Cape Peninsula 19, *pallidior* 18+ (incomplete), 14 Eronga Mountain 18.2 - 20.7; dorsal median length from frontals, *angusticeps* 18, Cape Peninsula 11.8, *pallidior* 17.7, 14 Eronga Mountain 17.5 - 18.5; zygomatic width, *angusticeps* 13.5, Cape Peninsula 13.7, *pallidior* 13.5, 14 Eronga Mountain 13.2 - 14.2; mastoid width, *angusticeps* 10.5, Cape Peninsula 9.5 (?), *pallidior* 10.5, 14 Eronga Mountain 10.2 - 11; width across molars, *benisoni* 8, *angusticeps* 9, Cape Peninsula 8.7, *pallidior* 8.3, 14 Eronga Mountain 8.1 - 8.8; width across upper canines, *angusticeps* 6.5, Cape Peninsula 6.9, *pallidior* 6.5, 14 Eronga Mountain 6.2 - 6.8, *benisoni* 5.8; upper c-m³ series, *angusticeps* 7.5, Cape Peninsula 7.5, *pallidior* 7.5, 14 Eronga Mountain 7.2 - 7.7, *benisoni* 6.8; mandible length, *angusticeps* 15.5, Cape Peninsula 15, *pallidior* 14.6, 14 Eronga Mountain 14 - 15.6, *benisoni* 13.6; c-m₃ series, *angusticeps* 8.5, Cape Peninsula 8.5, *pallidior* 8.5, 14 Eronga Mountain 8.1 - 8.7, *benisoni* 7.3.

Type: T.M. No. 9167, 6, Ncheu, Nyasaland, collected and donated by Mr C. W. Benson.

Eptesicus hottentotus portavernus Schlitter and Aggundey, 1986

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Holotype: Adult male, skin and skull, Carnegie Museum of Natural History, No. 89000, from Hell's Gate Canyon (= Njorowa Gorge), 20 km S, 14 km W Naivasha, Naivasha Dist., Rift Valley Prov., Kenya (0° 54' S, 36° 19' E); obtained 30 September 1985 by Duane A. Schlitter, original no. 6906.

Paratype: Adult female, body in alcohol with skull removed, United States National Museum of Natural History no. 375919, from Njorowa Gorge, near Lake Naivasha, Rift Valley, 6400 feet, Kenya (0° 55' S, 36° 19' E); obtained 20 September 1965 by A.D. Forbes-Watson, original number, 20 ix/l.

Measurements: Selected external measurements of the holotype and paratype respectively are: Total length, 125, 112; length of tail, 54, 48; length of hind foot, 12, 10; length of ear from notch, 19, 16; length of forearm 49., 52.3; and weight, -, 15. Cranial measurements are listed in table 2.

Diagnosis: External and cranial size large, palate short, and braincase deep for species.

[tables removed - eds.]

Description: Skull large for species; braincase relatively deep; rostrum short, flattened, lacking pronounced concavity in dorsal aspect; pterygoid region relatively narrow; inner upper incisor shallowly bifid in unworn paratype; outer upper incisor small and single cusped. Externally large for species; ear relatively small and with anterior margin nearly straight leading to rounded tip; tragus with notch at posterior base and posterior margin rounded to rounded tip; calcar keeled. Color of dorsal pelage of holotype medium fawn basally with light brown tips on hairs; ventral pelage dark brown basally with light tan tips. Color of dorsal surface of wing and interfemoral membranes medium brown with color lightening on uropatagium; ventral surface of wing membranes dark brown, lightening to fawn on propatagium. Ears same color as wing membranes.

Comparisons: *Eptesicus hottentotus portavernus* can be distinguished from the geographically adjacent subspecies *E. hottentotus benisoni* by the combination of large size of forearm and skull and short palate and deep braincase in the new subspecies. The

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new subspecies is paler in color than *E. h. benisoni*. From the nominate subspecies, the new subspecies can be separated by a deep braincase and short palate relative to overall similar size of the cranium, *E. h. portavernus* can be distinguished from *E.*

serotinus from North Africa and Eurasia by the long and broad palate of the latter species. *E. platyops* differs from the new subspecies by being small in length of forearm and skull.

Remarks: In *E. serotinus* and *E. hottentotus* individual variation was found in the degree of the bifid condition of the inner upper incisors. This variation would render this character less certain as a distinguishing character for species of large serotines than was used by Hayman and Hill (1971: 42) to key *E. hottentotus* from *E. platyops* and *E. serotinus* in Africa.

Etymology. - This new subspecies is named after the English name of Hell's Gate which is commonly used for Njorowa Gorge.

Eptesicus megalurus pallidior Shortridge, 1942

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Specimens from Goodhouse.

A relatively large buff-coloured bat with cranial characters as in *Eptesicus*; agreeing with the type of *Eptesicus megalurus* in skull and skin dimensions, but not altogether in colour.

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General colour above and on sides of neck pale rufous-buff; under parts drabby buff-white: all hairs above and below ashy-slate at base. Ears dusky brown: membranes horn-brown, rather translucent.

These Goodhouse specimens agree with Temminck's description of *megalurus* in that the hair is long, smooth, silky, and bicoloured throughout; but in typical *megalurus* the hair in front of the neck and abdomen is described as "cedar-brown" as far as the tip; on the flanks "dove-coloured," and on the pubic region quite white from base to tip.

Type: an adult female (Coll. No. 1157): H. & b. 74, Tl. 48, Hf. (s.u.) 11, Ear 19 mm.

A weak flyer, coming out at late dusk, about half an hour after *Cistugo* and *Platymops*. Apparently not plentiful around Goodhouse, about half a dozen observed in all.

The specimens collected were shot whilst "planing" very slowly in wide circles around a cattle kraal, to which they were attracted by swarms of flies.

These bats have a wide and broad wing span and, owing to similarity in size, were at first mistaken in flight for *Scotophilus*.

Eptesicus melckorum Roberts, 1919

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With the general characteres of *E. capensis* (A. Smith), but differing there from in its much longer tail and larger size; the tail measures 40-45 mm. as against only 30-34 mm. in *E. capensis*, the forearm 35.5-37 as against 32-36, and the skull 14.5-15 as against 14. In colouration the upper parts are externally "avellaneous" (Ridgway) with the base of the hair brown, and the under parts paler, tending to white, with the base of the hair darker brown.

Type, adult female, T. M. no. 2283, taken at Kersfontein, Berg River, Cape Province, 21st November, 1917. Also four more adults and a very young specimen captured at the same time in a loft.

Dimensions: Head and body 53, tail 40, hind foot (c.u.) 7, ear 15 mm. Forearm 37; pollex (c.u.) 5.5; 3rd digit, metacarpal 36, 1st phalanx 13.2, 2nd 12.8, 3rd 9.9; 4th digit, metacarpal 35, 1st phalanx 12.2; 5th digit, metacarpal 35.8, 1st phalanx 10.5. Tibia 14. Tragus (dry) on the outer margin 6.3, breadth 2.

Eptesicus phasma G.M. Allen, 1911

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Type. - Skin and skull, male, no. 8279, M.C.Z., collected August 6, 1909, on the Meru River, British East Africa, by Glover M. Allen.

General characters. - A small pallid species (forearm 34), with white membranes; possibly related to *E. tenuipinnis*.

Description. - Color of fur above from nose to root of tail pale dust color, between buff and cream-buff of Ridgway's Nomenclature (1886), slightly darker about the muzzle. Below, the hairs of the chin are buff to the bases paling on the throat to cream-buff and on the belly to pure white. The extreme bases of the hairs of the back, chest, and belly are dark slaty but those of the flanks ventrally are pure white throughout.

Wing membranes from the base of the first metacarpal, extremely thin and delicate; both interfemoral membrane and the wings are whitish throughout, the latter, however, becoming slightly clouded at their tips. Ears, forearms, feet, and tibia pale

brownish. The hair of the body extends on to the dorsal side of the interfemoral membrane and on the base of the tail slightly beyond a line joining the knees. Calcar long and slender with a well-marked low and elongate lobe; the calcar does not terminate in a lobule. Tail vertebrae entirely included within the membrane.

Ears small, triangular, their apical margins nearly straight, narrowly rounded at the tips; a prominent basal lobe at nearly right angles to the long axis of the ear at its inner margin; externally the basal notch is well developed and marks off a low rounded basal lobe. Tragus short, of nearly equal breadth throughout, bluntly tapering at its apex; its inner margin straight, its outer margin with a slight notch about opposite the inner base, thus producing two minute lobes at this point. Muzzle with rather conspicuous glandular swellings.

Measurements. - The following measurements of the type were taken in the flesh: - total length, 94 mm.; tail, 40.5; hind foot, 6; ear, 11.5; tragus, 5; forearm, 34; tibia, 12.5; calcar, 14. Skull: greatest length, 13.5; basal length, 10.5; palatal length, 5.4; interorbital constriction, 3.8; zygomatic width, 9; mastoid width, 7.8; upper cheek teeth, front of canine to back of m³, 4.8; lower cheek teeth, 5.2; mandible, 10.

Skull. - The skull is rather broad, with the anterior edges of the orbits conspicuously ridged so as to produce a rather squarish rostrum, tipper incisors directed sharply inward; the inner is simple, about twice the length of the outer; outer incisor with a conspicuous cingu-

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lum cusp externally. The two teeth are placed in a line nearly at right angles to the long axis of the skull. Canine with an anterointernal cingulum cusp. Crowns of the lower incisors trifid, slightly overlapping; first lower premolar about three fourths the length of the second.

Remarks. - This white-winged bat does not seem referable to any of the described species. It is possibly related to *E. tenuipinnis* of West Africa, but is larger, paler, the tail is entirely included in the membrane, and the tragus has two minute pointed lobules. It is equally distinct from *E. rendalli* (Thomas) from Gambia, which also has white membranes. The latter has large hind-foot pads, the calcar ends in a projecting point, and there is a penis bone present, which *E. phasma* does not have. The upper incisors are also different, being provided with two cusps in *rendalli*.

This bat was first noticed on the Guaso Nyiro at its junction with the stream from Meru, in the arid country to the northwest of Mt. Kenia. Here it was seen but once, when at dusk a single individual, conspicuous for its white coloration, was observed flying about over the river in company with numbers of small dark bats, probably *E. minutus somalicus*. A few miles farther up the Meru River (above the ivory-nut palm region) it was much commoner. At our camp by the side of this small stream numbers were observed and several were shot in the early evening as they flew past following the course of the rivulet, which here flows through a sun-baked plain with few trees except along the water course. Its flight is very steady, direct and rather slow, so that it was easily shot. In the gathering dusk, however, the white wing membranes were almost invisible against the sky, thus producing a curious ghostly effect, as only the body and forearms could be clearly seen.

Eptesicus rectitragus Wettstein, 1916

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Am ähnlichsten *E. pumilus* (Gray), aber kleiner, mit anders geformtem Tragus. Tragusform und Größe ähnlich der von *E. bicolor* (Bocage). Die kleinste bis jetzt bekannte Art dieses Genus. Kopf und Rücken dunkelbraun, "raw umber", seidig glänzend. Das einzelne Haar tief schwarzbraun mit "wood brown" gefärbter Spitze. Unterseite tief schwarzbraun, alle Haare mit lichten Spitzen. Jene auf Kinn und Kuhle mit licht bräunlichen auf der Brust mit grauweißen und am Bauche mit reinweißen Spitzen. Postcalcanallobes klein, gestreckt; Interfemoralmembran unterseits weißlichbraun, beginnt am Fußgelenk. Ohr reicht angelegt bis zur Schnauzenspitze. Tragus lang, schmal und fast gerade, oben viel weniger stumpf wie bei anderen Arten abgerundet. Seine größte Breite liegt in der Höhe der Basis des Innenrandes und beträgt 1.4 mm. Innenrand gerade, Außenrand an der Stelle der größten Breite schwach konvex. Lg. v. Kopf und Körper 35.5, Schwanzlg. 28.5, Unterarm 27, Ohrhöhe 10, Tragus 4 x 1.5 mm.

Typus: 1 M junior, Dilling, Südkordofan, 20.III.1914.

Eptesicus somalicus malagasyensis Peterson, Eger and Mitchell, 1995

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Holotype: ROM 42713, peau et crâne de femelle adulte de Maninday, 220 km au N.-E. de Tuléar [sous-préfecture de Sakaraha, sur la route nationale 7], 22°47'S, 44°43'E (13.V.1967).

Diagnose. - Petite forme proche de *E. somalicus* d'Afrique Orientale, à fourrure dorsale longue (environ 7 mm), assez dense, fortement bicolore: brun sombre à la base et Brun Bruxelles sur environ les trois cm apicaux, l'ensemble ayant une couleur uniforme et un aspect doux. Pelage ventral fortement bicolore, les 2/3 basilaires sombres et le tiers apical d'une valeur comprise entre l'Argile et l'Olive ou le Brun Cuir. Oreilles translucides et brun moyen. Queue relativement courte. Calcar à forte carène.

Crâne à rostre relativement long et aplani dorsalement à la jonction avec

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une boîte crânienne basse, faiblement arquée et un peu aplatie, comme chez *E. s. somalicus* (fig. 49). Mesures, cf. tableau 17.

Comparé à *E. s. somalicus* d'Afrique Orientale (Soudan, Ouganda et Nord du Kenya) l'holotype a un pelage un peu plus foncé sur le dos. Le pelage ventral est plus foncé (plus chamois, moins grisâtre) et n'est pas pâle ou blanchâtre dans la région pelvienne. Toutes les mesures se situent à l'intérieur de la gamme de mesures de *E. s. somalicus* sauf les largeurs zygomatique et crâniale qui sont un peu plus fortes.

Comparé à *E. matroka*, la tête et le pied sont plus courts, le pelage dorsal est plus pâle et plus nettement bicolore, le pelage ventral plus pâle, la portion apicale pâle est plus étendue. Oreilles plus pâles et plus translucides. Crâne à rostre plus étroit, région supra-orbitale moins renflée; boîte crânienne relativement plus longue, plus aplatie, moins arquée en profil dorsal et moins sphérique (cf. fig. 51 et tableau 18).

Comparé à *E. capensis* du Zimbabwe, la fourrure est plus longue et plus foncée, avec le ventre seulement un peu plus clair que le dos, au lieu d'être brun grisâtre pâle en dessus et d'avoir la partie terminale des poils ventraux blanc à chamois pâle. Crâne plus court; boîte crânienne plus profonde, plus arrondie, et légèrement arquée derrière le rostre, sans cimier postérieur, au lieu d'être entièrement plate dorsalement et se terminant par un cimier postérieur.

Comparé à *E. zuluensis*, la fourrure est plus courte, plus serrée et plus lisse et non longue et lâche; bourre brun foncé et non noirâtre. Partie terminale des poils du dos Brun Bruxelles et non Olive-Brun; celle des poils ventraux Olive-Brun et non blanchâtre, gris clair ou chamois ocre très pâle. Calcar à carène plus marquée. Crâne plus grand, à boîte crânienne plus large, moins bulbeuse et moins relevée; rostre plus long.

Distribution. - Connue seulement par l'holotype, cette sous-espèce est, peut-être, localisée à la partie sub-aride du Sud-Ouest malgache. C'est la seule espèce d'*Eptesicus* connue de la région sub-aride.

Variabilité systématique. - La parenté avec *E. somalicus* demandera à être précisée sur un matériel plus abondant. Comme le tableau 18 le montre,

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les sexes sont très dimorphes chez *E. matroka*, comme ils le sont chez *E. s. somalicus*, les femelles étant, en moyenne, beaucoup plus grandes, pour la plupart des caractères.

Biologie. - L'holotype a été pris au filet au-dessus d'un ruisseau bordé de Pahniers, dans une savane ouverte. Cette sous-espèce semble, comme *E. s. somalicus*, occuper un milieu plus ouvert et plus sec que ne le font la plupart

[table removed - eds.]

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des autres petits *Eptesicus* qui paraissent. comme *E. matroka* à Madagascar, rechercher plutôt des stations boisées et plus humides.

L'holotype femelle était bien gras et ne montrait pas trace d'activité reproductrice (ni gravide, ni en lactation).

Remarques. - Comme nous l'avons dit plus haut, la validité de *Vesperus humbloti* A. Milne Edwards, 1881, ne peut être établie. Aucun holotype ne semble plus exister et la description originale n'établit de comparaison qu'avec *V. minutus*, autre nom qui ne répond à aucune description satisfaisante. *V. humbloti* est dit plus grand que *V. minutus*, avec les vertèbres caudales inférieures enfermées dans la membrane interfémorale. Une telle description peut s'appliquer à de nombreuses espèces mais ne correspond pas à un taxon particulier.

Eptesicus ugandae Hollister, 1916

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Type from Ledgus, Uganda. United States National Museum No. 166520, adult female in alcohol (skull removed). Collected February 15, 1910, by J. Alden Loring. Orig. No. 9022.

Description. - A small brown species related to *Eptesicus capensis somalicus* (Thomas), but with larger, flatter skull. Ears broad, somewhat evenly triangular, with rounded tip; when laid forward reaching to tip of muzzle. Tragus comparatively long, broadest at center, the tip bluntly rounded and not turned forward; inner side nearly straight for two-thirds its length; outer side evenly convex from tip to opposite anterior notch, where there is a small sharp lobe, beneath which is a sharply concave margin and a larger triangular basal lobe, immediately above the outer notch. Wing from base of toes; tail vertebræ entirely within the interfemoral membrane, but a small fleshy tip extending very slightly beyond; postcalcaneal lobe comparatively small, sharply emarginate anteriorly and evenly sloping posteriorly. Hair extending only slightly on to wings but thinly covering the interfemoral membrane to line of ankles and, along tail, to slightly beyond. Color (from alcoholic specimens) apparently much as in *somalicus* but somewhat darker throughout; wings dark grayish brown, faintly-lined along posterior edges with buffy; interfemoral membrane slightly lighter than wings. Skull decidedly larger than in *somalicus* (as represented by specimens from the Northern Guaso Nyiro) with relatively and actually broader rostrum and braincase, and relatively much lower sinciput. Upper inner incisors broad and heavy, distinctly bifurcate at tip (except in a much worn specimen); outer upper

incisors small, barely reaching beyond cingulum of inner incisors. Lower incisors all trifold, considerably crowded in the row. Cheek teeth essentially as in *somaticus* but slightly more robust.

Measurements. - Type: Forearm, 29.6 mm.; outer height ear, 12.3; greatest breadth ear, 8.1; tragus from outer notch, 5.4; third finger metacarpal, 26.8; first phalanx, 10.3; fourth finger metacarpal, 27.7; first phalanx, 9.2; fifth finger metacarpal, 28.1; first phalanx, 7.3; tibia and foot, including claws, 16. Skull: Greatest length, 12.8; condylobasal length, 11.8; breadth of braincase, 6.8; depth of braincase, 4.6; mastoid breadth, 7.3; postorbital constriction, 3.5; mandible, 8.8. Teeth: Upper maxillary row, 4.3; breadth across upper canines, 3.9; entire lower row, 5.8.

Remarks. - In addition to other characters, this species may readily be separated from the other small forms of *Eptesicus* known in East

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Africa by its dark-colored wing membranes [distinguishing from *tenuipinnis*, *rendalli*, and *phasma*], short outer upper incisors [distinguishing from *grandidieri*], large, flattened skull, and distinctly bifurcate inner upper incisors [distinguishing from *somaticus*]. In addition to the type there are two topotypes and three other specimens from Gondokoro in the collection.

Eptesicus zuluensis Roberts, 1924

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Eptesicus zuluensis spec. nov. (text-fig. 1).

In external appearance very similar to *Eptesicus capensis* taken in the same place, but with a rather longer tail, and the skull smaller and deeper in the cranium even when compared with *E. capensis gracilior* Thomas and Schwann (from Eshowe, Zululand). In its deep cranium it somewhat resembles *Glischropus nanus* (Peters), but differs therefrom in the absence of the small upper premolar and in dimensions of both body and skull. In the size of the skull it is about the same as in *Pipistrellus kuhli fuscatus* Thomas, but differs from this also in the number of teeth, narrower muzzle and deeper cranium. It is altogether an interesting animal, whose affinity to our known species is not at all clear, the longer tail seeming to point to its affinity to the longer tailed

-Fig 1 [removed]-

-Fig 2 [removed]-

species *E. smithi* Wagner (Schreber, *Sauget. Suppl.* v, p. 747, footnote) and *E. melckorum* Roberts (*Ann. Transvaal Mus.* vi, p. 113), but for the skull being quite differently shaped. The following table of dimensions will serve for comparison of the type specimen with the dimensions given for the type of *gracilior*, of another specimen of the same form from Mokeetsi (N.E. Transvaal), and a series of *E. capensis* from a wide range of localities, including the type locality of *zuluensis*:

Type of *zuluensis*; Type of *gracilior*; Mokeetsi specimen; Series of *capensis*

Head and body: 45; 47; 44; 49-55

Tail: 37; 28; 30; 30-34

Forearm: 30.5; 29; 30.3; 32-35

Canines to occiput: 12.8; 13.2; 13.2; 13.7-14.5

Basilar skull length: 9.5; 10.1; 10; 10.4-10.8

Mastoid width: 7.5; 7.6; 7.8; 7.6-8

Width across M2: 5.1; -; 5.6; 5.6-6.2

Palate length: 4.6; -; 5.5; 5.3-5.5

Upper tooth row (canine to M3): 4.4; -; 4.7; 4.7-5.

Lower tooth row: 4.9; 5.1; 5; 5-5.5

Posterior height at bullae: 6.1; -; 5.3; 5.3-5.8

The slightly raised cranium is the most marked character of the species, all members of the *Eptesicus capensis* group having the cranium on a level with the muzzle.

Type: Adult female, T.M. No. 3024, taken in the White Umfolosi Game Reserve, on June 29th, 1922. It was shot in the evening while flitting past

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my camp. A specimen of the typical *E. capensis* (text-fig. 2) was taken in an aloe at the same place, and has rather shorter hair, whereas another specimen of *capensis* taken on higher ground at Ntambanana has the hair and same external appearance, even to the shape of the ears, as the type of *zuluensis*.

Glauconycteris alboquittatus J.A. Allen, 1917

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Text Fig. 7, B

Type, No. 49317 (and only specimen) adult female in alcohol, Medje, March 14, 1914; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 2351.

Upperparts seal brown; underparts slightly paler. A patch of lengthened pure white hairs on each shoulder at base of humerus, about 8 mm. long by 5 mm. in width, separated for about 10 mm. by dark fur from a second patch of white hairs of similar extent but the area longer and narrower. They are symmetrical on both shoulders and conspicuous when the fur is

floated out in clear alcohol. Muzzle, chin and sides of face nearly naked and whitish. Propatagium and outer border of wing whitish; forearm and phalangeal bones light brown above and nearly white below. Ears blackish externally, whitish internally. Membranes and feet blackish brown, interfomoral lighter below than above.

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Total length, 94; head and body, 52; tail, 42; forearm, 41.5; third metacarpal, 38.6; thumb, 5.6; tibia, 18.3; foot, 8; calcar, 11; ear from crown, 7; from notch, 13; tragus, 5 mm. high by 2.5 in greatest width, broad lunate, with a small angular lobe at posterior base.

Skull, total length, 13.2; condylobasal length, 13; zygomatic breadth, 10; breadth of braincase, 7.4; interorbital breadth, 4.8; maxillar breadth, 6.5; breadth at base of canines, 4.8; upper toothrow (c-m³), 4.5; length of mandible, 9.5; angle to condyle, 2; depth at coronoid, 2.9; toothrow (c-m₃), 5.2.

Similar to *Glauconycteris humeralis* but larger (the forearm 4 mm. longer and the skull fully one half larger in volume), and with two clear white spots on the shoulder (one behind the other) instead of one, and a very much broader tragus. In size it is similar to *G. congicus* (Noack) but differs from it widely in coloration.

Glauconycteris beatrix Thomas, 1901

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A small blackish species of the group with brown unicolor wings.

Size rather less than in *G. poensis*. General colour above and below uniform blackish brown without lighter markings; wings and interfemoral membranes uniformly brown throughout. Muzzle broad and tumid; lobes at corner of mouth well developed. Inner margins of ears extremely convex forwards, forming, from the rounded basal lobe to the tip, approximately the half of a circle; tip scarcely perceptible; outer margin straight above, then convex, with a well-marked angular antitragal lobe.

Tragus short, fairly broad, its inner margin straight, its tip rounded, its outer margin evenly convex, with a well-marked basal lobule.

Wings to the base of the toes. No postcalcareal lobule. Tail included in membrane to its extreme tip.

Skull, as compared with that of *G. poensis*, smaller, more delicately built, and with a narrower muzzle.

Inner upper incisors with the secondary cusp longer and more widely separated from the main one than in *G. poensis*; outer incisor fairly large, pressed against the canines behind, and reaching vertically about half the height of the secondary cusp of *i*¹; in transverse section it appears to be about half the area of the same tooth. In *G. poensis* it is shorter, smaller in area, and is separated on each side from the canine. Lower incisors bifid, slightly overlapping. Anterior lower premolar with a long well-defined main cusp half the height of the posterior premolar.

Dimensions of the type (measured in spirit): -

Head and body 45; tail 43; ear 10; tragus on inner edge 3; middle finger, metacarpal 38, first phalanx 13.5, second phalanx 23; lower leg 19.5; hind foot (c. u.) 7.5; calcar 13.

Skull: occiput to gnathion 11.1; interorbital breadth 4.2; breadth across brain-case 7; front of canine to back of *m*³ 4.2.

Hab. Benito River, French Congo, fifteen miles from mouth.

Type. Female. B.M. no. 98.5.4.19. Collected January, 1898, by Mr. G. L. Bates.

The only near ally of this species appears to be *G. poensis*, and from that it is readily distinguishable by its smaller size,

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darker colour, and the dental and cranial characters above described. *G. congicus*, Noack, is probably synonymous with *G. poensis*, the describer having been misled by Dobson's erroneous statement that the inner upper incisors of the latter species were unicuspid.

Glauconycteris egeria Thomas, 1913

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M. No. 22. Bibundi. B.M. no. 12.12.13.3. Type.

Brown-winged; the ears very large.

Size about as in *G. poensis*, though the skull is longer. General colour uniform dusky brown, not lighter below. Membranes also all dark brown, the reticulations not specially prominent. Ears much larger than in the allied species, subquadrangular,

the anterior basal lobule long, projecting backwards; anterior margin with its basal 6 mm. straight, then strongly projecting forward, almost to an angle; the

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upper half again straight; tip broadly rounded off; outer edge straight above, convex below; the outer basal lobe well developed, rounded. Tragus fairly large, its inner edge concave, outer convex; a well-developed angular external basal lobule. Lobes at angles of mouth rather small. Wings from the base of the toes. Calcar of medium length, a rudimentary postcalcareal lobule present.

Skull large in proportion to the size of the animal, its brain-case of the usual swollen shape, though not to so exaggerated an extent as in some species.

Inner upper incisors long, their distinct secondary cusp near their tips. Outer incisors minute, their tips reaching barely halfway to the secondary cusp of the inner pair. Lower incisors not markedly overlapping.

Dimensions of the type (measured on the spirit-specimen): -

Forearm 38 mm.

Head and body 43; tail 41; ear, from notch 13.3, from anterior projecting angle 11; greatest breadth 13.5; tragus, length on inner edge 4; third finger, metacarpus 36, first phalanx 14.5; lower leg and foot (c. u.) 24; calcar 15.5.

Skull: greatest length 13; basi-sinual length 10; zygomatic breadth 9.2; interorbital breadth 3.8; breadth of brain-case 7.5; palato-sinual length 5.3; front of canine to back of m^3 4.5; front of p^4 to back of m^2 2.2; breadth between outer corners of m^2 5.5.

Hab. and Type as above.

This well-marked species is readily distinguishable from the other two brown-winged *Glauconycteris*, *G. poensis* and *beatrix*, by its much larger and less evenly rounded ears and by the length of its skull, which is 13 mm. in length as against 12 mm. in *poensis* and 11 mm. in *beatrix*.

Peters's *Vesperus kraussi*, from Yoruba, Lagos, appears to agree with *G. poensis* in all essential respects. Its locality is almost identical, as the type of *G. poensis* came from the Lower Niger, and not from Fernando Po.

Glauconycteris Floweri de Winton, 1901

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AMONG the mammals collected in the Soudan by Mr. H. F. Witherby in the spring of this year and acquired by the late Dr. John Anderson is a small bat which appears to be new to science. The specimen, preserved as a skin, was obtained at Wad Mariun about 12 miles from Khartoum on the White Nile on May 12th, and will be taken as the type of the species. A second specimen, which has just been put into my hands by the kindness of Mr. O. Thomas of the British Museum, is preserved in alcohol; it was collected by Capt. S.S. Flower, Director of the Zoological Gardens at Cairo, on March 14th, at Abu Zeit on the White Nile some 200 miles south of Khartoum, during his recent trip with the party sent to inspect the Sud-cutting operations.

Although the front of the head is much damaged by shot, this "spirit specimen" is valuable in enabling a fuller description of the animal to be given than would have been possible if the dried skin alone had been available.

I have great pleasure in associating the name of so keen a worker in zoology, who also collected the first specimen, with this fine new species.

Dobson made a subgenus for the African bats allied to the Australian genus *Chalinolobus*, but I think it will be much more convenient to look upon these two geographically separated groups as distinct genera; the species from the two

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regions being so distinct from one another in general appearance, and have, besides the characters given by Dobson, the very important difference in the incisor teeth.

The first upper incisor in *Glauconycteris* is bifid, having an outer or rear cusp.

Glauconycteris Floweri, sp. n.

Colour above pale fawn, the bases of the fur greyish; the lower side cream-colour, yellower on the throat, paler on the belly. The fur on the forehead ends in a point barely in advance of the eyes, and the furred area is limited in extent in all directions, barely encroaching on to the upper arms, and leaving the tail and legs entirely bare. The skin of the face and ears is nearly black and naked, with the exception of a few short hairs more conspicuous round the lips. The wing-membrane is pale and transparent, with the veins and lines showing slightly darker; the posterior portion in front of the tibiae and the distal portion of the interfemoral membrane is yellowish white. The upperside of the forearms, legs, and tail, and all the proximal portion of the wings and interfemoral membrane, are thickly studded with dark warty papillae.

The measurements of the type taken by the collector from the animal in the flesh, an adult male, are: Head and body 47 millim., tail 34, hind foot 5.5, ear 12, forearm 35.

The specimen in spirit gives the following measurements : -
Head and body 42, tail 34, hind foot 5.8, ear 12, forearm 36.5.

In size and general colour this bat somewhat resembles *Scotophilus Schlieffeni*: the body-colour is, however, paler above and brighter on the lower parts, and the furred area is less extensive; but the paler wing-membrane, the dark face, and the extraordinary shagreening on the forearms, tail, and surrounding membranes distinguish this new species at a glance.

The skull is readily distinguished from that of *Scotophilus* by its rounded form and short facial portion; the lower jaw is likewise more rounded or blunter.

Glauconycteris gleni Peterson and Smith, 1973

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Holotype - ROM 57125, adult female, skin and skull collected 16 June 1970 by Robert Glen and V. Ngam in a tropical forest region near Lomie, Cameroun, 3°10'N, 13°37'E.

Geographic distribution - Known only from the type locality in Cameroun and from Malabigambo Forest near Sàngo Bay, Masaka District, southern Uganda, 1°05'N, 31°30'E.

Diagnosis - Wings and uropatagium white with dark brown pigment outlining the bones of the wings, hind limbs, and tail, the basal portions of the calcars and veinations of the interfemoral membrane, and to a lesser extent the proximal areas of the wings between humeri and hind limbs (Fig. 1). The ears are relatively large for the genus and have broad tragi. The pelage of the dorsum is quadricoloured with a pale clay-coloured terminal band, a narrow and dark-brown subterminal zone, a pale, buffy, sub-basal zone, and a dark slate-gray basal band. There are no distinct dorsolateral bands of pale pelage as in *G. argentata*. Ventral hairs are bicoloured, with the base gray and the whitish terminal band varying from pure white to a wash of pale yellowish buff. The length of the tibia relative to forearm length is short compared with other species of the genus (Tables 1 and 2).

The size and general shape of the skull of *G. gleni* is similar to that of *G. variegata* but differs from the latter by the combination of a more rounded braincase (equal in

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[Tables removed - Eds.]

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width to that of *G. variegata*) and a narrower mastoid width. The upper inner incisor is usually bilobate, but the development of the posterior lobe varies considerably in individual specimens. The upper premolar has a large hypocone with its lingual margin lying in line with the lingual margins of the canine and the first molar (M^1) in occlusal view (Fig. 2). The lower incisors are typically trilobate (Fig. 3), but the outer incisor (I_3) has a well-developed cingulum extending along its anterior surface below the central lobe. The occlusal surfaces of I_3 and I_2 are each characterized by the development of a posterior projection of the cingulum to form a heel-like lobe (Fig. 4).

Measurements (mm) of the holotype - *External*: Total length 100, tail vertebrae 46, hind foot 8, ear 16, tibia 16.4, forearm 41.1, third digit - metacarpal 40.2, first phalanx 15.2, second phalanx 21.3, fourth digit - metacarpal 37.8, first phalanx 10.9, second phalanx 9.7, fifth digit - metacarpal 34.9, first phalanx 7.8, second phalanx 6.9, wing span 305, weight 9.25 grams. *Skull*: Greatest length 14.2, condylobasal length 13.6, palatal length 4.1, zygomatic width 9.9, mastoid width 8.4, breadth of braincase 8.2, height of braincase 5.9, supraorbital process width 5.3, interorbital width (from front) 5.2, postorbital constriction 4.5, width across molars (M^3-M^3) 6.4, length of toothrow (C- M^3) 4.75, width across upper canines (C-C) 4.6, length of mandible (condylo-incisive) 9.9, length of lower tooth row (C- M_3) 5.2, width across lower canines (C-C) 3.0.

Specimens examined - *Glauconycteris gleni* sp. N. - The holotype from Cameroun (ROM); plus nine adult and two subadult males, five adult and one subadult females all from Malabigambo Forest near Sàngo Bay, Masaka District, southern Uganda in the collection of the Los Angeles County Museum. *G. argentata* (Dobson) - Cameroun, 8; French Congo, 2; Uganda, 6; Kenya, 49; Tanzania, 5. *G. variegata* (Tomes) - Cameroun, 2; Congo, 11; Dahomey, 2; Ghana, 2; Nigeria, 2; Zambia, 2; Sudan, 1; Uganda, 9; Malawi, 1; South Africa (Zululand), 1. *G. alboguttatus* J.A. Allen - Cameroun, 1; Congo, 1. *G. beatrix* Thomas (including *G. humeralis* J.A. Allen) - French Congo, 1; Gabon, 1; Congo, 5; Uganda, 9; Kenya, 6. *G. egeria* Thomas - Cameroun, 1; Uganda, 4. *G. machadoi* Hayman - Angola, 1. *G. superba* Hayman - Congo, 1; Ghana, 1; Ivory Coast, 1.

Comparisons - *Glauconycteris gleni* appears morphologically intermediate between *G. argentata* and *G. variegata*. Externally, *G. gleni* resembles a pale-coloured *G. argentata* in the venation of the interfemoral membrane, but markings are considerably darker and the membrane much whiter. *G. gleni* lacks the dorsolateral bands of pale-coloured pelage on the back, characteristic of *G. argentata*, and the white ventral pelage contrasts sharply with the dark-greyish or brownish coloration of that species. The ear of *G. gleni* is larger, the tragus is wider, and the tibia is distinctly shorter than in *G. argentata*. *G. gleni* is significantly larger in 11 of 16 external measurements in both sexes (Tables 1 and 2). In cranial characters, *G. gleni* is significantly larger ($P < .001$) without overlap in measurements of greatest length, condylobasal length, length of tooth row (C- M^3), and length of mandible, while the postorbital constriction of *G. gleni* is significantly narrower ($P < 0.001$) in both sexes compared with *G. argentata* (Tables 1 and 2). Of the 16

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cranial characters compared, *G. gleni* and *G. argentata* males differed significantly in 12 and females in 10. In dental morphology, specimens of *G. gleni* tended to differ from those of *G. argentata* in having a larger hypocone on the upper premolar and in the development of the cingula on the anterior surface of the crown of 13, as well as on the posterior portion of both hand 13 (Figs. 2 - 4).

When compared with *G. variegata*, *G. gleni* lacks the heavily pigmented markings on the wings, and the markings on the interfemoral membrane are less complex (typically, lines are laterally oriented and anteroposterior orientation of cross-veination is limited). The coloration of the pelage is strikingly different from any known example of *G. variegata*. The dorsal pelage is distinctly quadricoloured instead of tricoloured as in *G. variegata*. Although similar in shape, the ears are longer and the tragi are broader in the new species. The tibia of *G. gleni* is distinctly shorter (17.0 or less) than that of *G. variegata* (18.3 or more; see Tables 1 and 2). Of the 16 external measurements (Tables 1 and 2), *G. gleni* males are significantly smaller in nine and females in eight when compared with *G. variegata*. Although the crania of the two species are similar in size, differences are significant in 13 of the 16 measurements in males and 10 of the 16 in females. In dental morphology, *G. gleni* also differs from *G. variegata* by the shorter first commissure and longer third commissure of M^3 and by the greater development of the hypocone of P^4 (Fig. 2), as well as by minor differences in the development of the cingula of the lower incisors (Figs. 3 and 4).

Each of the remaining known species of *Glauconycteris* can be distinguished from *G. gleni* as follows:

G. poensis (Gray) - wings pale gray instead of white, general pelage coloration darker, and greatest length of skull less than 13.0.

G. albugutattus Allen - pelage coloration much darker, wing membranes dark instead of white, tibia greater than 17.0, and greatest length of skull shorter (13.1 and 13.2).

G. beatrix Thomas - pelage and wing membranes dark, ear shorter (12.0 or less), skull shorter (greatest length, 12.0 or less), and length of mandible less than 9.0.

G. egeria Thomas - pelage and wing membranes quite dark combined with striking whitish dorsolateral stripes on the back, larger square-shaped ears, greatest length of skull less than 13.5, and length of mandible less than 9.5.

G. machadoi Hayman - darker pelage coloration, as well as wing and interfemoral membranes with sharply defined veinations; otherwise differs in the same way as *G. variegata*.

G. superba Hayman - pelage black and white above, black below, forearm length greater than 44.0, greatest length of skull more than 16.0.

Glauconycteris humeralis J.A. Allen, 1917

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Text Fig. 7, A.

Type, No. 49013, F ad, (skin and skull), Medje, Belgian Congo, Sept. 7, 1910; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 956..

Size medium. A conspicuous tuft of white hairs at the shoulder. Pelage full and soft.

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Upperparts hair-brown, the hairs darker on basal half; a conspicuous tuft of white hairs on the shoulders; underparts similar to back but paler; membranes dark brown; limb bones pale brown on both surfaces; wing membranes attached to the whole length of basal joint of outer toe; no postcalcaneal lobe. Ears small, yellowish brown; tragus short, broad, straight on inner border, convex on outer border, expanded at middle. Tail wholly included in the interfemoral membrane.

Represented by 5 specimens, the type and 3 topotypes (2 of them in alcohol) from Medje, the other from Avakubi, collected Feb. 21, 1914.

Collectors' measurements of type: Total length, 82 mm.; head and body, 42; tail, 40; foot, 8; ear, 9.

Forearm, type (from skin), 36.8; third metacarpal, 35.8; tibia, 26.8; foot, 7; ear, 5 X 6; tragus, 3 X 1. The forearm length in 2 alcoholic topotypes is, M 35.8, F 35.3; the Avakubi specimen, 38.8.

Skull (type), total length, 11.3; zygomatic breadth, 8.2; breadth of braincase, 7.3; interorbital breadth, 3.9; maxillar breadth, 5.1; breadth at base of canines, 3.6; upper toothrow (with canine), 3.6; length of mandible, 7.9; height at coronoid, 2.5; lower toothrow, 3.9. Skull short and very broad, the braincase greatly expanded.

Inner incisor large, with the secondary column about one fourth lower than the main cusp; outer incisor inserted posterior to the inner one, minute, about one third of the height of the inner, and wedged in between inner incisor and canine. The single upper premolar is approximately the size of m^3 , and quite similar to it in form.

The pure white shoulder tuft is a conspicuous feature in the type and topotypes; it is present in the Avakubi specimen, but only the tips of the hairs are white (yellowish white instead of pure white). The collectors' measurements and the forearm are somewhat larger than in the type, but the skull measurements are practically the same in all the specimens.

Glauconycteris kenyacola Peterson, 1982

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Figs. 1 - 4

Holotype

ROM 71305 adult female skin and skull, collected 30 December 1970 on the Galole Road, 8.5 km N Garsen, Kenya (02°12' S, 40°05' E) by Judith Eger and Robert Glen.

Geographical distribution

Known only from the type locality.

Diagnosis

Similar in size to *G. argentata* but with distinctive whitish facial markings on nose and chin and at base of ears (Fig. 1); skull with narrow postorbital constriction, short rounded braincase (Fig. 2); inner upper incisor simple (without secondary cusp but with a slight posterior bladelike ridge) resembling that of *G. variegata* (Fig. 3B); third upper molar with relatively long first commissure, with narrow angles between first and second and between second and third commissures, the lingual termination of the latter not extending conspicuously beyond level of the junction of the first and second commissures (Fig. 4B),

Description

A whitish tuft of hair extends dorsally on the nose from between the nostrils (tuft length about 3 mm and width about 2 mm); a distinct thick band of white hairs (about 1.5 mm long) extends from the antitragal lobe to the base of the ears (band about 2.0 mm x 5.0 mm); tufts of finer, longer, and darker hairs occur on the posterior surface of the lower outer base of the ears as well as the

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posterior outer surface of the base of the leading edge of the upper (inner) portion of the ear; inner surface of the ear and base of tragus with a thin covering of long pale hairs.

Dorsal pelage long (about 5 - 6 mm), soft and bicoloured, Tawny-Olive basally and Mummy Brown at tips; head Dresden Brown, paler than back Except for the lateral flanks, which are Yellow Ocher, the ventral pelage is pale Ochraceous Buff with a thin wash of Dresden Brown.

Wing membranes of dried skin Light Ochraceous-Buff with venation outlined in Dresden Brown; uropatagiurn Drab. Skin covering wing, leg, and tail bones strongly outlined with Blackish Brown dorsally but not ventrally.

See Table I for measurements of the holotype.

Comparisons

Compared with *G. argentata*, *G. kenyacola* differs in its distinctive facial markings; dorsal pelage darker and lacking light lateral flank markings (dorsolateral bands); skin over wing, leg, and tail bones sharply outlined with Blackish Brown dorsally; a shorter second phalanx of third digit; skull with shorter palate; slightly narrower postorbital constriction; absence of secondary cusp on unworn first (inner) upper Incisor (Fig. 3); M³ more compressed (anteroposteriorly) with relatively longer first commissure and smaller angles between it and the second as well as between the second and third.

Compared with *G. gleni*, the holotype of the new taxon differs in the presence of distinctive facial markings; wing membranes light Ochraceous-Buff instead of white (skin covering wing, leg, and tail bones similarly strongly outlined dorsally); dorsal and ventral pelage much darker; longer tibia, tail length, fifth digit metacarpal, and second phalanx of fifth digit; shorter second phalanx of third digit; lighter weight; shorter greatest length of skull, condylobasal length, palatal length, C-M³, C-M₃, and mandible lengths; narrower, lower C-C width; M³ with a more acute angle between the first and second commissures (Fig. 4B).

Compared with *G. variegata*, *G. kenyacola* lacks the characteristic wing membrane pattern of dark outlining of the venation (above and below); differs in the presence of the distinctive facial marking; pelage darker dorsally and ventrally; wing size smaller; weight skull smaller throughout (see Table I); a more acute angle between the first and second commissures of M³ (Fig. 4B).

Compared with a series of 14 female and 21 male specimens of *G. poensis* from western Africa, *G. kenyacola* differs in its facial markings and pelage colouration in much the same way as from *G. argentata* except that *G. poensis* averages much darker grey than *G. argentata*. In size the new taxon has a longer tibia (20.0 vs 18.8 maximum), forearm (40.5 vs 39.7 maximum), fourth digit metacarpal (38.7 vs. 38.5 maximum), fifth digit metacarpal (37.3 vs 34.8 maximum). Cranially it has a wider zygomatic Width (ca. 9.4 vs. 9.1 maximum) and a wider braincase (8.0 vs. 7.6 maximum). The upper incisor lacks the strongly developed posterior cusp that is even more highly developed in *G. poensis* than in *G. argentata* (Fig. 3A).

Remarks

G. kenyacola is so uniquely distinct from all known taxa of *Glauconycteris* from Africa that it cannot be confused with any other species. It is most comparable with *G. argentata* and *G. poensis* in size, and needs further comparison with only *G. gleni* and *G. variegata*.

The range of normal geographical, sexual, and age variation within this new taxon will remain unknown until additional specimens become available. The name *Glauconycteris* as a genus is provisionally retained and follows Peterson and Smith (1973) and Corbet and Hill (1980).

Only a few specimens of *Glauconycteris* have been taken near the type locality of *G. kenyacola* and these consist of *G. argentata* and *G. variegata*. As there has been rather extensive collecting in other regions of Africa without its discovery, it appears possible that that new taxon may be restricted to the lower coastal plains of Kenya and adjacent regions. The holotype was netted over a seasonal waterhole.

The etymology of the name *kenyacola* is derived from the Latin *cola* (= to inhabit), thus "Kenya inhabitant."

[*Glauconycteris papilio*](#) Thomas, 1905

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Allied to *G. variegatus*, Tomes, with which it has hitherto been confounded ; but the acquisition by the British Museum of the co-types of that species in the Tomes collection enables me to clear up the confusion.

Size smaller than in *variegatus* (forearm 41 instead of 45). General colour of the ends of the hairs above uniform clay-colour from head to rump, exactly matching the posterior back of *variegatus*, but in the latter the head and shoulders are whitish, the clay-colour being only present posteriorly. Individually the hairs are pale slaty basally, then silvery cream-colour, the tips being pale brownish clay-colour. Under surface uniform cream-buff from chin to anus, the corresponding part of *C. variegatus* being white. Wings and interfemoral membrane variegated as in the allied species. Inner basal lobe of ear rounded.

Skull as in *G. variegatus*, but shorter, while nearly as broad. Inner upper incisors similarly unicuspid ; outer minute.

Dimensions of the type (measured in skin): -
Forearm 41 mm.

Third finger, metacarpus 40, first phalanx 15, second phalanx 27 ; fifth finger, metacarpus 38, first phalanx 9, second phalanx 9 ; lower leg and foot (c.u.) 27.

Skull: greatest length 13.2 ; median basal length 10.2 ; breadth of brain-case 8.2 ; front of canine to back of m^3 5.

Hab. (of type). Entebbi, Uganda. Other specimens from Gambaga, Gold Coast (*Giffard*), and Tette, Zambesi (*Boyd Alexander*).

Type. B.M. no. 99. 8. 4. 14. Collected 10th April, 1895, and presented by F.J. Jackson, Esq.

Mr. Tomes's collection contains six specimens of "*Scotophilus variegatus*," among which two labelled in his hand-writing can be identified as those of which he gave the dimensions in his original description*. Dobson's statement that the type of this species was in the Berlin Museum was therefore erroneous.

The beautiful "butterfly-bat" now described has been

* P. Z. S. 1861, p. 36.

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hitherto considered by Winton* and myself as *G. variegatus*, but it may be readily distinguished by the characters above described.

* Ann. & Mag. N. H. (6) xx. P. 317 (1897, and (7) iv. P. 354 (1899).

[*Glauconycteris phalaena*](#) Thomas, 1915

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Near *G. variegata*, but teeth smaller and incisors more slender.

Size as in *G. variegata*. Colour, of a spirit-specimen, apparently about as in that animal, the head and belly similar whitish, but the back hardly so strongly buffy. The colour is therefore still further from that of the neighbouring *G. variegata papilio*, in which both head and belly

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are more or less strongly buffy. Wings strongly reticulated as in the allied species.

Skull essentially as in *variegata*, though the brain-case is perhaps, a little longer and narrower and the muzzle shorter but the difference is very slight.

Incisors smaller and lighter than in *variegata*; i^1 slender, unicuspid in the type, but the presence or absence of a secondary cusp in this group appears to be variable. i^2 considerably smaller than in *variegata*, not crushed between and touching i^1 and the canine in front and behind, but clearly separated from each of them. Molars distinctly smaller than in *variegata* and *papilio*, the combined length on outer edge of m^1 and m^2 in eleven specimens of those forms, including types of both, being 2.7 - 2.8 mm., while in the type of *phalæna* it is only 2.4 mm.

Dimensions of the type (measured on the spirit-specimen): -

Forearm 44 mm.

Head and body 54; tail 44 ; ear 11.5; third finger, metacarpal 40, first phalanx 16; lower leg and hind foot (c. u.) 28.

Skull: greatest length 13.6; condylo-basal length 13.2; interorbital breadth 4.6; breadth of brain-case 8.1; palato-sinual length 5; front of canine to back of m^3 4.8.

Hab. White Nile, near Fashoda.

Type. Adult female. B.M. no. 15.3.6.67. Original number 71. Collected 27th January, 1914, by Willoughby P. Lowe.

Glauconycteris superba Hayman, 1939

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Glauconycteris superba, sp. n.

Diagnosis. - A *Glauconycteris* distinguished from all other members of the genus by its larger size, longer,

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flattened rostrum, and, above all, by its bold black and white colour-pattern, as figured. (N.B. - The figures are semi-diagrammatic representations of the pattern, and all membranes, including ear-tips, are left uncoloured.)

Type. - In Congo Museum, R.G. no. 14.765. Adult probably ? (the sexual organs were apparently cut away by the collector in opening the body), in alcohol. Collected at Pawa, about 30 miles E.N.E. of Medje, Ituri district, Belgian Congo, by Dr. Dubois of the Congo Red Cross.

Description. - General colour of head and body above and below, limbs, ears, and all membranes, black. The hairs of the body are dark brown basally, and black for the terminal half of their 5 - 6 mm. length. The body colour is relieved by several pure white patches and bands, the hairs forming which are the same length as the black hairs. These white markings are arranged as follows: - Above (fig. 1), a roughly circular patch, 4 - 5 mm. in diameter, on the top of the muzzle between the eyes; a patch on each side of the crown at the base of each ear, about 6 mm. long and a little less wide (these patches appear to encroach on the outer surface of the ear, but do not actually do so, the proximal half of the outer surface of each ear being clothed with short black hairs); two lanceolate stripes, one on each side of the median dorsal line on the fore part of the back, each about 18 mm. long, approximately 5 mm. wide anteriorly, and tapering rapidly to their termination about the middle of the back; two longer, narrower stripes, each about 29 mm. long and 2 mm. wide, one on either side of the body, each commencing a little in advance of, and exterior to, the ends of the scapular stripes last described, and each terminating just short of the root of the tail. On each shoulder, close to the junction of the antebrachial membrane with the body, another stripe begins, about 2 mm. wide, arising exterior to and on a level with the anterior end of the long flank stripe. This shoulder stripe runs forward over the shoulder, and below (fig. 2), joins a wider band, approximately 4 mm. wide, which then turns back and runs without interruption or narrowing along the side of the under-surface of the body, widening to about 6 mm. on the perineal region, where it joins the

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corresponding stripe of the opposite side of the undersurface. Most of the throat is occupied by a white band, 8 mm. wide at the centre, which narrows laterally to about 3 mm. as it passes on either side of the throat

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to the posterior outer surface of the lower part of the ear conch. On the side of the neck this band has a narrow connection joining it to the shoulder patch.

Other External Features. - In general form *G. superba* does not differ from other species of the genus. The ears are less rounded and more subquadrangular in outline than in other forms. *G. variegata papilio* being nearest to it in this respect.

Skull and Teeth. - The skull is proportionately longer and slightly less broad than in any other *Glauconycteris*. A most marked feature is the flattening of the rostrum, so that the profile shows an angle at the junction of the brain-case and the rostrum. The occiput is high, with a minute crest, and the total height of the skull from bulla to occiput is more than half the total length

instead of slightly less than half, as in all other species known. The teeth are characteristics of the genus: i^1 is not bicuspid, although there is a minute accessory cusplet on the internal angle near the cingulum. i^2 is closely crowded between i^1 and the canine. The lower incisors are closely crowded, and there is a greater proportionate difference in size between i_1 and i_3 than in other species.

Measurements. - It should be noted that, as a result of the unique type having been preserved originally apparently in formalin, the body has set in such a way that an accurate head and body measurement would probably be nearly 10 mm. More than the figure given here. Head and body 49, tail 30, hind foot 8.3, tibia 21.3, ear 13, tragus 7.5, forearm 47.5. Second digit 44.5; third digit metacarpal 45.5, first phalanx 20.7, second phalanx 24.3; fourth digit, metacarpal 44, first phalanx 13.3, second phalanx 10.8; fifth digit, metacarpal 40, first phalanx 8.2, second phalanx 8.2. Calcar approximately 11. Skull; greatest length 16.2, condylobasal length 15.5, palatal length 5.9 to posterior notch, 6.5 to tip of median process, height from bullae to occiput 8., interorbital width 5, zygomatic width 11, mastoid breadth 9.5, width of brain-case 8.5, width across canines 6, width across m^3 - m^3 7.9, upper toothrow (c- m^3) 6, lower toothrow 6.6, length of mandible 12.2.

Remarks. - Although in certain other species of *Glauconycteris*, namely *argentata* Dobson, *poensis* Thomas,

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alboguttatus J. A. Allen, and *humeralis* J. A. Allen, all of which are either fawn or brownish in colour, there is a more or less well-defined buff or whitish stripe on the lateral margin of the upper surface of the body, none shows a close approach either to the black and white colour or the pattern of spots and stripes found in *superba*, a pattern suggesting in a modified form that found in the spotted skunk, *Spilogale*. The nearest approach to this pattern is found in another Vespertilionid bat, *Scotomanes ornatus*, from S.E. Asia, a bat of a rich rufous general colour having a single median dorsal stripe and other white markings roughly corresponding in position to those of *Glauconycteris superba*, but much less well defined. The Emballonurine *Saccopteryx bilineata*, from S. America, has a pair of narrow white dorsal stripes on a black ground, but no other white markings, while the North American *Euderma maculata*, another Vespertilionid, is black with three well-defined white patches on the dorsal surface, one on each shoulder and one on the rump. That such a conspicuous new species should be found in a region which has received considerable attention from museum collectors of proved ability such as Lang and Chapin of the American Museum Congo Expedition of 1909 - 1915, Dr. Christy, and others, is somewhat surprising. It seems that much more collecting needs to be done before we can claim a complete knowledge of the mammalian fauna of tropical Africa.

Glauconycteris superba sheila Hayman, 1947

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Glauconycteris superba Sheila, subsp. nov.

Type. - B.M. no. 47.10. Adult ?, skin and skull (originally in alcohol), collected December 6th, 1946 at Oda, Oda Province, Gold Coast, by Mr. G. S. Cansdale. Collector's number 807.

Diagnosis. - Similar to *Glauconycteris superba* in size, external form and in the main features of the vivid pattern of white patches and lines on a black ground. It differs, however, in the addition of two white spots on each shoulder adjacent to the base of the humerus, in having conspicuous unpigmented areas on the upper surface of the elbow, knee and ankle joints, and in the paly grey colour of the under-surface of the interfemoral membrane, contrasting with the black of all the other membranous surfaces.

Description of the Type. - The general colour of head and body, above and below is black, the dorsal hairs being

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dark brownish at the base, as in *superba*. The white markings above, as shown in the diagrammatic figure (all membranes left uncoloured) consist of a median nose spot, a large patch on each side of the crown close to the base of each ear, two elongated patches lying one on each side of the median line on the upper shoulder, two long narrow lines, one along each flank, and on each lower shoulder, just above the root of the humerus, two spots of approximately equal size. These latter spots are the only feature of the body pattern distinguishing the new form from typical *superba*. Immediately in front of the two shoulder spots is the beginning of the white area which passes below to the broad U-shaped white band almost enclosing the black abdominal area. The broad white throat band terminates laterally immediately behind the base of each ear. The entire pattern of the underside of the head and body is thus exactly as in *superba*.

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Contrasting sharply with the completely black skin of the limbs and black upper surface of all membranes are the unpigmented round patches on upper surface of elbow, knee and ankle joints. These are described by Mr. Cansdale as flesh coloured in the fresh specimen, and are quite conspicuous in the dry skin.

The entire under-surface of the interfemoral membrane, in the alcoholic specimen as received, was pale grey, in sharp contrast with the black membranes elsewhere. In the dry skin this area has become brownish rather than grey, but still contrasts with the adjoining membranes, particularly since the numerous transverse veins in the under-surface form a prominent pale network in the dry state.

Skull and Teeth. - In size and proportions the skull agrees well with that of the type of *superba*. The flattening of the rostrum produces a marked angle at its junction with the brain case. In addition the length of the rostrum is markedly greater in proportion than in other species of the genus, and has brought about a considerable lengthening of the infraorbital foramen. In

other species the anterior opening of the foramen is situated over p^3 , in *sheila* it is over the canine root, and the foramen is thus much longer. The basisphenoid pits are somewhat deeper proportionately than in other species examined.

The teeth are normal, except that m^3 is considerably reduced, resembling that of *Scotophilus* in this respect. An examination of the same tooth in other species shows that it is best developed in the smallest forms and becomes proportionately weaker as the skull becomes larger.

Measurements. - All figures in parentheses are those for the type of *G. superba*. Head and body 63 (49); tail 49 (39); hind foot 10 (8.3); ear 13 (13); tragus 7. Forearm 47 (47.5). In comparing the two sets of figures it should be remembered that owing to the type of *superba* having been stiffened by preservation in formalin it was not possible to give fully accurate measurements, and it was suggested (1939) that it might be found later that the figures given then might not be fully reliable. The almost exact correlation in size between the skulls of the two forms suggests that this is the case.

Skull: greatest length 16.1 (16.2); condylobasal length 15.7 (15.5); palatal length to tip of median process

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7.7 (6.5); rostrum (incisive alveolus to front of orbit) 4.1; zygomatic breadth 11.9 (11); breadth of brain-case at posterior root of zygomata 9.1 (8.5); height of brain-case from auditory bulla 9.6 (8.8); interorbital breadth 4.8 (5); breadth across m^3 - m^3 8.1 (7.9); upper tooth row c - m^3 6.2 (6); length of mandible 12.9 (12.2); breadth across canines 6.2 (6).

Remarks. - This striking discovery, which I am pleased to name for Mrs. Cansdale, is obviously a distinct West African representative of *G. superba* of the eastern Belgian Congo. Its distinguishing features are very well marked; but in view of a tendency towards variability in the much simpler pattern in another species of the genus noted by the present author (1946) it is thought best to give the Gold Coast form subspecific status at present. Mr. Cansdale is to be congratulated on the addition of such a remarkable bat to the Gold Coast fauna. He informs me (in litt.) that the animal was found alive on the ground and brought in by natives, to none of whom was it previously known. In the absence of any field observations on the animal itself the significance of such a pattern remains to be explained.

Hipposiderus. *gigas gambiensis* K. Andersen, 1906

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Diagnosis. - Nose-leaves larger: width of horseshoe and posterior leaf about 13 mm.

In the specimen examined there are three supplementary leaflets only, no trace of a fourth.

Type. - F ad. (in alcohol). Gambia. Presented by the Earl of Derby. Brit. Mus. no. 42.9.27.36.

Hypsipsignathus *monstruosus* H. Allen, 1862

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Fur fine and short, very thin upon the face, interspersed with a few long stiff hairs. A well-marked white line extends from the facial protuberance to between the eyes. Top of head and nuclear region light ash brown, dark in the centre, lighter upon the sides, and narrowing to a whitish band which encompasses the inferior part of the anterior surface of the neck. The hair between this band and the chin is very thin and scattering, and of a pale brown. Dorsum delicate plumbeous tipped with grey, thicker above than below. The fur runs to a purer brown upon the posterior part of interfemoral membranes and thighs. The fore extremities to near the carpal joint are clothed with a thin coating of fur, while the interbrachial surface and that contained between the fourth finger and the sides of the body are studded in different directions with interrupted lines of minute hairs. The pectoral region and sides of belly of the prevailing hue - plumbeous grey - with a tendency of that in the centre of the abdomen to become whitish. Pubic region brownish. At the point of junction of the interbrachial membrane with the body a row of white hair is seen. The membranes beneath have upon them the same thin lightish hairs as above, but more extensive.

The dimensions of the skull are as follows:

Length from occiput to extremity of nose: 2.9 in.

" of cranium: 1.0

" " nose: 1.6

Height of nose: 0.9

Breadth of nasal bones: $0.3\frac{1}{2}$

" palate between canines: 0.6

" " molars: 0.9

Distance between zygomas: 1.6

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Length of lower jaw: 2.3 in.

Breadth of symphysis: 0.6

Distance between condyles: 1.3

Height of coronoids: 0.9

Distance between first premolar and canine: 0.2

Dimensions of the body:

From snout to interfemoral membrane: 12.0

" " ears: 3.4

" " eyes: 2.0

" anterior part of eyes to ears: 1.4

Length of mouth: 1.4

Breadth of snout: 1.4

Height of snout: 0.8

" face: 1.1

Breadth of chin: 0.9

Length " : 0.4¹/₂

Breadth between eyes: 1.3

" " ears: 1.6

Distance from snout to wing membrane: 5.7

Length of ulna: 4.6

" thumb: 2.0

" first joint: 0.9

" second joint: 1.3

" index finger: 3.9

" third " : 9.6

" fourth " : 7.1

" fifth " : 6.6

" inferior extremity: 3.6

" tibia: 2.1

" foot and claws: 1.3

Expanse of wing membranes: 27.0

Hab. Western Africa. Discovered by M. Duchailu

Hipposideros abæ J.A. Allen, 1917

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Type, No. 49123, M ad. (skin and skull), Aba, Uele district, Belgian Congo, Dec. 13, 1911; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 1715.

Upperparts (type, an average specimen of the dark phase) heavily washed with bistre from the shoulders posteriorly; lighter anteriorly through the shorter dark tips of the hairs, which nearly disappear on sides of head, neck and shoulders, which are thus much lighter than the back; underparts buffy gray, much lighter on throat; ears light brown, rather small, tip obtusely rounded; feet and ventral surface of limbs light brown; membranes dark brown.

Type, total length (collectors' measurements), 104 mm.; head and body, 66; tail, 38; foot, 12; ear (from outer base), 20. Forearm (from skin), 58.5; tarsus, 23.2; foot, 11; ear (from inner base), 16. Skull (type), total length, 23; zygomatic breadth, 13; width of braincase, 11; maxillar breadth (across m³), 8.8; breadth at base of canines, 6.1; upper tooththrow (c-m³), 8.2; length of mandible, 14; lower tooththrow, 9.4.

Posterior border of nose-loaf rounded; three secondary cutaneous leaflets, the outer one slightly developed; a frontal sac in the males, absent or much less devel-

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oped in females. P² uniformly present on both sides in each of the 35 skulls, inserted on the outer border of the tooththrow, closely crowded in between the canine and p³.

Represented by 35 specimens (16 males, 19 females, all adult, skins with perfect skulls), all taken at Aba, December 13, 1911.

The collectors' measurements from the fresh specimens (16 males, 19 females), are as follows:

Total length (MM; FF): 101.2 (97 - 107); 101.4 (95 - 105); Head and body: 65.5 (60 - 70); 65.4 (60 - 70); Tail: 34.2 (32 - 39); 36 (32 - 40); Foot: 12 (11 - 13); 12.2 (11 - 13); Ear: 21.4 (20 - 22); 21.5 (20 - 22)

The forearm and skull measurements (total length and zygomatic breadth) of the same specimens (MM; FF): Forearm: 58 (56 - 60); 57.8 (54 - 60.5); Skull: 23.1 (22.5 - 23.7) X 13.4 (13.1-13.9); 22.7 (21.9 - 23.3) X 13.2 (12.6 - 13.7)

This large series of specimens, all taken the same day at the same locality, and also all fully adult, is especially interesting from the wide range of color variation it presents, which, as shown by the skulls, is wholly independent of age, and probably of sex, although of the 16 specimens in the red phase 11 are females and 5 are males, and none of the males are nearly as red as are the majority of the red phase of the females. The dark phase, on the other hand, includes 11 males, while only 8 females can be assigned to it; and only two of the dark males are strictly referable to the norm of the dark series of females, the others, being distinctly intermediate between the dark and red phases.

The *dark phase* may be described as follows: Upperparts, in general effect, washed with drab-brown, lighter on the neck and shoulders, whitish prevailing on the sides of the shoulders; fur at extreme base dusky, with a broad intermediate zone of whitish, the tips of the hairs dark drab-brown, giving a dark brown superficial tone to the dorsal aspect. The extent of the dusky tipping varies greatly in different specimens, and maybe almost absent over the shoulders and especially on the sides of the shoulders, giving there a prevailing whitish tone. In general the white middle zone of the fur shows more or less at the surface. Underparts uniform dingy pale buff with a faint superficial wash of yellowish. Basal portion of the fur very pale brown, showing more or less through the pale buffy tips.

Red phase. Upperparts cinnamon-brown, often darkening on the middle of the back to nearly chestnut-brown. Fur at extreme base slightly darkened, the broad middle zone, pale cinnamon, the tips of the hairs dark ruddy brown; sides of shoulders light yellowish white. The underparts vary in different specimens from pale buffy to deep reddish ochraceous; in average female specimens, pale ochraceous. This is the 'red' phase as

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shown in females; the males in the red phase are much less rufous, both above and below, the upperparts being of a much darker and less reddish tone, and the ventral surface is seldom brighter (yellowish buff).

Many of the males and some of the females are so nearly intermediate between the two phases that it is difficult to assign them satisfactorily to either. None of the specimens is young enough to indicate the character of the immature or juvenile pelage.

Hipposideros abæ is allied to the *H. caffer* group, from which it differs in considerably larger size, and lighter and more buffy underparts. *H. caffer centralis* occurs with *H. abæ* at the type locality of the latter. Average length of forearm in *H. abæ* (35 specimens), 58 mm.; of *H. caffer centralis*, 50; total length, *H. abæ*, 101, *H. c. centralis*, 92. Skull, *H. abæ*, 23 X 13.3; of *H. c. centralis*, 18.5 X 10.

[*Hipposideros braima*](#) Monard, 1939

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Collection: N° 602 F ad., Bagingara.

Description. Très petite: espèce à longue queue du groupe de *caffer-beatus-nanus*.

Feuillet nasal (Fer-à-cheval) assez large (5 mm) muni sur les côtés de deux feuillets accessoires (largeur avec ces feuillets: 6 mm). Partie dressée aussi large que le fer-à-cheval avec ses feuillets latéraux, à marge supérieure en arc surbaissé, presque horizontale, sans trace de sillons et de crêtes verticaux. Immédiatement derrière le feuillet vertical, un petit sac frontal muni d'une touffe de poils noirs, à ouverture transversale; il est à présumer qu'il est mieux développé chez les mâles. Oreilles courtes mais larges, du type de *caffer* ou de *fuliginosus*. Marge interne d'abord

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très convexe, s'approchant beaucoup du feuillet nasal: puis à peu près droite et horizontale (l'animal en position verticale). Pointe de l'oreille en angle droit. Marge externe formant dès l'origine un large lobe convexe (antitragus) à peine séparé de la conque par une légère sinuosité concave: puis convexe, enfin concave vers la pointe de l'oreille. Celle-ci dirigée en arrière.

Membranes normales naissant de l'extrémité du tibia. Fémur et tibia proportionnellement allongés; pied étroit, plus long que le pouce. Queue allongée, la dernière vertèbre libre. Membrane interfémorale en forme d'arc.

Pelage complètement noirâtre.

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On voit par ces dimensions que les caractères principaux de notre nouvelle espèce sont l'allongement de la queue et des membres postérieurs:

nanus ALLEN, *beatus* AND., *braima* n. sp.

Longueur de la queue mm.: 22, 20.5 - 22, 26

" du tibia: 16.5, 15.2 - 16, 18

" du pied: 7.9, 7.8 - 8.5, 9

En outre l'avant-bras est proportionnellement plus court, les doigts ayant des dimensions comparables.

Le crâne n'a pas été tiré de notre unique exemplaire.

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Mœurs. L'espèce vivait en compagnie de nombreux *Nycteris* de diverses espèces dans le comble de la maison que le Régulo de Sama met à la disposition de ses hôtes, à Bagingara. Mais elle y était beaucoup moins nombreuse que les *Nycteris* et deux individus seulement furent capturés, le second ayant réussi à s'enfuir.

Notre exemplaire femelle porte, immédiatement en avant de l'éminence sexuelle, deux petits appendices, de signification douteuse, qu'on signale parfois dans les femelles de ce genre

Hipposideros caffer niapu J.A. Allen, 1917

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Type, No. 49414 (alcoholic), M ad., Niapu, Belgian Congo, Jan. 27, 1914; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 2322.

Similar in general coloration to *H. caffer centralis*, but much larger; it has a dark phase and a rufous phase, like the other forms of the *caffer* group.

Type (in rufous phase), upperparts uniform chestnut-brown; underparts cinnamon-brown; ears and membranes dark brown.

In the dark phase the upperparts are hair-brown, below lighter, with the tips of the hairs lighter than the basal fur. Most of the specimens are in the dark phase, which is similar to the corresponding phase of *H. caffer centralis* and *H. abæ*, but none of the specimens in the rufous phase approach nearly to the intensity of coloration seen in many examples of either of these forms.

Measurements: Type, total length, 88; head and body, 58; tail, 30; tibia, 21.8; foot (c.u.), 10.8; ear, 11.2 X 14.6. Forearm, 54.5. In the ten topotypes (including; type) the forearm averages 54.7 (53-56).

Skull (type), total length, 20.3; condylobasal length, 17.8; zygomatic breadth, 10.6; mastoid breadth, 10.2; interorbital breadth, 2.9; maxillary breadth, 6.8; breadth at base of canines, 4.8; maxillary tooththrow (c-m³), 6.6; length of mandible,

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12.4; mandibular tooththrow, 7.2. Four male topotype skulls (including type), total length, 20.1 (19.8-20.3); zygomatic breadth, 10.5 (10.3-10.0); interorbital breadth, 2.9 (2.8-3); length of mandible 12.4 (12.2-12.5).

H. caffer niapu seems clearly to represent a large race of the *caffer* group, larger than any hitherto described. In Andersen's table of measurements of the forms of the *caffer* group, based on 98 specimens (including 75 skulls), the maximum length of the forearm is given as 53.8 for 24 specimens of *centralis*, as against 54.7 in 10 specimens of *niapu*, while the maximum in 40 specimens of typical *caffer* is 51.8. He gives (l. c., p. 270) the average forearm length for 40 specimens of *H. caffer caffer* as 48.6, and for 50 specimens of *centralis-guineensis* (which do not differ in size) as 50.6, as against 54.7 in 10 specimens of *niapu*, the latter exceeding the *centralis-guineensis* series by 4 mm. The skulls bear out the evidence of larger size for the *niapu* series, in which the average total length of the skull is 1 mm. greater than in the *centralis-guineensis* series, which is equal to the average difference between the latter and typical *caffer*.

H. c. niapu is represented by 10 adult specimens (6 males, 4 females), all from the type locality, and all collected the same day. It is clearly specifically different from the much larger *H. abæ* described below from a series of 35 skins and skulls, all from Aba, and all collected also during one day.

Hipposideros camerunensis Eisentraut, 1956

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Material: 4 Exemplare

Buea, 29. April 1954, 2 MM (505, 510), 2 FF (506, 511).

Die hier neu beschriebene Art, die in 4 in den wesentlichen Merkmalen übereinstimmenden Exemplaren vorliegt, ist am nächsten verwandt mit *H. cyclops* und gleicht ihr in den äußeren Merkmalen, zeichnet sich aber durch ihre bedeutende Größe aus, die durch keinen gleitenden Übergang mit den für *cyclops* vorliegenden Maßen verbunden ist. Man könnte vielleicht die Ansicht vertreten, daß die neue Form nur eine Unterart von *cyclops* darstellt, und zwar in diesem Fall eine Gebirgsform, doch gibt als Fundort für *H. cyclops* DOBSON "Cameroon-Mountains" und MATSCHIE (1891) ausdrücklich Buea an, von wo auch die vorliegenden Stücke stammen, so daß eine artliche Trennung als gegeben erscheint. Es liegt hier ein ähnlicher Fall von Größendifferenz zweier nächstverwandter Formen vor, wie wir ihn z. B. von *Nyctalus noctula* und *Nyctalus lasiopterus* (*maximus*) kennen. Wie weit die neue Form in Kamerun und darüber hinaus verbreitet ist, wissen wir nicht. Die 4 vorliegenden Exemplare wurden in einer Höhle oberhalb des Eingeborenen-town Buea gefangen.

Hipposideros camerunensis besitzt wie *H. cyclops* ein außerordentlich dichtes und wolliges Fell. Oberseits ist der Grundton schwärzlich, doch gewinnt das Fell durch die grauweißlichen Haarspitzen einen graumelierten Ton. Am Unterarm, der oberseits bis zur Hälfte dicht behaart ist, nehmen diese Haare eine bräunliche Färbung an; auch am Vorderkopf ist das Fell etwas mehr graubräunlich getönt. Die Färbung der Unterseite ist eine Nuance heller als die der Oberseite und geht in den vorderen Partien ein wenig in Grau, an den hinteren in Braun über. Auch hier sind die Haare am Ende heller als am Grunde. Der Unterarm ist auf der Ventralseite in seinem proximalen Drittel schwach behaart. Flughäute und Ohren sind bräunlich-schwarz. Das Plagiopthagium schließt den Metatarsus der 1. Zehe völlig

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mit ein und inseriert ein Stück oberhalb des proximalen Endes des 1. Zehengliedes. Der Schwanz ist relativ kurz. Der letzte Wirbel steht frei aus der Flughaut.

Der Schädel von *H. camerunensis* ist nicht nur absolut, sondern auch relativ länger als der von *cyclops*. Die Gegenüberstellung der Körper- und Schädelmaße läßt den Größenunterschied zwischen beiden Arten deutlich erkennen (vgl. auch Abb. 6). Der Hinterfuß ist gegenüber dem von *cyclops* relativ klein.

Der Nasenaufsatz zeigt im wesentlichen die gleiche Form wie bei *cyclops* und ist durch den keulenförmigen Fortsatz in der Mitte des oberen Nasenblattes ausgezeichnet (Abb. 7). Die Breite des unteren Nasenblattes (Alkoholstück 511 F) - ohne die sekundären Seitenlappen - beträgt 14.8 mm, die des oberen Nasenblattes 10.1 mm gegenüber 12.9 und 9.4 mm bei *cyclops*. Der sich hinter dem Nasenblatt longitudinal öffnende Drüsenschlitz läuft am unteren Ende in eine mäßig lange Zunge aus. Die MM besitzen einen mit langen borstenartigen rotbraunen Haaren besetzten Analdrüsensack, der sich unmittelbar über dem After nach außen öffnet und mit der Afteröffnung zusammen in einen kloakenartigen, von der Hautfalte überdachten Vorhof mündet. Außerdem öffnet sich ein kleiner Drüsengang separat im Zentrum der Hautfalte nach außen. Wie bei *cyclops* kann der Drüsensack ausgestülpt werden. Die FF besitzen Afterzitzen, die bei dem einen Tier nur klein, bei dem anderen dagegen von

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[table removed - eds.]

ansehnlicher Länge sind. Die Vaginalgegend ist vorn und seitlich mit starren borstenartigen Haaren besetzt.

Die Zahnformel (vgl. auch Abb. 8) lautet: $\frac{1 \cdot 1 \cdot 2 \cdot 3}{2 \cdot 1 \cdot 2 \cdot 3} = 30$. Der 1. obere Prämolare ist sehr klein und völlig nach außen gerückt, so daß sich Caninus und 2. Prämolare fast oder ganz berühren. Der 2. Prämolare ist sehr kräftig entwickelt und überragt die Spitzen der Molaren. Die unteren Incisivi sind dreigeklappt. Der 1. Prämolare ist nur etwa $\frac{1}{3}$ so hoch wie der 2. (bei *H. cyclops* erscheint er etwas höher).

Die kurze Zeit in Gefangenschaft gehaltenen Tiere waren außerordentlich bissig.

Der Typus (M, Tagebuch-Nr. 505, Katalog-Nr. s 194) und die 3 Paratypen befinden sich im Staatlichen Museum für Naturkunde in Stuttgart

Hipposideros Commersoni mostellum Thomas, 1904

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Size small, as in *Commersoni*. Supplementary leaflets four, with rudiment of a fifth. General colour whitish, the brown Y-shaped marking of the back well defined; under surface creamy whitish, a brown line across each shoulder separating off a white patch at the insertion of the antebrachial membranes.

Skull and teeth as in true *Commersoni*, the cheek-teeth conspicuously smaller than in *gigas*.

Dimensions of the type (measured in skin): -

Forearm 92 mm.

Skull: length from cingulum of canine to back of occipital crest 32; basal length to cingulum of canine 26.5; zygomatic breadth 18; mastoid breadth 15; upper cheek-teeth, front of p^4 to back of m^3 8.4; front of lower canine to back of m_3 13.

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Hab. (of type). Tana R., British East Africa. Other specimens from Zanzibar.

Type. Male. B.M. no. 89.3.8.3. Presented by H. C. V. Hunter, Esq.

Hipposideros curtus G.M. Allen, 1921

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Hipposideros curtus sp. nov.

Type. - A female in alcohol, 19305, Museum of Comparative Zoölogy, from Sakbayeme, Cameroons. Rev. George Schwab, collector, 1920.

Description. - A small species, about the size of *H. beatus*; female without the frontal glandular sac; tail very short, barely exceeding the extended femur; wings from the distal end of tibia; calcar short, about one half the length of tibia; thumb markedly small and weak as contrasted with *H. beatus* and *H. caffer guineensis*; forearm about as in the latter but wings shorter, the combined length of metacarpal III and its proximal phalanx exceeding the forearm measurement by only 1 mm. as against 6 mm. in the two other species; metacarpals III and V subequal, IV a trifle longer, whereas in the two contrasted

species metacarpals III and IV are about equal, the fifth shorter. Ears noticeably larger than in either *H. beatus* or *H. caffer guineensis*; tibia longer than in the former but shorter than in the latter.

The noseleaf is essentially similar in *H. beatus* and *H. caffer* but in the new species is of a very different appearance. The horseshoe is noticeably broader and thicker, with a slight median emargination anteriorly, the edges of which are a trifle upturned; the small cushion-like expansion of the nasal septum, nearly obsolete in the two other species, is here considerably developed, and, with the small projections at the outer border of the nasal openings, forms a partial roof over the nares.

The erect "sella" is slightly thickened, densely covered with short stiff hairs, more as in *H. beatus* rather than nearly naked as in *H. caffer*. The posterior crescent is simple and evenly convex in outline, and is divided by three vertical ridges on its anterior face. There are two lateral accessory leaflets at each side of the horseshoe, but these are very small and poorly developed.

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the outer one a mere wart and so concealed by the hair of the lip that is not easily made out; the larger supplementary leaf is only about half the lateral extent of the horseshoe, instead of nearly as long, as in *H. caffer* and *H. beatus*.

The color is not definable from the alcoholic specimen, but seems to be a uniform dark smoky brown.

Skull shows no striking peculiarities. Contrasted with that of the other two species mentioned, it more resembles that of *H. caffer guineensis* in that the width across m^3 about equals rather than exceeds the length of the maxillary tooth row; it differs from both in having more terete upper incisors, their outer sloping border without indication of the notch obviously present in the two other species; p^3 is in contact with canine and p^4 , very small but barely external to the median axis of the tooth row; upper canine with a minute posterior basal cusp; lower canine very slender, nearly equalled in height by p_4 ; sagittal crest weakly developed, extending to the back of the parietals; nasal inflation prominent.

Measurements. - Length (circa) 67 mm., tail from anus 17, tibia 20, with the foot 27, ear from meatus 17.5, width of ear 15, calcar 10.5, forearm 47, third metacarpal 32.5, with proximal phalanx 48, fourth metacarpal 35, with proximal phalanx 45.5, fifth metacarpal 33, with proximal phalanx 45, greatest width of horseshoe 6.5, of posterior crescent 7.5.

Skull: total length 17.5 mm., zygomatic breadth 9, width outside m^3 , 5.8, upper maxillary teeth 5.7, lower tooth row i_1 to m_3 , 6.8.

Remarks. - Externally this small species is at once distinguishable from *H. beatus* and *A. caffer guineensis*, of similar size and inhabiting the same region, by its very short tail which barely exceeds the extended femur, by its much larger ears, by the short calcar combined with the long slender leg, and by the shorter fingers in comparison with the forearm. The noseleaf is very different, with its broad horseshoe, emarginate anteriorly, reduced

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supplementary lappets, expanded internarial septum, and hairy sella. These characters will also serve to distinguish it from *H. nanus*, of the Belgian Congo, apparently a close ally of *H. beatus*. For additional notes on the type of *H. nanus*, as well as for carefully executed drawings of its noseleaf, thanks are due M^r Herbert Lang of the American Museum of Natural History.

Hipposideros gigas niangaræ J.A. Allen, 1917

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Plate LI, Fig. 1.

Type (and only specimen), No. 49103, F ad., skin and skull, Niangara, Uele district, Belgian Congo, June 2, 1913; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 2069.

A small member of the *H. gigas* group. Dorsal hair-brown Y-patch of short

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dense fur well defined; upperparts anterior to the Y-patch with the fur much longer and looser and of a pale brownish gray, the extreme tips of the hairs, particularly on the posterior third of this area, whitish; still paler on the top and front of the head; sides of the head lighter than the top, and joined by a narrow still lighter band enclosing the nose-leaf area; sides of back grayish posteriorly, like the anterior back, separated from the Y-patch by a whitish transverse band about 13 by 6 mm. in extent; a patch of clear white on the sides beneath the junction of the propatagium with the body and extending slightly in front of it, bounded ventrally by a band of dark hair-brown fur, a lateral extension of the dorsal Y-patch; general color of underparts yellowish gray, with a broad median band of dusky gray, the fur darker basally; ears, feet and membranes blackish. Ears long and tapering, the extreme tip slightly rounded. Membranes attached about 5 mm. above tarsal joint. Supplementary leaflets three, well-defined.

Collectors' measurements: Total length, 137 mm.; head and body, 107; tail, 30; foot, 25; ear (from outer base), 35.

Forearm (in skin), 104; third metacarpal, 72; tail, 30; lower leg, 40.5; breadth of nose-leaf, 11.7.

Skull, total length, 35.3 (39.8); zygomatic breadth, 20 (22.2); mastoid breadth, 15.8 (18.2); maxillary breadth, 14 (14.2); antorbital breadth, 11 (11); across cingula of canines, 10.5 (11.6); mandible, 24.4 (27.8); upper teeth, 13.5 (13.8-14.5); lower teeth, 15.5 (16-16.3).

The single specimen, of *H. niangaræ* is a fully adult female, with a strongly developed sagittal crest, but the teeth are not appreciably worn. It has the coloration and general characters of *H. gigas*, from which it differs in much smaller size, the forearm measuring only 104 mm. as against 108-116 mm. given by Andersen (*l. c.*, p. 48) for 6 specimens of *H. gigas*; while the total length of the skull is 35.3 as against 39.8 given by Andersen for *gigas*, with other skull measurements proportionally less.

The type locality of *H. gigas* is the Benito River in Angola, and the 8 specimens cited by Andersen are likewise all from Angola. The type, and apparently the only recorded specimen, of *H. gigas gambiensis* is from Gambia, in the same general region. On the other hand the type of *niangaræ* is from the Uele district of the Belgian Congo, a region of quite different physical conditions and more than a thousand miles westward.

Hipposideros gigas viegasii Monard, 1939

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KNUD ANDERSEN: Ann. Mag. Nat. Hist, 7. 17. 1906. p. 42 (sur *H. gigas*)

Collection: N^{os} 908 à 926, 19 MM et FF; en plus 14 crânes de MM et FF; Madina Boé.

Les *Hipposideros* du groupe *commersoni* ont été revues par KNUD ANDERSEN dans le travail cité plus haut; il en distingue trois espèces et deux sous-espèces: *commersoni typicus* GEOFFROY, *commersoni marunqensis* NOACK, *thomensis* BOCAGE, *gigas typicus* WAGNER, *gigas gambiensis* KNUD ANDERSEN.

L'examen de notre abondant matériel, auquel il faudrait joindre une cinquantaine d'exemplaires que nous avons refusés ou qui se sont montrés trop détériorés pour la préparation, nous a démontré qu'il s'agissait d'une nouvelle sous-espèce de *gigas*, très nettement caractérisée par sa coloration, son dimorphisme sexuel, la brièveté de la queue et le faible développement de l'uropatagium. Tous nos exemplaires, sans aucune exception et sans atténuation montrent ces caractères distinctifs avec une grande netteté.

Description. Couleur: les mâles, plus grands que les femelles, plus fortement armés et plus agressifs, ont tous une couleur d'un beau jaune, parfois plus pâle, parfois plus vif; les oreilles, les appendices nasaux, les membranes sont brunes. Il n'y a pas trace des détails de coloration que donne KNUD ANDERSEN comme caractéristiques de *gigas gambiensis*: le dessin dorsal foncé en Y

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n'existe pas et ne laisse pas deviner sa présence; la tête a la même coloration que le dos, un peu plus claire seulement; pas trace non plus de poils bicolores: ils sont jaunes de la base à la pointe; le dessous est de même couleur que le dessus; pas trace encore d'une ligne claire longitudinale de chaque côté du dos, ni de tache blanche à l'insertion du propatagium (toutefois, à cette place, le jaune est plus vif). Il ne s'agit pas non plus d'une "russet phase" dont le style est le même que dans *gigas*, mais où le poil est fortement lavé, de roux.

Les femelles, plus petites, ont une coloration si différente qu'on pourrait les prendre au premier abord pour une autre espèce. La teinte jaune des MM est remplacée par un orangé extrêmement vif, même ardent. Aucune trace de tache en Y, de raie claire, de poils bicolores, de touffe blanche aux épaules, de différence entre le dos et la face ventrale, entre la tête et le tronc.

Le second caractère de notre sous-espèce est la réduction de la queue et de l'uropatagium qui en est la conséquence. Les individus de *commersoni* et *gigas* que nous avons examinés et qui proviennent des Musées de Bâle et Genève ont une queue longue de 30 mm et plus; ANDERSEN donne 34.7 à 35.5 (*commersoni*) et 31 à 39 (*gigas*). Nos exemplaires ont tous la queue du tiers plus courte, mesurant environ 20 mm seulement rarement plus; le nombre des vertèbres n'a pu être comptés, car nous n'avons pas conservé les corps, mais il nous a paru réduit à 4 ou 5? La queue est plus courte que la moitié du fémur et dépasse un peu l'uropatagium.

Le fer-à-cheval nasal est bien conformé; il est large de 11 à 12 mm, donc plus petit que dans *gigas gambiensis*; il y a trois folioles supplémentaires bien conformées et une quatrième rudimentaire.

L'oreille est du type de *gigas*, relativement petite et pointue.

Le crâne, qui chez les vieux mâles, est plus grand que dans *gigas gambiensis*, lui ressemble parfaitement. Les canines sont très longues, de section triangulaire, chaque face sillonnée, avec un faible tubercule accessoire postérieur. La première prémolaire est petite, arrondie, rejetée en dehors de la ligne dentaire, coincée dans l'angle formé par la canine et la seconde prémolaire. Celle-ci tranchante, plus élevée que la première molaire. Les deux premières molaires sont normales, la dernière réduite. Les crêtes

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sagittales et lambdoïdes sont extrêmement saillantes, la première presque aussi haute que la boîte crânienne dans certains exemplaires. Incisives inférieures trifides, canines triangulaires, les prémolaires et molaires comme dans *gigas*.

Cette nouvelle et remarquable forme de Chiroptères est dédiée à M. CARVALHO VIEGAS, gouverneur de Guinée.

[table removed - eds.]

Distribution géographique. Les espèces et sous-espèces du groupe de *commersoni* se distribuent selon le schéma suivant:

Gambie: *gigas gambiensis*
 Guinée portugaise: *gigas viegasii*
 Angola, fleuve Muni, Benguela: *gigas typicus*
 S. Tomé: *thomensis*
 Tanganyika, Moçambique, Brit. East Afr., Zanzibar: *commersoni marungensis*
 Madagascar: *commersoni typicus*

Mœurs. Ces chauves-souris nous furent apportées un soir à notre campement du Boé par un indigène qui en avait rempli trois

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sacs. Nous n'avons pas compté le nombre des individus, mais il pouvait y en avoir en tout une centaine. Féroces et agressives, leur maniement n'était pas sans danger. L'heure étant très tardive et mes boys absents, je me suis fait aider par l'indigène. Nous en attachâmes une partie aux branches avoisinantes, en laissâmes quelques-unes dans leurs sacs et en libérâmes une moitié environ. Celles qui étaient renfermées dans les sacs se battirent toute la nuit en poussant des sifflements aigus; elles finirent par se tuer mutuellement et furent pour la plupart inutilisables à cause des blessures qu'elles s'étaient faites. Il n'était pas question de les conserver en alcool; l'épaisse couche de graisse dont elles étaient toutes munies, eût rendu impossible la pénétration du liquide et la bonne conservation des sujets; la préparation des peaux fut rendue répugnante et difficile par la même raison.

Ces Chiroptères vivaient en grande société dans les trous des arbres de la forêt du Boé, toutes au même endroit. L'indigène qui nous les a amenées a dû épuiser la colonie, dans l'espoir d'une plus forte récompense. Les individus libérés s'enfuirent immédiatement et s'allèrent loger dans l'épais bosquet où j'avais établi mon campement. Toutefois, je n'en vis plus trace les jours suivants, et il est possible qu'ils aient, à la manière des pigeons voyageurs, rejoint leur station d'origine

Hipposideros jonesi Hayman, 1947

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Type. - B.M. No. 47.629, collector's number 42, F in alcohol, collected with two other FF, B.M. Nos. 47.627, 47.628, at Makeni, Sierra Leone (in cave), 12 December, 1946.

Diagnosis. - A medium-sized *Hipposideros* (forearm 48 mm.), distinguished by the combination of very large ears with a very distinct form of nose-leaf, in which the internarial septum is greatly expanded and the posterior transverse leaflet is greatly lengthened (see figure). Skull of the *H. bicolor* type, with mastoid-breadth-considerably exceeding zygomatic breadth.

Description. - Ears very large, length from meatus 23 - 23.5 mm., greatest breadth 20 - 21 mm., rounded, with a small fold on the lower part of outer margin. Nose-leaf quite distinct in form, characterised by (a) the wide main horseshoe with broad free margin, covering the whole of the muzzle, and having no median emargination, (b) the single wide lateral leaflet, (c) the greatly expanded internarial septum, which is developed into a broad fleshy oval pad, with slight median longitudinal groove, concealing the nares from view, (d) the wide sella with slightly undulating upper margin and with four small warts on the upper part of the anterior surface, and (e) the greatly lengthened posterior leaflet with strongly curved upper margin and having on its basal portion

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three weak vertical ridges dividing the basal portion into four shallow cells. The visible portion of this posterior leaflet occupies one-third of the total length of the whole nose-leaf. All surfaces of the nose-leaf are well haired. The measurements of the nose-leaf of the type are: greatest length from anterior margin of horseshoe to upper margin of posterior leaflet 15; breadth of horseshoe and of posterior leaflet 8; breadth of internarial septum 4.7, length 3. (All measurements in millimetres.)

The thumb is weak, metacarpals III. and V. subequal, IV. slightly longer. Legs are long, with wing attached at end of tibia. Terminal tail vertebra free of membrane.

There is no frontal sac in the three females examined.

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The colour above is a uniform smoky brownish grey, the hairs being dark basally, with pale grey median zone, and terminally as described. Below the general colour is paler, the hairs being without the terminal smoky zone.

Skull and teeth: the skull has the general proportions of a typical member of the *H. bicolor* group, particularly in the wide expansion of the brain-case and narrow zygomatic breadth. The upper incisors are slightly bilobed, upper canines with weak basal cusp, the small p_2 separating and in contact with canine and p_4 . Lower incisors are trilobed, canines slender, p_2 well developed and about half the height of p_4 .

Measurements of type and two paratypes (latter in parentheses). - Head and body 52 (52, 53), tail 25 (22, 27), hind foot 7.5 (8, 7), ear-length from meatus 23 (23, 23.5), ear, greatest breadth 21 (20, 20), forearm 48 (49, 48), tibia 22 (22, 22), metacarpal III. 35 (35, 36), metacarpal IV. 36 (36, 38), metacarpal V. 34 (33, 35); skull of type and one paratype, greatest length 18.2

(19.1), condylobasal length 17.2 (17.4), palatal length 7 (6.9), mastoid breadth 10 (10), zygomatic breadth 8.5 (8.3), interorbital breadth 2.8 (2.9), breadth across canines 3.6 (3.8), breadth across m^3 - m^3 6 (5.9), upper tooth-row c- m^3 6 (6.2), height of rostrum 5 (5.2), height of cranium from bulla 7.7 (7.5), mandibular length 10.9 (11.7).

Remarks. - *Hipposideros jonesi* stands apart from any of the groups defined and keyed by Tate (1941). Although in size and form of ears and in mastoid breadth exceeding zygomatic breadth it approaches the *bicolor* group, in other respects, such as the well-developed single lateral leaflet and absence of frontal sac it differs widely from that and the other groups. The expanded internarial septum is far better developed than in *H. curtus* from the Cameroons (breadth 4.7 against 1.7), and the greatly developed posterior leaflet removes it from other groups.

Hipposideros lamottei Brosset, 1985

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Parmi les Chiroptères collectés au Nimba figurent six spécimens se rapportant à une forme d'*Hipposideros* apparemment inédite que nous décrivons ci-après comme *Hipposideros lamottei*, en hommage à M. Lamotte qui organisa la mission au Nimba.

Type M ad. - collect.: Brosset, Pierre Richaud, déc. 1983, Muséum de Paris, CG 1984-487.

Cotypes 2 M, mêmes date et provenance, Muséum de Paris, CG 1984-488 et 489; 1 F et 1 M, mêmes date et provenance, Mus. de Genève; 1 M ad., déc. 1983, grotte de Blandé, Mus. de Genève.

Matériel de comparaison: 5 *Hipposideros ruber*: 1 M et 2 F ad., Mus. de Genève; 1 M CG 1984-490 et 1 F CG 1984-491, du Mus. de Paris. Tous provenant de la grotte de Blandé (collect. Lamotte-Aellen 1963 et Brosset 1983).

Diagnose.

Hipposideros de taille moyenne, du groupe *caffer-ruber*, plus grand que *caffer* et différent de la sous-espèce sympatrique de *ruber*, *H. ruber guineensis*, par des dimensions corporelles plus grandes et un crâne plus petit; autres différences concernant les oreilles, la feuille nasale, la forme du crâne et les dimensions des dents.

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Morphologie externe de *H. lamottei*.

Ressemble à celle des autres *Hipposideros* du groupe *caffer-ruber*. Diffère de *ruber* par des oreilles plus courtes de 2 mm; le processus postérieur de la feuille nasale forme un bourrelet rectiligne se terminant par un angle presque droit aux extrémités (Fig. 1), sa longueur étant égale ou supérieure à la largeur de la feuille nasale (inférieure chez *ruber*). Toutes les dimensions corporelles

[Table removed - eds.]

dépassent celles des formes décrites du groupe *caffer-ruber* (Tabl. 1). Présence d'un sac frontal chez les deux sexes. Pelage brun (rougeâtre chez les adultes de *ruber guineensis*).

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... corporelles, toutes les dimensions du crâne sont inférieures chez *lamottei* à celles relevées chez la forme sympatrique de *ruber* (Tabl. 2). Le crâne de *lamottei* est plus étroit et d'apparence

[Table removed - eds.]

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générale moins robuste. La distance entre C^1 - C^1 et M^3 - M^3 est plus faible, d'où un palais plus étroit chez *lamottei*. Le profil de la capsule crânienne présente une dépression caractéristique en son milieu, alors que celle de *ruber* réalise une courbe assez régulière (Fig. 2). La denture est considérablement plus faible chez *lamottei*.

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Cette forme a peut-être été déjà collectée au Nimba, où d'importantes séries d'*Hipposideros* ont été constituées (Verschuren, 1976; Wolton et al, 1982), mais elle n'aurait pas été distinguée de *ruber guineensis*. Peut-être s'agit-il d'une espèce montagnarde, encore qu'elle soit présente à Blandé, à 500 m d'altitude.

Répandus dans l'ouest du Maroc et toute l'Afrique tropicale, les *Hipposideros* du complexe *caffer-ruber* sont probablement les mammifères les plus nombreux du continent africain, et aussi les mieux représentés dans les collections scientifiques. Cependant, l'individualisation des espèces de ce groupe, composé de populations géographiques variables, est très compliquée, et la systématique en serait à reprendre à l'aide de critères de distinction non exclusivement morphologiques: étude des formes sympatriques et allopatriques, des caractères chromosomiques, de l'écologie et du comportement.

La validité d'une distinction spécifique entre *caffer* et *ruber*, fondée sur des caractères ténus et géographiquement variables, avait été mise en doute (Aellen et Brosset, 1968) jusqu'à ce que Adam et Hubert (1972) aient montré qu'au Sénégal les deux

formes vivent en association dans de mêmes gîtes, ce qui prouve leur isolement génétique. L'extension à d'autres régions de l'étude des formes sympatriques du complexe *ruber-caffer* permettrait probablement d'isoler d'autres espèces, non décrites ou considérées comme des sous-espèces de *ruber* ou de *caffer*.

Observations "in natura".

Cinq spécimens d'*H. lamottei* furent capturés près du Pierre Richaud (1 450 m), un autre à la grotte de Blandé (500 m). Ces deux cavités sont distantes de 6 km à vol d'oiseau.

La grotte de Blandé est située au pied du Mont Nimba, près de Ziéla, dans une vallée encaissée, en forêt dégradée. Elle se présente comme un abri sous roche assez vaste, se prolongeant au fond par un réseau de galeries et sous-bassements exigus. Ces galeries étaient connues pour abriter une colonie d'*Hipposideros ruber* (Aellen, 1963). Nous y avons capturé 3 représentants de cette dernière espèce et 1 *H. lamottei*. Une colonie de *Rousettus aegyptiacus* qui habitait le porche de Blandé (M. Lamotte, *viva voce*), n'a pas été revue en décembre 1963, bien que de nombreuses traces témoignent de sa présence récente.

La galerie de mine de la face N.E. du Pierre Richaud, où furent collectés 5 spécimens, dont le type de *H. lamottei*, s'ouvre sur une pente abrupte, dépourvue de végétation arbustive. La galerie, creusée dans le minerai de fer, est rectiligne, haute et large d'environ 2 m, profonde d'environ 100 m. La colonie d'*Hipposideros* comptait 130 individus, en majorité du type brun (non rouge, comme *ruber*). Dix individus, examinés à l'extérieur, répondaient au type *lamottei*. Les deux sexes étaient représentés (5 M - 5 F) mais il n'y avait aucun jeune ni aucune femelle visiblement gravide. Le cycle de reproduction est probablement boréal. 4 *Lissonycteris* cohabitaient avec ces *Hipposideros*.

Hipposideros langi J.A. Allen, 1917

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Text Figs. 4-6.

Type, No. 49098 (skin and skull), M ad., Avakubi, Jan. 24, 1914; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 2481.

A large species, with long, soft, woolly pelage, allied in general features to *Phyllorhina cyclops* Temminck.

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Upperparts, in general effect, blackish grizzled with white, the pelage being brownish black at base and apically with a narrow subapical zone of white; head grayish brown, much lighter than the back; with dusky eyerings; underparts uniform dusky brown, the tips of the hairs lighter, giving a decidedly grayish general effect, considerably lighter than the back. Proximal half of forearm heavily furred. Ears and membranes naked, blackish brown, the wing and leg bones and the feet much lighter than the membranes, especially on the ventral aspect.

Ears narrow and attenuate, tapering apically to a sharp point. Interfemoral membrane deeply hollowed; tail very short, about one-third of the length of head and body, the terminal vertebra free. Thumb short. A large frontal sac in both sexes, lined with silky straight hairs, wholly white, or white at base with light brownish tips. When the sac is everted these white hairs form a conspicuous white tuft. A glandular pouch is present in males just in front of the anus, lined with long, rusty brown bristly hairs, which when the pouch is everted form a conspicuous tuft, the hairs being more or less obvious even when the pouch is not everted. The females appear to lack the anal pouch, but have in its place a pair of elongated nipple-like papillæ, usually clubshaped or thickened apically. Those are conspicuous in alcoholic specimens and usually readily observed in dry skins. They are situated about 5 mm. apart and have a length of about 8 mm.

Nose-leaf very broad (about 20 mm. wide); posterior leaflet narrow (about 10 mm. wide), with a narrow central point 2 mm. high and 0.5 mm. wide. The nostrils are enclosed posteriorly and laterally by a fleshy conch-shaped border. There is a prominent fleshy knob at the posterior border of the nose-leaf, and two supplemental leaflets, the inner strongly and the outer weakly developed.

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Type, total length (collectors' measurements), 114 mm.; head and body, 87; tail, 27; foot, 20; ear, 33. Collectors' measurements of 4 adults (3 females and the male type), total length, 113 (110-116); head and body, 85.5 (83-87); tail, 28 (27-30); foot, 19.5 (19-20); ear, 33.8 (33-35).

Forearm in 10 specimens (6 skins and 4 alcoholics), 68 (66.2-68.8).

Type skull, total length, 28.1; condylobasal length, 25.1; zygomatic breadth, 14.0; mastoid breadth, 12; interorbital breadth, 2.8; breadth at base of canines, 8.4; maxillary breadth, 10.7; maxillary tooththrow (c-m³), 10.1; length of mandible, 18.6; mandibular tooththrow (c-m₃), 10.0.

Six adult skulls (5 females and the male type), total length, 28.4 (28-29); condylobasal length, 25.2 (24.5-25.7); zygomatic breadth, 15.4 (14.9-15.9); mastoid breadth, 12.1 (11.8-12.4); interorbital breadth, 2.9 (2.8-3.1); breadth at base of canines, 7.8 (7.2-8.4); breadth of rostrum (at point of greatest inflation), 8.2 (8-8.5); maxillary breadth, 11 (10.7-11.3); maxillary tooththrow (including canine), 10.1 (9.8-10.4); length of mandible, 19 (18.4-19.4); mandibular tooththrow (including canine), 11 (10.9-11.2).

P² is minute, and stands outside of the toothrow, the cingula of the canine and p³ being in close contact.

Posterior border of palate broadly U-shaped, with a slight median point, and extends to a line passing through the posterior border of m².

Represented by 14 specimens, of which 5 are alcoholic, collected as follows: Avakubi, 4, of which 3 were taken Jan, 24-26, 1914, and 1 in October, 1913; Niangara, 2, Dec. 3 and 6, 1910; Medje, 7, April 27 and Sept. 3, 1910; Niapu, 1, Oct. 28, 1914. Two of the Medje specimens are half-grown young, and two others are young

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adults, not fully mature as regards size. The others are fully adult, all with perfect skulls except the Niapu specimen, of which the skull is too much broken for measurement.

The specimens represent two seasonal periods, April, and September-January. The April specimens (from Medje), in worn pelage, are browner and the light tips of the hairs are stained faintly yellowish in contrast with the clear blackish brown ground-color and whitish hair-tips of the Septem-

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ber-October and January specimens. That the difference is probably purely seasonal is indicated by a Medje specimen taken in September, which is dark with white hair-tips, like the October-January specimens from other localities.

In its soft woolly pelage and other external features *Hipposideros langi* appears to resemble closely *Phyllorhina cyclops* Temminck, from the Boutry River, Guinea, but it is a much larger species and darker in coloration, being blackish brown with white hair-tips instead of dull brown with yellowish hair-tips. *H. cyclops*, however, appears to be little known, the only original references to it being Temminck's description and Dobson's account, based on two specimens, including one from the Cameroon Mountains, the other without definite locality. Neither of these descriptions is very satisfactory, the character of the skull not being mentioned in either. Dobson's measurements indicate a total length of about 93 mm., and Temminck's a total length of about 89 mm., as against an average of 113 mm. for 4 specimens of *H. Langi*. Dobson gives the length of the forearm as 62.2, as against an average of 68 in 10 specimens of *H. langi*. Temminck gives the length of the forearm as 38 mm. ("antibrachium 1 police 7 lignes"), which is obviously an error (possibly a misprint for 2 pouces 6 lignes = 63 mm.). The type localities of the two forms are widely separated. Dollman, however, has recorded two specimens from Avakubi (Rev. Zool. Africaine, IV, fasc., 1, 1914, p. 70), as "*Hipposideros cyclops*, Temm.," which probably are referable to the present form.

H. langi differs greatly from all the African species of the genus *Hipposideros* except *cyclops*, from which it is distinguishable by much larger size and darker coloration.

Hipposideros marisae Aellen, 1954

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TYPE. - M adulte, collection personnelle; n° original 400; Duékoué (Côte d'Ivoire), rocher de la Panthère Blanche, 13.5.1953.

DIAGNOSE. - *Hipposideros* de petite taille (avant-bras 41 mm.), voisin de *curtus* du Cameroun (avant-bras 44-47 mm.) dont il a les mêmes proportions externes, mais s'en distinguant par la feuille nasale plus petite qui ne possède qu'une seule foliole secondaire très rudimentaire (deux dans *curtus*) et par la forme du crâne dont la largeur zygomatique est subégale à la largeur mastoïde, alors que cette dernière est plus grande chez *curtus*.

DESCRIPTION. - Les oreilles sont grandes, presque aussi larges que longues, bien séparées sur le sommet de la tête, de même forme que chez *curtus*, c'est-à-dire que le bord interne est régulièrement convexe; le sommet à angle droit est arrondi et le bord externe présente une échancrure à 1.5 mm. du sommet; le lobe antitragal est séparé du pavillon par une deuxième échancrure moins importante; à ce niveau, il n'y a pas de pli interne ("internal fold") comme chez les espèces du groupe *bicolor*, auquel la nouvelle espèce pourrait s'apparenter.

Le sac frontal est très net et s'ouvre transversalement à 1 mm. en arrière du processus postérieur de la feuille nasale.

Celle-ci est du type *curtus* et ne diffère essentiellement que par ses dimensions nettement inférieures: la largeur du processus postérieur représente le 13 % de l'avant-bras pour 16-16.5 % chez *curtus*. Il n'y a qu'une seule foliole secondaire très petite et peu visible; en réalité, il s'agit plutôt d'une verrue allongée que d'une vraie petite feuille.

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Les plis palataux sont au nombre de six; les trois premiers sont nettement courbes et les trois derniers presque droits; le 3^e est situé entre les dents P⁴-M¹, le 6^e au milieu de M².

L'aile présente les mêmes proportions que celle de *curtus*. Les métacarpes III et V sont subégaux, le IV^e légèrement plus long.

Les tibias sont longs (44 % de l'avant-bras), de même que les pieds (env. 18 % de l'avant-bras). La queue est remarquablement courte (48 % de l'avant-bras) et libre sur les deux derniers millimètres.

Le patagium est inséré à l'extrémité distale du tibia, comme chez *curtus*.

La coloration est très uniforme: gris foncé dessus et dessous; les oreilles et le patagium sont brun-noir; la feuille nasale est noirâtre sur les bords et au processus postérieur, brun clair au processus médian (selle).

La forme générale du crâne est la même que chez *curtus* et *bicolor*. On peut remarquer la petitesse du rostre et la grande largeur zygomatique: le renflement nasal (mesuré au-dessus de l'insertion antérieure des arcades zygomatiques) n'est que le 46 % de la largeur zygomatique, alors que chez *curtus*, il représente le 54 % et chez *bicolor* le 50 %. Ce faible renflement nasal donne au crâne un profil supérieur peu concave, au-dessus des orbites. Les bulles tympaniques sont relativement petites. La largeur mastoïde est légèrement inférieure à la largeur zygomatique. La crête sagittale est faiblement développée.

Les dents ne présentent aucune particularité et sont tout à fait comparables à celles de *curtus*. Les incisives supérieures sont simples, sans trace de lobe postérieur et la couronne est dirigée vers l'intérieur. La canine est nettement séparée de la 2^e prémolaire (P⁴) par la 1^{re} prémolaire (P²) qui est très petite et située au bord externe de la rangée dentaire. La dernière molaire (M³) est réduite dans les mêmes proportions que chez les espèces du groupe *bicolor* et chez *curtus*. Les incisives inférieures sont tricuspidées, I₁ chevauchant I₂. La canine est étroite et la 1^{re} prémolaire inférieure (P₂) mesure environ la 1/2 de la hauteur de la 2^e (P₄).

REMARQUES SUR LA POSITION SYSTÉMATIQUE DE *H. marisae*. - Cette nouvelle espèce n'entre dans aucun des groupes définis par

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[Tables removed - Eds.]

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TATE (1941). Elle se rapproche le plus de *curtus* du Cameroun, que son descripteur, G. M. ALLEN (1921) compare à *beatus* et *caffer guineensis*. TATE place *curtus* dans le groupe *caffer* qu'il fait dériver du groupe indo-australien *galeritus*. Cependant, j'ai déjà attiré l'attention sur les affinités possibles de *curtus* avec le groupe indo-australien *bicolor* (ALLEN, 1952).

Il semble bien que l'on ne peut pas admettre *curtus* dans le groupe *caffer*. TATE a principalement étudié les rapports entre les formes orientales du genre *Hipposideros*; il dit précisément (1941, p. 353): "African species of *Hipposideros* haven been studied only in order to learn their relationships to Oriental species. No study has been made of sub-relationships within African groups, such as *caffer* and its allies."

Dans le tableau comparatif suivant, je résume les principaux caractères distinctifs de *marisae*, de *curtus* et des espèces des groupes *bicolor* et *caffer*. Le groupe *galeritus*, duquel le groupe *caffer* semble dérivé, présente généralement les caractères indiqués ci-dessous pour *caffer*.

Avec ces données, on se rend compte que les espèces *marisae* et *curtus* s'écartent notablement du groupe *caffer*; elles de rapprochent beaucoup du groupe *bicolor*, mais les différences me

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[table removed - eds.]

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semblent suffisantes pour justifier la création d'un nouveau groupe qui est ainsi défini:

Groupe *curtus*, n. gr.

Oreilles grandes (40-45 % de la longueur de l'avant-bras), plus longues que larges, à bord externe concave sans pli interne à la base. Sac frontal présent chez le mâle, parfois absent chez la femelle (type de *curtus*). Feuille nasale de taille variable, mais présentant une cloison internariale élargie, claviforme; 1 ou 2 petites folioles secondaires. IV^e métacarpe plus long que les III^e et V^e. Indice de largeur 19-22, indice digital 1.15-1.17. P² petite, située au bord externe de la rangée dentaire, séparant C de P⁴.

Hipposideros nanus J.A. Allen, 1917

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Type No. 49426 (alcoholic), F ad., Faradje, Uele district, Belgian Congo, Oct, 24, 1912; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 1689.

A very small species of the *H. beatus* type.

Upperparts dusky brown, the hairs dark at base and tips with a broad median zone of white; underparts lighter, the hairs with grayish tips; ears and membranes dark brown.

Total length, 68 mm.; head and body, 46; tail, 22; forearm, 43.4; third metacarpal, 31; tibia, 16.5; foot, with claws, 7.9; ear, length, 9.2, width, 12; greatest breadth of horseshoe, 5; of posterior leaf, 5.5.

Skull, total length; 16.2 (16.8) ; zygomatic breadth, 8.2 (9.7); mastoid breadth, 8.5 (9.2); maxillary breadth, 5.5 (7), breadth at base of canines, 4.5 (4.3-4.7); mandible, 9.5 (10.7-11.3); upper tooththrow (c-m³), 5.2 (5.9-6.2); lower tooththrow, 5.7 (6.2-7).

Represented by only the type.

Hipposideros nanus is nearly related to *H. beatus* Andersen (l. c., p. 279), from near Benito River, Guinea, and may be considered as its geographical representative in the Uele district of northeastern Belgian Congo.

Hipposiderus beatus K. Andersen, 1906

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Smaller than *H. caffer*, with very short tail and tibia. Skull small and very broad-jawed.

In all forms of *H. caffer* the maxillary width (across the antero-external corners of *m*³) is practically equal to the length of the maxillary tooth-series; in *H. beatus* the former is markedly greater than the latter (7 mm. as against 5.9-6.2); the great maxillary width, combined with the small size of the skull, makes the cranium of *H. beatus* easily distinguishable from that of any race of *H. caffer*. The zygomatic width is larger than the mastoid width, as in *H. c. centralis* and *guineensis*.

The teeth are of the same size as in the small-toothed *H. c. typicus*. The dentition, although in all essential respects

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as in *H. caffer*, seems to be a trifle more advanced: *p*² is in all the three specimens examined exceedingly small, so small indeed as to be very easily overlooked.

Externally this bat is readily distinguished from *H. caffer* by its small size and very short tail and tibia (see measurements below, p. 282). The wing-membrane is inserted on the middle or distal part of the metatarsus or on the base of the phalanges; in *caffer* it is never produced further backwards than the base of the metatarsus, and this but very rarely.

Type: - F ad. (in alcohol). 15 miles from Benito River; Feb. 1899. Collected by G. L. Bates, Esq. Brit. Mus. no. 0.2.5.45. - A second specimen (Brit. Mus. no 5.5.23.11), also obtained by Mr. Bates, is from Efulen, Cameroons. A third specimen +, from Mt. Coffee, Liberia is preserved in the Washington Museum (no. 83857) ++.

Judging from this, *H. beatus* is distributed over the countries bordering the Gulf of Guinea, from Benito River to Liberia. This it inhabits the same region as the large-skulled and large-toothed *H. c. guineensis*.

* Dentition in 76 skulls of *H. caffer* (all races): - *p*₃ always wanting. *p*₂ and *p*₄ never separated; in 10 specimens in simple contact, in 66 overlapping each other at bats. *p*² always external to the series and always easily observable. Upper canines and *p*⁴ in 21 specimens distinctly separated; in 39 extremely slightly separated or almost in contact; in 3 completely in contact on one side of the jaw only, in 13 on either side.

+ For the loan of this specimen I am indebted to the Authorities of the United States National Museum. It is one of the *Hipposiderus caffer* mentioned by Gerrit S. Miller in his paper on a collection of small mammals from Mount Coffee, Liberia (Proc. Wash. Acad. ii (1900) p. 647; forearm 42 mm.).

++ A fourth specimen, not examined by me, is in a Continental (probably Swedish or German) Museum: - In his "Säugethiere aus Kamerun, West-Afrika" (Bih. Kgl. Svenska Vet.-Akad. Handl. xxiii. Afd. iv. no. 1, p. 18; 1897) Dr. Yngve Sjöstedt gives some external measurements of 10 "*H. caffer*"; no. 1 is Sundevall's type, from Port Natal; nos. 2-8 and 10, all from Cameroon, are probably *H. c. guineensis*; no. 9, also from Cameroon, with the forearm measuring 44 mm., the tail 24, and the tibia 16, is undoubtedly a *H. beatus*.

Hipposiderus caffer centralis K. Andersen, 1906

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Large-toothed, large-skulled, and broad-jawed.

The skull is larger and in every respect more heavily built than in the typical form. The zygomatic width is almost invariably slightly larger than, or at least equal to, the mastoid width, which gives the skull in upper view a very characteristic aspect as compared with that of *H. c. typicus*. The maxillary width is markedly larger: 6.8-7.1 mm., as against 6-6.2 in the typical form; in conformance with this the width across the canines and the anteorbital width are slightly larger. The teeth are considerably larger, the mandible longer.

Externally, this form is on an average larger than *H. c. typicus*, but the difference is practically far less well marked than in the skulls and teeth (for details, see *H. c. typicus*). The colour of the fur, both in the dark and red phase, is darker than in the typical form, but lighter than in *H. c. guineensis* (see this latter, below).

Type: - M ad. (skin). Entebbi, Uganda. Presented by F. J. Jackson, Esq. Brit. Mus. no. 99.8.4.8.

26 specimens have been examined, from: - Takaungu, Mombasa, British East Africa (4); Dar es Salam, German East Africa (1); Zomba, Nyasa (1) §; Entebbi, Uganda (9); Stanley Falls, Upper Congo (3); Leopoldville, Lower

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Congo (2)*; Wathen, Lower Congo (1); 75 miles up the Congo River (4)*; Caijala, Bihé, Angola (1)+. - 16 skulls, from all the localities enumerated.

According to this, *H. c. centralis* is distributed in a broad belt across the Equatorial region of Africa, from British and German East Africa and Nyasaland in the East, through Uganda and the whole of the Congo Valley, to the western coast of the continent; like the typical form it extends to Angola.

Hipposiderus caffer guineensis K. Andersen, 1906

The extreme in the maxillary width of the skull and the intensity of the colour of the fur.

The skull and teeth of this form are of the same size as in *H. c. centralis*; but the maxillary width on an average decidedly larger: 7-7.7 mm., as against 6.8-7.1.

External dimensions as in *H. c. centralis*.

The colour of the fur is markedly darker than in any other race: - Back approaching "seal-brown," base of hairs scarcely lighter; upperside in front of the shoulders "hair-brown," base of hairs next to "bistre"; underside dull "drab," base of hairs next to "bistre." - I have seen no very young specimens of this form.

Also the red phase is darker than in the other races: - Upperside "cinnamon-rufous," in some individuals so dark as to approach "chestnut"; underside "cinnamon-rufous" or "hazel." Different at a glance from the corresponding phase of the typical form.

Type: - F ad. (skin). Como River, 70 miles from Gaboon, almost sea-level; June 3rd, 1897. Collected by G.L. Bates, Esq. Brit. Mus. no. 97.12.1.11.

27 specimens examined, from: - Como River (4); Gaboon (1); Benito River (4); Fernando Po (9)++; Cameroon Mts. (1); Efulen, Bulu Country, Cameroon, 1500-1800 ft. (4); Old Calabar (1); Mt. Coffee, Liberia (3) §. - 23 skulls, from all the localities enumerated.

According to this, *H. c. guineensis* is distributed from

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Como River westwards, through the countries bordering the Gulf of Guinea (including the island of Fernando Po), at least as far as Liberia.

Hypsignathus H. Allen, 1862

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Head large, massive. Face greatly developed, anterior portion humped. Nose very high, blunt, corrugated. Nostrils produced; opening laterally; bounded internally and inferiorly by a projection of the coriaceous membrane, which by its continuation externally forms a lateral fold. This expansion, after descending from the posterior part of the nostril to the lip, runs along the line of the upper jaw, forming the outer wall of a distinct groove, the inner wall of which is made by the true lip. Just as this remarkable membrane turns down from the anterior nares, it also extends forward, clearly defining the boundaries of the snout. From the lower anterior part of each nostril a leathery ridge extends to the mouth. These divide the muzzle into three distinct sulci. The chin is peltated, and indistinctly divided into halves by a mesial line. Ears small, naked, without tragus and tufted at base. Wings thrown very far back. Basal joint of thumb small. Index finger clawed. Interfemoral membrane small, ecaudate. Lower incisors closing anterior to the upper.

Dental Formula, $m \frac{3}{5}, c \frac{1}{1}, i \frac{4}{4}, c \frac{1}{1}, m \frac{3}{5} = 28$.

Skull remarkably high. Nose broad at summit, occasioned by the development of the nasal bones. Extending from the canines to near the top of the face is an irregular ridge for the insertion of the curious nasal membrane already noticed. The infra-orbital foramen opens posteriorly to the entire dental series. The skull is broader between the eyes than in other Pteropines. Post-orbital processes stout, short and pointed outwards and backwards. Cranium small, comprising but a third of the entire head. A parietal crest is present for about two-thirds its length, when it abruptly terminates at the apex of a small triangle which is formed by the want of approximation of the hinder

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portion of the temporal fossæ. Occipital ridge well marked and leaning backwards. Foramen magnum orbicular. Palatal bones united with the deeply arched mouth by a greater angle than that of *Pteropus*. The foramen incisivum is cordate.

The *lower jaw* is flattened and irregular, depressed at symphysis. The two halves unite at a less acute angle than in *Pteropus*. The coronoid process is low, and the angle which it forms with the alveolar ridge is so slight that the distance from the top of the process to the last molar is equal to one half the distance from the same point to the small pre-molar. The condyloid process is about one-half the distance between the base of the jaw and the top of the coronoid.

The superior incisors are small unicuspid, regular and separated from one another. The distance between the canines and the laterals is greater than the distance from one incisor to another. The canines are slender, convex anteriorly, point slightly backward, and, when the jaws are closed, nearly touch the plane on which the skull rests. The first molar is pointed; the second and third are much alike, the posterior being smaller and less trenchant, the external cusp being the larger.

The inferior incisors are small, separated, the space between the centrals exceeding that between the laterals. The canines are smaller and blunter than those above, and lean strongly backwards. The premolar, which is absent above, is here present; it is very minute, shaped, like an incisor, is nearer the canine than the second molar, is directed backward and outward, and, when the jaws are closed, is observed to be placed *anterior* to the superior cuspidatus. The first and second molars assume the carnivorous type; the second has two cusps divided as usual by a longitudinal groove; the third and fourth have their cusps much worn, the latter being little more than flattened tubercles.

As mentioned above, the articulation is curious; the inferior incisors close in front of the superior, so as to completely hide them.

Kerivoula africana Dobson, 1878

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Ears and tragus almost quite similar to those of *K. hardwickii*, but longer than the head; thumb shorter; wings from the base of the toes.

The fur extends upon the base only of the interfemoral membrane and along the legs to the backs of the feet; a few fine hairs extend along the calcanea and margin of the interfemoral membrane, but not so thickly as to form a fringe. The face is nearly naked between the eyes, but long hairs arise from the glandular eminences in front, and also form a fringe along the margin of the lips.

On the upper surface the fur is dark brown at the base, the extremities greyish brown; beneath similar, but paler.

Inner upper incisor on each side long, with a distinct outer cusp; outer incisor unicuspidate, exceeding the outer cusp of the inner incisor, and nearly equalling that tooth in vertical extent. First upper premolar smaller than the third and equal to the second premolar; lower premolars equal.

Length (of the type, a M, preserved in alcohol), head and body 1".35, tail 1".35, head 0".5, ear 0".5, tragus 0".3, forearm 1".1, thumb 0".2, third finger 2".15, fifth finger 1".6, tibia 0".45, calcaneum 0".5, foot 0".25.

Hab. East coast of Africa (Zanzibar).

Type in the collection of the Paris Museum.

This is the smallest species of *Kerivoula* as yet described. It resembles *K. hardwickii* very closely in general form, but may be at once distinguished by the upper incisors.

Kerivoula argentata Tomes, 1861

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In the Proceedings of this Society for 1858 I described at some length the peculiarities of the present group of Bats, giving my reasons for regarding these peculiarities as generic. All that was stated respecting the several species then enumerated may with equal exactness be said of the present one; and I may add that I have since that time again examined the fine collection of Bats in the Leyden Museum, and that, with the exception of identifying my specimens of *K. papillosa* with the *Vespertilio papillosus* of M. Temminck, and examining a specimen of my *K. aërosa*, labelled "Gorontalo," I saw nothing which in any way either confirmed or modified my previous opinions of the genus.

The present species is of larger size than either of the African species before described, fully equal in size to the *K. papillosa*, and in the colour of its fur it differs from all the other species.

Compared with the African species which it most nearly resembles, it possesses some of the characters of both *K. lanosa* and *K. aërosa*.

The top of the head is elevated in about the same degree as in *K. aërosa*; and it greatly resembles this species in the shape of the muzzle and the distribution of the hair on the face; but in the shape of the ears it approaches more nearly to *K. lanosa*, differing from it only in having the inner rounded margin toward the top of the ears more prominent even than in that species. The tragus is remarkably narrow, and tapers evenly to an exceedingly acute point; near the bottom of its outer edge is a narrow notch, or rather slit, and below it a small and pointed process, which is placed, in fact, immediately above what may be

called the foot-stalk or narrow root of the tragus. Inside the ear, and vertically beneath the tragus, is a well-defined fleshy tubercle, of a flattened form, and about one line in length.

In the quantity and distribution of the fur on the membranes, this species is intermediate between *K. lanosa* and *K. aërosa*, but it has fewer adpressed hairs on the wings than either. The fur of the back extends on to the membranes of the flanks a little, and on to the interfemoral membrane in a scattered manner, but more thickly on to the tibiæ and feet, especially on to the latter, which are well clothed. Beneath, it extends a little on to the membranes near the sides of the body. The *os calcis* is well clothed with short adpressed hairs; and between it and the tail-tip the membrane is fringed with closely-set hairs, which curve downwards and have a comb-like appearance, as in *K. lanosa*.

The fur is everywhere long and silky; that of all the upper parts is of four colours - at the root very dark grey for a fourth of its length, then yellowish, passing into a pale but bright rust-colour, and the tips of the hairs of a shining and silvery white. There is very little variation in the colouring of the different parts of the upper surface. Beneath, the fur is unicoloured and dirty-white, on the sides of the neck and on the cheeks-tinged with rust-colour.

The teeth, as far as may be gathered from inspection without re-

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moving the skull from the specimen, are like those of *K. lanosa*, the upper incisors being, as in that species, nearly of the same length. The outer incisors in the lower jaw have the singular and prominent cusp, which I have mentioned as peculiar to the genus, quite as much developed as in any of the species.

""
Length of the head and body, about: 2 0
----- of the head: 0 8
----- of the ears: 0 5
Breadth of the ears: 0 5½
Length of the tragus: 0 4
Greatest breadth of the tragus, barely: 0 1
Length of the fore-arm: 1 6
----- of the thumb and its claw: 0 4½
----- of the first finger: 1 6
----- of the second: 3 6
----- of the third: 2 6
----- of the fourth: 2 4
----- of the tibia: 0 7½
----- of the foot and claws: 0 4½
----- of the os calcis: 0 10
Expanse of wings, about: 12 0

Hab. "Otjoro, December 1st, 1859. Female."

[*Kerivoula brunnea*](#) Dobson, 1878

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Ears slightly shorter than the head, laid forwards the extremities do not extend quite to the end of the muzzle; ear-conch very similar in outline to that of *K. picta*; tragus with a slight indication of a lobe at the base of the outer margin, which, however, is not succeeded by an emargination, and the projection opposite the base of the inner margin is obtuse.

Fur, above, yellowish brown, the basal half of the hairs very dark brown or black; beneath paler or grayish brown, the base of the hairs also dark-coloured; very long and dense, covering nearly half the upper surface of the interfemoral membrane, and extending along the legs to the backs of the feet; the posterior margin of the interfemoral membrane has a comb-like fringe of curved hairs of equal length, as in *K. picta* and *K. lanosa*. Membranes dark brown.

Upper incisors nearly equal in length and in cross section, the inner incisor on each side with a small postero-external basal cusp visible from the front; of the lower incisors the outer on each side has a blunt cusp rising considerably above the others; second upper premolar slightly smaller than the first, with a more oblique cusp inclined forwards.

Length (of a not full-grown M), head and body 1".5, tail 1".9, head 0".55, ear 0".5, tragus 0".35, forearm 1".3, thumb 0".25; third finger - metacarp. 1".25, 1st ph. 0".65, 2nd ph. 0".8; fifth finger - metacarp. 1".2, 1st ph. 0".4, 2nd ph. 0".3; tibia 0".55, foot 0".3.

Hab. Uncertain.

This species is at once distinguished by the very dark colour of its fur, by the comb-like fringe along the interfemoral membrane behind, and by the large size of the upper outer incisors.

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The type specimen has suffered from long soaking in weak alcohol, which has denuded it of much of its fur, and it is possible that the anterior extremities may have had short hairs clothing them as *K. lanosa*, especially as some hairs are found still adhering to the outer margin of one wing.

A. M (not full-grown), al. Madras or South Africa. Sir A. Smith [P.].

[*Kerivoula cuprosa*](#) Thomas, 1912

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A small speckled brown species with short incisors.

Size decidedly less than in the related species *K. aërosa* and *lanosa*. Fur soft and fine (hairs of back 6 - 6.5 mm. in length), extending on to the forearm, thinly along the pollex and terminal part of the third digit, and down the upper side of the legs on to the feet; proximal part of interfemoral thinly haired, naked distally, the hinder margin with a

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number of fine hairs not forming a fringe; under surface of legs and interfemoral nearly naked. General colour above dark bistre-brown, the tips of many of the hairs conspicuously contrasted silvery buff, those on the forearms, rump, and hind limbs more ochraceous buff; under surface duller brown, the bases of the hairs dark slaty, the tips of some of the hairs whitish.

Ears of medium length, anterior border strongly convex, posterior with a sharp concavity just below the tip. Tragus long, curved outwards, its base with a small lobule externally succeeded above by an emargination.

Upper incisors unusually short, their enamel-covered portion but little longer than the projecting part of their root, the inner one bicuspid, its posterior cusp as thick as and half the height of the anterior. Outer incisor also bicuspid, owing to its basal ledge being raised up postero-internally as a second cusp half the height of the main cusp. Middle premolar about two-thirds the height and area of the anterior one. Outer lower incisors tricuspid, the outer cusps half as large as the median one. Middle lower premolar rather smaller than the subequal first and last.

Dimensions of the type (the starred measurements taken in the flesh by the collector): -

Forearm 32 mm.

Head and body *45; tail *45; ear *13.5; tragus on inner edge 5.5; third finger, metacarpal 3.1, first phalanx 15; lower leg and foot (c. u.) 21.

Front of upper canine to back of m^3 5.1; front of lower canine to back of m_3 5.5.

Hab. Bitye, Ja River, S.E. Cameroons. 2000'.

Type. Adult male. Original number 564. Collected 17th October, 1911, by Mr. G. L. Bates.

This well-marked species resembles *K. aërosa* in colour, but is much smaller, the forearm of that animal being 37 mm. in length. By the characters used in Dobson's synopsis it comes nearest to *K. lanosa*, but differs both by size and colour, and, as from every other, by its peculiarly short and deeply bicuspidate upper incisors. At Bitye Mr. Bates also obtained the little *K. muscilla* and an additional specimen of *K. smithii*, described by me in 1880. The latter is slightly larger than *K. cuprosa* and has practically unicuspid outer lower incisors.

[*Kerivoula harrisoni*](#) Thomas, 1901

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Most nearly allied, according to the characters used in Dobson's synopsis, to *K. lanosa* A. Smith, but considerably smaller. Fur very long, soft, and woolly, the hairs of the back about 9 mm. in length. Muzzle thickly hairy. Back of ears hairy, except at the edges, these and the whole inner surface practically naked. In shape the ears are very much as in *K. lanosa*; their inner margin slightly convex below, more strongly so above; tip inconspicuous, behind and below the level of the most convex point of the anterior edge; below the tip a marked concavity, after which the hinder edge is evenly convex to its base. Tragus, so far as can be seen in the dried skin, very like that of *K. lanosa*; its basal lobe small.

Fur of back not extending on to the wing-membranes, but the interfemoral is clothed basally with scattered long hairs; the legs are thickly clothed down to the toes; the upperside of the tail is well haired to the tip, and the interfemoral membrane has a thick fringe of hairs along its posterior edge. The forearm is irregularly tufted with line golden hairs, which are continued on the thumb to the base of its claw, and again along the final edge of the second digit, edge of wing-membrane, and terminal part of third digit to the extreme tip of wing.

The general colour of the back is a dull "old gold," the woolly hairs being blackish basally, then dull huffy orange with their tips blackish, longer hairs tipped with silvery yellow. The hairs on the legs, feet, tail, and calcars, like those on the forearm, golden yellow, but the interfemoral fringe is a dull pale brown. Belly-hairs blackish basally, dull buffy white terminally.

Skull very fine and delicate, on the whole very like that of *K. brunnea* Dobs., though the nasal notch is not so deep.

Upper outer incisors nearly as long as the inner ones, the former with a low postero-internal secondary cusp, the latter with a posterior one at about the level of the tip of i^2 .

Forearm 31.5 mm. Front of canine to back of m^3 5.2.

Hab. Walamo, between Lakes Suai and Margherita, N.E. Africa. Alt. 6700 feet.

Type. B.M. No. 0.11.4.1. 21st February, 1900. Collected and presented by Mr. J. J. Harrison.

No species hitherto described can be confused with *K. harrisoni*. *K. lanosa*, is larger and of a different colour; *K. smithi* has no interfemoral fringe; *K. ærosa* has minute outer incisors; and *K. brunnea* and *K. africana* have the same teeth unicuspidate, and the tragus of a different character.

Kerivoula harrisoni bellula Aellen, 1959

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Type. - M adulte, en alcool. Adiopodoumé, près d'Abidjan, Côte-d'Ivoire, 20.7.1953; coll. V. Aellen, n° orig. 622. Muséum d'Histoire naturelle de Genève, n° 965.38.

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Matériel examiné. - De la Côte-d'Ivoire, j'ai rapporté un autre spécimen provenant aussi d'Adiopodoumé, 21.12.1953, leg. U. Rahm (F n° 965.39 Mus. Genève). Les exemplaires suivants ont encore été examinés :

1 M subadulte, Hill-Town, Du Queah River, Libéria, 25.2.1887. Musée de Leyde. Signalé par JENTINK (1888) sous le nom de *Kerivoula africana*.

1 M; 1 ?, Ntronang, Ghana, 2.9.1945. British Muséum n° 46.118 et 46.119.

1 M, Mukonje, Cameroun anglais, 1905. Musée zoologique de Strasbourg, n° 20g. Signalé par AELLEN (1957) sous le nom de *Kerivoula* sp.

1 F, Pawa, district Kibali-Ituri, Congo belge. Musée royal du Congo belge, Tervuren n° 15729. Indiqué par SCHOUTEDEN (1944) sous le nom de *Kerivoula ? argentata*.

Diagnose. - *Kerivoula* de petite taille (avant-bras de 28,5 à 33 mm) à frange interfémorale très nette. Les premières incisives supérieures (i^1) sont bicuspidées. Le rostre est long, c'est-à-dire que la rangée dentaire C-M³ est égale ou plus longue que la largeur maxillaire (M³-M³). Diffère de *K. h. harrisoni* par ses oreilles qui sont plus longues: 11 à 12,5 mm au lieu de 8,8 mm; le tragus est plus long également: 5,8 à 6,5 mm au lieu de 5,3 mm.

Remarques. - Depuis quelques années, j'ai entrepris la révision des formes africaines du genre *Kerivoula*. Cette étude m'amène à croire que *K. lucia* Hinton n'est qu'une forme de *K. harrisoni* Thom. et non de *K. lanosa* (A. Smith), comme le pensent ROBERTS (1936) et ELLERMAN, MORRISON-SCOTT et HAYMAN (1953). En effet, chez *K. harrisoni* et *K. lucia*, le rostre est relativement long (C-M³ = ou > M³-M³) et les premières incisives supérieures (i^1) sont bicuspidées, alors que chez *K. lanosa*, le rostre est court (C-M³ = ou < M³-M³) et les premières incisives unicuspidées ¹.

K. harrisoni n'est connu que par la description originale de THOMAS (1901), basée sur l'examen d'un seul spécimen (British

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Muséum n° 0.11.41, peau sèche). Il existe au British Muséum un second spécimen, provenant de Taveta, Kenya (n° 9.6.12.18, en alcool), dont M. R. W. Hayman m'a aimablement communiqué les principales mensurations.

K. lucia, par contre, est mieux connu; j'en ai examiné une demi-douzaine d'exemplaires du SE du Congo belge, du Ngamiland et du Natal. Très semblable à *K. harrisoni*, il n'en diffère guère que par une oreille et un tragus plus longs, des proportions légèrement différentes du crâne.

Si mes conclusions sont correctes, la répartition géographique de *K. harrisoni* se présenterait ainsi:

K. h. harrisoni: Ethiopie, Kenya.

K. h. lucia: SE du Congo belge, N Rhodésie, Nyasaland, Ngamiland, Natal.

K. h. bellula: région forestière du Libéria à l'E du Congo belge: Libéria, Côte-d'Ivoire, Ghana, Cameroun, Congo belge.

Description. - Les oreilles mesurent de 11 à 12,5 mm; le tragus de 5,8 à 6,5 mm au bord interne. Le lobule de la base du bord externe du tragus, qui sert de principal caractère dans le synopsis des espèces de DOBSON (1878), semble variable d'un spécimen à l'autre: chez le type, il est à peine indiqué, alors que chez le paratype, provenant du même lieu, il est petit, mais bien marqué; il est présent sur les spécimens du Cameroun et du Congo, peu marqué chez celui du Libéria, à peine indiqué ou absent chez ceux du Ghana. Les avant-bras, les doigts et les jambes sont de dimension et de proportion semblables à celles de *harrisoni* et de *lucia*. L'avant-bras est peut-être à peine plus court: *bellula*, moyenne 6 spécimens: 30,75 mm; *lucia*,

moyenne 8 spécimens: 31,33 mm; *harrisoni*, moyenne 2 spécimens: 31,55 mm. La frange interfémorale du bord libre de l'uropatagium est très bien marquée, comme d'ailleurs chez *harrisoni* et *lucia*.

La coloration n'offre rien de particulier pour un *Kerivoula*. Les descriptions qu'en donnent THOMAS (1901) et HINTON (1920) s'appliquent presque mot pour mot au type de *bellula*. Le dessus est gris clair un peu brunâtre; les poils sont gris souris à la base, brun très clair au sommet; beaucoup présentent une

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pointe blanc brillant, ce qui donne un reflet argenté surtout visible au niveau des jambes. Les avant-bras, le pouce, le deuxième doigt, la base du métacarpe du cinquième, les jambes, les pieds et la plus grande partie de la queue sont couverts de poils plus ou moins longs de couleur jaune paille; ces poils sont particulièrement longs sur les jambes et donnent à celles-ci un reflet doré. Il y a quelques longs poils épars sur la moitié proximale de l'uropatagium. Le dessous est assez semblable au dessus, mais moins brunâtre; le reflet argenté s'étend surtout à la poitrine; les avant-bras, les doigts, les jambes et la queue sont nus; il y a quelques poils très courts, groupés par deux ou trois, sur les lignes glandulaires transversales de la moitié proximale de l'uropatagium. Les oreilles sont brun clair, un peu translucides; la moitié basale et le tragus sont crème. Les glandes faciales, bien que recouvertes de poils, sont très apparentes et de couleur orange vif; cette couleur est due à leur sécrétion grasse.

Les premières incisives supérieures (I^1) sont bicuspidées; la pointe postérieure atteint presque la hauteur de la seconde incisive. Celle-ci mesure environ les quatre cinquièmes de la première, en hauteur. La première incisive inférieure est quadricuspidée, la seconde tricuspidée, la troisième tricuspidée avec la pointe médiane plus développée que les latérales. Ces caractères dentaires se présentent, non seulement chez le type, mais aussi chez tous les *K. h. bellula* examinés.

Dans le tableau de mesures suivant, j'ai indiqué, à titre de comparaison, celles du type de *K. h. harrisoni* et d'un autre spécimen du Kenya (mesures communiquées par R. W. Hayman) et les minima et maxima de *K. h. lucia*, déduits des données de la littérature et d'examen personnels.

A l'échelle spécifique, *K. harrisoni* se distingue des autres *Kerivoula* africains à frange interfémorale nette par les caractères de la clé suivante:

1. Taille grande: avant-bras + troisième doigt de 109 à 128 mm. Long. totale du crâne (aux incisives) de 15 à 16 mm. I^1 unicuspidée (parfois une deuxième pointe postérieure basse). M^3-M^3 généralement $> C-M^3$. --> *argentata* Tomes

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- Taille moyenne ou petite: avant-bras + troisième doigt de 88 à 106 mm. Long. totale du crâne de 12,7 à 13,6 mm. --> 2

2. I^1 bicuspidée. Rostre long: $C-M^3$ généralement $> M^3-M^3$. --> *harrisoni* Thom.

- I^1 unicuspidée. Rostre court: M^3-M^3 généralement $> C-M^3$. --> 3

3. Taille moyenne: avant-bras de 30 à 35,5 mm. Troisième doigt de 65 à 72 mm. Long. totale du crâne de 13,2 à 13,6 mm. --> *lanosa* (A. Smith)

- Taille petite: avant-bras de 29 à 31 mm. Troisième doigt environ 60 mm. Long. totale du crâne 12,7 mm. --> *muscilla* Thom.

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Kerivoula harrisoni bellula: Côte d'Ivoire: Mus. Genève 965.38 M type, 965.39 F, Liberia: Musée Leyde M subad., Ghana: British Mus. 46.118 M, 46.119 sex ?, Cameroun: Musée Strasb. 20 g M, Congo: Musée Tervuren 15729 F;
K. h. harrisoni: Ethiopie, Kenya: British Museum 0.41.4.1 sexe ? type, 9.6.12.18 sexes ?;
K. h. lucia: (S Afrique 8 spécimens min - max)

Avant-bras: 29.5, 31.5, 29.5, 28.5, 33, 30, 32; 31.5, 31.6; (30 - 33)
3e doigt, métacarpe: 28.5, 30.5, 27, 30.5, 31.5, 29.5, 31.5; 31, 30; (29.5 - 33.5)
" 1^{re} phalange: 14, 15.5, 15.5, 14, 15.5, 14.5, 16; -, 15.7; (15 - 16.5)
" 2^e phalange: 20.5, 22, 17.5, 19, 17.5, 19.5, 20.5; -, 20.5; (18 - 23)
4e doigt, métacarpe: 28.3, 30, 26.5, 29.5, 31.5, 29.5, 30; 29.2, 30; (29 - 33)
" 1^{re} phalange: 10, 10.5, 10, 10.5, 10.5, 10, 11.5; -, 11; (9.5 - 12)
" 2^e phalange: 8, 8.5, 8.5, 8, 9, 8.5, 9.5; -, 8; (8 - 10.5)
5e doigt, métacarpe: 28, 29.5, 26.5, 29, 31.5, 29, 30; 29.8, 29.8; (29 - 33)
" 1^{re} phalange: 8.5, 9, 9, 9, 9, 10; -, 9; (8.5 - 12)
" 2^e phalange: 8.5, 8, 8, 7.5, 8.5, 8, 8.5; -, 6.8; (7 - 9.5)
Tibia: 13.5, 14, 11, 13, 13, 13, 14; -, -; (11.5 - 13.5)
Pied (avec griffes): 6.5, 6, 7.5, 6.5, 6.5, 7, 6.5; -, 6; (5.5 - 7)
Queue (de l'anus): 39, 36, 36, (34), -, 33, 38.5; -, 34; (36 - 42)
Oreille (longueur): 12.5, 11.5, 11, 12, -, 11, 12.5; -, 8.8; ((12) - 13.5)
Tragus: 6.5, 6.5, 6, 6.1, -, 5.8, 5.8; -, 5.3; (6.7 - 7.8)

Crâne:

Long, tot. (avec incisives): 13.4, 13, -, 13, -, 12.7, 12.8; -, 12.8; (13 - 13.5)
" (au prémaxill.): 13.1, 12.7, -, 12.6, -, 12.5, 12.5; -, -; (12.9 - 13.4)
" condylobasale: 12.1, 11.9, -, 11.8, -, 11.7, 11.8; -, -; (12.2 - 12.5)

Larg. zygomatique: 8.2, 8, -, 8, -, 7.8, 8; -, 8; (7.7 - 8.4)
 " mastoïde: 7, 6.9, -, 6.9, -, 6.7, 6.9; -, -; (6.8 - 7.5)
 " C-C (externe): 3.1, 2.8, 2.5, 3, -, 3.2, 3; 3, 3; (3 - 3.2)
 " M³-M³ (externe): 5.1, 5, 4.5, 5.1, -, 4.8, 5; 5.2, 5; (5 - 5.4)
 Long, rangée dent. I-M³: 6, 5.9, 6, 5.9, -, 5.9, 5.8; -, -; (5.9 - 6.1)
 " " " C-M³: 5.3, 5.2, 5.1, 5.1, -, 5.1, 5.2; 5.2, 5.3; (5 - 5.4)
 " mandibule: 9.6, 9.2, -, 9.2, -, 9.1, 9.6; -, -; (9.6 - 10)
 " rangée dent. I-M₃: 6.3, 6.2, 6.1, 6.2, -, 6.2, 6.2; -, -; (6.4 - 6.5)
 " " " C-M₃: 5.5, 5.3, 5.2, 5.4, -, 5.3; -, -; (5.5 - 5.6)

Footnote 1. Ce dernier caractère m'a été confirmé par R. W. Hayman, qui a réexaminé le crâne du type (in litt. 31.1.1955).

Kerivoula lucia Hinton, 1920

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Kerivoula lucia, sp. n.

Type. - An adult male collected at N'dola on Sept. 26, 1919 [by Capt. Guy C. Shortridge (see page 239)]; original number 472. No other specimen seen.

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This species closely resembles *K. lanosa* in general appearance, but it is distinguished by its rather smaller size, smaller and less hairy ears, greyer colour, and by some characters of the skull.

Size small, forearm 30.5 mm. Fur on body and top of head very long, dense, and woolly, closely resembling that of *lanosa* in quality. Cheeks in front of ears nearly naked. Ears very sparingly haired on outer surface, nearly naked within. Forearm, thumb, and outer edge of dorsal surface of wing clothed with tufts of hair; upper surface of tibia, hind foot, and tail similarly clothed. Interfemoral membrane with tufts of hair along the veins on dorsal surface, similar but smaller tufts of hair on ventral surface; with a well-developed posterior fringe. Ears smaller than in *lanosa*, with a somewhat deeper, though narrower, lateral emargination towards the tip; tragus normal.

General colour of back between "sepia" and "dusky drab," passing to a light grey on top of head and muzzle. Dorsal hairs with slaty bases, mostly with long yellowish-brown tips; in many the tips are silvery, and these silver tips produce a quite conspicuous "lining" on the back and rump. Under surface silvery grey, darkened irregularly by the partly visible slaty bases of the hairs. Hairs on forearm, wings, legs, tail, and the upper surface of the interfemoral membrane yellowish; those on the ventral surface of the membrane are silver.

Skull about as large as that of *K. lanosa*, from which it is distinguished by its relatively narrower brain-case, more boldly convex frontal region, narrower rostrum, and more nearly parallel tooth-rows, the width between the outer borders of the last molars markedly less in proportion to the width across the canines. Dentition not essentially different. Outer upper incisor about three-fourths the height of inner incisor, rather stouter than the latter in cross-section, and with a well-developed internal basal cusp. Inner incisor with a posterior secondary cusp, the summit of which is a little less lofty than the outer incisor. Middle upper premolar smaller in cross-section than the anterior premolar.

Collector's measurements (taken on the flesh). - Head and body 39 mm.; tail 40; hind foot 6; ear 12.

Wing-measurements (from skin). - Forearm 30.5; third digit, total length 66, its metacarpal and phalanges 1 and 2 being respectively 32, 16, and 18; fifth digit 46.5, the metacarpal and phalanges being 30.5, 9, and 7.5.

Skull measurements (those of 7.1.1.538, a cotype of *lanosa*, being added in parentheses for comparison). - Greatest length 13 (13.4); condyle to canine 11.5 (11.5); canine to m³ 5.1 (5.2); width across canines

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3 (3.2); width across outer .borders of m³-m³ 5.1 (5.5); zygomatic breadth 7.7 (7.9); interorbital breadth 3 (3) ; width of brain-case 6.6 (7.1).

This pretty little bat is named in honour of Miss Wilson, to whom I am indebted for much intelligent assistance.

Kerivoula muscilla Thomas, 1906

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Kerivoula muscilla, sp. n.

Size very small. Ears short, laid forward in the spirit-specimen they barely reach to the tip of the muzzle; their breadth equal to their length; inner margin strongly convex, outer with a well-marked emargination above, convex below. Tragus as in Dobson's second group, a small basal lobule present, very much as in *K. Smithii*; inner margin evenly convex, outer slightly

concave. Limbs short, less strikingly delicate than usual; upper surface of forearms and proximal part of the digits and of hind limbs and feet clothed with fine golden-brown hairs. Wings to the base of the toes. Posterior edge of interfemoral with a fringe of short curved hairs growing from its under surface. Prepuce tufted with long hairs.

Fur long, brownish grey above and below, so far as can be made out on a spirit-specimen.

Inner upper incisors slender, practically unicuspid, a rudiment of a secondary cusp present about halfway up the hinder aspect; outer incisors long, nearly as long as the inner ones, each with a minute external basal secondary cusp. Two small upper and three lower premolars subequal *inter se*.

Dimensions of the type (measured on the spirit-specimen): -

Forearm 27 mm.

Head and body 37; tail 33; head 14; ear 10; tragus on inner edge 6; third finger, metacarpus 26.5, first phalanx 13.5, second phalanx 15; lower leg and hind foot (c. u.) 17.5; calcar 16.

Hab. Ja River, Southern Cameroons.

Type. Adult male. Collected 22nd December, 1905, by Mr. G. L. Bates. One specimen.

This little *Kerivoula* is readily distinguishable from any African species hitherto described by its small size, the presence of an interfemoral fringe, and its long outer incisors. Dobson's *K. africana* agrees with it in some respects, but is said to have a tragus as in *K. hardwickei* and ears "longer than the head".

* By this expression Dobson appears always to have meant that the ears when laid forward extended beyond the tip of the muzzle.

Kerivoula nidicola zuluensis Roberts, 1924

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Kerivoula nidicola zuluensis subsp. nov.

In dimensions almost identical with a specimen of *K. nidicola* from Boror, and therefore larger than *K. lanasa*, but in colour much darker, the underparts of body being smoky greyish white, instead of buffy white, the upper parts darker, buffy brown instead of golden buffy as in *nidicola*. In *K. lanasa* the subterminal coloration of the hair is of a reddish brown, merging into the dark basal colour, whereas in *nidicola* the dark base of the hair is well defined from the subterminal golden buffy, which occupies the greater part of the hair; in *zuluensis* the dark base is even more pronounced, owing to the presence of a buffy line immediately above it, this becoming darker towards the tip of the hair, which becomes brown and produces a general dark effect. The hair of the back is long, as in *lanasa*, about two millimetres longer than in *nidicola*. White tips to the hairs of the back are present as in the other species.

Type: Adult male, T.M. No.3025, taken from the nest of a weaver bird, together with two companions, on the White Umfolosi River, Zululand, on July 17th, 1922.

Kerivoula phalaena Thomas, 1912

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A small pale brown species with subequal incisors.

Size as in the smallest members of the genus. Fur long soft, and fine; hairs of back about 7 mm. in length. General colour above uniform pale reddish brown - like "Mars-brown" of Ridgway, but much paler; the hairs of this color all through, except that on the posterior back they have inconspicuously darker bases. Extreme tips of some of the rump-hairs silvery buff. Under surface similar but rather paler, and with more blackish at the bases of the hairs. Membranes and wings almost naked, the base and edge of the interfemoral with thinly scattered hairs, not forming a fringe, the upper surface of the legs thinly hairy, the feet well haired.

Ears with inner margin strongly convex, a distinct concavity below the tip. Tragus slender, straight, a well-marked projection at its outer base, succeeded above by an emargination, above which there is again a projecting point, forming the broadest part of the tragus; in front of the middle of the base there is a wart clothed with long hairs, forming a loose tuft; a particularly prominent tragoid projection present facing the tragus on the inner side of the outer base of the ear.

Skull very light and delicate, with narrow brain-case. Upper incisors subequal in length, the outer rather shorter, and practically unicuspid, a small secondary cusp at the extreme posterior base of the inner one and at the internal base of the outer. First and second lower incisors tricuspid, third with a single large rounded cusp with minute anterior and posterior secondary cusps.

Dimensions of the type (the starred measurements taken in the flesh): -

Forearm 29.5 mm. (28 mm. in the male).

Head and body *33; tail *40; ear *13; third finger metacarpus 29.5, first phalans 12.5; lower leg and foot 18.8.

Skull: greatest length 12.1; basi-sinual length 9.1; zygomatic breadth 7.1; breadth of brain-case 6.1; front of upper canine to back of m^3 5.1

Hab. Bibianaha, inland of Denkwa, Gold Coast. Alt. 720'.

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Type. Adult female. B.M. no. 12.6.20.3. Original number 224. Collected 24th April, 1912, and presented by Dr. H. G. F. Spurrell. Male and female skins, female and young in spirit examined.

This delicate little *Kerivoula* belongs to Dobson's second group of the genus, and would seem to be allied to *K. lanosa* and *smithii*, but is markedly smaller than either.

Perhaps its nearest relative is the Kamerun *K. muscilla*, Thos., which is, however, distinguishable by its more inflated brain-case and its interfemoral fringe.

[*Læphotis*](#) Thomas, 1901

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Most nearly allied to *Vespertilio*, but the ear and tragus enlarged as in *Histiotus*.

Skull, as compared with that of the allied form, long and narrow, flattened above, very smooth and little ridged, the crests scarcely perceptible. Palate narrow, its posterior part unusually produced backward. Bullæ rather large.

Dental formula as in *Vespertilio*.

Upper incisors close to canines, instead of being well separated from them, the tip of the lower canine biting on to the top of, or outside, the outer incisor, instead of between it and the upper canine. In correlation with this the lower canine is unusually short and feeble, its length from cingulum to tip not exceeding the outer horizontal length of m^1 . Detailed proportions of teeth as described below.

Type and only species *Læphotis Wintoni*

[*Laephotis angolensis*](#) Monard, 1935

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Voir A. M. N. H. 7, VII, 1901, p. 460.

Le genre *Laephotis*, créé par THOMAS sur une espèce du British East Africa, est voisin d'*Eptesicus* et en diffère surtout par les oreilles très grandes, plus longues que la tête et par quelques détails de dentition: les incisives supérieures externes placées tout près de la canine, la canine inférieure venant buter contre elles au lieu de se placer dans l'intervalle qui les sépare de la canine. Le crâne est allongé et étroit, aplati, lisse, les crêtes à peine perceptibles. Le palais est étroit, sa partie postérieure remar-

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quablement prolongée en arrière, L'espèce type, *L. wintoni*, mesure 37^{mm} d'avant-bras.

Du Tyihumbwé, nous avons rapporté une Chanve-souris qui nous paraît appartenir à ce genre. Elle possède la dentition d'*Eptesicus* mais se distingue à première vue par la grandeur des oreilles, plus longues que la tête. Les détails de la dentition sont ceux de *Laephotis*, les incisives supérieures, les canines correspondent à ce qui a été rappelé plus haut. Malheureusement le crâne a été cassé, et je n'ai pu en vérifier l'aspect et la grandeur des bulles auditives. Les détails de forme et de coloration correspondent en partie, si bien que nous pensons avoir à faire à une forme voisine; de taille un peu plus faible.

Voici du reste la description détaillée.

Pelage de couleur foncée dessus, la base des poils noirâtre, la pointe brune; poils de 6^{mm} de longueur. A la face ventrale, la couleur est plus claire, la base des poils toujours foncée mais la pointe blanchâtre. Région pubienne blanche. Les membranes sont presque nues en dessus, sauf dans le voisinage immédiat du corps. En dessous, il y a des poils blancs vers le corps et le long de l'avant-bras. Dans la partie interfémorale, il y a quelques poils dessus et dessous, surtout dans le voisinage de la queue.

Oreilles grandes, triangulaires en forme générale, les deux bords supérieurs, en position normale, placés sur une même ligne horizontale. Marge antérieure munie d'un repli bien développé, long de 5^{mm} et formant un angle droit avec la marge, qui est un peu convexe. Pointe de l'oreille arrondie. Bord externe replié, le pli rectiligne, donnant la forme triangulaire générale; marge à peu près droite. Antitragus en demi-cercle, séparé de la marge externe par une profonde incision. Tragus grand, à marge interne bien concave, à sommet arrondi, ayant son maximum de largeur un peu au-dessus du point de la base interne. Sa

marge externe commence par un petit lobe arrondi en cornet, séparé de la partie supérieure par une échancrure arrondie, l'un des oreillons présente en outre une petite échancrure triangulaire au dessus du lobe basal; mutilation? La longueur de l'oreille, comptée de l'angle antérieur

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de l'antitragus est de 18^{mm}; la largeur est de 12^{mm} (mesurée sur l'oreille aplatie); le tragus mesure 5.5^{mm} sur sa marge interne.

Ailes normales; métacarpes III à V égaux; dernière phalange du troisième doigt terminée en T. La membrane s'insère à la base du cinquième orteil. Eperon long de 15^{mm} environ; un lobe éperonnier arrondi bien développé et, en plus, à l'extrémité de l'éperon un très petit lobe supplémentaire. Extrême pointe de la queue dépassant la membrane. Celle-ci d'une couleur générale brune, mais une très étroite bordure blanche du cinquième doigt à la cheville.

Le crâne cassé ne permet pas une description générale. L'échancrure palatale nous paraît normale et non étroite, comme dans *wintoni*; elle atteint en arrière le niveau postérieur de la canine; le palais est allongé et profondément concave. Les incisives médianes sont convergentes et munies d'une petite pointe supplémentaire près de leur extrémité; le cingulum forme, derrière la dent, un petit tubercule. Les incisives externes sont placées très proches des canines, qu'elles ne touchent cependant pas; les canines inférieures viennent buter contre une concavité de celles-là. La canine supérieure est bien développée; la première prémolaire, grande, touche la canine; les deux premières molaires sont normales, avec les tubercules bien développés; la dernière branche de W de la dernière molaire manque et la crête se termine par un tubercule. Dents mandibulaires normales; la dernière molaire comptant un petit tubercule postérieur, donc six au total.

Dimensions: avant-bras 35^{mm}; métacarpes 34; doigt III 62; tibia 14; pied 6; queue 38; tête et corps 50.

Cette forme diffère de *wintoni* par une taille plus petite, par les oreilles moins développées et d'une forme un peu différente.

Localité: Tyihumbwé, septembre 1932; une femelle

Laephotis botswanae Setzer, 1971

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Holotype: Adult female, skin and skull, U.S. National Museum no. 425349, from 50 mi. W, 12 mi. S Shakawe, Botswana. Obtained 24 May 1967 by S. W. Goussard, original no. 691.

Referred Specimens: British Museum (Natural History) 55.1134 and BM(NH) 55.1135 from Solwezi Boma, Zambia, obtained 28 April 1952 by W. H. F. Ansell; BM(NH) 57.436 and BM(NH) 57.438 from Mumene, 40 km. E Elizabethville, Congo (Kinshasa), obtained 28 December 1955 by the Hygiene Service of the Congo.

Measurements: The measurements of the holotype are: Total length 96; length of tail 41; length of hind foot 8; length of ear from notch 21; length of forearm (dry) 37.3; greatest length of skull 14.5; greatest breadth across zygomatic arches 8.3; least postorbital breadth 3.4; greatest breadth of braincase 7.0; depth of braincase 4.7; length of maxillary tooththrow 4.7; postpalatal length 5.2; breadth of palate across M²-M² 5.4; breadth of palate across C-C 4.4.

Diagnosis: Upper parts near Buffy Brown; flight membranes near Clove Brown; underparts somewhat paler than dorsum, individual hairs black at base (5 mm), a narrow band (1 mm) of pure color and tipped (2 to 3 mm) with buffy. Ears moderate in size; tragus and antitragus moderately developed. Skull average in size for the genus; relatively narrow; rostrum relatively long and narrow; zygomatic arches fragile;

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[table removed - eds.]

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tooththrows diverging slightly posteriorly; palate markedly arched dorsally; auditory bullae moderately sized.

Comparisons: From *Laephotis wintoni* as known by a specimen in the Royal Ontario Museum (ROM 36368) from Namanga, Kenya, *L. botswanae* differs in lighter color throughout; smaller ears; somewhat shorter forearm; and smaller, less curved tragus. The skull is smaller in all respects; auditory bullae smaller; palate more concave; upper tooththrows more divergent posteriorly; and bony palate less projecting posteriorly.

Laephotis botswanae differs from the holotype and paratype of *L. namibensis* in smaller size; darker color; markedly smaller ears and tragus; and in having a shorter forearm. The skull is markedly smaller with a broader, shorter rostrum; occipital region more rounded; palatal region shorter, broader and more concave; auditory bullae less inflated; and foramen magnum markedly smaller.

Compared with *Laephotis angolensis* as known by a specimen from 35 mi. E Dande, Angola (AMNH 87244), *L. botswanae* differs in somewhat paler color; larger ears; and longer forearm. The skull is markedly larger; rostrum more massive; occipital crest less projecting posteriorly; palate broader but not as deeply concave; upper tooththrows more divergent posteriorly; and auditory bullae, relatively as well as actually, smaller.

Remarks: With the recognition of these two new taxa, four species are now known in the genus *Laephotis*. There is reasonably close relationship between *L. wintoni* and *L. botswanae* but the relationship of *L. angolensis* and *L. namibensis* with the others is somewhat obscure.

When the skulls are arranged according to their greatest length, *namibensis* is largest, followed by *wintoni*, *botswanae*, and finally *angolensis*. The same applies when greatest breadth of braincase is examined; *wintoni* is the largest, *namibensis* next, followed by *botswanae* and *angolensis* (Table 1).

Forearm measurements are rather distinctive for *L. namibensis* and *L. angolensis*, the measurement of the former being over 38 while the measurement of the latter is not known to exceed 33. The same measurement overlaps between *L. botswanae* and *L. wintoni*. If, however, one adds to the preceding differences, the size and configuration of the tragus (Fig. 1), *L. wintoni* and *L. namibensis* show the closest relationship while *L. angolensis* and *L. botswanae*, which might possibly occur together, show striking differences in the tragi.

If one assumes *L. namibensis* to be the most specialized in shape and size of the ears; shape and size of the tragus; size of the auditory bullae; and in the shape and size of the rostrum; one is tempted to group the other taxa on the same morphological bases. When this is done it seems reasonable to place *L. wintoni* and *L. namibensis* as being closely allied, even though they are separated from each other by some 24° of latitude and some 30° of longitude. The other two species, occupying this hiatus of latitude and longitude can be readily separated on the basis of length of tragus, length of forearm and greatest length of skull. It would seem

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that an arrangement from least to most specialized based on several morphological features *L. angolensis* might be the least specialized; *L. botswanae* next with *L. wintoni* and *L. namibensis* in that order.

Peterson (MS, 1971) suggests that the populations of *Laephotis* represented by specimens from Botswana, Zambia and the southeastern Congo (Kinshasa) might represent a new species if female specimens of true *L. wintoni* were known. I suggest that since the holotype of *L. angolensis* is a female and that the measurements of a single male of the same taxon from 35 km. E Dande, Angola (AMNH 87244) show no real differences, there is no significant sexual variation involved. Examination of the external and cranial measurements of the series of specimens in the British Museum from Mumene, Musonge, Congo, as presented by Peterson, reveals that sexual variation is slight but that females do average a bit larger than males.

Laephotis namibensis Setzer, 1971

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Holotype: Adult female, skin and skull, U.S. National Museum no. 342152, from Namib Research Station, Gobabeb, South West Africa; obtained 22 November 1963 by Ronald E. Cole, original no. 270.

Specimens examined: Two from the type locality.

Measurements: The measurements of the holotype with those of a female paratype (USNM 342153) in parentheses are: Total length 106 (104); length of tail 47 (46); length of hind foot 8 (8); length of ear 25 (24); length of forearm (dry) 38.2 (38.6); greatest length of skull 16.5 (16.5); greatest breadth across zygomatic arches 9.0 (-); least postorbital breadth 3.2 (3.6); greatest breadth of braincase 7.5 (7.6); depth of braincase 4.7 (4.9); length of maxillary tooththrow 4.9 (5.0); postpalatal length 5.9 (6.0); breadth of palate across M²-M² 5.2 (5.4); breadth of palate across C-C 4.0 (4.0).

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Diagnosis: Upper parts Light-Drab; flight membranes near Olive Brown; underparts noticeably paler than dorsum, individual hairs black at base, a narrow band (1-2 mm) of pure color (near Light-Drab), and tipped (2-3 mm) with white. Ears exceptionally large; tragus and antitragus well developed. Skull large for the genus; relatively long and narrow; upper tooththrows nearly parallel; palate relatively long and narrow; auditory bullae remarkably inflated ventrally.

Comparisons: Compared with the holotype of *Laephotis wintoni*, *L. namibensis* is markedly paler in color; markedly larger ears, especially across the base (10.9 dry); and markedly larger tragus. The skull is longer; the postorbital constriction is less; the braincase is somewhat more domed; the auditory bullae are more inflated; the zygomatic arches are more markedly bowed dorsad; and the maxillary tooththrows are more nearly parallel.

From a specimen of *Laephotis angolensis* from 35 km. E Dande, Angola, in the American Museum of Natural History (AMNH 87244), *L. namibensis* differs in markedly larger size; paler color; and strikingly larger ears. The skull, in all respects is decidedly larger and more robust.

Remarks: These specimens were taken in a mist net over a water hole in the bed of the Kuiseb River near the Namib Desert Research Station.

The name proposed for this new taxon is in reference to the Namib Desert of South West Africa.

Læphotis Wintoni Thomas, 1901

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Size rather less than in *Histiotus velatus*; general appearance very much as in that species, although the ears are not so large. Fur close and fine, the hairs of the back about 6 - 7 millim. in length. General colour above coppery brown, the basal halves of the hairs sooty, the terminal halves clear

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coppery or cinnamon (Dr. Hinde speaks of the colour as "bronze"). Under surface paler brown, the tips of the hairs becoming almost white in the pubic region.

Fur above not extending on to arms or wing-membranes, though there are a few fine hairs on the thumbs; but the legs are clothed as far as the knee, the backs of the feet are finely hairy, and the basal half of the interfemoral is thinly covered with fine and inconspicuous hairs. Below the wing-membranes are finely clothed nearly as far outwards as a line from the elbow to the knee, but the interfemoral membrane is more nearly naked than above. No trace of an interfemoral fringe.

Ears large, much larger than in *Vespertilio*, but not so large as in *Plecotus*, *Otonycteris*, or *Histiotus*; not connected across the forehead with each other. Inner margin with a distinct basal fold, ending below in a narrow lobe; its basal third is strongly convex forward, so that the basal line is at right angles to the upper two thirds, which are very slightly convex and pass gradually into the rounded tip; outer margin nearly straight above, slightly convex below; antitragus low, half-oval, marked by a distinct notch behind. Tragus unfortunately damaged in the type on both sides, but enough is left to show that it is long, broad at base, with a slightly concave inner margin and a broadly rounded tip.

Wing of normal Vespertilionine proportions, the metacarpals of the third, fourth, and fifth digits approximately equal in length. Insertion of wing-membrane at base of the digits, so far as can be made out in the dry skin. Calcar reaching about halfway towards the tip of the tail; postcalcaneal lobule present, but very narrow. Tip of fourth finger T-shaped, or even slightly spatulate. Membranes and ears uniformly brownish grey, except that the wing-membrane from the tip of the fifth finger to the ankle is edged with whitish.

Skull very thin and papery, long and narrow, low, smooth, and scarcely ridged at all. Nasal notch comparatively shallow. Intertemporal region not strongly constricted, the intertemporal but little less than the interorbital breadth. Palatal notch penetrating to the level of the middle of the canines. Palate narrow, the greatest breadth across the outer corners of m^2 going nearly three times in the total length of the skull; its posterior part produced behind the molars a distance equal to the combined lengths of p^4 and m^1 .

Upper incisors slightly convergent, of about the same relative proportions as in the Serotine, the inner one long, with a secondary postero-external cusp near its tip and a small

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postero-internal cingular cusp at its base; outer incisor quite small, its main cusp about half the height of the inner incisor, also with a small internal basal cusp. Large premolar pressed close up against the back of the canine; last upper molar triangular, with a well-developed posterior lobe, in correspondence with which the talon of the last lower molar is large and tricuspid. Lower incisors trifid, overlapping; anterior lower premolar about one half the height and one fourth the area in cross-section of the posterior one.

Dimensions of the type (measured in skin): -

Forearm 37 millim.

Head and body (approximate) 50; ear (dry and no doubt contracted) 21; tragus, length on inner edge 6.3, breadth basally 3.5; metacarpus of third finger 35; first phalanx 13; tibia 13.5; hind foot, s. u. 7, c. u. 7.7; calcar (c.) 15.

Skull: greatest length 15.7; basipalatal length in middle line 12.8; interorbital breadth 5; intertemporal breadth 3.6; breadth of brain-case 7.8; palate length 6.6. Front of lower canine to back of m_3 5.6.

Nab. Kitui, British East Africa. Altitude about 3500 feet.

Type. Male. Original number 64. Collected 7th January, 1901, by Dr. S. L. Hinde.

I have ventured to name this interesting bat in honour of my friend Mr. W. E. de Winton, to whose labours on the small mammals of Africa all naturalists, and I especially, have been so constantly indebted for assistance.

Lavia frons affinis K. Andersen and Wroughton, 1907

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Diagnosis. - Average size smaller.

Details. - Comparative measurements are given above under *L. frons frons* (p. 139).

Type. - M ad. skin. Kaka, White Nile. Collected by R. M. Hawker, Esq. B.M. no. 1.8.8.3.

Specimens examined. - 8 (7 skins) and 6 skulls, viz, White Nile (5), Lake No (1), Lado (1), Wadelai (1).

The British Museum possesses a somewhat damaged skull without skin from Cape Coast Castle (Gold Coast) which seems to belong to this race, and Miller's specimen of "*L. frons*". referred to above under *L. f. frons* (p. 139) appears also to belong to the present race; so it is possible that the range of *affinis* extends westward to the coast.

[*Lavia rex*](#) Miller, 1905

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1892. *Megaderma frons* True, Proc. U.S. Nat. Mus., XV, p. 469, October 26, 1892 (part).

Type from Taveta, German East Africa. No. ¹⁸⁹⁹³/₃₈₁₉₇, United States National Museum. M adult (in alcohol). 1889. Dr. W. L. Abbott.

Characters. - Like the West African *Lavia frons* (Geoffroy) but considerably larger (forearm 60 instead of 56, mandible 17.8 instead of 15.2), and with disproportionately heavier teeth.

Color. - (Skin of topotype, No. 18,992, not sexed): Fur everywhere drab-gray (that of belly a little darker than that of back) tipped with ochraceous-buff. On middle of back and neck and on posterior half of belly the ochraceous-buff is so inconspicuous that it scarcely modifies the ground color, but on face, sides of neck, entire chest and throat, and along border of interfemoral membrane it strongly predominates. At shoulder the wood-brown fades to buffy white, forming an inconspicuous light shoulder-spot. The type does not appear to differ appreciably in color from the dry specimen, though it has been subject to the action of alcohol for more than fifteen years.

Ears, membranes, etc., - Probably not different from those of *Lavia frons*. Dr. F. W. True has already noted the peculiar broadened, serrated form

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of the secondary lobe of the tragus in the type specimen as compared with that figured by Dobson and represented by a Sierra Leone specimen in the United States National Museum. In the skin from Taveta this lobe has a form intermediate between the others and closely resembling that of *Lavia frons* as figured by Geoffroy. The variation is probably individual.

Skull and Teeth. - As compared with those of an adult male *Lavia frons* from Sierra Leone (No. 38,196, United States National Museum) the skull and teeth of *Lavia rex* are readily distinguishable by their uniformly greater size and massiveness. In actual form of either skull or teeth there are no striking differences between the two species; but the larger animal has the auditory bullæ relatively larger and the interpterygoid space narrower, while the teeth, particularly the upper canines and upper molars, are very considerably increased in size.

Measurements. - Type: Head and body, 70 (69);** tibia, 34 (29); foot, 16 (15); forearm 60 (56); thumb 14 (11); second finger, 62 (55); third finger, 110 (105); fourth finger, 78 (72); fifth finger, 83 (75); ear from meatus, 43.6 (40); ear from crown, 39 (33); width of ear, 28 (26); tragus, 29 (25); noseleaf, 22 (22); greatest width of noseleaf (flattened), 16 (16). Skull: Greatest length, - (23); basal length, - (18); basilar length, - (16), median palatal length, 6.6 (4.8); greatest palatal width including molars, 9.2 (8.2); distance between tips of upper canines, 5.4 (?); mandible, 17.8, (15.2); maxillary tooththrow, 9.2 (8.2); mandibular tooththrow, 11 (9).

Footnote:

** Measurements in parentheses are those of an adult male of *Lavia frons* from Sierra Leone (No. ¹¹⁵³⁹/₃₈₁₉₆, United States National Museum).

[*Leiponyx*](#) Jentink, 1881

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Muzzle as in *Pteropus*. Nostrils slightly projecting, rather deeply emarginate between. Upper lip with a distinct vertical groove in front.

Index finger without a claw. Metacarpal bone of the middle finger smaller than the index finger. Wings from the sides of the back and from the base of the first toe. Tail short, for its greater part free from the membrane.

Dentition. Inc. ²⁻²/₄, c. ¹⁻¹/₁₋₁, p.m. ²⁻²/₂₋₂, m. ²⁻²/₃₋₃ = 32.

Leiponyx büttikoferi Jentink, 1881

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Ear longer than the muzzle (the distance between the tip of the nose to the foremost corner of the eye). Earconch with a rounded tip, the outer margin in its upper part concave, convex further on, inner margin slightly curved. Shoulder-glands absent in the female, the single specimen we possess.

The palate-ridges are seven in number; the first four are undivided; the first is nearly straight and placed between the canines, the second is slightly curved and placed immediately behind the first premolars, the third ridge is broader than the second and unites the second premolars; the fourth again is more arched and unites the space between the two molars on each side. The three remaining ridges form oval elevations on each side resembling those in *Epomophorus comptus*. They are succeeded by ill-defined semicircular toothed lines similar to the same parts in *Epomophorus franqueti*.

The four unicuspidate upper incisors are placed at a distance from the canines, in pairs, with a rather wide space between those pairs. The foremost incisors are the smallest. The lower incisors are also unicuspidate and nearly equal in

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size and placed in a semicircular row between the smooth canines. The canines in the upper and lower jaw are followed at a small distance by the little developed first premolars, which are separated from the much stouter second premolars by a rather large interval. The third premolar in the lower jaw is separated by a space nearly as large as the interval between first and second premolar. This is the most developed premolar, not in vertical extent but in circumference. Molars very small, slightly raised above the level of the gum, especially the lower molars.

Fur short, yellowish brown, darker on the head. Ears and wings blackish brown. Claws black, strongly arched and acutely pointed.

Measures of the single specimen. (mm)

Head and body: 195.

Tail: 13.

Tail free from membrane: 7.5

Ear: 25.

Thumb with claw: 46.

Forearm: 125.

Index finger: 85.

Third finger, metacarp: 82.

" " 1st phalanx: 50.

" " 2nd phalanx: 89.

Fourth finger, metacarp: 82.

" " 1st phalanx: 43.

" " 2nd phalanx: 52.

Fifth finger, metacarp: 78.

" " 1st phalanx: 33.

" " 2nd phalanx: 33.

Tibia: 52.

Foot with claw: 35.

Hab. Liberia, St. Paul's River (Millsburg).

Liponycteris Thomas, 1922

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WHEN, in 1915, I wrote my "Notes on *Taphozous* and *Saccolaimus*", and recognized the latter as a distinct genus from the former, as had Hollister previously under another name, I did not sufficiently weigh the characters which separate *Taphozous nudiventris* and its allies from the typical members of *Taphozous*.

On reconsidering the subject, I now think that these remarkable half-naked bats should be separated as a distinct genus from the ordinary hairy species, and would suggest that the whole group might be synoptically arranged as follows: -

A. Bullæ imperfect, the inner side of each incomplete. A radio-metacarpal pouch.

a. Frontal concave; upper profile of brain-case rising from it in a strong curve. Occipital "helmet" scarcely developed. Body haired as usual..... I. *Taphozous*, Geoff.
Genotype: *T. perforatus*, Geoff.

b. Frontal almost flat, the cranial profile scarcely rising above it behind. A strongly developed occipital helmet.

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Body partly naked behind, both above and below II. *Liponycteris*, gen. nov.

Genotype: *L. nudiventris* (*T. nudiventris*, Cr.).

B. Bullæ perfect internally. No radio-metacarpal pouch. Body hairy..... III. *Saccolaimus*, Less.

Genotype: *S. saccolaimus* (*T. saccolaimus*, Temm.).

Besides *nudiventris*, *Liponycteris* would contain only *kachhensis* and its two subspecies - *magnus*, Wettstein (1914, syn. *babylonicus*, Thos., 1915), and *nudaster*, Thos.

Lissonycteris angolensis goliath Bergmans, 1997

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Rousettus (Lissonycteris) angolensis; Harrison, 1960: 65, 1961: 287; Meester et al., 1986: 30.

Rousettus angolensis; Fenton, 1975; Hutton, 1986: 227.

Rousettus angolensis angolensis; Smithers et al., 1976: 42, 1983: 65 (in part: records from Zimbabwe and Mozambique).

Rousettus (Lissonycteris) angolensis angolensis; Hayman et al., 1971: 12 (in part: records from Rhodesia); Smithers et al., 1979: 27 (in part: not the record from the Haroni/Lusitu River confluence); Koopman, 1994: 20 (in part: the record from Zimbabwe).

Material examined

ZIMBABWE. Holotype specimen: 1 F, skin, skull, 19-III-1970, Gleneagles, Inyanga, collected by M. Stuart Irwin and party (NHMBZ 59831; field number T1854); paratype specimens: 4 MM, 1 imm. M, 2 FF, 1 imm. F, skins, skulls, same data as für holotype specimen (HBM 16.7321, field number GNA/MP 1; NHMBZ 59826/29, -32/33 & ZMA 24.719, field numbers T1848/50, -52/53, -55/56).

Other material: Umtali: 1 M, skin, skull, 18-VI-1960, R. H. Smithers (HBM 15.3219).

Referred material (not examined)

MOZAMBIQUE.

(Western boundary of Vila Pery District, 1832Bd, 1832Dc, 1833Cc.)

ZIMBABWE.

(Birchenough Bridge at Sabi River.)

Diagnosis: As for the species, but with large body, skull, and teeth dimensions, with fal ranges of 81.9-87.5 in 4 MM and 82.1-89.6 in 3 FF and gsl ranges of ≥ 43.3 -46.0 in 4 MM and ≥ 42.2 -45.8 in 3 FF, and (antero-posteriorly) lang cheek teeth with rather distinct heels anterior and posterior to the outer cusps.

Measurements: Table 3. Same ratios are:

rl MM 37.2 -38.0 % of gsl (n = 2),
 FF 38.2 % of gsl (n = 1);
 C¹-C¹ MM 17.4 -20.4 % of gsl (n = 2),
 FF 19.0 % of gsl (n = 1);
 M²-M² MM 29.1 -30.1 % of gsl (n = 2),
 FF 29.4 % of gsl (n = 1).

Distribution: Fig. 5.

Related species: See the account of the typical subspecies.

Remarks

Taxonomy: The seven known localities are all in or near the border area of Zimbabwe and Mozambique, between 18 and 20° S and at a distance of about 1000 km from the nearest known populations of *L. a. ruwenzorii*. Ansell (1967, 1974) recorded representatives of the species, here included in *ruwenzorii*, from extreme northwest Zambia and, in 1978 the same author remarked that, apart from northeast Zambia it was perhaps unlikely to occur elsewhere in the

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[table removed - eds.]

country. Other *ruwenzorii* specimens are known from southeast Zaïre (fig. 5). With fal ranges of 72.3-84.4 in MM and 72.8-83.0 in FF and gsl ranges of 39.3-43.8 in MM and 39.1-43.2 in FF, *ruwenzorii* averages very distinctly smaller than *goliath*. No fruit bats have been recorded from the northeast zone of Zimbabwe (Bergmans, 1988, fig. 1), and the new subspecies' characteristics strongly suggest its prolonged isolation from the Central and East African populations of *angolensis*. Apart from

its larger size, its premolars and molars, especially P^3 , P^4 , M^1 and P_4 , M_1 and M_2 , are relatively longer, with in lateral view more distinct heels anterior and posterior to the outer cusps, which themselves are relatively obtuse. The difference in size between M^1 and M^2 is generally smaller than in the smaller subspecies *smithii* and *petraea*. As a consequence of the longer cheek teeth, the interstices between them are relatively short.

Harrison (1960), who described the first specimen of *Lissonycteris angolensis* from this region, already remarked on its exceptional size when compared to specimens from Kenya, and re-

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frained from describing a new subspecies only because of the limited material. Smithers et al. (1979) gave same rounded measurements of specimens from Zimbabwe but did not comment on them.

The palatal ridge pattern has been preserved only in the specimen from Umtali and is typically 3 + 4 + 2.

Distribution and geographical variation: Harrison recorded a specimen from Umtali (= Mutare; square code 1832Dc, or 1832D3), Zimbabwe. Smithers et al. (1976) recorded specimens from along the western boundary of Vila Pery District in Mozambique. They did not mention specific localities (nor where their specimens have been deposited), but on their map the squares 1832Bd, 1832Dc, 1833Cc and 2032Bb have been marked. (From the localities mentioned in their gazetteer, only Machipanda corresponds with 1832Dc and Chemezi with 1833Cc.) Smithers et al. (1979) added Birchenough Bridge on Sabi River (1932Cd) and Gleneagles (1832Bd), both in Zimbabwe, as collecting localities for *Lissonycteris angolensis*. They also mentioned material from the Haroni/Lusitu River, but this actually represents *Myonycteris relicta* Bergmans, 1980 (see present paper).

The distribution appears to be confined to the escarpment region in the border area of Zimbabwe and Mozambique, between 17 and 21 S, with known occurrences from between 200 and 500 m (Birchenough Bridge) to 1800 m (Gleneagles). Smithers et al. (1979) reported that the specimen from Umtali had been taken in an old, established suburban garden with trees and shrubs, where it was feeding on guavas among many *Epomophorus wahlbergi* (Sundevall, 1846), but that two of the other localities were associated with evergreen forest (one of these probably the Haroni/Lusitu River confluence from where the species was incorrectly recorded). According to White (1983) the localities are all associated with forest mosaics, forest transitions or woodland: 2 are in Undifferentiated Afromontane vegetation (type 19a), 1 is in Drier Zambezian miombo woodland (dominated by *Brachystegia* and *Julbernardia*) (type 26), 1 in *Colophospermum* mopane woodland and scrub woodland (type 28), and 3 in either type 19a, or 16, or 26, in which 16(a) is East African Zanzibar-Inhambane coastal mosaic. Its restricted distribution would suggest that geographical variation within this subspecies is unlikely.

Etymology: The specific epithet *goliath* has been chosen in honour of Dr. David L. Harrison, who would have 'mastered' this Goliath long ago if he had had more specimens. Of course, it also refers to the subspecies' exceptionally large dimensions.

Lissonycteris angolensis petraea Bergmans, 1997

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Rousettus aegyptiacus aegyptiacus (not of E. Geoffroy-St. Hilaire, 1810); Dorst et al., 1972: 394 (in part: the specimens from 10 km from Agaro and from Ghimbi).

Rousettus aegyptiacus (not of E. Geoffroy-St. Hilaire, 1810); Largen et al., 1974: 228 (in part: the records from 10 km from Agaro and from Ghimbi).

Rousettus angolensis; Largen et al., 1974: 229, 255.

Rousettus angolensis ruwenzorii; Largen et al., 1974: 230.

Material examined

ETHIOPIA. Holotype specimen: 1 M, alc., skull, 10/13-VI-1968, 10 km from Agaro, at the road to Jimma, J. Dorst and party (MNHN CG 1972-482; field number 4017); paratype specimens: 5 MM, 2 imm. MM, 1 F, 2 imm. FF, alc., skulls, same data as holotype specimen (MNHN CG 1972-482, field numbers 4018/19, -24, -27, -32, -44/46, -51 and ZMA 25.180, field number 4050).

Other material: Didessa River: 1 F, 2 MM, alc. (BMNH 71.2446/47, 72.638). Ghimbi: 1 imm. M, 2 imm. FF, alc., 27/30-IX-1971, J. Dorst and party (MNHN CG 1984-2074/76). Nur Moha-med Cave: alc. material (BMNH).

Referred material (not examined)

ETHIOPIA. Didessa: 2 specimens, alc., 1-1972, J. S. Ash (USNM 497488/89); 1 specimen, 25-1-1973, J. S. Ash (USNM 497490).

(Doki River Bridge.)

Diagnosis: As for the species, but with small average body and skull dimensions, with a fal range in 8 MM' of 72.7-76.3 (mean 74.2) and in 2 FF 71.7-73.5, and a gsl range in 6 MM of 38.5-41.3 (mean 39.7), and (antero-posteriorly) short and low canines and cheek teeth, the latter with relatively long outer ridges.

Compared with *L. a. angolensis* and *L. a. ruwenzorii*, smaller average body and skull dimensions and (antero-posteriorly) relatively shorter upper canines and shorter and lower upper cheek teeth with longer and more diverging outer cusps, or ridges, and P⁴ and M¹ more squarish; larger lower cheek teeth equally low and also with relatively long outer cusps. Compared with *L. a. smithii*, slightly larger in average body and skull dimensions, upper incisors narrower and not sharply curved backward, upper and lower cheek teeth with relatively long outer cusps, P⁴ with a well-developed inner cusp.

Measurements: Table 2. Some ratios are:

rl MM 37.1 - 38.3 % of gsl (n = 6);

C¹-C¹ MM 17.9 - 18.7 % of gsl (n = 5);

M²-M² MM 27.3 - 30.5 % of gsl (n = 5).

Distribution: Fig. 5.

Related species: See the account of the typical subspecies.

Remarks

Taxonomy: The Ethiopian populations of *L. angolensis* are geographically nearest to those in South Sudan, at a distance of about 600 km, identified as *L. a. ruwenzorii* and including the largest specimens of that subspecies: the fal range in 8 MM is 76.5-84.4 and in 8 FF 76.5-80.9; the gsl is c. 41.2 in 1 M and 40.1-41.3 in 3 FF.

The next nearest subspecies is *L. a. angolensis*, possibly represented at about 1500 km to the West, in the northern Central African Republic (fal of 1 M 74.4, of 1 F 75.1; Schlitter et al., 1982), and still further west in Cameroun where specimens average larger (fal range in 4 MM 73.4-80.0 and in 12 FF 74.5-83.0; gsl in 1 M 42.1 and in 4 FF 38.7-40.5) and, like *ruwenzorii*, differ in teeth morphology. In overall size *L. a. petraea* comes nearest to *L. a. smithii*, but it is separated from this by *L. a. angolensis* and differs in teeth morphology. The cheek teeth in the new subspecies show specific trends in their development. They are relatively weak by being shorter and lower, and at the same time retain long outer ridges (or cusps) instead of the short, more pointed cusps in the other sub-

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[table removed - eds.]

species, and the inner cusp in P⁴ is less obsolete than in *smithii*. The overall weakening of the dentition may indicate that this subspecies represents an old branch of the species, while the long outer ridges, which are probably a primitive character, may have persisted as an adaptation to specific food sources.

The palatal ridge pattern in *petraea* is 3 + 4 + 2 in nine, and 4 + 3 + 2 in two (including the holotype) out of 11 specimens. The sixth ridge is much reduced in two specimens. The specimen with field number 4018 has an M³ measuring 0.9 x 0.95 on the left side, and the one with number 4024 an M⁴ of 1.0 x 0.9 on the right side.

Distribution and geographical variation: The six known localities are all in or near montane rain forest in the southern Ethiopian highlands, at both sides of the Central Rift, with altitudes between 1190 and 2600 m (only of Doki River Bridge no altitude is known). Of the six localities, three are in East African evergreen and semi-evergreen bushland and thicket, two in

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Undifferentiated Afromontane vegetation, and one in Undifferentiated Ethiopian woodland (types 38, 19a and 29b in White, 1983). The distribution area is separated from the areas of the subspecies *ruwenzorii* and *angolensis* by less favourable and probably uninhabitable vegetation types: towards the west mainly by a Transition from undifferentiated Ethiopian woodland to Acacia deciduous bushland and wooded grassland, and Edaphic grassland in the Upper Nile basin (types 35b and 61 in White, 1983), and towards the south mainly by Somalia-Masai *Acacia-Commiphora* deciduous bushland and thicket (type 42 in White, 1983).

With the exception of the type series, the samples are insufficient to establish possible geographical variation.

Four specimens were infested with typical examples of the nycteribiid fly *Dipseliopoda biannulata* (Oldroyd, 1953).

Etymology: The new subspecies has been named in honour of Dr. Peter J. H. van Bree, former Curator of Mammalogy of the Institute for Systematics and Population Biology (Zoologisch Museum) of the University of Amsterdam, under whose guidance the author laid the basis for his studies of African fruit bats. The names Peter and *petraea* are both derived from the Greek *petra*, in its meaning of rock in the metaphorical sense of a solid and trustworthy basis, but also referring to the subspecies' apparent confinement to the tormented relief of the Ethiopian plateau.

[*Macronycteris*](#) Gray, 1866

An updated description is given by Foley *et al.* (2017: 10).

Megaloglossus Pagenstecher, 1885

[Eds.: This is from Jahrbuch der Hamburgischen Wissenschaftlichen Anstalten, 1885, 2: 125 - 128. This paper has the same title as the one in Zoologischen Anzeiger, which is generally considered to be the original description]

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Megaloglossus: Schwanz mit zwei gegen das Os sacrum und unter einander beweglichen, aber gänzlich versteckten Wirbeln, der zweite verkümmert, Flughaut mit 2 Fältchen von der Basis der zweiten und der dritten Zehe.

In dem einen Merkmal vermittelt also die Gattung *Megaloglossus* zwischen *Macroglossus* und *Melonycteris*, im anderen eutfernt sie sich weiter von *Macroglossus* als *Melonycteris*. Sie nähert sich in der Anordnung der Gaumenfalten, indem die zwei letzten gespalten sind, mehr *Melonycteris*.

Es ist nicht unwarscheinlich, daß auch die neue Art allein in ihrer Gattung steht und man wird dann Gattungscharakter und Artcharakter vielleicht überhaupt nicht zu trennen in der Lage sein. Jedenfalls wird man für jetzt über das obige mit Sicherheit nicht hinaus gehen können.

Nach nachfolgender Tabelle hält die neue Art in der Größe die Mitte zwischen *Macroglossus minimus* Geoffr. und *Melonycteris melanops* Dobs.

Macroglossus minimus M incl. Schwänzchen; *Megaloglossus Woermanni* F; *Melonycteris melanops* M
Körperlänge von Nasenspitze an: 82 mm; 90 mm; 120 mm
Länge des Schädels: 26; 29; 38
Entfernung von Augenmitte bis Schnauzenspitze: 12; 25; 19.5

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Entfernung von Augenmitte bis zum unteren Winkel der Ohröffnung: 12; 11; 16.5
Länge der Zunge: 21; 28; 28
Höhe der Ohröffnung: 12; 13; 16
Vorderarm: 38; 45; 62
Daumen mit seinem Metacarpus: 15; 19; 25
Dritter Finger: 70; 80; 130
Fünfter Finger: 55; 56; 85
UnterSchenkel: 17; 20; 28
Fuß ohne Sporn: 10,5; 12; 18

Rechnet man alle genommenen Maße jeder Fledermaus zusammen und dividirt durch die Zahl der Maße, so hat *Megaloglossus* das 1.21, *Melonycteris* das 1.54 fache von *Macroglossus*. Über diesen Durchschnittsfaktor gehen bei *Megaloglossus* hinaus die Zunge mit 1.83, die Schnauze mit 1.33 und der Daumen mit 1.26, bei *Melonycteris* der dritte Finger mit 1.85, der Fuß mit 1.71, der Daumen mit 1.66, der Unterschenkel mit 1.65, der Vorderarm mit 1.63. *Megaloglossus* ist hiernach die größzüngige Macroglosse und zum Klettern gut geeignet. Der Fersensporn ist größer als bei *Macroglossus*.

Was die Zähne betrifft, so hat die neue Fledermaus die Formel der beiden anderen Gattungen:

$$2+3+1+4+1+3+2 / 3+3+1+4+3+3$$

Die oberen Schneidezähne sind fast aequidistant, in der Mitte wenig weiter von einander entfernt, die Krone der unteren ist in zwei körnerartige Lappen getheilt, der obere Eckzahn hat, wie bei den anderen Gattungen die vordere Rinne, in welcher der untere gleitet. Der erste obere Lückzahn ist vom Eckzahn deutlich getrennt, wie bei *Macroglossus*, während er bei *Melonycteris* dichter an diesen rückt. Die beiden folgenden Lückzähne sind im vorderen Theile gut entwickelt und etwas hakig gespitzt, während bei *Macroglossus* der vordere, hakige Teil früh abschleift und bei *Melonycteris* der Zahn mehr im Ganzen und plumper sich zuspitzt. So ist namentlich der zweite obere Lückzahn recht kräftig. Die Jochbogen sind weniger nach außen gedrängt als bei *Macroglossus* und setzen sich vorne etwas höher an.

Die Umrandung der Naslöcher steht im ganzen minder vor als bei *Macroglossus*. Die Naslöcher werden theilweise überdeckt von einem

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deutlichen Zipfel des oberen oder inneren Randes. Die Oberlippe ist breiter, plumper als bei *Macroglossus*, ihre Mittelkerbe eher tiefer.

Die Zunge, indem sie die gleiche Länge hat, wie die des erheblich größeren *Melonycteris*, auch ähnlich breit und dicker, viel massiger als bei *Macroglossus* ist, hat mich zu der Gattungsbenennung *Megaloglossus* veranlaßt. Sie ragt schon beim ungeborenen Jungen aus dem Munde hervor. Ihr Mittelfeld mit einem Pflaster von Platten, jede mit drei nach hinten gerichteten Zähnen, hat fast die Ausdehnung wie bei *Melonycteris*, namentlich eine größere Breite als bei *Macroglossus*. Haar dicht, weich, dunkler als bei *Macroglossus*, umbra-braun, auf der Unterseite graubraun, Flughäute dunkelbraun. Zitzen groß, an den Brustseiten, ungefähr in der Höhe des Ellenbogens.

Nach dem um unser Museum, insbesondere für west-afrikanische Thiere, so hoch verdienten Herrn *Adolf Woermann* habe ich diese Fledermaus *Megaloglossus woermanni* zu nennen mir gestattet. Sie wird als ein Beweis einer gewissen alten Faunalbeziehung zwischen westafrikanischen und malayischen Gegenden betrachtet werden können, wie er ähnlich in den anthropomorphen Affen vorliegt, hier mit einer Ausdehnung nach Polynesien, wie sie für Fledermäuse möglich ist.

Wahrscheinlich wird sich das Thierchen weiter verbreitet finden, da die westafrikanische Fauna sich von der ostafrikanischen im allgemeinen erst am Tanganika-See und Albert-Nianza trennt.

Megaloglossus woermanni Pagenstecher, 1885

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Fruchtfressende langzungige Fledermäuse sind bisher nicht weiter westlich als im Himalaja gefunden worden. Unser Museum hat eben eine solche durch Herrn Soyaux von Ssibange-Farm in Gabon erhalten. Diese große Veränderung in unserer tiergeographischen Erfahrung berechtigt zu einer vorläufigen Mittheilung. Unser Thierchen gehört in die zweite Gruppe der Macroglossen bei Dobson: Zeigefinger mit Nagel, Zwischenkiefer vorn verbunden. Sie hat das volle Gebiß mit $\frac{2}{2} + \frac{1}{1} + \frac{3+2}{3+3}$ jederseits. Wäre nicht *Melonycteris* von *Macroglossus* geschieden, so hätte man diese Art auch unter *Macroglossus* lassen können. Wie in der Größe, so vermittelt sie mit einem Theile der Eigenschaften zwischen diesen beiden Gattungen; mit einem Theile aber entfernt sie sich weiter von *Macroglossus* als *Melonycteris*. Der Schwanz, bei *Melonycteris* fehlend, hat hier zwei Wirbel, bei *Macroglossus* drei. Die Flughaut am Fuße bei *Macroglossus* von der vierten Zehe, bei *Melonycteris* von der dritten entspringend, thut dies hier mit Bändchen gleichmäßig von der dritten und zweiten. Die Schnauze ist eher noch einfacher als bei beiden Gattungen; der zweite Lückzahn ragt oben und unten über das Niveau der übrigen: zwei hintere Gaumenfalten sind getheilt, wie bei *Melonycteris* während sie das bei *Macroglossus* nicht sind. Da die Zunge so lang ist wie bei dem erheblich größeren *Melonycteris melanops* Dobson, *alboscapulatus* Ramsay und zugleich breit, schlage ich den Gattungsnamen *Megaloglossus* vor.

Die Art ist dunkelbraun, am Leibe etwas heller, Gesamtlänge von der Schnauze zur Schenkelhaut 90, Unterarm 45, dritter Finger 80 mm.

Die genaue Beschreibung und Abbildung behalte ich mir vor für die Beilagen des Jahresberichts über das Museum für 1885.

Micropteropus intermedius Hayman, 1963

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(Fig. 8)

Type: F adult, 5060, in alcohol. Dundo, Lunda. N.E. Angola. Collected June or July 1953 by Dr. A. DE BARROS MACHADO. Brit. Mus. No. 62.2073.

Native name. - *Tshingunzu* (tshokwe).

Diagnosis. - Larger than *M. pusillus*, with forearm 59 to 63.5, thus falling within the bodily size range of *M. grandis* SANBORN, but cranial measurements notably less, particularly in the rostrum, palate and tooth row. Palatal ridges five (six in *M. grandis*), the first undivided, the remaining four paired, widely separated anteriorly, converging posteriorly so that the last pair are almost in contact, so that the pattern is quite distinct from the two parallel rows of divided ridges in *M. grandis* (see fig.). The number of ridges is as in *M. pusillus*, but the clear separation into distinct pairs is as in *M. grandis*, so that it can be said that not only the skull size but the palate ridge pattern indicates that this species is intermediate between the two other forms in this genus. Principal measurements of the type; forearm 63.5; skull greatest length 33.2, upper toothrow 10.4. (For full comparative measurements see table).

General description. - In all external characters closely akin to other *Micropteropus*. General colour dull brown above, hairs pale for two-thirds their length with dark brown tips. Ventral colour paler. The whole impression is of an animal darker than *M. grandis*. At the anterior and posterior borders of the ears are the normal Epomophorine pale tufts. The interfemoral membrane is well haired. Skull with rostrum less elongate than in *M. grandis*. Teeth slightly built.

The discovery that three distinct species of *Micropteropus* are sympatric at Dundo is of special interest, as until 1950, when SANBORN described *M. grandis*, the genus had been believed monotypic. The new form, *M. intermedius*, is quite clearly intermediate in character between *M. pusillus* and *M. grandis*, and one effect of its discovery is that the generic distinction of *Micropteropus*, as defined by ANDERSEN (1912), is no longer so clearly recognisable as compared with *Epomophorus*. The strong shortening of the palate ridge pattern indicates that this species is intermediate between the two other forms in this genus. *M. pusillus*, in both of which the rostrum is longer in relation to total skull length, and the highly specialised palate ridges of *M. pusillus* lead, in *M. grandis* and *M. intermedius*, to a type which, while still peculiar, is approaching that found in *Epomophorus*. Thus it can be said that these two species tend to link *Micropteropus* with *Epomophorus*.

In this connection it may be mentioned that although the smallest *Epomophorus*, *E. labiatus minor* DOBSON, has closely similar overall measurements to *Micropteropus grandis*, its very distinct palatal ridges (the first four undivided), together with its longer narrower rostrum and longer, narrower teeth, remove any possibility of confusion.

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[table removed - eds.]

Mimetillus thomasi Hinton, 1920

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Mimetillus thomasi, sp. n.

Type. - An adult female, collected at N'dola on Oct. 4, 1919 [by Capt. Guy C. Shortridge (see p. 239)]; original number 481.

This interesting bat, represented by a single specimen, is distinguished from its West-African relative, *M. moloneyi*, chiefly by its larger size, duller colour, and still flatter skull.

General outward appearance and all the essential characters of the genus as in *moloneyi*. Size larger, the forearm measuring 31 instead of 27 - 29 mm. Third digit of wing relatively a little longer, its total length equalling 165 % (instead of 146 - 159 %) of the length of the forearm. Fur on back slightly longer. General colour, above and below, dark brown, deepening almost to black on head, dull, lacking both the gloss and the deep chestnut tinge seen in *moloneyi*.

Skull considerably larger (condyle to canine 14.1, instead of 13 mm.), with the characteristic depression and flattening of the brain-case even more pronounced than in *moloneyi*; interorbital region relatively broader. Dentition without especial peculiarities.

External measurements (taken in flesh by collector). - Head and body 56 mm.; tail 38; hind foot 7.5; ear 13.

Measurements of wing (made on skin). - Forearm 31; third digit, total length 51, its metacarpal and phalanges 1 and 2 being respectively 32.5, 9, and 9.5; fifth digit 36, its metacarpal 29.5. Revilliod's index of width 49.

Skull. - Extreme length 14.6; condyle to canine 14.1; canine to m^3 5.2; width of brain-case in mastoid region 9.5; median occipital depth 4.7; interorbital breadth 5.3; breadth across preorbital swellings 8; width across outer borders of m^3 - m^3 7.8.

Mimetillus moloneyi was originally described from Lagos by Mr. Thomas. Many specimens were collected subsequently in Fernando Po; and we have lately received an example from Sierra Leone, collected by Mr. Willoughby P. Lowe. Hitherto no representative of the genus has been found inland or away from the West-African coast. Captain Shortridge's discovery in Rhodesia is therefore of considerable interest. I have great pleasure in naming the second species of this genus in honour of Mr. Oldfield Thomas, to whom I am indebted in so many ways.

Miniopterus Bonaparte, 1837*Miniopterus*

In description of *Vespertilio emarginatus*:

Introducendoci ora nel sottogenere *Vespertilio*, il cui tipo inevitabile è il *V. murinus*, L. lo vediamo corrispondere a quella sezione de *Vespertilioni* detta de' Murinoidi da Federico Cuvier, e tanto distinto dai nostri *Pipistrelli*, che se non si frapponesse il nuovo sottogenere *Miniopterus* lo riguarderemmo come genere confacendoci agli epistolari desiderii del dottissimo prof. Savi.

Miniopterus africanus Sanborn, 1936

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Type from Sanford's Ranch, Mulo, northwest of Addis Ababa, Shoa District, Ethiopia. Altitude 8,000 feet. No. 28769 Field Museum of Natural History. Adult female. Collected October 25, 1926, by A. M. Bailey. Orig. No. 73.

Diagnosis. - In general like *natalensis* but much larger. Skull and teeth larger. Zygomatic width greater than mastoid width.

Color. - Type, general color above very light Fuscous, bases of hairs Fuscous with light tips. Chin and throat dark where worn.

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Rest of under parts light, hairs Fuscous basally with gray tips. A few light gray hairs in pubic region.

A male and female topotype are practically like the type. Three other female topotypes have the head and shoulders near Natal Brown and the back near Clove Brown, some hairs with gray tips. The under parts are as in the type but browner.

Skull. - Large, with zygomatic width greater than mastoid width. Sagittal crest low. Teeth large.

Measurements. - Type and topotypes: forearm 50.4 (48.4 - 50.5). Skull: greatest length 16.9 (16.6 - 17); condylo-basal length 16.1 (16 - 16.5); palatal length 7.8 plus (8.2 - 8.8); interorbital width 4.2 (4.1 - 4.2); zygomatic width 9.4 (9 - 9.5); mastoid width 8.9 (8.7 - 9.1); width of brain case 8.3 (8.4 - 8.5); width across canines 5 (5 - 5.1); width across m^2 - m^2 , 7.1 (6.9 - 7.1); upper tooth-row c - m^3 , 6.6 (6.4 - 6.6); lower tooth-row c - m^3 , 7 (7 - 7); length of mandible 12.5 (12.2 - 12.7). Two alcoholic specimens from Kenya Colony: forearm 49.6 - 49.7; second digit metacarpal 45.9 - 45.9; third digit metacarpal 45.6 - 46.5, first phalanx 11.4 - 11.8, second phalanx 32.1 - 33.5, tip 6 - 6.5; fourth digit metacarpal 44.8, first phalanx 9 - 9.4, second phalanx 17 - 17.1; fifth digit metacarpal 40.6 - 41, first phalanx 9.8 - 10.6, second phalanx 7.6 - 7.6. Tibia 21.3 - 21.5; foot 11 - 11.

Specimens examined. - Type and five topotypes (six skins with five skulls) and two specimens in alcohol from Ulukenia Hills, Kenya Colony (U. S. Nat. Mus. Nos. 166595 - 96).

Remarks. - The large size and the color added to the more widely expanded zygoma will separate this form from all other African *Miniopterus*. Heuglin (Reise in Nord. Afrika, 2, p. 32, 1877) records a *Miniopterus* from Lake Tana which was identified by Peters as *M. dasythrix* of south and southwest Africa. His specimen may belong here but it is not recognizable from the description.

Miniopterus breyeri Jameson, 1909

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Miniopterus breyeri, sp. n.

I name this bat after my friend and former colleague Dr. H. G. Breyer, Chairman of Senate of the Transvaal University College, who accompanied me on the last of my collecting-expeditions in the Transvaal, when the series under description were captured.

It differs so markedly from the form inhabiting eastern South Africa that it requires a name to itself.

I may here say that it is probable that all the South African *Miniopteri*, with the exception perhaps of *M. minor*, Peters, will prove to be geographical subspecies of *M. schreibersi*, a form originally described from Southern Europe.

Sclater's 'Fauna of South Africa' treats the common South-African *Miniopterus* as *M. schreibersi*, but this form, which is of a light grey colour, does not appear to extend into South Africa at all.

Description. - Size as in *M. natalensis*, but lighter in colour. The hairs on the dorsal surface are reddish brown or reddish grey, the tips with a slight tinge of red-gold. On the ventral surface the tips of the hairs are rather lighter, becoming almost white in the pubic region.

The bases of the hairs are never black, as in *M. natalensis*.

Dimensions of the type: -

Head and body 56 mm.; tail 53; ear 11; tragus 6; forearm 46.

Type F. No. 398 (B.M. no. 9.7.2.6).

These examples were found in the great cave at Gatkoppies, in the Waterberg District of the Transvaal. There must have been many hundreds in one small chamber, the roof of which was black for many yards with a continuous cluster of bats.

All the examples examined were females.

Miniopterus breyeri vicinior J.A. Allen, 1917

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Type, No. 49019, M ad., skin and skull, Aba, northeastern Belgian Congo, Dec, 16, 1911; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 1770.

Differs from *M. breyeri breyeri* in slightly smaller size and in the absence of white on the pubic region.

Upperparts dark drab, the hairs unicolor from base to tip; underparts paler, grayish drab, the hairs dark brown basally with grayish brown tips; the pubic region fulvous; ears and membranes dark brown. Limb bones ventrally light yellow brown; calcar yellowish white.

Wing membranes attached at or slightly above ankle; no post-calcular lobe. Ears very small, uniformly convex on outer and upper border, posterior border nearly straight. Tragus long, narrow, with parallel edges, 4.8 mm. long by about 1.5 mm. wide.

Collectors' measurements of type: Total length, 100 mm.; head and body, 52; tail, 48; foot, 11; ear, 11. Forearm (from skin), 43.2; third metacarpal, 40; tibia, 28; foot, 8.8; ear, length, 6.5, width, 7.5.

Skull (type), total length, 14.8; zygomatic breadth, -; breadth of braincase, 7.5; interorbital breadth, 3.8; maxillary breadth, 5.6; breadth at base of canines, 3.8; maxillary tooththrow (with canine), 5.1; length of lower mandible, 10.4; height at coronoid, 3.2; lower tooththrow, 6.3.

Represented by 17 specimens (skins and skulls), all adult, and all taken at Aba, Dec. 10, 1911.

Collectors' measurements, 8 males and 8 females (MM; FF): Total length: 101 (94-105); 103 (96-108); Head and body: 54.5 (51-59); 54.0 (50-57); Tail: 46.3 (43-49); 49.0 (46-50); Foot: 10.6 (10-11); 10.5 (10-11); Ear: 10.6 (10-11); 10.7 (10-11).

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Forearm, from skins, same specimens, M 43.6 (42-44.7), F 44.2(43.4-44.9).

Skull, 5 males, 6 females: Total length, M 14.6 (14.3-15), F 14.7 (14.4-14.9); breadth of braincase, M 7.4 (7.2-7.6), F 7.5 (7.3-7.7).

The present form is similar in general coloration to *M. breyeri breyeri*, from the Waterberg District of the Transvaal, with two topotypes of which the present series has been compared. The pubic region is pale fulvous, however, instead of white, and the average size is considerably less, the forearm in *breyeri* having a length (3 specimens) of 46 mm. and in *vicinior* (16 specimens) 43.5, the largest specimen of the latter being smaller than the smallest of the *breyeri* specimens. Jameson gives the length of the skull in *breyeri* as 15.5; the average in *vicinior* is 1 mm. less, only one in a series of 12 skulls reaching 14 mm.

M. breyeri vicinior is readily separable from the *M. natalensis* group by its paler, much browner coloration and by other features, as shown by direct comparison with typical *natalensis* and with topotypes of *natalensis arenarius*.

[*Miniopterus fraterculus*](#) Thomas and Schwann, 1906

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M. 1073, 1077. F. 1079. Knysna

Closely similar in every respect to *M. dasythrix*, agreeing with that species absolutely in colour, even to the peculiarity in the respective coloration of the two sexes. Thus the back of the male is smoky blackish, the head and whole of the under surface dark brown (darker than Prout's brown); the female is dark brown above and below, rimmed with black along the hinder part of the back. But the skull and teeth are very markedly smaller and the forearm rather shorter; though the latter is not so much shorter than in *dasythrix* as might have been expected from the difference in the skulls.

Dimensions of the type, the starred measurements taken in the flesh: -

Forearm 43.7 mm. (other specimens, M 43, F 44).

*Head and body 54 mm.; *tail 53; *ear 9; *tragus 4; third finger 79; lower leg and foot (c.u.) 27.5.

Skull - greatest length 14.7; basal length in middle line 11.2; breadth of brain-case 7.5; height of brain-case from basion 6.3; palate length 5.7; front of canine to back of m^3 5.5; greatest breadth of palate, including molars, 5.6; front of lower canine to back of m_3 5.7.

Type. Male. B.M. No. 5.5.7.18. Original number 1073.

Taken 3 October, 1905.

"In cave on sea-coast."

Like as all the eight specimens of *Miniopterus* found at Knysna are to each other in most respects, we have come to the conclusion that they cannot be referred to one species, as in size they fall into two groups, without intermediates. We have therefore described the smaller form as new, while the larger, as already noted, may be referred to *M. dasythrix* Temm.

"Fairly common. *Miniopteri* were taken in both the caves where *Rhinolophus augur* and *Rousettus collaris* were secured." - C.H.B.G.

[*Miniopterus gleni*](#) Peterson, Eger and Mitchell, 1995

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(MMAD, fig. 60 et 61)

Holotype: Royal Ontario Museum (ROM) 42567, mâle adulte, crâne et peau récoltés le 9 mai 1967 dans une grotte marine entre Sarodrano et Saint-Augustin, à 20 km au Sud de Tuléar.

Description. - Pelage long, soyeux, sombre; poils du dos longs de 7 à 8 mm: Brun Girofle à Brun Foncé (Ridgway 1912); poils du ventre longs de 5 à 6 mm, couleur générale un peu plus pâle que sur le dos. Oreilles basses, arrondies, à large teagus uniforme (fig. 60). Museau large et poilu. Crâne grand et large avec la boîte crânienne haute, longue et arrondie (fig. 61). Formule dentaire I 2/3, C 1/1, P 2/3, M 3/3.

Diagnose. - Le plus grand des *Miniopterus* malgaches. Diffère de *M. inflatus* d'Afrique (*M. i. inflatus*, MIIN; *M. i. rufus*, MIRU) par les tibias nettement plus longs; les 3^{ème} et 4^{ème} métacarpiens nettement plus longs (3DME et 4DME), la calotte crânienne plus haute (HBC), le rostre plus large (ROSW et CCUP); les rangées dentaires (M^3 - M^3) et C- M^3) plus longues et plus larges et par le plus fort rétrécissement post-orbital (POC). Il diffère de *M. africanus* (MXAF) par le tibia significativement plus court, le 5^e métacarpien plus court (5DME), la plus grande largeur du rostre (ROSW, M^3 - M^3 , CCUP), la plus grande longueur du palais

(PALL et PALN). mais la partie post-palatale plus courte (PPL). la plus grande largeur de la rangée dentaire (C-M³) et de la mandibule (MANL). Le tableau 26 permet de comparer *M. gleni*, *M. inflatus* et *M. africanus*. *M. gleni* est significativement différent

[table removed - eds.]

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des autres *Miniopterus* malgaches par tous les caractères, sauf celui de la première phalange, qu'il partage avec *M. majori* (MSMD).

Distribution. - Répandu dans la plus grande partie de Madagascar, et, en particulier, dans les régions côtières.

Nous avons étudié 77 spécimens de cette espèce.

MADAGASCAR NORD: Cap Diégo (ZMK).

MADAGASCAR-EST: 20 km au N.-O. de Maroantsetra. - 40 km au N.-O. de Maroantsetra (MNHN et FMNH).

MADAGASCAR CENTRE: Maninday (ROM). - Beroroha (BMNH).

MADAGASCAR SUD-OUEST: Sarodrano, Saint-Augustin, 20 km au S. de Tuléar (ROM).

Variation systématique. - Etant donné que l'analyse multivariable ou univariable n'a pas permis de trouver de différences entre les populations du Nord-Est et celles du Sud-Ouest, traitées séparément, l'ensemble de l'échantillon malgache a été regroupé en un seul OTU.

Les mesures sur le terrain des exemplaires capturés par nous dans le Sud-Ouest malgache, de l'holotype * et des valeurs moyennes et extrêmes de huit mâles et de quatorze femelles étaient les suivantes:

Poids: 15*, mâle 14.25 (11 - 16); femelle 13.6 (11 - 17).
Longueur totale: 124*, mâle 120.4 (110 - 126); femelle 117.6 (112 - 126).
Longueur de la queue: 56*, mâle 55.2 (53 - 57); femelle 55.2 (51 - 60).
Longueur du pied: 11*, mâle 11.4 (11 - 13); femelle 11.3 (10 - 13).
Longueur de l'oreille au-delà de l'entaille: 15*, mâle 15.0 (14 - 16); femelle 14.8 (14 - 16).
Envergure alaire: 358*, mâle 356.0 (344 - 360); femelle 356.7 (342 - 369).

Les longueurs de l'avant-bras, du tibia et les mesures du crâne de l'holotype et les statistiques basées sur une série de 58 *M. gleni* (MMAD) figurent tableau 26.

Biologie. - On trouve, nichant avec *M. gleni*, dans la région de Saint-Augustin, les espèces suivantes: *M. majori*, *M. manavi*, *Trienops furculus*, *Myotis goudoti* et *Otomops madagascariensis*. On l'a trouvé en association sympatrique avec *M. fraterculus*; dans des grottes à 20 km au N.-O. de Maroantsetra en juin 1936 (MNHN à Paris). Sur la base des données actuelles, *M. gleni* n'a pas été trouvé associé à *M. manavi* dans la région centrale, dans la savane de l'Ouest ou à la Montagne d'Ambre. A Madagascar, le même gîte est souvent partagé par deux ou quelque fois trois espèces de *Miniopterus*.

Miniopterus inflatus Thomas, 1903

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Colour of *M. scotinus*. Size larger than in *M. Schreibersi*.

Fur of back about 4 mm. in length, its texture about as in *M. scotinus*, not extending on the interfemoral or wing-membranes except close to the sides below. Colour uniform dark "seal-brown" above, rather paler below. Ears and membranes black.

Skull markedly larger than in *M. Schreibersi*, therefore still more exceeding that of the similarly coloured *M. scotinus*.

Dimensions of the type: -

Forearm 46 mm.

"Head and body 65; tail 48; hind foot 7; ear 11" (G. L. B.).

Skull: greatest length 16.7; basal length in middle line 12.3; front of incisors to back of m³ 7.8

Hab. Efulen, Cameroons.

Type. Adult skin (male). B.M. no. 3.2.4.8. Collected 24th July, 1901, by Mr. G. L. Bates.

This *Miniopterus* is at once distinguishable from *M. Schreibersi* by its dark colour and large head, and from *M. scotinus* and *M. Newtoni* by its much larger size.

Miniopterus inflatus villiersi Aellen, 1956

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?*Miniopterus inflatus* J. ALLEN, 1917; SANBORN, 1936; HAYMAN, 1954.

8 spécimens, n° 54-1-1 (type) à 54-1-8. 3 M. 5 F. Grotte du Marché, Dalaba, 16-4-1954.

Aucun *Miniopterus* n'est signalé jusqu'à présent en A.O.F. En effet, MONARD (1939), VEIGA-FERREIRA (1949), CANSDALE (1948), ROSEVEAR (1953), etc. n'en citent pas. Ce genre n'a pas été trouvé plus à l'Ouest qu'au Cameroun anglais où SANBORN (1936) indique "*M. inflatus*".

Avant d'examiner ces spécimens de Guinée française, j'avais déjà été surpris de constater que divers auteurs rapportaient à *inflatus*, avec quelques doutes d'ailleurs, des Chauves-Souris de taille nettement inférieure.

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Grâce au matériel rapporté par J. L. PERRET du Cameroun français, je suis maintenant en mesure de confirmer la diagnose de *inflatus* de THOMAS (1903). En effet, je possède 6 spécimens de Nkoétyé, près d'Ebolowa, qui correspondent parfaitement à *inflatus*, en particulier par leurs grandes dimensions. Ce sont presque des topotypes, puisque la localité typique de *inflatus* est Efoulen, situé à quelque 90 km. à l'Ouest de Nkoétyé, dans le même biotope.

Je suis persuadé que les "*inflatus*" de J. ALLEN (Thysville, Bas Congo), de SANBORN (Mont Cameroun) et de HAYMAN (Thysville: Kakontwe; Mont Homa, Congo belge) appartiennent à une autre forme. Les spécimens de Guinée française semblent correspondre à ces derniers et je propose de les nommer *Miniopterus inflatus villiersi*. Il sera peut-être permis d'élever cette forme au rang spécifique, s'il s'avère qu'elle cohabite avec *M. inflatus* typique.

TYPE. - M ad., grotte du Marché, Dalaba, Guinée française, 16-4-1954, collecté par A. VILLIERS. N° du collecteur 2839. N° 54-1-1 des collections zoologiques de l'IFAN, Dakar. 7 paratypes don't 2 M et 5 F.

DIAGNOSE. - *Miniopterus* de taille moyenne, voisin de *inflatus* du Cameroun, mais s'en distinguant par un avant-bras, un pouce et des mesures crâniennes plus petites.

DESCRIPTION. - La coloration de la fourrure est brun noir uniforme. Le patagium et les oreilles sont brun grisâtre. L'extrémité distale du tragus est blanche, dépigmentée; ce caractère s'est révélé constant chez les 8 spécimens.

Le crâne présente la particularité d'avoir la largeur zygomatique plus grande que la largeur mastoïde, ce qui distingue cette nouvelle forme de tous les autres *Miniopterus* africains, à part *africanus* (1) et *rufus* (2). SANBORN (1936 : 111) dit que *inflatus* a des largeurs mastoïde et zygomatique subégales ou que la première est plus grande que la seconde. Ce caractère n'est pas signalé dans la description originale et les auteurs subséquents qui citent *inflatus* n'en parlent pas non plus. Chez les *inflatus* typiques rapportés par J. L. PERRET, la largeur zygomatique est nettement plus grande que la largeur mastoïde.

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Dans le tableau suivant, on trouvera les mesures de la nouvelle, forme et, à titre de comparaison, celles de *inflatus* typique.

[Table removed - eds.]

Les "*inflatus*" mesurés par J. ALLEN, SANBORN et HAYMAN sont, tout à fait comparables à nos *inflatus villiersi*:

avant-bras : (A) 44.2 - 46; (S) 42.2 - 45.4; (II) 44.4 - 44.6;
crâne. Long. Tot.: (A) 15.3 - 15.9; (S) 15 - 15.3;
- larg. Zygom.: (A) 8.6 - 8.7;
- rang. Dent. I-M³: (S) 6.9.

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ECTOPARASITES. - 4 spécimens d'une nouvelle espèce de Nyctéribiidé ont été pris sur ces *Miniopterus*. Cette espèce sera décrite par O. THEODOR sous le nom de *Nycteribia exacuta*, dans son prochain article sur les Nyctéribiidés éthiopiens.

Un spécimen de *Raymondia huberi* FRAUENF. examiné par B. JOBLING a également été trouvé, de même que quelques Acariens non encore déterminés.

Miniopterus Majori Thomas, 1906

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Size large, slightly exceeding that of the largest South-African members of the genus. Fur fairly long, hairs of back about 6 mm. in length. Colour dark glossy black, with a slight brownish suffusion, the head not or scarcely lighter than the back; the hairs black for their basal half, then dull yellowish brown, their terminal fourth glossy black. Under surface little lighter, the ends of the hairs dark brown instead of black. Fur extending slightly on to interfemoral membrane both above and below.

Dimensions (see below).

Hab. Madagascar. Type from Imasindrary, N.E. Betsileo. Other specimens from Vohimar.

Type. Adult female. B.M. no. 97.9.1.33. Original

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number 457. Collected 3rd July, 1895, by Dr. C. I. Forsyth Major, after whom the species is named.

This large Malagasy *Miniopterus* differs from *M. dasythrix*, Temm., its South-African ally, by its glossy black colour and the dull yellowish rings on the dorsal hairs, that animal being of a "noir mat," as stated in the original description, confirmed by fresh specimens recently received from Knysna (Rudd Collection).

Miniopterus manavi Thomas, 1906

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Size small, much smaller than in *M. Majori*, which occurs in the same localities. Fur of medium length, hairs of back about 5 mm. Fur of body extending thinly over nearly half the upper surface of the interfemoral membrane and on the proximal fourth below. General colour blackish, not so glossy as in *M. Majori*, the hairs blackish for two thirds their length, with dull brown tips. Under surface similar, but rather paler.

Some specimens are dull reddish throughout, representing a rufous phase of the species.

Skull comparatively small and delicate, with a smooth and narrow brain-case.

Dimensions (see table below).

Hab. Madagascar. Type from Imasindrary, N.E. Betsileo. Other specimens from Vinanitelo.

Type. Male. B.M. no. 97.9.1.37. Original number 453. Collected 3rd July, 1895, by Dr. C. I. Forsyth Major.

This is the bat called by Dobson *Miniopterus scotinus*, Sund., but Sundevall's species has a forearm 44 mm. in length, and differs also by its much paler colour and its African habitat. *M. Majori* again was referred by Dobson to *M. Schreibersi*, from which, to use his own words, it "only differs in colour and in the forearm being constantly longer."

Peters's *M. minor* is presumably nearly related to *M. manavi*, but from its continental habitat (Zanzibarà) is no doubt specifically distinct.

Dimensions of the two *Miniopteri* above described: -

M. majori; *M. manavi*
mm
Forearm: 46.3; 39.2
skull:
Greatest length: 15.5; 13.8
Basal length in middle line: 12; 10.4
Front of canine to back of m^3 : 6.4; 5.0
Breadth of palate outside m^2 : 6.6; 4.9
Front of lower canine to back of m_3 : 6.7; 5.4

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The difference in size between these closely allied species affords an instance of the curious tendency of Chiroptera to sort themselves into couples, a larger and a smaller form of so many different groups being often found, as in this case, inhabiting the same locality.

Miniopterus minor Peters, 1867

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In der Färbung, dem Ansatz der Flughäute, der Bildung der Spornen u. s. w. mit *Miniopterus Schreibersi* übereinstimmend, aber beträchtlich kleiner als diese Art.

Totallänge 0^m.086; Kopf 0^m.0153; Ohr 0^m.010; Schwanz 0^m.042; Vorderarm 0^m.037; Mittelfinger 0^m.072; Tibia 0^m.014.

Ein einziges ausgewachsenes Männchen von der Küste von Zanzibar.

Miniopterus minor griveaudi Harrison, 1959

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in honour of its discoverer, M. Paul Griveaud.

Type: Adult female. Grande Comore Island, Comoro Islands, Indian Ocean. 10 August, 1958. Obtained by M. Paul Griveaud. In my collection, No. 3.2803.

Description: Differs from *Miniopterus minor* Peters in the decidedly narrower braincase and rostrum of the skull (See Photo) and also in colour, being a darker shade of brown on the back and belly. It differs from *Miniopterus minor manavi* only in colour, there being two colour phases in that form, either blackish on the back or dull reddish, as described by Thomas (1906). The dorsal surface of the Type of *M. minor griveaudi* closely approximates to Bison (Plate Maerz and Paul, 1950). On the belly it is slightly

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paler brown but again darker than the examples of *M. m. minor* and different from both colour phases of *M. m. manavi*.

Measurements of the Type: Forearm 37.8, hind foot 7.2, ear 7.8 mm.

Cranial measurements (in mm.): Condylbasal length 13.1, zygomatic breadth 6.7, interorbital constriction 2.9, breadth of the braincase 6.2, mandible 10.5, maxillary dentition c-m³ 5.0, mandibular dentition c-m₃ 5.4, width of the palate from the external alveoli across m²-m² 4.9.

Remarks: In contrast to the material of *M. m. manavi*, the paratypical series of *M. m. griveaudi* are very uniform in colour and the five paratypes, three of which are now in the Museum d'Histoire Naturelle, Paris, differ little from the Type. At this stage little definite can be stated about the exact shade of colour found in *M. m. minor*, which was not described in detail by Peters and the two examples here recorded had been immersed for a short while in alcohol and their tint may thus have been slightly altered, but it is clearly different. However, all three skulls of *M. m. griveaudi* which have been examined differ strikingly from *M. m. minor* in their narrow, delicate braincases, whilst the difference in colour from

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M. m. manavi is equally striking. It is interesting to note that the fur of the body extends thinly over the proximal half of the upper surface of the interfemoral membrane in all the examples of *M. m. manavi* in which this character can be ascertained, as stated by Thomas in his description. In two of the three examples of *M. m. griveaudi* this part of the membrane is naked, while in one it is haired thinly as in *M. m. manavi* but less marked. In the case of the two examples of *M. m. minor* both have a thin hairy covering of this part of the interfemoral membrane. It is clear therefore that this character does not distinguish the Madagascar form specifically, but that it is of some value in subspecific determination. No differences in dentition have been noted.

The measurements of all the material examined are tabulated below, compared with those of the Type of *M. minor* after Peters.

[table removed - eds.]

Miniopterus natalensis arenarius Heller, 1912

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Æquatorial Broad Winged Bat

Type from the Guaso Nyuki, Northern Guaso Nyiro River, British East Africa; adult female; No. 181811, U.S. Nat. Mus.; collected by Edmund Heller, October 4, 1911; original number, 4413.

Characters. - Size of *natalensis*, but color paler, pelage shorter; the skull with broader brain-case and with inner incisor greatly exceeding outer in size.

Coloration. - Dorsal color seal-brown, the head darker, clove-brown, and the rump lighter walnut-brown, the hair only slightly darker at the base; underparts washed with drab-gray, the hair basally dark slaty; ears and membranes blackish, the latter narrowly white bordered.

Measurements. - Head and body, 55 mm.; tail, 47; hind foot, 9; ear, 11; forearm, 45.

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Skull: Greatest length, 14.8; condylo-incisive length, 14.3; basilar length, 11; zygomatic length, 5.6.; upper cheek teeth, 4.8; condylo-incisive length of mandible, 10.9; coronoid-angular depth of mandible, 3.5.

The type was secured in a large crevice among granite boulders on the summit of a small kopje. One other specimen was shot at dusk among some acacia trees in the same vicinity.

Miniopterus Newtoni Bocage, 1889

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De la taille à peu-près du *M. scotinus*. Pelage d'un beau brun-marron, plus pâle, tirant au roux, sur la partie inférieure de l'abdomen; membranes alaires et caudale noires. Museau court, glanduleux, ayant de chaque côté de la face, au-dessous de l'œil, un sillon horizontal profond. Oreilles beaucoup plus courtes que la tête, d'une forme à peu-près quadrangulaire, à contour supérieur droit; tragus allongé, étroit, arrondi à la pointe. Ailes se prolongeant jusqu'à l'extrémité inférieure de la jambe. Pieds longs et étroits. Queue entièrement comprise dans la membrane interfémorale, de la même longueur que la tête et le tronc réunis. Une bande étroite de poils, de la couleur du dos, sur la face dorsale de la membrane alaire, limitée en dehors par une ligne tirée du tiers supérieur de l'humerus au milieu du fémur; en dessous tout l'espace compris entre l'humerus et le fémur est revêtu de poils plus clairsemés; la membrane interfémorale nue sur ses deux faces.

Tête et corps 45 mm., queue 43 mm., tête 16 mm., oreille 95 mm., tragus 45 mm., avant-bras 39 mm., pollex 5 mm., 3^e doigt - metac. 34 mm., 1^e phal. 10 mm., 2^e phal. 28 mm., jambe 16 mm., pied 9 mm.

Nos individus de St. Thomé me semblent bien distincts du *M. scotinus*, de Madagascar et de la côte orientale d'Africa, le seul avec lequel il serait possible de les confondre, par son système de colora-

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tion et surtout, par une disposition tout-à-fait différente des poils sur les membranes alaire et interfémorale. Je suis arrivé à cette conclusion non pas par suite d'une comparaison directe des deux espèces, car le *M. scotinus* ne se trouve pas représenté dans nos collections, mais en me servant de la description de cette espèce, publiée par M. Dobson dans son *Catalogue des Chiroptères du Muséum Britannique*. "Le *M. scotinus*, suivant cet auteur, est d'un brun-roux ou brun-foncé, presque noir, avec les parties inférieures d'une teinte plus pâle. En dessus l'aile porte une bande distincte de poils courts entre le coude et le cou-de-pied, tandis que l'espace compris entre la moitié externe de l'humerus, le tibia et le genou est presque entièrement nu; la moitié de la membrane interfémorale est couverte de poils courts et clairsemés, qui forment aussi une bande sur le bord interne de la jambe jusqu'à la base du calcanéum".

Miniopterus rufus Sanborn, 1936

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Type from Katobwe, 40 miles below Bukama on the Lualaba River, Tanganyika-Moero, Belgian Congo. No. 29416 Field Museum of Natural History. Adult male. Collected November 28, 1926, by J. T. Zimmer. Orig. No. 566.

Diagnosis. - About the size of *M. africanus* but with shorter forearm and tibia and of a reddish brown color.

Color. - Reddish brown, between Kaiser and Carob Brown (Ridgway, 1912), slightly lighter below. Hairs uniform to bases.

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Skull. - In general like *africanus* but narrower and with lambdoid and sagittal crests more developed and mandible heavier.

Measurements. - Forearm 47.4. Skull: greatest length 16.9; condylo-basal length 15.9; palatal length 8.7; interorbital width 4.1; zygomatic width 9.4; mastoid width 9.1; width of brain case 8.2; width across canines 5; across m²-m², 6.7; upper tooth-row c-m³, 6.4; lower tooth-row c-m³, 7; length of mandible 12.8.

Remarks. - While there is but one specimen, the type, the character of the skull and the very different color seem to be of enough importance to separate this form from other African *Miniopterus*. There seems to be no red phase in this genus as in *Hipposideros*, *Myotis*, and others, but there has been another *Miniopterus* described which is of about this color, *M. rufus* Sody, of central Java.

Mops (Allomops) faradjius J.A. Allen, 1917

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Type, No. 49222, M ad. (in alcohol), Faradje, northeastern Belgian Congo, Nov. 1, 1913; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 3015.

Similar to *A. osborni* in general size and in coloration, but with a shorter forearm (44 instead of 49 mm.), a vestigial p² on the right side only, in the toothrow (visible only by aid of a strong lens), less developed sagittal and lambdoid crests (doubtless due to the fact that it is much younger than the type of *osborni*), darker upperparts and a more decided brownish band across the pectoral region.

Upperparts blackish brown, much darker than in *osborni*; underparts blackish brown along the sides and across the throat, forming a darker and broader pectoral band, the central area broadly whitish. Ears and membranes blackish, and in all other respects similar to those of *osborni*. The broad patch of glandular hairs between the ears deep black, fuller and longer than in *osborni* and covering the membrane connecting the ears, but not lengthened into a crest as in *Lophomops*.

Total length, 108 mm.; head and body, 76; tail, 32; forearm, 43.5; third metacarpal, 42.5; third finger, 82; ear from crown, 10; from notch behind antitragus, 17; expanse from tip to tip of both ears, 32.8; tibia, 15; foot, 9.

Skull, total length, 22.2; zygomatic breadth, 12.9; mastoid breadth, 11.5; breadth of braincase, 9.3; interorbital breadth, 4; maxillar breadth, 8.8; breadth at base of canines, 6.7; upper tooththrow (c-m³), 7.6; length of mandible, 14; angle to condyle, 4.5; depth at coronoid, 3.7; lower tooththrow, 8.7; transverse extent of lambdoid crest at upper border, 6.7.

Represented by only the type, taken at Faradje, and labelled by the collectors

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as "from the; stomach of a *Machærhamphus* hawk." The specimen (preserved in alcohol) is practically uninjured for study.

As noted above, *Mops (A.) faradius* closely resembles *osborni*, but it has a much shorter forearm; a slenderer body, and though a male, is somewhat smaller in both external and cranial measurements. It is also markedly darker in general coloration. The type localities of the two forms (Kinshasa and Faradje) are far apart and in regions of widely different environment.

Mops (Allomops) nanulus J.A. Allen, 1917

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Type, No. 48864, M ad., skin and skull, Niangara, Belgian Congo, Dec. 12, 1910; Herbert Lang and James P. Chapin, American Museum Congo Expedition. Orig. No. 1318.

Similar in cranial characters and in dentition to *Mops (A.) occipitalis*, but much smaller and much lighter in coloration.

Upperparts cinnamon-brown; below with the whole median area lighter, the tips of the hairs dull gray with a slight yellowish or buffy tone, the sides much like the back but rather darker; extreme lower back naked or with only a slight pubescence; a thin fringe of bristly hairs from the posterior border of the dorsal pelage, the longer of these reaching to the base of the tail. Wing membranes from the sides of the back, light brown above, somewhat paler apically and on the lower surface. Interfemoral membrane palish brown above and below. Ears of medium size and thickness connected by a low membrane, from the back of which (most prominent in males) is a tuft of lengthened hairs, as in its larger congeners; inner base of ears heavily furred and otherwise agreeing closely in form and structure with those of *occipitalis*. Tragus small, pointed, black. Antitragus of medium size, broad basally, convex on upper border. Tail short, rather more than the apical half free.

Skull and dentition similar to that of *A. occipitalis*.

Type, collectors' measurements: Total length, 82 mm.; head and body, 61; tail, 21; foot, 9; ear, 15. Forearm (from skin), 28.5. (The type is the largest of a series of 6 specimens.)

Type and 5 topotypes (3 males, 3 females), collectors' measurements: Males, total length, 78.7 (75-82), females, 76 (76-77); head and body, males 58 (56-61), females 55.7 (54-58); tail, males, 20.3 (19-21), females, 20 (19-21); foot, males 8 (7-8), females 9 (all 9); ear, males 15.5 (15-17), females 17 (16-18). Forearm (from skin), males 28.6 (28.3-28.7), females, 27.4 (27.3-27.0).

Skull (type), total length, 16.8; condylobasal length, 15.5; zygomatic breadth, 10.9; mastoid breadth, 9.6; interorbital breadth, 3.3; maxillar breadth, 7.3; breadth at base of canines, 3; depth (including pterygoids), 5.7; upper tooththrow (with canine), 6.1; length of mandible, 11.2; height, angle to condyle, 3.5; depth at coronoid, 2.9; length of tooththrow, 6.8.

Skull, type and 5 topotypes (3 males, 3 females): males, total length, 16.5 (16.0-16.8), females 15.4 (15.3-15.0); zygomatic breadth, males 10.3 (10.5-11), females; 9.9 (9.8-10). The females are thus smaller than the males, and they also lack the extreme development of the supraoccipital region shown by the males.

Represented by 7 specimens, all collected the same day at Niangara.

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This species is readily distinguished from all other known African molossid bats by its extremely small size, aside from any other characters.

Although the premaxillæ are slightly emarginate behind the incisors, the other characters both external and cranial, agree completely with the larger forms here referred to *Allomops*, even in the sexual difference, and especially the sexual difference in the size of the lower premolar, p₂ being larger than p₄ in the males and smaller than p₄ in the females.

Mops (Allomops) occipitalis J.A. Allen, 1917

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Text Fig. 15

Nyctinomus thersites THOMAS, Ann. and Mag. Nat. Hist. (7), XII, p. 364, Dec. 1903. Efulen, Cameroons.

Type, No. 48851, ad. M, skin and skull, Avakubi, Belgian Congo, Feb. 13, 1914; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 2487.

Pelage short, soft and velvety, about 1.5 mm. in length on the back to about 2.5 on the sides of the neck, throat, and sides of body below. External base of ears and whole space between them thickly covered with slightly lengthened glandular black hairs, which also cover the back of the low connecting membrane, but do not rise much above it.

Fur of upperparts dark chocolate-brown; the sides of the back, the lower back from a line joining the hips posteriorly, and the hind limbs naked and blackish, the central furred portion of the back, from considerably in front of the shoulders to hips, of about the same width as the naked area each side. A scanty fringe of long bristly hairs (blackish or mixed with a few whitish ones) from the hips, directed backward, the longest reaching to or beyond the middle of the uropatagium (scanty or nearly wanting in some specimens). Underparts blackish laterally, the median area buffy brown; a ventral area (corresponding ventrally with that of the lower back) and limbs naked, pale rusty brown (the denuded space much reduced in some specimens). Ears near together, blackish brown, united by a low membrane which extends forward as a swollen obtuse projection reaching nearly half-way to the nose; front and upper border of ears thickened, forming a sharp angle at the anterior inner border; tragus merely a minute dusky point, about 2 mm. in height; antitragus brownish, small, about 4 X 4 mm., bluntly convex above. Face naked, blackish; nostrils prominent, upper lip weakly corrugated; chin naked, brownish. Wings from middle of tibia. Forearm and membranes naked. Wing membranes above, including antebrachium, blackish brown, lighter, grayish brown apically; below simi-

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lar, the humerus and forearm whitish, in strong contrast with the membrane. Uropatagium dark brown and naked on both surfaces.

Collectors' measurements of type: Total length, 98; head and body, 68; tail, 30; foot, 11; ear, 18. Additional measurements from the skin: Forearm, 38; tibia, 15.5; foot, 10; ear from crown, 9; from notch behind antitragus, 16.

Collectors' measurements of 4 specimens (2 males, 2 females) from Medje (near type locality): Total length, 99 (98-101); head and body, 70 (68-73); tail, 29.2 (28-30); foot, 9.6 (8-11); ear, 18 (all 18). Forearm (from skins), 38 (36-39).

Measurements of the type skull: Total length, 20.4; condylobasal length, 18; zygomatic breadth, 12.7; mastoid breadth, 11.5; breadth of braincase, 9.5; interorbital breadth, 4.1; maxillar breadth, 8.2; breadth at canines, 5.8; depth (including pterygoids), 7.8; length of upper toothrow (c-m³), 6.9; length of mandible, 13.4; angle to condyle, 4.5; depth at coronoid, 4; length of lower toothrow, 7.4 (Fig. 15).

Four skulls from Medje: Total length, 2 young adult males, each 91.8; 2 young adult females, 18.3, 18.8; zygomatic breadth (2 males, 1 female), all 12.5.

In the adult males the skull has the occipital region enormously developed, the lambdoid crest being high and broad with a marked backward overhang. In females and young males (subadult) it is much less developed. The mandible is noticeably massive in even young specimens.

Represented by the type, from Avakubi, and 6 specimens from Medje, of which 2 are about half grown and the others young adults, collected Sept, 15-16, 1910.

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Judging from the description, *Nyctinomus thersites* Thomas, greatly resembles the present species but whether they are identical cannot be satisfactorily determined without direct comparison of the types. The description of *thersites* makes no direct mention of the coloration of the underparts, which in *occipitalis* are very unlike the upperparts, the coloration of *thersites* being described as "Colour brownish chestnut, glandular hairs back of ears black." Other features mentioned point strongly to a form greatly resembling *occipitalis*. In *thersites* the premaxillæ are thus described: "Premaxillæ separated, but opening between them small," for which reason the species has been regarded as a *Nyctinomus*. In six skulls of *occipitalis* the premaxillæ are fully ossified in four of them and in the other two (one of them the type and the most mature specimen) there is a slight opening behind and between the incisors. In a skull of *A. faradjius* (the only one available for examination) the premaxillæ are fully ossified. In five adult skulls of the much smaller *A. nanulus* there is a very small opening at the inner base of the incisors in all, but in all other characters, external and cranial, including color pattern and femoral bristles, *nanulus* is obviously only the little brother of the three larger forms here referred to *Allomops*.

Mops (Allomops) osborni J.A. Allen, 1917

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Text Fig. 12-14, and 26.

Type, No. 49230, M ad. (in alcohol), Kinshasa (near Leopoldville), Belgian Congo, Dec. 22, 1914; cotype, No. 94244, F ad. (alcoholic), same locality and date. Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. of type 2570.

Upperparts dark hair-brown with a grayish sheen, the hairs whitish at extreme base; underparts, sides paler brown than back, the hairs grayish-tipped, the central area from chest posteriorly broadly white, throat and an indistinct pectoral band; grayish brown; wing membranes and limb bones above dark brown, below lighter, becoming whitish toward the body, including the propatagium and a narrow posterior edging; uropatagium dark brown above, whitish below; ears united by a deep band, the back of the membrane (in both sexes), covered with lengthened hair-brown hairs which barely reach its upper border (Fig. 14). Tragus minute; antitragus small (about 4 X 4 mm.), evenly rounded above. Sides of back, lower back and hind limbs above naked and dark brown; under side of limbs and anal region naked and yellowish white. Face dark brown; upper lip heavily corrugated. No gular pouch in either sex. Feet heavily clothed with spoon hairs (see Fig. 20, p. 556).

Total length, male type, 111.3, female cotype, 106.6; head and body, 73.5, 70, tail, 37.8, 36.6; forearm, 49, 47; third metacarpal, 47.2, 46.5; tibia, 17.5, 16.5; foot 10.5, 10.2; ear from crown, 8.5, 9; length from notch at posterior base of antitragus, 14.7, 12.5; expanse (tip to tip), 33.8, 32.

Skull in the old-adult male type with a highly developed sagittal crest, as in *Mollossus*, but differing from the latter in having also the lambdoid crest enormously developed, as in some other species of the *Mops* group. In the male type the sagittal crest has a height of 1.8 mm. and extends from the interorbital region to the lambdoid; in the female cotype neither the sagittal nor the lambdoid is more developed than in some species of *Nyctinomus* (e.g., *N. ochraceus* and *N. leonis*). Rostral and interorbital regions broad (as in other species of the group), the braincase wide and rounded; premaxillæ fully ossified; palate deeply concave; basisphenoid pits small and shallow; p² vestigial, on the outer edge of the toothrow and difficult to see without the aid of a strong lens. The type has only one upper incisor, the right having been lost in life, (Figs. 12 and 13).

Measurements of type (male) and cotype (female) skulls: Total length, M 22.9,

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F 20.7; condylobasal length, M 19.6, F 18.6; zygomatic breadth, M 13.4, F 12.6; mastoid breadth, M 12.2, F 11.8; breadth of braincase, M 10.4, F 9.9; interorbital breadth, M 4.4, F 4.5; breadth at base of canines, M 6, F 5.7; maxillary breadth (between outer borders of m³), M 9, F 9.2; depth of braincase medially (without the crest), M 7, F 6.8; upper toothrow (including canine), M 7.7, F 7.2; length of mandible, M 15, F 13.3; angle to condyle, M 5.2, F 4; depth at coronoid, M 4.2, F 3.4; lower toothrow (including canine), M 8.5, F 8.1. Height of sagittal crest in male, 1.8, in female, 0.5.

Represented by 2 alcoholic specimens, type (M) and cotype (F), collected at Kinshasa, 6 miles from Leopoldville, lower Congo, December 22, 1914.

The only species with which the present form needs comparison is *Mops (Allomops) faradjius*, which differs from it in smaller size, less highly developed sagittal crest, less concave palate, and in certain other features indicated under *M. (Allomops) occipitalis*.

Mops angolensis niveiventer Cabrera and Ruxton, 1926

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B.M. 2 M, 4 F (En maison.)

Type. Adult female, obtained in Saint Joseph de Luluabourg, April 16th, 1924. Collector's no. 409.

General colour dark vandyke-brown; under surface entirely pure white; without any buffy or brownish tinge.

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Dimensions of type: -

Head and body 72 mm.; tail 34; forearm 45; hind foot 11; ear 21.

Skull: condylo-basal length 19.3; zygomatic breadth 13.4; breadth of brain-case 10.5; upper tooth-row from canine 7.8; lower tooth-row from canine 9.2.

Specimens of *M. angolensis* from different parts of Africa appear to differ constantly among themselves by the coloration of the ventral surface. Those coming from the Angola coast have pale reddish-white bellie; others from the Shire River (Portuguese East Africa) present also the lower parts strongly suffused either with brown or buff, while in specimens from Bagamoyo the belly is white only in the centre. These differences may be of subspecific significance; at least, we think it convenient to describe under a trinomial the form inhabiting the Lulua region, with its quite pure white underside.

In connection with *M. angolensis*, it is interesting to state that this specific name was given by Günther, and not by Peters as generally said. Peters described the typical Angolan bat in the 'Jornal' of the Lisbon Academy, vol. iii. 1870, p. 124, as

"*Nyctinomus*, nov. sp.," the name *angolensis* being originally published, and wrongly attributed to Peters, by Günther in the 'Zoological Record' for 1870 (dated October 1871), p. 8, with the quotation of Peters's description as a basis.

Mops angolensis orientis G.M. Allen and Loveridge, 1942

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6 M 4 F (M.C.Z. 38826-35) Kitaya, T. T. 3.iv.39.

Type. Museum of Comparative Zoölogy No. 38829, an adult male, skin and skull, from Kitaya, Rovuma River, southeastern Tanganyika Territory. Collected by Arthur Loveridge, April 3, 1939.

Description. One of the stout-bodied forms with white underside, slightly smaller than *Mops (Allomops) angolensis osborni* of the Great Lakes region, to which it is related, and with the upper surface faintly tinged with tawny ochraceous instead of being uniform drabby brown; skull slightly smaller and with less development of the sagittal crest.

Distribution of the fur as in *M. a. osborni*, the wing above naked, but with a narrow line of whitish hairs on the anterior and posterior sides of the humerus; hind legs practically naked as well as the posterior part of the rump and the anal region, where, however, minute scattered hairs are visible under a lens. Face, chin and ears blackish, with minute sparse hairs; toes with the usual longer stiff hairs as well as lateral hairs on outer edge of the first and fifth toes. Entire under surface of the body pure white to the edge of the membrane, with

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slight individual variation, so that in some specimens the sides of the neck are pale drab while in others this tint is more extensive, from the axilla back along the side halfway to the groin. Wings dusky brown, becoming whitish lateral to the forearm; this pale area varies individually but may extend to the tip of the third finger.

Measurements. The collector's measurements of the type are: head and body, 86 mm.; tail, 40; hind foot, 13; ear, 19; spread of wings, 340.

The cranial measurements of the skull of the type are: greatest length, 21.8 mm.; condylobasal length, 19.2; palatal length, 9.9; zygomatic width, 13.0; mastoid width, 11.5; width outside molars, 9.5; upper cheek teeth, 7.7.

In the adult-male skull the occiput is produced behind and squarely truncate, with a transverse angular crest, while the knife-like sagittal crest extends forward to the interorbital level. In the upper jaw the anterior premolar is minute and crowded into the outer angle between the canine and large premolar, which are in contact in their median line. In the lower jaw the anterior pair of incisors overlaps the posterior pair, and all four are bifid. In males the two lower premolars are of nearly equal height, but in females the anterior one is distinctly the shorter.

Remarks. The series is uniform in the decided tint of russet above instead of the dull drab of *osborni*, while the much clearer and more extensive white area of the lower side and the whitish wings, distinguish it further and altogether probably reflect the somewhat different climatic conditions of this part of the coast as compared with the Tana River and Lake region. The form, *Mops faradjius*, is a darker representative found in the northeastern Congo forest; probably all should be regarded as races of *M. angolensis*.

Habitat. These bats were roosting under the galvanized roof of the baraza house. I set up the net at 6.30 p.m.; the bats began emerging at 6.45 p.m. and by 7 p.m. I had removed ten of this species and one of the much smaller *C. limbatus*. Naturally their stomachs were empty, none of the females held embryos. The natives apply the same name *kiputiptiti* to them as for other small bats.

Mops angolensis wonderi Sanborn, 1936

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Type from Sotuba, 7 km. east of Bamako, French Sudan. No. 42138 Field Museum of Natural History. Adult female. Collected April 13, 1934, by Frank C. Wonder. Orig. No. 814.

Diagnosis. - Like *M. angolensis* but smaller and with lighter-colored under parts.

Color. - Type: upper parts dark brown, lighter on head and very strongly flecked with gray. A broad, almost hairless collar across shoulders. Below brownish gray on throat and chin; center of belly and sides of body from femur to groin white. Between these white areas, dark gray. Other specimens show more dark gray below and in one there is a rather spotted line of brown, but in all the sides of the body are white from femur to groin.

Skull. - Probably smaller than in *angolensis*, for which I can find no measurements, and shorter, with a narrower zygomatic width, and a much shorter lower tooth-row than in *M. a. niveiventer* Cabrera and Ruxton. Sagittal and lambdoid crests well developed and preorbital processes distinct. Anterior upper premolar minute.

Measurements. - Forearm 45.7 (44.8 - 48.5; 5 skins 44.8 - 47.8, 3 skeletons 46 - 47.5, one alc. 48.5). Skull: greatest length 20.9 (20.1 - 21.8); condylo-basal length 17.9 (17.9 - 18.8); palatal length 8.6 (8.6 - 8.8); interorbital width 4.5 (4.5 - 4.7);

zygomatic width 12.6 (12.4 - 13.1); mastoid width 11.8 (11.7 - 12.2); width of brain case 10.4 (10.3 - 10.5); width across canines 6 (6 - 6.5); across m^2 - m^2 , 8.7 (8.6 - 9.1); upper tooth-row c - m^3 , 7.3 (7.3 - 7.8); lower tooth-row c - m^3 , 8 (8 - 8.6); length of mandible 13.5 (13.5 - 14.6). Measurements of alcoholic specimen: forearm 48.5; second digit 47.3; third digit metacarpal 50.9, first phalanx 22.9, second phalanx 23.6, tip 12; fourth digit metacarpal 49, first phalanx 20.5, second phalanx 18.8; fifth digit metacarpal 33.5, first phalanx 13.2, second phalanx 6.4. Ear, from meatus 15, from crown 12.6. Tibia 17.1, foot 13.

Mops chitauensis J.Eric Hill, 1937

p. 3

TYPE. - No. 88116, Amer. Mus. Nat. Hist.; M adult, skin and skull; from Chitau, Angola, alt., 4930 ft. Secured by the Phipps-Bradley Expedition, Mr. Lee S. Bradley, collector, orig. no. 676, February 10, 1933.

This species is known only from the type specimen.

GENERAL CHARACTERS. - A blackish brown, grizzled species of *Mops*; skull with high lambdoidal crest. Uropatagium reaches to heel, wing membrane attached to lower third of tibia.

COLORATION. - Upperparts near fuscous-black, shading into black on the head, grizzled with whitish hairs and tips; bases of hairs paler. Underparts dirty whitish. Wings darker than fuscous, as are the ears. Upper arms and legs pale.

SKULL. - Short and broad; supraoccipital region produced more than usual for *Mops*, but sagittal crest poorly developed.

DENTITION. - Upper incisors closely approximated, separated from canines by a gap. P^2 minute, situated laterally to axis of tooth row; P^4 and canine in contact medially. M^3 about half size of M^1 and M^2 , with a V-pattern.

MEASUREMENTS. - Head and body, 75; tail, 41; forearm, 46.3; digit III, 47.0 X 19.4 X 19.9 X 8.4. Skull: greatest length, 23.3; basilar length, 17.6; palatilar length, 8.2; zygomatic breadth, 14.5; interorbital breadth, 4.5; mastoid breadth, 12.7; outside breadth across M^2 - M^2 , 9.7; maxillary alveoli, 9.1; crowns P^4 - M^3 , 7.0; width M^1 , 2.9.

Except for coloration, this species resembles the description of *Mops angolensis* (Peters). A specimen from Chitau, agreeing in color with Peters' description, collected by the Pulitzer Angola Expedition of the Carnegie Museum, has less extensive uropatagium and the wing membranes attach to the middle of the tibia. Cranially the occipital region is much less produced.

Mops congicus J.A. Allen, 1917

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Plate LV.

Type, No. 48893, F ad. (skin and skull), Medje, Belgian Congo, Sept. 8, 1910; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 966.

Above uniform deep chestnut-brown, the extreme base of hairs rufous; sides of back and lower back naked; below snuff-brown, passing into bistre laterally, the extreme tips of the hairs on the middle area buffy; a naked band at base of uropatagium, continuous with the wholly naked hind limbs; wing membranes naked, blackish brown above, a little lighter brown below; interfemoral membrane naked, dull brown above and below. Ears joined by a low membrane, which extends forward as a protuberance in front of the ears; ears thick and leathery, black, with a strong frontal keel; tragus minute, difficult to distinguish in dry skins; antitragus small, low, evenly rounded above, about 5 by 3 mm. Face and chin naked, dark brown; nostrils prominently projecting; upper lip heavily corrugated; thumb with a conspicuous basal pad. A very small patch of fur near outer base of humerus, and a dense patch on inner upper border of ear; outer base heavily furred.

Collectors' measurements of type: Total length, 140 mm.; head and body, 91; tail, 49; foot, 17; ear, 22. Type and 12 adult topotypes; Total length, 138 (131-14x); head and body, 93.4 (86-100); tail, 45.3 (41-49); foot, 15.8 (15-17); ear, 22 (20-24). Additional measurements from skins: Type, forearm, 55; third metacarpal, 54; thumb (with claw), 9.3; foot, 15.8; length of ear on outer border, 19; breadth of ear near upper border, 9.5. Forearm, type and 14 topotypes, 55.8 (54.7-57.4).

Skull narrow and deep, heavily ossified; a low continuous median crest from front of interorbital constriction, joining the rather heavy lambdoid; preorbital process weak; braincase with a strong transverse preoccipital depression; basisphenoid pits very deep and large, circular in outline. Upper incisors slightly separated and parallel; lower incisors, middle pair bifid, the outer slender, posterior to the middle pair; canines not specially modified; first upper premolar minute, in the toothrow, separating the canine from p^4 ; first lower premolar about equal in size to the second.

Skull of type: Total length, 25; condylobasal length, 22.4; zygomatic breadth, 15, mastoid breadth, 13.2; interorbital breadth, 4.7; maxillary breadth, 10.3; breadth

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at canines, 8.7; breadth of braincase, 11.7; depth of braincase at condyles, 9.5, at bullæ, 10.7; upper toothrow (with canine), 8.9; length of mandible, 19.4; angle to condyle, 5.6; depth at coronoid, 4.6.

Skull, type and 11 topotypes (1 male, 10 females), total length, 25 (24.4-25.6); zygomatic breadth, 15 (14.5-15.0).

Represented by 15 specimens (of which 3 are alcoholic), all taken at Medje, September 1910.

Chærephon congicus is easily distinguished by its size, and dark coloration. The length of the forearm averages 56 mm. in a series of 12 adults, the skull 25, zygomatic breadth 15. It greatly exceeds the size of *C. angolensis* from which it also differs in other important characters. The forearm in *angolensis* is given by Peters as 48 mm., and by Dobson as 47 mm., while the average in *congicus* is 55.8. It differs from *C. midas* in being smaller (forearm 55.8, in *midas* 60.4), with much smaller ears and much darker coloration.

Mops niangaræ J.A. Allen, 1917

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Type (and only specimen), No. 48901, M ad. (skin and skull), Niangara, northeastern Belgian Congo, Dec. 12, 1910; Herbert Long and James P. Chapin. American Museum Congo Expedition. Orig. No. 1313.

Similar to *M. congicus* but smaller and much paler.

Upperparts uniform cinnamon-rufous, extreme base of hairs much lighter; underparts tawny-brown, lighter and more yellowish on the pectoral area; ears, membranes and the broad naked space along sides of back and posterior margin of body dark brown; ears not joined at base, thick, and otherwise as in *M. congicus*. A. large crown-patch of lengthened hairs, similar in color and texture to the surrounding pelage, but much longer, forms a broad, low crest covering the interaural portion of the head.

Collectors' measurements: Total length; 125 mm.; head and body, 91; tail, 34; foot, 13; ear, 22. Forearm (from skin), 52; third metacarpal, 52; foot, 13; ear from crown, 8.6; from notch at antitragus, 21; transverse breadth, 13.2; tragus very small, breadth at base and height about equal.

Skull, total length, 23.2; condylobasal length, 21.6; zygomatic breadth, 14.4; breadth of braincase, 11.1; interorbital breadth, 4.0; maxillar breadth, 9.8; breadth at base of canines, 7.1; upper toothrow (with canine), 8.2; length of mandible, 16; depth at condyle, 5.4; at coronoid, 4.5; lower toothrow, 9.4.

Mops niangaræ is structurally closely similar to *M. congicus*, but is smaller and much paler, and the ears are not joined by a membrane. The forearm is 4 mm. shorter and the cranial measurements are correspondingly less.

This is the only species of molossid bat in the present collection, numbering 19 species, in which the ears are not joined across the forehead by a strongly developed band, if we regard the high convex lappet between the ears, but not distinctly joined to them, in *Chærephon* (*Lophomops*) *abæ* as

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morphologically a connecting ear-band. In the present species (represented by the single type specimen) there is not even a vestigial band, but the position it should occupy if present is outlined by a line of slightly lengthened dark brown hairs, giving the visual effect of a vestigial band till it is carefully examined and found not to be a slight ridge of membrane.

Mops osborni occidentalis Monard, 1939

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ALLEN, LANG and CHAPIN: Bats From the Belgian Congo. Bull. Amer. Museum, 37, 1917. p. 473 (sur *M. osborni*)

Collection: N^{os} 215 à 227, 275. 13 MM, une F, Mansoa; 494 à 498, un M, 4 FF, Contubo-el; 775 à 780 et 788 à 790, 4 MM et 5 FF, Pitche. En tout 29 exemplaires,

Le sous-genre *Allomops* a été créé par ALLEN sur 4 espèces du Congo belge. Il est caractérisé par la forme du crâne, qui, chez les vieux mâles, présente une crête lambdoïde extrêmement saillante et prolongée au-dessus du trou occipital. Les autres caractères indiqués par ALLEN sont soit ceux du genre *Mops*, soit d'ordre plutôt spécifique. Le sous-genre *Allomops*, plus heureux que son voisin, *Lophomops*, subsiste, quoique la forme caractéristique du crâne n'apparaisse nettement que chez les vieux mâles.

Une description détaillée de *Mops* (*Allomops*) *osborni* nous a permis de conclure avec certitude que les *Mops* ramenés de Guinée où l'espèce est très abondante et forme même une véritable plaie dans les maisons européennes de Mansoa, Bafatá, Contubo-el, Pitche et certainement d'autres lieux encore, sont extrêmement proches de cette espèce. Nous ne notons guère, comme différence, que la petitesse du tragus qui ne mesure dans nos exemplaires que 3 X 3 mm, tandis qu'il en mesure 4 X 4 dans *osborni* typique, et la plus grande dimension de la queue, ordinairement 40 mm dans nos exemplaires contre 37 dans ceux du Congo. En outre, le pelage est un peu différemment réparti.

Description. Pelage d'une couleur gris-noirâtre sur le dos; il débordé légèrement sur la membrane qui, dans cette espèce, ne part pas des côtés du corps, mais prend son insertion sur la face dorsale, de sorte que par dessous, existent deux poches dépourvues de poils entre le corps et la membrane, côté dorsal. En vue dorsale,

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le pelage s'étend jusqu'à la base de la queue et sur les fémurs. Du côté ventral, la couleur est plus variable: les côtés du corps sont foncés, de même qu'une bande pectorale. Mais ces plages sombres ont des développements très inégaux d'un individu à l'autre, si bien que la face ventrale peut se montrer presque entièrement blanche, ou entièrement foncée. La lèvre inférieure et le menton sont ordinairement noirâtres et tranchent parfois nettement sur la couleur de la gorge. Les membranes sont brunâtres par dessus, plus pâles par dessous: le propatagium, une plage s'étendant du radius et de l'humérus jusqu'au milieu du tibia et l'uropatagium sont blanchâtres. Une touffe de poils existe derrière le bandeau qui unit les deux oreilles; les pieds sont garnis de soies courbées, les lèvres de courts bâtonnets sensitifs, les deux glandes du menton d'un long poil chacune. Les membranes sont quasiment nues, surtout dessus; dessous l'uropatagium montre de fins poils blancs. Quelques longs poils naissent des fémurs, face dorsale, et se dirigent en arrière.

Le museau est très proéminent. dépassant de 7 mm la fente buccale; la lèvre supérieure est fortement plissée verticalement; les oreilles épaisses ont un antitragus séparé du pavillon par un sinus profond; l'antitragus lui-même est petit arrondi et mesure 3 X 3 mm.

Crâne. Il présente, très nettes, les caractéristiques du sous-genre: le développement postérieur de la crête lambdoïde. La dentition est celle de *osborni*, avec le caractère sexuel secondaire indiqué par ALLEN: la première prémolaire inférieure est aussi grande que la seconde chez les MM, plus petite chez les FF (ALLEN dit même: plus grande chez les MM, mais dans nos exemplaires il y a très peu de différence). La première prémolaire supérieure est très petite et logée dans l'angle externe formé par la canine et la seconde prémolaire qui sont contiguës.

[table removed - eds.]

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Distribution géographique et mœurs. *Mops osborni* est connu par un mâle et une femelle, venant de Kinshasa, à 6 miles de Léopoldville (Congo belge). Nous ne savons s'il a été retrouvé depuis. Sa présence en Guinée portugaise, à l'extrême Occident du continent est très remarquable et laisse prévoir d'autres découvertes dans les espaces intermédiaires.

En Guinée portugaise, l'espèce pullule dans les établissements européens, à Mansoa (local des Postes), à Bafatá (Immeuble de la Compagnie française), à Contubo-el, dans toutes les maisons de commerce, à Pêche, dans la maison du poste. Elle y forme de vastes colonies dont l'épouvantable odeur musquée et douceâtre se perçoit à distance, surtout à la tombée de la nuit, et rend les habitations malpropres et même inhabitables. Elle préfère les habitations européennes aux indigènes, car la présence de combles au-dessus des plafonds lui assure la tranquillité.

Son vol est aisé et rapide: elle pousse, lorsqu'elle est dérangée, de petits cris aigus.

[*Mops osborni occidentalis f. fulva*](#) Monard, 1939

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Collection: N^{os} 228 à 237, 5 MM et 5 FF, Mansoa; 781 un M, Pêche.

En tous points semblables par la forme, la distribution du pelage. les dimensions, à *Mops occidentalis*, mais en différant par la couleur du poil. Le dos est brun vif, le ventre varie du brun plus clair au jaune ou à l'orange, avec le milieu du ventre plus clair. La bande pectorale est parfois indiquée. La lèvre inférieure est noirâtre; les membranes, les oreilles et le museau sont de la couleur des exemplaires gris.

Les dimensions du crâne N° 781 sont données plus haut

[*Mops trevori*](#) J.A. Allen, 1917

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Plate XLVIII, Fig. 2.

Type, No. 49250, F ad. in alcohol, Faradje, Sept. 29, 1912; H. Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 1954.

Pelage short, soft and fine, with the grayish sheen in certain lights seen in many other molossid bats.

Upperparts cinnamon brown, with the usual naked transverse zone across front of shoulders; underparts pale brown, darker on the sides and lighter medially, with the hairs of the pectoral region conspicuously tipped with grayish white; caudal end of body not denuded, but with the projecting bristly hairs seen in allied forms. Membranes and wing bones above blackish and naked, except for a small patch of short brown fur near the center of the propatigium; below much lighter and the wing bones whitish; naked portion of face and ears, and the naked hind limbs and tail blackish, interfemoral membrane above blackish,

below much paler, as is also the ventral surface of the hind limbs. A low crest (the hairs about 5.5 mm. in length) arises from the back of the membrane connecting the ears and extends back over the front half of the interaural area and the basal half of the backs of the ears. The crest area is of the same color as the surrounding pelage.

Total length, 121.5 mm.; head and body, 82.3; tail, 39.2 (free portion 24); forearm, 53; third metacarpal, 53.5; thumb to base of pad, 8.4; tibia, 20; foot, 13.6; ear from notch, 20; breadth (near front border), 12; tragus minute, quadrate, 2 X 15; antitragus convex, broad at base and low, 6.5 X 4.

Skull, total length, 24.2; condylobasal length, 22; zygomatic breadth, 14.6; interorbital breadth, 5; mastoid breadth, 13.8; maxillar breadth, 10.4; breadth at base of canines, 7; upper toothrow (c-m³), 8.6; length of mandible, 15.8; angle to condyle; 5.2; depth at coronoid, 4.5; lower toothrow, 9.7.

Represented only by the type.

Mops trevori is of nearly the same size as *M. congicus*, but it is slightly smaller in all measurements, except that the ears are larger. The coloration is radically different, the upperparts in *congicus* being deep chestnut, and dull cinnamon brown in *trevori*, with still greater difference in the color of the underparts. It differs from *M. midas* in much smaller size, while the color of both upperparts and the ventral surface in the two species is as different as between *trevori* and *congicus*. In cranial characters *trevori* is too different from either *congicus* or *midas* to render comparison necessary.

Mormopterus setiger Peters, 1878

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(Taf. 1. Fig.2.)

Kopf sehr platt und breit. Schnauze oben flach; in der Mitte ein wenig vertieft, kahl, mit ganz kurzen Härchen bekleidet, welche die Haut nicht verdecken. Nasenlöcher queroval, um mehr als ihren doppelten Durchmesser von einander entfernt und unmittelbar unter dem scharfen Endrande der Schnauze stehend; zwischen ihnen eine wulstige gelappte Längsleiste. Die breite wulstige, aber nicht quergefaltete Oberlippe ist jederseits mit vier bis fünf Längsreihen kurzer stachelförmiger dicker Borsten bekleidet, zwischen denen sparsame feine kurze und längere Härchen hervortreten. Die Unterlippe zeigt noch kürzere und schwächere sparsame Borsten und Haare. Die wulstigen Augenlider der kleinen Augen sind ebenfalls mit sparsamen Härchen bekleidet, welche sich nach oben und vorn zu einem grösseren Haufen zusammendrängen. Die Ohren sind dreieckig abgerundet und stehen weit von einander ab; die Ohrklappe ist im ganzen viereckig, am verdickten Endrande sparsam behaart.

Die oberen Schneidezähne sind deutlich zweispitzig, mit äusserer kürzerer Spitze. Sonst zeigt das Gebiss keine auffallende Verschiedenheit von dem der anderen Arten: $3^{1/3} 2^{1/1} 1^{1/6} 1^{1/1} 3^{1/2} 3$.

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Der Körper erscheint plump und etwas platt. Kurze rostbraune, an der Basis etwas hellere Haare bedecken die obere Körperseite, die Halsseiten und die Seite der Brust und des Bauches; auch setzen sie sich noch auf die Basis der Lendenflughaut fort. Die Mitte der Brust und des Bauches ist von rostgelben noch kürzeren Haaren bekleidet, welche in der Analgegend die Haut durchsehen lassen. An der Kehle befindet sich eine Querfalte, welche in eine sackförmige Grube führt.

Der Schwanz ragt zur Hälfte aus der Schenkelflughaut hervor. Die langen Spornen sind sehnig. Die Gliedmaßen erscheinen im Verhältniss zu dem plumpen Körper kurz und gedungen. Die Oberseite des Vorderarms und eines Theils der Finger ist durch zugespitzte Hautwarzen ausgezeichnet. An der Basis der Daumensohle befindet sich eine grössere, an der Basis der Fusssohle eine kleinere rundliche Wulst. Der Daumen und die Zehen sind durch sparsame lange borstige Haare ausgezeichnet. Die Flughäute sind ziemlich derbe, dunkelbraun, die Lendenflughaut am Rande weiss.

Maße eines weiblichen Exemplars in Weingeist:

Meter
 Totallänge: 0.090
 Kopflänge: 0.0215
 Abstand der Ohren: 0.009
 Ohrhöhe: 0.017
 Vorderer Ohrtrand: 0.013
 Ohrbreite: 0.012
 Länge der Ohrklappe: 0.004
 Oberarm: 0.028
 Vorderarm: 0.035
 L.1.F. Mh. 1. Gl. 2.Gl.: 0.006
 L.2.F. - 0.0315; - 0.001: 0.0325
 L.3.F. - 0.034; - 0.014; 0.012; Kpl. 0.006
 L.4.F. - 0.033; - 0.013; - 0.010; - 0.002
 L.5.F. - 0.025; - 0.075; - 0.007; - 0.002
 Schwanz: 0.028
 Oberschenkel: 0.015
 Unterschenkel: 0.011
 Sporn: 0.017

Fuss: 0.008

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Aus Ndi (Taita).

Diese Art unterscheidet sich, abgesehen von ihrer viel plumperen Gestalt, den Lippenborsten, der warzigen Beschaffenheit der Oberseite der vorderen Extremität und der viereckigen Gestalt der Ohrklappe, sehr wesentlich von den anderen bisher bekannten Arten dieser Gattung durch die weit von einander abstehenden Ohren, welche bei jenen einander auf 2 bis 3 Millimeter genähert sind. In der Form der Ohren nähert sie sich mehr dem *M. jugularis* Ptrs., während *M. acetabulosus* Hermann (*D. natalensis* Smith) durch die zarte schlanke Gestalt und die sehr spitzen Ohren sehr verschieden erscheint.

[*Mormopterus Whitleyi*](#) Scharff, 1900

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Mormopterus Whitleyi

Three specimens of a bat, which I was unable to identify with hitherto published descriptions, were sent to me by Dr. J.C. Whitley from Benin City in West Africa. Before describing them I forwarded one of them to Mr. Oldfield Thomas, of the British Museum, who was good enough to confirm my opinion that it belonged to a new species. I have great pleasure therefore in naming the bat after its discoverer.

Dr. Whitley, and herewith give a description of its most salient external features.

Ears shorter than the head and approaching one another on the forehead to within 2 millim. Both the ears and their short tragus are rounded off above. The antitragus is well developed and separated posteriorly by a distinct notch. The extremity of the muzzle projects very much beyond the mandible, and the end of the nose is distinct from the upper lip. The nostrils open almost laterally. The upper lip is only very indistinctly folded and covered with thick bristles and fine hairs. On the lower lip the bristles are confined to the angle of the mouth, but the hairs extend all over it. The gular sac is small.

The dentition being $\frac{1}{3} \frac{1}{1} \frac{1}{2} \frac{3}{3}$, *M. Whitleyi* differs from all other species of the genus in the possession of four lower incisors instead of the usual six.

The fur is dark brown above, the bases of the hairs being pale. The underside is of a light reddish-yellow colour. Nearly half the tail projects beyond the wing-membranes, which reach to the ends of the tibiae.

The dimensions of the type, now in the British Museum (adult M in formalin), are: - Total length 81 millim.; head 22; body 30; tail 23; length of forearm 36, of lower leg 27.

The height of the ear is 15 millim., the width 10.

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Three other species of *Mormopterus* are known to science, viz., *M. albiventer*, *M. acetabulosus*, and *M. setiger*. The first of these is confined to Madagascar, the second has a wide range from South-east Africa to Madagascar and Mauritius, while the last has been taken in German East Africa.

M. Whitleyi differs from *M. albiventer* in being somewhat smaller, in having a well-defined antitragus in the ear-conch, and from all the three in having only four lower incisors instead of six.

M. Whitleyi differs from *M. acetabulosus* and *M. setiger* in having the ears close together on the forehead, in the more regularly triangulate shape of the ears, and in the squareness of the tragus.

Finally, *M. Whitleyi* differs from *M. setiger* in being smaller and in having the tragus rounded off above.

[*Myonycteris leptodon*](#) K. Andersen, 1908

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Diagnosis. - m^2 about twice the bulk of p^1 . Molariform teeth short and narrow. Forearm about 61-62 mm. Hab. Sierra Leone; Liberia.

Differential characters. - Skull similar to that of *M. wroughtoni*, but rostrum narrower in front (distance between inner bases of canines 3.3 mm., against 3.7-4 in *M. wroughtoni*)

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and *torquata*), coronoid process higher and much slenderer. All molariform teeth conspicuously shorter and narrower than in the two other species of the genus: m^1 , length 1.9 mm., against 2.2, breadth 1.3 mm., against 1.6-1.7; p_4 , length 2.4 mm., against 2.7-3, breadth 1.7 mm., against 1.8; m^2 relatively of the same size as in *M. wroughtoni*, less reduced than in *M.*

torquata. - Ears smaller than in the allied species, length from notch 14 mm., against 15-15.5 in *M. torquata* and 16.5-17.5 in *M. wroughtoni*; length of tibia and hind foot as in *M. wroughtoni*, but forearm, metacarpals, and phalanges somewhat shorter. Distribution and colour of fur as in the allied species.

Type. M ad. (skin and skull), Sierra Leone; presented by J. Hickman, Esq.; B.M. 91.2.13.1. - The species is represented in the Leyden Museum from Liberia.

Myonycteris relict Bergmans, 1980

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Myonycteris relict n. sp. (figs. 1-4)

Holotype: adult male, skin and skull, collected 30-VII-1978 by Mr. Robert N. Kyongo (field number RK 594), in the Shimba Hills, southeast Kenya (RMNH 27909). "Caught over water, about 1.20 m high, at about 10 p.m. local time, Mukanda River, Lukore area, Shimba Hills; along the river: big thorn trees and fig trees" (R. N. Kyongo in litt.).

Paratypes: adult male and adult female, in alcohol, skulls separate (and damaged to some extent), collected in January 1900 (collector unknown) at Ambangulu (5°5'S, 38°26' ?), Usambara Mountains, northeast Tanzania (ZMB 54936 and 54937, respectively

[Table removed]

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Diagnosis. - *Myonycteris relict* is a large member of its genus (forearm lengths 69.3 - 75.1 mm and greatest skull lengths 36.4 - 39.2 mm in the three specimens known), with deflected facial axis, relatively slender rostrum, dental formula $\frac{2.1.3.2}{2.1.3.2}$, elongate teeth, and reduced M^2 .

Description of holotype (notes concerning soft parts checked on paratypes). - Snout relatively long, with linear dorsal profile. Posterior part of head rather domed. Nose skin brown, nostrils pointed forward and outward, separated by a shallow but distinct groove. Chin pad narrow, notched at its proximal end in the median plane, with small and narrow offshoots along lower lips. Upper and lower lips quite thick, upper with a row of small pointed papillae on inner margin, extending backward from close behind the nose, lower with same from about halfway. Tongue with long and narrow field of toothed papillae on anterior part. Ears moderately long, anterior margin thickened, posterior margin convex with slight emargination just below tip; tip rather distinctly pointed; conchs darkish brown, with about ten transverse folds; antitragal lobe moderate, angular. Top and sides of head furred with soft woolly hair, very short and sparse near nose and somewhat longer and very dense backward from in between and under eyes, near Ecru-Drab, tipped with near Verona Brown on snout, forehead and cheeks, and with orange-brown on top of head. Same light short hair in zone along lower lip (centre of chin furred like ventral body side; see below). Sides of rostrum, a narrow zone around eyes, and lower lips furnished with additional, mostly dark brown, tactile hairs: many shorter ones, a few longer ones.

Dorsally, body densely furred with bicoloured hair (6-10 mm long), Light Cinnamon-Drab with Brüssels Brown to Tawny tips, the tips dominating the overall - light reddish brown - colour impression. Hair of same colours extending on arm, down to (and much shorter on) first half of forearm, on small areas of patagium in axillar region and below elbow, on hind legs almost down to ankles, and on adjoining parts of uropatagium, surpassing its caudal margin at either side of tail.

Ventrally, body covered with dense, short hair (length 3.5 - 5 mm), Fawn Colour with slightly darker bases; also on upper arm and proximal half of forearm, along upper legs to slightly below knee, and on proximal part of uropatagium. Holotype (and male paratype) with throat ruff of relatively coarse hair; foremost point of ruff on chin between mouth corners; ruff gradually widening to greatest width at throat level, with 'crowns' on shoulders, and ending on breast, about on level with antebrachial membrane insertion; ruff hairs with short thin bases and thick shafts, wavy and growing

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in clusters of about 12 in centre of ruff, but less coarse, less wavy and less obviously clustered towards ruff margins; ruff hairs in holotype from about Light Ochraceous Buff to Tawny Olive, with darkest hairs on chin. (Ruff colour in paratype ZMB 54936 very light, but possibly somewhat bleached.) Female paratype without ruff. Propatagium near furred part of arm, and endopatagium between furred part of arm and body, thinly covered with Fawn Coloured or in places weakly reddish brown hairs.

Wings rather large (see table 1 for proportions), darkish brown. Endopatagium attached near middle of first phalanx of second toe. Toes 'webbed' to middle of first phalanges. Calcar length about 7 mm. Caudal margin of stretched uropatagium almost linear between end of calcar and a point on the dorsal side of the tail at about 3 mm from its external base. Soft palatum not preserved in the studied specimens.

Skull (figs. 1 - 4) lightly built. Facial axis deflected. Alveolar line, if projected backward, passing slightly above upper edge of occipital condyle. Premaxillae slender, fused anteriorly (only clearly co-ossified in specimen ZMB 54937). Rostrum relatively long, with nearly linear dorsal profile. Lateral pair of frontal sinuses quite strongly inflated, medial pair somewhat less. Behind these, a third pair of externally slightly inflated interorbital sinuses, roundish in outline, in between postorbital foramina and not extending beyond postorbital constriction, well visible in holotype and male paratype, indistinct in female paratype. Postorbital processes well developed, slender. Postorbital foramina small, distinct. Braincase rather domed, inflated behind interorbital sinuses, somewhat constricted at level of posterior zygomatic arch insertions, elongated posteriorly. Temporal ridges distinct

but weak, not fusing into a sagittal crest. Supraoccipital crest low. Zygomatic arches slender, curving down about as low as the alveolar line when viewed from the side. Orbit large. Tympanic bullae of moderate size (greatest length in specimen ZMB 54936 4.6 mm). Bony palate concave, with greatest depth at level of M^1 ; postdental margins rather straight and weakly converging backward.

Mandible delicate, with narrow rami and low coronoid processes; angle between alveolar line and anterior margin of coronoid process about 145° .

Upper incisors minute, with tips curved backward, crowded (distance I^1 - I^1 slightly smaller than I^1 - I^2); I^2 at about 1 mm from canine. Upper canine short (reconstructed height from cingulum in specimen RMNH 27909 3.5 mm), with only weakly recurved tips; basal outline rectangularly ovate; antero-internal face flat or slightly concave, without longitudinal groove; anterior keel distinct, not sharp; labial face quite evenly rounded; posterior face sharply defined, narrow, concave; lingual face concave except in the middle,

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its orientation inward and backward, with trace of horizontal shelf at its base. Anterior upper premolar (P^1) small, about 2 to 3 times the bulk of an upper incisor, wider than long, with rounded anterior and flattish posterior face and an obtuse cusp. Second upper premolar (P^2) oblong, basal outline more or less triangular, with a pointed (outer) cusp just before the middle; lateral (outer) profile sub-triangular; with very weakly rounded labial face, obtuse anterior keel, rather flat antero-internal face and well-defined, slightly concave lingual (postero-internal) face; greatest width at level of keel separating antero-internal and postero-internal face; orientation of latter face either parallel with, or converging backward toward, labial face (thence the variable tooth outline). Third upper molar (P^4) oblong, subrectangular, with a pointed (outer) cusp before the middle; rather obtuse anterior (outer) and posterior outer ridges from cusp tip to cingulum; a weak commissure from tip to lingual side; antero-internal and postero-internal faces concave. First upper molar (M^1) essentially as P^4 , but smaller and lower, with less rounded anterior side and strongly reduced cusp. Second upper molar (M^2) small, ovate to triangularly ovate in outline, greatest width in front, even less differentiated than M^1 , at some distance from posterior margin of anterior zygomatic arch insertion.

Lower incisors small, outer ones (I^2) slightly larger than inner ones (I^1); tips bicuspid, inner cusp the larger in both I^1 and I^2 ; distances I^1 - I^1 and I^1 - I^2 minimal, distance from I^2 to lower canine same or slightly larger. Lower canine short (height from cingulum in holotype specimen 1.8 mm); basal outline broadly ovate, with rather straight posterior side; tip hardly recurved, pointing outward; no distinct keels; antero-internal and labial faces rounded, lingual (postero-internal) face straight or concave, with a narrow shelf at its base. First lower premolar (P_1) relatively large; outline squarish or more rounded; cusp rather obtuse, separating rounded vertical antero-external face from sloping postero-internal face; latter ending almost horizontally. Second lower premolar (P_2) somewhat shorter than lower canine, with pointed cusp well in front of the middle; basal outline ovate (with truncated posterior side in holotype); labial face rather straight or weakly rounded; no anterior keel; antero-internal face rounded; posterior face somewhat concave, sloping, ending almost horizontally; postero-external longitudinal ridge distinct, obtuse. Third lower premolar (P_4) largest of all teeth; basal outline oblong, with about parallel labial and lingual sides and rounded anterior and posterior sides; the single cusp well in front of the middle, with ill-defined anterior keel and blunt postero-external ridge running from it; both labial and lingual faces weakly rounded and gradually passing into essentially round and sloping anterior face; postero-internal face an even,

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weakly concave slope without a clear horizontal posterior basal part. First lower molar (M_1) long, narrow, low; labial and lingual faces parallel; anterior and posterior faces somewhat truncated; outer longitudinal ridge higher than inner one (and with traces of antero-median cusp and commissure from cusp to lingual ridge in holotype); upper surface concave. Second lower molar (M_2) rectangularly ovate in outline, reduced, low, undifferentiated; surface flat or weakly concave.

Salient aspects of upper tooth rows when viewed from below (fig. 3) are the relatively heavy third premolars (P^4) and the more or less coulisse-like arrangement of second and third premolars and first molar, with longitudinal axes pointing forward - in the line of the row - but also somewhat inward; and of lower tooth rows when viewed from above (fig. 4) the large third premolars (P_4) and the absence of M_3 .

Measurements: table 1. The only known weight is that of the adult male holotype: 48 g.

Myonycteris wroughtoni K. Andersen, 1908

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Diagnosis. - m^2 about twice the bulk of p^1 . Skull and teeth heavy. Forearm about 65-67 mm. Hab. Welle District, N.E. Congo.

Differential characters. - Skull larger and more heavily built than in *M. torquata**: total length 34 mm., against 31-31.8; rostrum longer and broader: from front of orbit to tip of nasals 10.5-10.7, against 9-9.2, lachrymal width (across lower edges of lachrymal foramina) 9.8-10 mm., against 8.8; length of combined orbital cavity and temporal fossa, measured on underside of skull from front of glenoid fossa to back of maxillary process of zygomatic arch, 10.7-11 mm., against 9-9.8; frontal region broader: interorbital breadth 6 mm., against 5-5.2. Cheek-teeth on the whole slightly broader; m^2 less reduced in size, being about twice the bulk of p^1 (in *M. torquata* subequal to p^1), actual length of tooth in two specimens 0.9 mm., against 0.5-0.7 in two *M. torquata*, breadth 0.6-0.7, against 0.4-0.5. - Wings, tibia, and foot conspicuously longer; forearm 65-67 mm. (57-60.5 in *M. torquata*), third metacarpal 44.5-45 (36-39), lower leg 25-25.5 (22), foot with claws 17-19 (14). Distribution and colour of fur as in the allied species.

Type. M ad. (skin and skull), River Likandi, Welle District, 18th April, 1906; Alexander-Gosling Expedition; B.M. 7.7.8.25. Two specimens examined.

Named in honour of Mr. R. C. Wroughton, who assisted in working out the mammals collected during the Alexander-Gosling Expedition.

[*Myopterus*](#) E. Geoffroy Saint-Hilaire, 1818

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MYOPTÈRE. *Myopterus*.

Dents incisives ²₂; canines ¹₁; molaires ⁴₅-⁴₅

Nez simple; chanfrein méplat.

Oreilles larges, isolées et latérales ; oreillon intérieur.

Membrane interfémorale moyenne.

Queue longue, à demi enveloppée, et libre au-delà.

OBS. On n'en connoît que la seule espèce publiée par Daubenton, sous le nom de rat-volant.

[*Myopterus albatus*](#) Thomas, 1915

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A large whitish and white-winged species with the upper body-colour arranged in a lineated pattern.

Size large, the forearm about half as long again as in *M. whiteyi*, and rather larger than that of *M. daubentoni* would be if in the same relative proportion to the skull (that is to say, about twice the length) as in *M. whiteyi*. Fur soft and fine, hairs of shoulders about $4\frac{1}{2}$ mm, in length, of lower back $3\frac{1}{2}$. Fur not extending at all on the membranes, and stopping on the lower back a little way short of the tail, just as in *M. whiteyi*. General ground-colour above drabby brown ("dusky drab"), the hairs white at their bases, brown terminally; but on the median area of the nape, from occiput to withers, and along two broad lines running down the back on each side from the withers, the white extends nearly to the tips of the hairs, so as to show through above, and to form broad whitish lines, the brown along the side of the body and that in the middle line from the withers backwards, therefore, appearing as three broad brown longitudinal lines separated by whitish. Fur of whole of under surface pure creamy white, sharply contrasted on the sides of the neck with the brown of the upper surface. Wing-membranes white throughout, except that by the side of the body which is spotted with brown granules, just as in *M. whiteyi*; interfemoral membrane brown. Upper surface of forearms, digits, legs, and feet also brown.

Ears apparently similar in structure to those of *M. whiteyi*, separated in the middle line, their inner keels low and little developed. Tragus short and broad. Nasal septum without a mesial ridge, clothed with fine whitish hairs. Edges of lips with a close mixture of spoon-shaped and normal bristle-hairs. Lips practically without wrinkles. Wings to the distal end of the tibiae.

Dimensions (measured on the skin): -

Forearm 55 mm.

Head and body (probably stretched) 84; tail 40; ear (dry) 19; third finger, metacarpal 54.5, first phalanx: 19.5, second phalanx 19.5; fifth finger, metacarpal 35, first phalanx: 15; hind foot (c. u.) 13.

Hab. R. Welle. Collected by M. Hutereau.

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Original number 17. Congo Museum, no. 2911.

The coloration of this beautiful bat is quite unique, but of course it assimilates, in the brown upper and white lower surface, with that of *M. daubentoni* and *whiteyi*.]

[*Myotis blythii punicus*](#) Felten, 1977

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Holotypus: SMF 44104, M ad.; Höhle El Haouaria, Cap Bon, Tunesien; NAGEL, SCHUBERTH & VESMANIS leg. 25.III.1971. S + A.

Paratypen: SMF 18946-52, 18955-64, 22214, 22217-20, 43386-92, 43961-68, 44097-103, 44618; 6 M, 39 F vom gleichen Fundort.

Weiteres Material: Die auf S. 31 angeführten Exemplare von anderen tunesischen Fundorten, aus Algerien und Marokko.

Diagnose: In den Schädelmaßen (Mittelwerte; s. Tab. 14) meist größer als *omari* und *oxygnathus*. Mittelwerte von Länge, Breite und Kronenfläche (= Länge X Breite) des M² deutlich größer als bei *omari* und *oxygnathus*. Jedoch Breite C¹-C¹ (Alv) im Mittel so groß wie bei *oxygnathus* und deutlich kleiner als bei *omari*.

Myotis Bocagei cupreolus Thomas, 1904

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Essential characters as in true *M. Bocagei*, but the colour much darker, owing to only the terminal millimetre instead of 2 - 3 mm. of the dorsal hairs being reddish; the reddish is also of a more coppery tone. A blackish patch at the base of each humerus. Under surface dark smoky brown, the hairs being dark smoky with brownish tips; inguinal region not or scarcely lighter. In true *Bocagei* the under surface is pale buffy brown.

Dimensions of the type: -

Forearm 39 mm.

"Head and body 60"; "tail 40" ; "ear 15."

Skull: greatest length 15.

Hab. Efulen, Bulu Country, Cameroons.

Type. Male. B.M. no. 3.2.4.6. Collected 14th August, 1901, by Mr. G. L. Bates.

Myotis Goudoti anjouanensis Dorst, 1960

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Les Vespertilionidés du genre *Myotis* sont représentés dans la région malgache par une espèce assez bien différenciée, *Myotis Goudoti* (A. Smith), qui habite Madagascar, probablement la partie orientale comme la partie occidentale de l'île, et vraisemblablement aussi le groupe des Comores, du moins Anjouan où elle a été collectée par Humblot.

Nous avons eu l'occasion d'étudier de longues séries de *Myotis Goudoti* provenant de diverses localités de Madagascar et de les comparer à de nombreux spécimens rapportés d'Anjouan par Humblot. De cet examen, il résulte que si les populations d'Anjouan appartiennent manifestement à l'espèce malgache, comme l'attestent notamment les similitudes dans la morphologie dentaire, elles se différencient cependant suffisamment de celles de Madagascar pour être distinguées subspécifiquement. Il existe en effet des différences constantes dans la taille et dans la coloration des parties supérieures.

Les différences de taille apparaissent avec une netteté particulière dans les longueurs de l'avant-bras. Les individus malgaches, provenant de plusieurs localités, ont en effet des avant-bras dont la longueur varie entre 36 et 40,5 mm avec une fréquence maximum de 38 mm environ, tandis que leurs homologues d'Anjouan ont des avant-bras dont la longueur varie entre 42,5 et 45, avec une fréquence maximum de 44 mm environ. Des différences du même ordre se retrouvent dans les autres mesures, notamment la longueur des phalanges et celle de l'oreille. Les dents, du même type chez les deux formes, sont nettement plus fortes chez les spécimens provenant d'Anjouan, leur robustesse se traduisant notamment par une plus grande largeur, surtout apparente quant aux molaires (M¹ et M³).

Nous avons comparé par ailleurs ces caractères aux descriptions originelles concernant les populations malgaches: cette espèce a été en effet décrite sous trois noms différents : *Vespertilio Goudoti* A. Smith (1834), *Vesp. madagascariensis* Tomes (1858) et *Vesp. sylvicola* A. Grandidier (1870), ces deux derniers noms étant synonymes du premier. Or les dimensions de l'avant-bras que

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donnent A. Smith (38 mm) et Tomes (38,5 mm), (Grandidier ne donne pas les dimensions de l'avant-bras de son type, mais les autres mensurations montrent clairement qu'il s'agit d'un spécimen de petite taille), concordent parfaitement avec la moyenne de nos spécimens malgaches et se séparent par contre nettement de ceux provenant d'Anjouan.

Par ailleurs la coloration des parties supérieures paraît d'une manière générale plus foncée chez les spécimens d'Anjouan. Leur pelage est en effet roux foncé alors qu'il est plutôt roux clair chez les spécimens malgaches. Nous proposons en conséquence le nom de

Myotis Goudoti anjouanensis subsp. nov.

pour les populations d'Anjouan.

Type : 1 ? rapporté par M. Humblot, d'Anjouan, conservé dans les collections du Muséum de Paris, CG. 1886-1536.

Il existe donc deux sous-espèces de *Myotis Goudoti* dont les diagnoses sont les suivantes:

- *Myotis G. Goudoti* (A. Smith). Taille relativement faible; avant-bras variant de 36 à 40,5 mm, avec une fréquence maximum de 38 mm. Parties supérieures brun roux clair. Madagascar.

- *Myotis G. anjouanensis* subsp. nov.

Taille relativement forte; avant-bras variant de 42 à 45 mm, avec une fréquence maximum de 44 mm environ. Parties supérieures brun roux foncé. Anjouan.

Nous ne disposons pas de spécimens provenant d'autres parties de l'archipel des Comores. Il est cependant possible que cette espèce existe dans d'autres îles.

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The vespertilionids of the genus *Myotis* are represented in the Malagasy region by a fairly well differentiated species, *Myotis Goudoti* (A. Smith), which inhabits Madagascar, probably the eastern part as well as the western part of the island, and probably also the Comoros group, at least Anjouan where it was collected by Humblot.

We had the opportunity to study long series of *Myotis Goudoti* from various localities of Madagascar and to compare them with many specimens brought back from Anjouan by Humblot. From this examination, it follows that if the populations of Anjouan clearly belong to the Madagascan species, as evidenced in particular by the similarities in dental morphology, they nevertheless differ sufficiently from those of Madagascar to be distinguished subspecifically. There are in fact constant differences in the size and coloring of the upper parts.

The differences in size appear with particular clarity in the lengths of the forearm. Malagasy individuals, coming from several localities, have forearms whose length varies between 36 and 40.5 mm with a maximum frequency of around 38 mm, while their counterparts in Anjouan have forearms whose length varies between 42.5 and 45, with a maximum frequency of approximately 44 mm. Similar differences are found in other measurements, including the length of the phalanges and that of the ear. The teeth, of the same type in the two forms, are clearly stronger in the specimens coming from Anjouan, their robustness being reflected in particular by a greater width, especially apparent as regards the molars (M^1 and M^3).

We have also compared these characters with the original descriptions concerning the Malagasy populations: this species has indeed been described under three different names: *Vespertilio Goudoti* A. Smith (1834), *Vesp. madagascariensis* Tomes (1858) and *Vesp. sylvicola* A. Grandidier (1870), these last two names being synonymous with the first. Now the dimensions of the forearm that

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give A. Smith (38 mm) and Tomes (38.5 mm), (Grandidier does not give the dimensions of his type forearm, but the other measurements clearly show that it is a specimen small), agree perfectly with the average of our Malagasy specimens and on the other hand clearly separate from those from Anjouan.

In addition, the coloring of the upper parts generally appears darker in specimens from Anjouan. Their coat is indeed dark red while it is rather light red in Madagascan specimens. We therefore propose the name of

Myotis Goudoti anjouanensis subsp. Nov.

for the populations of Anjouan.

Type: 1 ♂ reported by M. Humblot, from Anjouan, kept in the collections of the Muséum de Paris, CG. 1886-1536.

There are therefore two subspecies of *Myotis Goudoti* whose diagnoses are as follows:

- *Myotis G. Goudoti* (A. Smith). Relatively small size; forearm varying from 36 to 40.5 mm, with a maximum frequency of 38 mm. Light reddish brown upperparts. Madagascar.

- *Myotis G. anjouanensis* subsp. Nov.

Relatively large size; forearm varying from 42 to 45 mm, with a maximum frequency of approximately 44 mm. Dark reddish brown upperparts. Anjouan.

We do not have specimens from other parts of the Comoros archipelago. However, it is possible that this species exists on other islands.

Myotis Hildegardae Thomas, 1904

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A beautiful and brightly coloured species allied to *M. Bocagei*.

Size medium. Ears small, narrow; inner margin evenly convex, tip very narrowly rounded, outer margin concave above, convex below, a marked angular antitragal lobule at the outer base thickly covered with fur. Tragus rather short, its inner

margin slightly but evenly convex, its greatest breadth opposite the lower third of its inner margin, whence it slopes evenly to the narrow but not sharply pointed tip; basal lobe large, rounded. Feet large; wings to the metatarsi; calcars long, reaching nearly three-fourths towards the tip of the tail and ending in a distinct lobule.

Fur soft, thick and fine; hairs of back about 5 - 6 mm. in length. Wing-membranes naked, except for a few hairs on the under surface between the humeri and the flanks. Interfemoral furry above at the base, a narrow band passing outwards behind the legs nearly halfway down the tibiae. Top of toes hairy.

General colour of upper surface bright "tawny-ochraceous," the head rather paler than the back. Individually the hairs are blackish brown for about 2 mm. at their bases, then pale tawny, darkening to their tips. Below the general colour is "pinkish-buff," the hairs blackish at their bases. Membranes dark throughout, contrasting strikingly with the bright colour of the body.

Skull considerably larger than in *M. Bocagei*, broader and lower than in *M. Goudoti*. Small upper premolars in the tooth-row, subequal in horizontal section, and less unequal in height than usual.

Dimensions of the type (measured in skin): -

Forearm 37 mm.

Head and body (c.) 53; tail 37; ear (dry) 13; tragus on inner edge (dry) 4.6; thumb clear of membrane 5; third finger, metacarpus 35, 1st phalanx 15.5, 2nd phalanx 10.7; fifth finger 53; tibia 17; foot from back of calcar (c. u.) 9.8; calcar 17.

Skull: greatest length 15.2; basal length 11.3; breadth of brain-case 8; front of canine to back of m^3 5.7.

Hab. Fort Hall, Kenya District. Alt. 4000 feet.

Type. Male. B.M. no. 3.3.2.2. Original number 115. Collected 17th Oct., 1902, by Mrs. Hinde. Two specimens.

This very beautiful bat I have much pleasure in naming in honour of its discoverer Mrs. Hildegard Hinde, to whom

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the British Museum is indebted for so many interesting Chiroptera and Rodentia.

M. Hildegardæ is readily distinguishable from any of its allies by its striking coloration, as it is far brighter in tone than either *M. Bocagei* or *M. Goudoti*, the species most similar to it.

Myotis morrisi Hill, 1971

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(Plates i, 2 (a, b))

SPECIMEN. HOLOTYPE. B.M. 70.488. Adult female. A flat, card-mounted skin with skull; collector's number A107. "Forward Base Three", mouth of Didessa River, Blue Nile Gorge. 10° 05' N, 35° 38' E, alt. c. 1,000 m. 28 August 1968.

OTHER MATERIAL: none.

TAXONOMIC NOTES. DIAGNOSIS. Similar to *Myotis tricolor* (Temminck, 1832) of eastern Africa but differing from this species in its generally more orange dorsal coloration; unicolored and not bicolored ventral pelage; elongate, narrower skull with uninflated braincase and supraorbital region, the braincase shorter, more globular and markedly narrower than in *M. tricolor*, the rostrum proportionately longer and less broadened than in that species. Dentition less massive than in *M. tricolor*; second upper premolar (pm^3) relatively larger and incompletely intruded from the toothrow; second lower premolar (pm_2) relatively larger, not compressed between pm_2 and pm_4 ; posterior upper premolar (pm^4) with narrower lingual shelf, separated from the lingual shelf of m^1 by a wider interspace.

DESCRIPTION. Of moderate size (length of forearm 45.4) for the genus; anterior margin of ear smoothly convex, posterior margin concavely emarginated in its distal half, the proximal half convex. Tragus long, its length equal to one half of the length of the ear, tapered, with slender, rounded tip directed slightly posteriorly; anterior margin faintly convex, especially distally, posterior margin concave distally, convex proximally, slightly serrated, a small rounded lobe at base beneath an acute, angular emargination just below widest point of tragus. Wing inserted at base of first toe; calcar strongly developed, extending along almost one half of the posterior margin of the tail membrane; no obvious post-calcareal lobe. Pelage woolly, dorsally overall orange brown, individual hairs tricolored, the basal quarter blackish brown, most of remainder creamy white, hairs tipped terminally with bright orange brown. Dorsal pelage extending narrowly on to the wing membrane and on to the tail membrane for nearly one half of its width. Ventral pelage unicolorous dull creamy white, tinged faintly with brown on chin and flanks. Wing membrane generally black but antebrachium and endopatagium pale orange yellow, a narrow band of the same colour extending across the membrane immediately behind the forearm; anterior edge of membrane and area between first and second metacarpals and tail membrane similarly coloured. Tibia flanked by a narrow band of orange hairs on wing membrane and a wider band of similar hairs on tail membrane. Toes with a sparse covering of long orange hairs.

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Skull (Plates 1, 2(a)) elongate, the braincase not especially inflated; rostrum narrow, the supraorbital region not expanded, the supraorbital ridge forming an uninterrupted curve; a shallow, narrow median rostral depression. Narial emargination narrow, V-shaped posteriorly, its apex rounded. Palate long, narrow, anterior palatal emargination rounded posteriorly, extending almost to a line joining the centres of the canines; narrow post-palatal extension; shallow basioccipital pits. Inner upper incisor (i_2) longer than wide, bicuspid, with strong angular anterior cusp; smaller posterior cusp extending for two thirds of the height of the anterior cusp. Outer upper incisor (i_3) wider than long, bicuspid, closely appressed to inner tooth, cusps lying transversely to line of toothrow. Outer cusp the larger, rising from a narrow cingulum shelf to a height equal to that of the posterior cusp of inner tooth; supported internally by a smaller secondary cusp for two thirds of its height, the tooth somewhat hollowed internally, separated from the canine by a short diastema. Anterior upper premolar (pm^2) about as high as i_3 , a little larger at base, in contact with canine, a slender pointed cusp with wide, strong cingulum. Second upper premolar (pm^3) rising slightly above cingulum of pm^2 , slightly less than one half its basal area, visible externally, a simple cusp rising from a narrow cingulum, slightly intruded from row but separating pm^2 and the posterior upper premolar (pm^4), in contact with these teeth. Posterior upper premolar (pm^4) with strong cusp and short, narrow lingual shelf with a wide interspace, separating it from the lingual shelf of the first upper molar (m^1). Lower incisors imbricated, first (i_1) and second (i_2) with four cusps, outer cusp of i_1 incipient, of i_2 well developed but lower than inner cusps. Lower premolars not especially compressed: second (pm_3) rather more than one half the height of anterior tooth (pm_2) and approximately one half its basal area, in contact with pm_2 and posterior lower premolar (pm_4), but not compressed or displaced, its length and width equal. The measurements of the new species are compared with those of *M. tricolor* in Table 4.

REMARKS. In some respects *M. morrisi* resembles *M. bocagei* but is larger, has unicolored and not bicolored ventral pelage, a relatively longer rostrum lacking supraorbital inflation, narrower narial and anterior palatal emarginations and relatively narrower palate. Dentally the two species are closely similar but the teeth of *M. morrisi* are more generally massive than are those of *M. bocagei* and *M. morrisi* lacks a protoconule on the anterior ridge of the first and the second upper molars (described in *M. bocagei* by Harrison (1964b: 135).)

Tate (1941 : 539) reviewed the subgenera of *Myotis* and (p. 552) referred *tricolor* to the subgenus *Selysius*. However, both *tricolor* and *morrisi* approach the subgenus *Chrysopteron* in dichromatic wing pattern, although this is less evident in *tricolor*, and in the presence of four lobes on the inner (i_1) and second (i_2) lower incisors, but retain a relatively high braincase and concave frontal profile. I have much pleasure in associating with this new species the name of my co-author, Dr. Pat Morris, of Royal Holloway College, University of London, in recognition of his many services to the study of the Ethiopian fauna while with the Great Abbai Expedition.

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[table removed - eds.]

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COLLECTION AND FIELD NOTES. The specimen was caught by Messrs. Yalden, Largen and King of the Great Abbai Expedition, using a hand-held mist net. The bat was flying after dark over the river Nile, near its north bank, at a height of about one metre. The surrounding habitat comprises mainly maize plots and thick bush. The specimen was photographed whilst still alive (Plate 2 (b)).

N[lycteris] albiventer Wagner, 1840

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Die weißbäuchige Hohnase.

N. supra dilute brunea, pilis basi alibidis; subtus pulchre alba, pilis basi brevissime griseis.

Aus dem frankfurter Museum hat schon für längerer Zeit das unferige vier Exemplare einer Hohnase erhalten, die, den äußern Formverhältnissen nach, mit den Beschreibungen von *N. thebaica* übereinkommen, so daß man sie unbedenklich dieser Art zuzählen könnte, wenn nicht in der Färbung ein Unterschied sich ergäbe. Die Oberseite ist nämlich zwar ebenfalls lichtbraun, wie bei jener Art, aber die Unterseite ist nicht aschgrau, sondern rein weiß, wobei die hellbraune Farbe der Oberseite nur die Spitzen einnimmt, während die viel längere Wurzelhälfte graulichweiß ist, was am Hinterhals am hellsten ausfällt. Auf der Unterseite ist bloß der kurze Wurzeltheil etwas graulich, so daß die graue Farbe auf der Oberfläche gar nicht sichtlich wird; an dem einen Exemplar sind aber die Haare des Unterleibs, fast ihrer ganzen Länge nach, durchaus weiß. Der Anfang des Oberarms und der Flügelansaß am Leibe ist oben braun, unten weiß behaart. Backenzähne sind jederseits $\frac{4}{4}$ vorhanden. Ob ich diese weißbäuchige Hohnase mit Fug und Recht von der graubäuchigen *N. thebaica* als selbständige Art trennen darf, muß ich der Beurtheilung Deren überlassen, welche Gelegenheit haben, die Geoffroy'sche Art zu vergleichen.

N. thebaica; *N. albiventer*; *N. discolor*; *N. hispida*

Länge des Körpers in gerader Linie: 1" 10"; 1" 10"; 2" 2"; 1" 5"

----- des Schwanzes: 1 11; 1 9; 1 8; 1 10; 1 2

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----- des Kopfes: 0 10; 0 9; 0 10; 0 5

Breite der Nasendeckel: 0 $\frac{1}{4}$; 0 $\frac{3}{4}$

Länge der Ohren: 0 11; 1 $\frac{1}{2}$; 1 0; 1 $\frac{1}{2}$; 0 9

Flugweite: 9 0; 8 9; 9 0; 10 0; 7 4

Nach unserem Kataloge ist die Heimat Nubien, wo Rüppell diese Hohltnase auffand.

Nycteris discolor Wagner, 1840

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Die zweifarbige Hohltnase.

N. fusca, subtus sordide albida, pilis omnibus basi nigro-schistaceis, auriculus amplissimis.

? *Nycteris capensis*. SMITH zool. journ. IV. p. 434 - SMUTS mamm. cap. p. 7.

Von Ecklon ist uns ein ausgestopftes Exemplar zugekommen, daß vielleicht zu *N. capensis* gehören könnte; indeß ist die Ohrlänge des unserigen beträchtlicher, auch Smith's Angabe der Färbung so ungenau, namentlich von der Zweifarbigkeit der Haare sogar keine Rede, daß ich es nicht wagen mag, selbiges mit *N. capensis* zu identifiziren. Die Formenverhältnisse sind die der vorigen Art; auch die Furche an der Spitze des Unterkiefers scheint nicht zu fehlen; die breite abgerundete Klappe ist and ihrem obern Ende mit Haaren gewimpert. Der letzte Schwanzwirbel vor der

footnotes:

20) Smith's Diagnose lautet: *N. cervice et dorso nigro-fuscis, colli lateribus sordide albis, pertibus inferioribus subcinereis, membranis rubro-fuscis; apice tragi semicirculari cum crista villi albescentis*. Die Eindigung des Schwanzes ist nur schwach gabelig, der letzte Wirbel kurz. Die Höhe der Ohren außen vom Pelze bis zur Spitze giebt er aus $\frac{7}{8}$ ", die Breite über $\frac{6}{8}$ an, die Länge über 2"; als Heimat das Innere von Südafrika und die Ostküste.

21) Smith stellt noch eine Art als *N. affinis* auf: "*N. cervice et dorso rubro-fulvis, colli lateribus rubro-albis, .. terge aurium subrufa; partibus inferioribus fulvo-albis; membranis nigro-fuscis; dentibus primoribus maxillae in paria dispositis*." Die beiden Paare der obern Schneidezähne sind durch einen deutlichen Zwischenraum getrennt; die Endigung des Schwanzes ist stark gegabelt, der letzte Wirbel einer der längsten. Die Länge des Körpers ist 2", die Ohren ziemlich länger und breiter als bei *N. capensis*. - Smuts hält dies *N. affinis* mit Unrecht für identisch mit *N. thebaica*; viel eher könnte mit letzterer *N. capensis* zusammen gestellt werden. Färbung, wie die Stellung der obern Schneidezähne deuten auf eine eigne Art hin; vielleicht möchte sie aber auch mit *N. hispida* zusammen gehören.

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Spalte ist einer der kleinsten. Die Farbe der Oberseite ist im Allgemeinen ein etwas rußiges Relkenbraun, der Unterseite schmutzig weiß, wobei alle Haare, welche sehr lang sind, in ihrer ganzen Wurzelhälfte schieferschwarz sind. Die Ohrwurzel ist hinten und an den Seiten mit weißen Haaren besetzt. Die Wangen sind bräunlich; ein großer brauner Fleck findet sich unten an der Einlenkung der Flügel, während weiterhin die Behaarung an dem Anfang de Oberarms und an dem Flügelansaß am Leibe weißlich, oben braun ist. Die Flughäute und Ohren sind dunkelbraun. Die Heimath dieser Art ist die Südspitze von Afrika.

Nycticeius adovanus Heuglin, 1877

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Die Originaltype von *N. Adovanus* des Stuttgarter Museums gehört nach Professor Peters zu *N. Schlieffenii*, der, wie es scheint, in Kordofan heimlich ist.

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Ich gebe hier die nach dem Thiere im Fleisch von mir entworfene Beschreibung wörtlich aus meinem Notizbuch;

Im Oberkiefer $\frac{2}{3}$ eckzahnartige Schneidezähne, unten $\frac{3}{4}$, deren jeder oben dreispitzig; der obere, dem Eckzahn zunächststehende, sehr klein, wohl abortiv. Ohren start von halber Kopflänge, ihre Außenseite ziemlich gradlinig; Tragus schmal, gegen die Augen zu nach vorwärts geneigt; Nasenlöcher etwas rührig, auswärts und vorwärts gerichtet; Wangen vor den Augen wulstig aufgetrieben; Schwanz bis zum sehr rudimentären Spitzglied von der Schenkelhaut eingeschlossen; der lange Sporn trägt einen äußeren Hautlappen; die Schenkelflughaut reicht bis zum Fersengelenk, die Bauchflughaut dagegen noch weiter am Fuß herab; Obersiete der Füße, Ohren und Flughäute schwärzlich, die letztere mit deutlichem, aber feinem, weißem Saum; Gesicht und Oberseite bräunlich mausfarben; Unterseite graulichweiß; *Patagium interfemorale* mit regelmäßigen Querreihen von weißen Haaren auf der Unterseite; ist aber auch äußerlich spärlich behaart; Nägel weißlich. Körperlänge bis zum After 1" 7^{mm}. Schwanz von After bis zum Spitze 1" 2 $\frac{1}{2}$ ^{mm}. Vorderarm 1" 1^{mm}.

Das beschriebene, den Zahnverhältnissen nach jüngere Thier wurde von uns im November am Asam-Fluß bei Adowa in Tigré erbeutet.

Nannugo Kolenatii Müller, 1858

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Die weissgesäumte Buschfledermaus.

Vesperugo Kolenatii, Zelebor in litt. 6. Oct. 1856.

Eine der allerkleinsten Nordafrikanischen Fledermäuse. Die Schnauze sehr stumpf und abgerundet; die Nasenlöcher hersförmig, vorn etwas seitlich gestellt, der innere Herzflügel etwas weiter und in den Nasenrücken wulstig

[pages 76 and 77 missing - eds.]

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Länge der 1. Phalange am Mittelfinger: 0,0105

" der 2. Phalange am Mittelfinger: 0,0086

" der 3. Phalange am Mittelfinger: 0,0063

" des Metacarpus am 4. Finger: 0,0278

" der 1. Phalange am 4. Finger: 0,0099

" der 2. Phalange am 4. Finger: 0,0056

" der 3. Phalange am 4. Finger: 0,0008

" des Metacarpus am 5. Finger: 0,0281

" der 1. Phalange am 5. Finger: 0,0072

" der 2. Phalange am 5. Finger: 0,0038

" der 3. Phalange am 5. Finger: 0,0012

" des Schenkels: 0,0111

" des Schienbeins: 0,0120

" des Fusses: 0,0056

Vorkommen: In Nord-Afrika, besonders Aegypten, in Gesteins- Mauer- und Baumritzen (Zelebor). Original Exemplare: in der Sammlung des k. Hofnaturalienkabinetts zu Wien, des Hrn. Zelebor, des Prof. Dr. Kolenati in Brünn, Parasiten u. z. 1. Körperhautschmarotzer: Ichoronyssus foveolatus Kolen. Die grubentragende Schmutzmilbe. 2. Ohrmuschelschmarotzer: Otonyssus orthotrichus Kolen. Die geradborstige Ohrmilbe. 3. Flughautschmarotzer: Monostaspis hexastigma Kolen. Die sechsgrubige Einschildborstenmilbe.

Anhang. Mit *V. macuanus* Peters (Säugeth. Mossamb. 61. Taf. 16. Fig. 1.) hat unsere Art keine Aehnlichkeit, da bei *macuanus* der Tragus oben erweitert und die Flugweite = 0.36 ist; eben so wenig mit *V. Rüppeli* Fischer (Synops. 108), bei dem die Ohren rund sind, und die Flugweite = 0.193 beträgt. Bei *V. nanus* Peters (63. Taf. 16. Fig. 2.) ist der Aussenrand des Ohres gebuchtet, und beide oberen Schneidezähne gleich lang, Flugweite 0.22. Sundevall's *V. subtilis* hat zwar oblonge ganzrandige Ohren, aber einen kurzen halb ovalen Tragus. Smiths Mammal. Cap. 107 *V. platicephalus* hat eine an der Rückenseite bis zur Hälfte behaarte Schwanzflughaut, und eine Flugweite von 0.23. Temminck's (Monogr. Mamual. II. 309) und Smiths (Illustrat. S. Afrik. Tab. 51.) *V. minutus* hat den innern obern Schneidezahn zweispitzig, sonst wäre er in der Grösse und Ohrbildung unserem ziemlich ähnlich, doch ist das Patagium bei ihm nicht weiss gesäumt. Smith's *V. hesperida*, 211 hat auch mit unserem hinsichtlich der Grösse viel Aehnlichkeit, aber eine schwarze Schnauze. *Vesperugo Kuhlii* Natterer hat 6 untere Schneidezähne und 8 Gaumenfalten, *Vesperugo ursula* Wagner dagegen einen winkelig vorspringenden Spornlappen und Haflappen an den Fusskrallen, *Vesperugo marginatus* Cretschmar in Rüppel's Atlas 74 Tf. 29 Fig. a. hat zwar den weissen Saum an der Flughaut, aber eine Flugweite von 0.22, und wird von dem

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kritischen Chiropterologen Prof. Blasius selbst als Synonym zu *V. Kuhlii* Natterer gezogen, daher hier ein Verkennen nicht wohl denkbar ist. Er wäre einzureihnen zu *Vesperugo albolimbatus* Cretschmar, *V. subtilis* Sundevall und *V. hesperida* Smith

Neoromicia Roberts, 1926

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Neoromicia gen. nov., type *Eptesicus zuluensis* Rbts.

Since describing *E. zuluensis* in these *Annals* (x, p. 60, 1924), I find that the species ill fits the genus *Eptesicus*, and there being no other to which it can be referred, I have no option but to place it in a new genus as above. It differs from *Eptesicus* in having the cranium slightly raised above the level of the muzzle, as in *Pipistrellus*, from which it differs in having no small upper premolar. In examining the generic arrangement of these little bats, I find it advisable to recognize other genera. Thus, while *Glischropus*, to which I tentatively refer *P. nanus* (Peters), has much in common with the typical *Pipistrellus*, it has a lighter skull, and might therefore be removed if necessary, though it would seem to represent the genus in S. Africa. *Romicia* Gray, type *R. kuhli* (Kuhl), has the heavy skull of *Pipistrellus*, but the upper incisors as in neither, the outer one very small, much less than half the height of the inner, instead of being nearly as high. *Neoromicia* comes nearest to *Romicia*, but lacks the small upper premolar. *Pipistrellus rusticus* (Tomes), which is commonly associated with the above-mentioned genera, has really nothing to do with them and in the shape of the skull shows affinity to *Eptesicus*, from which it differs in its smaller size

and presence of the small upper premolar. I therefore place it in a new genus *EPTESICOPS*, type *V. rusticus* Tomes. With this last genus I tentatively associate *Vespertilio subtilis* Sundeval.

[*Neoromicia vanson*](#) Roberts, 1932

Neoromicia vanson

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A small form similar to *N. zuluensis* and its allied forms in colour and general characters, not so small as *pusillus* Noack, but smaller than *zuluensis* Roberts. Colour above tawny olive for the terminal part of the hair, dark slate basally, the hairs rather long and soft, about 7 mm. in

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length on the middle of the back; these hairs extend slightly on to the membrane at the sides and interfemorally; face and throat like the back, but remaining under parts white, these hairs dark slate for the basal two-thirds. Skull rather lighter than in *capensis*.

Type: T.M. 6553, adult F. Zweizwe Waterhole north of Tsotsoroga Pan, Northern Bechuanaland, 7th July, 1930, No. 882, Vernay-Lang Kalahari Expedition. Length of head and body 38, tail 36, hind foot (c.u.) 5.6, ear 10. Skull: canines to occiput 12.2, zygomatic width 7.3, width of brain case 6.1, of palate across outside of m^2 4.5, length of upper tooth row (c- m^3) 4, of mandible 8.2 mm.

[*Noctula*](#) Bonaparte, 1837

Noctula ??

La presente specie, che a prima giunta crederesti il giovane del *Pipistrellus serotinus*, appartiene certamente al suo medesimo gruppetto di *Noctula* per l'encefalo molto depresso, per la gran disuguaglianza degli incisivi, e per la forma del trago. Si distingue specificamente però dal detto *P. serotinus*, non solo per la piccola statura. ma per la tinta altresì del vello, i cui peli son bicolori, e per l'acutezza delle orecchie; carattere questo che unito a quelli del cranio serve a non farla confondere col *Pipistrellus vispistrellus*, col quale ha più simile il colore. Le vien da noi dato il nome di *Alcythoe*, la più solenne delle tre suore, nella cui denominazione conviene anche Ovidio, che nelle altre da Eliano discorda.

[*Nyctalus verrucosus*](#) Bowdich, 1825

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... and, that the bat is more than specifically distinct from all those which have, as yet, been described, for it has four pointed incisors above (two by the side of each canine, with a large interval between), and six small incisors below, with three indentations in each. It forms a new sub-genus between *pharopus* and *cephalotes*, and may be named, *nyctalus verrucosus* ^h.

Footnote:

^h Νυκταλος, noctem amans. *Suid* : de *Diogen. Cyn.* The lower canines have a heel. The muzzle and *oreillettes* are simple; the ears are equal to the depth of the head in length, and present clusters of orange warts on the outer part, and a few within. It has a nail, and extra joint to the forefinger; three joints to the middle finger, two to the others. The interfemoral membrane (not notched, but triangular) reaches to within one line of the end of the tail, which is within it: - width, from the tip of one wing to the other, $11 \frac{3}{4}$ in., from the muzzle to the tip of the tail $4 \frac{1}{4}$ in., colour, dusky brown.

[*Nycterikaupius*](#) Menu, 1987

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(Second Ensemble dentaire)

(Pl. 4, fig. 1)

Origine du nom: du grec *Nycteris* (chauve-souris) et de J. Kaup, mammalogiste allemand. Il est établi en hommage à ce dernier pour ses travaux sur les Vespertilionidae.

Espèce-type: "*Eptesicus*" *pumilus* GRAY, 1841.

Le genre *Nycterikaupius* est odontologiquement homogène, et notre investigation est

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limitée à cette seule considération. Il est vraisemblable que ce taxon fera plus tard l'objet d'une révision, et que les raisons de le scinder seront autres que celles liées aux structures dentaires. Il semble prématuré de s'engager présentement dans cette démarche, qui nous obligerait à utiliser d'autres méthodologies que l'odontologie. Cette probable mise au point comporte d'ailleurs un préalable: la révision des types spécifiques et leur comparaison avec les pièces muséologiques, afin de limiter la confusion qui règne actuellement à ce niveau.

Diagnose odontologique:

I1/: première voie évolutive; types B' et B" (I,80);
 I2/: type B (I,84); processus de régression engagé;
 C. sup.: type B (I,91) très modifié; la crête mésiale s'estompe, elle laisse place à un arrondi antérieur; la face plane interne devient moins importante; par ailleurs pas de renfort sur la face interne, comme cela est le cas sur les dents du type C (I,91);
 P2/: type B (I,99) quand la dent est présente;
 P4/: type D (I,102), dent morphologiquement fluctuante;
 M1-M2/: type A (I,112) peu souvent; type B (I,114) communément; dans les deux cas la partie linguale de la dent est fluctuante;
 M3/: intermédiaire entre les types A et B (I,122);
 I1-I2-I3: nombreux intermédiaires entre les morphologies B et C (I,86-87); disposition imbriquée ou non;
 C. inf.: type B1 (I,96);
 P/2: type B (I,106);
 P/4: type B (I,108); fusion des racines parfois amorcée; type uniradiculé quelquefois réalisé (I,111);
 M1-M2: myotodontes (I,119);
 M/3: myotodonte, types B et C (I,124-125).

Composition spécifique du genre *Nycterikaupius* gen. nov.: *brunneus*; *capensis*; *inexpectatus*; *matroka*; *melkorum*; *nasutus*; *pumilus*; *rendalli*; *somalicus*; *tenuipinnis*. Cette liste n'est pas limitative

Nycteris aethiopica Dobson, 1878

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Ears as in *N. javanica*, but the tragus is narrower and more curved inwards (Plate XI, fig 3).

Fur, above, brown, paler at the base of the hairs; beneath, the chest and abdomen vary in colour from yellowish white to pure snow-white.

Upper incisors bifid; the second lower premolar as large as in *N. hispida*, standing in the tooth-row, but drawn half inwards.

Length, ear 1".15, thumb 0".6; third finger - metacarp. 1".4, 1st ph. 1", 2nd ph. 1".15; fourth finger - metacarp. 1".5, 1st ph. 0".55, 2nd ph. 0".5; fifth finger - metacarp. 1".55, 1st ph. 0".55, 2nd ph. 0".6; tibia 0".85, foot 0".45.

Hab. N.E. Africa (Kordofan; Senaar).

a. ad. sk. (type). Kordofan. Purchased.
 b, c. ad. sk. Kordofan. Purchased.
 d. ad. sk. Senaar. Purchased.
 e. skull of d.

Nycteris aethiopica guineensis Monard, 1939

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Collection: N^{os} 282 F, 283 M, 284 M, 285 F, 319 M, Mansoa.

Cette nouvelle sous-espèce fait partie du groupe *aethiopica*, mais diffère du représentant occidental du groupe, *macrotis* DOBSON par la taille des oreilles et la forme du tragus; de *aethiopica* DOBSON, elle diffère par la longueur du tibia.

Description. Oreille: grande et large, moins grande toutefois que dans *macrotis*, plus longue que la tête; les deux oreilles se touchent sur la ligne médiane du crâne par un petit lobe accessoire, visible de derrière, mais sans se souder. Tragus de *aethiopica*, marge interne concave, sommet arrondi, marge externe convexe sur toute sa longueur, lobe accessoire replié en avant. Appendices nasaux: lobes supérieurs arrondis en croissant; lobes moyens larges et arrondis; lobes inférieurs dépassant peu les moyens en arrière et seulement légèrement échancrés; fosse postérieure profonde, sans crête médiane; lobe postérieur très proche du lobe interne des oreilles. Membranes normales. Tibias plus longs que dans *aethiopica* et *macrotis*.

Pelage: noirâtre sur le dos, la base des poils claire. Faces ventrales blanchâtres; côtés du museau, nombreux poils des lèvres, base des oreilles par derrière blancs. Membranes noirâtres.

Crâne: incisives supérieures bifides, séparées par paires au milieu. séparées aussi des canines. Deuxième prémolaire inférieure petite, située à peu près dans la ligne dentaire, légèrement en dedans. Pointes accessoires des canines supérieures basses. Des-

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sin formé par les naso-frontaux terminé en arrière par un arc en ogive, les expansions latérales larges.

[table removed - eds.]

Nycteris aethiopica guineensis f. aurantiaca Monard, 1939

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Collection: N° 717 M, Pitche.

Cet unique exemplaire possède tous les caractères de *guineensis* et des mensurations comparables: longueur du corps 56 mm; queue 53; oreille, marge externe 30; avant bras 49; doigt III, 36, 26, 26; doigt IV, 40, 14, 13; doigt V, 42, 13, 14; tibia 25; pied 12. Seule la couleur diffère: elle est d'un bel orangé, avec les pointes des poils plus foncées sur le dos, de même teinte sur le ventre. Le museau et la base des oreilles, par derrière sont blancs. Les appendices nasaux ont la même forme que dans *guineensis*. Une autre forme orangée de *Nycteris* a été signalée: *Nycteris thebaica aurantiaca* DE BEAUX, dans la colonie du Kenya. Les mêmes altérations de couleur se trouvent dans d'autres genres, tels que *Hipposideros*, *Mops*, etc.

Nycteris aethiopica luteola Thomas, 1901

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Similar to the typical form in essential characters, but slightly larger, with longer posterior extremities, and of more yellowish coloration.

Fur soft, fine, and straight, the hairs of the back about 9 millim. in length, considerably longer than those of true *aethiopica*. Humeri and proximal halves of forearms clothed with fur, which also extends on to the wing-membrane. Base of interfemoral membrane and proximal halves of femora also hairy. Below, the fur extends further back on the wing-membrane, but less on the interfemoral than above.

General colour above dull buffy, not far from Ridgway's "clay-colour," browner posteriorly and on the humeri. Individually the hairs are glossy buff, with brown tips. Below, the colour throughout is clear buff, very near Ridgway's pl. v. fig. 13.

Ears and tragus as in true *aethiopica*. Forearms rather longer; tail, lower legs, and feet decidedly longer.

Skull similar to, but rather larger than, that of true *aethiopica*.

Dimensions of the type (in skin): -

Forearm 56 millim.

Tail (vertebræ) 57; lower leg and foot, including claws, 36.

Skull: greatest length from occiput to tip of canines 22.2;

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zygomatic breadth 13; breadth of frontal shield 7.8; upper tooth-row, from front of canine to back of m^3 , 7.5.

The measurements of an adult spirit-specimen are given by Dobson (P. Z. S. 1879, p. 718) under the name of *N. aethiopica*. In that specimen the tail is 59 millim. long and the lower leg and foot 37, just about the same as in the type. On the other hand, in a series of true *aethiopica* from Shendy, Soudan, therefore practically topotypes, the tail does not exceed 53 millim., and the lower leg with foot 33 millim., these measurements being very uniform in all.

Hab. Kitui, British East Africa, alt. 3500 feet. Another specimen from Zanzibar.

Type. Female. B.M. no. 1.5.6.4. Collected 14th February, 1901, and presented by S. L. Hinde, Esq.

The original series of *N. aethiopica*, from Kordofan, consists of three faded skins, with imperfect skulls; but fortunately, just as Mr. Hinde's specimen needed determination, the Museum has been presented by Messrs. N. C. Rothschild and A. F. R. Wollaston with a very nice set, both skins and spirit-specimens, from Shendy, on the Upper Nile, and these have enabled me to satisfy myself as to the constancy of the distinguishing characters of the new form.

As already indicated, the example from Zanzibar, referred to by Dobson (P. Z. S. 1879, p. 718), proves to belong, as might be expected, to *N. æ. luteola*.

Nycteris affinis A. Smith, 1829

p. 434

Nycteris affinis, mihi.

N. cervice et dorso rubro-fulvis; colli lateribus rubro-albis; a tergo aurium subrufa; partibus inferioribus fulvo-albis; membranis nigro-fuscis; dentibus primoribus maxillæ in paria dispositis.

Neck above and back reddish brown or tawny, sides of neck before wings reddish white; behind ears somewhat rufous; beneath tawny white, membranes blackish brown. Incisors of upper jaw in pairs which are separated from each other by a distinct open space immediately in the front of the jaw. Length from nose to root of tail, two inches; ears rather longer and broader than in the last species; tragus short, and its apex semicircular; termination of tail deeply forked, with the last vertebra but one the longest.

The arrangement of the incisors of the upper jaw, the marked difference in respect to the last joint but one of the tail, added to the

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depth of the fork, and the greater proportion of transverse veins, in the portion of the interfemoral membrane connected with the joint named, tend, with other characters, to establish this as a distinct species from the last.

Nycteris angolensis Peters, 1871

p. 903

(Fig. 5.)

Durch die Güte des Hrn. Barboza du Bocage habe ich verschiedene Exemplare einer *Nycteris* zur Untersuchung erhalten, welche ich für identisch mit *N. fuliginosa* aus Moçambique gehalten habe. Eine genauere Untersuchung hat mir aber gezeigt, dafs, obgleich sie in der Färbung mehr mit dieser letztern übereinstimmt, sie durch die Entwicklung des kleinen zweiten untern falschen Backzahns und auch durch eine etwas geringere Länge des Sporns der *N. thebaica* näher steht und dafs sie von dieser nur durch eine etwas stärkere Entwicklung dieses äufserst kleinen

p. 904

Zalme von ihr verschieden ist. Der Tragus zerfällt, wie gewöhnlich, in zwei Abtheilungen und die obere abgerundete Abtheilung hat, wie bei *N. thebaica* und *capensis*, den vordern Rand convex.

Meter

Totallänge: 0.113

Kopf: 0.0195

Ohrhöhe: 0.030

Ohrbreite: 0.020

Ohrklappe: 0.008

Schwanz: 0.066

Oberarm: 0.023

Vorderarm: 0.045

L. 1. F. Mh. 0.0058; 1 Gl. 0.005; 2 Gl. 0.0022: 0.013

L. 2. F. - 0.040; - 0.0015: 0.0115

L. 3. F. - 0.0368; - 0.026; - 0.024; Kpl. 0.005

L. 4. F. - 0.038; - 0.015; - 0.0115; - 0.0012

L. 5. F. - 0.038; - 0.0145; - 0.0125; - 0.0025

Oberschenkel: 0.024

Unterschenkel: 0.023

Fufs: 0.012

Sporn: 0.017

Diese Art ist in Caconda, Biballa und Rio Coroca VOn Hrn. Anchieta gefunden worden

Nycteris arge Thomas, 1903

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Allied to *N. thebaica*, but with much smaller tragus.

Size medium. General colour dark brownish above and below. Nose-leaf of normal structure, but unusually thickly

p. 634

hairy. Ears of average size, the projecting lobule at their outer bases very strongly developed, inverted, deeply concave externally, convex internally. Tragus with its free portion, as in *N. thebaica*, expanded above and convex on its inner margin, but the free portion itself is barely one third its size in the allied species, the distance from its inner base to its tip considerably less than the distance from the same point to the base of the outer margin.

Upper incisors deeply bifid; second lower premolar nearly half the size of the first, in the tooth-row.

Dimensions of the type (measured in spirit): -

Forearm 45 mm.

Head and body 52; tail 48; lower leg and foot (s.u.) 32.5; head 21.5; ear 28; inner margin of tragus 2.5.

Hab. Efulen, Cameroons.

Type. Adult male. Collected by Mr. G. L. Bates. Two specimens.

This *Nycteris* is readily distinguishable by the extremely small size of the free portion of the tragus, which is otherwise similar in shape to that of *N. thebaica*.

Nycteris avakubia J.A. Allen, 1917

p. 426

Type (and only specimen, in alcohol), No. 49403, M ad., Avakubi, Belgian Congo, September, 1913; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 2623.

A member of the *N. hispida* group, intermediate in size between *N. hispida* and *N. grandis*. Upper incisors trifid, p_4 large, fully one third the size of p_3 , and similar to it in general form. Coloration not distinctive, but very dark including ears and membranes, in the single specimen in alcohol.

Total length, 137 (150, 144.7); head and body, 63 (–, 71); tail, 65 (70, 73.7); ear, 25 (32, 30.5); forearm, 51 (59.5, 57); third finger, metacarp, 40 (44, 43.2); tibia, 30 (25.5, 31.8); foot, 12 (18.5, 15.2); calcaneum, 20 (23, 22.9).

Skull, total length, 22.4; zygomatic breadth, 12.9; upper toothrow (c-m³), 7.7; length of mandible, 15.1; lower toothrow (c-m₃), 8.4.

Represented only by the type (in alcohol) taken at Avakubi.

Nycteris benuensis Aellen, 1952

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COLLECTION

2 exemplaires.

N° 1481 a (type) M ad., 1481 b (tête seulement), Rei Bouba, 16.9.1947.

DESCRIPTION

Caractères externes. - D'allure générale, cette nouvelle espèce est semblable à *nana nana*. Les mesures externes sont un peu plus fortes que chez cette dernière. L'oreille et la feuille sont comme chez *nana nana*. Le lobule de la base du bord externe est grand, presque plat, beaucoup moins concave que chez *nana* ou *arge*. Le tragus est falciforme; le bord externe est convexe; le bord interne est sinueux, concave à la base, convexe au sommet; la partie libre est égale à la moitié de la longueur totale (fig. 12, p. 50). La queue est relativement plus courte que chez *nana nana*. Le pelage s'étend sur le 1/3 basal de l'avant-bras, à la face supérieure, un peu moins dessous.

Coloration. - La teinte générale est gris brunâtre, légèrement plus clair à la face inférieure et à la base des oreilles. Les oreilles et le patagium sont bruns.

Crâne. - Il ne présente aucun caractère particulier.

Dentition. - Les incisives supérieures sont à peine bifides, probablement très usées. La 2^{me} prémolaire inférieure est dans la rangée dentaire; elle est petite et atteint juste le talon de la première prémolaire dans un des spécimens (N° 1481 a), elle est encore plus petite dans l'autre.

Mensurations

Type, N° 1481 a, M ad.

Long, tête + corps: 46

Avant-bras: 38
 Tibia: 18.2
 Pied: 9.2
 Oreille: 19.5
 Queue: 44
 3^{me} doigt, métac: 30.5
 " 1^{re} phal: 21
 " 2^{me} phal: 23
 4^{me} doigt, métac: 32
 " 1^{re} phal: 11.8
 " 2^{me} phal: 8
 5^{me} doigt, métac: 31.5
 " 1^{re} phal: 11
 " 2^{me} phal: 11.5

Paratype, N° 1481 b, ad.
 Crâne, long, totale: 17.4
 Long, condylobasale: 14.5
 Larg.. Zygomatique: 10.2
 Larg. Cérébrale: 7.9
 Larg. Interorbit: 4.8
 Larg. Bords ext. de M³-M³: 6.6
 Rang. Dent. Sup: 5.6
 Rang. Dent. Sup. (alv.): 5.2
 Long, mandibule: 10.9
 Rang. Dent. Inf: 5.9

N. benuensis est voisin de *nana*, mais s'en distingue principalement par la forme du lobule du bord externe de l'oreille et par la taille de la 2^{me} prémolaire inférieure.

BIOLOGIE

Les deux exemplaires obtenus par la Mission Suisse ont été apportés par un indigène en même temps que des *Rhinolophus foxi*. Mais, il n'est pas certain que ces deux espèces aient été trouvées ensemble.

Nycteris Capensis A. Smith, 1829

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Nycteris Capensis, mihi.

N. cervice et dorso nigro-fuscis, colli lateribus sordido-albis; partibus inferioribus subcinereis: membranis rubro-fuscis; apice tragi semicirculari cum cristâ villi albescentis.

Neck above and back blackish brown, sides of neck dirty white, below cinereous white with shades of blackish; membranes reddish brown; height of ears without from fur to tip, $\frac{7}{8}$ of an inch, width better than $\frac{6}{8}$; tragus short, apex semicircular and its upper edge clothed with a tuft of long, white, woolly fur; termination of tail but slightly forked, last vertebra but one if any thing the shortest. Length from nose to root of tail, better than two inches; expanse of wings, ten inches. Found in the interior parts of South Africa as well as upon the Eastern coast.

Nycteris damarensis Peters, 1871

p. 905

(Fig. 7.)

Aus dem Damaralande haben das Berliner, das Stockholmer und das British Museum Exemplare einer *Nycteris* erhalten, welche der *capensis* Smith äusserst nahe steht, durch die ganz schneeweiße Unterseite, ohne bräunliche Schattirung an der Seite der Brust vor der Schulter, auffällt und sich durch eine etwas stärkere Entwicklung des kleinen zweiten untern falschen Backzahns auszeichnet. Ich lasse dieser Art den Namen, unter welchem sie im British Museum und in dem Catalogue of Mammalia von 1843 (p. 24) aufgeführt ist, obgleich sie niemals beschrieben wurde.

Meter
 Totallänge: 0.118
 Kopf: 0.0222
 Ohrlänge: 0.035
 Ohrbreite: 0.023
 Tragus: 0.009
 Schwanz: 0.056
 Oberarm: 0.022
 Vorderarm: 0.040

L. 1. F. Mh.0.005; 1 Gl. 0.005; 2 Gl. 0.0025: 0.013
 L. 2. F. - 0.040; - 0.0: 0.040
 L. 3. F. - 0.0372; - 0.0265; - 0.025; Kpl. 0.005
 L. 4. F. - 0.0408; - 0.014; - 0.0117; - 0.0015
 L. 5. F. - 0.0408; - 0.0137; - 0.0117; - 0.002
 Oberschenkel: 0.0235
 Unterschenkel: 0.023
 Fufs: 0.012 - 0.013
 Sporn: 0.017

Wir haben diese Art durch Hrn. Hahn aus Otjimbingue

Nycteris Daubentonii E. Geoffroy Saint-Hilaire, 1813

p. 19

Pelage roussâtre; blanc-sale sous le ventre. Oreilles oblongues.

p. 20

Vesp. hispidus. SCHREB., pl. 56.

HABITE le Sénégal.

Nycteris fuliginosa Peters, 1852

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Tafel X.

N. fuliginosa, *subtus ex griseo fuscenscens; alis nigris; pilis cervicis, laterumque colli in basi albis, reliquis in basi nigroschistaceis; auriculis dimidio capite longioribus.*

Longitudo tota 0,120; caudae 0,055; antibrachii 0,045; volatus 0,300.

Habitatio: Africa orientalis, Boror, 17° Lat, Austr.

Die Ohren sind außerordentlich lang, entfaltet um die Hälfte länger als breit und um ebenso viel länger als der Kopf, über der Stirn durch eine niedrige wulstige Hautfalte verbunden, von elliptischer Gestalt, oben an der Vereinigung der verdickten bogenförmigen Ränder stumpfwinklig zugespitzt, am vorderen Rande und an der Basis mit wolligem weifslichen Haar bekleidet, am übrigen Theil mit zerstreuten Härchen bewimpert. Der Tragus ist durch einen hintern Einschnitt in zwei Lappen getheilt; der obere Lappen ist eiförmig, länger als breit, am obern Rande mit einem Haarbüschel geziert; der Antitragus ist abgerundet, deutlich vom hintern Ohrande abgesetzt und verlängert sich nach vorn durch eine Hautfalte über die Schläfe. Der hintere Ohrand spaltet sich, wie bei allen andern Arten, in eine innere nach dem Gehörgang gerichtete Lamelle und in den äufsern mit dem Antitragus verbundenen Theil. Die Augen sind klein und stehen in der Mitte zwischen dem Schnauzenende und der Wurzel des vorderen Ohrandes. Die Schnauze ist stumpf und abgerundet; die Maulöffnung ist bis unter die Basis der Nasenklappe gespalten, und an den Seiten in einem nach unten flach convexen Bogen geschwungen. Der Rand der Oberlippe ist vorn ein wenig vorstehend und verdickt. Die Unterlippe zeigt eine mittlere breite dreieckige sich nach unten verschmälernde Furche, die zu beiden Seiten durch einen wulstigen Rand, nach oben hin durch eine warzige Erhöhung begrenzt ist. Die Bildung des Nasenbesatzes, die Furchung des Kinnes und die Bildung der Lippen zeigt nichts von den andern Arten Abweichendes. Die Schleimhaut des Gaumens bildet sechs Querfalten. Die Behaarung des Körpers ist sehr reich, an der Rückseite doppelt so lang wie am Bauche. Sie erstreckt sich aber über die Flughäute nicht weiter als bei *Nycteris thebaica*, Geoffroy, etwas über die Hälfte des Oberschenkels, an der Bauchseite etwas weiter als an der Rückenseite. Die Verhältnisse der Gliedmafsen gehen aus den unten angeführten Mafsen hervor, sie zeigen nichts Eigenthümliches. Das letzte Schwanzglied ist bei verschiedenen Exemplaren von verschiedener Länge, bald äufserst kurz, bald länger, immer aber viel kürzer als die beiden Zweige des Endknorpels zusammengenommen.

Die Farbe der Rückenseite ist rufsbraun, die Bauchseite grau mit einem bräunlichen Schein. Die Haare des Nackens und der Halsseiten sind am Grundtheile weifs, die übrigen schwarzgrau, schieferfarbig. Die Flügel und Ohren erscheinen blauschwarz, bei durchfallendem Schein haben sie eine Beimischung von Braun. Bei jungen Individuen ist die Farbe etwas

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heller, immer aber noch viel dunkler als bei den verwandten Arten. Die Nägel sind von weifser Hornfarbe.

Der Schädel und das Gebifs zeigen dieselbe Form wie von *Nycteris thebaica*; auch in den relativen Verhältnissen finde ich keinen bemerkenswerthen Unterschied. Der untere zweite kleine Lückenzahn ist bei allen Exemplaren vorhanden, und ein ganz wenig gröfser als bei den jungen ägyptische, denen er im späteren Alter zu fehlen scheint. Die Zahl der Wirbel ist constant an mehreren Skeleten 35: hiervon sind 7 Halswirbel, 10 Rückenwirbel, 6 Lendenwirbel, 5 Kreuzbeinwirbel und 7 Schwanzwirbel.

Die Zunge ist ganz gleichmäßig von kleinen platten Schüppchen bedeckt, welche auf der Grundhälfte etwas weniger gedrängt stehen, und nahe der Basis zeichnen sich zwei von einer ringförmigen Vertiefung umgebene Wärcchen durch ihre Größe aus. Die Speiseröhre geht, sobald sie das Zwerchfell durchbohrt hat, in einen sackförmigen rundlichen Magen über, der sich durch eine geringe Abschnürung von dem etwa 120 Millimeter langen Darm scheidet, welcher einfach, nach mehreren Windungen, in das etwas erweiterte Rectum übergeht. Die Luftröhre macht unter dem Kehlkopf zwei seitliche Anschwellungen und enthält bis zu ihrer Theilungsstelle 23 knorpelige Halbringe. Die Lungen bilden jederseits nur einen unregelmäßig quadratischen Lappen. Die Nieren sind ungelappt, bohnenförmig. Die Nebennieren haben eine platte Gestalt und sind sehr klein. Die Gestalt der Milz ist zungenförmig. Die Hoden liegen aufserhalb der Bauchhöhle. Die Ruthe enthält in ihrer Endhälfte einen 3 Mm. langen Knochen, der mit einer breiten, ausgehöhlten Basis dem Ende der Corpora cavernosa aufsitzt. Die Hörner am Uterus des Weibchens sind sehr lang.

Ich fand diese *Nycteris* nur ein einziges Mal, aber in großer Anzahl, in einem dunkeln Raume meiner Wohnung, welcher durch einen Schornstein mit der freien Luft in Verbindung stand. Es war dies im März 1846, in Boror, etwa 12 Meilen nordwestlich von Quellimane. Diese Thiere nähren sich wol nur von Insecten, obgleich die Einwohner ihnen Schuld geben, daß sie das Fleisch in den Vorrathskammern angreifen. Wenigstens fand ich nur Theile von Insecten in ihren Verdauungsorganen. Die Eingebornen nennen diese Fledermaus *nantûtu*.

Mafse in Millimetern.

M; F

Ganze Länge von der Schnauze bis zur Schwanzspitze: 120; 115

Flugweite: 300; 300

Länge des Kopfes: 21; 21

Länge oder Höhe des ganzen Ohres: 32; 32

Breite des Ohres: 20; 21

Länge des Oberarms: 20; 20

Länge des Unterarms: 45; $43\frac{1}{2}$

Länge des Daumens (Mittelh. 5, 5. 1.Gl. $5\frac{1}{2}$, $5\frac{1}{2}$. 2.Gl. $2\frac{1}{2}$, $2\frac{1}{2}$): $12\frac{1}{2}$; $12\frac{1}{2}$

Länge d. 2ten Fingers (Mittelh. 39, $35\frac{1}{2}$. 1.Gl. 1, $1\frac{1}{2}$): 40; 37

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Länge d. 3ten Fingers (Mittelh. 35, 32. 1.Gl. 24, 23. 2.Gl. $23\frac{1}{2}$, 23. 3.Gl. $3\frac{1}{2}$, $3\frac{1}{2}$, $3\frac{1}{2}$): 86; $82\frac{1}{2}$

Länge d. 4ten Fingers (Mittelh. 37, 35. 1.Gl. $13\frac{3}{4}$, $13\frac{1}{4}$. 2.Gl. 10, $10\frac{1}{4}$. 3.Gl. 1, 1): $61\frac{3}{4}$; $59\frac{1}{2}$

Länge d. 5ten Fingers (Mittelh. $38\frac{1}{2}$, 36. 1.Gl. $1\frac{1}{2}$, 13. 2.Gl. $11\frac{1}{4}$, 11. 3.Gl. $2\frac{1}{2}$, 2): $65\frac{3}{4}$; 62

Länge des Oberschenkels: 22; 22

Länge des Unterschenkels: $23\frac{1}{2}$; $23\frac{1}{2}$

Länge des Fusses: $10\frac{1}{2}$; $10\frac{1}{2}$

Länge des Schwanzes: 55; 52

Länge des Sporns: 18; 18

Länge des Schädels: 19; 19

Länge der Wirbelsäule vom Atlas bis zum Ende des Kreuzbeins: 35; 35

Nycteris capensis und *Nycteris affinis* sind nur nach Smiths kurzen Diagnosen bekannt; hiernach unterscheidet sich die erste von vorstehender Art durch rothbraune (rubrafusca) Färbung der Flughäute, schwache Gabelung des Schwanzes und die im Verhältniß zum Länge größere Breite der übrigens kürzeren Ohren (Länge $\frac{7}{8}$ Zoll, Breite $\frac{6}{7}$ Zoll); die zweite ist ganz anders gefärbt "cervice et dorso rubro-fulvis, colli lateribus rubro-albis; a tergo aurium subrufa; partibus inferioribus fulvo-albis: membranis nigro-fuscis;" und soll die oberen Schneidezähne paarweise getrennt haben. Außerdem wird zwischen beiden Arten noch eine Verschiedenheit in der Kürze oder Länge des letzten Schwanzgliedes gesucht, was nach meine Beobachtungen bei einer und derselben Art variirt, *N. discolor*, welche ich durch die zuvorkommende Güte meines hochverehrten Freundes A. Wagner vergleichen konnte, hat eine breitere Ohrklappe und verhältnismäßig breitere Ohren, außerdem ist die Farbe des Thieres und der Flughäute viel heller braun. *Nycteris hispida*, Schreber, ist oben röthlichbraun, unten gelblich weifs. *Nycteris thebaica*, Geoffroy, hat kürzere und verhältnismäßig breitere Ohren, die Ohrklappe ist breiter und abgerundeter, die Farbe der Rückenhaare ist an der Basis schmutzig weifs, und der zweite kleine untere Lückenzahn ist ganz nach innen gedrängt und fällt bei den älteren Individuen aus. *Nycteris javanica*, Geoffroy, ist durch ihre Größe, durch die roströthliche Farbe und die Größe des zweiten untern Lückenzahns hinreichend von allen anderen Arten unterschieden.

Nycteris Geoffroyi Desmarest, 1820

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(Non figuré dans l'Encycl.) *Description d'une chauve-souris étrangère*, Daub. Œuvres de Buffon, descript. du cabinet, n. DCDX et DCDXI. - *Nyctère de la Thébaïde*, Geoff. Mém. de l'Institut. d'Egypte, hist. nat. tom. 2. pl. 1 et 2.

CAR. ESSENT. *Oreilles très-grandes; opercules des environs des narines assez développés et en spirale; lèvre inférieure ayant une forte verrue à son extrémité, située entre deux bourrelets alongés, non réunis et en forme de V; pelage d'un gris-brun en dessus; gris plus clair en dessous.*

DIMENS. Longueur du corps mesuré en ligne droite, depuis le bout du museau jusqu'à l'origine de la queue: 1 pouc. 10 lig.

- de la tête, depuis le bout du museau jusqu'à l'occiput: 1 pouc. 10 lig.

- de la fosse du chanfrein: 5 lig.

Largeur des opercules: $1 \frac{2}{7}$

Longueur des oreilles: 11 lig.

Envergure des ailes: 9 pouc.

Longueur du pouce de l'aile: $5 \frac{2}{7}$

- de la queue: 1 pouc. 11 lig.

DESCRIPT. Tête grosse, fort prolongée en avant; crâne volumineux, très-arrondi en arrière; museau renflé; bouche très-fendue; lèvre supérieure haute et très-entière; lèvre inférieure comme bifurquée, et offrant deux bourrelets ou replis de la peau épais et nus, formant un angle entr'eux, et étant séparés par un sillon qui se prolonge sous la mâchoire; un tubercule entre-deux, formant la terminaison de la lèvre; canines assez fortes; incisives très-petites et bilobées ou trilobées; langue alongée, arrondie au bout, et ayant sa surface parsemée de petits grains élevés qui paroissent être des papilles cornées extrêmement fines; nez très-compiqué, composé, 1°. des deux ouvertures nasales fort rapprochées et situées à la partie antérieure d'une grande fosse du chanfrein qui se porte depuis le haut de la lèvre jusqu'à la base du crâne proprement dit; 2°. d'un repli mince de la peau, recouvert de poil, bordant extérieurement, cette fosse, et ne s'apercevant que lorsqu'on le soulève avec l'extrémité d'un instrument aigu; 3°. de deux replis plus minces, longitudinaux, sans poils, situés parallèlement l'un à l'autre dans le fond de la fosse du chanfrein; 4°. de deux espèces de pièces de

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forme arrondie, un peu en spirale, tenant au repli extérieur de la peau, et recouvrant en partie le milieu de la fosse du chanfrein, mais non les ouvertures des narines, qui sont situées en avant; oreilles placées à peu près au tiers postérieur de la longueur de la tête, d'une hauteur presque double de la sienne, ayant l'ouverture de la conque de forme ovale oblongue, dirigée en avant et les contours entiers; les bords internes des deux oreilles étant assez rapprochés l'un de l'autre, et même réunis sur le front par une perire cloison membraneuse, transversale; bords externes commençant sur les côtés de la tête et fort bas, où ils forment un assez grand repli; conque velue près de la tête, n'offrant en dehors qu'un seul pli droit, partant de sa base et se portant presque à son extrémité, et assez près du bord externe, ce pli étant indiqué par une nervure saillante postérieurement, et garnie d'une seule rangée de petits poils disposés comme des cils; des poils rares sur les deux faces de la conque, et dont les bulbes forment autant de points moins transparens que le reste de la membrane; oreillon petit, appliqué au bord interne du dedans de la conque, de forme arrondie ou en cuiller, et étant deux fois aussi large que haut, sa face antérieure étant velue; yeux petits, une fois plus près de l'oreille que de la pointe du museau; cou court, mais bien marqué; corps très-épais et très-muscleux antérieurement; ligne moyenne du dos entre les épaules, offrant un sinus longitudinal très-profond; poitrine très-renflée et très-large; ventre mince; ailes grandes et larges; pouce grêle avec un ongle foible; muscles des avant-bras très-forts; membrane interfémorale très-ample, soutenue par des osselets cartilagineux presque aussi longs que la jambe, et embrassant la queue, qui est formée de sept vertèbres, et terminée par un cartilage en forme de T, dont les branches partent à droite et à gauche de l'extrémité de la dernière; pelage doux et fin, brun en dessus et gris-brun clair en dessous.

Nota. Telle est la description détaillée d'un nyctère qui nous a été rapporté du Sénégal par M. Huzard fils, habile médecin vétérinaire. Ayant comparé ce cheiroptère avec le nyctère de la Thébaïde de M. Geoffroy, nous n'avons pu trouver de caractères distinctifs assez tranchés pour l'établir en titre d'espèce; mais les proportions de diverses parties de son corps nous ont présenté quelques différences que nous relaterons ici.

Nycteris Geoffroyi Var. *Senegalensis* Hartmann, 1868

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N. Geoffroyi Desm. (Mumm. p. 127, 190). Var. *Senegalensis* (das.). Sennar.

Nycteris grandis Peters, 1865

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In der Färbung und Behaarung ganz mit *N. fuliginosa* übereinstimmend, aber beträchtlich gröfser, indem sie selbst noch *N. javanica* an Gröfse übertrifft. Die Ohren sind so lang wie der Kopf. Die vier oberen Schneidezähne sind dreilappig und der zweite untere falsche Backzahn ist zwar klein, indem seine Gröfse kaum ein Drittel des vorhergehenden Zahns ausmacht, aber er ist nicht von vorn nach hinten zusammengedrückt. Auch bei dieser grofsen Art habe ich vergeblich nach einer ??bula gesucht.

Länge vom Hinterhaupt bis Ende der Schenkelflughaut ungefähr 0^m,110; Länge der Schenkelflughaut ungefähr 0^m,0??, Kopf 0^m,029; Ohr 0^m,029; Vorderarm 0^m,055; Tibia 0^m,0??; Sporn 0^m,023.

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Aus Guinea; dem einzigen getrockneten Exemplar, welches sich im Reichsmuseum zu Leiden befindet, ist der Schwanz ausgezogen, so dafs sich die Länge desselben nicht genau bestimmen läfst.

Nycteris intermedia Aellen, 1959

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Type. - F adulte, en alcool. Adiopodoumé, près d'Abidjan, Côte-d'Ivoire, 20.7.1953; coll. V. Aellen, n° orig. 623. Muséum d'Histoire naturelle de Genève, n° 923.94.

Matériel examiné. - Trois paratypes ont encore été obtenus en Côte-d'Ivoire, à Cosrou (W d'Adiopodoumé) le 24.7.1953 (2 F, n°s 923.95 et 923.97 Mus. Genève) et le 19.8.1953 (F n° 923.96). Je rapporte, d'autre part, à cette espèce le spécimen n° 277 de la Mission scientifique suisse au Cameroun, que j'avais déterminé *Nycteris nana* et dont la forte taille m'avait déjà intrigué (AELLEN, 1952).

Diagnose. - *Nycteris* de petite taille (avant-bras 37 mm) voisin de *N. nana* (Anders.) et de *N. arge* Thom., c'est-à-dire à P_4 grand (égal ou supérieur à la moitié de P_2) et à I^1 bifide; de proportions très semblables à *N. arge*.

Description. - Par tous ses caractères, cette nouvelle espèce est voisine de *N. nana* et *N. arge*. Les oreilles ont la même proportion que chez ces derniers, c'est-à-dire qu'elles mesurent environ les 60 à 75 % de la longueur de l'avant-bras. La profonde poche située à la base externe de l'oreille (lobe antitragal) est présente comme chez *N. nana*, *arge* et *major*. Le tragus est de forme semblable chez les quatre spécimens: type liguli-pyriforme; le bord interne est droit ou légèrement convexe, le bord externe est généralement échancré au milieu, parfois presque droit. Le patagium est inséré environ au milieu du métatarse, alors que chez *N. nana* et *N. arge*, il est fixé au tiers distal du métatarse, presque à la base de la première phalange. Le tibia est de même proportion que celui de *N. arge*; il dépasse la moitié de la longueur de l'avant-bras (plus court chez *N. nana*).

La coloration du spécimen type était la suivante, au moment de sa capture: sommet de la tête et dos brun un peu roussâtre; cou et surtout base externe des oreilles brun fauve; sur le dos les poils sont bruns de la racine à la pointe, au cou ils sont gris blanchâtre à la base et brun fauve au sommet. La face ventrale présente la même coloration que le dessus. La feuille nasale est

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brune; les côtés du museau brun clair, les oreilles et le patagium brun foncé.

Le crâne est de forme générale très semblable à celui de *N. arge*. La largeur zygomatique représente les 56,8 à 58,6 % (moy. 57,45) de la longueur totale du crâne (chez *N. arge*: moy. 58,6 %, chez *N. nana*: moy. 55 %). Les premières et secondes incisives supérieures (I^1 et I^2) sont bicuspidées. La deuxième prémolaire inférieure (P_4) atteint la moitié ou les deux tiers de la hauteur de la première prémolaire (P_2); en aucun cas, elle n'arrive à la hauteur de la pointe antéro-interne de la première molaire.

Remarques. - La répartition géographique de *N. intermedia* englobe très probablement toute la zone forestière du golfe de Guinée. Connu jusqu'à présent seulement en basse Côte-d'Ivoire et dans le S du Cameroun, ce Chiroptère a peut-être été signalé par quelques auteurs sous le nom de *N. nana* ou éventuellement de *N. arge*. Du fait qu'il cohabite avec ces deux espèces, on doit le considérer comme espèce autonome et non comme sous-espèce de l'une ou l'autre.

De taille exactement intermédiaire entre *N. nana* et *N. arge*, il fait partie d'un groupe de *Nycteris* qui demande une nouvelle définition. Mon intention, dans ce court article descriptif, n'est pas de faire l'historique du genre *Nycteris* et de ses divisions subgénériques. Cependant, je rappellerai qu'ANDERSEN (1912) a proposé un arrangement des espèces de *Nycteris* en quatre groupes constitués comme suit:

1° groupe **javanica**: *javanica*, *tragata* (espèces indo-malaises);
arge, *nana*, *major* (espèces africaines);

2° groupe **hispida**: *hispida*, *aurita*, *grandis*;

3° groupe **aethiopica**: *aethiopica*, *macrotis*;

4° groupe **thebaica**: *thebaica*, *revoili*, *capensis*, *damarensis*, *gambiensis*.

Si l'on considère seulement le premier de ces groupes, on constate que l'une des espèces se sépare nettement des autres par ses incisives supérieures tricuspidées: il s'agit de *N. javanica* (cf. ELLERMAN ET MORRISON-SCOTT, 1953). Les autres espèces

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ont toutes les incisives supérieures bicuspidées. Dès lors, il me semble nécessaire de créer un nouveau groupe qui se trouve défini dans la clé suivante. Le nom de l'espèce la plus anciennement décrite a été choisi pour désigner ce groupe:

1. P_4 grand (égal ou plus grand que la moitié de P_2). --> 2

- P_4 petit (plus petit que la moitié de P_2). --> 3

2. I¹⁻² bicuspides (I² parfois tricuspidé). --> groupe **arge** gr. nov.

- I¹⁻² tricuspides. --> groupe **javanica**

3. I¹⁻² bicuspides. --> 4

- I¹⁻² tricuspides. --> groupe **hispida**

4. Tragus semi-luné. --> groupe **aethiopica**

- Tragus pyriforme (partie libre plus étroite à la base). --> groupe **thebaica**.

Clé des espèces du groupe arge:

1. Avant-bras de plus de 39 mm. Long, du crâne dépassant 19 mm. --> 2

- Avant-bras de moins de 38 mm. Long, du crâne inférieure à 19 mm. --> 4

2. Rangée dentaire supérieure C-M³ de plus de 8 mm. Avant-bras de 46 mm environ. --> *tragata* Anders. 1912

- Rangée dentaire supérieure C-M³ inférieure à 8 mm. --> 3

3. Avant-bras de 47,3 à 50 mm. Long, du crâne de 20,9 à 22,4 mm. Oreilles mesurant les 50 à 65 % de la longueur de l'avant-bras. --> *major* Anders. 1912

- Avant-bras de 39,5 à 46 mm. Long, du crâne de 19,3 à 20,9 mm. Oreilles mesurant les 63 à 77 % de la longueur de l'avant-bras. --> *arge* Thom. 1903

4. Avant-bras de 36,5 à 37 mm. Long, du crâne de 18 à 18,7 mm. Tibia plus long que la moitié de la longueur de l'avant-bras. --> *intermedia* sp. nov.

- Avant-bras de 32 à 36,2 mm. Long, du crâne de 15,9 à 16,6 mm. Tibia plus court que la moitié de la longueur de l'avant-bras. --> *nana* Anders. 1912

Dans le tableau de mesures suivant, j'indique à titre de comparaison les minima et maxima pour *N. nana* et *N. arge*.

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Ces derniers chiffres ont été publiés par divers auteurs et sont complétés par des mensurations personnelles et inédites.

Nycteris intermedia (923.94 F type, 923.95 F, 923.97 F, 923.96 F); *N. nana* (min - max); *N. arge* (min - max):

Avant-bras: 37, 37, 37, 36.5; (32 - 36.3); (39.5 - 46)
 3e doigt, métacarpe: 29.5, 30, 30, 29.5; (25 - 28.5); (30 - 36)
 " 1^{re} phalange: 19, 19.5, 18.5, 18.5, (15.5 - 16.5); (20 - 23.1)
 " 2^e phalange: 23, 22.5, 23, 23; (18); (25 - 27)
 4e doigt, métacarpe: 31.5, 33.5, 33, 32.5 (27.5 - 29.7); (33 - 38.2)
 " 1^{re} phalange: 10.5, 11, 10.5, 10; (9); (11 - 13.4)
 " 2^e phalange: 9, 9, 9, 9.5; (8); (11 - 13.2)
 5e doigt, métacarpe: 32, 34, 33.5, 32.5; (27 - 30.3); (35 - 38.6)
 " 1^{re} phalange: 10, 10.5, 10.5, 10; (9); (11.5 - 13.8)
 " 2^e phalange: 11, 11, 11.5, 11.5; (9); (12.5 - 14.5)
 Tibia: 21, 21, 21, 21; (14.5 - 17.3); (22 - 24.5)
 Pied (avec griffes): 9, 9, 9, 9.5; (6.5 - 8); (9.5 - 11)
 Queue (de l'anus): 53, 51, 52, 53; (38 - 44); (50 - 60)
 Oreille (longueur): 24, 23.5, 24, 24; (17 - 21); (25 - 32)
 Tragus (bord interne): 2.1, 2.2, 2.2, 2.3; (1.7); (2.5)
 " (bord externe): 5.4, 5.3, 5.5, 5.5; (3.2); (6.2)

Crâne:

Long, totale (à la canine): 18.7, 18, 18.3, 18.2; (15.9 - 16.6); (19.3 - 20.9)
 " condylobasale: 16, 15.2, 15.7, 15.7; (13.4 - 13.9); (16.8 - 18.2)
 " losange nasal: 8, -, 8.1, 7.2; (6.8); (9.2)
 Larg. losange nasal: 7.8, -, 7.1, 6.6; (6.3); (7.4 - 7.8)
 " zygomatique: 10.7, 10.3, 10.4, -; (8.6 - 9.5); (11 - 12.6)
 " mastoïde: 8, 8.1, 7.8, -; (7.2 - 8); (8.6 - 9.6)
 " C-C (externe): 4.6, 4.3, 4.5, -; (3.7); (4.7 - 5.2)
 " M³-M³ (externe): 6.9, 6.5, 6.8, -; (5.5); (7.4 - 7.7)
 Long, rangée dent. C-M³: 6.4, 6.1, 6.3, 6.1; (5 - 5.7); (6.7 - 7.3)
 " mandibule: 12.3, 12, 12.3, -; (9.8 - 10.7); (12.9 - 14)
 " rangée dent. C-M₃: 6.9, 6.4, 6.8, 6.7; (5.6 - 5.8); (7.3 - 8.2)

Nycteris labiata Heuglin, 1861

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Fissura frontis membranis utrinque quatuor ornata, quarum superiore biloba; labro trisulcato, labio bipartito; gastreo, pileo naribusque albidis; regione ophthalmica, fronte, cervice et dorso murinis; colli lateribus humerisque intense griseo-rubente indutis; auriculis pallide fuliginosis, minutissime nigro-marginatis; patagiis nigricantibus.

Auf der gespaltenen Nase vier Hautfalten; die Oberlippe als eine ihrem Rande parallele Querfalte aufgetrieben, welche jederseits in der Eckzahngegend durch eine Furche gespalten ist; die Unterlippe in der Mitte einfach getheilt und die ganze Schnauze dicht und ziemlich kräftig und lang behaart - Ohren sehr gross und breit, elliptisch, am Innenrand fein und mässig lang behaart; *Tragus* klein und gerundet, letzterer sowie die Aussenseite des Ohrenrandes mit kleinem Lappen; Gaumenfalten sechs, die vorderste ungeliebt, zwischen ihr und der nächstfolgenden in der Gaumenmitte eine sticknadelkopfgrosse Warze; Schwanz sehr lang, an der Spitze getheilt und mit den Gabelgliedern einen

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Theil des untersten Randes des *Patagium interfemorale* umfassend; die Lippen sind nackt fleischfarb, ins Gräuliche mit feinen violetten Punkten besetzt - Zahnsystem regelmässig; oben je zwei, unten je drei Schneidezähne. Die Länge der Ohren ist bei verschiedenen Individuen ziemlich verschieden: die mittlere durchschnittliche Länge 1" 1" französ. Mass auf 9" Breite; Körper 2" lang, Schwanz 2", Flugweite 10"-11".

Ein Exemplar zeigt auf Rücken und Schultern einfach mausgraue Färbung ohne Beimischung von röthlichgrau.

Nycteris macrotis Dobson, 1876

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Length (of an adult M preserved in alcohol), head and body 2".25, tail 2".4, head 0".85, ear 1".4, tragus 0".3 x 0".18, forearm 1".9, thumb 0".5, second finger 3".6, fourth finger 2".8, tibia 0".95, calcaneum 0".75, foot and claws 0".4.

Hah. Africa (Sierra Leone).

Type in the collection of the British Museum.

Nycteris madagascariensis G. Grandidier, 1937

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Vient de la vallée du Rodo; il y en a deux exemplaires dans nos collections. Bien que représenté par de nombreuses espèces dans toute la région chaude, de l'Ancien monde, des îles de la Sonde au continent africain, ce type de chauve-souris n'a pas encore été signalé à Madagascar; nos spécimens bien que présentant tous les caractères du genre, diffèrent des *Nycteris* connus jusqu'ici par leurs proportions, et la teinte de leur fourrure notamment de *N. hispida*, *javanica*, *arge* et *angolensis* avec lesquelles *N. madagascariensis* y a quelques points de ressemblance; nous devons cependant les considérer comme les types d'une espèce nouvelle.

Le pelage est très clair, gris brun sur le dos et gris ardoisé clair sur le ventre; les oreilles qui sont à peu près de la longueur de la tête et très développées sont glabres, le tragus est arrondi et très petit, les pieds sont grisâtres. La queue qui est très importante et qui est complètement prise dans la membrane, se termine par une petite vertèbre en forme de T et non pas en pointe comme chez la plupart des *Nycteris*.

Le crâne montre tous les caractères de ceux des *Nycteris* avec sa dépression naso-frontale qu'entourent des expansions osseuses foliacées; il est cependant remarquablement fort et puissant en, proportion des dimensions des deux exemplaires de nos collections qui sont des adultes approximativement de même taille, l'un est une femelle, peut-être en état de gestation.

Voici les dimensions moyennes de ces deux chauves-souris:

Envergure 260 mm.; longueur totale 99 mm.; corps 45 mm.; queue 54 mm.; hauteur de l'oreille 27 mm.; largeur de l'oreille 18 mm.; hauteur du tragus 3 à 4 mm.; avant-bras 51 mm.; partie inférieure de la jambe 24 mm.; pouce 16 mm.; 2^e doigt 63 mm.; 3^e doigt 85 mm.; 4^e doigt 70 mm.; 5^e doigt 71 mm.

Crâne: longueur totale maximum 22 mm.; largeur zygomatique extérieure 13.5 mm. ; longueur C. m³ 8 mm.; Mâchoire inférieure: longueur maximum 14 mm.; longueur C. m³ 8 mm.

Nycteris marica Kershaw, 1923

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Nycteris marica, sp. n.

A large bat, equal to *N. grandis* in size, but with a smaller skull.

Colour drab-grey above and below, rather darker in the posterior dorsal region. Wing-membranes broccoli-brown. Lower surface of wing-membranes on each side of the body thickly furred to a distance of about 20 mm. from the body.

In the skull the fourth lower premolar is much larger than in *grandis*, though not so large as in the *javanica* group.

Type. Adult. B.M. no. 23.8.2.1. Collected on 24th Jan., 1922.

Type-locality. Tindiga, Kilosa.

Measurements from the dried skin: -

Forearm 60 mm.; head and body 93; tail 70; ear 27;

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thumb 18.7; third finger, metacarp. 48, first ph. 32, second ph. 32; tibia and foot with claws 44.

Skull: greatest length to tip of canine 26; zygomatic breadth 16; breadth of frontal shield 10.2; p_4 comparatively large and standing in the tooth-row.

N. marica, a member of the *hispidia* group, is, with *grandis*, larger than any other species in the genus. It differs from *grandis* in colour and in the smaller skull with the larger p_4 .

Nycteris nana tristis G.M. Allen and Lawrence, 1936

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Type. Museum of Comparative Zoölogy, No. 31,156. An adult female, skin and skull, from Kaimosi, Kakamega district, Kenya Colony, collected by Arthur Loveridge, February 13, 1934.

Description. Compared with skins from Lolodorf, Cameroon, representing typical *N. nana* (type locality, Benito River, French Congo), the East African race lacks the warm russet coloration of the fur of both surfaces, and is instead a uniform dark drab gray both above and below. The fur is of the same color from tip to base except on the nape and especially about the bases of the ears, where it is slightly paler, a soiled grayish, basally. On the membranes the fur extends out on the propatagium from the axilla to about the end of the first third of the fore arm, and on the plagiopatagium to a line joining the elbow and the first third of the tibia. On the uropatagium the fur extends out as far as a line connecting the proximal ends of the tibiae. On the under side the extent is about the same, except that it does not quite reach the knees.

Measurements. The specimen has practically the same dimensions as those of the West African race. The type measures: fore arm, 35.8 mm.; tibia, 15.7; foot, 6.5; tail (about) 45; thumb, 11.5; third metacarpal, 28.5; first phalanx, 16.5; fourth metacarpal, 29.7; fifth metacarpal, 30.3 mm.

The skull measures: greatest length, 16.6 mm.; basal length, 12.5; palatal length, 3.6; zygomatic width, 9.3; mastoid width, 8.0; width across frontal plate, 6.6; width outside last molars, 6.2; upper cheek teeth, 5.3; lower cheek teeth, 5.8 mm.

Remarks. Through the kindness of Mr. J. Kenneth Doult, of the Carnegie Museum, we have had the loan of two specimens representing typical *Nycteris nana*, from Lolodorf, Cameroon. Both agree in their pronounced russet tint, contrasting with the dull gray hue of the

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eastern animal. It is a rather rare species, for, in addition to the original specimen from Benito River, it has apparently been recorded but twice: by Hollister, in 1918, who mentions two in the United States National Museum from Yala River, Kenya Colony, as forming a considerable extension of the known range into eastern Africa, and again by Cabrera and Ruxton (1926, Ann. Mag. Nat. Hist. (9), 17, p. 591), who had a specimen from Luluabourg, Belgian Congo, that flew into a room. The Cameroon specimens, referred to above, are now recorded for the first time.

Nycteris oriana Kershaw, 1922

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Nycteris oriana, sp. n.

F. 57, 312. Chiromo.

A member of the *aethiopica* group, with long ears and tail, approaching *N. luteola*, Thos., in size.

The body is clothed with long soft hairs, a dirty white colour on the dorsal surface for the greater part of their length, tipped with brown, the general result being a light pinkish brown. The hairs of the ventral surface are ecru-drab throughout their length.

The edge of the wing-membrane from the ankle for about 21 mm. is fringed with light-coloured hairs. The shape of the tragus is as that figured by Dobson in the 'Catalogue of Chiroptera' for *N. macrotis*.

Type. Female. B.M. no. 22.4.2.5.3. Original number 312. Collected on June 7th, 1918, and presented by Mr. Rodney C. Wood.

Type-locality. Chiromo, Shiré Valley, Nyasaland.

Dimensions of the type: -

Forearm 53 mm.; head and body 67.5; tail 63; ear 33; thumb 15.2; third finger - metacarp. 44, 1st ph. 24, 2nd ph. 29; fourth finger - metacarp. 42.5, 1st ph. 16, 2nd ph. 15; fifth finger - metacarp. 45, 1st ph. 15, 2nd ph. 16.2; tibia 25; foot 11; tibia and foot (including claws) 37.

Skull: greatest length to tip of canine 22.3; zygomatic breadth 13.7; breadth of frontal shield 8.4; length of upper tooth-row 8.1; p_4 minute and internal to the tooth-row, not in it, as in *N. æthiopica* and *N. luteola*.

N. oriana does not appear to be nearly related to any of the species in the *æthiopica* group. In size it approaches *N. luteola*, Thos. (which Mr. Thomas now agrees should be elevated to specific rank), but differs widely from it in other respects, such as in the length of the fur (13 mm. in *oriana*,

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9 in *luteola*), and in its colour and texture, the dimensions of body and skull, the heavier dentition, the position of p_3 , and the much larger frontal shield.

[*Nycteris pallida*](#) J.A. Allen, 1917

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(Type, No, 49144, M ad., Faradje, Belgian Congo, March 1, 1912; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 1858.

A pale small form of the *N. hispida* group, differing strongly from typical *hispida* in its pale coloration and also slightly in smaller size.

Above, pale brown, the tips of the hairs on the lower back and sides pale fulvous; underparts grayish brown; head back to the ears very pale brown, in strong contrast with the body; a narrow whitish band bordering the nose pad in front and laterally; ears light brown terminally, much paler basally; membranes much paler than in *hispida*.

Collectors' measurements of type: Total length, 92; tail, 45; foot, 10; ear, 21 (in skin from anterior border, 15.5),

Additional measurements from type; Forearm, 39.5; tibia, 19; foot, 9.2. Forearm in 5 alcoholics (Faradje, 3, Vankerckhovenville, 2), 37.8 (36.5-39.5).

Skull (type), total length, 17.4; zygomatic breadth, 10.2; upper toothrow (c-m³), 6.5; lower jaw, 11.8; lower toothrow, 6.8. Type and three topotypes, total length, 17.2 (16.9-17.4); zygomatic breadth (2 skulls), 10.2 (10.1-10.3).

Represented by 12 specimens (3 skins, 9 alcoholics, part immature), of which 8 are from Faradje, and 4 from Vankerckhovenville.

Present material indicates that *N. pallida* differs from *N. hispida*, its nearest ally, in much paler coloration, including especially the ears and membranes, and smaller size, averaging about 4 mm., less in the length of the forearm, with other measurements proportional. *N. pallida* is thus much smaller than *N. aurita* Andersen.

[*Nycteris poensis*](#) Gray, 1843

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Fernando Po Nycteris.

a. Africa, Fernando Po. - Presented by Capt. Edw. Downes, R.N.

[*Nycteris proxima*](#) Lönnberg and Gyldenstolpe, 1925

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A member of the *N. hispida*-group approaching in some respects to *N. grandis* PETERS, but differing in several dimensions.

Upper incisors trifold, p_4 in the tooth-row, smaller in size than the corresponding tooth of *grandis* but similar in shape to the same. The colour (in spirit) appears to be comparatively dark, a little paler below. Membranes and ears also dark.

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Type: M ad. coll. at Kartoushi, Sembiliki valley, ¹¹₃ 1921 (orig. n:o 813).

Dimensions: forearm 58; third finger, metacarpal 38; third finger, first phalanx 27; tibia and foot with claws 39; tibia 25; calcaneum 19; tail 57; ear 30 mm. Tragus very hairy, almost lingulate, broadly rounded at the tip and of almost equal breadth along its whole length.

Skull: Greatest length to tip of canines 24; condyles to canine 20.5; zygomatic breadth 13.5; breadth of frontal shield 7; upper tooth-row, $c-m^3$ 8.9, length of mandible 16.5; length of lower tooth-row, $c-m_3$ 10 mm.

From *Nycteris avakubia* ALLEN, 1917, the new species differs by its considerably longer forearm, somewhat shorter metacarpal of third finger and considerably shorter tail as well as its generally larger cranial dimensions. *N. grandis* PETERS and *N. marica* KERSHAW, 1923, are larger in every respect, the remaining species of this group are on the other hand much smaller.

Nycteris Revoilii Robin, 1881

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Oreilles beaucoup plus longues que

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la tête; tragus élargi en spatule à l'extrémité présentant un lobe externe arrondi, deuxième prémolaire inférieurs très petite et située dans la rangée dentaire; tête raccourcie (longueur 20^{mm}, largeur 12^{mm}); troisième phalange cartilagineuse du 5^e doigt ligé du côté externe; incisives supérieures séparées sur la ligne médiane. Couleur gris à peine centré sur le dos, blanchâtre sur le ventre.

Longueur totale 63^{mm}, queue 51^{mm}, bras 19^{mm}, avant-bras 44^{mm}, cuisse 22^{mm}, jambe 22^{mm}.

Nycteris Thebaïcus E. Geoffroy Saint-Hilaire, 1818

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NYCTERE DE LA THEBAÏDE. *Nycteris thebaïcus*.

Planche I, N.° 2.

Les nyctères forment un genre très-différent des deux précédents.

C'est encore le même nombre d'incisives, mais non la même disposition: plus petites, sur-tout les inférieures qu'on distingue à peine à la vue simple, elles ne sont plus en haut (comme dans les makis) écartées par paire, mais garnissent, au contraire, sur une ligne continue, tout le bord de l'intermaxillaire.

Cet os, subordonné aux variations de l'organe de l'odorat, quoiqu'appuyé sur les maxillaires, jouit d'un mouvement propre; il est soulevé ou abaissé, oscillant comme sur un axe, par la lèvre supérieure, qui est d'une épaisseur et d'une consistance propres à l'entraîner: aminci à ses points d'articulation, il ne pouvoit participer à la fixité de toutes les autres parties osseuses.

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C'est sans doute parce que l'intermaxillaire est ainsi maîtrisé par les organes qui l'entourent, qu'il est très-petit: il ne fait pas de saillie au-delà des canines d'où il arrive que la mâchoire supérieure est plus courte que l'inférieure, et paroît comme tronquée; il en résulte aussi que les incisives des deux mâchoires ne se correspondent pas, et que posant à faux, elles n'usent point leurs sommets, lesquels restent à deux lobes en haut et à trois crénelures en bas.

Si l'on commence par examiner dans le crâne les fosses nasales des nyctères, on les juge d'abord sans profondeur, parce que les planchers qui en circonscrivent l'étendue sont très-bornés: le plancher inférieur ou la lame palatine ne se prolonge pas au-delà de la deuxième molaire, et l'externe ou les nasaux maxillaires sont des pièces réduites à des dimensions rudimentaires. Mais on prend, au contraire, une autre opinion de ces fosses nasales, en les voyant recouvertes de leurs parties molles. Les arrières-narines s'ouvrent beaucoup au-delà du point où se termine l'os maxillaire; et les méats extérieurs ont leurs larges entrées remplies et pour ainsi dire encombrées de lobes et d'appendices cutanés: un repli du derme naît du milieu de chaque conduit. On diroit que les conques nasales, en saillie chez les vespertiliens, et dans une cavité chez les nyctères, ne sont devenues aussi voisines, et ne sont ainsi descendues dans une sorte d'en tonnoir, que parce qu'elles auroient été contractées, repliées sur elles-mêmes et tirées à travers le crâne. Un lobe qui a la forme d'une tête de clou, et qui n'est autre que le cartilage de la narine, se voit de chaque côté, et concourt comme opercule, avec le repli intérieur, à fermer hermétiquement l'orifice nasal.

Il n'est pour cela besoin d'autre effort de la part de l'animal que de froncer toutes ces parties, et peut-être même de les abandonner à leur élasticité naturelle.

La cavité des narines se prolonge en arrière sur le chanfrein; première circonstance déjà remarquable. Mais ce qui ne l'est pas moins, c'est la grandeur et la forme canaliculée de cette dernière partie: elle donne aux nyctères cette physionomie sombre et farouche qui les caractérise.

Le chanfrein s'étend en effet au-delà de ses dimensions habituelles, et ce, au moyen de lames osseuses qui naissent des côtés de l'os coronal, et se réunissent au vertex: le canal ou fente longitudinale qui résulte de la saillie de ces crêtes, verse sur les narines; seule relation, en dernière analyse, que ces parties aient entre elles.

Cependant le chanfrein aurait-il subi ces étranges métamorphoses pour suppléer à la petitesse extrême des ouvertures nasales, et seroit-il une sorte d'entonnoir où se recueilleraient les fluides odorans? Les bords de la fente sont hérissés de poils longs et abondants qui la remplissent; mais ce n'est pas quand les muscles labiaux soulèvent les opercules, détendent les plis intérieurs et entr'ouvrent les conduits nasaux: ces bords, par la tension de la peau, sont ramenés en dessus, et avec eux les longs poils qui les garnissent.

Des narines qui sont habituellement fermées, et qui, pour entrer en communication avec les corps ambiants, exigent la volonté de l'animal et le jeu de quelques

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quelques muscles, fournissent sans doute une considération intéressante en elle-même.

Les nyctères ne peuvent manquer d'en tirer avantage; et il se trouve, en effet, qu'ils établissent leur demeure en des lieux d'où de fortes exhalaisons repousseroient d'autres animaux. Mais que la disposition des conduits nasaux son dans un ordre inverse pour les soustraire ainsi aux inconvénients d'odeurs infectes, c'est ce que je n'ai pu croire. Cet arrangement suppose ailleurs une autre modification, et j'ai dû m'en proposer la recherche.

Le vol des chauve-souris a souvent ramené à l'idée de les comparer aux oiseaux; et l'on a trouvé que ceux-ci se distinguoient toujours par plus d'aisance et de grâces dans les allures, parce qu'indépendamment de plus de perfection dans les organes directs du vol, ils jouissent encore de la faculté de se gonfler d'air et de se rendre plus légers. En s'exprimant ainsi, on étoit loin de penser qu'on retrouveroit la même faculté dans les chauve-souris, dont en effet les fonctions pulmonaires sont si différentes de celles des oiseaux.

C'est toutefois ce que les nyctères m'ont montré, des vésicules aériennes semblables, encore plus grandes, et que l'animal remplit, quand il le veut et autant qu'il le veut. Mais, comme on le pense bien, les nyctères y portent l'air en vertu d'un mécanisme particulier, et au moyen d'une organisation qui, dans ses anomalies, dérive néanmoins du plan primordial et classique des mammifères.

On pressent peut-être déjà les résultats d'un mode si nouveau d'organisation; les moyens qui les donnent, sont d'une simplicité parfaite.

La peau n'a d'adhérence au corps qu'en quelques endroits, où elle est retenue par un tissu cellulaire très-lâche et très-écarté: l'air s'y introduit, et en séjournant ainsi, comme on le dit, entre cuir et chair, donne à l'animal l'apparence de ces veaux soufflés dans les boucheries. Il n'y a de brides aponévrotiques ou de tissu cellulaire que dans le voisinage des méats et sur les côtés du tronc: ainsi la peau se soulève entière sur le dos, à la poitrine et à l'abdomen; ce qui met les nyctères dans un bain d'air, ou, si l'on veut, dans une sorte de manchon que leur forme ce fluide élastique.

Jusque-là, quelque extraordinaire que soit un pareil fait, on ne voit pas qu'il soit en rien dérogé à l'essence du type des mammifères: il n'y est pas dérogé davantage quant aux moyens de souffler cette unique, mais bien vaste cellule.

Au fond de chaque abajoue est une ouverture de deux millimètres de large; et c'est tout simplement par-là que le sac aérien communique avec la bouche.

L'animal en ouvrant ses naseaux fait que l'air ambiant entre et gonfle sa poitrine: en abandonnant, au contraire, un moment après, toutes les membranes nasales à leur élasticité propre, et en tenant simultanément la bouche close, il force le gaz expiré à se rendre dans les abajoues, et de là dans le grand sac aérien.

Quoiqu'il y ait, à l'entrée de ce sac, un sphincter très-apparent, ce n'est pas lui, ou lui seul du moins, qui s'oppose au retour de l'air: il y a de grandes valvules, sur le cou et le dos, qui en sont chargées.

L'air ne suit de route qu'à partir du sphincter: il se rend, en passant,

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au-devant de l'oreille, dans le sinus du chanfrein, d'où il gagne le vertex, l'occiput, et le col supérieur: c'est là qu'il est versé dans le grand sac.

Ainsi, le nyctère se conduit exactement comme le tétrodon; il porte, à volonté, une gorgée d'air dans son sac, puis une seconde, et ainsi de suite. Il souffle comme nous pouvons le faire nous-mêmes, et de la même manière, avec cette seule différence qu'il souffle dans sa bouche, dont il tient la cavité sans issue à l'extérieur. Sa peau devient une véritable vessie, au-dedans de laquelle le tronc se trouve comme déposé. Les nyctères agissent presque à son égard de même que si elle étoit un hors-d'œuvre, puisqu'ils la remplissent au point de lui faire prendre une forme sphérique. Dans cet état, tout l'animal ressemble à un ballon auquel on auroit attaché des ailes, une tête et des pieds.

Plus heureux que le tétrodon, qui ne recourt à la même industrie qu'en se réduisant à n'être plus qu'une masse inerte sur le miroir des eaux, il conserve toutes ses facultés, ou mieux il en augmente l'énergie, en devenant plus léger et susceptible de plus de vitesse dans le vol.

J'avois cru apercevoir que les étranges anomalies des conduits olfactifs peseroient sur un autre système d'organe, et occasionneraient peut-être ailleurs d'autre changemens; et il se trouve en effet qu'un grand sac modifié, dans les nyctères, ou plutôt procure à leur organe respiratoire, un précieux appendice. Si cet appareil, qui est si bien adapté à ce système, n'est pas le motif des modifications des fosses nasales, et n'en donne pas une explication entièrement satisfaisante, du moins on ne sauroit nier qu'il n'y ait entre toutes ces parties des relations réciproques et nécessaires.

C'est aux différences que je viens de signaler que se borne l'énoncé des caractères distinctifs des nyctères: les dents canines et molaires de ces chauve-souris ressemblent à celles des vespertiliens; il en est de même des viscères abdominaux.

Les tégumens offrent seulement plus d'étendue; les oreilles sont plus longues que la tête, sans que l'oreillon qui borde aussi le méat auditif soit agrandi en même proportion. Cette étendue se fait sur-tout remarquer entre les jambes, ou la membrane caudale surpasse dans ses deux sens la longueur de l'animal.

La dernière vertèbre de la queue est bifurquée; séparation singulière, puisqu'elle se trouve dans tous les nyctères, et n'existe dans aucun autre genre de chauve-souris.

On n'a fait mention que d'une seule espèce de nyctère, le campagnol-volant de Daubenton, dont Linnéus a fait son *Vesp. hispidus*. Le nyctère de la Thébaïde en diffère, ainsi qu'une autre espèce qui a été rapportée de Java, et qui m'a été remise par M. Leschenault.

Les dimensions de ces chauve-souris forment un de leurs traits distinctifs: le nyctère de Daubenton a trente-huit millimètres de long, de la tête à la naissance de la queue; le nyctère de la Thébaïde cinquante-quatre, et celui de Java soixante-sept.

L'oreille a plus d'ampleur dans l'espèce d'Égypte, et le poil y est non moins long et touffu.

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Le pelage du nyctère de la Thébaïde est brun-clair en dessus, et cendré en dessous: c'est presque la même teinte dans le nyctère de Daubenton; mais elle passe davantage au roux sur le dos, et à un blanc sale sur le ventre, où se voit aussi un mélange de fauve: l'espèce de Java a les parties supérieures d'un roux-vif, et le poil inférieur cendré-roussâtre.

Le nyctère anciennement décrit avoit été rapporté du Sénégal: ainsi tout le genre habite les contrées chaudes de l'ancien continent.

Je présume qu'il en existe deux espèces au Sénégal; du moins Daubenton en a décrit deux variétés qui lui avoient toutes deux été données par Adanson: la seconde, qu'il ne constata que sur un individu desséché (voyez H. N. G., tome X, page 91), différoit de la première en ce que la couleur blanchâtre du dessous du corps étoit mêlée d'une teinte de cendré, et que la membrane des ailes n'avoit point de roussâtre.

J'ai sous les yeux le crâne et les principales parties osseuses du même individu; et ces parties ne s'accordent, ni pour les dimensions plus fortes, ni pour quelques détails de forme, avec les os, dans les nyctères de Daubenton et de la Thébaïde.

Nycteris villosa Peters, 1852

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Tafel XL.

N. colore corticinofusco, subtus ex griseo fusciscente; alis fuscis; auriculis longitudine capitis, retro exisis, disjunctis; dentibus primoribus superioribus distincte trifidis; alis supra multo latius quam infra villosis.

Longitudo tota 0,094; caudae 0,44; antibrachii 0,038; volatus 0,235.

Habitatio: Africa orientalis, Inhambane, 24° Lat. Austr.

Bei dieser kleinen ausgezeichneten Art sind die Ohren im Verhältnifs zu denen bei den andern Repräsentanten dieser Gattung sehr kurz, kaum so lang wie der Kopf; ihr äufserer

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Rand zeigt einen flachen Ausschnitt, und ihre inneren Ränder sind auf der Stirn nicht, wie bei allen andern Arten, durch eine Hautfalte miteinander verbunden; die Ohrklappe, welche ziemlich tief sitzt, ist an ihrer oberen Hälfte mit langen Haaren besetzt, verlängert, doppelt so lang wie breit. Der Nasenbesatz und die Bildung der Lippen zeigen nichts von den andern Arten abweichendes. Am Gaumen befinden sich sechs quere Schleimhautfalten; die erste ist grade und liegt zwischen den Eckzähnen; die zweite, welche V förmig ihre Mitte nach hinten in einem spitzen Winkel zurückgezogen hat, liegt zwischen den Lückenzähnen; die drei folgenden bilden in der Mitte einen stumpfen nach hinten gerichteten Winkel und haben ihre Lage zwischen dem ersten Paar der wahren Backzähne; die sechste und letzte grade erhebt sich zwischen den vorderen Enden des zweiten Paares der wahren Backzähne.

Die Behaarung des Rückens ist beträchtlich länger als die der Bauchseite, und ein dichter Flaum setzt sich viel weiter als bei irgend einer andern Art über die Flughäute fort, so daß die Schenkelflughäute bis zwischen den ersten zwei Fünfteln des Vorderarms und dem Unterschenkel, und die Schenkelflughaut über die Hälfte wollig erscheint. Auf der Bauchseite dehnt sie sich dagegen nicht so weit aus, während umgekehrt bei den andern bekannten Arten die Behaarung der Rückenseite die am wenigsten ausgedehnte ist. Die Bildung der Flughäute und Gliedmaßen ist dieselbe wie bei anderen Arten. Von den sehr langen Schwanzgliedern sind die beiden ersten von gleicher Länge, und die vier folgenden nehmen progressiv an Länge ab. Die Gabelung des Endknorpels übertrifft die Länge des damit verbundenen letzten knöchernen Gliedes.

Die Farbe der Oberseite ist rufsig nelkenbraun, und die Haare sind hier in der Mitte weißlich, an der Basis schwarzbraun gefärbt. Die Bauchseite ist bräunlich weißgrau, die Basis der Haare ebenfalls schwarzbraun. Die Ohren sind an ihrer äußeren Fläche bis zur Hälfte mit einer weißlichen Wolle bekleidet. Der Haaranflug der Bauchseite der Flügel ist weißlich. Die Flughäute und Ohren sind dunkelbraun, die Nägel weiß.

Der Schädel zeigt in seiner Form nichts Abweichendes. Die vier oberen Schneidezähne stehen zusammen, und sind, was ich bei keiner andern Art bemerken konnte, dreilappig, größer als die sechs unteren, welche quer zum Rande des Kiefers gestellt sind. Der zweite untere kleine Lückenzahn ist mehr entwickelt wie bei den andern mir bekannten africanischen Arten und steht in der Reihe zwischen dem ersten Lückenzahn und dem ersten Backzahn. Die Zahl der Wirbel beläuft sich auf 35; darunter sind 7 getrennte Halswirbel, 10 Brustwirbel, 6 Lendenwirbel, 5 kurze Kreuzbeinwirbel und 7 Schwanzwirbel nebst dem knorpeligen gespaltenen Anhang. Das Brustbein, welches aus drei Stücken besteht, ist an seiner vorderen Seite durch einen Längsriegel ausgezeichnet, welcher bis auf die Pars xiphoidea herabgeht. Es befestigen sich fünf Paar Rippen an dasselbe, während die übrigen fünf als falsche Rippen zu betrachten sind. Der Magen ist sackförmig und geht in einen einfachen 85 mm. langen Darm über. Die Leber zerfällt in drei Hauptlappen, in einen linken größeren und zwei rechte kleinere: zwischen und unter den letzteren liegt die birnförmige Gallen-

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blase. Die Milz, welche hinter dem linken Ende des Magens liegt, hat eine Länge von 6 Millim.; eine Breite von $1\frac{1}{4}$ Millim. Von den Nieren ist die linke größer als die rechte, $5\frac{1}{2}$ Millim. lang und 3 Millim. breit, von einfacher, bohnenförmiger Gestalt. Der ganze Kehlkopf ist nur zwei Millimeter lang; der Schildknorpel ist an seinem vorderen Winkel, der Ringknorpel längs der Mitte seiner hinteren Seite stark gekielt. Die Luftröhre bildet unter dem Kehlkopf eine ringförmige Auftreibung, und enthält bis zur Theilungsstelle 24 knorpelige Halbringe. Die Lungen bestehen jederseits aus einem einzigen, unregelmäßig quadratischen Lappen. Das Herz hat eine eiförmige Gestalt, eine Länge von 7 und eine Breite von 4 Millimetern.

Maße in Millimetern.

Ganze Länge in krummer Linie von der Schnauze bis zur Schwanzspitze: 94

Flugweite: 235

Länge des Kopfes: 18

Höhe des Ohres: 17

Breite des Ohres: 13

Länge des Oberarms: $15\frac{1}{2}$

Länge des Vorderarms: 38

Länge des Daumens (Mittelh. 5. 1.Gl. $4\frac{1}{2}$, 2.Gl. 2): $11\frac{1}{2}$

Länge d. 2ten Fingers (Mittelh. 34. 1.Gl. 1): 35

Länge d. 3ten Fingers (Mittelh. 29. 1.Gl. $21\frac{1}{2}$, 2.Gl. $22\frac{1}{2}$, 3.Gl. 3): 76

Länge d. 4ten Fingers (Mittelh. 32. 1.Gl. 11. 2.Gl. 8. 3.Gl. $1\frac{1}{2}$): $52\frac{1}{2}$

Länge d. 5ten Fingers (Mittelh. $30\frac{1}{2}$. 1.Gl. $11\frac{1}{4}$. 2.Gl. $9\frac{1}{2}$. 3.Gl. $1\frac{3}{4}$): 53

Länge des Oberschenkels: 18

Länge des Unterschenkels: 19

Länge des Fußes: 9

Länge des Schwanzes: 44

Länge des Sporns: 16

Länge des Schädels: 17

Länge der Wirbelsäule vom Atlas bis zum ersten Schwanzwirbel: 30

Alle anderen bekannten Arten von *Nycteris* unterscheiden sich von dieser durch bedeutendere Körpergröße, viel längere Ohren, geringere Behaarung der Flughäute und endlich durch die zweilappige Form der oberen Schneidezähne.

Das Material für diese Beschreibung besteht in einem einzigen Exemplar, einem Männchen, welches in Inhambane, im 24° Südl. Br. gefangen wurde. Die Nahrung besteht dem Inhalte des Magens und Darms nach, ausschließlich aus Insekten.

Nycteris vinsoni Dalquest, 1965

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Type. - Adult female, skin and skull, no. 18739, Walter W. Dalquest, Midwestern University Collection, taken on the south bank of the Save River, 212 km SSW Beira, Mozambique.

Diagnosis. - A very large *Nycteris*, similar in general proportions to *Nycteris thebaica* but larger and stouter. Forearm more than 50 mm and length of skull from canines more than 21 mm. Ears relatively short, probably less than 25 mm in life.

Comparisons. - This species needs comparison only with *Nycteris thebaica*, the only species of *Nycteris* ranging into southern Africa that is nearly as large. Fortunately two *Nycteris thebaica capensis* were taken in a hollow tree a few hundred yards from the site where the type and paratype of *N. vinsoni* were found. These *N. thebaica* are apparently typical and their

measurements agree closely with those given by Roberts (1951). *N. vinsoni* differs from *N. thebaica* principally in its much larger size, more massive skull and heavier dentition but shorter ears. Both *N. thebaica* are in the gray color phase, while the type of *vinsoni* is in the orange phase. However, the paratype of *vinsoni* is of the gray phase.

The size difference between *thebaica* and *vinsoni* is too great to be individual variation within a species. The rostrum of *vinsoni* is 20 % heavier than that of *thebaica*, and the superorbital flanges overhang the orbits to a much greater degree; breadth across the flanges in *thebaica* is 6.8 mm versus 9.0 mm in *vinsoni*.

Measurements. - External measurements of the type are: total length, 125; length of tail, 55; length of hind foot, 13; height of ear from notch, 22. Cranial measurements are

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those used by Roberts (1951): length of skull from canines, 22.0; frontals to occiput, 17.3; zygomatic breadth, 13.3; braincase breadth, 10.0; width across upper M3's, 8.8; width across upper canines, 5.8; canine-upper M3 length, 7.8; height at bullae, 10.3; mandible length plus incisor, 15.0; canine to lower M3, 8.4; forearm length, 50.6. Every measurement exceeds, usually by a considerable margin, the maximum given by Roberts for *N. (thebaica) capensis*, as well as the measurements of the specimens of *N. thebaica* from the Save River.

Remarks. - The type and paratype were occupying a small hollow, not more than 6 inches in diameter, at the base of a large baobab tree. A fire was built at the base of the hollow and two bats were stupified by the smoke. One, in the gray color phase, fell into the flames and was so singed the skin could not be saved. The ears and membranes of the other, the type, were somewhat shrivelled. It is likely that the height of the ear in the living bat was greater than the 22 mm recorded, although some allowance was made for the shrivelling of the ear. In any event, the ear was much shorter than that of the *N. thebaica* (35 mm) taken nearby.

This species is named for Mr. Jerry Vinson, the sponsor of the Save River expedition.

Nycteris woodi K. Andersen, 1914

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A member of the *N. aethiopica* group (see Ann. & Mag. N. H. (8) x. p. 549, Nov. 1912), differing from the other representatives of the same group by its much smaller size and relatively longer ears, and from all other forms of the genus by having the fur of the underparts pure white, without any trace of darker bases to the hairs.

Forearm 42.5 mm.; ear from base of inner margin (relaxed) about 29. Skull, total length to front of canine 18.2; condylo-canine length 15.8; maxillary tooth-row (crowns) 6.

Type, skin and skull of an adult, Chilanga, N.W. Rhodesia, 4100', Nov. 1913, presented by R. C. Wood, Esq. B.M. 14.4.22.2.

Nycteris woodi sabiensis Roberts, 1946

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Differs from *Nycteris woodi* K. Andersen (*Ann. Mag. Nat. Hist.* (8), XIII, 563; 1913: Chilanga, Northern Rhodesia) in its longer ears (33 - 34 mm. as compared with 29) and in colour, having the underparts of body greyish white, browner on the sides, instead of pure white.

Measurements of the type: Head and body 49, tail 49, hind foot c.u. 9; ear 35, Forearm 40.5; 3rd finger metacarpal 33; 1st phalanx 21.5; 2nd phalanx 25; 4th finger metacarpal 35; 5th finger metacarpal 36. Tibia and foot (c.u.) 30.5. Skull, greatest length from canines 18 (in *woodi* 18.2); c-m³ series 6 (in *woodi* 6).

Type: T.M. No. 8578, adult F, Birchenough Bridge, Southern Rhodesia. Also five others, adult and immature, collected at the same time.

Nycterops Gray, 1866

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NYCTEROPS. Ears separate, as long as the head. Africa.

N. pilosa

Nycticeius africanus G.M. Allen, 1911

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Type. - Skin and skull, male, no. 8272, M.C.Z., collected August 11, 1909, on the Meru River, British East Africa, by Glover M. Allen.

General characters.- A typical *Nycticeius*, slightly smaller and much paler than the *N. humeralis* of the southeastern United States, with shorter closer fur. Postcalcarian lobe well developed.

Description. - Fur of the upper surface of head and body, short and close, about 4 mm. long on the lower part of the back, of a uniform pale wood-brown quite to the bases of the hairs; below cream-buff

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to the bases of the hairs. The muzzle in front of the eyes is nearly naked.

Wing membranes dark, and attached at the base of the first digit of the foot. Ears pale brownish. Hair of the body does not extend on to the interfemoral membrane. Calcar well developed with a conspicuous, low, rounded lobe at about 3 mm. behind the heel. Tragus with a distinct external basal lobe." Tip of tail free from the interfemoral membrane.

Skull. - The skull of the type is scarcely to be differentiated from that of the American species, *N. humeralis*, except by its slightly smaller size. The upper incisors, however, are set much more nearly at right angles to the long axis of the skull, so that viewed from above they hardly project beyond the premaxillae, whereas in *humeralis* they extend prominently forward.

Measurements. - The following measurements were taken in the flesh: total length, 85 mm.; tail, 35; hind foot, 6.5; tibia, 13; calcar, 17.5; ear, 12; forearm, 33. Third metacarpal, 29.7.

Skull: greatest length, 13.6; basal length, 10; palatal length, 5; interorbital width, 4; zygomatic width, 9.4; mastoid width, 8; upper cheek teeth, front of canine to back of m³, 4.5; lower cheek teeth, 5.3; mandible, 10.2.

Remarks. - The discovery in East Africa of this genus hitherto known only from the southeastern United States and Cuba is one of the most important and interesting results of our expedition. Mr. Gerrit S. Miller, Jr., who examined the specimen, first called my attention to the fact that it is in all respects a typical *Nycticeius*, agreeing with the American species in all essential characters. The postcalcarian lobe is well developed in the African but scarcely at all in the American bat, and the incisors of the former are less projecting, but otherwise they differ hardly at all except in color and size.

The single specimen on which this species is based was shot shortly after sunset as it was circling about in an open space among the ivorynut palms by the Meru River. The river here runs through a dry plain, and no doubt the pale coloration, of the species is due to the arid conditions under which it lives.

From a zoögeographic point of view the discovery of this genus in Africa is of extraordinary interest. Its case is somewhat paralleled by that of *Mormopterus* which occurs in Cuba, South America, Africa, and Madagascar. Possibly *Nycticeius* may yet be found to occur in South America.

Nycticeius eriophorus Heuglin, 1877

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Nycticeius eriophorus, Heugl. - Fitz. u. Heugl. Säugethiere p. 11.

Klein; die Ohrspitze scharf; der äußere Ohrtrand bis tief unter den Mundwinkel herablaufend und hier eine scharfe Ecke bildend; Tragus schmal, lanzettförmig, fast so hoch als die Ohrspitze; 8 bis 9 Gaumenfalten. Behaarung reichlich, lang, eigenthümlich gekräuselt; Oberseite hell erdgrau mit weißlichen Haarspitzen; Unterseite schmutzig weiß, Weichen mehr ins Braungrauliche; Flughaut braunschwärtlich. Körperlänge 1" 6". Kopflänge gegen 7". Außenrand des Ohres 6" hoch. Schwanz 1" 3". Vorderarm 13". Mittelfinger 2" 4 $\frac{1}{2}$ ". Der vierte Finger 1" 11 $\frac{1}{2}$ ". Der fünfte Finger 1" 10".

Von dieser Art sammelte ich drei Exemplare im Belegaz-Thal zwischen Semién und Wogara in Abessinien ine. Sie befanden sich im Hängenest einer SperlingS ober WeBervogelart.

Nycticeius flavigaster Heuglin, 1861

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Supra brunneo-olivaceus, subtus pure sulfurcus; capitis lateribus nitide pistacino tinctis. Unguinibus podii postici virente griseis, apice intense-flava.

Das Ohr ist 7" hoch, ziemlich breit und gerundet, mit an der Basis wagrecht abgeschnittenem Innenrande und tief herab und nach vorne gezogenem Aussenrande; hier hat letzterer einen halbkreisförmigen Lappen mit umgeschlagenem Rande. Der sichelförmige, 3 $\frac{3}{4}$ " hohe Tragus nach vorne gerichtet, mit halbzirkelförmigem Ausschnitt oder Klappe an der Oeffnung des Gehörganges.

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In der Ohrmuschel 5 - 6 undeutliche Querfalten. Nasenkuppe etwas aufgeworfen, die Nasenlöcher etwas röhrenförmig mündend und nach vorne und seitlich gerichtet; die Unterlippe schwach gespalten; die Gegend zwischen Auge und Nase aufgetrieben, breit und stumpf, mit verhältnissmässig gröberen wenigen Haaren besetzt.

Zwei eckzahnartige Schneidezähne an dem Zwischenkiefer. Die vordere Spitze des ersten oberen Backenzahns auch kegelförmig und sehr verlängert.

Das unterste Daumenglied und der Sporn des Hinterfusses haben nach Aussen zu einen Hautrand.

Ohrmuschel schwärzlich-braun, fast nackt, an der Basis des Aussenrandes mit sehr feinen grünlichen Haaren bekleidet. Lippen schwärzlich fleischfarb; Flughaut fast schwarz und an der Innenseite des Oberarms, wie auf verschiedenen Partien der ganzen Innenseite mit feinen grünlich-gelben Härchen. Auffallend sind die licht-grüngelben Nägel der Füsse.

Am Schwänze, in den die Seitenfittige spitz verlaufen, nur das äusserste Glied, sowie eine feine, scheinbar nur aus Hautmasse bestehende Spitze frei. Diese freien Theile messen zusammen nur $2\frac{1}{4}$ " . - Körperlänge 2" 10". Schwanz 1" 9". - Flugweite 13". Ohrenlänge 7".

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??

The ear is 7" long, rather wide and rounded, with an on the base horizontally cut off inner margin and deep down and forwards drawn outer margin; the latter has a semicircular lobe with a turned edge. The sickle shaped, $3\frac{3}{4}$ " high tragus is pointed forwards, with as semicircular cutting or flap on the opening of the auditory canal.

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In the pinna there are 5 - 6 unclear ridges. Top of nose slightly raised, the nostrils somewhat pipeshaped ending and forwards and sideways pointed, the lower lip weakly split; the area between eyes and nose vigorous [?], wide and blunt, with proportionally larger fewer hairs.

Two canine like incisors in the middle jaw [actually "between jaw"]. The anterior tip of the first upper molar also cone shaped and extremely elongated.

The lower thumb part and the spur of the feet have on the outside a membrane.

Pinna blackish-brown, almost naked, on the base of the outer margin with very fine greenish hairs covered.

Lips blackish fleshcolour; wing membrane almost black and the inside of the upper arm, like various parts of the entire inner side with fine greenish-yellow small hairs. Remarkable are the light green-yellow nails of the feet.

On the tail, in which the "Seitenfittige" [???] tips develop, is only the extreme vertebra, as well as a fine, apparently consisting of only skin mass, tip free. These free parts together measure only $2\frac{1}{4}$ " - Length of body 2" 10". Tail 1" 9". Wingspan 13". Length of ear: 7"

[Translated by Victor Van Cakenberghe, 2008]

Nycticejus leucogaster Cretzschmar, 1826

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Weißbäuchiger Nachtflatterer.

Diagnos. *Nycticejus facie nuda*; frontis, verticis et corporis supra colore lucide fuscescente, toto gastraco albescente.

Ausmessungen.

Fuß. Zoll, Lin.

Länge von dem Scheitel bis zu dem Schwanz: - 2 2

Länge des Schwanzes: - 1 3

Länge des Kopfes: - - 9

Länge der Ohren: - - 5

Flügelweite: - 10 5

Beschreibung.

Das von Raffinesque aufgestellte Geschlecht *Nycticejus* *) ist eine sehr wohl begründete, durch die Gleichförmigkeit in Schädel- und Zahnbildung aller Arten natürlich sich aufwerfende, vom *Genus vespertilio* zu trennende Gattung. Wir haben daher, um diese Ueberzeugung zu versinnlichen, nächst dem Schädel unseres hier aufgestellten *Nyctic. leucog.* (unter No. 1.2.) auch den Schädel des *Nyctic. novemboracensis* (unter No. 3.4.) abbilden lassen, welche in aller Hinsicht von gleicher Beschaffenheit sind. Die Zähne sind *dent. incis. 2/6*. Die oberen 2 liegen dicht an den Eckzähnen, während sie

*) Von ... Nacht und ... ich tödte. Wir haben ganz willkürlich den Namen Nachtflatterer gewählt, anstatt einer gezwungenen Uebersetzung dieser Geschlechtsbezeichnung.

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stark von Gestalt, nach innen convergiren und durch einen großen leeren Raurn getrennt sind. *Dent. canin* $1^{-1}/1-1$, diese kehren ihre Spitzen nach außen. *Dent. Molares* $4^{-4}/4-4$; diese bestehen aus einem vorderen falschen Backenzahn, so wie drei darauf folgenden normalen, wobei zu bemerken ist, daß der letzte der oberen Kinnlade als ein flacher, schmaler Zahnkörper, beinahe ganz transversal gestellt ist. Der digit. Index hat zwei Phalangen, der digit. medius vier und die übrigen drei. Da übrigens Herr Temminck in seinen *monographies de mammalogie* (siehe pag. 316.) eine umständliche Bearbeitung dieses in die systematische Darstellung der Säugthiere aufgenommenen Geschlechtes angezeigt hat, so haben wir uns nur auf diese wenigen Angaben von demselben beschränken wollen, um den Bemühungen unseres um die Naturgeschichte der Cheiroptern so sehr verdienten Freundes nicht von zugreifen.

Die Gesichtsfläche unseres *Nyctic. leucog.* ist nackt und fleischfarbig; die Ohren nackt und hellbraun; der Ohrdeckel von hinten nach vorn gerichtet; die Nase ziemlich wulstig, mit seitlichen runden Nasenlöchern. Auf der Stirne, dem Scheitel und dem ganzen Oberkörper ist das Haar wollig, dicht, bräunlich von Farbe. Der ganze Unterkörper weiß, mit gelblichem Anflug. Die Flügelhäute röthlich braun, so wie die Intercrural-Membran, über welche das Schwanzende etwas hinausragt.

Vaterland. Kordofan.

Aufenthalt. Bewohnt die Höhlen den Adansonia, welche in Kordofan den Namen *Taboldi* führt und wurde am Brunnen Nedger gefunden. Heißt bei den Arabern *Abu Righ.*

Nycticejus murino-flavus Heuglin, 1861

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Similis *N. flavigastro*, sed minor; supra murino-olivaceus, subtus olivaceo-flavus; dentibus laniariis violascente tinctis.

Diese Art unterscheidet sich ausser durch die in der Diagnose bezeichneten Merkmale noch von *N. flavigaster* durch viel breitere Nasengegend und etwas kürzere, mit einer stumpfen Spitze versehene Ohren. Ohr- und Zahnbildung ist im übrigen dieselbe. Körperlänge 2" 6""; Schwanz gegen 2", Flugweite 11 1/2", Ohr kaum 6 1/2"" hoch. - Der olivengelbe Ton des Unterleibes ist an der Kehle und der Innenseite der Schenkel am intensivsten.

Nur ein Exemplar, ein altes M, bei M'kullu an der abyssinischen Küste eingesammelt, wo wir diese Art übrigens in Gesellschaft von *Dysopes pumilus* öfter zu sehen Gelegenheit hatten.

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Auffallend ist der olivenbraune und gelbe Ton verschiedener ost- und südafrikanischer Schwirrmäuse, während die asiatischen Arten meist rostbraun oder rostgelb überflogen sind. *Nycticejus leucogaster*, Rüpp. aus Kordofan und dem südlichen Sennaar hat häufig einen gelblichen Anflug, bei *N. Nigrita* von Senegambien fehlt er auch nicht und bei *N. Dingani*, *N. planirostris* und *N. viridis* ist er vollends deutlich ausgesprochen. Möchte diese Färbung nicht durch äussere Einflüsse entstanden und etwa dem umstande zuzuschreiben sein, dass die *Nycticejus*-Arten meist hier hohle Bäume bewohnen, deren Holz entweder direct mechanisch färbende Eigenschaften besitzen, oder in welchen durch Verwesungsprocesse Humus-, Humin- oder ähnliche Säuren sich gebildet haben, die bei längerer Berührung, verbunden mit Feuchtigkeit, auf den feinen, sehr wenig Marksubstanz enthaltenden Pelz ähnliche Wirkung zur Folge haben? - Diese letztgenannte Species könnte möglicher Weise mit den vorhergehenden zusammenfallen; ein dieser Tage erlegtes Exemplar von *N. flavigaster* (kleiner und dunkler gefärbt) dürfte vielleicht den Übergang dazu bilden.

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This species distinguishes itself except by the in the diagnosis mentioned characters also from *N. flavigaster* by the much wider nose area and the somewhat shorter, ane ears with a blunt tip. Shape of ears and teeth is for the rest identical. Length of body 2" 6""; tail almost 2", wingspan 11 1/2 ", ear hardly 6 1/2"" high. - The olive yellow tint of the belly is at the throat and inside of the thigh the most intens.

One single specimen, an adult M near M'kullu on the Abyssinian coast collected, where this species was frenquently observed together with *Dysopes pumilus*.

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Remarkable is the olivebrown and yellow tint in several east- and south-african bats, whereas the Asiatic species have primarily redbrown or russet-yellow glares. *Nycticejus leucogaster* Rüpp. from Kordofan and southern Sennaar has a yellowish spark, in *N. Nigrita* from Senegambia it isn't lacking either and in *N. Dinganii*, *N. planirostris* and *N. viridis* it is clearly prominent. Could it be possible that this colour is the result of external influences and attributed to circumstances, such as the *Nycticejus* species live mainly in hollow trees, whose wood has direct mechanical colouring properties, or in which decomposition processes result in humus, humin or similar acids, which as a result of extended contact, linked with humidity, might have such a result on the little marksubstance [???] containing pelage?

[I am not completely sure about the correct translation of this sentence, but my feeling is that Heuglin attributes the yellowish colour to external circumstances, e.g. colouring properties of the wood directly or of their decay processes] This last mentioned species could possibly be identical to the previous one [= *N. flavigaster*]; a recently deposited specimens from *N. flavigaster* (smaller and darker coloured) could possibly be an intermediary. - Translated by Victor Van Cakenberghe, 2008]

Nycticejus nidicola Kirk, 1865

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Nycticejus nidicola, sp. nov.

Fur brown; the base of the hairs blackish, beneath yellowish. Ears ovate, acute, with a well-developed rounded process at the front part of the outer or lower edge. Tongue linear lanceolate, acute, rather more than half the length of the ear. Face depressed, bristly. Wings elongate, thin, bald, rather hairy above and below, close to the body; forearm-bone nearly $1\frac{1}{2}$ inch long; the thumb compressed, rather elongate, slender, of a single joint. Tail as long as the body. The interfemoral membrane very large, broad, with nearly regular, almost parallel, transverse muscular bands, which are hairy on the upper and lower surface. The spur elongate, strong, nearly as long as the foreleg and foot; the spur and the end of the membrane fringed with short, rather rigid hairs. The legs rather elongate; the lower part of the thigh slender; the shank slender, not quite half the length of the arm-bone; the toes moderate, slender, compressed, covered with short adpressed hairs.

Expanse of wings 10 inches, of forearm-bone 1.5 inch, of foreleg 8 inches, of foot 3 inches, of spur $9\frac{1}{2}$ lines.

Shupanga, near the Zambesi.

Four specimens were obtained; they had taken possession of the nests of Weaver-birds (*Euplectes*). Having accidentally found a pair in one of these hanging nests; others were soon discovered in similar positions near by.

Nycticejus planirostris Peters, 1852

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Tafel XVII. Fig. 4

N. oliveofuscus, subtus flavidoalbus; auriculis capite tertia parte brevioribus, trago elongato, ensiformi; calcare crure longiore; rostro depressissimo; naribus in labii superioris margine positis.

Longitudo tota 0,120; autibrachii 0,052; caudae 0,055; volatus 0,340.

Habitatio: Africa orientalis, Tette, 17° Lat. Austr.

Die Schnauze ist breit und sehr platt, so dafs die Nasenlöcher ganz am vorderen Rande der Oberlippe ausmünden. Die Ohren sind um ein Drittel kürzer als der Kopf, eiförmig, am hintern Rande flach ausgeschnitten; ihr vorderer Rand bildet unten einen nach hinten vorspringenden Lappen, ihr hinterer Rand setzt sich über die Wange bis 4. Mm, hinter und unter dem Mundwinkel fort; der Tragus ist von sickelförmiger Gestalt, mit seiner Spitze nach vorn gekehrt, und halb so lang wie das Ohr. Die Schleimhaut des Gaumens bildet sieben Querfalten, von denen die beiden ersten grade und ungetheilt sind. Vor der Mitte der Unterlippe tritt die Schleimhaut in Form einer zweilappigen breiten Platte hervor. Der Körper ist dicht behaart, ohne nackte Stellen; die Haare sind an den Seiten der Brust am längsten, 15 Mm. lang. Die Flughäute sind sehr grofs, gehen bis zur Basis der Zehen herab und stehen über der Lendengegend nur 9 Mm. von einander entfernt; neben den Seiten des Körpers sind sie dicht behaart, sonst auf der Rückenseite nackt; an der Bauchseite dagegen zeigt sich eine leichte wollige Behaarung auf der Halsflughaut, und langs des ganzen Vorderarms bis zum vierten Finger hin. Der Vorderarm reicht, neben dem Körper angelegt, eben so weit wie das vordere Ende der Schnauze. Das Mittelhandglied des Daumens wird nach innen nur zur Hälfte, nach dem zweiten Finger hin ganz von der Flughaut umfaßt. Die beiden letzten Glieder des Schwanzes, welcher den Unterarm an Länge übertrifft, treten frei aus der Schenkelflughaut, deren Seiten durch lange Spornen gestützt sind, hervor. Nur das Anfangsviertel der Schenkelflughaut, besonders an der Bauchseite, ist leicht behaart, der übrige Theil nackt.

Die Farbe des alten Männchens ist an der Rückenseite dunkel olivenbraun, an der Bauchseite weifs mit gelblichem Anfluge, und vor dem Schultergelenk befindet sich ein grofser braunlicher Fleck. Die Flughäute und Ohren sind dunkler braun, die Nägel gelblich weifs.

Der Schädel zeigt nichts in seiner Form von anderen Arten dieser Gattung Abweichendes Gebifs $3\frac{2}{3}2\frac{1}{1}\frac{1}{4}\frac{1}{1}\frac{2}{3}\frac{3}{2} = 30$. Die Backzähne der *Nycticejus* unterscheiden sich in ihrer Gestalt sehr wesentlich von denen der eigentlichen Fledermäuse: die W Form der Schmelzfalte an den oberen Backzähnen ist wegen der Kleinheit des mittleren Winkels sehr undeutlich, so dafs auch statt fünf nur vier Höcker auf der Kaufläche hervorspringen, und die hintere Abtheilung

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der unteren Backzähne, welche bei den eigentlichen Fledermäusen die breiteste und gröfste ist, bildet hier nur ein schmales Anhängsel, welches kaum ein Drittel des ganzen Zahns ausmacht. Den ersten obern Backzahn mufs man seiner ganzen Bildung nach als Lückenzahn betrachten. Was das übrige Skelet anbelangt, so besteht die Wirbelsäule aus 39 Wirbeln; diese sind 7 Halswirbel, 12 Rückenwirbel, 5 Lendenwirbel, 5 Kreuzbeinwirbel und 10 Schwanzwirbel. Der Kiel des Brustbeins ist sehr niedrig und läuft oben vor dem Manubrium in einen dreieckigen Stachel aus. Das Wadenbein ist vollständig verknöchert.

Die vordere Hälfte der Zunge ist mit kleinen rückwärts gekehrten spitzigen Papillen bedeckt, welche in der Mitte gröfser und zwei- bis dreispitzig sind; auf der hintern Hälfte stehen linsenförmige rundliche Warzen und zwei grofse Papillae vallatae. Der Magen ist bohnenförmig und geht in einen einfachen 95 Mm. langen Darm über, welcher vor seinem Ende sehr erweiter erscheint. Die Leber ist rechts mit einem tiefen Einschnitte versehen, unter welchem hinten die kleine rundliche Gallenblase

liegt. Die Milz ist platt, gestreckt, 8 Mm. lang und 2 Mm. breit. Die Nieren sind einfach bohnenförmig, 4, Mm. lang; die linke liegt viel tiefer als die rechte. Die Ruthe ist weich und häutig. Der Kehlkopf hat eine Länge von 5 Mm.; der Schildknorpel ist bandförmig schmal, der Ringknorpel hinten mit einer starken Crista für die Anheftung der Giefsbeckenknorpel versehen. Die Luftröhre ist allenthalben gleich weit und enthält bis zum Spaltungsstelle 18 Knorpelringe. Die Lungen bestehen jederseits aus einem einzigen Lappen, die linke Lunge ist wegen des mehr nach links gerichteten Herzens etwas kleiner als die rechte.

Diese Art stammt aus dem Innern des Landes, aus Tette, ungefähr 120 Meilen von der Küste, im 17° Südl. Br.; das einzige Exemplar, ein ausgewachsenes Männchen, wurde im März 1845 im Fluge gefangen.

Mafse in Millimetern.

N. planirostris; N. viridis.

Ganze Länge von der Schnauze bis zur Schwanzspitze: 120; 95

Flugweite: 340; 290

Länge des Kopfes: $22\frac{1}{2}$; 20

Länge des Ohres: 14; 11

Breite des Ohres: 9; 7

Länge des Oberarms: 32; 28

Länge des Vorderarms: 52; 46

Länge des Daumens (Mittelh. $3\frac{1}{2}$, 3. 1.Gl. 4, 4. 2.Gl. 2, 2): $9\frac{1}{2}$; 9

Länge d. 2ten Fingers (Mittelh. 47, $42\frac{1}{2}$, 1.Gl. $5\frac{1}{2}$, $4\frac{1}{2}$): $52\frac{1}{2}$; 47

Länge d. 3ten Fingers (Mittelh. $48\frac{1}{2}$, $43\frac{1}{2}$, 1.Gl. 18, 15. 2.Gl. $10\frac{1}{2}$, $11\frac{1}{2}$, 3.Gl. $7\frac{1}{3}$, $8\frac{1}{2}$): $84\frac{1}{3}$; $78\frac{1}{2}$

Länge d. 4ten Fingers (Mittelh. $46\frac{1}{2}$, $42\frac{1}{2}$, 1.Gl. 14, $11\frac{1}{2}$, 2.Gl. 8, $7\frac{1}{2}$, 3.Gl. 2, 2): $70\frac{1}{2}$; $63\frac{1}{2}$

Länge d. 5ten Fingers (Mittelh. 42, 39. 1.Gl. $8\frac{1}{2}$, $7\frac{1}{2}$, 2.Gl. 5, $5\frac{1}{4}$ • 3.Gl. 2, 2): $70\frac{1}{2}$; $63\frac{1}{2}$

Länge des Oberschenkels: 19; 19

Länge des Unterschenkels: 21; 19

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Länge des ganzen Fusses: $10\frac{1}{2}$; 11

Länge des Schwanzes: 55; 40

Länge des Sporns: $20\frac{1}{2}$; 18

Länge des Schädels: $19\frac{1}{2}$; $17\frac{1}{2}$

Länge der Wirbelsäule vom Atlas bis zum ersten Schwanzwirbel: 53; 47.

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???

The snout is wide and very flat, so that the nostrils are opening at the frontal edge of the upper lip. The ears are about one third shorter as the head; egg-shaped, at the posterior edge evenly cut (?), their anterior edge forms at the bottom a lobe pointing to the back, their posterior edge runs over the cheek up to 4 mm behind and under the corner of the mouth: the tragus is sickle-shaped, with its top pointed forward, and half as long as the ear. The mucous membrane of the palate forms seven ridges, of which both first are straight and undivided. In front of the middle of the lower lip, the mucous membrane forms a two-lobed wide plate. The body is densely haired, without naked patches; the hairs are the longest at the sides of breast, 15 mm long. The wing membranes are very large, run to the base of the toes and are in the lumbar region only 9 mm separated from one another; close to the body, they are densely haired, but on the backside they are naked; on the belly side there is a light woolly fur on the neck membrane, and along the entire forearm up to the fourth finger. The forearm, pressed against the body, reaches the top of the snout. The middle hand bone of the thumb is on the inner side up to half and to the second finger completely enclosed in the wing membrane. Both latter vertebra of the tail, which is longer than the forearm, are free of the tail membrane, which sides are supported by long spurs. Only the first quarter of the tail membrane, especially on the belly side, is faintly haired, the remaining part is naked.

The colour of the old male is on the back dark olive brown, on the belly white with a yellowish shine, and in front of the shoulder joint there is a large brownish spot. The wing membrane and the ears are dark brown, the nails are yellowish white.

The shape of the skull is not different from any other species in this genus. Dentition 3 - $2\frac{2}{3}$ - 2 $1\frac{1}{2}$ 1 - $1\frac{1}{4}$ $1\frac{1}{2}$ 2 - $3\frac{2}{3}$ - 3 = 30. The molars of Nycticejus can be clearly distinguished by their shape from the proper bats: the W form of the enamel of the upper molars is due to the small size of the middle corner very unclear, so that instead of five there are only four bends on the chewing surface,

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and the posterior part of the lower molar, which is the widest and largest in the proper bats, forms here only a narrow appendage, which barely accounts for one third of the complete tooth. The first upper molar should be considered false molar (?) due to its build. Concerning the remaining skeleton, the spinal column consists of 39 vertebrae, which are 7 cervical vertebrae, 12 back vertebrae, 5 lumbar vertebrae, 5 sacral vertebrae, and 10 tail vertebrae. The keel on the breastbone is very low and ends above in front of the manubrium (?) in a triangular sting. The fibula is completely turned into bone.

The anterior half of the tongue is covered with small backwards turned pointed papillae, which are largest in the middle and have two or three tips; on the posterior half there are lens shaped roundish warts and two large Papillae Vallatae. The stomach is bean shaped and runs into a simple 95 mm long intestine, which before its end seems much widened. The liver has on the right side a deep incision, under which posterior end a small roundish gall-bladder is located. The spleen is flat, stretched out, about 8 mm long and 2 mm wide. The kidneys are simple bean shaped, 4 mm long, the left one situated much deeper as the right one. The rod [=penis?] is weak and skin-like (?). The larynx has a length of 5 mm, the thyroid is band shaped narrow, the ringcartilage (?) has at the back a strong crista for attaching the "Giefsbeckenknorpel" [no idea what this is, some sort of cartilage?]. The trachea is everywhere as wide and contains 18 cartilage rings to the split. The lungs have a single lobe on each side, the left lung is due to the left oriented heart slightly smaller than the right one.

This species came from the inside of the country, from Tette, about 120 miles from the coast, at 17° South; the only specimen, a fully grown male, was caught in flight in March 1845

Measurements in mm.

Total length from snout [=nose] to tailtip: 120; 95

Wingspan: 340; 290

Length of head: 22 1/2; 20

Length of ear: 14; 11

Width of ear: 9; 7

Length of forearm: 32; 28

Length of upper arm: 52; 46

Length of thumb (Meta1, 1ph1, 1ph2): 3 1/2, 3. 1.Gl. 4, 4. 2.Gl. 2, 2): 9 1/2; 9

Length of second finger (Meta2, 2ph1): 47, 42 1/2. 1.Gl. 5 1/2, 4 1/2): 52 1/2; 47

Length of third finger (Meta3, 3ph1, 3ph2, 3ph3): 48 1/2, 43 1/2. 1.Gl. 18, 15. 2.Gl. 10 1/2, 11 1/2. 3.Gl. 7 1/3, 8 1/2): 84 1/3; 78 1/2

Length of fourth finger (Meta4, 4ph1, 4ph2, 4ph3): 46 1/2, 42 1/2. 1.Gl. 14, 11 1/2. 2.Gl. 8, 7 1/2. 3.Gl. 2, 2): 70 1/2; 63 1/2

Length of fifth finger (Meta5, 5ph1, 5ph2, 5ph3): 42, 39. 1.Gl. 8 1/2, 7 1/2. 2.Gl. 5, 5 1/4. 3.Gl. 2, 2): 70 1/2; 63 1/2

Length of the upper leg: 19; 19

Length of lower leg [=Tibia]: 21; 19

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Length of entire foot: 10 1/2; 11

Length of tail: 55; 40

Length of spur [=calcaneum]: 20 1/2; 18

Length of skull: 19 1/2; 17 1/2

Length of the spinal column from Atlas to the first tail vertebra: 53; 47.

[Translated by Victor Van Cakenberghe, 2008]

Nycticejus Schlieffenii Peters, 1859

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supra rufescens, subtus ex albo rufescens; alis fuscis.

Long. tota 0.075; cap. 0.015; aur. 0.013; tragi 0,005; caudae 0.032; antibr. 0.031; exp. alar, 0.200.

Nycticejus serratus Heuglin, 1877

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Ziemlich groß; Kopf schmal und etwas zugespitzt. Nasenlöcher klein, durch einen schwachen Spalt getrennt, der oben und unten sich erweitert und in seiner Mitte eine kleine aufgedunsene Membran zeigt; Unterlippe einfach gespalten, jederseits dieser Spalte in ein spitziges Läppchen herabgezogen; unter dem Kinn eine nackte, dreiseitige Vertiefung; Ohr klein, nach hinten und aufwärts gerichtet, dreiseitig mit abgerundeten Ecken, innen, der Außenrand mit 5 bis 7 Querfalten, am Borderrand 6 bis 7 kerbenartige Einschnitte; die Lappe an der Wurzel des äußeren Ohrflügels bogig bis gegen den

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Mundwinkel hin verlaufend; Tragus kürz, platt, oben mit einer rundlichen, blattförmig ausgebreiteten Spitze, Außenrand doppelt, Innenrand einfach eingekerbt; auf der Kehlmittle ein kahler Fleck.

Im Oberkiefer 2 kleine, im Unterkiefer 2 + 2 ebenfalls kleine und dreizackige Schneidezähne; die Eckzähne mit deutlichem Löcker an der Basis, also zweitheilig. Der kurze Schwanz theilweise vom Patagium interfemorale eingehüllt, jedoch seine Spitze frei. Zeigefinger eingliedrig, die drei folgenden dreigliedrig.

Olivengrünlich ins Graue; Unterseite heller olivengrünlich-grau; die Behaarung an der Spitze gelblich; Schnauze und äußere Fläche der Flughäute rufbraun, die unteren Theile der Spannhaut um die Schenkelgegend sowie der etwas aufgedunsene Seitenrand des Patagium gelblich ins Fleischröthliche; die ganze Innenseite der Spannhaut bläulichgrau bis grau-grünlich, nach der Unterhälfte hin mehr ockergelblich genetzt; Ohr sehr wenig behaart; von einer Ohrbasis zur anderen führt ein etwa so obsoletes Halsband über die Kehle, dasselbe ist olivengrünlich, wie der Rücken. Körperlänge stark 3" 5". Schwanz 1" 2". Vorderarm 2" 6" bis 2" 7". Flugweite fast 16". Ohrhöhe 8". Tragus 2 1/2". Sporn 9".

Ein jüngeres, kleineres Exemplar ist obenher dunkler umbrabraun ins Graue, am Unterleib mausgrau; im Ohr 10 bis 11 Falten.

Häufig im sogenannten Scherq el Aqabah, am Araschkol und den benachbarten Granitgebirgen.

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Rather larger, head narrow and somewhat pointed. Nostrils small, separated by a faint groove, which above and below widens and has in its middle a small swollen membrane; lower lip with a single split, on each side of the split with a pointed small lobe; below the chin a naked three-sided depression; ear small, backwards and upwards directed, three-sided with rounded corners, inside the outer margin has 5 to 7 ridges, on the border edge 6 to 7 notchlike incisions; the lobe at the base of the outer earwing curved until near the corner of the mouth

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Tragus short, flat, open with a roundish, leaf shaped spread tip, outer margin double, inner margin once carved; on the middle of the throat a naked spot.

In the upper jaw 2 small, in the lower jaw 2 + 2 likewise smaller and trilobed incisors; the canines with an obvious hump at their base, also bilobed. The short tail partly enclosed in the interfemoral membrane, but its tip free. Index finger with one member, the three following with three.

Olivebrownish to greyish; lowerside brighter olivebrown-grey; the fur on the tips yellowish; snout and outer sides of the wing membrane russetbrown, the lower parts of the wing membrane in the area of the thighs as well as somewhat swollen edge of the patagium yellowish to fleshreddish; the entire inner side of the wing membrane blueish-grey to greyish-greenish, towards the lower half more ochre-yellowish netted; ear little furred, from one base of the ear to the other there is a somewhat obsolete collar over the throat, which is the same olivebrownish as the back. Length of body robust 3" 5", Tail 1" 2". Forearm 2" 6" to 2" 7". Wingspan almost 16". Height of ear 8". Tragus 2 ½". Spur: 9"

A young, smaller specimen is above dark umbra-brown to grey, on the belly mouse-grey; in the ear 10 to 11 ridges.

Common in the so-called Scherq el Aqabah, in Araschkol and in the neighbouring Granit mountains.

[Translated by Victor Van Cakenberghe, 2008]

Nycticejus viridis Peters, 1852

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Tafel XVII. Fig. 2.

N. olivaceoviridis, subtus viridiflavus; auriculis capite dimidio brevioribus, trago elongato; calcare crure brevior.

Longitudo tota 0,095; antibrachii 0,046; caudae 0,040; volatus 0,290.

Habitatio: Africa orientalis, Insula Mossambique, 15° Lat. Austr.

Mit breiter, dicker und wulstiger Schnauze, an deren vorderem Ende über dem Rande der Oberlippe die Nasenlöcher stehen. Die Entfernung der Nasenlöcher ist nicht ganz so groß wie die Hälfte ihres Abstandes von den Augen. Die Ohren sind halb so lang wie der Kopf, oval, am hintern Rande oben schwach ausgeschnitten, an ihrer äußeren Fläche bis zur Hälfte dicht behaart. Der hintere Ohrtrand geht nach vorn in einen Lappen über, der in gleicher Höhe mit dem Mundwinkel, 3 Mm. hinter demselben und durch eine warzenförmige Hervorragung von ihm getrennt; endigt. Der vordere Ohrtrand endigt mit einem freien Lappen, welcher spitzwinkelig nach hinten vorspringt. Der Tragus ist sichelförmig, am vorderen Rande fast grade und mit seiner Spitze nach vorn und innen gerichtet. Schnauze und Kinn sind nur schwach behaart und mit großen warzigen Drüsenhervorhebungen versehen. Die Schleimhaut der Unterlippe bildet vorn eine große, abgerundet viereckige, freiliegende Platte. Am Gaumen bemerkt man sieben Querfalten der Schleimhaut, von denen die erste grade, die zweite in der Mitte eingeknickt, die beiden folgenden tief V förmig zwischen der getheilten fünften und sechsten eingezogen sind, und die letzte und siebente mit ihrem stumpfen mittleren Winkel nach hinten vorspringt. Die Behaarung läuft am Körper keine Stellen unbedeckt, ist am Rücken, Halse und an den Seiten des Bauches gleich lang, etwas länger als in der Mitte des Bauches. Die Flughäute stehen in der Lendengegend viel weiter, 15 Mm., von einander ab, als bei der vorhergehenden Art, und gehen bis an die Basis der ersten Zehe herab. An der Rückenseite sind sie nackt bis auf den schmalen Saum, welchen die Ausdehnung der Körperbehaarung bedeckt; an der Bauchseite verdeckt langes zerstreutes Haar die Seiten bis zur Gegend zwischen Knie und Ellbogen, und eine noch viel spärlichere Behaarung macht sich an der Halsflughaut und längs des Unterarms bis zum vierten Finger bemerkbar. Das Ende des Vorderarms reicht bis zur Mitte zwischen Auge und Eckzahn. Das Mittelhandglied des Daumens ist nach innen nur zur Hälfte, nach dem zweiten Finger hin ganz von der Flughaut

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umsäumt. Der Schwanz, welcher kürzer als der Unterarm ist, steckt bis zur Spitze in der Schenkelflughaut, welche nur an ihrer Basis behaart erscheint.

Die Farbe der Rückseite ist ein schönes dunkles Olivengrün; die Haare der Bauchseite sind grünlichgelb. In Weingeist verändern sich diese Farben, so daß das Grüne weniger hervortritt. Die Flughäute und Ohren sind braunschwarz, die Nägel weiß.

Schädel und Gebiß stimmen außer einem geringen Größenunterschiede mit der vorigen Art überein, wie aus der Abbildung zu ersehen ist. Die Wirbelsäule besteht aus 7 Halswirbeln, 11 Rückenwirbeln, 4 Lendenwirbeln, 5 Kreuzbeinwirbeln und 9 Schwanzwirbeln, also aus drei, 1 Rücken-, 1 Lenden- und 1 Schwanzwirbel, weniger als bei der vorigen Art. Die Eingeweide stimmen in ihrem Bau mit der vorigen Art überein, nur sind die Ringe der Luftröhre etwas zahlreicher, indem man deren 24 zählt.

Auch diese Art scheint selten zu sein, da ich nur ein einziges Exemplar, ebenfalls ein altes Männchen, von derselben erhalten habe, welches im Februar 1844 des Abends in einer Wohnung auf der Insel Mossambique gefangen wurde. Der Magen enthielt Skelettheile von Insecten.

Sie unterscheidet sich von der vorigen Art, abgesehen von der Färbung, durch kürzere Ohren, kürzere Spornen und kürzeren Schwanz, durch eine dickere Schnauze, etwas robustere Füße, weiteren Abstand der Flughäute, viel schmalere Umsäumung der Spornen, und innerlich durch eine geringere Zahl der Wirbel und Rippen.

Scotophilus Dinganii (Smth, *Illustrations of the zoology of South Africa. Mammalia. Tab. 53*), welcher zur Gattung *Nycticejus* zu ziehen ist, scheint mir durch Folgendes von beiden Arten unterschieden zu sein: Die Ohren sind breiter als lang; die Nasenlöcher stehen näher beisammen; die Schnauze und Lippen sind dicker; die Flughäute gehen nur bis an die Fußwurzel herab; die unteren Schneidezähne sind zweilappig, während sie bei den beiden Arten aus Mossambique deutlich dreilappig sind; der Vorderarm ist viel länger, 2 Zoll 3 Linien oder 60 Mm., und die übrigen Maße stimmen mit keiner von beiden, indem die ganze Länge der von *Nycticejus viridis* entspricht, der Schwanz aber in seiner Länge mit dem von *Nycticejus planirostris* übereinstimmt. Wie sich die Spornen und der innere Bau verhalten, namentlich, was die Zahl der Wirbel anbelangt, darüber gibt weder die Abbildung noch die Beschreibung Auskunft. Vielleicht gehört auch eine von Sundevall mitgetheilte Art von eigentlichen Fledermäusen, *Vesperugo Dinganii* (*Öfversigt af Kongl. Vetenskaps - Akademiens Förhandlingar. 3. Jahrgang. 1846 pag. 119*) hieher, aber ich wage es nicht, anzunehmen, daß einem so ausgezeichneten Zoologen der Unterschied zwischen dem Bau der Zähne der eigentlichen Fledermäuse und der *Nycticejus* entgangen sei.

Die übrigen bekannten africanischen Arten wird man nicht leicht mit den von mir beschriebenen verwechseln können.

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Tafel XVII. Fig. 2.

N. olivaceoviridis, subtus viridiflavus; auriculis capite dimidio brevioribus, trago elongato; calcare crure brevior.

Longitudo tota 0,095; antibrachii 0,046; caudae 0,040; volatus 0,290.

Habitatio: Africa orientalis, Insula Mossambique, 15° Lat. Austr.

With wider, thicker, and more swollen snout, on which anterior end the nostrils are located over the edge of the upper lip. The distance between the nostrils is not entirely as large as half the distance between them and the eyes. The ears are half as long as the head, oval, on their posterior edge weakly cut (?), on their outer side up to their half thickly haired. The posterior ear margin runs to the front in a lobe, which at the same height of the corner of the mouth, 3 mm behind the same and separated from it by a wart-like projection. The anterior ear margin ends with a free lobe, which turns forward with [or possibly "in"] a sharp corner. The tragus is sickle shaped, its anterior margin is almost straight and with its top directed to the front and inwards. Snout and chin are only weakly haired and with a large wart-like glandular projection. The mucous membrane of the lower lip forms a large, rounded four-sided, free-lying plate. In the palate there are seven ridges in the mucous membrane, from which the first almost, the second in the middle bent inwards, both following ones deep V shaped drawn between the split fifth and sixth, and the last and seventh with its blunt middle angle sticking out to the back. The pelage leaves no spot uncovered on the head, and is on the back, neck and sides of the bell evenly long, slightly longer as on the middle of the belly. The wing membranes are in the lumbar region much wider, 15 mm, separated from one another as in the previous species [= *Nycticejus planirostris*], and run to the base of the first toe. On the back side, they are naked to the very narrow border, which covers the extension of the body, on the belly side long, scattered hairs cover the sides to the area between the knee and the elbow, and an even more scarcer pelage makes itself visible on the neck membrane and along the forearm to the fourth finger. The end of the forearm reaches until the middle between the eye and the canine. The metacarpus of the thumb is on the inner side only until its half and on the outer side entirely included in the wing membrane.

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The tail, which is shorter than the forearm, is enclosed until its tip into the tail membrane, which seems only haired on its base.

The colour of the back is a beautiful dark olivegreen; the hairs on the belly side are greenish-yellow. In spirit, these colours change, so that the green is less prominent. The wing membrane and the ears are brownish-black, the nails are white.

Skull and dentition agree with the previous species, except for a small size difference, as can be seen on the picture [I haven't seen any pictures of this]. The spinal column consists of 7 cervical vertebrae, 11 back vertebrae, 4 lumbar vertebrae, 5 sacral vertebrae, and 9 tail vertebrae, so 1 back, 1 lumbar and 1 tail vertebra less than the previous species. The intestines agree in their build with the previous species, only the rings on the trachea are slightly more numerous, since there are 24.

Also this species seems to be rare, since I have only obtained one specimen, also an old male, from the same, which in February 1844 was captured in the evening in a house on Mozambique islands. In its stomach, there were skeletal parts of insects.

This species distinguished itself from the previous species, besides its colour, by the shorter ears, shorter spurs and shorter tail, by a thicker snout, somewhat more robust feet, larger distance of the wing membrane, much narrower edging of the spurs, and on the inside by the lower number of vertebrae and ribs.

Scotophilus Dinganii (Smth, *Illustrations of the zoology of South Africa. Mammalia. Tab. 53*), which can be assigned to the genus *Nycticejus*, seems to be separated from these two species by the following: the ears are wider as long; the nostrils are closer together, the snout and lips are thicker, the wing membranes end at the base of the feet, the lower incisors are bicuspid, whereas they are in both species from Mozambique clearly tricuspid, the forearm is much longer 2 inches, 3 lines or 60 mm, and the other measurements do not agree with any of the other species, the total length agrees with the one of *Nycticejus viridis* [??? at least this is what I understood here ???], the tail however agrees with that of *Nycticejus planirostris*. How the spurs and internal built correspond, more specifically the number of vertebrae, can not be determined from the picture nor the description. Possibly a species of proper bats reported by Sundevall, *Vesperugo Dinganii* (*Öfversigt af Kongl. Vetenskaps -*

Akademiens Förhandlingar. 3. Jahrgang. 1846 pag. 119) might belong here, but I do not dare to accept that a so eminent zoologist might have missed the difference in the built of the teeth of the proper bats and Nycticejus.

The other known African species will hardly be confused with both described by me.

[Translated by Victor Van Cakenberghe, 2008]

Nyctinomia Fleming, 1822

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Nyctinomia. Four incisors in the lower jaw. Upper lip deeply notched.

Nyctinomus E. Geoffroy Saint-Hilaire, 1818

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NYCTINOME. *Nyctinomus*.

Dents incisives ²₄; canines ¹₁; molaires ⁴₅-⁴₅

Nez confondu avec les lèvres; celles-ci largement -et profondément fendues.

Oreilles grandes, réunies et couchées sur la face; oreillon extérieur.

Membrane interfémorale moyenne et saillante.

Queue longue, à demi enveloppée, et libre au-delà.

OBS. Les espèces de ce genre sont le nyctinome d'Égypte, celui du Bengale, et le nyctinome de Bourbon.

Nyctinomus (Dysopes) ventralis Heuglin, 1861

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Vellere brevi, sericato, brunneo-rufo, subtus pallidiore; stria ventrali, lata, mediana, conspicua rubente-flava; fibrissis faciei obscure brunneis; fascia lata subbrachiali villosa pallide fulvescente; pilis digitorum (ciliis) podarii setosis, valde elongatis et unguines ex parte superantibus.

Nasenlöcher rüsselartig aufgetrieben, nach vorne durch eine schmale aber sehr deutlich ausgesprochene Leiste geschieden. Oberlippenrand hängend und faltig; auch das nicht sehr kleine Auge in einer langen, fast unter dem Ohre

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versteckten Querfalte, in welch letzterer vor dem Auge hin sich auch eine scharfe thränengrubenähnliche Furche befindet. Unterlippe ganzrandig, durch eine Querfurehe vom Kinn geschieden, an dessen Seiten zwei kurze Falten der Länge nach herablaufen. Gesicht im allgemeinen rauh und unbehaart, nur über der Nase bis zwischen den Ohren und um die Oberlippe stehen büstenartige Haare, aus denen einzelne, theils über 2" lange Barthaare sprossen.

Die rundlichen breiten Ohren auf der Stirn durch eine niedrige, kurze Hautfalte verbunden und mit ihrem Aussenrande die Mundspalte fast erreichend; der genannte Rand an seiner Basis mit nach oben gerichtetem halbrundem Lappen; *Tragus* länglich-oval, niedrig, etwas tief in den Grund der Ohrmuschel herabgerückt, dort ebenfalls mit einem zum Verschlusse des Gehörgangs geeigneten Lappen. 8 - 12 Querfalten in der Ohrmuschel; die der faltigen Partie gegenüberstehende innere Längshälfte der Muschel innen gänzlich bis gegen den Rand, aussen mit einem schmälere Streifen von Haaren bedeckt.

Gaumen mit 7 in ihrer Mitte getheilten Querfalten. Schneidezähne ¹₂, die oberen zwei ziemlich eng stehend und eckzahnartig verlängert, die unteren vier sehr klein und oben schaufelförmig und mit je einer Kerbe versehen. Obere Eckzähne normal, die unteren an der Basis der Krone je gegen den zunächst gelegenen Schneidezahn hin mit zackenförmig erweitertem Zahnkranz, so dass zwischen Schneide- und Eckzahn ein dritter abnormer Zahn zu stehen scheint. Der Zeigfinger besteht nur aus einer Phalange. Flughaut den Vorderfuss am Handgelenke umschliessend, letzteres mit kräftigem *Calcaneus*, der an seiner Basis nach aussen einen der Spitze zu immer schmaler werdenden Hautrand und eine Andeutung von nagelartiger Verlängerung hat. Zehenseiten und Nägelglieder mit weissen langen, halbrund gebogenen büstenartigen, bis zu 2" langen Haaren besetzt. Das *Patagium interfemorale* sehr faltig und dehnbar, etwa ¹₃ des dicken, faltigen Schwanzes erreichend, von welcher Vereinigung aus sie die Spitze als Scheide einschliesst. Genannte Scheide sehr muskulös und sie kann an der Schwanzröhre weit auf- und abgeschoben werden.

Pelz kurz und sammtartig, ein 3" - 5" breiter Streif von Behaarung zieht sich auf der Innenseite der Flughaut längs des ganzen Oberarms hin.

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Länge des Körpers: 3" 6"

..... Schwanzes: 1" 11"
 Tolallänge: 5" 7"
 Flugweite: 15½"
Antibrachium: 2" 6"
 Ohr hoch: 9"
 Kopflänge: 1" 1".

Nyctinomus (Nyctinomus) tongaënsis Wettstein, 1916

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Aschgrau, mittelgroße Art, dem *N. aegyptiacus* Geoffr. am nächsten stehend, aber etwas größer. M mit Kehlsack, Ohren nicht durch ein Band verbunden, aber am Grunde zusammenhängend. Ohrmuschel sehr groß mit starkem Kiel, in ausgebreitetem Zustande ungefähr quadratisch, mit schwach konvexem Innenrande. breit abgerundetem Ende und zuerst geradem, dann stark konvexem Außenrand. Antitragus triangulär, oben stark abgerundet, durch einen tiefen Einschnitt vom Ohrande getrennt. Tragus klein, breit und stumpf abgerundet, ungefähr rechteckig, Innenrand sehr stark konvex, Außenrand fast gerade. Oberlippe stark wulstig mit fünf bis sieben deutlichen Furchen. Oberseite hell aschgrau, das einzelne Haar dunkel "drab-gray", am Grunde lichter, mit hellgrauer ("10. Gray") Spitze, Unterseite der Oberseite ähnlich, aber die Haarspitzen weißlich und der "drab"-farbige Ton lichter. Ein breiter Haarstreif auf der Unterseite der Flughaut entlang den Körperseiten graubräunlich-weiß, Kopf und Körper. 73 und 71, Schwarz 44 und 42.5. Unterarm 52 und 51, Ohrhöhe 23 und 22, Ohrbr. a. d. Basis 19 und 18, Traguslg. 6 mm.

Typen 2 MM, Tonga am oberen Weißen Nil, 16.IV.1914

Nyctinomus [(Mormopterus)] albiventer Dobson, 1877

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Inc. $\frac{2}{6}$; Pm. $\frac{1-1}{2-2}$. Ears separate, tragus triangular.

(Subgen. *Mormopterus*, Peters.)

20. *Nyctinomus albiventer*, n. sp.

Ears triangular, shorter than the head, inner margins arising from distinct points of origin from the forehead; inner margin of the ear-conch almost straight, slightly concave in upper third, tip rounded off, outer margin straight; antitragus scarcely defined, separated posteriorly by a very shallow notch; tragus nearly as broad opposite the base of its inner margin as high, irregularly triangular, rounded off above, very similar in shape to that of *N. norfolcensis*, but less triangular and broader above, an obtuse projection about the middle of its outer margin. Extremity of the muzzle projecting very much beyond the mandible, the end of the nose very prominent and distinct from the upper lip, nostrils opening almost laterally. Sides of the upper lip with vertical grooves not well defined. Gular sac small.

Wings from the lower third of the tibiae or from the ankles; fifth toe somewhat smaller than the first.

Dentition similar to that of *N. acetabulosus* (to be described), but the lower incisors are not crowded.

Fur, above, white at the base, the remaining part dark brown; beneath, the greater part of the chest and abdomen dirty white, the sides brown. Wing- and interfemoral membrane nearly naked; a narrow band of fur extends outwards behind the forearm and along the proximal third of the fifth metacarpal bone.

Length (of an adult ♀, the type): head and body 2"2; tail 1"25, tail free from membrane 0"7; head 0"85; ear 0"65, tragus 0"15; forearm 1"45; thumb 0"35; second finger - metacarp. 1"4, 1st ph. 0"55, 2nd ph. 0"35; third finger - metacarp. 1"4, 1st ph. 0"5, 2nd ph. 0"35; fourth finger - metacarp. 0"9, 1st ph. 0"35, 2nd ph. 0"15; tibia 0"4; foot and claws 0"3.

Hab. Madagascar.

Type in the British Museum.

Nyctinomus africanus Dobson, 1876

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Nyctinomus africanus n. sp.

Upper lip smooth, without vertical wrinkles. Ears from perfectly distinct points of origin though close together, their inner margins arising from the upper surface of the muzzle, though not so near the extremity of the muzzle as *N. Cestonii*; tragus somewhat similar to that of *N. Cestonii*, but longer, and the upper margin quite evenly rounded off; antitragus quadrilateral, the shortest side in front, the upper margin slightly concave.

Fur bright orange-chestnut above and beneath.

Incisors $2/4$; the lower incisors not crowded: premolars $2-2/2-2$; the first upper premolar very short and blunt, its base occupying the whole space between the canine and second premolar.

Length (of an adult male) 3.6 inches; tail 2.2; ear 1; tragus 0.3; forearm 2.5; thumb 0.4; second finger - metacarpal bone 2.4, first phalanx 1.1, second phalanx 1.3; fourth finger 2.4; tibia 0.75; foot and claws 0.45.

This species resembles *N. midas*, from North Africa, in size and in general appearance. The large tragus, however, at once distinguishes it from that species.

Hab. South Africa (Transvaal Republic). Type in the collection of the British Museum, presented by R. B. Sharpe, Esq.

Nyctinomus aloysii-sabaudiae Festa, 1907

p. 1

NYCTINOMUS ALOYSII-SABAUDIAE, nov. spec.

(diagnosi preventiva)

Orecchie grandi, arrotondate, riunite alla base del loro margine-interno da una spessa fascia cutanea, formante sulla sua parte anteriore una protuberanza carnosa, sulla parte anteriore della quale nascono radi peli setolosi. La carena interna è elevata, coll'orlo superiore ingrossato nella parte inferiore, e gradatamente assottigliantesi nella sua parte superiore.

L'antitrigo è piuttosto alto, largo alla base, col margine interno obliquo e quasi rettilineo, col margine superiore leggermente convesso.

Il trago è assai piccolo, rivolto allo innanzi, col margine interno un po' concavo, e l'esterno convesso.

Il muso è troncato obliquamente. Il labbro superiore è molto ampio, solcato da profonde rughe, delle quali quelle sulla parte anteriore del labbro stesso sono leggermente oblique allo indietro, e quelle sulla parte posteriore, leggermente oblique allo innanzi.

La membrana alare è inserita alla metà della tibia sul lato anteriore; l'uropatagio comincia alla parte posteriore della tibia un po' al disotto del punto, dove è inserita la membrana alare.

La coda è compresa nell'uropatagio per un quarto circa della sua lunghezza.

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Il calcaneo è breve.

Il pelame è di colore castagno chiaro (*Russet* del Ridgway (*)) colla base dei peli di color fulvo chiaro.

Denti: Incis. $1/2$, Can. $1/1$, Premol. $2/2$, Mol. $3/3$.

Il cranio è mediocrementemente allungato. Il 1° premolare superiore è molto piccolo, situato nella linea dentale, in contiguità del canino. Gli incisivi inferiori sono bilobati. I canini hanno le basi vicine, più alte degli incisivi.

Le dimensioni dell'esemplare tipo della specie, una femmina di Toro sono le seguenti:

Lunghezza del capo e del corpo: mm. 69

" del capo: " 27

" dell'avambraccio: " 51

" della tibia: " 18

" della coda: " 42

" della porzione libera della coda: " 31.5

" delle orecchie, dall'intaccatura dell'antitrigo all'apice: " 18

" del trago, dalla base del margine interno all'apice: " 2

p. 1

NYCTINOMUS ALOYSII-SABAUDIAE, nov. spec.

(preliminary diagnosis)

Ears large, rounded, join together at the base of their inner margin by a thick cutaneous band, forming on its front a fleshy protuberance, on the front of which grow sparse bristly hairs. The internal keel is elevated, with the upper edge enlarged in the lower part, and gradually tapered into its upper part.

The antitragus is rather tall, broad at the base, with an oblique and almost rectilinear internal margin, with a slightly convex upper margin.

The tragus is very small, facing forward, with a slightly concave inner margin, and a convex exterior.

The snout is obliquely truncated. The upper lip is very broad, furrowed by deep wrinkles, of which those on the front of the lip itself are slightly oblique to the back, and those on the back, slightly oblique to the front.

The wing membrane is inserted in the middle of the tibia on the front side; the uropatagium begins at the back of the tibia a little below the point, where the wing membrane is inserted.

The tail is included in the uropatagium for about a quarter of its length.

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The calcar is short.

The hair is light chestnut (*Russet* of the Ridgway (*)) with a light tawny hair base.

Teeth: Incis. ¹ / ₂, Can. ¹ / ₁, Premol. ² / ₂, Mol. ³ / ₃.

The skull is slightly elongated. The 1st upper premolar is very small, located in the dental line, in proximity of the canine. The lower incisors are bilobed. The canines have bases close to each other, higher than the incisors.

The dimensions of the typical specimen of the species, a female from Toro are the following:

Length of head and body: mm. 69

" of the head: " 27

" of the forearm: " 51

" of the tibia: " 18

" of the tail: " 42

" of the free portion of the tail: " 31.5

" of the ears, from the indentation of the antitragus to the apex: " 18

" of the tragus, from the base of the internal margin to the apex: " 2

Nyctinomus Anchietae Seabra, 1900

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a, b. MF, Quibula, Angola, 1891. *Anchieta*; (alc. coll. d'Afr.).

c, d. MM, Galanga, Angola, 1893; *Anchieta*; (alc. coll. d'Afr.).

A presente especie foi ainda vista e estudada pelo professor Barboza du Bocage. Infelizmente, n'essa occasião, já a falta de vista principiava a dificultar sobremaneira os seus trabalhos de observação, e os preciosos apontamentos contendo a diagnose original d'esta nova especie extraviaram-se parece que de vez.

Procuraremos pela nossa parte evidenciar quanto possivel os seus caracteres, conservando-lhe a denominação especifica com a qual o illustre Professor a havia já determinado em homenagem a esse inolvidavel naturalista, a quem o Museu deve por assim dizer todas as riquezas que possui da nossa África occidental.

Descrição: - Orelhas separadas, muito redondas e bem desenvolvidas; anti-tragus elevado, carnoso e ovoide, separado da orelha por um sulco bem distincto; tragus mediocrementemente desenvolvido, um pouco anguloso; labio superior pouco rugoso proximo da commissura, mas profundamente nas proximidades das narinas; estas salientes; sacco gular indistincto; membrana da aza partindo do terço inferior da tibia. O pello não muito abundante, curto, pardo escuro, ligeiramente mais claro e acinzentado no ventre, e amarellado no rebordo superior e anterior das orelhas. Membrana ante-brachial muitissimo curta, inerme pela parte inferior, coberta de curta e sedosa pubescencia pela parte superior; a inter-femoral, que é sustentada pelas fortes apophyses cartiliginosas do calcaneum, inerme pela parte superior, e apenas com alguns raros pellos proximo da cauda pela parte inferior; aquella que se estende dos membros anteriores e posteriores coberta em toda a volta do corpo pela parte inferior por uma larga faixa de pelles bastantes longos e um pouco mais claros que os do ventre. Estes pellos cobrem a região articular do autebraço bem como a do femur.

As membranas são bastante amplas e conservam approximadamente a côr geral do pello.

Os pés notavelmente guarnecidos de longos pello acinzentados.

Dentição: in. ¹/₂ c. ¹/₁ pm. ²/₂ m. ³/₃. Maxilla superior: incisivos mediocrementemente desonvolvidos, um pouco recurvados; caninos longos e quasi direitos; primeiro premolar pequenissimo, um pouco interior

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á linha normal e ao meio do espaço deixado entre o canino e segundo premolar, que ó beni desenvolvido; molares normaes. Maxilla inferior: incisivos bióbadados; caninos bem desenvolvidos; primeiro premolar curto; segundo e molares normaes.

Paladar concordando com o typo do genero já exposto.

Dimensões: cabeça e corpo 70 mili.; cauda 40; parte livre 21; cabeça 24; orelha 19 (da base do anti-tragus á orla superior); tragus 5 X 2.3; ante-braço 51; pollegar 6; terceiro dedo: metacarpo 50; primeira phalange 20; segunda 24; quarto dedo: metacarpo 46; primeira phalange 17; segunda 10; quinto dedo: metacarpo 27; primeira phalange 14; segunda 7; tibia 17; pe 7.5.

Os dois exemplares de Galanga são de côr mais escura do que aquelles que vimos de descrever.

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The species presented was seen and studied by Professor Barboza du Bocage. Unfortunately, on this occasion, his lack of sight was beginning to greatly affect his observation work, and the precious notes containing the original diagnosis of this new species seems to have gone astray.

We searched on our part for as much evidence as possible for his/their characters, saving the names which the Professor had given in honour of this unforgettable naturalist, to whom the Museum must say all the riches that our Occidental Africa possess.

Description: Separated ears, rounded and well developed; high anti-tragus, fleshy and ovoid, separated from the ear by a distinct groove; underdeveloped tragus, slightly angular; upper lip slightly wrinkled near the commissura, but deep near the nostrils; protruding; distinguished gular pouch; wing membrane starting from the bottom third of the tibia. There is not an abundance of hair, it is short, dark brown in colour, lighter grey colour towards the belly, and yellowish by the upper edge and above the ears. Anti-brachial membrane is very short, largely at the bottom, covered in short and silky pubescence hair on the top; the inter-femur, that is supported by the strong cartilaginous apophyses of the calcaneum, largely on the top part, and with a few rare hairs near the bottom part of the tail; those extending from the anterior and posterior of the membranes covers around the whole back of the body from the bottom with a range of long hairs and a little lighter than those of the belly. These hairs cover the joint region of the forearm as well as the femur.

The wing membranes are very large and are approximately the same colour as the hair.

The feet are notably trimmed with long grey hair.

Dentition: in. $\frac{1}{2}$ c. $\frac{1}{1}$ pm. $\frac{2}{2}$ m. $\frac{3}{3}$. Maxilla superior: poorly developed incisors, slightly curved; long and relatively straight canines; small first premolar, slightly inward.

The normal line goes until the middle of the space left between the canine and the second premolar, which is well developed; normal molars. Inferior maxilla: incisors are bi-lobed; canines well developed; first premolar is short; second premolar and molars are normal.

The palate corresponds with the general type already stated.

Measurements: head and body 70 mili.; tail 40; free part 21; head 24; ear 19 (from the base of the anti-tragus to the superior oral); tragus 5 x 2.3; forearm 51; thumb 6; third finger: metacarpal 50; first phalange 20, second 24; fourth finger: metacarpal 46; first phalange 17; second 10; fifth finger: metacarpal 27; first phalange 14; second 7; tibia 17; foot 7.5.

The two examples from Galanga are a darker colour than those we've been describing.

Nyctinomus angolensis Peters, 1870

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Nyctinomus angolensis. Nov. sp.

N. supra ferrugineus, subtus pallidior,. ventre médio ferrugineo-albus; patagio lumbari subtus pilorum stria alborum, a corporis vellere spatio calvo sejuncta, vestito; auriculis connatis, trago angulato minuto, antitrigo quadrangulati-rotundato; pollice disco nudo magno; digitis posticis setosis; incisivis superioribus approximatis; proemolari primo superiore externo mimitissimo.

Long. tota 0^m,1200
 " capitis 0^m,0280
 Alt. auris 0^m,0150
 Lat. " 0^m,0135
 Long. tragi , 0^m,0015
 " caudae 0^m,0450
 " humeri 0^m,0305
 " antibrachii 0^m,0480
 " digitl. (mtc. 0,0040; ph. 1.0,0050; ph. 2. 0,0045) 0^m,0135
 " " 2. (" 0,0445; " 0,0020) 0^m,0465
 " " 3. (" 0,0470; " 0,0220; ph. 2. 0,0217; cart. 0,0110) ... 0^m,1017
 " " 4. (" 0,0450; " 0,0175; " 0,0160; " 0,0023) ... 0^m,0808
 " " 5. (" 0,0310; " 0,0140; " 0,0055; " 0,0020) ... 0^m,0525
 " femoris 0^m,0170
 " tibiae 0^m,0150
 " plantae c. ung 0^m,0125

Mas adultus ex Angola. (Toulson)

Nyctinomus ansorgei Thomas, 1913

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Allied to *N. cisturus*, Thos., with which alone it shares a somewhat *Chærephon*-like shape of skull combined with typical *Nyctinomus m*³ and premaxillæ.

Size about as in *N. cisturus*; distribution of fur as in that species, except that the tail and interfemoral are less hairy; a partly naked patch on the crown behind the junction of the ears, and another across the nape just in front of the shoulders. General colour above dark chocolate-brown, the tips of the hairs paler. Below, succeeding the naked chin, the throat is blackish brown, considerably darker than any other part of the animal, and suggesting the blackish beard of certain species of *Taphozous*; chest and belly lighter brown, the tips markedly lighter than the bases, but not anywhere white. Ears of medium size, rounded; basal keel not specially thickened; antitragus triangular, rounded above, about half as high as long, separated behind by a deep notch. Tragus small, subquadrangular, very different from the triangular pointed tragus of *N. cisturus*. Tail apparently without special glands.

Skull on the whole very similar to that of *N. cisturus*, but rather larger throughout; upper profile not so convex in the frontal region. Teeth as in *cisturus*; anterior upper premolar in the tooth-row, not crowded, but just filling up the space between the canine and *p*⁴. Lower incisors 4. Anterior lower premolar broader than the posterior.

Dimensions of the type (the italicised measurements taken in the flesh): -

Forearm 47 mm.

Head and body 70; tail 32; ear 19; tragus (dry) 1.5; third finger, metacarpus 46, first phalanx 18.6, second phalanx 19; fifth finger, metacarpus 28.

Skull: greatest length 19.2; zygomatic breadth 11.4; interorbital breadth 4.2; mastoid breadth 10.7; front of canine to back of *m*³ 7.4.

Hab. Malange, North Angola. Alt. 1150 m.

Type. Adult male. B.M. no. 10.4.8.4. Original number 5. Collected 17th February, 1908, by Dr. W. J. Ansorge.

The Angolan species described by Sr. de Seabra all have the low skull of typical *Nyctinomus*, *N. cisturus* being the only known species combining a high skull with the complete *m*³ and divided premaxillæ which distinguish *Nyctinomus* from *Chærephon* and *Mops*. From *N. cisturus* the

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new species is readily recognizable by its dark beard and non-glandular tail.

Nyctinomus Bemmeleni Jentink, 1879

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Nyctinomus Bemmeleni, n. sp.

Dentition, Inc. ²/₄, c. ¹⁻¹/₁₋₁, p. m. ²⁻²/₂₋₂, m. ³⁻³/₃₋₃.

At a glance this species is distinguished from all the other species of the genus *Nyctinomus* by the length of the metacarpal bone of the third and the fourth finger, by the peculiar manner in which the interfemoral membrane and the wings are attached to the tibiae, and by the very long end of the tail free from the membrane.

Muzzle as in the other species; upper lip very expansible and thick, deeply grooved by vertical wrinkles.

Ears united by a low band in front, but connected by the bases of their inner margin, which form a sharp angle. Earconch triangular, rounded towards the tip, the inner and outer margins being oval. Tragus also triangular, very small. Antitragus well developed, broad, obtusely rounded towards the top, with a large base, separated from the earconch by a very deep notch.

The female possesses no gular sac.

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In the other species of the genus *Nyctinomus* the forearm always exceeds in length the metacarpal bone of the third and the fourth finger: in our species, however, the case is just the contrary, for the metacarpal bone of the third finger exceeds the forearm and that of the fourth finger equals the forearm in length.

Comparatively the tail is not longer than in the other species, but its part included in the interfemoral membrane is particularly short and therefore the free end of the tail is extraordinarily long.

In the other species of this genus according to rule the wings are attached to the outside of the tibiae or ankles, and the interfemorale membrane to the inside. Now in the species in question the wings and the interfemoral membrane proceed from the same point, to be found on the middle of the tibia just between the out and inside.

The thumbs, the first and the fifth toe are very thick; especially the latter, and further the other toes are ornated with long rigid, white colored, overhanging hairs - as commonly in the other species of this genus.

Fur dark smoke-brown above, yellowish brown beneath. Ears, wings and tail colored as the back.

Upper incisors well developed, close together, separated from the canines by a rather large interval. Lower ones very little, bifid, crowded. First upper premolar very small, acute, not filling up the space between the canine and second premolar: first lower premolar about half the size of the second premolar.

Type in the Leyden Museum.

Measures of the single specimen, being a female, preserved in alcohol.

Head and body: 54

Tail: 35

Tail free from membrane: 26

Ear: 15

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Tragus: 2 X 2

Antitragus= 3.5 X 5

Forearm: 41

Third finger-metacarp: 42

" " 1st phal.: 18

" " 2nd " : 16.5

" " 3rd " : 8

Fourth finger-metacarp: 41

" " 1st phal.: 14

" " 2nd " : 11.5

Fifth finger-metacarp: 24

" " 1st phal: 12

" " 2nd " : 5

Tibia: 14

Foot with claws: 11

Hab. Liberia.

Mr. A. A. van Bemmelen, Director of the Zoological Gardens at Rotterdam has been so kind as to present this curious Bat to the Leyden Museum: I therefore propose to name it in honor of its donor.

Nyctinomus bivittatus Heuglin, 1861

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Medius, auriculis apice vix rectangulare, fronte connatis; trago acuminato brevissimo; taenia frontale aures conjungente distinctissima, medio plica antice producta (projectione W-forme) - calcaneo indistincto, patagio interfemorale et lumbare circumdato; pollice intus disco orbiculare instructo; parte basale ($\frac{1}{4}$) caudae patagio vestita, antice pilis rigidiusculis. - Dent incis. Utrunque $\frac{1}{2}$, supra obtusis, subtus bi-cuspidibus minimis; caninis inferioribus apice vix cestiforme simpliciter incisa; molaribus utrinque duabus incurrentibus mandibulae. - Corpore supra obscure umbrino; pectore paulo pallidior; abdomine fulvescente, regione femorale ferrugineo-ventre medio griseo indutis; vittis duabus occipitalibus albis; humeris, pectoris lateribus et regione subauriculare delicate albido stictis; labio maxillare, auriculis et patagio nigricantibus, mandibula pallidior, vix carnicolore, nuda.

Dieser weitläufigen Diagnose habe ich beizufügen, dass die mittelgrossen Ohren 6 - 7 deutliche Querfalten zeigen; der Gaumen deren 5, und eine sechste minder entschiedene. Der sehr kurze, weit hinabgerückte *Tragus* mit seitwärts und nach aussen gebogener Spitze. Nasenlöcher wenig röhrenförmig aufgesetzt, seitlich geöffnet, durch eine kleine behaarte Furche getrennt. Die Lippen sehr faltig, die obere am Rande gekerbt; der Unterkiefer fast kahl und fleischfarb, weit zurücktretend, im Kinn mit rundlicher Vertiefung. Obere Eckzähne mit erhabenem Kronrande und einer die Spitze nicht erreichenden Furche auf der Vorderseite, die unteren mit starkem Höcker gegen die Schneidezähne hin und sie, an Grosse weit überragend. Im Unterkiefer kann ich 2 falsche Backen-

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zähne unterscheiden. Am Fusse reicht die Flughaut bis unter die Mitte der *Tibia*, der Schwanz fast zu $\frac{3}{4}$ frei, innen, so weit er im *Patagium interfemorale* steckt, wenig aber rauh behaart: letzteres ist heller als die übrige Flughaut, ins Bläulich-fleischfarbe

spielend, innen dicht mit weisslichen Hautwärtchen besetzt; die Zehen mit borstigen Haaren auf dem Nagelgliede und Hände der innersten und äussersten. Die Flughaut in den Winkeln, die der Oberarm an seiner Basis mit ihr bildet, behaart. Auf der Vorder- und Hinterseite des Ohres etwa parallel mit seinem Innenrande ein behaarter Streif, der namentlich auf der Rückseite sehr deutlich ist und nach der Basis zu breiter wird. Am Daumenballen eine runde, etwas erhabene hornige Platte oder Scheibe. Occiput, Rücken, Kehle und Halsseiten umbräun, die beiden letzteren etwas heller; Unterleib bräunlich-grau, nach den Seiten etwas rost-röthlich angefliegen. Hinter der Ohrbasis längs des Ober- und Hinterkopfes jederseits ein schmaler, weisser Streif; Schultern, Brustseiten und Kehlmittle, sowie die untere Basis des Ohres fein weissgescheckt. Ganze Länge des alten M 4", davon kommen auf den Schwanz 1" 5"; Ohr 8" hoch, Vorderarm 1" 10", Flugweite 12½". Das F ziemlich schwächer. Ein jüngeres M ist auf der Oberseite mehr dunkel-röthlich-braun, die weissen Streifen an den Occiputseiten weniger deutlich entwickelt. Die unteren Eckzähne spitziger und an der Spitze selbst noch nicht gekerbt.

Diese Art trafen wir unfern Kérén mit einbrechender Dämmerung zuweilen zu 3 - 4 Individuen längs Regenbeeten und Hochbäumen hinziehend.

Nyctinomus Bocagei Seabra, 1900

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5 M, 1 F ad., Galanga, Angola. Anchieta; (alc. coll. d'Afr.).

Um grande numero de particularidades que procuraremos evidenciar distinguem esta especie de todas aquellas que temos observado e de que conhecemos a descripção. As orelhas obedecerem ainda aos caracteres do grupo das especies até aqui citadas e descritas: separadas nas suas bases, opacas, com um engrossamento na sua base proximo da orla esterna, occupando o terço inferior d'esta e separada do anti-íragus por um profundo sulco. São bem desenvolvidas, de forma um pouco irregular, angulosas e guarnecidas anteriormente por uma serie rnuíio pronunciada d'aquellas lacineas tegumentares cuja ausencia citámos para, auxiliar a distincção do *Nyctinomus brunneus*. Anti-tragus pequeno, semi-ovoide como nas outras especies; tragus quadrado e mais estreito. As rugas anteriores dos labios profundissimas, particularmente as duas medianas; as narinas salientes; sacco gular indistincto. Pello abundante, mesmo sobre as orelhas, que cobre em grande parte; pardo escuro sobre o dorso, pouco amarellado, e, segundo a re-

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gra, mais claro na região ventral. A sua distribuição sobre as membranas é semelhante áquella que temos observado nas outras especies, sómente é mais escasso na ante-brachial superior, nullo pela parte inferior d'esta, contrariamente á inter-femoral, onde é bastante abundante pela parte inferior e quasi nullo pela parte superior, excepto junto á base da cauda. As apophyses cartilagosas do calcaneum approximando-se notavelmente da cauda, que é delgada e curta. Os pellos que guarnecem os dedos nos pés estendendo-se sobre o rebordo da inter-femoral até mais de metade da apophyse cartilaginosa do calcaneum, diminuindo gradualmente de comprimento.

Dentição: in. ½ c. ½ pm. ½ m. ⅓. Na maxilla superior os incisivos bastante convergentes; caninos bem desenvolvidos e pouco curvos; primeiro premolar muito pequeno e ao meio do espaço deixado entre o canino e o segundo premolar; este bem desenvolvido e excedendo notavelmente os molares, que são normaes. Na maxilla inferior os incisivos são bilobados e os restantes dentes não offerecem tambem nada de particular. O paladar concordando ainda com o typo generico.

Dimensões: cabeça e corpo 64 mill.; cauda 30; parte livre 13; cabeça 20; orelha (da base do anti-tragus á orla superior) 15; tragus 4 X 1.5; ante-braço 45; pollegar 4; terceiro dedo: metacarpo, 43; primeira phalange 18; segunda 22; quarto dedo: metacarpo 41; primeira phalange 14; segunda 10; quinto dedo: metacarpo 26; primeira phalange 12; segunda 6; tibia 13; pé 7.

Procurando ligar a todas as valiosas collecções do Museu de Lisboa o nome do seu Fundador, determinamos como fica esta nova especie.

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A large number of features were sought for evidence to distinguish this species from all those we have seen and whose descriptions we know. The ears still obey the characters of the group of this species which have been cited and described: separated at the base, opaque, with a thickening at the proximal base of the edge of the esterna, occupying a third of the lower esterna and separated from the anti-tragus by a distinct groove. They are well developed, somewhat irregular, angular and finished anteriorly by a series of many pronounced tegumental lacineas whose absence we have mentioned, assists in the distinction of *Nyctinomus brunneus*. Small anti-tragus, semi-ovoid like those in other species; square tragus and more narrow. The anterior wrinkles of the lips are deep, particularly the two medians; protruding nostrils; distinct gular pouch. A lot of hair, even on the ears, which covers the most part; dark brown on the back, yellowish, and, second to the rule,

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lighter on the ventral region. The distribution on the membranes is similar to that which we have observed on other species, only that it is scarcer on the top of the anti-brachial, no hair on the lower part, contrary with the inferior-femur, where there is an abundance of hair on the lower part and almost no hair on the top part, except near the base of the tail. The cartilaginous apophyses approaches noticeably from the tail, which is thin and short. The hairs which cover the toes of the feet extend over the inter-femoral rim to more than half of the calcaneum cartilaginous apophyse, gradually shortening in length.

Dentition: in. ½ c. ½ pm. ½ m. ⅓. On the superior maxilla the incisors are very convergent; the canines are well developed and slightly curved; first premolar is very small and until the middle of the space left between the canine and the second

premolar; well developed and significantly exceeding the molars, which are normal. On the inferior maxilla the incisors are bi-lobed and the remaining teeth do not offer anything in particular. The palate corresponds with the genus type.

Measurements: head and body 64 mill.; tail 30; free part 13; head 20; ear (from the base of the anti-tragus to the superior edge) 15; tragus 4 x 1.5; forearm 45; thumb 4; third finger: metacarpal 43; first phalange 18; second 22; fourth finger: metacarpal 41; first phalange 14; second 10; fifth finger: metacarpal 26; first phalange 12; second 6; tibia 13; foot 7.

Trying to connect all the valuable collections of the Museum of Lisbon to the name of its Founder, we determined how this new species fits in.

Nyctinomus brunneus Seabra, 1900

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a,b,c. 2 MM e 1 juv. Quissange, Angola, Anchieta; (alc. coll. d'Afr.).

A primeira vista distingue-se esta especie da precedente pela cor de um pardo muito escuro das suas membranas. Mais particularmente é assim caracterizada: orelhas separadas, redondas, *lisas no rebordo anterior e posterior*, isto é, desprovidas d'aquella serie de pequenas lacineas tegumentares que caracterisam grande numero das especies d'este genero; anti-tragus semi-ovoide mediocrementemente desenvolvido, separado da orelha por um distincto sulco; tragus quadrado e bem desenvolvido; o labio superior finamente rugoso proximo da commissura, seguindo-se-lhe tres pregas distinctas e irregulares nas proximidades das narinas; sacco gular do macho indistincte. Membrana

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das azas partindo quasi do meio da tibia. Pello abundante, bastante longo, avelludado sobre o dorso, e mais particularmente o da cabeça, que se estende muito sobre as orelhas, onde toma uma brilhante cor de sepia; o do dorso e mesmo o da região occipital é mais claro em consequencia da cor amarelada das extremidades dos pellos; aquelles que cobrem ainda superiormente as azas junto ao dorso, bem como as membranas brachial e inter-femoral, são como fias orelhas de cor de sepia. Pela parte ventral é mais claro, mais acinzentado e mais grosseiro; notavelmente longo no pescoço, disribuindo-se ainda fias membranas tambem, bastante longo e abundante junto do ventre; na interfemoral menos abundante que nas especies já descriptas.

A dentição, muito semelhante á da especie precedente, distingue-se comtudo na maxilla superior pelos incisivos, menos desenvolvidos, contrariamente aos caninos, muito mais longos que n'esta outra especie, e pela posição do primeiro premolar, que se encontra encostado ao segundo, ao passo que no *Nyctinomus Anchietae* está ao meio do espaço deixado entre este dente e o canino. Na maxilla inferior os incisivos, em consequencia do pequeno espago que existe entre os caninos, acham-se irregularmente dispostos, mas symetricos; estes são bilobados como nas outras especies e em numero de quatro. O paladar concordando com o tipo do genero já descrito.

Dimensões: cabeça e corpo 78 mill.; cauda 43; parte livre 26; cabeça 25; orelha (da base do anti-tragus á orla superior) 17; tragus 4.5 x 3.5; ante-braço 52; pollegar 5; terceiro dedo: metacarpo 56, primeira phalange 20, segunda 26; quarto dedo: metacarpo 52, primeira phalange 17, segunda 9; quinto dedo: metacarpo 30, primeira phalange 14, segunda 7; tibia 16; pé 7. Os pellos que guarnecem os pés são pouco abundantes e curtos.

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At first sight this species can be distinguished by the dark brown colour on its wing membranes. More particularly are the following characteristics: separated ears, round, *smooth at the anterior and posterior edge*, this is lacking from the series of small mucocutaneous lacineas which characterise a number of species of this genera; semi-ovoid anti-tragus which is underdeveloped, separated from the ear by a distinct groove; square tragus which is well developed; the lip is finely wrinkled near the commissura, followed by three distinct and irregular folds near the nostrils; the gular pouch of the male is distinct.

Membranes...

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of the wings start almost from the middle of the tibia. Abundant hair, very long, velvety on the back, more particularly that of the head, which extends over the ears, where the hair becomes a brilliant sepia colour; the hair on the back and occipital region is more clear in contrast with the yellow colour of the ends of the hair; those covering the wings and the back, as well as the brachial membranes and inter-femoral, are a sepia colour as in the ears. On the ventral part it is lighter, more greyish and coarse; noticeably longer on the neck, distributed on the membranes as well, very long and abundant together with the belly; less abundant on the inter-femoral than on species already described.

The dentition, very similar to that of the preceding species, distinguishes itself by incisors on the upper maxilla, less developed, contrary to the canines, much longer than the other species, and by the position of the first premolar, which is against the second, whereas in *Nyctinomus Anchietae* it is in the middle of the space left between this tooth and the canine. On the lower maxilla the incisors, in consequence with the small space between the canines, they are irregularly organised, more symmetrical; these are bi-lobed like in other species and in numbers of four. The palate corresponds with the type of genus described.

Measurements: head and body 78 mill.; tail 43; free part 26; head 25; ears (from the base of the antitragus to the superior edge) 17; tragus 4.5 x 3.5; forearm 52; thumb 5; third finger: metacarpal 56, first phalange 20, second 26; fourth finger:

metacarpal 52, first phalange 17, second 9; fifth finger: metacarpal 30, first phalange 14, second 7; tibia 16; foot 7. There is not a lot of hair covering the foot, and these hairs are short.

Nyctinomus cisturus Thomas, 1903

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A medium-sized species, with imperfect premaxillæ, united ears, and a peculiar swollen gland at the base of the tail.

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Ears thin, united at their bases, but not forming a prominent folded lobe at their junction; their anterior edge evenly but slightly convex, not folded back, without minute warts; tip broadly rounded off; outer edge strongly convex; antitragus high, broadly triangular; keel low, not flattened or thickened externally; tragus minute, triangular. Lips with vertical wrinkles. Apparently no gular sac. Pad at base of thumb not specially large. Wings to the distal third of the tibiae, on their inner aspect. Tail involved in membrane for about three fifths of its length; at its base, on the upper surface, occupying the middle of the interfemoral area, there is a large glandular swelling, 8 millim. Long by 6 broad, covered with fine fur; this swelling is caused by the presence of a pair of glands, presumably of a scent-secreting nature, which open on the under surface of the interfemoral by two large mouths, each over 3 millim. In length, placed diagonally on the two sides of the tail some 5 millim. Behind the anus; the area round these mouths well covered with fine silky white hair; internally some sort of dark-coloured glandular secretion is present.

Colour above chocolate-brown, rather paler below, the tips of the belly-hairs greyish. Wings and interfemoral membranes above brown, below brown outside of the elbow, white near the body and on the interfemoral.

Skull rather small in proportion, smoothly rounded, a low median crest present running from the level of the intertemporal constriction to the occiput. Premaxillæ imperfect mesially, but the opening between them unusually small.

Incisors $2/4$, the upper slender, not far apart, nearly parallel. Anterior upper premolar small but not crowded, standing in the line of the tooth-row, its conical point higher than the cingula of the neighbouring teeth.

Dimensions of the type (measured in the flesh): -

Forearm 47 millim.

Head and body 63; tail 35; ear 15 ; tragus 1.3; height of antitragus 2.2; third finger, metacarpal 46, first phalanx 19, second phalanx 18; fifth finger, metacarpal 27, first phalanx 14, second phalanx 5; lower leg and foot (s.u.) 24.

Skull: greatest length 18.1; basal length in middle line 13.8; zygomatic breadth 11.2; anteorbital breadth 6.5; constriction 3.7; mastoid breadth 10.3; front of canine to back of m^3 6.8; front of lower canine to back of m_3 7.1.

Hab. Mangala, on the east bank of the Nile, 25 miles N. of Gondokoro.

Type. Adult male. B.M. no. 2.7.4.4. Original number 2805. Collected and presented by W. L. S. Loat, Esq.

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This bat differs from all known members of the genus by its possession of the unique caudal gland above described. In other respects it is a true *Nyctinomus*, belonging to the typical group with imperfect premaxillæ.

Nyctinomus Condylurus A. Smith, 1833

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Nyctinomus Condylurus, (Knob-tailed Nyctinome). Fur very short and silky; above the surface colour is brown beneath dull tawny white, verging upon dull brown on the sides; ears black and rounded at tips, each with a narrow stripe of fur on its outer surface, extending from the base almost to the circumference; lips with four or five deep furrows; wing membranes blackish brown; interfemoral membrane enveloping more than half of the tail, above it is thinly covered with short fur towards its base and below till near its posterior margin tail enlarged and wrinkled at the point. Length from nose to base of tail about three inches, the latter an inch and a half. - When the fur is reversed the hairs of the back and sides are found to be tawny towards their bases; those of the middle of the body beneath dull brown.

Inhabits South Africa, - about Port Natal.

Nyctinomus demonstrator Thomas, 1903

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A medium-sized thick-set species with united premaxillæ, conjoined ears, and small scent-glands on each side of the penis.

Ears rather small, rounded, united at their inner bases for a height of about 2.5 millim., but without any folded lobe: keel slightly thickened in lower third; antitragus higher than long, rounded above, with a broad base; tragus minute, broader above than below, with an incurved upper angle and concave anterior margin. Lips wrinkled. No gular sac. Pad at base of thumb rather small. Wings to the distal third of the tibia. Tail involved in membrane for about half its length. Penis unusually short, or at least appearing short, as it projects by its tip only from the swollen mass of a pair of scent-glands surrounding its base, their two openings separated by a narrow septum situated halfway along the penis on its lower surface.

Fur very short and close, that on the forehead behind the junction of the ears appearing to be suffused with some oily secretion.

Colour dark brown above, more blackish on head; below also brown, the median area more greyish brown. Wing-membranes brown above and below; interfemoral membrane brown above, whitish below.

Skull low, stout and strong; brain-case small; muzzle narrow. Median crest not developed except in the interorbital region; premaxillæ united round the small palatine foramina. Upper incisors long, straight, almost parallel, not far apart; lower incisors four, bilobate, the outer pair rather smaller than the inner; anterior upper premolar present, but exceedingly minute, placed in the outer angle between the canine and large premolar, which touch one another.

Dimensions of the type (measured in spirit): -

Forearm 44 millim.

Head and body 64; tail 32; ear 17; tragus 2.0; height of antitragus 3; third finger, metacarpus 43, first phalanx 18, second phalanx 18; fifth finger, metacarpus 27, first phalanx 12.4, second phalanx 5; lower leg and foot (s. u.) 22.

Skull: greatest length 19.7; basal length in middle line 15.8; zygomatic breadth 12.7; anteorbital breadth 7; con-

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striction 3.8; mastoid breadth 11.6; front of canine to back of m^3 7.5; front of lower canine to back of m_3 8.5.

Hab. Mangala, N. of Gondokoro.

Type. Adult male. B.M. no. 2.7.4.3. Collected and presented by W. L. S. Loat, Esq.

I cannot find any known species to which this *Nyctinomus* can be assigned. None of Heuglin's show any relation to it, even apart from the peculiar gland above described, which he might have overlooked. It is curious that the two Mangala species should each be distinguished by a glandular structure unique of its kind.

Nyctinomus dubius A. Smith, 1833

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Nyctinomus dubius. (Caffer Nictinome.) Fur rather long; above the surface colour is a dull deep brown, beneath a light tawny brown inclined on the sides to blackish brown. Ears brownish black and pointed; wing membranes dull black with a faint brownish tint; interfemoral membrane enveloping nearly two-thirds of the tail, the last third very slender and pointed; hairs on the feet dull white; furrows of upper lip very indistinct. Length from nose to base of tail about two inches, of tail an inch and a half. - On the back, each of the hairs is tawny white towards its base; beneath towards the middle of the body, tawny white throughout; on sides tawny white towards the base.

Inhabits South Africa, - between the Cape Colony and Natal.

Nyctinomus Emini de Winton, 1901

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Ears united by a broad band, probably as in *N. pumilus*.

Size rather larger than *N. pumilus* and *N. limbatus*. General colour dark sooty brown, the centre line of the belly grey or pure white; a tuft of snow-white hair is also found at the base of the humerus, and these white hairs continue

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down the side to the groin. The ears, face, and entire wing-membranes are black.

Forearm 43 millim.

Type B.M. No. 90.6.8.15, M. Mosambi. Collected by Dr. Emin.

It is impossible to give a detailed satisfactory description of bats from dried skins.

Skull broader than that of *N. limbatus*, but not so broad as that of *N. major*, and distinguished from both by its prominent preorbital processes. The premaxillæ are joined, but are less perfectly ossified than in the allied species. The upper incisors are close together and parallel; the upper premolar is not in any way crowded, directly in the toothrow. Lower incisors not markedly bifurcate.

Nyctinomus fulminans Thomas, 1903

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General characters of *N. africanus*, Dobs., but with whitish belly and conspicuously smaller skull.

Size large, though rather smaller than in *africanus*. Ears

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large, evenly rounded, rising from the same spot on the muzzle; their anterior edge with four or five minute and inconspicuous warts; their keel moderately developed, not broadened or folded externally; antitragus as in *africanus*; tragus large, quadrangular, with rounded corners; a small angular projection present halfway down its outer margin. Wings to the ankles. Callosity at base of thumb unusually large. No gular sac.

Colour of body above dark chestnut-brown; below brown laterally, the median area of throat, chest, and belly white, becoming rather browner distally.

Skull smaller and conspicuously narrower than in *N. africanus*. A median crest developed along parietal suture, more developed than in *africanus*, but disappearing on interparietal, where the bone is smooth and transparent. Anteorbital and mastoid projections comparatively little developed. Premaxillæ incomplete mesially. Incisors $2/4$. Anterior upper premolar small, in general line of the tooth-row.

Dimensions of the type (measured in spirit): -

Forearm 60 millim.

Head and body 80; tail 53; ear 22; tragus on outer edge 7, breadth 4; height of antitragus behind 4; diameter of pollical callosity 3.2; third finger, metacarpal 57, first phalanx 25, second phalanx 7; lower leg and foot (s.u.) 28.

Skull: greatest length 22.5; basal length in middle line 17.7; zygomatic breadth 14; anteorbital breadth 8.5; interorbital constriction 4.7; mastoid breadth 13.2; front of canine to back of m^3 8.; front of lower canine to back of m_3 10.

Hab. Fianarantsoa, E. Betsileo, Madagascar.

Type. Male. B.M. no. 82.3.1.34. Collected by the Rev. W. Deans Cowan.

This is the "rather smaller, white-bellied, and light-winged form" of *N. africanus* referred to in Mr. de Winton's paper on the group. It is, I think, fully distinct enough to be separated specifically, as the skull-differences are very strongly marked, in addition to those of colour and locality.

Nyctinomus gambianus de Winton, 1901

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Much resembling *N. limbatus* in size and general appearance, but differing in having the lower parts uniform brown. The skull also closely resembles that of the East African species; but in this new bat the first upper premolar is only vestigial, so small, indeed that it is not possible to see it without the aid of a strong lens. This minute tooth is placed close against the canine, in the outer angle formed by the meeting of the canine and second premolar. The first lower premolar is also smaller in this species.

Forearm 37 - 38 millim. (type).

The males are not furnished with the crest of long hair on the back of the membrane connecting the ears, which is such a characteristic feature in *N. limbatus* from E. Africa.

Gambia, Lagos, Nigeria.

Type, M, B.M. No. 89.10.7.3. Gambia. Collected by Dr. Percy Rendall.

A specimen referable to this species formed the basis of Dobson's description of *N. pumilus*.

Nyctinomus hindei Thomas, 1904

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A whitish-winged member of the *N. pumilus* group.

Essential characters of ears, tragus, skull, &c., apparently as in *N. limbatus*, Peters. A marked tuft of brown, hairs behind the joining membrane of the ears.

Colour of upper surface chocolate-brown, finely flecked with white; the bases of the hairs (which attain about 4 - 4.5 mm. in length) rather lighter. Under surface brown, more or less washed superficially with whitish, especially along the middle line of the belly; a creamy white line edging the junction of the wings with the flanks. Ears, forearms, hind limbs, and interfemoral membrane dark brown. Wing-membranes near the body whitish brown, paling to white on the middle part of the wing, and darkening again at the tips to brown.

Skull about as in *N. Emini*, though with less marked preorbital processes. Small upper premolar outside the middle line of tooth-row, less crushed than in *limbatus*, more so than in *Emini*. Middle lower incisors deeply bifid.

Dimensions of the type (measured in skin): -

Forearm 40 mm.

Head and body (c.) 61; tail 35; thumb close to membrane 6; third finger, metacarpal 39, 1st phalanx 15.5; fifth finger 39.

Skull: greatest length 17.6; basal length 14.6; zygomatic breadth 11.4; front of canine to back of m^3 6.7.

Hab. Fort Hall, Mt. Kenya district, British East Africa. Alt. 4000 feet.

Type. Adult male. B.M. no. 3.3.2.4. Original number 134. Collected 1st Jan., 1903, and presented by Mrs. Hinde. Two specimens.

This *Nyctinomus* is most closely related to *N. Emini*, de Wint., of Usambiro*, German E. Africa, but differs by its whitish wings and more closely crushed upper premolars.

* Not Mosambiro, as accidentally printed in the original description, Ann. & Mag. Nat. Hist. (7) vii. p. 41 (1901).

Nyctinomus leonis Thomas, 1908

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Nyctinomus leonis, sp. n.

Nyctinomus brachypterus, Peters, Dobson, P. Z. S. 1878, p. 722; Cat. Chir. B.M. p. 426 (1878) (*nec* Peters, Reis. Mossamb., Säug. p. 59, pl. xv. fig. 1, 1852).

External characters as described by Dobson.

Skull of the high, not flattened *Nyctinomus* type, with well-marked median crest; emargination between premaxillæ very narrow, about 0.75 mm. in greatest breadth. Four lower incisors.

Measurements of type: -

Forearm 37.5 mm. (For other external measures, see Dobson.)

Skull: greatest length 19.4; basal length 15.5; zygomatic breadth 12.1; intertemporal breadth 4; mastoid breadth 11.3; palatal length 8; front of canine to back of m^3 7.

Hab. West Africa. Type from Sierra Leone. Other specimens from Fernando Po (*Capt. Downes*), Cameroons (*C. Bovallius*), and French Congo (*G. L. Bates*).

Type. Adult male skin. B.M. no. 62.12.23.3. Specimen c of Dobson's catalogue. Presented by J. Brown, Esq.

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This species, referred to *N. brachypterus* by Dobson, is evidently not that described by Peters, for the skull of the latter is no less than 21.5 mm. in length, while the typical skull of *N. leonis*, 19.4 mm. long, is the largest of the five skulls of the western form examined by me.

On the other hand, Dr. Jentink's *N. bemmeleni* from Liberia has, as he has been so good as to inform me, a skull only 16.5 mm. in total length, with a tooth-row length of just 6.0 mm.

Nyctinomus leucogaster A. Grandidier, 1869

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Nyctinomus leucogaster

Parties supérieures d'un brun foncé, ainsi que la gorge et la poitrine. Abdomen blanc. Ailes médiocres à membrane blanchâtre. Les deux incisives supérieures sont fortes et pointues; les quatre inférieures sont très-petites. Lèvres très-fendues et ridées; la supérieure est fort développée. Oreilles larges, réunies par leur base et penchées en avant; elles naissent à l'angle des lèvres; l'oreillon est court et extérieur. Les deux doigts externes des pieds sont plus forts que les autres et garnis de poils longs. Queue longue et enveloppée dans la membrane interfémorale dans sa première moitié seulement.

Envergure, 21^c 1/2. Long. du nez à l'extrémité de la queue, 8^c 1/2. Long. des oreilles, 14^{mm}; et larg., 13^{mm}. Long. de la queue, 3^c.

Hab. Mahab, au Ménabé.

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Nyctinomus leucogaster

Upperparts of a dark brown, as well as the throat and chest. Abdomen white. Wings mediocre with whitish membrane. The two upper incisors are strong and pointed; the four inferiors are very small. Lips Very split and wrinkled; the superior is highly developed. Ears wide, united by their base and bent forward; they are born at the angle of the lips; the tragus is short and outward. The two outer fingers of the feet are stronger than the others and have long hairs. Tail long and wrapped in the interfemoral membrane for its first half only.

Wingspan, 21^c 1/2. Length from the nose to the end of the tail, 8^c 1/2. Length ears, 14^{mm}; and width, 13^{mm}. Length tail, 3^c.

Hab. Mahab, at the Menabe

Nyctinomus lobatus Thomas, 1891

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Nyctinomus lobatus, sp. N.

Allied to and of the same size as *N. tæniotis*, Raf. (*N. Cestoni*, Savi), but distinguished by its much larger ears, tragus, and antitragus, by the thinness of the ear-membranes and keel, and by its belly being pure white. Forearm 63 millim.

Hab. Turquel, Sük, inland British East Africa. Coll. F. J. Jackson, Esq.

A more extensive description was made in Thomas ([1891b](#)): 1. On a collection of small mammals made by Mr. F. J. Jackson in Eastern Africa -

Proc. Zool. Soc. Lond., 1891, II: 181 - 187 (which according to Duncan ([1937](#): 74) was published in August 1891):

Nyctinomus lobatus, sp. N. ¹

a. ?. Turquel, Sük. 1/90. Type.

Allied to *N. cestoni*, Savi, or rather *N. tæniotis*, Raf., as it ought to be called, and to *N. africanus*, Dobs.; but distinguished from both by its larger ears, larger tragus, higher antitragas (fig. p. 183), and coloration, while in other respects it agrees sometimes with one and sometimes with the other, and forms in some respects a connecting-link between them.

Ears very large, rounded, laid forward they extend quite a quarter of an inch beyond the tip of the muzzle; their inner bases united on the muzzle; their substance comparatively thin and transparent; keel of the conch scarcely thickened below; antitragus very high

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posteriorly, the notch behind it more than 5 millim. Deep, as compared to about 3 millim. In the two allied species. Tragus large and broad, quadrangular, its tip sharply angular, and its outer upper border long and straight. Lips apparently not deeply wrinkled. Gular sac apparently present. Pads at base of thumb distinct. Lower incisors 4. Colour everywhere above, and on the chin, shoulders, and sides of thorax below dark brown; chest, belly, base of tail, and thighs to below knees pure white.

Dimensions: - Head and body 77 millim., tail 56; tail free from membrane 30; ears, length from base of post-antitragal notch to tip 30; forearm 63 (=2-47 in.).

This fine new Bat is distinguished at once from all the other species of the genus, except *N. miarensis*, Grand., and the two above-mentioned, by its much greater size, none of them having a forearm exceeding two inches in length. *N. miarensis* is at once separated by its structural characters and is not really allied to *N. lobatus*. Of the two species to which it is allied, *N. africanus* is a native of the Transvaal, and *N. tæniotis* of Abyssinia, extending northwards over a great part of the Palæarctic region: the new form is therefore intermediate between them in habitat as in many of its characters; but there can be no question as to the specific distinctness of all three.

¹ Preliminary descriptions of the new species were published in Ann. & Mag. N. H. (6) Ti. P. 303 (1891)

Nyctinomus martiensseni Matschie, 1897

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In der Grösse übertrifft diese Fledermaus noch *N. africanus* Dobs und *cestoni* Savi, ebenso wie *lobatus* Thos. Sie unterscheidet sich von allen übrigen afrikanischen Arten durch die ungeheuer grossen Ohren, bei denen Tragus und Antitragus vollständig verkümmert sind. Praemolaren $2 \cdot \frac{1}{2} - 2$; Ohren ziemlich durchscheinend; der Kiel der Ohrmuschel sehr schmal; Tragus verkümmert; Antitragus nicht angedeutet; die Ohren sind über der Nasenmitte verwachsen; Lippen sehr fein gefurcht, dünn, überhängend; Gularsack vorhanden.

Färbung: rauchbraun; eine Binde über Hals und Oberrücken weisslich.

Körper: von der Schwanzspitze zum oberen Ohrrand: 135 mm; Schwanz vom After: 43; freie Schwanzspitze: 28; Länge des Ohres vom vorderen unteren Rande der Oeffnung bis zur Spitze: 37; Unterarm: 66 mm.

Eine genauere Beschreibung dieser sonderbaren Fledermaus werde ich demnächst gelegentlich der Herausgabe der Peters'schen Fledermaustafeln geben.

Herr Martienssen, dem das Kgl. Museum für Naturkunde schon eine grosse Reihe sehr seltener Arten aus Usambara verdankt, hat ein M dieser Art bei der Plantage Magrotto unweit Tanga im Handei-Gebiet im August d. J. gefunden

Nyctinomus mastersoni Roberts, 1946

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A member of the *N. africanus*, *fulminans* and *lobatus* group, in which the upper lips are not wrinkled, the skull not so flattened as in the *N. aegyptiacus* and *bocagei* group, size rather large, hair on the integument broadly present between the humerus and base of the forearm and extending thinly in a line parallel with, but a little distance from, the forearm; in the dentition, with the upper anterior premolar small, its cusps about level with the cingulum of the large premolar, and lying rather on the outer side but within the tooth row. In size it is practically the same as in *N. fulminans* Thomas (*Ann. Mag. Nat. Hist.* (7), XII, 501; 1903) of Madagascar, and therefore smaller than *N. africanus* Dobson (*Ann. Mag. Nat. Hist.* (4), XVII, 348; 1876), known only by the type from Transvaal, and *N. lobatus* Thomas (*Ann. Mag. Nat. Hist.* (6), VII, 303; 1891) of the Suk country, Kenya Colony. It differs from *N. fulminans* in having the white median line below not extending to the throat, where the hairs are for the greater part white, but with dark brown tips, the pure white hairs only distributed from the pubic region to the chest. The reddish brown or chestnut colour characteristic of both *N. africanus* and *N. fulminans* may prove to be inconstant, for although the type and two other specimens are dark chocolate brown above and below, in one other specimen the upper parts are more reddish brown, while still another is decidedly erythristic, the upper parts being reddish brown and the under parts of body suffused with a brighter reddish colour, which extends to the white hairs of the integument

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between the femur and humerus, toning outwards to light buffy on the edges; but this erythrism is not uniform, as there is a broad normal coloured brown line on the right-hand side of the body both above and below. Normal coloured specimens agree best with the description of *N. lobatus*, chocolate brown above and below, except for the white median line and white on the integument adjacent to the sides. *N. fulminans* is said to have minute warts on the anterior edge of the ears (a character seen in *N. aegyptiacus* and *bocagei*), but these are lacking in the specimens examined of this new species. The callosity on the thumb is large, as in *N. fulminans*, and the skull is similar in having a sagittal crest from the interorbital region to the occiput, and also in having preorbital processes very slightly developed; the interparietals are swollen and raised slightly above the level of the parietals; the dentition is apparently the same, the anterior upper premolar small, in the tooth row, but rather pushed outwards; the cingulum of the lower canine is very much developed so that the pair of canines are not far apart and push the lower incisors (of which there are two pairs) forward; the anterior lower premolar is only about half the height of the adjacent large premolar, the latter with the usual high outer anterior and pointed cusp. There are apparently no glands present, either on the chin, or near the root of the tail on the border of the integument as described of *N. cisturus*.

Type: T.M. No. 9976, adult M, Chikupo Caves, Masembura Native Reserve, Bindura district, Southern Rhodesia, collected and donated by Mr H. B. Masterson; also four more specimens.

Nyctinomus Miarensis A. Grandidier, 1869

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Nyctinomus Miarensis

Semblable au *N. leucogaster*, mais de plus grande taille. Parties supérieures brunes; parties inférieures grises. Queue enveloppée dans la

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membrane interfémorale dans son premier tiers seulement.

Envergure, 39° 1/2. Long, du nez à l'extrémité de la queue, 12° 1/2. Long. de la queue, 4° (la partie libre mesure plus de 2° 1/2). Long. des oreilles, 2°. 3^{mm}; et larg., 2°, 2^{mm}.

Hab. Miari, pays situé entre Soua-haze et Soua-hané.

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Nyctinomus Miarensis

Similar to *N. leucogaster*, but larger. Upperparts brown; underparts gray. Tail wrapped in the

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interfemoral membrane for its first third only.

Wingspan, 39° 1/2. Length from the nose to the end of the tail, 12° 1/2. Length of the tail, 4° (the free part is more than 2° 1/2). Length ears, 2°. 3^{mm}; and width, 2°, 2^{mm}.

Hab. Miari, a country situated between Soua-haze and Soua-hané.

Nyctinomus ochraceus J.A. Allen, 1917

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Type, No. 48821, F ad. (skin and skull), Medje, March 16, 1910; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 745.

Ears thick, united at base in front. Coloration unusual for a molossid.

Above, superficially, dark chestnut-brown, the hairs ochraceous-buff basally, showing, more or less at the surface, especially on the shoulders and sides of the neck where it often gives the prevailing tone to the coloration; below orange, varying in tone in different specimens from pale to deep orange. A tuft of lengthened dark chestnut-colored hairs on the front part of the crown behind the connecting membrane of the ears. Ears blackish brown. Membranes naked; wing membranes from upper part of tibia; face naked, brownish. Interfemoral membrane narrow, much more than half the tail free. Tragus small, narrow and pointed, a little higher than wide. Antitragus low, evenly rounded above, broader at base than high, about 5.5 X 4.5 mm.

Collectors' measurements of type: Total length, 105 mm.; head and body, 72; tail, 33; foot, 11; ear, 18. Forearm (in skin), 37.7.

Collectors' measurements of type and 10 topotypes; Total length, 103 (100-106); head and body, 71.6 (68-73); tail, 31 (29-34); foot, 11 (all 11); ear, 17.5 (17-19).

Forearm, same specimens, 38 (37.3-39.3).

Skull (type), total length, 18.6; condylobasal length, 16.6; zygomatic breadth, 11.7; mastoid breadth, 10.8; breadth of braincase, 9; maxillary breadth, 8.3; inter-

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orbital breadth, 4; breadth at base of canines, 5.6; length of upper toothrow (with canine), 6.7; length of mandible, 12.2; angle to condyle, 3.6; depth at coronoid, 3.1; length of lower toothrow, 7.4.

Skulls of type and 10 topotypes, total length, 18.8 (18.2-19.4); zygomatic breadth, 12 (11.4-12.5).

Braincase low and flat, sagittal and lambdoid crests slightly developed, the latter indicated only in old adults; preorbital process absent or barely indicated. Premaxillae emarginate behind base of incisors. Upper incisors small; slightly convergent apically, widely separated from each other and the canines. First upper premolar very small, in midline of toothrow and not crowded between the adjoining teeth. Lower incisors 2-2, small, crowded in a convex row between canines. First lower premolar about half the size of the second. Coronoid process small and low, barely reaching the level of the condyles, and directed outward.

This striking species is represented by 22 specimens, 18 skins with skulls and 4 in alcohol, all taken the same day at Medje, and all adult. Only 4 are males, and of the 14 female skins, 11 are labeled as "carrying an embryo in right uterus." One of the alcoholic specimens is in poor condition, the ears and membranes being abraded and light colored, and the orange of the underparts is faded to pale yellow. This specimen was taken at first to represent a different but (owing to its condition) an unidentifiable species, and was so listed in this paper, but Mr. Lang assures me that this is one of the 22 specimens of *N. ochraceus* taken at Medje, all on the same day, and that its present abraded and bleached condition is due to bad preservation due to an accident in transportation.

Nyctinomus ochraceus appears to be related to *N. thersites* Thomas but it is smaller (forearm 38, in *thersites* 41; skull 18.8 X 12; in *thersites* 20 X 12.5), and the lower premolars are not subequal as in *thersites*. *N. thersites* is described as "colour dark brownish chestnut, the glandular hairs behind the junction of the ears black." This applies fairly well to the upperparts of *N. ochraceus*, except that the lengthened hairs behind the ears in males are not black but dark chestnut. It is hardly possible, however, that this brief description could apply if the lower parts had been ochraceous-orange, as in the species here described.

Nyctinomus pusillus Miller, 1902

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Nyctinomus pusillus sp. nov.

Type. - Adult female (in alcohol), No. 37852/20997^(*) United States National Museum. Collected on Aldabra Island, Indian Ocean, by Dr. W. L. Abbott.

Characters. - Externally similar to *Nyctinomus pumilus*. Skull and teeth noticeably smaller than in the related species.

External features. - In size as well as in details of external form *Nyctinomus pusillus* agrees so closely with *N. pumilus*, as to need no description.

Skull and teeth. - The skull is similar to that of *N. pumilus*, except that it is smaller and more lightly built, the interorbital profile is more concave, the basisphenoid pits are better defined, and the interpterygoid space is relatively wider. Teeth much smaller than in *N. pumilus* and inner segment of upper molars relatively broader, so that the protocone and hypocone of the first and second are actually further apart than in the larger species.

Measurements. - External measurements of type: total length, 84; head and body, 53; tail, 31; tibia, 11; foot, 7.8 (7); forearm, 46.6; first digit, 6.4; second digit, 35; third digit, 70; fourth digit, 58; fifth digit, 34; ear from meatus, 15.4; ear from crown, 10; width of ear, 14.

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Cranial measurements of type: greatest length, 15 (16.6);* basal length, 13.4 (14.8); basilar length, 12.8 (13.4); zygomatic breadth, 9.2 (10); interorbital constriction, 3 (3.6); greatest breadth of braincase above roots of zygomata, 8 (8.4); lachrymal breadth, 5 (6); mandible, 10 (11); maxillary toothrow (exclusive of incisors), 5.6 (6.6); mandibular toothrow (exclusive of incisors), 6 (6.8).

Specimens examined. - Two, both from Aldabra. Island.

* Measurements in parenthesis are those of an adult male *Nyctinomus pumilus* from Massowa, Red Sea (No. 38933/15546).

^(*) Poole and Schantz (1942: 120) mention "Type designated by No. 37852/20997, an error for 20991/37852".

Nyctinomus rhodesiae Roberts, 1946

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Seemingly related to *N. ansorgei* Thomas (*Ann. Mag. Nat. Hist.* (8), XI, 318; 1913) from Malange, Angola, in general characters of size, colour, skull and dentition, but lacking the 'partly naked patch on the crown behind the junction of the ears', which is thinly haired in this new form, and the naked patch 'across the nape just in front of the shoulders', of which there is no sign in the present form. There are no glands on the border of the interfemoral membrane at the base of the tail, but a small gular gland, surrounded by a few short hairs, is present in the middle of the naked area of the chin. Ears in contact at the base. The skull has the cranium slightly raised above the dorsal level, the premaxillae not in contact, a slight sagittal crest from the interorbital constriction to the occiput, and a well-developed process interorbitally, near the anterior corner of the orbits; the upper incisors lie parallel but not in contact, the upper anterior premolar in the tooth row and small, its height about equal to the level of the cingulum of the large premolar; the lower canine has a much developed cingulum, so much so that the canines are either very close together or actually (as in the type) in contact; the two pairs of lower incisors are therefore rather crushed and pushed forward; the anterior lower premolar is as large as the posterior premolar, or even, as in the type, rather larger, and its cusp nearly as high as that of the large premolar. The upper lips are wrinkled with a few short bristles on the ridges, the ears without nodules on the anterior upper edges and the tail protrudes for at least two-thirds of its length (tail length 36 mm., free portion 24 mm.).

In colour this species is like *N. ansorgei* in being dark brown above, the throat broadly blackish and the underparts of body paler than the upper, the tips of the hairs tending to be lighter coloured and even whitish in some specimens, and the integument at the femurs tending to be white. The chin is very sparsely haired, almost naked.

Type: T.M. No. 9977, adult M, Chikupo Caves, Masembura Native Reserve, Bindura district, Southern Rhodesia, collected and donated by Mr H. B. Masterson; also five more specimens from the same source.

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Measurements of the above two new species are as follows:

(4) *N. mastersoni*; (4) *N. rhodesiae*
 Head and body: 80 - 85; 65 - 67
 Tail: 55 - 57; 36 - 36
 Free portion of tail: 26 - 28; 24 - 25
 Ear: 20 - 23; 19 - 21
 Tibia and feet, c.u.: 31 - 33; 23 - 24
 Skull: greatest length: 22 - 23; 19 (1)
 median dorsal length: 18.3 - 19; 16.5 (1)
 basilar length: 17.2 (1); 15.2 (1)
 zygomatic width: 13.2, 14 (2); 11.5 - 12 (3)
 mastoid width: 12.7, 13 (2); 10.5 - 11 (3)
 width of brain case: 11.5 (3); 10 (3)
 anterior interorbital width: 7.1 - 8.5; 6.3 - 7.1
 interorbital constriction: 4.5 - 5.2; 4.2 (3)
 posterior median height: 7.8, 8 (2); 6.5 (1)
 height at m³: 8 - 8.5; 7 - 7.5
 c-m³ length: 8.5 - 9.2; 7 - 7.5
 mandible length: 16 - 16.5; 13 - 13.4
 c-m₃ length: 9.5 - 10.1; 8.1 - 8.5

Nyctinomus spillmanni Monard, 1933

Nyctinomus Spillmanni

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Trois individus, une femelle, un mâle et un très jeune de Vila da Ponte.

Le synopsis de Dobson conduit à *pumilus*; mais une série de caractères ne concordant pas, je donne du male une description complète.

Museau prolongé de 4 mm. au-devant de l'ouverture buccale, à bord supérieur tranchant. Lèvres sillonnées verticalement de

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plusieurs plis, les plus profonds au milieu de la distance séparant les commissures des narines. Celles-ci séparées par un sillon.

Oreilles jointes, commençant à 4 mm. en arrière des narines. La bande qui les unit est couchée sur le crâne, formant une protubérance au-dessus du museau, large de 4 mm., sillonnée et échancrée au milieu. Les oreilles rabattues en avant atteignent presque les narines; elles sont rondes et épaisses. Le pli qu'elles présentent est bien marqué, aplati, saillant, arrondi proximement, et dirigé en dehors.

Antitragus trapézoïdal, commenç. ni par une oblique à 3 mm. de la commissure des lèvres, séparé du pavillon par une profonde échancrure; la petite base est de moitié plus courte que la grande base. Tragus très petit, carré, à angles supérieurs vifs, avec un très petit lobe basal extérieur. L'oreille est nue, à l'exception d'une bande de poils roux le long du pli interne, de la bande commissurale, et d'une plage située à la base et derrière la conque.

Le museau porte des touffes de poils raides devant la commissure des oreilles; celles-ci portent près de leur sommet quelques points cornés.

Queue épaisse, dépassant longuement la membrane (13 mm. + 18 mm.). Membranes partant du quart inférieur du tibia.

Pelage entièrement brun roussâtre, un peu plus clair à la face ventrale.

Mâle: membranes brunâtres dessus; dessous, une bande de poils blancs, large de 8 mm. environ, borde les flancs. Humérus et radius blanchâtres dessous. Les membranes de la femelle sont blanchâtres dessus et dessous, avec la même bande de poils blancs.

Chez le mâle, les incisives de lait ne sont pas tombées encore; elles se trouvent devant les incisives médianes et sont très crochues. La première prémolaire est petite et est située entre la canine, petite, et la deuxième prémolaire. Les autres dents sont normales.

L'attribution spécifique de cet individu est douteuse. La forme trapézoïdale de l'antitragus, et la fourrure, brune partout, ne permettent pas de l'attribuer à l'espèce *limbatus* de Peters, de laquelle il se rapproche aussi. Il paraît plus voisin de *brachypterus* de Peters, mais ni la couleur du pelage, ni la forme de l'antitragus ne correspondent exactement.

Les espèces décrites par Seabra ont les oreilles séparées et sont voisines de *aegyptiacus*. Les quelques *Nyctinomus* d'Oldfield Thomas, dont nous possédons la description (*Hindei*, *fulminans*, *cisturus*, *demonstrator*, *thersites*) ne concordent pas.

Femelle:

Longueur de la tête et du corps: 78 mm.

Longueur de la queue: 14 + 24 mm.

Avant-bras: 49 mm.

Tibia : 17 mm.

Nyctinomus thersites Thomas, 1903

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A medium-sized species, with proportionally short limbs, united ears, separated premaxillæ, and very small lower cheek-teeth.

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Body large as compared with the short forearms and legs. Muzzle short, upper lip distinctly furrowed. Ears short, opaque, their tips broadly rounded; inner margins united at base; anterior margin without minute horny points; antitragus as high as long, with a deep notch behind it; keel thickened below, but not flattened externally. Tragus minute, almost linear; a separate, very distinct, external basal projection developed halfway between its outer base and the inner side of the antitragus. Fur short, close, and velvety; hairs of back barely $2\frac{1}{2}$ mm. long, tufts* of long hairs (about $\frac{1}{2}$ inch in length) placed on each side of the rump; fur continued to body throughout, except a narrow rim along each side below. Colour dark brownish chestnut, the glandular hairs behind the junction of the ears black. Penis projecting but little from the surrounding flesh, without special scent-glands. Premaxillæ separated, but the opening between them small; anteorbital crests scarcely developed; sagittal crest low; lambdoid crest strong, forming a prominent projection on each side of the middle line. Lower jaw unusually thick and heavy.

Upper incisors thick, vertical, parallel, well separated. Anterior premolar small, but standing in the tooth-row, and separating the canine from the large premolar. Lower incisors four, bifid, overlapping. Lower premolars very abnormal in being subequal, the anterior scarcely shorter than the posterior. Molars unusually low-crowned, their vertical height considerably less than that of the thick mandibular ramus.

Dimensions of the type (measured on the spirit-specimen): -

Forearm 41 mm.

Head and body 75; tail 32; ear 17; tragus on inner edge 1.2; third finger, metacarpus 40, first phalanx 17, second phalanx 16; fifth finger, metacarpus 26, first phalanx 9, second phalanx 4; lower leg 15, lower leg and foot (s. u.) 23.

Skull: greatest length 20; zygomatic breadth 12.5; front of canine to back of m^3 7.3.

Hab. Efulen, Cameroons.

Type. Old male. Collected by Mr. Gr. L. Bates. Three specimens.

By Dobson's synopsis this bat comes near *N. pumilus*, but may be readily distinguished from that species by its larger size and the many peculiarities, external and cranial, described above.

* These tufts appear to occur in many species, though seldom so well developed as in the present animal.

Nyctinomus unicolor A. Grandidier, 1870

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Nyctinomus unicolor

d'un brun assez foncé, qui est a peine plus clair sous l'abdomen. Queue libre dans plus de la moitié de son étendue. Oreilles fort grandes, qui sont réunies sur le front par une petite membrane. Lèvres épaisses et larges.

Longueur totale, 13 cent.; de la queue, 42 cent.; de la partie de la queue enveloppée dans la membrane interfémorale, 15 mill.; envergure, 41 cent. 1/2.

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Nyctinomus unicolor

a rather dark brown, which is scarcely clearer under the abdomen. Tail free for more than half of its extent. Ears very large, which are united on the forehead by a small membrane. Lips thick and wide.

Total length, 13 cent.; tail, 42 cent.; part of the tail wrapped in the interfemoral membrane, 15 mill.; wingspan, 41 cent. 1/2.

Otomops icarus Chubb, 1917

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Otomops icarus, sp. nov.

Like *O. wroughtoni* and *martiensseni* in the essential characters of skull and ear structure, including the practically complete suppression of both tragus and antitragus.

Colour very much as in *O. martiensseni*, the fur of the top of the crown, above the bases of the united ears, similarly pale brown and the back of the head dark brown, both with whitish bases to the hairs, in contrast to the deep uniform blackish-brown of the same parts in *O. wroughtoni*. But as compared with *O. martiensseni* the prominent whitish mantle so conspicuous in that animal is almost obsolete and there is but little whitish on the front of the neck. Lower part of body both above and below, deep blackish-brown, very much as in *O. wroughtoni*.

Skull more elongate than in *wroughtoni* or in the Congo specimen of *martiensseni*. Prof. Matschie does not give any cranial dimensions in his original description of the latter species.

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Dimensions of Type.

Forearm 66 mm.

Head and body 88 mm.; tail 45 mm.; ear 38 mm.; fifth finger, metacarpal 34 mm., first phalanx 20 mm., second phalanx 8 mm.; lower leg and foot (s.u.) 30 mm.

Skull: greatest length 27.5 mm.; basal length in middle line 24 mm.; zygomatic breadth 14 mm.; anteorbital breadth 10 mm.; interorbital constriction 5.7 mm.; mastoid breadth 14 mm.; front of canine to back of m_3 11 mm.

Hab. Durban, found helpless in gutter, apparently as a result of collision with street electric light.

Type. Male.

Otonycteris Peters, 1859

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Zwei Exemplare dieser neuen Gattung befinden sich im zoologischen Museum, welche aus der Sammlung der Hrn. Hemprich und Ehrenberg stammen sollen. Sie hat durch den Bau der Ohren und des Ohrdeckels die grösste Ähnlichkeit mit der Gattung *Plecotus* und war unter diesem Namen auch aufgestellt; jedoch sind die Nasenlöcher nicht nach hinten erweitert, noch auf der oberen Seite gelegen, sondern sie sind einfach sichelförmig und nach vorn gerichtet wie bei der Gattung *Vespertilio*. In der Gestalt des Schädels nähert sich diese Gattung am meisten den *Nycticejus* und ebenso stimmt sie auch hinsichtlich der Gestalt und Zahl der Zähne ganz mit *Nycticejus (planirostris)* Pet.) überein: $3^1/2 \cdot 1/2 \cdot 1/1^{1/1} \cdot 1/6 \cdot 1/1^{1/1} \cdot 1/3 \cdot 1/2 \cdot 3 = 30$.

Otonycteris Hemprichii Peters, 1859

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supra albescenti-brunneus, subtus albus, alis dilute brunneis.

Long. tot 0.110; cap. 0.025; aur. 0.030; tragi 0.015; caudae 0.045; antibr. 0.058; exp. alar. 0.320.

Ist diese Art übereinstimmend mit Gray's *Plecotus Christii*?

Pachyotus Gray, 1831

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The Bats, the Vespertiliones of Geoffroy, might for convenience be divided into three genera, the true Bats, *Vespertilio*, with thin ears and membranes and a hairy face, the *Pachyotus* with thick ears and membranes and bald swollen cheeks, including the genera *Nycticeius* and *Scotophilus* and the hairy-tailed species of America, (*Lasiurus*.)

Petalia (Nycteris) thebaica aurantiaca de Beaux, 1923

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(N. 1432) Tipo 1 F, in alcool, cranio estratto. Archers Post, I.1920, L. FRANCHETTI e L. TONKER.

Stato di conservazione buono, astrazione fatta per un processo arrestato di dissolvimento dell'integumento in corrispondenza del carpo e di alcuni diti.

Rivestimento peloso abbondante e soffice; singoli peli lunghi 12 mm. tra le spalle. Colorazione generale arancio-ocracea (ochraceous orange, Ridgw. XV) con brillantezza arancione (orange, III); i singoli peli lievemente più chiari alla base. Parti ventrali soltanto impercettibilmente più chiare delle dorsali. alla base dorsale di ciascun orecchio una indistinta mac-

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chia chiara color cuoio aranciato (orange buff, III). Tale colore hanno anche i peli sulla superficie ventrale della membrana alare in corrispondenza del $\frac{1}{4}$ prossimale dell'avambraccio. Patagio bruno fosco (fuscous, XLVI); orecchio lievemente più chiaro.

Unghia del pollice uniformemente grigia chiara. Unghie del piede color crema diafano nella metà basale, intensamente bruno-aranciato nella metà distale.

L'*orecchio* ed il *trago* in nulla differiscono dalle *Petalia thebaica*, Geoff., d'Egitto.

Il *cranio* corrisponde per forma e dimensioni alle *P. th.* suddetta.

I *denti* invece offrono alcune particolarità degne di nota.

Premetto che gli II sono in parte anormali. I_2 des. è spezzato alla base "ex vita". I_1 des. è largo et *trifido*. I_1 sin. è stretto e bifido, ma ha il lobo laterale troppo corto. I_2 sin. è bifido e di forma identica a quella del *P. th.*, Geoffr.

E peraltro indubitabile trattarsi di una *Petalia* con incisivi superiori bifidi!

P_4 è molto piccolo; resta nella linea dentale meglio che nella *P. th.*; è visibile dall'esterno, ed ancor meglio dall'interno, ma è appena visibile *dal disopra*, perchè P_3 et M_1 sono molto ravvicinati tra di loro e lo celano coi rispettivi cingoli, che si trovano a contatto o quasi l'uno dell'altro.

C è più grande e più decisamente uncinato che nella *P. th.* C è più alto, con profilo interno più rettilineo e punta mediale del cingolo più sviluppata. P_3 è più alto e mostra una piccola cuspidi anteriore particolarmente distinta. I MM sono tutti particolarmente alti e strettamente serrati l'uno contro l'altro. M_1 ha la cuspidi anteriore particolarmente alta. M_3 è particolarmente lungo. I MM hanno il lobo basale interno (mesostile) assai più largo e grosso, ed esteso all'indietro con una punta tondeggiante.

Uguali risultati ottengo dal confronto con *Petalie* d'uguale statura di Keren, Setit, Agordat, Lugh, Aden.

Misure del Tipo. Testa più tronco mm. 50. Coda 05. Altezza dell'orecchio, margine mediale 26, laterale 30. Larghezza massima dell'orecchio appiattito 20.5. Altezza del trago 7. Braccio 21. Avambraccio 44. Pollice con unghia 10.5. Dito terzo, me-

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tacarp 36, falange prima 23, fal. seconda 23. Coscia 20. Gamba 19. Piede con unghia 10.5. Sprone 14.5.

Cranio. Lunghezza massima con mandibola 20. Lungh. mass. C. -Occipite 10. Lungh. mass. della mandibola con II 12.4. Larghezza sull'arcata zigomatica 10.6. Larghezza della cassa cranica sulla squama temporale 8.4. File dentale $C-M_3$ 6.5. $C-M_3$ 7. Altezza massima C 2.3. C 2.1. M_1 2.6.

Reputo indispensabile far seguire una tavola sinottica in cui alcune delle precedenti misure sono messa a confronto con quelle di altre forme componenti il "gruppo *P. thebaica*".

[table removed - eds.]

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Dalla presente tavola risulta chiaramente che vis sono entro il "gruppo thebaica" due distinte forme, una piccola settentrionale ed una grande meridionale. Ambedue si estendono pressochè in ugual misura al di là dell'Equatore, la piccola scendendo fino a Zanzibar ed oltre, la grande salendo fino nell'Eritrea. Attribuendo a queste due forme valore specifico, la piccola dovrà chiamarsi *P. thebaica*, Geoff., la seconda *P. capensis*, Sm.

Risulta inoltre che le dimensioni dell'orecchio, la proporzione tra lunghezza dal braccio e dell'avambraccio, e la proporzione tra lunghezza e larghezza della testa sono tutt'altro che costanti entro una stessa forma della stessa località.

Di ciò resta infirmata la validità specifica della *P. revoili*, Rob. (Bull. Soc. philom. Paris, 1881, V, p. 90; e Ann. Sc. Nat. Zool. XIII, 1882, art. 2, p. 3, Tav. 12 Fig 5-6-7); molto più che nemmeno il carattere che dovrebbe "immediatamente distinguere la *P. revoili* dalle due specie *P. thebaica*, Geoffr. e *P. angolensis*, Pet." (= *capensis*, Sm.), e cioè la situazione di P_4 nella fila dentale, ove è visibile dall'esterno (l.c. fig. 5), ha valore specifico, giacchè riscontro un P_4 ben visibile dall'esterno e *dal*

disopra, e ben piantato nella fila dentale perchè P_3 e M_1 sono sufficientemente distanziati tra di loro, in esempl. di *P.* del Caio, Egitto, Lungh., Zanzibar, Aden, Keren, ed inoltre di *Saganeiti*, Harrar, Angola, Capo; mentre P_4 è scarsamente visibile dall'esterno, *poco o punto visibile dall'isopra*, perchè P_3 et M_1 sono molto ravvicinati tra di loro, e talvolta anche abbastanza energicamente spinto verso l'interno, in esemplari di Korosco, Setit, Agordat, *Saganeiti*, Harrar, Angola.

Gl. M. Allen (Bull. Mus. Comp. Zool. Harv. Coll. LIV, 9, 1911, p. 323) attribuì daltronde "only after much deliberation" 3 Petalie del Guaso Nyiro settentr. alla *P. revoili*, unicamente per le misure somatiche, che viceversa riscontro rientrare in quelle ottenute anche su Petalie egiziane soltanto.

Ed il Lönnberg (Arkiv Zool. Stockh. 10, 1916, art. 12 p. 5) attribuì 1 *P.* del gruppo thebaico alla *P. revoili*, pure unicamente per le dimensioni somatiche, appoggiandosi ad Allen (l.c.). Ma come nel caso precedente riscontro tali dimensioni presso *P. thebaica* egiziana, eccezion fatta per l'orecchio lievemente (3 mm.) più lungo.

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L'attribuzione della *P.* del gruppo thebaica dell'Eritrea, Africa orient. inglese ed Uganda alla *P. revoili* (K. Andersen, Ann. M. Nat. Hist. London 1912, X, p. 550) deve quindi sembrare alquanto arbitraria, tanto alle caratteristiche dentali precedentemente discusse.

Ma se, come credo, la *P. thebaica* dell'Eritrea non va distinta sottospecificamente da quella dell'Egitto (v. Senna, Arch. zool. Napoli, II, 3, 1905, p. 284) anche la sottospecie *adana*, K. A. (l.c.) resterà difficilmente distinguibile dalla thebaica tipica.

Credo infine, in base alla descrizione-tipo (Peters, Monatsb. Ak. Wiss. Berlin, 1870, p. 905, fig. 7), che la *P. damarensis* possa essere considerata tutt'al più come una sottospecie, di *P. capensis* Sm. e la stessa sorte seguirebbero quindi la *P. d. brockmani*, K. A. e la *P. d. media* K. A. (l.c.) non sempre distinguibili tra di loro (confr. tavola delle misurazioni).

Tornando ora alla *P. di Archers Post*, penso ch'essa *rappresenti in sostanza una varietà di colorazione della P. thebaica* (confr. Senna, l.c. p. 281 per l'*Asellia tridens*, Geoff), ma mi trattengono dal considerarla fin d'ora come tale le caratteristiche dentali rilevante, la quasi uniformità di colorazione tra parti super. ed infer., la singolare distribuzione di colore nelle unghie del piede.

Il quadro sinottico del gruppo "*P. thebaica*" (v. K. Andersen, l.c. 1912) andrebbe pertanto a mio avviso così modificato:

P. thebaica, Geoffr. Egitto, Sinai, Africa orient. inglese e tedesca, Zanzibar;

[*P. th. adana*, K. A.]. Aden.

[*P. th. revoili*, Rob.]. Somalia settentr.

P. th. aurantiaca, de B. Africa orient. Brit.: Archers Post.

P. th. gambiensis, K. A. Gambia

P. capensis, Sm. Africa merid. fino alla Zambesia ed Angola.

[*P. c. damarensis*, Pet. Damaraland, Namaqualand, Lago Ngami, Tette]

[*P. c. brockmani*, K. A. Eritrea, Somalia.

[*P. c. media* K. A.]. Abissinia (Harrar).

Petalia aurita K. Andersen, 1912

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Dentition (incisors and p_4), tragus, and external dimensions as in *P. hispida*, but ears much longer, skull larger, toothrows longer. Forearm of type 43 mm. (in forty-seven adult *P. hispida* 37.2-43), ear from base of inner margin 21.5 (16.5-18.7); skull, total length 18.7 (in thirty-two adult *hispida* 16.5-17.8), condyle to front of canine 16.1 (14-15.3), c-m³ (crowns) 6.5 (5.5-6.1).

Type, F ad. (alc.), Kilifi, British East Africa, collected and presented by G. D. Trevor-Roper, Esq., B.M. 89.1.11.1.

Other specimens from Maungu, B.E.A., and Burao, Somaliland.

Petalia damarensis brockmani K. Andersen, 1912

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Probably the north-eastern representative of *P. d. damarensis*, from which it is distinguishable only by the larger average size of the skull. Total length of skull (fourteen specimens) 21-21.5 mm. (20-21.2 in six *d. damarensis*), total length of lower jaw 13.7-14.2 (12.7-13.7), maxillary tooth-row (crowns) 7.2-7.7 (7-7.5). Forearm 47-51, ear from base of inner margin 29-31.5 mm.

Type, F ad. (skin), Upper Sheikh, British Somaliland, 4300', 11 Jan., 1910, collected and presented by Dr. R. E. Drake-Brockman, B.M. 10.3.27.4. The range of this form is known to extend to Erythrea. *P. d. damarensis* occurs in Damaraland and Namaqualand, through the Lake Ngami region, eastward to Tette.

Note. - *P. d. brockmani* should not be confused with the smaller, shorter-eared *P. revoili*, which belongs to the same section of the genus and occurs in the same region (forearm 41-45, ear 26-26.5, maxillary tooth-row 6.5-6.8 mm.).'

Petalia damarensis media K. Andersen, 1912

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Probably the Abyssinian representative of the foregoing form, from which it differs by its conspicuously smaller size: total length of skull of type 20.6 mm., of lower jaw 13, maxillary tooth-row 6.8, forearm 45.5, ear from base of inner margin 29.

Type, ad. (skin), Harar, Abyssinia, 19 Jan., 1912, collected by Hr. G. Kristensen, presented by the Hon. N. C. Rothschild, B.M. 12.2.28.1.

Petalia gambiensis K. Andersen, 1912

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Tragus of the *P. thebaica* type, but species differing from any other form of the *thebaica* section by its remarkably small size. Forearm (type and topotype) 39-40.8 mm.,

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third metacarpal 29, ear from base of inner margin 23-23.5, skull (total length) 18.5, maxillary tooth-row (crowns) 6.1-6.2.

Type, ad. (akin), Dialocote, French Gambia, 7 March, 1910, presented by G. Fenwick Owen, Esq., B.M. 11.6.10.10.

Note. - Gambia is inhabited by two other species of *Petalia*, *P. hispida* and *P. macrotis*.

Petalia major K. Andersen, 1912

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Similar to *P. arge* (p₄ large, tragus lingulate), but easily distinguished by its conspicuously larger size, being about equal in dimensions to *P. æthiopica luteola*. Forearm of type 49 mm. (39.5-45 in ten adult *arge*); skull, total length 22.2 (19.5-20.2), condyle to front of canine 19.7 (16.8-17.7), maxillary tooth-row 7.8 (6.8-7.3).

Type, F ad. (alc.), Ja R., Cameroons, 23 Jan., 1906, collected by Mr. G. L. Bates, B.M. 9.10.2.49.

Petalia nana K. Andersen, 1912

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Allied to *P. arge* (p₄ large), but considerably smaller, and with proportionately much smaller ears (tragus not differing in shape, being lingulate as in *arge*). Forearm of type 34 mm. (39.5-45 in ten adult *arge*), third metacarpal 25.7 (30.5-34.2), tibia 14.5 (20-23.7), ear from base of inner margin 15.5 (23-26.5), maxillary tooth-row (c-m³, crowns) 5.7 (6.8-7.3).

Type, M ad. (alc.), Benito R., French Congo, collected by Mr. G. L. Bates, B.M. 0.2.5.46.

This is the smallest known form of the genus.

Petalia parisii de Beaux, 1924

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(N. 1449) Tipo. F, Balli, Somalia it. merid., 6 IV 1922. In alcool. Cranio estratto.

Stato di conservazione. In generale buono. Manca il pelo nella porzione mediana delle parti inferiori e sul dorso eccettuate la spalle.

Appartiene al gruppo *P. aethiopica* (Kn. Andersen, Ann. Mag. Nat. Hist. X, 1912, 549). Si distingue nettamente dalla *P. aethiopica*, Dobs. e *P. aeth. luteola*, Thos. per le dimensioni molto minori; dalla *P. macrotis*, Dobs. e *P. oriana*, Kershaw (Ann. Mag. Nat. H. X, 1922, 179) per le dimensioni molto minori e la forma del *Tragus*; dalla *P. woodi*, K. A. (Ann. Mag. Nat. H. XIII, 1914, 513) per le dimensioni del padiglione molto minori (mm. 21 contro 29) e per il colore delle parti inferiori, che è uguale alle superiori, anziché bianco.

Tragus. Come quello rappresentato dal Dobson (Catal. 1878, Pl. XI, fig. 3) per la *P. aethiopica*, ma munito alla base del margine posteriore di un grosso lobulo orientato lateralmente.

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Dimensioni somatiche. Testa e tronco min. 47; coda 46; orecchio, margine interno 21; avambraccio 43; pollice (senza metacarpo, con unghia) 9; 3° dito: metacarpo 31.5, prima falange 22, seconda falange 22.2; 4° dito: metacarpo 33, prima fal. 11.8, seconda fal. 9; 5° dito: metacarpo 34, prima fal. 11.5 seconda fal. 10.8; tibia 19; piede senza unghia 8; unghie in media 2.

Cranio. Lunghezza massima colla mandibola in situ, II-occipite, mm. 19. Lunghezza massima C-occipite 18. Larghezza zigomatica 10.7. Larghezza massima sullo scudo frontale 7. Lungh. massima della mandibola, denti esclusi, 11.8. Lungh. massima della fila dentale superiore 6. Fila dentale inf. 6,9.

MM assai logori. CC relativamente grandi e robusti. P₃ assai piccolo, nella, fila dentale, prevalentemente mediale, ma pur riconoscibile anche nella veduta laterale.

Rivestimento peloso. Soffice e lungo (10 mm. sulle spalle) Tutto il braccio ed il terzo prossimale dell'avambraccio sono riccamente rivestiti. Il patagio è rivestito dorsalmente: nella metà prossimale della zona tra braccio ed avambraccio; su di una striscia larga 10 mm. lungo il tronco; nella zona tra la coscia e le prime due vertebre caudali. Ventralmente nella metà prossimale della zona tra braccio ed avambraccio, e, più scarsamente, nella zona tra gomito e ginocchio.

La colorazione d'insieme è buffy brown (Ridgw. XL) sulle spalle, più scura sul braccio; drab (XLVI) sulla testa e le parti inferiori. Patagio ed orecchio sono cinnamon drab (XLVI). I singoli peli sono lievemente più scuri e più grigi nel terzo o quarto basale. La zona nuda nasale è lateralmente orlata da una striscia di peli bianchi.

Unghie grige brune basamente; chiare e diafane apicalmente.

Dedico questa nuova specie al Dottor Bruno Parisi, Direttore della Sezione Zoologica nel Museo Civico di Storia Naturale di Milano.

Petaliidae Miller, 1910

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The generic name *Nycteris* as used for a group of Old World bats is currently assumed to date from Geoffroy and Cuvier, 1795. In reality it was not published in this sense until 1803, the name as it occurs in the earlier work being strictly a *nomen nudum*. But during this interval of eight years *Nycteris* had been applied by Borkhausen and Bechstein to a very different animal, the New York Bat of Pennant, now currently known as *Lasiurus borealis*. It is therefore necessary to substitute *Nycteris* Borkhausen 1797 for *Lasiurus* Gray 1831, *Petalia* Gray 1838 for *Nycteris* Geoffroy 1803, and the new family name *Petaliidia* for *Nycteridae* Dobson 1875

Phyllorhina commersoni var. *thomensis* Bocage, 1891

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Deux individus, M et F adultes.

Ces individus ressemblent à *Ph. Commersoni* par leur conformation générale, par la forme de leur fer-à-cheval et par leurs caractères de dentition; mais ils diffèrent de nos individus d'Angola et d'autres localités d'Afrique occidentale: 1° par leur taille, sensiblement plus petite; 2° par le nombre des plis cutanés latéraux du fer-à-cheval, trois au lieu de quatre; 3° par leurs couleurs, d'un brun-noirâtre foncé et uniforme en dessus, plus pâles en dessous, avec une tache blanchâtre sur le tiers supérieur de la face interne du bras. Les membranes des ailes et la peau nue des membres sont noires.

Chez la femelle de cette variété, comme chez la femelle du *Ph. Commersoni*, l'orifice du sac frontal est bien distinct, et ses appendices pubiens sont assez développés ainsi que les mamelles, les uns et les autres noirs.

Dimensions:

M; F

Longueur total (tête et tron): 106 mm; 102 mm.

- " de la tête: 34; 32
- " de l'oreille: 25; 35
- " de l'avant-bras: 82; 81
- " du 3° doigt: 126; 124
- " du pollex: 18; 17

" de la queue: 29;30

Habitat: la femelle a été recueillie à Roça Saudade, le mâle à Ribeira peixe.

L'espèce est connue à St. Thomé sous le nom de Guimbu.

Comme je l'ai déjà remarqué ailleurs, M. Greeff cite à peine deux chauves-souris de St. Thomé, *Cynonycteris straminea* et *Phyllorhina caffra*, l'une et l'autre non encore rencontrées dans cette île par M. Newton (V. Journ. Ac. Sc. de Lisboa, 2.^a série, I, p. 197).

Phyllorhina commersonii var. *marungensis* Noack, 1887

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(Vergl. Taf. X, Fig.31 - 33.)

5 Exempl., darunter 1 F, Juli und August, Qua Mpala, Marungu. "Fliegt in der Dämmerung."

PETERS in: M. B. d. Berl. Acad. 1871, S. 318, DOBSON Cat. of the Chiropt. of Brit. Mus., S. 133; DOBSON Report of Brit Association, 1880, S. 11, Proc. L. Z. S. 1872, S. 364; 1878, S. 8798; 1870, S. 769.

Der Grund, weshalb die vorliegenden Ex. als var. *marungensis* bezeichnet werden müssen, liegt hauptsächlich in dem Nasenblatt, welches bei *commersonii* unten gerade, bei allen 5 Ex. von var. *marungensis* unten genau wie bei *Phyll. cyclops* rundlich lanzettförmig ausgebogen ist, während das Blatt oben bei *cyclops* viel schmaler ist als unten. Das Blatt von *marungensis* sieht also so aus, wie wenn die untere Hälfte von *cyclops* mit der oberen Hälfte von *commersonii* zusammengesetzt wäre.

Diagnose von *commersonii* nach DOBSON:

Das Querblatt so breit wie das horizontale, starke Stirndrüse. Unterarm so lang wie Kopf und Körper, Schwanzende frei, oben Kopf und Nacken hellbraun, Extremitäten grau, ein mehr oder weniger breites Band von derselben Farbe fließt von der Schulter auf dem Rücken zusammen, Körperseiten grau, fast weiss bei manchen Exemplaren, unten weiss, ausgenommen ein brauner Fleck an jeder Schulter, am Bauch hat das Haar eine dunklere Färbung, manche Exemplare mit rötlichem Schimmer. Kopf und Körper 4 Zoll 1 L. Kopf 1 Zoll 6 L. Unterarm über 4 Zoll; doch schwankt die Länge, die bei einigen nur $3\frac{1}{2}$ Zoll beträgt. Gebiss wie bei *Ph. cyclops*.

Körperlänge des grössten M 95 mm, Schwanz 25, der 3 anderen M 80 - 85, des F 75, Schwanz 15 mm.

Kopf kurz, dick, Schnauze breit, Ohren weit von einander entfernt, Ohr = $\frac{2}{3}$ Kopflänge, Unterarm erheblich länger als der Körper, Schwanz mit der Spitze aus der Schenkelhaut hervorragend.

Das fein und dünn behaarte Nasenblatt ist vorn an den Seiten wie bei *commersonii* und *cyclops* von 3 übereinander liegenden Blättern eingefasst, der vordere Theil spitz hufeisenförmig, vom Rande der Oberlippe 3 mm entfernt. Der vordere Rand auf 1 mm frei über-

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hängend, in der Mitte ein Grat, der nach der mittleren Oeffnung als ein 1 mm langer Zacken hineinragt, vom oberen Rande ragen auf beiden Seiten des Grats zwei spitzige gewundene Hautlappen in die Vertiefung hinein, das obere Stirnblatt in der Mitte mit einem Grat, am oberen Rande etwas herzförmig ausgeschnitten und auf 2 mm beutelförmig an den Seiten überhängend, unter jedem überhängenden Zapfen eine Vertiefung. Der vordere wulstige Rand derselben, welcher die obere Begrenzung der Nasenöffnung bildet, in der Mitte mit einem in die Tiefe ragenden Grat, in den beiden seitwärts desselben liegenden rundlichen Lappen ebenfalls je zwei feine Oeffnungen. Die beiden mittleren gewundenen Hautlappen bilden mit diesem Grat eine herzförmige Vertiefung, die Nasengruben liegen unter dem hinteren Blatt und sind durch die herzförmige Klappe geschieden. Die Oeffnung der Glandulardrüse ein feiner, an den Rändern behaarter Spalt über dem hinteren Blatt der bei weitem nicht so deutlich sichtbar ist wie in der Abbildung von *Ph. comm.* bei DOBSON und fast ganz in den Stirnhaaren versteckt ist. Breite des Stirnlappens 9 mm, Länge 4, Länge des ganzen Hufeisens 10, Breite 8.5, Länge der mittleren Oeffnung 3 mm. Ich verweise übrigens auf die Zeichnung, welche die complicirte Bildung des Hufeisens deutlicher macht als viele Worte. Ohr an der Basis behaart, schlank zugespitzt, 13 Querfalten nach dem äusseren Rande zu; dieser wenig ausgebuchtet, der innere Rand rundlich, unten legt sich der innere und äussere behaarte Rand am, so dass unten an der Ohrbasis die Ohröffnung nur einen schmalen Spalt bildet. Tragus fehlt. 2 mm vom inneren Rande geht eine starke Sehne der Länge nach durch das Ohr bis auf 6 mm von der Spitze, von welcher der innere Ohrrand nach hinten umklappt. Ohrlänge 22 mm, grösste Breite unten 11, Länge des äusseren Basallappens 5 mm, Entfernung der Ohren 16 mm.

Alle Maasse dem grössten Exemplar entnommen. Oberlippe und Rand der Unterlippe mit warzigem Rande, in der Mitte der Oberlippe ein nackter Grat, Mundspalte 11.5 mm lang, AÜge klein, 2 mm lang, rund, 2 mm seitlich von dem die Nasenlöcher oben begrenzender Wulst. Unterarm sehr lang und stark, muskuloös, wie auch der Schenkel, Radius kräftig, 95 mm lang, Humerus 52.5. Vom Daumen a in der Flughaut eingehüllt, a = 8.5, b = 9, der stark gekrümmte braune Nagel 4.5. Finger I einer starken, oben 2 mm dicke, 75 mm lange Knochenleiste, b verkümmert, IIa = 72, b = 32.5, c = 52; IIIa = 69, b = 26, c = 16; IVa = 68, b = 26, c = 17. Die Enden der Phalangen nagelartig scharf. Tibia 39, Femur 32. Die zweigliedrige Fuss mit

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quergefalteten Ballen lang, mit starken, braunen Krallen. Sporn 8 mm. Die Flughaut reicht bis zu $\frac{2}{3}$ der Tibia. Metatarsus 6, Phalangen a= 6, b = 10, Nägel 6 mm. Penis behaart, 6 mm lang, Schwanzwirbel sehr dünn, 7 mm frei, an der Spitze 5 - 6 weisse lang Haare. Beim F misst der Radius 95, Humerus 43, die ersten Phalangen von II, IV 63, 62, 61.5; tibia 34, Femur 25. Humerus und Vorderarm innen und aussen schwach behaart, Flughaut weiss als schmaler Saum bis über das Tarsalgelenk, zwischen Humerus und Tibia schwach weisslich behaart, aussen nur am Rande der Tibia einige gelbe Haare. Flughaut zwischen Arm und Schenkel mit feinen Querfalten, die unter der Lupe aus sehr feinen warzigen Papillen bestehen, Behaarung zwischen den Schenkeln dünn gelblich graubraun mit hell Weissgrau melirt, da die Haare theils eine graubraune, theils eine weissliche Spitze haben. Bauch mehr gelblich weiss, in der Mitte mehr bräunlich. Der Oberkörper ist in der vorderen Hälfte ähnlich wie unten mit mehr oder weniger Braun, hinter variirt die Färbung. Bei dem kleinsten M ist sie ähnlich wie vorn, nur in der Mitte des Rückens und auf dem Oberarm mehr lichtbraun, Rand des Hinterkörpers an der Flughaut und hinterer Rand des Oberarms mehr weiss. Bei einem älteren M ist das Braun intensiver, gegen die Schultern hell streifig abgesetzt, ohne eigentliche Streifen zu bilden. Beim dem grössten M ist die hintere Partie oben mehr rothbraun, nach vorn heller quertstreifig abgesetzt, die Mitte des Nackens dunkler, bei einem M die vordere Partie hell graubraun, weiter nach hinten reichend, das Braun viel dunkler und nach vorn halbmondförmig abgesetzt. Allen gemeinsam aber ist die hellgelbliche Färbung des Bauches, die unter den Achseln lebhaft hellgelb wird und vorn durch einen nach hinten zugespitzten dreieckigen, dunkelumbraun streifigen Fleck begrenzt wird. Das Weissgelb der Achselgegend schneidet gegen das Braun der Oberseite scharf ab. Der weisse Fleck unten an der Schulter ist bei dem grössten M matt weissgelb, sonst besonders lebhaft an der Innerseite des Humerus. Bei dem F ist die Färbung oben vorn dunkler, das mit weisslicher Sprenkelung abgesetzte Braun zieht sich weiter nach vorn und ist vorn dunkel sepiabraun, hinten der Rand mehr gelbbraun. Die Färbung bei *var. marungensis* variirt ebenso wie bei *commersonii*. Die Längsstreifen von *vittata* PETERS fehlen. Vergl. die Abbildungen von *vittata* PETERS, Säugethiere Taf. 6, Schädel Taf. 13, 7 - 13, *commersonii* bei DOBSON Taf. 9, Fig. 2, *cyclops* Fig. 1. Ueber *gracilis* PETERS S. 35.

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Die Schädel dem von *vittata* (auch bei BRONN Taf. 53, Fig. 4) sehr ähnlich/

Zähne: I $\frac{2}{4}$ C $1^{+1}/_{1+1}$ P $2^{+2}/_{2+2}$ M $3^{+3}/_{3+3}$

Die jüngeren M haben von der Basis des Eckzahns gemessen die gleiche Sagittallänge von 32 mm, beim grössten M 33, beim F 31. Bei dem alten M fehlt der kleine Schneidezahn jederseits im Oberkiefer, den die übrigen besitzen. Beim F For. infraorb. länger, Jochbogen vorn und die hintere Schuppe schwächer, ebenso der ganze Oberkiefer und die Auftreibung der Nasenbeine, Massetergrube flacher, Eckzähne zarter und schwächer, der kleine Lückzahn niedriger.

Grösste Breite zwischen den Jochbogen bei F 17.5 mm, bei M adult 19.5, bei letzterem die grösste Breite am Hinterhaupt 15, die schmalste Stelle vorn an der Einschnürung 4.5, Breite des Oberkiefers 10, Schädelhöhe 14.5. Bullae aud, bestehen aus 2 Kapseln, die vordere bis dicht an den Condylus des Unterkiefers vorgezogen, je 3 mm breit, Hinterhauptloch 5.5 breit, 4.5 hoch, Zahnreihe von der Hinterseite des Eckzahns 10, Entfernung der C 8, Länge von C 7 mm, F 5. Der Unterkiefer besonders an der Sutura mentalis kräftig, Massetergrube tief; Eckfortsatz lang und schmal, nach hinten und aussen gerichtet, 2.5 lang; hinterer Ast schräg aufsteigend, Condylus 3 mm breit, Coracoidfortsatz dreieckig rundlich, hinten ausgebogen. Länge des Unterkiefers bis zum Condylus 24, Höhe des horizontalen Astes in der Mitte 3 mm. Zahnreihe 12.5. Entfernung hinten oben 8, unten 6 mm.

Zähne. C oben vorn und innen gerillt mit starker, übergreifender Basis, vorn und hinten an der Innenseite ein Zacken, P I ein kleiner, ganz nach oben gerückter, 0.3 hoher Stift, P II dreieckig, $\frac{1}{3}$ so hoch wie C, mit hinterem und innerem Nebenzacken, M I u. II mit den bekannten V-förmigen Höckern und niedrigem Seitenhöcker, M III V-förmig mit viel schwächeren Zacken. Im geschlossenen Zustande der Kiefer greifen die W-Höcker über die unteren M über. Die im knorpeligen Zwischenkiefer stehenden I, mit den Spitzen nach innen, sehr klein und rudimentär. Unten I klein, dreilappig mit langer Wurzel, C an der Basis mit wulstigem Rande, vorn nicht gerillt, hinten ausgekehlt, bei M ad. 6, bei F 4 mm hoch, P I = $\frac{1}{3}$ C, oben dreieckig zugespitzt, P II = $\frac{2}{3}$ C, vorn und hinten innen kleine Basalzacken, M W-förmig ohne innere Seitenzacken, M III verkleinert, Schon halb in dem aufsteigenden Aste sitzend. Sämmtliche Zähne oben und unten mit Basalwulst.

Phyllorhina rubra Noack, 1893

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20. *Phyllorhina rubra* n. sp.

"Ein Exemplar mit Schädel, M, Lugerrunjer-Fluss, 20.V.1890." E. (Schädel Taf. 18, Fig. 14 und 15.)

Diagnose: Kleine *Phyllorhina* von prächtig gelbrother, unten wenig hellerer Farbe (bright rufous), breitem, oben kurz zugespitztem Ohr, braunen Flughäuten, ohne Schwanz.

Beschreibung. *Phyllorhina rubra* weicht in der Bildung des Nasenblattes, des Ohrs, des Schwanzes und der Färbung erheblich von den bisher bekannten afrikanischen *Phyllorhina* ab und nähert sich mehr der asiatischen *Phyllorhina bicolor*, *var. fulva*, zeigt auch Anklänge an die australische *Rhinonycteris aurantia*, sie beweist also wie *Megaloglossus woermanni* für einen frühern Zusammenhang Afrikas mit Südasien und Australien.

Nasenblatt und Ohr sind am meisten denjenigen von *Phyllorhina fulva* ähnlich. Die obere Kante des Aufsatzes verläuft ziemlich gerade mit abgerundeten Ecken; an der Seite hat das Nasenblatt drei Falten, wie bei den afrikanischen *Phyllorhina*, während die Seiten bei *Phyllorhina bicolor* und ihren Varietäten glatt sind. Der vordere Theil des Nasenblattes ist viel schmaler als bei *Ph. bicolor*, an beiden Seiten und an dem die Nasenlöcher seitlich begrenzenden Wulst sitzen kleine, zipflige Anhängsel, wie bei *Rhinonycteris aurantia*, deren oberer Aufsatz aber gänzlich von dem der *Phyllorhina rubra* verschieden ist.

Die Augen stehen, wie bei *Ph. bicolor*, dicht neben der Basis des obern Aufsatzes. Das Ohr ist breit und verhältnissmässig kurz, der innere Rand convex, der äussere unterhalb der Spitze etwas eingebogen, der untere Rand mit rundem, nach oben gebogenem Antitragus vorgezogen. Aussen ist der Ohrand unbehaart, innen das Ohr am innern und obern Rande dünn roth behaart. Die rundliche ungespaltene Oberlippe ist braun-röthlich behaart, einzelne längere Haare hängen bartartig herab und besetzen die Seiten des Nasenblattes. Uebrigens ist das Gesicht dicht und zottig wie der Körper behaart. Vom Daumen p. 587

ist die letzte Phalanx frei, die braune Flughaut unbehaart, die Seitenflughaut im Fersengelenk angeheftet. Die Schwanzflughaut biegt sich nach hinten mit zwei runden, durch einen Einschnitt getrennten Lappen aus, die beiden Seiten der Flughaut sind statt des Schwanzes durch ein schmales, sehniges, sich bis zum Einschnitt erstreckendes Band getrennt, in welchem jede Spur von Schwanzwirbeln fehlt. *Phyll. bicolor* besitzt Schwanzwirbel, von denen das letzte Drittel des letzten frei ist.

Auch der Sporn von *Phyll. rubra* ist sehnig und nur im Ansatz verknöchert. Die zweigliedrigen Zehen des Fusses sind lang und schmal, die Nägel hell-hornfarben, auf der Oberseite mit röthlichem Anflug. Die Behaarung ist lang, dicht und flockig, oben gelb-roth mit bräunlichem Anflug, unten ohne denselben, also ein wenig heller. Die Haarfärbung ist gleichmässig, doch ist das Haar der Unterseite im basalen Theile etwas dunkler.

Maasse. Körper 50 mm, Ohr 15 lang und breit, Unterarm 52, III = 39 + 18 + 18 = 75, V = 34 + 14 + 10 = 58, Fuss 12, Metatarsus 2 breit; mittlere Sehne der Schwanzflughaut 12 lang, Sporn 7 mm.

Schädel. Die Stirn stark rundlich hervortretend, auch der Scheitel gerundet, zwischen beiden etwas eingesenkt; auch die Einschnürung im Profil deprimirt, Crista schwach. Die gerundeten Nasenbeine sind vorn halbkreisförmig eingebogen und hinten seitwärts rundlich aufgetrieben. Der sehr dünne Jochbogen verläuft fast horizontal. Bulla aud. klein, flach-rundlich, die vordere Partie von der hintern durch eine seichte Furche getrennt; Knochenring ziemlich gross. Von den 10 Gaumenfalten sind die vordem durch eine weite Lücke getrennt und leicht nach hinten umgebogen, die mittlern in der Mitte nach hinten eingeknickt, die hintern schmal und fast gerade, die zehnte in der Mitte nach hinten ausgebogen. Der knöcherne Gaumen endet kurz hinter M^3 mit flacher Einbuchtung. Flügelbeine schmal mit ziemlich langem, nach hinten gerichtetem Zacken.

Am Unterkiefer ist der horizontale Ast niedrig, unter der Symphyse etwas verstärkt, die vordere Partie bis M^2 gerade, die hintere etwas in die Höhe gebogen. Eckfortsatz mit stark nach aussen gerichtetem Zacken, Proc. coron. sehr niedrig mit schmaler, bis zum Condylus verlängerter Leiste, Massetergrube tief.

Maasse. Scheitellänge von der Basis der I bis zum Anfang des (defecten) Hinterhauptes 18 mm (annähernd), grösste Scheitelbreite 10, Einschnürung 3, hinten zwischen den Jochbogen 11, Schädelkapsel bis

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zur Einschnürung 11, Nasenbreite 5,5; Kieferbreite zwischen M^3 7, knöcherner Gaumen 6,5, Bulla aud. 2 mm.

Unterkiefer bis zum Condylus 12, Höhe des horizontalen Astes 1, unter dem Proc. coron. 2,5; Eckfortsatz 1,5; Zahnreihe 8 mm.

Gebiss. $I^{1-1}/_4 C^{1-1}/_{1-1} P^{2-2}/_{2-2} M^{3-3}/_{3-3}$.

I oben durch eine Lücke getrennt, nach innen gerichtet, klein, rundlich mit undeutlicher Spitze und schmalere Basis. C im basalen, nach vorn gerichteten Theile sehr stark, die wenig schräg gerichtete Spitze schlank, vorn mit Zacken, die vordere, gerade Kante gefurcht, die hintere und innere Seite concav, die Basis aussen hinten verdickt mit scharfem Zacken. P^1 sehr klein, rundlich, eng zwischen C und P^2 aussen gedrängt. $P^2 = 1/3 C$, die vordere Kante gerade, der Basalwulst hinten nach unten gezogen, die Innenseite concav, vorn mit Nebenzacken, M mit niedrigen Zacken, M^1 und ^{2} identisch, der Talon vorn hoch, abgerundet, bei M^3 der hintere W- Zacken verkümmert, der Rand des Talons gerade, hinten nicht hochgezogen.

I unten dreilappig, die mittlern schmal und mehr nach vorn gerichtet. C schlank, wenig gebogen, etwas nach rückwärts gerichtet, hinten innen mit rundem Basalzacken. $P^1 = 1/3 C$, breiter als P^2 ; letzterer = $1/2 C$, schlank, vorn innen mit rundem Basalzacken, innen stark concav. Bei M der vordere Aussenzacken so hoch wie P^2 , der hintere niedriger, M^1 und ^{2} identisch. Bei M^3 der vordere Aussenzacken etwas niedriger als bei M^1 und ^{2} , der hintere Innenzacken länger und mehr nach hinten gerichtet. Zwischen M fanden sich fein zerkaute Reste von sehr kleinen Insecten.

Phyllorrhina bicornis Heuglin, 1861

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Grisescence-albida, cervice, collo. dorsoque obscurioribus, pilis omnibus apice fusciscentibus; facie pallide murina; regione inguinale pure alba; auriculis griseis, intus helice versus violascentibus; prosthemate simplice replicato et cristam transversalem implicante; auriculis latis approximatis, margine interiore semilunare, exteriori falciforme, hic basi lobato.

Die Ohren sind um $1/3 - 1/4$ kürzer als der Kopf, 6^{mm} lang, innen grossentheils ziemlich dicht und fein behaart, der Aussenrand Störmig, an der Basis

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der Wurzel des Innenrandes sehr nahe gerückt und dort mit einem zugespitzten Ausschnitte versehen, innen mit 8 - 10 undeutlichen Querfalten. Die Nase mit drei halbkreisförmigen, aufeinander liegenden Blättern (Hufeisen) umgeben, deren zwei untere in der Oberlippengegend absetzen.

Zwischen den Nasenlöchern und dem nach vorne zu gerichteten Theile des obren Hufeisenblattes eine kleine fleischige Leiste, daneben jederseits ein kleiner hornförmiger Lappen, der auf- und rückwärts gebogen ist und wohl zum Verschliessen der Nasenlöcher dienen kann. Auf dem Nasenrücken steht ein niedriger, oben mit zwei flachen Kerben versehener Querkamm, auf dessen nach vorne gerichteter Seite vier in einer Linie liegende violette Wärrchen; dieser Querkamm ist umschlossen von einem an seiner hinteren Basis entspringenden, wenig erhabeneren ganzrandigen und oben und seitwärts umgefalteten, oben und hinten etwas behaarten *Prosthema*; und hinter letzterem endlich befindet sich ein ringförmiges, vorne mit etwas erhöhtem Rande versehenes, in der Mitte aber durchbohrtes Organ, das in eine Stirnrinne zu münden scheint.

Im Oberkiefer am beweglichen, etwas prolongirten Zwischenkiefer jederseits einen oben abgerundeten Schneidezahn. Im Gaumen 8 Querfalten. - Daumen vom ersten Gliede an frei. Der Zeigfinger besteht nur aus einer Phalange. Schenkelflughaut bis zur Zehenwurzel reichend. Schwanz mittellang mit wenig freier Spitze, das *Patagium anale* in sehr spitzem Winkel in ihn verlaufend. Zehen wie beim ganzen Genus zweigliederig.

Die Lippen sind mit kleinen violett-röthlichen Drüsen besetzt und fein behaart, der Pelz im allgemeinen graulich-weiss, alle Haare mit röthlich- bis rauch-grauen Spitzen, welche auf dem Rücken, Scheitel und Halse an Intensität der Färbung zunehmen. Gesicht hell mausgrau, Gegend um die Geschlechtstheile weiss. Ohrmuschel hell rauchgrau, im Innern ins Violette spielend. Flughäute schwärzlich. Körper 2" lang, Schwanz 11". Flugweite fast $10\frac{1}{2}$ ".

Phyllorrhina cyclops Temminck, 1853

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PHYLLORRHINA CYCLOPS.
PHYLLORRHINE CYCLOPE.

Cette espèce paraît habiter de préférence les rives

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boissées des eaux; elle vole, pendant le crépuscule, à la lisière des forêts voisines des rivières. Elle est facile à distinguer de ses congénères, par son pelage cotonneux, frisé et comme ébourifé partout. Le mâle, un peu en arrière de la feuille compliquée du nez, est muni d'un large syphon, qui s'ouvre sur le front; la femelle porte simplement le stigmate d'une ouverture, sans qu'il conduise à une bourse. La membrane interfémorale est découpée en demi cercle, et la courte queue qui s'y trouve engagée, n'a que sa pointe libre. Les oreilles sont longues, pointues et sans oreillon mobile.

Tout le pelage supérieur et inférieur est d'une même nuance, et il n'existe sous ce rapport aucune différence, qui puisse servir de moyen pour distinguer les sexes.

Le pelage supérieur est brun-noirâtre, mais le bout frisé de tous les poils est jaunâtre; en dessous il est brun-bistre, à bout des poils également jaunâtre. La feuille transversale qui prend naissance entre le pelage touffu du front, s'élève en deux lobes, séparés par une échancrure, de laquelle sort une petite feuille très-étroite; sur le bourrelet s'élève une autre petite feuille, à laquelle vient aboutir le large derme du fer-à-cheval, accompagné en dessous d'une seconde membrane. L'ouverture au front du mâle est entourée par le pelage laineux et enbourifé; la poche est également tapissée de poils onctueux. La moitié de l'antibrachium est couvert par le pelage.

Longueur du bout du museau à la pointe de la queue 3 pouces 6 lignes; hauteur des oreilles 1 pouce; queue 6 lignes; envergure 12 pouces 6 à 9 lignes; antibrachium 1 pouce 6 lignes.

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Patrie. L'on trouve cette espèce sur la rivière Boutry, côte de Guinée.

Phyllorrhina fuliginosa Temminck, 1853

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PHYLLORRHINA FULIGINOSA.
PHYLLORRHINE FULIGINEUX.

L'espèce nouvelle décrite ici sur un *sujet unique*, de sexe *féminin*, de toutes celles qui me sont connues est la moins affublée de membranes ou de feuilles nasales; les oreilles sont grandes et larges, et leur bord antérieur interne est garni de poils. La membrane interfémorale est ample et longue; la queue s'y trouve engagée totalement. Tout le système cutané est nu.

Parties supérieures du pelage d'un roux de rouille vif, mais la base de tous les poils est peinte de roux-doré; la tête, le devant du cou, toutes les parties inférieures ainsi que le ruban latéral, le long des flancs, sont d'un roux-doré vif. Pour toute membrane l'on voit seulement une feuille transversale, peu apparente, d'où se détache le petit fer-à-cheval en liséré, qui est un peu plus large vers le muffle, et qu'accompagnent deux petits plis, à peine visibles. La base postérieure, ainsi que le bord antérieur interne des grandes oreilles, sont couverts de poils. Les membranes sont nues et noires.

Longueur 3 pouces 4 lignes; dont la queue prend 1 pouce 3 lignes; envergure 11 pouces 6 lignes; antibrachium 2 pouces.

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Sur la vue d'un seul individu *féfelle*; ce qui fait qu'on ne peut indiquer, s'il y a ou non différence de coloration du pelage dans les sexes, ni s'il est certain que le *mâle* soit pourvu d'un syphon.

Patrie. La côte de Guinée.

Phyllorrhina gracilis Peters, 1852

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Tafel VII. Tafel XIII. Fig. 14. 15.

Ph. rufocorticina, subfus ex albo rufescens, patagiis umbrinis; auriculis capite quarta parte brevioribus, lobulo distincto; prosthemate simplice, concavo, margine reflexo, apertura frontali transversali; plicis accessoriis ad utrumque ferri equini latus binis; digito quarto et quinto apice bifidis; metacarpo digiti quarti brevioris quam digiti tertii; alis supra tarsum terminatis; crure capite longiore.

Longitudo tota 0,090; caudae 0,032; antibrachii 0,046; volatus 0,270.

Habitatio: Africa orientalis, Tette, 17° Lat. Austr.

Körper und Gliedmaßen schlank und zart. Die Ohren sind groß, um ein Viertel kürzer als der Kopf, entwickelt etwas breiter als lang, an ihrer äußeren Oberfläche bis auf das letzte Viertel behaart; sie ragen mit ihrem vorderen Rande bis nahe an die kleinen Augen, und sind auch an ihrer inneren Fläche bis zum Längskiel, welcher durch das zweite Fünftel des Ohr verläuft, mit Haaren besetzt; an ihrem hinteren Rande sind sie schwach ausgerandet, mit kurzer seitlich vorspringender Spitze; man kann 13 Querfalten zählen, deren mittlere sich bis zum Längskiel erstrecken; der Ohrlappen ist abgerundet und deutlich abgesetzt. Das Aug liegt der Ohrenbasis ein wenig näher als dem Schnauzenende. Das Hufeisen ist schmal, an den Seiten ein wenig ausgeschnitten, zu jeder Seite von zwei feinen Nebenfalten umgeben

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nach innen von den Nasengruben durch kleine am Ende zweizipflige Läppchen getrennt; die mittlere Abtheilung des Nasenbesatzes ist etwas breiter als lang, abgerundet dreieckig, am oberen Rande behaart; das Prosthemata ist einfach, am Rande nackt und nach oben umgeschlagen, durch keine Längsfalte mit der mittleren Abtheilung verbunden, so daß nur eine einfache breite Grube zwischen denselben erscheint. Zu beiden Seiten, gleich hinter dem Prosthemata, liegt eine drüsige Hervorragung, und nach der Mitte zu mehr zurück eine quere Oeffnung, aus der vorn ein kleines zungenförmiges Hautlappchen vorspringt, hinten ein paar längere Haarbüschel hervorkommen. Das Gesicht ist bis auf den nackten vorderen Theil der Lippen kurz aber dicht behaart. Von der Mitte der Oberlippe geht eine schmale nackte Leiste bis zum Nasenbesatz hinauf. Am Gaumen bemerkt man zehn quere Schleimhautfalten, von denen die sechs letzten gedrängter stehenden den Raum zwischen den beiden vorletzten Backzahnpaaren einnehmen. Der schlanke Körper ist doppelt so lang wie der Kopf. Die Mittelhandglieder nehmen vom zweiten bis fünften Finger progressiv an Länge ab. Die Phalangen des dritten Fingers zusammen sind kürzer als die Mittelhand desselben. Die kleinen Endglieder des dritten und vierten Fingers theilen sich in zwei Äste, auf dieselbe Weise, wie man es beim letzten Schwanzgliede der *Nycteris* beobachtet. Die Flughaut endigt am Unterschenkel, etwas oberhalb der Fußwurzel. Der Fuß ist nicht so lang wie der Unterschenkel; der Bau der Zehen und Krallen, wie bei der vorhergehenden Art. Die Schenkelflughaut ist zu jeder Seite zwischen dem Schwanz und dem Sporn flach ausgeschnitten. Der Schwanz ist mehr als ein halb Mal länger als der Unterschenkel, und ragt mit einem kurzen Ende frei aus der Schenkelflughaut hervor: sein vorletztes Glied ist das längste von allen.

Die Behaarung ist fein und lang, läßt den größten Theil des Oberarms und Schenkel frei, und geht an der Bauchseite nur ein wenig weiter als an der Rückseite auf die Flughäute über. Die Längslinien der Flughäute sind sowohl oben wie unten mit ganz kurzen Härchen versehen.

Die Farbe an der Rückenseite ist hell röthlich nelkenbraun, an der Bauchseite röthlich weiß, an der Brust mit stärkerem röthlichbraunen Aufzuge. Im Gesicht ist die braune Farbe gestätigter. Die Haare des Rückens, des Oberkopfes und der Seiten des Halses sind an der Basis und Spitze röthlich braun, am übrigen größten mittleren Theile weiß. Die Bauchhaare sind am Grundtheile braun, an der Spitze weiß. An der Brust finden sich Haare mit weißer und mit bräunlicher Spitze. Die Farbe der Flughäute ist hell umbrabraun. Die Nägel sind schmutzig weiß. Beim ausgestopften Thiere erscheint die Rückseite durch Auseinanderweichen der Haare gefleckt.

Der Bau der Eingeweide stimmt mit dem der vorigen Art ganz überein. Auch das Skelet weicht nur wenig ab. Man kann nur sechs getrennte Halswirbel unterscheiden; darauf folgt ein Knochenstück, das aus der Verwachsung des siebenten Halswirbels und der zwei obersten Rückenwirbel mit den beiden ersten Rippen und dem Manubrium sterni hervorgegangen ist, an dem man aber keine Spur der ehemaligen Trennung findet; dann kommen

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noch zehn rippentragende Wirbel, fünf verwachsene erste und zwei getrennte letzte Lendenwirbel; das Kreuzbein besteht aus drei, und der Schwanz aus zehn Wirbeln, von welchen der letzte nur $\frac{1}{2}$ Mm. lang ist, der vierte und fünfte Endwirbel dagegen am meisten gestreckt erscheinen. Die beiden ersten Rippenpaare sind, wie oben bemerkt, verwachsen. Die drei folgenden verbinden sich durch besondere Knorpel mit dem Brustbein. Die 6te, 7te und 8te Rippe stehen mit demselben durch ein

gemeinschaftliches breites Knochenstück in Verbindung, und die vier letzten sind frei. Der Schädel (Taf. XIII. Fig. 14, 15) zeigt keine so großen Knochenkämme und die Gehöröffnung ist mehr nach der Mitte gerückt; die Zähne zeigen im allgemeinen dieselbe Form und Anordnung, doch ist die Schmelzleiste des letzten oberen und unteren Backzahns etwas mehr entwickelt, und die oberen Schneidezähne sind undeutlich zweilappig.

Diese zierliche Art stammt aus dem Innern des Landes, aus Tette. Das einzige Exemplar, welches ich erlangte, ein Männchen, verlor sich, durch das Licht verlockt, in meine Wohnung. Die Nahrung besteht, wie aus dem Inhalte des Magens, Überbleibseln von Hymenopteren, hervorgeht, aus Insecten.

Mafse in Millimetern.

Länge von der Schnauze bis zur Schwanzspitze: 90
 Flugweite: 270
 Länge des Kopfes: 19
 Länge des Nasenbesatzes: 5
 Breite desselben: $4\frac{1}{2}$
 Höhe des Ohres: 14
 Breite des Ohres: $14\frac{1}{2}$
 Länge des Oberarms: 28
 Länge des Vorderarms: 46
 Länge des Daumens (Mittelh. $3\frac{1}{2}$, 1.Gl. $2\frac{1}{2}$, 2.Gl. $1\frac{1}{2}$): $7\frac{1}{2}$
 Länge des 2ten Fing. (Mittelh. $38\frac{1}{2}$, 1.Gl. $\frac{1}{2}$): 39
 Länge des 3ten Fing. (Mittelh. 36, 1.Gl. $14\frac{1}{2}$, 2.Gl. $16\frac{1}{2}$, 3.Gl. $1\frac{1}{2}$): $68\frac{1}{2}$
 Länge des 4ten Fing. (Mittelh. 35, 1.Gl. $9\frac{1}{4}$, 2.Gl. 8, 3.Gl. $\frac{1}{4}$): $52\frac{1}{2}$
 Länge des 5ten Fing. (Mittelh. 31, 1.Gl. $11\frac{1}{2}$, 2.Gl. $9\frac{1}{2}$, 3.Gl. $\frac{1}{4}$): $52\frac{1}{4}$
 Länge des Oberschenkels: 22
 Länge des Unterschenkels: 20
 Länge des ganzen Fusses: 8
 Länge der mittleren Zehen: 4
 Länge des Sporns: 11
 Länge des Schwanzes: 32
 Länge des Schädels: $17\frac{1}{2}$

Von den bisher beschriebenen Arten ist mir keine bekannt, welche mit dieser südostafrikanischen zu verwechseln wäre.

Phyllorrhina megalotis Heuglin, 1861

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Minima; auriculis rotundatis, maximis fronte connatis; membrana semi-

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orbiculare (ferrum equinum) simplice; vellere sericato, facie nitide et delicatissime flavo-ferruginea; stria parva nigricante, per oculos ducta; notae pilis apice lata chocolalina, dimidio basali cinerascante albido; gastraeo albo, pallide isabellino-induto.

Diese sehr aberrante Art gehört wegen ihrer zweigliederigen Zehen und in Folge der vollständig gleichen Bildung der Finger und Schneidezähne und theilweise auch des Nasenbesatzes zu *Phyllorrhina*, weicht jedoch hievon in einigen Stücken ebenso sehr ab, als von *Rhinolophus*: die Ohren sind nämlich sehr lang, breit und gerundet, auf der Stirn sich sehr genähert, dort durch eine niedrige Hautfalte verbunden, nach vorn stehend und ihr Aussenrand an der Basis über die des Innenrandes dem Maule zu vorgerückt; der breite Lappen des ersteren (Aussenrandes) kaum durch eine leichte Kerbe vom Rande geschieden, nach vorn tief und fast rechtwinkelig zum Grunde des Ohres eingesenkt Ohrmuschel mit ungefähr 20 feinen, aber sehr deutlich ausgesprochenen Querfalten, der Innenrand und ein ihm naheliegender und mit demselben parallel laufender Faltenrücken innen behaart. Das Hufeisen selbst besteht nur aus einer einzigen Falte, in der beckenartig vertieften Nasengrube seitwärts je ein und in der Mitte ein drittes, wohl theils zum Verschluss der Nasenlöcher dienendes Fleischlappchen, die zusammen einer Ankerform - - zu vergleichen sind; dahinter der mit vier nach vorne gerichteten Wärmchen oder Drüsen versehene ungekerbte Querkamm, der nach hinten strahlenförmig mit einem rundlichen, auf- und vorgerichteten niedrigen Hautblatt geziert ist. Die Flughautspitze erreicht kaum den Vorderfuss; die Fusswurzel ist ganz frei; das erste Daumenglied dagegen beiderseits von ihr umschlossen. Der Schwanz ganz vom *Patagium interfemorale* eingeschlossen; von der Calcaneusspitze zieht sich eine feine hornige, an den Schwanzspitzen zusammenlaufende Vorstreckung herab. An seiner Spitze einige feine Härchen; ebenso die Zehenglieder mit einzelnen kleineren borstenartigen Haaren bekleidet.

Die Zähne genauer zu analysiren, ist mir auf der Reise unmöglich, da ich das einzige Exemplar (M) nicht sichern möchte. Bezüglich der Stellung der Schneidezähne im oberen beweglichen Zwischenkiefer und was die Zahl der

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Schneide- und Eckzähne ($\frac{1}{2}$ - $\frac{1}{1}$ anbelangt, weicht diese neue Art gar nicht von *Phyllorrhina* ab. Die oberen Eckzähne nach innen und vorne und nach aussen und hinten mit einer Zahnkrone versehen, die etwas am Hauptzahn heraufläuft und oben eine ausgezackte Spitze hat. Gaumen mit 6 Falten, deren 2 erste fleischig, dick und breit und mit vielen Papillen oder Widerhaken besetzt sind.

Die Behaarung der Schnauze ist verhältnissmässig kräftig und dicht, an den übrigen Theilen des Körpers sehr fein und sammtartig. Gesicht fuchsroth ins Gelbe, welche Farbe nach hinten zu allmählich verläuft und am Kinn fast nicht ausgesprochen ist; durch die kleinen Augen ein schwärzlicher Streifen. Oberseite weisslich-grau, alle Haare mit breiter chokolade-brauner Spitze; Unterseite weisslich, namentlich seitlich fein rothgelblich überflogen; die Ohren sehr hell -, Flughaut wenig dunkler graulich. Im Innern der Ohren zwischen den Falten ja eine Reihe weisslicher, drüsenartiger Punkte.

Diese Art fliegt, wie es scheint, bei Nacht; in der Dämmerung habe ich sie nie gesehen. Sie ist äusserst behende und flink in ihrer Bewegung.

Phyllorrhina vittata Peters, 1852

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Tafel VI. Tafel XIII. Fig. 7 - 13.

Ph. cervina, *vittis quatuor dorsalibus albitis*, *cervice albide irrorata*, *gastroe canescente*, *lateribus albis*, *alis fuscis*; *auriculis ovatis*, *acuminatis*; *apertura frontali magna longitudinali*; *prosthema lato*, *foveis quatuor insigni*; *plicis accessoriis ad utrumque ferri equini latus quaternis*; *cauda crure breviori*.

Longitudo a rostro ad caudae apicem 0,150, *capitis 0,040*, *antibrachii 0,105*.

Habitatio: Africa orientalis, Insula Ibo, 12° 20' Lat. Austr.

Phyllorrhina vittata. W. PETERS, Mittheilung in der Gesellschaft naturforschender Freunde zu Berlin, am 21. August 1849.

Der Kopf dieser grossen Hufeisennase ist doppelt so lang wie breit und hoch; die Schnauze vorn stumpf und breit. Das Auge liegt in der Mitte zwischen Ohr und Schnauze; die Augenlidränder sind ringsum mit abstehenden Haaren besetzt. Die Ohren sind um ein Drittel kürzer als der Kopf, lang zugespitzt, am hintern Rande bogenförmig ausgeschnitten, und haben dreizehn Querfalten, von denen nur acht bis neun sehr hervorspringen; am vorderen Rande, am Längskiel, und am untern Drittel ihrer Aufsenfläche sind sie lang behaart; die Ohrecke ist klein und undeutlich abgegrenzt. Der Nasenbesatz ist ähnlich wie bei *Phyllorrhina (Rh.) insignis*, Horsfield; die vordere Abtheilung, das Hufeisen, ist einfach, viel breiter als lang, jederseits nach innen durch eine dreieckige Klappe von den Nasengruben getrennt, und in der Mitte vor ihnen mit einem abgerundeten Längswulst versehen: neben jeder Seite des Hufeisens erheben sich vier progressiv an Grösse abnehmende Hautfalten; die mittlere Abtheilung, deren Gestalt man mit einer Hübe vergleichen kann, welche ihr spitzes Wurzelende zwischen die aufgewulsteten inneren Nasenränder absendet, ist sehr kurz, drei Mal so breit wie lang; die hinterste Abtheilung, das Prosthema, ist so lang und breit wie die vorhergehende, hinten mit der Stirnhaut durch zwei, vorn mit der rühenförmigen Abtheilung durch fünf Falten, zwischen denen vier tiefe Gruben liegen, verbunden. Hinter dem Nasenblatt, sowohl bei dem Männchen wie bei dem Weibchen, liegt eine $3\frac{1}{2}$ Millimeter lange wulstig gerandete Spalte welche in eine geschlossene behaarte Höhle führt, und zu beiden Seiten derselben eine warzenförmige, durch Borsten ausgezeichnete Hervorragung mit zwei sehr feinen Oeffnungen, den Ausmündungen kleiner Drüsen. Die Lippen sind breit, an ihrer Aufsenfläche sparsam behaart und durch zahlreiche Drüsenhervorragungen ausgezeichnet. Die Schleimhaut der Unterlippe springt in der Mitte in Form einer dreieckigen glatten Fläche hervor, welcher eine kleinere körnige nackte Stelle der Oberlippe entspricht. Die Maulöffnung ist halbkreisförmig und reicht eben so weit nach hinten wie das Hufeisen. Die Schleimhaut des Gaumens bildet neun wulstige Querfalten. Der Körper ist stark und doppelt so lang wie der Kopf. Die Flughäute setzen sich ganz seitlich und entfernt von einander an. Der Oberarm ist um eine Hälfte länger

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als der Kopf; der Vorderarm und die Finger sind ebenfalls sehr lang, so dass die Flugweite nahe an zwei Fufs beträgt. Der Daumen ist kurz, und die Halsflughaut setzt sich nicht über sein Mittelhandglied hinaus an. Die Mittelhandglieder der folgenden Finger nehmen progressiv vom zweiten bis fünften ein wenig an Länge ab. Die Phalangen des dritten Fingers zusammen sind länger als der Mittelhandknochen desselben. Die beiden Phalangen des fünften Fingers sind um ein Geringes länger als die entsprechenden Glieder des vierten Fingers. Alle vier Finger sind mit einem kleinen knorpeligen Endgliede versehen, welches eine einfache Verlängerung derselben darstellt. Der Oberschenkel ist nur wenig kürzer als der Unterschenkel, welcher ebenso lang ist wie der Kopf. Die Flughäute gehen bis auf die Fufswurzel herab. Die Zehen sind kräftig; die erste und fünfte um ein Geringes kürzer als die mittleren; ihre Krallen sind merklich grösser als die Daumenkrallen. Der kurze Schwanz ragt mit seinen letzten beiden Gliedern aus der spitzwinklig ausgeschnittenen Schenkelflughaut hervor.

Die Behaarung ist reich, an der Bauchseite, zumal am Halse und an der Brust viel länger als am Rücken; auf den Oberarm dehnt sich die Behaarung vorn und hinten gleich weit aus; am Oberschenkel dagegen geht sie an der Rückenseite weiter herab als an der Bauchseite. Die Schenkelflughaut ist bis zur Mitte sparsam und lang behaart, die Seitenflügel zeigen dagegen nur zerstreute kurze Härchen auf den Querfalten.

Die Farbe des Rückens ist rehbraun, bei dem Weibchen ins Kästenbraune spielend, mit vier gelblichweissen Längsbinden, von denen zwei längs des Ansatzes der Seitenflügel, und zwei etwas verwischte vom Nacken herabsteigend neben der Wirbelsäule verlaufen. Oberkopf und Nacken sind mit Weiss besprenkt. Die Bauchseite ist graulich weiss, an den Seiten mit einem gelblichweissen Saum, der sich über den Ansatz der Halsflughaut nach dem Rücken herumschlägt. Das Gesicht ist schmutzig weiss. Die nackten Theile der Lippen, der Nasenbesatz und die Ohren sind von brauner Hautfarbe. Die Flügel sind dunkelbraun, die Nägel braunschwarz. Die braunen Rückenhaare sind zum Theil einfarbig, zum Theil weiss beringt, die des Oberkopfes und Nackens zum Theil mit weissen Spitzen versehen; die Haare des Bauches sind an ihrem Enddrittheile schmutzig weiss, im übrigen braun; die weissen Haare zur Seite der Brust und über dem Ansatz der Halsflughaut sind einfarbig.

Das Skelet zeichnet sich, wie das der Hufeisennasen überhaupt, durch die Stärke und Breite der Rippen aus. Der siebente Halswirbel, die beiden ersten Rückenwirbel, das Manubrium sterni und die beiden ersten Rippen sind zu einem Stücke mit einander verschmolzen, eben so sind die fünf ersten Lendenwirbel zusammengewachsen, wodurch der Brustkasten eine vogelähnliche Festigkeit erhält. Es sind 11 Rippenpaare vorhanden, von denen sich 7 mit dem Brustbein verbinden. An dem Brustbein kann man außer dem mit den ersten Rippen verwachsenen breiten Manubrium noch zwei Knochenstücke und den knorpligen Fortsatz des Processus xiphoideus unterscheiden. Die Wirbelsäule besteht aus 42 Wirbelkörpern. Diese zerfallen in 7 Halswirbel, 11 Brustwirbel, 7 Lendenwirbel, von denen der erste einen ganz kleinen zwei Millimeter langen Rippenfortsatz trägt, 4 verwachsene Kreuzbeinwirbel, und

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13 Schwanzwirbel. Die ersten Schwanzwirbel nähern sich in ihrer Gestalt noch sehr den Kreuzbeinwirbeln, wodurch es hier eben so wie bei andern Flederthieren schwierig wird, eine genaue Grenze zwischen Schwanz- und Kreuzbeinwirbeln zu ziehen. Die Länge des Schädels (Taf. XIII.) beträgt etwas weniger als ein Drittel der Wirbelsäule; die Längsgröße desselben ist außerordentlich stark und die Nähte sind so verwachsen, daß sich die Grenze der einzelnen Knochen nicht mehr bestimmen läßt. Gebiß: $3/2/3, 2^{1/1}, 1/4, 1^{1/2}, 2^{3/2}, 3/2, 3 = 30$. Die beiden oberen kleinen Schneidezähne stehen getrennt, jeder in dem vordem Ende des Zwischenkiefers seiner Seite, nach vorn gerichtet; ihre Gestalt ist abgerundet, von außen nach innen und unten schräg abgestutzt. Die vier unteren Vorderzähne stehen senkrecht und der Quere nach in zwei Reihen zwischen den Eckzähnen zusammengedrängt; ihre Krone ist dreilappig und zwar so, daß der äußere Lappen immer der kleinere ist. Die oberen Eckzähne sind sehr groß, an ihrer Außenseite doppelt gefurcht, vorn und hinten mit einem starken Absatz versehen. Die unteren Eckzähne sind kleiner, ohne Furchung und nur am hintern Rande des stark entwickelten Cingulums durch einen kleinen hakigen Absatz ausgezeichnet. Von den oberen Backzähnen ist der erste ein ganz kleiner rundlicher, nach außen gedrängter Lückenzahn; der zweite ist eckzahnförmig und hat drei Nebenzacken, einen vorderen sehr kleinen, einen hintern größeren, und einen inneren ganz oben an seiner Basis; der dritte und vierte sind an Größe und Form einander ähnlich, außer den fünf Höckern auf ihrer w-förmigen Schmelzleiste noch mit einem sechsten oben an ihrer innern Seite versehen: die Schmelzleiste des fünften und letzten, welche kaum halb so groß ist wie die vorhergehenden, ist unvollständig, V-förmig. Der erste untere Lückenzahn ist einwurzelig, an der Spitze zusammengedrückt, breit lanzettförmig schneidend; der folgende Lückenzahn ist zweiwurzelig, dreiseitig, langspitzig und in seiner Gestalt der ersten Hälfte der darauf folgenden Backzähne entsprechend; diese, der dritte und vierte haben eine nach innen offene W-förmige Schmelzfalte mit drei inneren und zwei äußeren Höckern, von denen der vordere äußere der größte ist; der fünfte untere Backzahn ist wie die vorhergehenden gestaltet, aber es mangelt ihm das letzte Viertel, so daß er nur vierhöckerig erscheint.

Das Oberarmbein trägt neben seinem äußern Condylus ein kleines besonders eingelenktes Knochenstück, an welchem die langen Streckmuskeln der Hand entspringen und welches allen Fledermäusen zuzukommen scheint. Die Ulna ist rudimentär und das Olecranon getrennt. Die Handwurzel besteht aus sieben Knochen, welche in zwei Reihen, drei in der ersten, vier in der zweiten, liegen. Die Fibula ist sehr fein aber vollständig (Taf. XIII. Fig. 13), wie auch bei allen übrigen Hufeisennasen, vorhanden. An dem Unterschenkel schließt sich die Fußwurzel an, welche eben so wie bei dem Menschen aus sieben Knochen in zwei Reihen, drei in der ersten und vier in der zweiten Reihe besteht. Darauf folgt die Reihe der fünf fast gleich langen Mittelfußknochen, und dann die Zehen, von denen jede aus einem sehr langen Basalgliede und aus dem krallentragenden gekrümmten Endgliede besteht.

Die Zunge ist mit platten hornartigen Papillen bedeckt, welche auf der Mitte ihre vordere Hälfte größer und zwei- bis dreispitzig sind. Der Oesophagus geht sogleich unter

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dem Zwerchfell in einen bohnenförmigen, 35 Millimeter langen Magen über, welcher links in einen größeren, rechts in einen kleinen Blindsack erweitert ist; der durch eine kreisförmige Schleimhautfalte abgegrenzte Pförtner liegt ganz nahe neben der Cardia. Der ganze Darm hat eine Länge von 38 Centimetern. Die Leber liegt mehr nach rechts, und besteht aus drei Hauptlappen, einem linken größeren, und zwei rechten kleineren; die Gallenblase, welche groß und von eiförmiger Gestalt ist, liegt in dem tiefen mittlern Einschnitte, wodurch der linke Lappen von dem rechten getrennt wird. Die Milz liegt schief an den linken Blindsack des Magens, ist sehr schmal, an dem untern Ende zugespitzt und 25 Millimeter lang. Das Pankreas ist vielfach gelappt, sehr ausgedehnt und dünn. Die Nieren sind bohnenförmig, ungelappt, 9 Millimeter lang, und bestehen im Innern aus drei in sehr lange Papillen endigenden Pyramiden. Die Nebennieren sind platt, abgerundet dreieckig, ihr größter Durchmesser nur $1/2$ Millimeter lang. Die Luftröhre ist unter dem Kehlkopf zu beiden Seiten blasig erweitert. Die Lunge bildet jederseits nur einen einzigen Lappen. Das Herz hat eine Länge von 14, an der Basis eine Breite von 10 Millimetern. Der Uterus des Weibchens ist zweihörnig. Die Hoden des Männchens liegen in der Bauchhöhle; die Ruthe desselben ist 12 Millimeter lang, die Öffnung der Harnröhre sehr weit, und die Eichel an ihrer untern Hälfte mit einem breiten Knorpel versehen. Sehr merkwürdig ist der Bau der Höhle vor der Stirn, in welche die Längsspalte hinter der Nase hineinführt; ihre Wandung besteht nämlich aus einer halbkugeligen Knorpelkapsel, die beim Durchschnitt vorn dicker als hinten erscheint; an ihrer concaven nach außen gekehrten Wandung münden zahlreiche Drüsen, welche besonders in der vordern Hälfte starker angehäuft und entwickelt sind; aus dem hintern Theile dagegen ragen lange vorwärts gerichtete Haare hervor, welche durch das bräunliche Drüsensecret angefeuchtet sind.

Der Inhalt des Magens bestand aus Sand und Überbleibseln von Insecten.

Die beiden einzigen Exemplare, welche ich von dieser großen Hufeisennase erhalten habe, wurden auf Ibo, einer der Querimba-Inseln, gefangen, und zwar wurde eins derselben auf einer Cocospalme erhascht, wo es sich durch Palmwein berauscht hatte.

Masse in Millimetern.

M; F

Länge von dem Ende der Schnauze bis zur Schwanzspitze: 150; 140

Flugweite: 620; 540

Länge des Kopfes: 40; 35

Länge des Nasenbesatzes: 14; 13

Breite desselben: 12; 10
 Länge des Ohrs: 26; 24
 Breite des Ohrs: 17; 15
 Länge des Oberarms: 60; 52
 Länge des Vorderarms: 105; 90

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Länge des Daumens (Mittelh. $8\frac{1}{2}$, 7. 1.Gl. $8\frac{1}{2}$, 7. 2.Gl. 6, $5\frac{3}{4}$): 23; 20
 Länge d. 2ten Fingers (Mittelh. 76, 67. 1.Gl. $2\frac{1}{2}$, 2): $78\frac{1}{2}$; 69
 Länge d. 3ten Fingers (Mittelh. 75, 65. 1.Gl. 34, 31. 2.Gl. 45, $38\frac{1}{2}$. 3.Gl. $1\frac{1}{2}$, $1\frac{1}{2}$): 156; 136
 Länge d. 4ten Fingers (Mittelh. 74, 63. 1.Gl. 27, 24. 2.Gl. $15\frac{1}{2}$, 12. 3.Gl. $1\frac{1}{2}$, 1): 118; 102
 Länge d. 5ten Fingers (Mittelh. 73, 62. 1.Gl. $27\frac{1}{2}$, $24\frac{1}{2}$. 2.Gl. $16\frac{1}{2}$, 15. 3.Gl. 1, 1): 118; $102\frac{1}{2}$
 Länge des Oberschenkels: 37; 29
 Länge des Unterschenkels: 40; $31\frac{1}{2}$
 Länge des ganzen Fusses: 27; 23
 Länge der mittlern Zehen mit dem Nagel: 16; 14
 Länge des Sporns: 18; 16
 Länge des Schwanzes: 30; 25
 Länge des Schädels: 36; 33
 Länge der ganzen Wirbelsäule: 115

Die einzige Art, welche durch ihre Größenverhältnisse mit der unsrigen übereinkommt ist A. Wagners *Ph. (Rh.) gigas* aus Benguella. Sie unterscheidet sich aber leicht durch die rufsbraune Farbe, durch nur drei falten zu jeder Seite des Hufeisens, durch den Mangel (oder die Kleinheit?) der Stirnöffnung, und durch die nicht bis zur Ferse herabreichenden Flughäute.

Pipistrellus (Pipistrellus) permixtus Aellen, 1957

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Type. - F adulte, Dar es Salam, Tanganyika, 1905 (Hürstel). Musée zoologique de Strasbourg, n° 12c. Spécimen en alcool; crâne extrait, incomplet.

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Description. - C'est une espèce très semblable à *P. nathusii* (Keys. Et Blas.), mais à oreilles plus petites, n'arrivant pas à l'extrémité du museau lorsqu'elles sont repliées en avant; le sommet de l'oreille est arrondi, son bord externe légèrement concave. Le tragus est falciforme; sa plus grande largeur est atteinte un petit peu au-dessus du niveau de la moitié de la hauteur de son bord interne. Le pouce est allongé. Le patagium, attaché à la base du 5^e orteil, englobe complètement la queue. Le pied est plutôt petit. Le lobe postcalcanéen, large, bien marqué, est du type *nathusii*. Les plis palataux présentent la disposition suivante: 1^{er} entier, droit, situé au milieu de C; 2^e entier, biarqué, au milieu de P⁴; les quatre suivants interrompus au milieu, incurvés en arrière et partant respectivement du milieu de M¹, du bord antérieur de M², du bord postérieur de M² et du milieu de M³; 7^e entier, biarqué, partant du bord postérieur de M³; cette disposition rappelle beaucoup celle figurée par KUZJAKIN pour *nathusii*.

A la face dorsale, les poils s'étendent à peine sur le cinquième proximal de l'uropatagium. La couleur primitive ne peut être indiquée, car après un séjour de plus de 50 ans dans de l'alcool, l'animal est fortement décoloré. Tout au plus, peut-on dire que les poils sont bicolores, foncés à la base, clairs au sommet.

La forme générale du crâne est assez semblable à celle de *nathusii*. Le rostre est toutefois moins déprimé et la boîte crânienne plus petite. Les processus préorbitaires sont moins développés. Comme chez *nathusii*, il n'y a pas de processus postorbitaires. La crête sagittale est bien marquée postérieurement, de même que les crêtes lambdaïdes. Les arcades zygomatiques sont fines. Le crâne est relativement plus large que chez les espèces du groupe *pipistrellus* (voir ci-dessous).

La dentition est du type *pipistrellus-nathusii*: I¹ est bicuspidé, la pointe postérieure arrive aux deux tiers de la hauteur de la pointe antérieure. I² atteint la hauteur de la pointe postérieure de I¹; une première pointe secondaire latéro-externe se voit parfaitement par-devant et arrive au tiers de la hauteur de la couronne; une deuxième pointe postéro-interne est plus basse. C a une pointe postérieure atteignant le sixième de la hauteur de la couronne.

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P² est situé sur le côté interne de la rangée dentaire et est bien développé, environ comme I²; sa pointe, qui arrive au niveau de la pointe postérieure de C, est bien visible en vue latérale. P⁴ est séparé de C et atteint environ les deux tiers de la hauteur de la couronne de C. Les molaires sont normales. Les incisives inférieures sont tricuspides et légèrement imbriquées. C est plus large que chez *nathusii*; sa longueur le long du cingulum est subégale à la hauteur de la couronne à son bord antérieur. P², dont la couronne est dirigée vers l'extérieur, mesure la moitié de la hauteur de C et les trois quarts à quatre cinquièmes de P⁴.

[Table removed - eds.]

Remarques. - Si cette nouvelle Pipistrelle ne présente aucun caractère singulier, elle ne s'en trouve pas moins différenciée par un ensemble de particularités qui ne se rencontrent chez aucune autre forme africaine.

Parmi les espèces à I¹ bicuspidé, à I² bien développé et à P² bien visible extérieurement, elle trouve sa place dans la clef suivante:

1. Taille relativement grande. Avant-bras de 32 mm et davantage. Long, du crâne de plus de 12.5 mm. Rangée dentaire supérieure (C-M³) de 4.6 à 5 mm. -> 2.

- Taille plus petite. Avant-bras de 32 mm environ ou moins. Long, du crâne de moins de 12.5 mm généralement.

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Rang. Dent. Sup. (C-M³) de 3.6 à 4.2 mm: *nanus*, *pipistrellus*, *helios*, *nanulus*, *stampfli*.

2. Lobe calcanéen présent. -> 3.

- Lobe calcanéen absent: *fuscipes*

3. Canine inférieure longue et étroite; son plus grand diamètre le long du cingulum dépasse à peine la moitié de la hauteur de la couronne à son bord antérieur. Rostre déprimé. Crâne allongé, rapport larg. Zygomatique X 100 / long, totale = 600 à 610: *nathusii*.

- Canine inférieure plus forte; son plus grand diamètre le long du cingulum est subégal à la hauteur de la couronne à son bord antérieur. Rostre moins déprimé. Crâne moins allongé, rapport larg. Zygomatique X 100 / long, totale = 666: *permixtus* sp. nov.

Il faut rechercher dans la faune paléartique et orientale des affinités plus précises avec cette nouvelle espèce.

Le synopsis de DOBSON (1878: 211) conduit à *abramus*, espèce à laquelle cet auteur rattache *nathusii* et *coromandra* comme synonymes. DOBSON signale "*abramus*" à Zanzibar (spécimen e", p. 228). *P. abramus* (Temm.) est considéré aujourd'hui comme une espèce uniquement extrême-orientale (à l'ouest, jusqu'en Birmanie, selon ELLERMAN et MORRISON-SCOTT, 1951); il est probable que l'exemplaire de Zanzibar, de DOBSON, appartient à la même espèce que le mien, de Dar es Salam.

TATE (1942) a fait une revision des formes paléarctiques et orientales du genre *Pipistrellus*. Parmi les subdivisions adop-

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tées (p. 235), la nouvelle espèce peut entrer dans les groupes *abramus*, *coromandra* et *ceylonicus*:

Rapport : ((larg. Zygomat. X 100) / long. Totale), selon les mesures de TATE:

groupe <i>pipistrellus</i>	
(qui comprend <i>nathusii</i>) . . .	De 601 à 610, moyenne 606
groupe <i>coromandra</i>	"603" 666" 633"
groupe <i>ceylonicus</i>	"664" 734" 688"
groupe <i>abramus</i>	"639" 734" 693
<i>P. permixtus</i>	666

Si l'on ne prend en considération que les groupes *coromandra* et *ceylonicus*, don't les représentants habitent les côtes de l'océan Indien, il semble, pour autant qu'un seul spécimen permette d'en juger, que c'est de *coromandra* (*sensu lato*) que *P. permixtus* est le plus proche. Parmi les formes ou espèces de ce groupe *coromandra*, on ne peut guère retenir que *aladdin* Thom. (Perse) et *coromandra* Cray (Indes) pour comparaison. Selon les mesures de TATE (1942: 291), ces formes sont toutes deux de taille inférieure à *permixtus*.

Il est certain que *P. permixtus* n'est pas un élément autochtone de la faune africaine. Ses affinités le rapprochent nettement des formes paléarctiques et orientales.

Pipistrellus abaensis J.A. Allen, 1917

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Type, No. 48979, F ad. (skin and skull), Aba, Belgian Congo, Dec. 18, 1910; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 1825.

Size of *P. nanus*, but much lighter in coloration, and with the sides of lower back broadly naked.

Upperparts light brown or pale isabella-color, the hairs nearly unicolor from base to tip; underparts pale buffy gray, the basal portion of the hairs nearly black, the tips light grayish, with a slight buffy tone. Wing membranes attached to the entire length of the basal joint of the outer toe. Ear small, pointed, about equally convex on both borders.

Type, total length (collectors'; measurements), 77 mm.; head and body, 44; tail, 33; foot 7; ear, 10. Additional measurements from skin: Forearm, 31.4; third metacarpal, 29.7; tibia, 12.4; foot, 6; ear 7.2. An adult female topotype is larger: Total length (collectors' measurements), 84; head and body, 53; tail, 31; foot, 8; ear, 12. Forearm (in skin), 32.1; third metacarpal, 30; tibia, 12.8; foot, 6.8; ear, 9. (The skull of this specimen, No. 48980, is lost.)

Skull (type), total length, 11.7; breadth of braincase, 6.2; interorbital breadth, 3.1; maxillar breadth, 4.6; upper toothrow, 4; length of mandible, 8.1; lower toothrow, 4.2. Incisors subequal in size, the inner one slightly bifid, P² exceedingly minute, inserted on extreme inner edge of toothrow.

Represented, by 3 specimens, the type and two topotypes; both of the latter lack skulls. Only two of the specimens, both females, are adult, one slightly larger than the other, but both indistinguishable in coloration. The third specimen is an immature male with the epiphyses not ankylosed but nearly adult in respect to size. It differs from the others in the pelage being nearly black above but somewhat lighter and more brownish below. All three of the specimens agree in having the sides of the lower back naked, the bare space being 5 to 6 mm. wide and 17 to 20 mm. long, extending from the base of the tail membrane anteriorly for more than half the length of the body. This feature, combined with small size, pale coloration and

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subequal upper incisors should render the present form easily recognizable. As said above, it is about the size of *P. nanus*, from which it differs strikingly in coloration, in the shape of the ears, and in the character of the upper incisors and upper premolars. It does not appear closely related to any described species.

Pipistrellus aero Heller, 1912

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Uaragess Pipistrelle

Type from summit of Mount Garguez, Mathews Range, altitude 7,000 ft., British East Africa, adult male; No. 181812, U.S. Nat. Mus.; collected by Edmund Heller, August 26, 1911; original number, 4110.

Characters. - Similar to *fuscatus* in color and in the position of the first upper premolar which is placed well inside of the tooth-row, but size of body much less, equal to that of *nanus*, which differs from it in the position of the minute first upper premolar which is in the tooth-row and can be seen from the outside. Skull larger than in *nanus*.

Coloration. - Dorsal coloration uniform vandyke-brown; underparts lighter wood-brown, the hair everywhere slate-black at base; membranes, feet, ears,, and tail black.

Measurements. - Head and body, 42 mm.; tail, 32; foot, 6; ear, 10.5; forearm, 31.5.

Skull: Condylar-incisive length, 12; zygomatic breadth, 8.5; mastoid breadth, 7; length upper tooth series to outer edge of canine, 4.8; condylar-incisive length of mandible, 9.

Two additional specimens, from the summit of Mt. Garguez are in the collection. They agree with the type in color, size, and dental characters. This species was seen only in the heavy forest on the summit of the mountain. Numbers of them were seen at dusk every evening but no other species was noted at so high an altitude.

Pipistrellus ariel Thomas, 1904

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Pipistrellus ariel, sp. n.

A pigmy species of a very pale colour.

Size very small, not exceeding that of *P. nanus*; form slender, limb-bones unusually light and delicate. Fur long, but not dense; hairs of back rather over 7 mm. in length. General colour above pale buffy, the slaty bases of the hairs showing through; below similar, but slightly paler. Membranes pale brown, without lighter edging; naked throughout, except quite close to the body. Ears rather short; inner margin strongly convex below, with very small basal lobule, slightly convex above; tip rounded off; outer margin convex, slightly above, strongly below, with a long, low, rounded antitragal lobe. Tragus rather short, broadest rather above its inner base, inner margin straight, tip rounded, outer margin evenly convex; basal lobule distinct, rounded.

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Wings to the base of the toes. Post-calcareal lobule very narrow. Tip of tail little projecting.

Skull, as compared with that of *P. nanus*, similar in size, but with a broader, flatter muzzle and smaller brain-case. Median palatal spine less developed. Base of skull between bullæ conspicuously narrower.

Incisors slender, conical, unicuspid terminally, though each has a minute basal cusplet on its cingulum behind; the outer two thirds the height of the inner. Small upper premolar unusually minute, hidden in the inner angle between the closely adpressed canine and large premolar, and lower than their cingula, so as to be quite invisible from without. Lower incisors trifid, overlapping, subequal in horizontal length, but the outer pair slightly thicker than the others. Anterior lower premolar two thirds the height of the second, small in section, the canine and large premolar almost touching each other on its inner side.

Dimensions of the type (the measurements in inverted commas taken by the collector in the flesh): -

Forearm 30 mm.

"Head and body 34"; "tail 34"; "hind foot 5"; "ear 10"; tragus on inner edge 3.1; third finger, metacarpus 27, first phalanx 9.3, second phalanx 10; fifth finger 36; tibia 12.5.

Skull: greatest length 11.3; basal length in middle line 8.4; breadth of brain-case 5.5; front of canine to back of m^3 3.7; least breadth of basioccipital between bullæ 0.6.

Hab. Eastern Egyptian desert, lat. 22° N., long. 35° E. Alt. 2000 feet.

Type. Adult female. Original number 28. Collected 12th August, 1903. Two specimens.

This most interesting little bat is widely different from any species yet described, the proportions of its upper incisors distinguishing it at once from most members of the genus. Its anterior upper premolars are so minute that they would probably be overlooked by any worker only examining spirit-specimens; but there is no recorded "*Vespertilio*" of so small a size in Egypt.

Pipistrellus crassulus Thomas, 1904

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Pipistrellus crassulus, sp. n.

A medium-sized species with disproportionally short forearms.

General build thick and heavy. Muzzle broad, swollen. Ears short, laid forward they do not nearly reach to the tip of the muzzle; inner margin straight below, convex above; tip evenly and broadly rounded; outer margin straight above, slightly convex below; basal lobe small, rounded. Tragus of medium length, its greatest breadth opposite its

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inner base; inner margin straight, tip rounded, outer margin gently convex, ending below in a small basal lobule. Thumbs short, with thickened but not enlarged basal pad. Wings from the base of the toes. Calcars about equal in length to the free border of the uropatagium; postcalcareal lobules distinct but narrow. Tail involved in membrane almost to the tip. Penis very long, slender.

Fur 3.5 - 4.0 mm. long on back. Uniformly dusky brown above, scarcely paler below. Membranes blackish brown throughout, without any trace of white margins.

Skull broad, stout and flattened, conspicuously broader and heavier, especially anteriorly, than in *P. pipistrellus*, which has a much longer forearm. Upper profile straight, the frontal region not inflated.

Inner upper incisors very thick, bifid; the postero-external cusp nearly as long as the main one; outer incisor slender, unicuspid, reaching about halfway from the cingulum to the tip of the inner tooth. Small upper premolar in the inner angle between the canine and large premolar, which touch one another outside it; not visible from without. Lower incisors broad, bifid. First lower premolar about three fourths the height of the second.

Dimensions of the type (measured in spirit): -

Forearm 28 mm.

Head and body 47; tail 27; ear 10; tragus on inner edge 3.5; thumb, free of membrane (c. u.) 4; third finger, metacarpus 26, 1st phalanx 9, 2nd phalanx 8.8; fifth finger 37; lower leg 12; hind foot, from back of calcar (c. u.) 7; penis 11.

Skull: greatest length 12.7; mastoid breadth 7.7.

Hab. Efulen, Cameroons.

Type. Adult male. Collected by G. L. Bates. One specimen.

This bat, with the short forearm of such pigmy species as *Pipistrellus Stampflii* and *minusculus*, has a very much larger body and head. The breadth and flatness of the skull are particularly noticeable.

Pipistrellus culex Thomas, 1911

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Pipistrellus culex, sp. n.

F. 100. Kabir. 2700'. B.M. no. 11.3.24.4. Collected 15th May, 1910. *Type*.

A very small pale species. Inner upper incisors practically unicuspidate.

Size rather greater than in *P. stampflii* and *minusculus*. Fur about 4.5 mm. in length on the back. General colour above near "wood-brown," below rather paler, the hairs both above and below blackish slaty basally. Ears with a well-marked squarish lobe at the base of their outer margin; tragus without basal lobule, broadest opposite the middle of its straight inner margin. Wings from the base of the outer toe. Calcar with a narrow calcareal lobule.

Skull very delicate, low and narrow; the brain-case narrower than in *P. stampflii*.

Upper incisors slender, unicuspid, the outer nearly as long as the inner. (If a secondary cusp has been present on the inner tooth it must have been very small and close to the tip of the tooth.) Small premolar half internal, visible from without. Lower incisors tricuspid, not overlapping.

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Dimensions of the type (the starred measurements taken in the flesh): -

Forearm 28.5 mm.

Head and body *39; tail *27; ear *9; tragus on inner edge 2.6; third finger, metacarpus 27, first phalanx 9; lower leg and hind foot (c. u.) 14.5.

Skull: greatest length 11.1; basi-sinual length 8.6; interorbital breadth 3.3; breadth of brain-case 5.9; palatosinual length 4.2; front of canine to back of m^3 3.6; front of p^4 to back of m^2 2.2; three lower molars 2.5.

Hab. and *Type* as above.

This little bat would seem to be readily distinguishable from the other small species of the genus by its pale colour and unicuspid upper incisors.

Pipistrellus deserti Thomas, 1902

p. 4

71. M. Mursuk. 30/5/1.

A small buff-coloured desert ally of *P. kuhli*, with a particularly small skull.

Size smaller than *P. kuhli*, but the forearm-length not so much less than in that form as to be in proportion with the much smaller skull. General structure, of ears, wings, and dentition, as in *P. kuhli*. Ears and tragus pale transparent buffy, little darker than the general colour. Wings dark brown, the usual white edging very conspicuous. Interfemoral paler brown, white posteriorly.

Colour of fur pale buffy, between cream and pinkish buff of Ridgway, strikingly different from the colour in ordinary *kuhli*. The hidden bases of the hairs dull slaty. Belly-hairs blackish slaty basally, whitish buff terminally. Skull very small and delicate; the total length, the breadth across brain-case, and the length of the tooth-series, all conspicuously, less than in *P. kuhli*, whether from Europe, Morocco, Tunis, or Egypt.

Dimensions of the type: -

Forearm 29.5 mm.

Head and body (measured in flesh) 43; tail (do.) 33; ear (do.) 10; third finger, metacarpal 29, first phalanx 10, second phalanx 8.5; lower leg and hind foot (c.u.) 22.

Skull - greatest length 11.6, median length above 10, median length below 9; interorbital breadth 4.1; intertemporal breadth 3.1; breadth of brain-case 62; front of canine to back of m^1 4.3.

Type. Adult male. Original number 71.

Although with the general characters of *P. kuhli*, I do not feel justified in calling this Bat only a subspecies of that animal, for other North-African bats of this group, while tending towards *P. deserti* in colour, show no approach to its conspicuous reduction in size of skull. Examples of *P. kuhli* from Morocco (*Dodson*), Tunis (*Anderson*), and Egypt (*Anderson*), all have skulls of the full normal size.

Two names might have been thought to refer to it. Cretzschmar's *V. marginatus* from Nubia is paler in colour than usual, but Dr. Andersen's specimens show that the form from there is of the usual size.

Pipistrella minuta Loche, on the other hand, is so far smaller as either to be a totally different form, or, more probably, the young of some indeterminable species. Its locality is in the Algerian range of *P. kuhli*.

Pipistrellus eisenrauti Hill, 1968

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Holotype: M. Eisenraut No. 498 (skin and skull, the right bulla and right first and second lower molars missing). Adult M. Camp V, Dikume-Balue, Rumpi Highlands. 18th. February 1967. Collected by Professor Martin Eisenraut and now in the collection of the Museum Alexander Koenig, Bonn, No. 68.5.

Other material: M. Eisenraut No. 198 (skin and skull). Adult F. Camp II, Mount Kupe, c. 1,100 metres. 30th. November 1966. B.M. 67.2129 (originally M. Eisenraut 643) (skin and skull). Old F. Buea, Mount Cameroon. 15th. March 1967. Both collected by Professor Martin Eisenraut: the first in the collection of the Museum Alexander Koenig, Bonn, the second presented to the British Museum (Natural History).

Diagnosis: A large pipistrelle (forearm length c. 33.5 - 35mm.) characterised by high braincase with elevated frontal region which is almost as high as the occiput; expanded supraorbital region with slight but clearly defined supraorbital ridges terminating in an angular junction with the orbital margin, the upper surface of the rostrum thus more or less pentagonal; high, massive rostrum; absence of basioccipital pits and by long, narrow upper incisors which are bicuspid and blade-like.

Description: Dorsal pelage dark brown, the hairs unicolorous: ventral pelage slightly paler, the hairs bicolored, basally blackish brown, tipped with dark brown. The specimen from Buea is slightly paler above and below than the other two and ventrally is a little tinged with buff. Ears of moderate size, rounded at

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the tip, margin smoothly convex anteriorly with a small basal lobe, posteriorly nearly straight in distal part, proximal part convex with small antitragal lobe. Tragus a little more than one third the length of the ear, its tip rounded, its greatest width at about one half its height; margin nearly straight anteriorly with slight basal concavity, posteriorly slightly convex for most of its length but nearly straight at the base. Wing inserted at base of first digit: a small post-calcaneal lobe or epiblemma.

Braincase broad, inflated, frontal region elevated almost to the occipital height; slight sagittal and lambdoid crests. Interorbital region wide; rostrum short, deep and massive, its upper surface nearly pentagonal in outline; supraorbital region inflated and swollen with a shallow median depression, the supraorbital ridges slight but clearly defined, terminating in an obtusely angular junction with the orbital margin which is nearly vertical anteriorly. Narial emargination short, narrow, rounded posteriorly, extending for approximately one half of the distance from the tip of the rostrum to the anterior orbital margin. Palate short, wide, with a narrow, U-shaped anterior emargination extending posteriorly almost to a line joining the posterior faces of the canines; broad, ligulate post-palatal spicule. Basioccipital pits lacking: width of bullae equal approximately to one and one half times their distance apart.

Inner upper incisor (i^2) long; narrow, blade-like, its longitudinal diameter approximately twice its transverse diameter, so orientated that posteriorly it is directed slightly inwards from the toothrow. It is strongly bicuspid, with a large anterior cusp and an equally massive but slightly lower posterior cusp: there is a low, undeveloped posterior cingulum cusp. Outer upper incisor (i^3) short, wide, its longitudinal diameter little more than one half its transverse diameter, lying laterally and slightly posteriorly to i^2 with its longer axis nearly transverse to the toothrow. Its principal cusp extends almost to the tip of the posterior cusp of i^2 : it is hollowed posteriorly and flanked by low lateral cingulum cusps. Canines slender and delicate but relatively massive at the base, with a low posterior cingulum cusp: the upper canine almost in contact with the posterior upper premolar (pm^4). Anterior upper premolar (pm^2) small, with well-developed cingulum and conical cusp, just visible externally, recessed into the angle between the posterior face of the canine and the anterior face of pm^4 . Lower incisors tricuspid and slightly imbricated: anterior lower premolar (pm_2) reduced, its height and basal area little more than one half that of the posterior lower premolar (pm_4).

Remarks: Apart from any other features, the large size and especially the large skull of *P. eisenrauti* readily distinguish it from many of the African species of *Pipistrellus* with bicuspid i^2 such as *nanus*, *nanulus*, *musculus* and *pipistrellus*. Its narrow tragus, high braincase and inflated supraorbital region disassociate it from *savii*: no specimens of *permixtus* Aellen, 1957 or *inexpectatus* Aellen, 1959 are available, but Dr. H.-J. Kuhn of Frankfurt/M. has kindly loaned photographs of the skulls of the type specimens so that comparison could be made. *Pipistrellus eisenrauti* is larger than either of these and has a higher braincase which is more elevated frontally while neither *permixtus* nor *inexpectatus* has the supraorbital region so expanded that the upper surface of the rostrum assumes a pentagonal outline. *Pipistrellus crassulus* approaches *eisenrauti* in size and also displays some lateral expansion of the supraorbital region: however, in *crassulus* the braincase is low and the frontal not elevated, the rostrum lower and less massive than in *eisenrauti* and shallow basioccipital pits are present.

Pipistrellus eisenrauti bellieri De Vree, 1972

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TYPE - KMMA (Tervuren) RG 35686, skin and skull, adult F. Adiopodoumé (5° 19-21' N - 4° 7-8' W), alt. 0 - 30 m, 19th November 1969. Collected by the author, but prepared and labelled in the Laboratoire d'Ecologie (L. BELLIER) of the ORSTROM, original number AX 0104.

DIAGNOSIS - Differing from *Pipistrellus eisenrauti* in its generally smaller size; hairs of the pelage unicolorous throughout their length and shorter; skull with less inflated frontal region, but with characteristic rostrum, with upper surface nearly pentagonal in outline.

DESCRIPTION - The colour of the dorsal pelage is dark brown; the ventral pelage is paler. The hairs, both above and below, are unicolorous throughout their length. The wing membranes are blackish-brown and there is no white hind-margin. The wing is attached to

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the base of the first toe. The interfemoral membrane is dark brown; a small postcalcaneal lobe is present. The ears are blackish-brown; the tip is rounded off, its external border straight in the upper part and convex in the lower part. There is a small semicircular antitragal lobe. The tragus has its greatest width at about one half the height of its internal border. The tip is rounded; the inner margin is straight, the outer convex in the upper part, nearly straight at the base and provided with a small basal lobule.

The skull is proportionately large. The frontal region is inflated and elevated, but not to the same extent as in *P. eisenrauti* from Cameroun. There is a slight sagittal crest and the lamboidal crests are clearly developed. The rostrum is short and wide; its upper surface is more or less pentagonal in outline. The supraorbital region is swollen, but there are no pronounced median or lateral depressions in the rostrum. The supraorbital ridges are very slight. The palate is longer than it is wide. There is a relatively broad and U-shaped anterior palatal emargination and a broad post-palatal spicule. The width of the bullae is equal to 1.4 times their distance apart.

The right upper incisors are broken and the whole dentition is unfortunately very worn. The left inner upper incisor is long, narrow and bicuspid, but it is impossible to determine the relative height of the cusps; there is a low posterior cingulum cusp. The outer upper incisor is short and wide, extending well beyond the cingulum of the inner upper incisor; its principal cusp is completely worn; there is a well developed posterior cingulum cusp. The upper canine is not in contact with the posterior upper premolar. The anterior upper premolar is small, lying in the toothrow and visible from the outside; it has a well developed cingulum and a conical cusp. The lower incisors are tricuspid. The anterior lower premolar is a little more than half the height of the posterior lower premolar. The lower canine has a low antero-lingual cingulum cusp.

REMARKS - This single specimen from Ivory Coast was carefully compared with the types or other specimens of all previously described African species of *Pipistrellus*, present in the British Museum (London), the Museum Alexander Koenig (Bonn), the Museum d'Histoire Naturelle (Genève) and the Koninklijke Museum voor Midden-Afrika (Tervuren). It is obviously a distinct west African representative of *P. eisenrauti* of Cameroun. Its distinguishing features are well marked, but in view of the poor knowledge of the variability

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of the characters on which the species are founded, such as size, colour, form of tragus and tooth proportions, it is thought best to give the Ivory Coast form subspecific status at present. It may be stressed that a critical revision of the species of *Pipistrellus* is highly desirable.

P. e. bellieri resembles *P. e. eisenrauti* HILL, but is smaller, has unicoloured and not bicoloured ventral pelage, a less inflated frontal region and a slightly smaller braincase. The hairs of *eisenrauti* are about 2 mm longer than in *bellieri*. It is of interest to remember in this connection that *eisenrauti* is a montane form, the collecting localities lying over 1000 m, while *bellieri* is a lowland form, the type locality lying at sealevel. The difference in size and in the length of the hairs is in all likelihood due to this difference in altitude.

Some points of comparison may be noted with other species possessing a bicuspid inner upper incisor, with which this new pipistrelle might be confused. *P. crassulus*, which is of about the same size and has also an unicoloured ventral pelage, is distinguishable by its lower braincase, less inflated frontal region and its dorsal profile being nearly straight. *P. crassulus* has the supraorbital region also somewhat expanded, but the rostrum is clearly smaller and lower than in *P. eisenrauti bellieri* (see table 1).

P. inexpectatus is of about the same external size, but has a bicoloured pelage; its skull is relatively smaller, the rostrum lower and the supraorbital region much narrower than in *P. e. bellieri* (see table 1). Moreover in *inexpectatus* there is a pronounced depression on each side of the rostrum, a character which it shares with *P. anchietae*.

P. e. bellieri is equal to or slightly larger than *P. nanus*, *P. nanulus* and *P. musciculus*, but its much larger skull, massive rostrum and expanded supraorbital region immediately distinguishes it from either of these species.

Pipistrellus fouriei Thomas, 1926

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F. 1390 (very old). Ukualukasi, N.W. Ovamboland. 3900'. 7 October, 1924. B.M. No. 25.12.4.20. Type.

A very minute species, pale with bicolor fur.

Size very small, the skull as small as in *P. ariel* and *culex*, although the forearm is not so short as in *nanulus*, *pusillulus*, and some others with larger skulls. Build light and delicate, not as in some of the minute Congo species. General appearance very like that of the East African *P. helios* Heller, the hairs of the upper surface similarly drab, the hairs dark slaty at base, then

drabby whitish, darker drab terminally. Under surface paler drab, the hairs blackish slaty for their basal three-fourths, those along sides and on the base of the interfemoral drabby whitish to their roots. Ears rather narrow, inner margin slightly convex, upper half of outer margin straight or even concave. Tragus of medium size, rather broader terminally, but by no-means as in the Noctule, as Peters says is the case in his *pusillulus*. Wing membranes greyish brown, with white edges; interfemoral with whitish transverse lines.

Skull low, light and delicately built, the brain-case flatter than in *ariel* and *helios*, more as in *culex*. Upper incisors apparently about as in *nanus*, but being much worn in the single specimen, their exact shape cannot be described, the outer incisor about three-fourths, the height of the inner. Small premolar well developed, visible from outside, the canine and large premolar not touching each other outside it.

Dimensions of the type:-

Forearm 29 mm.

Head and body 38 mm.; tail 30. Third finger, metacarpal 27, first phalanx 9; lower leg and hind foot 16.

Skull: greatest length 11 mm.; condylo-basal length 10.1; breadth of brain-case 5.8; palato-sinual length 4; breadth across molars 4; combined length of large premolar and first two molars 2.3; front of canine to back of m^3 3.5.

Hab. And Type as above.

The pale colour and small skull of this tiny Bat will readily distinguish it from any described species. It has, indeed, a smaller skull than any known species except *P. ariel* and *culex*. Bocage's *P. bicolor* from Angola is a dark-coloured form, while the skull of Peter's *pusillulus*, as figured in his plates of Chiroptera, is decidedly larger, almost equalling that of *P. nanus*.

Tome's *P. rusticus*, obtained by Andersson in this same region, also has a very much larger skull. The two co-types of it are in the British Museum, received in the Tomes collection, and of these No. 7.1.1.419, an old female, may be now selected as its lectotype.

Named in honour of Dr. Fourie, of Windhuk, who has rendered assistance to Capt. Shortridge in various ways, an assistance which has been the more effective because of his own very considerable knowledge of the Mammalogy of South-West Africa.

Pipistrellus fuscipes Thomas, 1913

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Near *P. rueppelli* and *pulcher*, but outer incisors larger.

General characters, including the striking contrast between the white or buffy underside and the greyish upperside, as in *P. rueppelli* and *pulcher*, but the membranes rather paler, while the forearms, hind legs, feet, and tail are darker, apparently black, so as to form a marked contrast with the pale

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membranes. Feet large in proportion. Calcar long, without post-calcareal lobule. Wings to the base of the fifth toe.

Skull rounder and more swollen than in *P. rueppelli*, the frontal region broader and more convex, and the brain-case more inflated. Supra-orbital edges more rounded, less ridged. Bony palate slightly shorter posteriorly.

Inner upper incisors not so long as in *P. rueppelli*, bicuspid, the secondary cusp well developed. Outer incisor much larger than in *rueppelli*, its longer cusp falling not far short of the outer cusp of the inner incisor, its base with two small secondary cusps, postero-internal and postero-external. Small upper premolar well developed, nearly half the height of the large premolar, quite visible from without, in the centre of the fairly large space between the canine and large premolar.

Dimensions of the type (the italicised measurements taken in the flesh): -

Forearm 36 mm.

Head and body 49; tail 41; ear 12; third finger, metacarpal 34.5, first phalanx 12.6, second phalanx 11; tibia 14; hind foot (c. u.) 10.

Skull: greatest length 13.5; basi-sinual length 9.8; front of canine to back of m^3 4.8.

Hab. Uganda. Type from 60 miles W. of Entebbe. Alt. 3700'.

Type. Adult male. B.M. no. 6.7.1.5. Original number 1. Collected during the Ruwenzori Expedition by R. E. Dent. Several specimens.

In both *P. rueppelli* and *P. pulcher* the outer incisor is minute, and in the type of the latter species, which in other respects seems nearest to *P. fuscipes*, there is no indication of the characteristic contrasted coloration of the limbs and membranes.

Pipistrellus helios Heller, 1912

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Samburr Pipistrelle

Type from Merelle Water, 30 miles south of Mt. Marsabit, British East Africa; adult male; No. 181813, U.S. Nat. Mus.; collected by Edmund Heller, July 22, 1911; original number, 3065.

Characters. - Related most closely to *nanus* but size much less and coloration much paler; size of *culex*, but inner upper incisors broad

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and bifid at tip; from *ariel* it differs in bifid inner upper incisor and large size of first upper premolar

Coloration. - Dorsal haired area ochraceous-buff; the hair at extreme base deep black; underparts lighter, buff in color, the hair deep black on the basal two-thirds of its length; ears naked, clay color; membranes blackish, narrowly edged by white.

Measurements. - Head and body, 45 mm.; tail, 31; hind foot, 5; ear, 9; forearm, 27.5.

Skull: Greatest length, 11; condylo-incisive length, 10.6; basilar length, 8.3; zygomatic breadth, 7; interorbital constriction, 3.2; palatilar length (nasal notch), 4.1; upper cheek teeth, 2.9; condylo-incisive length of mandible, 7.7; coronoid-angular depth of mandible, 2.8.

Three specimens of this diminutive Pipistrelle are in the collection. All were collected in the Northern Guaso Nyiro watershed.

Pipistrellus inexpectatus Aellen, 1959

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Type. - M adulte, en alcool. Ngaouyanga, vallée de la haute Bénoué, Cameroun, 11.7.1947; coll. A. Monard et V. Aellen, n° orig. 954. Musée d'Histoire naturelle de La Chaux-de-Fonds.

Cette espèce a été confondue jusqu'à présent avec *Pipistrellus marginatus* (Cretzschmar). La description originale de *Vespertilio marginatus* Cretzschmar (1826) est insuffisante pour une identification sûre; aucun caractère et aucune mesure du crâne ne sont donnés; la dentition, en particulier, n'est pas décrite. TEMMINCK (1837), qui donne une nouvelle description de *V. marginatus*, rapporte à celui-ci des spécimens de Tripoli et de Sardaigne; il précise que la première incisive est "longue et droite" et la seconde "accollée à la canine, très petite".

V. marginatus a été couramment mis en synonymie de *Pipistrellus kuhli*, avec des raisons plus ou moins valables. L'assertion la plus probante est fournie par MERTENS (1925), qui a revu le type de CRETZSCHMAR déposé au Senckenberg-Museum et qui le rapporte à *P. k. kuhli*.

Cependant, G. M. ALLEN (1914) avait réhabilité l'espèce de CRETZSCHMAR en lui identifiant un spécimen F obtenu à El Garef,

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près de Roseires (Soudan). L'auteur précise que sa Pipistrelle possède une première incisive supérieure (I^1) fortement bicuspidée et mesurant environ deux fois la hauteur de la seconde incisive. Comme chez *P. kuhli*, I^1 est unicuspidée et I^2 plus petit que la moitié de I^1 , il ne pouvait s'agir de cette espèce.

Lors de la Mission scientifique suisse au Cameroun, j'ai récolté un spécimen que je rapportais à *P. marginatus*, en acceptant la réhabilitation de ALLEN (cf. AELLEN, 1952). Grâce à l'amabilité du Dr H. Felten, j'ai obtenu les précisions désirables concernant le type de *Vespertilio marginatus*: le crâne étant resté dans la peau et vu l'état vétusté de celle-ci, il serait risqué de l'extraire. Par contre, le Senckenberg-Museum (Frankfurt) possède un autre spécimen, récolté par RÜPPELL au même endroit, la même année, donc un topotype: I^1 est unicuspidée et I^2 est plus petit que la moitié de I^1 . Il ne fait alors plus de doute pour moi que *Vespertilio marginatus* Cretzschmar (1826) = *Pipistrellus kuhli* (Kuhl, 1819).

La Pipistrelle trouvée dans le N du Cameroun ne peut être assimilée à aucune espèce africaine; il convenait donc de lui donner un nouveau nom.

Diagnose. - *Pipistrellus* de taille moyenne (avant-bras 31 mm, long, du crâne 13 mm). I^1 bicuspidée. I^2 environ de la moitié de la hauteur de I^1 . P^2 très petit, non visible du côté externe; C et P^4 sont en contact. I_{1-3} tricuspidées. Tragus ayant sa plus grande largeur au niveau du tiers inférieur de son bord interne; lobule de la base du bord externe présent. Oreilles de longueur moyenne (11,5 mm) à sommet largement arrondi. Un liséré blanc très net au patagium, de la pointe de la queue à celle du troisième doigt.

Description. - Je renvoie à la description détaillée que j'ai donnée, sous le nom de *Pipistrellus marginatus* (Cretzschm.) (AELLEN, 1952: 80-81).

Remarques. - Cette nouvelle espèce ne semble avoir comme plus proche parent que *Pipistrellus anchietae* (Seabra), d'Angola. Toutefois, cette dernière s'en distingue par la couleur,

par des oreilles plus petites et pointues, par les longueurs relatives des quatrième et cinquième métacarpes (cf. HILL et p. 228

CARTER, 1941). Ces différences sont peut-être seulement d'ordre subsppécifique; mais il faudra attendre une revision générale des *Pipistrellus* africaines pour tenter de tels regroupements de formes.

Le spécimen du Soudan, attribué par G. M. ALLEN à *Pipistrellus marginatus*, semble bien devoir être assimilé à *P. inexpectatus*: le patagium possède le liseré blanc, la dentition correspond parfaitement; seul le crâne est plus petit (voir mesures ci-dessous).

Parmi les espèces africaines à I¹ bicuspidé, à I² bien développé et à P² non visible extérieurement, la nouvelle espèce trouve sa place dans la clé suivante:

1. Taille très petite: avant-bras de 30 mm ou moins, long, du crâne inférieure à 11,5 mm
musculus Thom., ? *pagenstecheri* (Noack)

- Taille plus grande: avant-bras généralement de plus de 30 mm, long, du crâne 11,5 mm ou plus. --> 2

2. I₁₋₃ bicuspidés. *crassulus* Thom.

- I₁₋₃ tricuspidés. --> 3

3. Tragus ayant sa plus grande largeur au niveau du milieu de son bord interne. *savii* (Bonap.)

- Tragus ayant sa plus grande largeur au niveau du tiers inférieur de son bord interne. --> 4

4. Oreilles petites et pointues. Pas de liseré blanc au patagium. *anchietae* (Seabra), *rusticus* (Tomes)

- Oreilles plus grandes et largement arrondies. Un liseré blanc très net au patagium. *inexpectatus* sp. nov.

Je n'indique ci-dessous que les principales mesures. Pour plus de détails, je renvoie à ma description de 1952.

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Pipistrellus inexpectatus (Cameroun M Type; Soudan F cf. Allen, 1914)

Avant-bras: 31; 30

3e métacarpe: 30,8; -

4e métacarpe: 28,5; -

5e métacarpe: 29; -

Oreille (longueur): 11,5; -

Tragus (bord interne): 3,3; -

Crâne, longueur totale: 13; 11,8

Largeur zygomatique: 8,6; -

" M³-M³: 5,4

Longueur rangée dentaire C-M³: 4,5; 4

Pipistrellus Kuhlii fuscatus Thomas, 1901

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Similar in all essential respects, in size, proportions, shape of ears and tragus, and in dentition, to the true *P. Kuhlii*, but distinguished by its uniform dark colour. The body is dark smoky brown above and scarcely lighter below, and the ears and volar membranes are uniformly dark slaty grey, without trace of the white edging so characteristic of *P. Kuhlii*.

Dimensions of the type (an adult male in spirit): -

Forearm 35 millim.

Head and body 45; tail 34; ear 12.5; tragus on inner edge 4; third finger, metacarpal 32, first phalanx 12, second phalanx 10; lower leg 14.5.

Hab. Naivasha, British East Africa.

Type. Male. Collected 19th October, 1900, and presented by Sir H. H. Johnston

Pipistrellus marrensis Thomas and Hinton, 1923

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M. 633, 653, 656. Foot-hills of S. Jebel Marra. 4000'.

Essentially as in *P. deserti* Thos., but of smaller size and darker colour.

Colour comparatively dark and rich, much like that of Egyptian specimens of *P. kuhli*, the general hue of the upper parts being near "Dresden brown." of Ridgway. Ears noticeably darker than back. Wings dark brown, with usual whitish edgings; interfemoral paler. Tragus shorter and broader than in *deserti*, with

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broadly rounded tip and parallel borders; the inner border the longer. Forearm not exceeding 28 mm. (29.8 in *deserti*).

Apart from its slightly smaller size, the skull agrees with that of *P. deserti*. The outer upper incisor and the small upper premolar p^2 appear to be a little more reduced than in *deserti*, but the available material shows a rather wide range of variation in these respects. In the type-skull i^2 is both absolutely and relatively much smaller than in the only known skull of *deserti*; and the point of the tooth does not rise above the cingulum of i^1 . The small premolar is also greatly reduced, and so crowded between the canine and p^4 that it is not easy to detect. But in the fragments, all that is left of the two other skulls from Jebel Marra, the teeth in question, though rather smaller than in *deserti* are considerably larger than in the type.

Dimensions of the type: -

Head and body 37 mm.; tail 21; hind foot 5.5; ear 12. Forearm 26.5; third finger 46 (m.c. 24.5; phalanges 8.7 - 7.5 - 5.5); lower leg and hind foot (c.u.) 14.5.

Skull: greatest length 11.2; median length above 9.6; median length below 8.1; interorbital breadth 4.3; intertemporal breadth 3.4; breadth of brain-case 6.7; canine to m^3 3.8.

Hab. Foot-hills, S. Jebel Marra; altitude 4000'.

Type. Adult male. B.M, No. 23.1.1.15. Original No. 633. Collected April 3, 1921.

This interesting little bat is, no doubt, closely related to *P. deserti*, described from Tripoli. Its smaller size, darker colour, shorter tragus, and possibly more reduced dentition seem to warrant its receiving distinct specific rather than subspecific rank. The individual variation noticed in the dentition is of some interest in a group in which, normally, even minute dental characters are surprisingly constant.

Both *marrensis* and *deserti* are apparently closely allied to *P. kuhli*; and we can see no good reason for placing them in the genus *Scotozous* (cf. Miller, Fam. & Gen. Bats, p. 206, 1907).

Pipistrellus minusculus Miller, 1900

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Type. - Adult female (in alcohol no. 84500 U. S. National Museum. Mount Coffee, Liberia, May, 1894. O. F. Cook. collector

Characters. - Similar to *Pipistrellus stampflii* (Jentink), but smaller, the tail relatively longer and color "more reddish brown."

Ears. - The ears are rather short; laid forward they extend to extremity of muzzle. Anterior margin of conch abruptly convex at base, then slightly convex to narrowly rounded tip. Posterior border concave to middle, then somewhat more strongly convex to deep notch separating antitragus from rest of ear. Antitragus small but sharply defined, subquadrate in outline, its height to its width. Inner surface of conch with five or six ill-defined cross ridges.

Tragus blunt, slightly curved forward, broader above than at base.

Membranes. - The membranes are very thin and delicate. Wings from base of toes. Free border of uropatagium longer than calcar.

Feet. - The feet are short, scarcely one-half as long as tibiae. Calcar nearly double as long as foot, terminating in a distinct lobe, and noticeably keeled posteriorly.

Fur. - Fur short, that on middle of back about 5 mm. in length. It is closely confined to body, barely extending on membranes except as a thin sprinkling of scattered hairs.

Color. - Dorsal surface raw umber, slightly more yellowish posteriorly than anteriorly. Ventral surface yellowish wood-brown, in pubic region tinged with gray. The hairs are everywhere slaty black through basal half or more. Ears and membranes blackish brown,

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the wings and uropatagium with a very narrow pale border along free edge.

Skull. - The skull of *Pipistrellus minusculus* (figs. 43b), which I am unable to compare with that of *P. stampflii*, is considerably smaller than that of the European *P. pipistrellus* (fig. 43a). The reduction in size is chiefly due to shortening of the rostrum and flattening of the braincase, but, aside from the general difference in size and form, there are no very striking characters to

distinguish the skull of the African animal. The premaxillaries in *P. minusculus* are more abruptly bent downward than in *P. pipistrellus*, the anterior nares thus appearing to open more distinctly forward and less upward. Interpterygoid space *slightly broader than long*. Audital bullæ relatively smaller than in *P. pipistrellus* and space between them actually as well as relatively greater.

Teeth. - The teeth are essentially as in *P. pipistrellus*, though much smaller. Crown of middle upper molar narrower and with more narrowly conical protocone. Hypocone of first and second molars minute but distinct. The mandibular teeth present no peculiarities.

Measurements. - External measurements of type: total length 70; tail vertebræ 31; tibia 9.4; foot 5.4; calcar 9; forearm 26.6; thumb 4.8; second digit 23; third digit 45; fourth digit 40; fifth digit 35; ear from meatus 8; ear from crown 7; width of ear 7; tragus 4.4.

Cranial measurements of type: greatest length 11; basal length 10; basilar length 8; median palatal length 4.6; lachrymal breadth 4.8; least interorbital breadth 3.2; zygomatic breadth 7; mastoid breadth 6.2; occipital depth 3.6; upper toothrow (exclusive of incisors) 4; mandible 7.4; mandibular toothrow (exclusive of incisors) 4•

Specimens examined. - Three, all from the type locality.

Remarks. - I should hesitate to separate this species from *Pipistrellus stampflii* had not Dr. Jentink made comparison of one of the Mount Coffee specimens with the type of the latter. After pointing out an

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error in the original description of *P. stampflii* in which the length of forearm is recorded as 32 mm. instead of 27 mm.. he continues (in letter under date of April 22, 1900) : "your Liberian bat is still smaller [than *P. stampflii*], but its tail is longer. I think it is a new species, particularly as it is much browner-red colored than *stampflii*. As far as I can see the dentition is the same, though all the teeth appear to be smaller." The more important measurements of the type of *Pipistrellus stampflii* are: total length 62; tail vertebræ 24; forearm 27

Pipistrellus musciculus Thomas, 1913

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A very minute species with unicolor fur.

Size excessively small, smaller than in any known bat. General colour perfectly uniform umber-brown, slightly darker than Ridgway's "burnt umber," the hairs of the same colour from base to tip; under surface similar, though appearing slightly lighter owing to the glossy tips to the

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hairs. Ears and membranes very dark, practically black. Inner margin of ears strongly convex at base, straight above; outer margin concave above, convex below. Tragus of medium size, its inner margin slightly concave, its outer convex, with an unusually large angular basal lobe. Wings to the base of the toes. Post-calcareal lobule well developed.

Skull very small, rounded, with short broad muzzle and proportionally high brain-case, not so flattened as in *P. minusculus* and other pigmy species. Palate extended posteriorly further than in any of the allied forms.

Upper inner incisor bicuspid, outer incisor probably also bicuspid, but too worn down in the type for exact description, its tip about equalling the posterior cusp of the inner incisor. Anterior premolar small, not visible externally and not exceeding in height the cingular cusps of the canine and large premolar. Lower incisors small, tricuspid, not crowded or overlapping.

Dimensions of the type (the italicised measurements taken in the flesh): -

Forearm 24.4 mm.

Head and body 40; tail 24; ear 9.5; third finger, metacarpus 23.5, first phalanx 10.6; lower leg and hind foot (c.u) 15.2.

Skull: greatest length: 10.7; condylo-basal length 10.4; basi-sinual length 8; zygomatic breadth 7.5; intertemporal brain-case 5.5; palato-sinual length 4.4; front of canine to back of m^3 3.5; p^4 - m^2 , length on outer edge 2.5.

Hab. Bitye, Ja River, S.E. Cameroons. Alt. 2000'.

Type. Old male. B.M. no. 13.2.8.1. Original number 622. Collected 3rd September, 1912, by Mr. G. L. Bates.

"Caught in hollow palm leaf stalk - another got away." - G. L. B.

This minute species appears to be the very smallest bat as yet described, its forearm being less than an inch in length. It is readily distinguishable from the other small African species by its short stumpy skull, elongated palate, and unicolor fur, the allied species having the fur dark basally and pale terminally. In *P. pusillulus*, Peters, which was described as having a

forearm only 25 mm. in length, the skull is as large as in *P. nanus*, to which Dobson was probably right in referring it; its fur, as in the other small species, was bicolor.

Pipistrellus nanulus Thomas, 1904

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A pigmy species with very short forearm. A bone in the penis.

Size very small. Form fairly stout, the body large in proportion to the short forearms, which are shorter than any hitherto recorded in this genus. Fur of medium length; hairs of back about 4 mm. long. Muzzle thick and broadly rounded, the glandular prominences well developed. Ears rather short, their inner margin straight basally, gradually curving round to form the broadly rounded end to which there is no defined tip; outer margin slightly concave mesially, slightly convex below; basal lobe low, well defined posteriorly. Tragus of medium length, fairly broad, its broadest part opposite the middle of its inner margin; the latter straight or faintly concave, tip blunt, outer margin convex; basal lobe sharply angular. Limbs and feet short; wings to the base of the toes; postcalcareal lobule narrow, distinct; extreme tip of tail projecting; penis with a bone, its prepuce hairy to the tip.

Colour (in spirit) blackish brown above and below. All the membranes also very dark, without lighter edging.

Skull broad and rounded, with large low brain-case and short muzzle; upper profile line practically straight. Base of skull between bullæ broad, its bony edges running outwards quite to the bullæ.

Upper incisors prominently bicuspid; the inner one with its large postero-external secondary cusp falling but little short of the main cusp, and exceeding by about the same distance the top of the outer incisor; secondary cusp of the latter posterior, about halfway up its main cusp. Anterior upper premolar comparatively large, filling up the triangle between the canine and second premolar, which are separated from each other external to it; its tip visible externally just behind the hinder edge of the canine. Lower incisors tricuspid, overlapping.

Dimensions of the type (measured in spirit): -

Forearm 25.5 mm.

Head and body 41; tail 25; ear 8; tragus on inner edge 2.7; third finger, metacarpus 23, first phalanx 9, second phalanx 12; fifth finger 33; lower leg 10; hind foot 5.6.

Skull: greatest length 11.1; occiput to bottom of nasal

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notch 9.5; breadth of brain-case 6.1; front of canine to back of m^3 3.9.

Hab. Efulen, Cameroons.

Type. Male. B.M. no. 4.2.8.8. Collected by Mr. G. L. Bates.

This bat shares with the larger *P. crassulus* of the same region, also discovered by Mr. Bates, a disproportionately short forearm, the length of this member being less than has been hitherto recorded in any Pipistrelle, and nearly the least of any known bat. From *P. minusculus*, Miller, apparently its nearest ally, *P. nanulus* may be readily distinguished by its rounded ears and other differences in detail.

Pipistrellus Rüppelli senegalensis Dorst, 1960

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Type: 1 F, ad, Richard-Toll, Sénégal; septembre 1958; capturé dans un filet tendu près d'un point d'eau.

Cette forme se distingue des *Rüppelli* typiques par une taille supérieure, perceptible entre autres dans la longueur de l'avant-bras. Le crâne comporte un rostre plus fort, plus renflé à son extrémité. Les dents présentent un aspect semblable à celles de *Rüppelli*, mais sont de dimensions nettement plus fortes; la rangée dentaire est légèrement plus longue surtout en ce qui concerne les molaires dont chacune prise individuellement est plus grande, surtout plus large que son homologue chez *Rüppelli*. La première prémolaire supérieure est très petite, pour ainsi dire invisible chez *Rüppelli*. Elle est exclue de la rangée dentaire, le diastème relativement grand chez *Rüppelli* n'existant pas chez la forme du Sénégal; la 2^e prémolaire et la canine se trouvant en contact, la 1^{re} prémolaire est en effet comprimée dans l'angle intérieur de ces deux dents. Ceci peut dans une certaine mesure être interprété comme une conséquence du plus grand développement des dents qui ont repoussé la première prémolaire vers l'intérieur. Les dents de la mandibule inférieure sont elles aussi plus fortes que chez *Rüppelli*.

Notons par ailleurs que les bulles tympaniques sont plus longues et plus volumineuses chez la forme du Sénégal. Voici à titre d'in-

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dications les principales mensurations de notre spécimen par comparaison avec celles de spécimens provenant du Soudan (région du Haut-Nil).

Sénégal; Soudan

Avant-bras: 34.4; 32, -, -

Longueur maximale du crâne: 13.9, 13.6, 13.6, 13.2

Longueur de la rangée dentaire supérieure (C-M³): 5; 4.5, 4.6, -

Le pelage des parties supérieures paraît plus foncé chez *senegalensis*, principalement du fait de la couleur de la base des poils qui est brun noirâtre très foncé chez *senegalensis*, chez qui seule l'extrémité du poil est claire comme la totalité du poil chez *Rüppelli*. Le dos est en fort contraste avec le dessous du corps, recouvert d'un pelage blanc pur. Les oreilles et les membranes sont foncées.

La forme du Sénégal est dans l'ensemble étroitement apparentée à celle de l'Est Africain. Mais plusieurs de ses caractères sont suffisamment distincts de ceux de *Rüppelli* pour justifier la séparation. Cela est particulièrement vrai pour la morphologie et la dimension des dents. *Senegalensis* est manifestement le représentant "occidental" de *Rüppelli*, répandu depuis le Soudan (ex Soudan anglo-égyptien) jusqu'à l'Uganda. Le type de *Pipistrellus* possède donc une aire de répartition qui s'étend largement vers l'Ouest. Sa présence dans toute la bande sahélienne allant du Sénégal à la mer Rouge est probable.

Pipistrellus vernayi Roberts, 1932

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Pipistrellus vernayi sp. nov.

A large species having the raised, but rather swollen, brain case of typical forms of the genus, the outer upper incisor about half the height of the inner, the small anterior upper premolar wedged between and separating the canine from the large premolar. The skull shows grooves on the frontal median line, the cranium swelling up and outwards in the anterior portion, descending behind to the normal height, but with two slight swellings near the occipital and above the mastoid, and swellings again in the occipital region; bullae about equal in diameter to the length of the three upper molars; the larger upper premolar with a canine-like anterior cusp, raised high above the level of the posterior cusps; the tips of the upper canine, large premolar and foremost cusp of the first molar in about the same plane. The small outer upper incisor shows traces of cusps about midway on the inner and outer edges; inner incisors decidedly bifurcate, the two points diverging and under low magnification resembling two teeth, but the outer cusp lower than the inner. The tips of the lower canine, large premolar and two highest points of the m₁ are on the same plane and at an obtuse angle to the line of the tooth-row, but the anterior lower premolar is at the same level as the posterior outer cusps of the large premolar and molars; the lower three pairs of incisors are normally placed, at right angles, and are trifold.

Colour of the upper pale mouse grey and under parts pure white; base of hair above for two-thirds dark grey, but below the hairs are white to the base. Ears naked and dark brown, the muzzle covered with fine bristles, also darker than the back.

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Type: T.M. No. 6546, adult ♂, Maun, Ngamiland, 29th May, 1930, No. 648, Vernay-Lang Kalahari Expedition. Length of head and body 56, tail 41, hind foot (c.u.) 9. ear 12, span of wings 278, tragus 4, penis 12 mm. Forearm 35.3, 1st phalanx of 3rd digit 35.5, 2nd 12.5, 3rd 9.5. Skull: length of canine to occiput 14.6, zygomatic width 9, width of brain case 7.6, width of palate across outside of m² 6, width across upper canines 4, length of upper tooth-row (c-m³) 5, height at bullae 6.7 mm.

Platymops Thomas, 1906

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Ears widely separated, as in *Mormopterus*. Lips without Wrinkles, covered with short stout bristles. Forearms shagreened. Wings short. Metacarpal of third finger only one third longer than that of fifth; first phalanx of fifth finger much shortened, one third the length of the metacarpal and not longer than the second phalanx. A small untufted gular sac present in both sexes.

Skull of the general outline in upper view of that of *Mormopterus*, but extraordinarily flattened vertically. Crests much reduced, the sagittal absent and the lambdoid not

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meeting in the middle line. Premaxillary notch narrow, not expanded behind. Mesopterygoid fossa without median bony ridge. Ramus of lower jaw very low vertically.

Incisors 1^{-1/4}; the upper pair long, nearly parallel, bicuspid, the lower ones subequal, all bicuspid. No trace of a small upper premolar, though the large p⁴ does not touch the canine.

Type *Platymops Macmillani*.

This most remarkable bat is readily distinguishable from any member of the *Nyctinomus* group by its extraordinarily flattened head, unusual digital proportions, shortened wings, and dental formula. In this group it has a curious analogy with the Vespertilionine genus *Mimetillus*, being modified in its proportions and the shape of its head almost exactly in the same way, and having a strong superficial resemblance to it.

Platymops barbatogularis Harrison, 1956

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Amongst a number of bats received from Mr. John G. Williams of Nairobi, which he obtained in the extreme south-eastern part of Sudan during 1953, were two small Flat-headed Bats belonging to the genus *Platymops* Thomas.

Platymops barbatogularis, sp. nov.

As type specimen I nominate an adult male (No. 2.1897) in my collection obtained on June 10th, 1953.

Type Locality. - Lokomarinyang Marsh, Ilemi Triangle, S.E. Sudan, 5° 02' N., 35° 33' E.

Paratype : an adult female (No. 1.1867) in my collection obtained on June 10th, 1953 at the same locality.

Description of the Type.

This specimen is similar to *P. macmillani* in size and in general characteristics. Whereas in *P. macmillani* a small untufted gular sac is present in the lower throat region of both sexes, in this animal the sac contains a prominent tuft of brownish hairs. These hairs can be seen protruding from the semicircular orifice of the sac and when this is everted they are seen to average about 2 mm. in length (see fig. 1). Such a tufted gular sac is not known in Molossid bats.

A further striking character is provided by the distinctive pattern on the chest and abdomen of this animal (see fig. 1). The central part of the chest and abdomen is creamy white in colour extending up to the throat to surround the gular sac. The lateral areas of the chest and abdomen from the axillae backwards are also creamy white, in contrast to *P. macmillani* in which this area is blackish (vide Thomas, 1906). In *P. barbatogularis* the central and lateral whitish areas are sharply divided by a broad brown stripe on each side commencing anteriorly behind the antitragal lobe of the ear and extending backwards, at first curving outwards to the antebrachial membrane and then inwards to extend straight down the belly as a stripe about 3 mm. in width as far as the lower abdomen, which is practically naked. The exact shade of colour of the type is a little doubtful as the specimen has been immersed in alcohol for a time, but it is remarkable that the thoraco-abdominal stripes are clearly a darker shade of brown than the dorsal surface of the body.

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Direct comparison with the colour of *P. macmillani* is not possible as the only available specimen of that species, the type, has been in alcohol fifty years and as a result it is greatly faded. Thomas's description was based on eight examples and from it *P. barbatogularis* clearly differs greatly in colour as well as in pattern. Thomas stated that *P. macmillani* is smoky blackish on the back, dull whitish below in the middle and blackish along the sides. The type of *P. barbatogularis* is a uniform light brown colour above approximating to Drab of Ridgway (plate 3, No. 18) while the darker brown thoraco-abdominal stripes approximate to Sepia of Ridgway (plate 3, No. 3). These colours may have been affected somewhat by immersion but clearly differ greatly from *P. macmillani*.

(Furthermore it may be noted here that a third specimen of this bat in the Coryndon Museum, Nairobi, was prepared very shortly after it was collected and Mr. Williams tells me that this one also was pale light brown in the flesh.* Therefore little doubt remains that the colour of this species is as described above in the type).

* Since going to press this specimen has been sent to the author for examination and its colour is almost identical with that of the type. This specimen also shows a narrow whitish margin to the wing membrane between the foot and the 5th digit, a feature present in the Type but less well marked.

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The skull of the type is slightly shorter than that of the type of *P. macmillani*, with a more squat relatively shortened rostrum. Until more material of both species becomes available however, the significance of this must be considered doubtful. No differences in dentition have been detected. It may be noted here that no trace of a small upper premolar can be found in the type but a minute one is present on the left side in the paratype. A minute upper premolar is present on one side in the type of *P. macmillani* so that its presence must be regarded as a variable feature in the genus.

The paratype (No. 1.1867) closely resembles the type in all the above-mentioned features but the thoraco-abdominal stripes are partially interrupted by a small whitish patch just below the nipples.

Flesh measurements of the type and paratype (in mm.).

Total Length; Forearm; Hind; Foot; Ear

Type No. 2.1897: 80 (fresh); 31.8; 7; 11

Paratype No. 1.1867: 87 (fresh); 31.1; 7; 12.2

(Measurements from spirit, relaxed, except for the overall length, which was measured by the collector in the fresh specimens.)

Skull measurements of the type and paratype (in mm.).

Condylbasal length; Lacrymal breadth; Breadth of Braincase; Depth of Braincase; Mandibular tooththrow (c-m₃; Maxillary tooththrow (c-m₃; Mandible

Type No. 2.1897: 14.9; 7.3; 9.8; 3.7; 5.8; 5.6; 11.3

Paratype No. 1.1867: 14.2; 7.2; 9.7; 3.9; 5.6; 5.1; 10.5

This remarkable little Flat-headed Bat is at present only known from the type locality. Mr. Williams has kindly forwarded some field notes. The species was found over a small permanent waterhole and marsh immediately N.N.E. of Lokomarinyang, which is the north-eastern extension of a block of mountainous country overlooking a vast plain to the north. Immediately above the marsh were cliffs in which there were long, narrow, horizontal fissures but these were so inaccessible that he was unable to confirm whether they were the bats' day-time retreat, but this seems more than likely from the recorded habits of other members of the genus. Although short winged they had a rapid erratic flight when hunting over the marsh about thirty feet up or lower. They appeared when it was nearly dark and they were feeding upon small beetles.

This species is immediately recognizable amongst the known members of the genus by its tufted gular sac and brownish thoraco-abdominal stripes. It is approximately the same size as *P. macmillani*, but considerably smaller than the two species known in S. Africa which are *P. petrophilus* Roberts (Forearm given by Roberts 1951 as 38 - 42 mm.) and *Platymops haagneri* Roberts (Forearm 36 - 37 mm.). Further material of this rare and interesting genus is clearly needed for a more detailed comparison of their coloration and size.

Platymops barbatogularis parkeri Harrison and Fleetwood, 1960

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Type: Adult female, No. 2. Lualeni Borehole, Maktau, southern Kenya Colony (03° 25' S., 38° 10' E.). 12 February, 1958. Collected by Ian Parker. In the collection of the British Museum (Nat. Hist.), South Kensington, London.

Description: Similar in all essential features to *barbatogularis* but larger both in external and cranial dimensions and differing significantly in colouring.

Gular sac present and tufted as in the typical race, its orifice lying transversely in the lower throat region and measuring 3.6 mm. across in the spirit specimen. The tuft of coarse brownish glandular hairs which it contains measures about 1.6 mm. long and becomes obvious when the sac is everted.

The thoraco-abdominal stripes are present as in the typical race and are darker than the dorsal surface of the body, there being a small bare area just distal to the nipples, interrupting the stripe centrally, as in the typical form. The thoraco-abdominal stripe in this animal, however, is broader distally than in *P. b. barbatogularis*, extending further laterally and leaving only a narrow band of pale creamy hairs lateral to it, which extends along the medial edge of the wing-membrane from the axilla to the knee (vide Plate 4). The internal borders of the stripes have just the same disposition as in the typical form, leaving a creamy white central area, commencing at the chin, surrounding the gular sac and widening on the upper thorax and narrowing again gradually towards the lower abdomen, which is naked of hairs at its extremity, as in the typical form.

Although the colour of this specimen may have been modified by immersion in alcohol, it is clear that both the dorsal surface and the thoraco-abdominal stripes are darker than in the typical race. The colour of the back approximates nearly to Grey 31 (Plate 15 C.6, Maerz and Paul, 1950) while the thoraco-abdominal stripes approximate nearly to Castor (Plate 16 A.8, Maerz and Paul).

Measurements of the Type: Total length 91, forearm 34.5, hind-foot 7, tail 29, ear 17 mm.

Cranial Measurements of the Type: Condylbasal length 16, zygomatic breadth 12, breadth of braincase 10.1, depth of braincase 4.2, mandible 12 mm.

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Apart from the marked difference in cranial size from the typical form no essential difference in cranial or dental characters is to be noted.

In view of the great rarity of these bats the paratypical series of *P. b. parkeri* has been studied in great detail in comparison with *P. b. barbatogularis*.

Colour: The colour of the dorsal surface is in all instances much darker than the typical form and varies only a little in the series. The three immature specimens tend to be a slightly greyer brown on the back, and the adult male a shade browner than the three adult females. The pale central area of the throat and abdomen is whiter in the immature animals, more creamy in the adults. The thoraco-abdominal stripes do not differ significantly in shape or colour in the series, save that the bare nipple spots are absent in the adult male and three immature animals. In the adult male the narrow band lateral to the stripe from axilla to knee is brownish instead of creamy or white, although not so dark as the stripe itself. In all the specimens in our series the stripes are darker than the dorsal surface of the body.

Size: In both external and cranial dimensions all the adults exceed the typical form, while one immature female with the upper milk incisor still in situ has dimensions equal to those of the adult Type of *P. b. barbatogularis*. The Tables below bring together the measurements of all known specimens of the species.

[table removed - eds.]

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Gular Sac: The sac is present in all specimens of this series and it is macroscopically as already described in *P. b. barbatogularis*. No difference can be detected between its condition in the adults of both sexes. The transverse opening in the lower throat region measures from 3.6 to 5.5 mm. across, the largest dimension being that of the adult male. The pocket itself is about 3.5 mm. deep and in all the specimens contains the tuft of coarse brownish glandular hairs as described in the typical race. This tuft can be seen in the sac and protrudes from it when the sac is everted. The hairs are from 1.2 to 3 mm. long in the four adults. The tuft is present in the three immature animals, but it is scantier, the hairs measuring 1.2 to 1.6 mm. in length. When the skin of the lower throat region is examined from its internal aspect it is seen that there is a fan-shaped glandular body, creamy white in colour in the fixed specimen, which is flattened dorso-ventrally and which extends to a depth of about 3.5 mm. from the sides and floor of the pocket. In the adult male specimen the greatest width of the sac with its associated glandular body is 13.2 mm., while its greatest length from the orifice of the sac to the caudal extremity of the gland is 6.8 mm., its greatest thickness anteriorly being 3.5 mm. The anterior (i.e. ventral) and surfaces of the pocket appear devoid of glandular tissue macroscopically. While there is no apparent difference in the appearance of this glandular body in the adult male and females, it is noticeably

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less well developed, although present in the three immature specimens. The opportunity has been taken to study the histological structure of the gular sac and its associated gland.

The entire sac of the adult male specimen, with its orifice and the attached glandular body described above, was excised and sections cut across its long axis obliquely and stained with haematoxylin and eosin. Unfortunately fixation of the tissue was imperfect and the haematoxylin staining is consequently rather poor, but the basic structure of the sac is, nevertheless, quite clear.

The actual cavity of the sac is lined with stratified squamous epithelium with a thin layer of desquamating keratin. Hair follicles are seen growing into the sac from all sides. Three types of glandular structure are seen to be associated with the sac (vide Fig. 1). Along that aspect of the pocket which is macroscopically not covered by the fan-shaped glandular body it is seen that the hair follicles are associated with simple flask-shaped glands penetrating deeply below the hair follicle roots. These glands closely resemble those described elsewhere in the gular sac of *Otomops martiensseni icarus* (vide Harrison, 1957) and are regarded as a primitive type of sebaceous gland. In the part of the sac which is covered by the fan-shaped body, however, these glands are seen to be entirely replaced by large complex lobulated glands of sebaceous type. The two types of gland have quite different structure and staining reactions. The primitive glands stain deeply with eosin, they contain no lobules and, as in *Otomops*, their cells appear to proliferate inwards and break down to form a granular eosinophilic debris which discharges into the hair follicles. The complex sebaceous glands are much larger and there are twenty-three separate ones visible in these sections grouped along the central part of the fan-shaped body. They tend to be triangular in shape with their bases distal to the pocket, the largest glands being about 100 μ by about 50 μ in size. Each gland consists of numerous lobules of typical foamy sebaceous cells which discharge their contents into a large intraglandular reservoir. This is situated in the centre of each gland and it has tributaries extending into the larger lobules. The reservoir leads to a wide and nearly straight duct, which is itself lined by sebaceous cells and which is about as long as or a little longer than the gland itself, that is about 100 μ in length and about 10 μ in width. Both the reservoirs and the ducts of these glands contain an oily secretion produced by the breakdown of the sebaceous cells. Each duct discharges into a hair follicle.

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The third type of gland associated with the pocket lies on each side laterally, forming the corners of the fan-shaped glandular mass. In this situation there are six to nine large lobules, irregularly shaped, which are seen to consist of a mass of coiled tubules and which are clearly apocrine glands. Their tubules are lined by a cubical or cubo-columnar epithelium. In an adult female in which the whole sac was similarly excised and examined the histological structure is similar to that of the male, and part of the glandular body removed from the Type is also histologically similar.

It is clear, therefore, that the gular sac of *P. barbatogularis* is a highly elaborated glandular structure; that it contains three distinct types of gland and that it differs materially both in macroscopical anatomy and in microscopical structure from the gular sac as described in other species of *Molossidae*: *Cheiromeles* (Harrison and Davies, 1949), *Otomops martiensseni* (Harrison, 1957), *Molossus* (Schaffer, vide Grassé, 1955, p. 1732). It is interesting to note that the gular sac of *Molossus* differs in many respects from that of *P. barbatogularis*. Thus it occurs only in the males, in which the pocket itself is small without any tuft of glandular hairs. A large mass of gland tissue surrounds the whole pocket, consisting of a deep apocrine layer and a superficial one of sebaceous glands. It is interesting to note that the sebaceous glands of *Molossus* have intra-glandular reservoirs as in *P. barbatogularis* but they open by pores into the pocket, unassociated with any hairs. It is clear that the detailed structure of the gular sac, which occurs in several genera of *Molossidae*, provides important characteristics of the various genera and species, exhibiting a marked diversity of structure in the various species possessing it.

Wing membranes, Ears and Facial Bristles: The narrow pale border of the wing membrane between the tarsus and the tip of the fifth digit is present in all these specimens just as described in the typical race, but it is not well marked in the immature specimens. There is no apparent difference from *P. barbatogularis* in the distribution of the warty excrescences on the forearms, thumbs and metacarpals, and these are present in the adults and immature animals (vide Plate 2); nor can any other differences in the wing membranes be detected, nor in the ears, tragi or facial bristles.

Feet: All the toes of adult and immature animals alike are adorned with long hairs as in the typical race, while the outer aspect of the first and fourth toe is similarly densely covered with short recurved hairs. The long hairs reach about 10 mm. in length, the short ones about .1 mm.

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Body Form: One of the most striking features of the bats of the Genus *Platymops* is the marked flattening, not merely of the head, as indicated by their popular name, but also of the body (*vide* Plate 5). A section through the thorax of an adult female is shown in Fig. 2, made at the level of the mid-point of the sternum, with the skin removed, to illustrate the marked flattening of the thorax. This is without doubt an adaptation to the creature's habitat and represents an extreme degree of the body form towards which all the crevice-dwelling *Molossidae* tend and also some other bats, notably *Tylonycteris*.

Skull and Dentition: As already noted in the description of the Type there is no essential difference in the cranial or dental characters of this race as compared with the typical one, apart from the greater size of the skull in *P. b. parkeri*, and the main features of the skull shown in Fig. 3 are those of the Genus, already described in detail elsewhere (Miller, 1907, p. 255). It may be noted, however, that there is marked individual variation in the development of the lambdoid crest, this being strongly developed in the adult male, less so in the adult females and absent in the immature skulls. Even in the adult male, however, there is no trace of a sagittal crest, the surface of the cranial vault being actually slightly hollowed out in the mid-line.

The dentition also shows no constant difference. It is of interest to record that in this series, as in the typical race, the small upper premolar is inconstant. It is absent on both sides in the adult male and all three adult females, while it is bilaterally present in two of the immature animals and lost on one side in one of them. This confirms the impression that this minute functionless tooth is deciduous and in process of complete loss in *Platymops* and is of no systematic importance; the Sub-Genus *Sauromys* Roberts being untenable on this account. It appears, however, that the southern African representatives of the Genus possess no gular sac, a fact which has received little emphasis hitherto, and it seems that the Sub-Genus might be maintained on the basis of that character rather than on the inconstant dental feature on account of which Roberts first erected it.

It is of interest that one of the immature specimens retains the upper milk incisors *in situ*. The milk tooth lies to the outer side of the permanent incisor and slightly behind it (*vide* Fig 4.), and its tip just reaches the secondary cusp of the permanent tooth. The milk incisor is a tall spicule in form, strongly recurved caudally

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towards its conical tip, and it possesses on its lateral side a faint indication of a secondary cusp corresponding to that of the permanent tooth.

External Genitalia: The opportunity is taken to describe and figure the external genitalia of this species, which have not hitherto received attention.

The penis of the adult male is situated pre-anally and no scrota swelling is visible externally. The penis measures 6.8 from the base of the shaft dorsally and the shaft is 2.1 mm. in width. A prepuce is present, and when this is retracted the glans penis is revealed as a sub-rectangular body when viewed from above (*vide* Fig. 5). The glans measures in length 1.8, by maximum width 1.6, by .6 mm. in depth. It is flattened dorso-ventrally and bluntly expanded distally with a slight ridge-like elevation in the mid-line dorsally. The urethral meatus is situated in the centre of an oval smooth area on the distal part of its ventral surface, which is hollowed out when seen from the lateral aspect. The remainder of the glans is covered by minute horny excrescences. A few long hairs project over the glans from the prepuce dorsally and the shaft is covered with short hairs.

The vulva of the female (*vide* Fig. 6) is slightly elevated above the surrounding skin and the skin around the vaginal and anal orifices is pigmented blackish. The vaginal orifice is transverse, measuring 2.8 mm. in width, and the mucosa at the orifice is smooth. The glans clitoridis, which is oval and rather flattened, projects from an opening between the vaginal orifice and an elevated clitorideal pad anterior to it. This pad is surmounted by two blunt elevations laterally, between which arise a few long hairs projecting over the glans clitoridis. The anal orifice is puckered by mucosal rugae and it is separated from the vaginal orifice by a small perineal space.

Breeding Biology: The nipples were prominent in all three of the adult females, lying below and slightly anterior to the axilla and measuring from 2.2 to 3.8 mm. long and from .9 to 1.2 mm. in width; their long axis lying parallel to that of the thoraco-abdominal stripe. The mammary glands of all three were hypertrophied measuring from 11-15.8 in greatest length and from 6-7.1 in width and about 3.8 mm. in maximum thickness. The mammae were situated over the lateral aspect of the thorax and upper abdomen, only just extending on to the ventral and dorsal surfaces of the trunk, differing in this respect from the condition noted in the lactating *Tadarida (Chaerephon) pumila websteri* (Harrison, 1958). Histological sections were prepared of one mammary gland from each of the adults, and these were stained with haematoxylin and eosin.

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These fully confirm the condition of lactation, the glandular acini being distended with globular secretion and the epithelium actively secreting, while many of the ducts are distended with milk. This is consistent with the presence of immature animals in the series, one with upper milk incisors *in situ*. The uteri of these adult females were also examined, but although there was a slight swelling of the right cornu in the Type, measuring 2 in width by 3 mm. in length, no evidence of a new pregnancy was found in the sections prepared. It is clear, therefore, that in this series a season of parturition had occurred a little before the time of capture in mid-February.

Habits: These remarkable creatures were found beneath a stone, where they shared their retreat with scorpions. It is of interest to note that in South Africa *Platymops* have occasionally been found by zoologists who were in fact searching for scorpions beneath flat slabs of rock (Roberts, 1951, p. 103). It is clear that the whole body form of these bats is remarkably well adapted for such narrow recesses, and it may be concluded that the glandular secretion of the gular sac in *P.*

barbatogularis becomes smeared on the floor of the retreat as the bat crawls about within it, and perhaps thus marks the home of the colony with a characteristic odour. It may also be that the tuft of glandular hairs in the sac may act as a kind of scent paint brush. Whether there is any special significance in the association with scorpions is a point that might well claim the further attention of zoologists.

SYSTEMATIC REMARKS

The occurrence of this form in southern Kenya led to a careful consideration of the description of *Mormopterus setiger* Peters, 1878, amended in 1881 (type-locality: Ndi, Teita, Kenya Colony). It is clear from certain points in the description that this form is a *Platymops*. Thus the upper incisors are bicuspidate, the skull very flattened (unfortunately no cranial measurements were given), the upper surface of the forearms is covered with warts and a gular sac is present, which is apparently untufted. In all these features it resembles *P. macmillani*, from which, however, it differs in being larger and also different in colour. It is larger than typical *P. barbatogularis* and approximately the same size as *P. b. parkeri*, and differs from both these forms in colour and in lacking the thoraco-abdominal stripes, which are darker than the dorsal surface of the body in *P. barbatogularis*. It also differs by lacking the glandular tuft in the gular sac. It is unfortunate that the Type of *P. setiger* has probably been lost, according to Prof. Dr. K. Zimmermann, (*in litt.*), so that the further elucidation of the status of this form must await the collection of further material.

Platymops haagneri Roberts, 1917

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Platymops haagneri spec. nov.

A small species having an affinity to *P. petrophilus* in the dental formula, flattened skull, absence of a definite antitragus, shortness and number of the bristles on the toes, and in the horny points on the upper part of the ears being situated near the horizon of the ear; but differing therefrom in its smaller size, the horny points on the ears only two in number; the lower P2 much smaller than the P4, which is higher, the tips of M₁, P₄ and C forming an almost straight line at a slight angle rising forward; the upper incisors are also comparatively shorter than in *petrophilus*, and a cingulum can be seen in P2 with the aid of a lens. The lower incisors are bifid. The bony palate extends well behind the molars.

Dimensions of the type in spirits: Length of the head and body 50 mm.; of the tail 32, of the free portion of the tail 13.5; ear, height 10, breadth 13.5; height of the tragus 2.3; length of forearm 37; pollex (c.u.) 5; third digit, metacarpal 37, 1st phalanx 14, 2nd without cartilage 12; fourth digit, metacarpal 36, 1st phalanx 12.5, 2nd phalanx without cartilage 6; fifth digit, metacarpal 25, 1st phalanx 10, 2nd 4.7; tibia 10; hind foot, length (c. u.) 6.5. Skull: Greatest basal length 16.2; zygomatic width 10; width at mastoid 9.8; width of brain case 9; interorbital constriction 3.7; basilar length 13.1; palate length 5.5; length of dental series, from front of C to back of M3, 6; width of palate taken outside the molars 7; greatest length of mandible 12.

Type: Adult male from Keetmanshoop, Damaraland, presented to the Museum by Mr. Sigmund Haagner.

These two species apparently belong to the genus *Platymops* Thos., but differing as they do in the dental formula, the P² being present, I propose to place them in a new subgenus bearing the name of SAUROMYS, of which *P. haagneri* maybe taken as the genotype. The following figures will indicate the difference in the height of the skull of *N. aegyptiacus* and the present new species, the measurements having been taken from the tips of the P⁴ vertically parallel to the top of the premaxilla:

N. aegyptiacus 6.5 mm., *P. petrophilus* 4.2, *P. haagneri* 4.

Platymops haagneri umbratus Shortridge and Carter, 1938

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Platymops (Sauromys) haagneri umbratus subsp. n.

One specimen from Kliphuis, northern spur of the Cedarberg.

A Cape subspecies of *P. haagneri*, distinguished by its dark coloration.

Colour above smoky seal-brown, a shade paler than in *Nyctinomys bocagei*; under surface smoky-brown, several shades lighter than the

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upper surface; ears and flying membranes almost black - as opposed to comparatively pale horn-brown in typical *haagneri*.

In typical *haagneri* (series from Goodhouse, lower Orange River, examined, and a single specimen from Berseba, Great Namaqualand) the colour above is drabby brown, paler and less cinereous than the under surface of *umbratus*, and pale burly, approaching whitish, below.

Type (in the Kaffrarian Museum). - Adult male, original number 2827, collected 7th December 1937 at Kliphuis, Pakhuis Pass, 11 miles N.E. of Clanwilliam, N.W. Cape Province.

Dimensions of Type. - H. and b., 61; tl., 31; hf., 8.5; ear, 15; forearm, 37 mm.

Skull. - Basal length, 16.2; zygomatic width, 10; width of brain case, 9; interorbital constriction, 3.7; length of palate, 5.5; upper dental series (from front of canine), 6; mandible, 12.5 mm.

Genus new to the Cape Province.

Platymops Macmillani Thomas, 1906

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Size about as in *Mormopterus acetabulosus*, the abnormally short forearms giving a deceptive idea of the general bulk. Fur short; hairs of back about 3 mm. in length. General colour above smoky blackish, below dull whitish along the middle area, blackish along the sides. Limbs dark above, whitish below; membranes dark throughout, except that the surface near the body below is lighter, as is the extreme edge of the plagiopatagium. Ears widely separated, triangular, their anterior edge evenly convex, their tip rounded; antitragal notch scarcely perceptible. Tragus fairly large, quadrate, its inner edge about equal to its breadth at tip. Upper side of forearm, wrist, and basal half of third metacarpal thickly and coarsely shagreened with small round warts, very much as is the case in *Vespertilio Floweri*, de Wint. Wing-membrane attached to the distal third of the tibia. Upper side of the short lower leg flattened, finely granulated. Proximal plantar pad very prominent. Penis short, without special modifications.

Skull and teeth as described above.

Dimensions of the type (measured on the spirit-specimen): -

Forearm 32 mm.

Head and body 51; tail 27; tail free from membrane 15; ear 15; tragus on inner edge 2; third finger, metacarpus 31, first phalanx 12.5, second phalanx 11; fifth finger, metacarpus 23, first phalanx 6.5, second phalanx 6.7; lower leg 11; calcar 16.

Skull: condylo-basal length 16; basal length in middle

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line 13; zygomatic breadth 11.2; breadth between preorbital processes 7.4; interorbital constriction 3.8; greatest mastoid breadth 10.2; height from basion to top of brain-case 4; palatal length in middle line 5.9; front of canine to back of m^3 6; front of lower canine to back of m_3 6.6.

Hab. N.E. Africa, between Adis Ababa and Lake Rudolf.

Type. Adult male. Presented by W. N. McMillan, Esq. Collected by Ph. C. Zaphiro. Seven specimens examined.

The peculiar flattening of the head of this bat - even to the abolition of the sagittal crest and the reduction in the vertical thickness of the lower jaw - probably indicates that it either lives in small cracks in trees or rocks, into which it may creep beyond reach of enemies, or else that it frequents some large-leaved plant (such as banana or cactus), between whose leaves the other bat with a similarly shagreened forearm (*Vespertilio Floweri*) was found by Mr. N. C. Rothschild.

I have named this most interesting bat in honour of Mr. W. N. McMillan, to whose liberality science is indebted for the exploration of which it is part of the outcome.

Platymops petrophilus Roberts, 1917

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This bat has a superficial resemblance to *Nyctinomus aegyptiacus* (S. African specimens), but upon close examination is found to differ in having the ears more widely separated at the inner base; the antitragus represented by a mere thickening of the lobe; horny points on the upper part of the ear commencing near the rim of the ear and not far down as in *aegyptiacus*; the thumb very thick; the first and fifth toes also very thick - equal to double that of either of the other three - and thickly covered above and on the outer surface with fine whitebristly hairs, but with hardly any of the very long vristles found in *aegyptiacus*; the ears are haired as in *aegyptiacus*, but the area around the fibulae is more sparsely covered with white hairs. The ears arise about 4mm. apart to a height of a little more than 1 mm. and the upper margin thence for about 13 mm. is practically horizontal, thence curving downwards little way, increasing the breadth of the ear to 15 mm., then downwards and very slightly inwards for about 9.5 mm. to a point whence an increasing tendency inwards for 4 mm. carries the lower margin in an almost straight line to the outer base; the antitragus is represented by a broadening and thickening of the lobe, flattened or slightly folded on the upper surface, with only a slight emargination beyond its outer edge; the tragus is like that of *aegyptiacus*, quadrate, rather narrower at the base than the tip, with a slight notch in the outer edge. The horny points on the upper margin of the ear are five in number just below the horizon, the rest on the edge of the ear, producing a saw-like horizon. The lips are wrinkled, the nostrils sub-lateral and no glands appear to be present. The wings arise on the tibia at about one-third of the length of the tibia from the feet.

The skull of the type is, unfortunately, fragmentary, having been crushed when the rock was overturned under which it had taken up its abode. Nevertheless, from what there is of it, the dentition is clearly seen to be numerically identical, with that of *N.*

aegyptiacus, but the skull is flattened as in *Platymops* Thos. (Ann. & Mag. N.H., ser. 7, vol. 17, p. 499, 1906); the premaxillae do not meet anteriorly, and the incisors are large and convergent; the foremost premolar (P 2) is small but easily seen without the aid of a lens, and is situated between the C and P4, well within the tooth row; the lower anterior premolar is slightly smaller than the posterior premolar.

Dimensions (taken from a spirit specimen): Length of the head and body 53 mm., of the tail 38, of the free portion of the tail 15; diameter of the ear, vertically 14, horizontally 15; height of tragus 2.5; length of snout to the inner base of the ear 7; forearm 42; thumb 7; third digit, metacarpal 43, 1st phalanx 16, 2nd without the cartilage 15, with the

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cartilage 22; fourth digit, metacarpal 41, 1st phalanx 15, 2nd phalanx without cartilage 8.5, with cartilage 11.5; fifth digit, metacarpal 27, 1st phalanx 11, 2nd with the cartilage 7.5; tibia 9; hind foot length (c.u.) 8, breadth 4; breadth of ankle pad 2. Skull: Length of dental series, from front of C to back of M3, 7.4; greatest length of mandible 13.5.

Type: Adult male, in spirits, taken by G. van Dam at Bleskop, near Rustenburg, 2nd February, 1917.

Platymops petrophilus erongensis Roberts, 1946

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Of about the same size as *P. p. petrophilus* Roberts (Ann. Transv. Mus. VI, 4; 1917: Bleskop, Rustenburg district), but in colour pale brownish grey above (instead of tawny olive), below greyish white (instead of pale tawny olive), and ears light brownish instead of dark brown.

Measurements of eighteen adults (including the type) from Eronga Mountain, and, in brackets for comparison, of two specimens, including the type, of which the skull is broken, of *P. p. petrophilus*: Head and body 55 - 62 (53, 53); tail 33 - 40 (38, 35); hind foot (c.u.) 7 - 8.5 (8); ear 13 - 17 (14, 13.5). Wing span in ten measured in the flesh 275 - 302. Forearm 38 - 41.5 (42, 39.5); 3rd digit, metacarpal 39.5 - 43 (43, 42); 1st phalanx 15 - 16 (16, 17.5), 2nd phal. 16.5 - 19.5 (22, 20); 4th digit, metacarpal 38.5 - 42.5 (41, 42); 1st phal. 12.5 - 14.5 (15, 14.5), 2nd phal. 8.5 - 10.5 (11.5, 11.2); 5th digit, metacarpal 26 - 29 (27, 27.5). Tibia and foot (c.u.) 18 - 19.5 (19, 19). Skull, greatest length 17 - 18 (17.5); nasal cavity to occiput 15 - 16.5 (15.6); basilar length 13.2 - 14.8 (15); zygomatic width 10.5 - 11.6 (11.5); mastoid width 9.8 - 10.8 (11.3); width of brain case 8.2 - 8.9 (9); posterior median height 4.6 - 5 (4, 6); interorbital, anterior, width 6.3 - 7.1 (7.2); interorbital-constriction 3.5 - 4 (3.7); c-m³ series 6.1 - 6.5 (6.4, 6.4); width across m³ 7.2 - 7.7 (7.3); width across upper canines 4.3 - 4.7 (4.7); mandible length 11.8 - 12.5 (12.8, 12.8); c-m₃ series 6.7 - 7.3 (7.1, 6.8).

Type: T.M, No, 9494, adult M, Ombu Farm, Eronga Mountain, Omaruru district, South-west Africa; also nineteen more skins and skulls and others in alcohol.

Platymops petrophilus fitzsimonsi Roberts, 1946

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In size about the same as *P. p. petrophilus* Roberts, but in colour paler and more greyish, especially about the head and neck, which are lighter than the back, and below paler and more whitish; the ears are blackish, contrasting with the hair of the head.

Measurements of the type and another F captured at the same time: Head and body 60, 57; tail 40, 35; hind foot (c.u.) 9, 9; ear 15, 14; wingspan 292, 272. Forearm 40, 38; 3rd digit, metacarpal 41, 40; 1st phal. 15.5, 14.5 and phal. 19, 18; 4th digit, metacarpal 41.5, 41.5; 1st phal. 13.5, 13; 2nd phal 10.5, 9.5; 5th digit, metacarpal 10.5, 9, Tibia and foot (c.u.) 19, 19. Skull, greatest length 17.8, 17.2; nasal cavity to occiput 16.2, 15.5; basilar length 15, 14; zygomatic width 11, 10.5; mastoid width 11, 10; width of brain case 9, 8.5; posterior median

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height 4.7, 4.6; interorbital, anterior, width 6.7, 6.8; interorbital constriction 3.6, 3.7; c-m³ series 6.1, 6.1; width across m³ 7.5, 7; width across upper canines 4.5, 4.4; mandible length 12.3, 11.8; c-m₃ series: 6.5, 6.3.

Type: T.M, No, 8968, adult F, Mitchell's Pass, near Ceres, Cape Province; also another F from the same place; captured by Dr V. FitzSimons under a cleft rock.

Plecotus aethiopicus Heuglin and Fitzinger, 1866

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Von dieser bis jetzt noch unbeschriebenen Art überbrachten Heuglin's Leute ein Exemplar vom Bahr-el-abiad.

Plecotus balensis Kruskop and Lavrenchenko, 2000

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Holotype. Adult male No. S-164904 (collection of the Zoological Museum of the Moscow State University), Harenn Forest, Bale Mountains National Park, southern Ethiopia, 6°45' N, 139°44'E, 2760 m ASL, December, 29, 1995, collected by S. V. Kruskop. Dry skin, skull, and carcass in alcohol (Figs 1, 5, 7).

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Other specimens studied. Two adult males, Nos. 8825, 66732 (collection of the Zoological Institute, St. Petersburg), locality unknown ("Abissinia" = Ethiopia; probably vicinity of Addis-Ababa), specimens in alcohol; adult female, No. HFE 244 (collection of the Bale Mountains National Park), Harenn Forest, Bale Mountains National Park, southern Ethiopia, August, 15, 1986, collected by the Harenn Forest Expedition, skin only; two adult males and two females, Nos. LM.1986.212.61-64 (collection of the Liverpool Museum), same locality, August, 4 - 19, 1986, collected by HFE, three specimens in alcohol, one dry skin with skull.

Diagnosis. A typical medium-size representative of *Plecotus* s. str. Differs from all subspecies of *Plecotus austriacus* by dark-brownish coloration of fur, small skull size, relatively short rostrum, noticeably smaller and rounded tympanic bullae, inflated muzzle behind nostrils and baculum shape. *Plecotus balensis* differs from *Plecotus auritus* in having more inflated and slightly larger tympanic bullae, more

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slender tooth row, narrower rostrum, small protuberances in front of eyes, smaller baculum, and absence of yellowish component in fur coloration.

Measurements of the holotype (in mm). External: body length - 45, tail length - 48, ear length - 37, tragus length - 15.2, tibia length - 17.4, foot length (without-/with claws) - 7.1/8.3, FA - 36.3, DI (without-/with claws) - 5.4/7.0, Mc2 (metacarpal of the second digit) - 33.5, Mc3 - 35.0, Ph3.1 (first phalanx of the third digit) - 14.0, Ph3.2 - 12.8, Ph3.3 - 7.2, Mc4 - 34.4, Ph4.1 - 9.0, Ph4.2 - 10.5, Mc5 - 33.3, Ph5.1 - 9.0, Ph5.2 - 10.0. Skull: CBL - 15.21, CCL - 14.64, W - 8.92, BCH - 5.81, IOW - 3.71, WR - 4.13, LR - 3.38, CM3 - 5.42, C - 0.98, P4M3 - 4.18, CC - 3.54, M3M3 - 5.76, LMD - 10.15, MCM3 - 5.76, ABL - 4.28.

Etymology. The name *balensis* is derived from the Bale Mountains, the area from where the type specimen originated.

Description. Medium-sized *Plecotus*, noticeably smaller than typical *Plecotus austriacus* and *Plecotus teneriffae*. Average forearm length is 38.5 mm (n=6) which is less than in all forms of the *Plecotus austriacus* complex and in *Plecotus teneriffae*, and agrees with measurements of European *Plecotus auritus*. Skull on the whole (Fig. 1) and tooth-row visibly smaller than in European and North African specimens of *Plecotus austriacus*.

The most important skull features are the size and shape of the auditory bullae. In the two measured skulls from Harenn forest they have a maximum length of 4.28 and 4.16 mm which is less than in any *Plecotus austriacus* subspecies. In this species the maximum length of bullae according to Strelkov (1988) is never less than 4.3 mm (4.41 mm is the minimum among specimens measured for this study). In two specimens of *Plecotus austriacus christiei* from Israel (H.Z.M.1.3511) and Egypt (ZIN.8822), the bullae have a length of 4.88 and 4.62 mm. Thus, the size of the auditory bullae of the new form seems to be more similar to *Plecotus auritus* than to *Plecotus austriacus* (Fig. 2). However, the bullae of *Plecotus balensis* look more inflated than those of *Plecotus auritus* and some specimens of *Plecotus austriacus*.

The rostrum looks slightly shortened compared to the of the typical *Plecotus austriacus*. Small premolar displaced lingually from the midline of the tooth-row but not reduced in size. All teeth, including the canines, look relatively less robust than in *Plecotus austriacus* and in this respect are similar to those of *Plecotus auritus* and *Plecotus teneriffae* (Ibanez & Fernandez 1985).

The scatter plot of the first two principal components (Fig. 3) shows that *Plecotus austriacus* and *Plecotus auritus* are clearly separated, whereas the two specimens of the new species are obviously closer to *Plecotus auritus*, lying on the periphery of its range.

The two "large" species of long-eared bats are well segregated by the stepwise discriminant function analysis. Based on the 9 metric cranial parameters chosen, 100 % of *Plecotus austriacus* and 98 % of *Plecotus auritus* were allocated correctly. In respect of cranial shape on the whole, the specimens from Bale associate closer with the latter species but are still far from its centroid (Fig. 4).

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Externally the new form shows features somewhat intermediate between the two Palearctic species. Protuberances in front of the eyes are relatively small, the distance between them in all specimens seen is more than three times their diameter which is characteristic of *P. austriacus*. The frontal part of the muzzle is thinly covered with hairs. Like in *P. auritus* (Strelkov 1985, 1988; Fig. 5), the muzzle behind the nostrils is noticeably inflated. Thumb length without claw (ca. 5.8 mm) is similar to that of *P. austriacus* (both *austriacus* and *christiei*; 4.7 - 6.3) and distinctly smaller than in Mediterranean *P. auritus* and *P. teneriffae* (5.9 - 7.6; 6.0 - 6.5) (dePaz 1994).

Coloration of *balensis* is noticeably darker than in all North African, Arabian and European forms of *P. austriacus* (Fig. 6). Dorsal hairs have dark brown bases, covering more than half of their length, a pale ring in the upper third and a brownish-grey tip, visibly darker than in specimens of *P. austriacus* from Israel, Algeria and southern Europe. Ventral hairs have long whitish or pale greyish tips and dark greyish bases. As in *P. austriacus*, there is practically no brownish component in the fur coloration, in contrast to that of *P. auritus*. Wing membranes and hairless parts of the head are brown; fingers, arms and hind

feet are dark blackish brown. It should be noted that relatively dark-coloured subspecies of *P. austriacus* are known only from Asian mountain areas, while the closest neighbour of the Ethiopian form is *P. austriacus christiei* which has a very pale coloration and almost colourless membranes and naked body parts.

The karyotype of the new form (2N=32, NF=50) shows no differences to Palearctic long-eared bats (Fedyk & Fedyk 1971).

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The shape of the baculum is considered as a good diagnostic trait distinguishing *P. auritus* and *P. austriacus* (Topal 1958, Strelkov 1988, 1989). The baculum of *P. balensis* to some extent combines characters of *P. austriacus* and *P. auritus*. Its general shape looks more similar to that of *P. teneriffae* (Ibanez & Fernandez 1985). It is noticeably more slender than the baculum of *P. austriacus*. Lateral projections on its proximal end are relatively thin with slightly bulbous tips, similar to those of *P. auritus*, but proportionally longer than in both *P. auritus* and *P. austriacus*. Thus, maximum width of the baculum of *P. balensis* is subequal to its maximum length (Fig. 7). The last feature is similar to the baculum of *P. teneriffae*. The basal projections in the latter, however, are thin, while in *P. balensis* they are thicker than the main portion. Another specific feature of the *P. balensis* penial bone is a perforation of its distal portion, not present in other *Plecotus*. Size of the baculum of the new species (0.9 mm) is noticeably smaller than in *P. auritus* (1.02 - 1.47 mm; Strelkov 1988) and slightly larger than in European *P. austriacus* (0.7 - 0.85 mm; Strelkov 1988) and *P. teneriffae* (0.8 mm; Ibanez & Fernandez 1985).

Distribution and remarks on natural history. The Ethiopian long-eared bat was reported by Largen et al. (1974) from only two localities. The Eritrean specimen may be *P. austriacus christiei*, and the second record has no exact locality ("Shoa"). Specimens deposited in St. Petersburg also have no precise geographical data, and probably came from somewhere on the Ethiopian upland north or north-west of

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Addis-Ababa. The only fully documented records of the new species are from the Harennna Forest, Bale Massif (Yalden et al. 1996; our data) and represent the southernmost point of the known distribution of the genus. According to the available data, *P. balensis* is confined to the upper belts of the Harennna Forest, Bale Mts. It is possible that the new species occurs also in some other forested areas of Ethiopia. Nevertheless, we failed to find any *Plecotus* in two other humid Afromontane forests on the opposite side of the Ethiopian Rift Valley (western plateau): the Beletta Forest (07°32'N 36°33'E, 2250 m ASL) and the Sheko Forest (07°04'N 35°30'E, 1930 m ASL).

According to our observations, the long-eared bat in the Bale Mountains inhabits humid forest belts at elevations from about 2500 - 3000 m ASL and seems to be a relatively common species. The preferred foraging places are open parts of forests or edges of clearings where both perch hunting and aerial hunting activity were observed. Although the bats emitted audible (social?) calls, we failed to detect their echolocation signals with a QMC-mini narrow-band heterodyne detector, suggesting that the signals are of very low intensity, a characteristic of *P. auritus* hunting in cluttered spaces.

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Discussion

At present, 32 mammalian species endemic to Ethiopia are known (Yalden et al. 1996; Lavrenchenko et al. 1998a, 1998b). Most of them have evolved from Afrotropical ancestors (Yalden & Largen 1992). Only three endemic species (*Capra walie*, *Lepus starcki* and *Canis simensis*) are putative derivatives of Palearctic lineages (Yalden et al. 1996). It is obvious that *Plecotus balensis*, being a member of the essentially Palearctic species complex, belongs to the latter group of endemics.

Within *Plecotus* s. str., *P. austriacus* is the most widely distributed species. It inhabits a wide range of habitats which results in a high level of variation among populations. Such variation concerns coloration, which is whitish or pale-grey in desert and semi-desert forms, and darker in specimens from mountain areas or from the European forest zone. However, *P. austriacus* seems to be less a forest inhabitant than *P. auritus* and its coloration is generally paler. Differences in rostrum proportions, bulla size and shape of muzzle are probably connected to the habits of the species; *P. austriacus* is reported to be mainly an aerial forager, while *P. auritus* is a typical gleaner (Andersen & Racey 1991, Kruskop 1999). Inflations behind the nostrils as well as the smaller size of bullae characteristic of *P. auritus* may be associated with some parameters of the echolocation signals used during foraging in highly cluttered space. While one would expect similar transformations in populations of the *P. austriacus* group confined to forested areas, the only population which shows such modifications is the Ethiopian long-eared bat, which is probably a derivative of the *P. austriacus* lineage.

The range of the Ethiopian long-eared bats is separated from the European forest subspecies *P. austriacus austriacus* by the "belt" of *P. austriacus christiei*, a pale semi-desert subspecies with greatly inflated bullae and no inflations on the muzzle. That subspecies occurs in Egypt, Syria and Israel (Harrison & Bates 1991) and has also been reported from Eritrea (Largen et al. 1974) and Sudan (Koopman 1975). Possibly the ancestor of the Ethiopian *Plecotus* belonged to a lineage of *P. austriacus* which penetrated into montane Ethiopia in one of the inter-glacial periods, when the climate of northern Africa was more humid and the vegetation more abundant. There this population evolved in partial or complete isolation. The closest sibling (*christiei*) evolved in a different habitat, bordering the Ethiopian uplands from Mediterranean, Europe and West Asia. It has been suggested (Lavrenchenko 2000) that the small mammal fauna of the Harennna Forest was formed rather recently and mainly through a recruitment of species from adjacent altitudinal zones with their subsequent adaptive speciation. Apparently, this process was prompted both by an ecological vacuum in the forest communities and the relative richness of the non-forest Ethiopian fauna. According to this hypothesis, the ancestor of *P. balensis* must have come from savanna or any other arid habitat. The forest island conditions shaped the Ethiopian long-eared bat to a typical forest species including some features which are convergent to *P. auritus*. A parallel exist in another island population, *P. teneriffae*, which also inhabits relatively humid woodlands in the Canary Islands (Ibanez & Fernandez 1985).

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An alternative hypothesis would assume that the craniometric similarity between the Ethiopian long-eared bat and *P. auritus* is accounted for by phylogenetic proximity rather than parallelism. In this case we may suppose that the range of the immediate ancestor of the latter species was much wider in the past covering areas adjacent to Ethiopian highlands but later underwent substantial reduction. As *P. austriacus* replaced *P. auritus* in North Africa, some isolated populations of the latter could remain in peripheral refuges giving origin to *P. balensis*. In the framework of the considered hypothesis an analogous scenario might be suggested for the three other forms, similar to *P. auritus* and separated from the main distribution area of the latter (Fig. 8): *P. teneriffae*, *P. "auritus" homochrous* Hodgson, 1847 (with the synonym *puck* Barrett-Hamilton, 1907, shown in fig. 8 as Himalayan section of *P. auritus*) and the recently described *P. taivanus* (Yoshiyuki 1991).

A third hypothesis may suppose that *P. balensis* is a pre-glacial relict, a sister group of the common ancestor of both *P. auritus* and *P. austriacus* which has survived at the periphery of the core distribution of the genus. This view is supported by the "intermediate" morphology of this species. It is important to notice that intermediate characters exist not only in external or cranial traits which may be a result of adaptive evolution, but also in those (baculum) not known to be affected by environmental factors. The similarity of the baculum between *P. balensis* and *P. teneriffae* may also be interpreted as a shared relict character. To test these three competing hypotheses additional studies using molecular techniques will be necessary.

Plecotus teneriffae Barrett-Hamilton, 1907

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Plecotus teneriffae sp. n.

This bat resembles *P. auritus* of Britain, but has much larger wings. The colour is apparently darker, but cannot be taken as reliable from the old dried skin upon which this description is based: it is, above, near Ridgway's "hairbrown," the hair-tips lighter; below, dirty yellowish white, the dark basal portions of the hairs not evident.

The type has no skull.

The *dimensions* (in millimetres) of the type are as follows: -

Head and body 52; tail 45; ear from the notch 35; tragus 14; thumb without claw 6; longest digit 73; basal joint of fifth digit 35; basal joint of second digit 37; forearm 44; tibia 18.5; hind foot (without claw) 8.

Hab. Teneriffe.

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The type, a dried skin, is No. 87.4.18.1 of the British Museum collection, and was taken at Orotava, Teneriffe, by Señ. R. Gomez on the 3rd of April, 1887.

The large wings of this bat mark it as distinct from any other known *Plecotus*.

Propottininae Butler, 1984

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Subfamily PROPOTTININAE nov.

Diagnosis: Lower molars with low relief and poorly differentiated cusps, but posterior trigonid crest retained, and talonid not shortened; M/3 less reduced than in other Pteropodidae, and M3/ probably present; mandible fairly robust.

Propotto Simpson, 1967

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Type-species: *Propotto leakeyi* new species.

Known distribution: Miocene of Kenya; Songhor and probably Rusinga.

Diagnosis: Lorisids with variable lower cheek teeth closely resembling those of *Perodicticus*. P3-M3 rounded, ovate in outline with low, blunt, poorly defined cusps. P4 not oblique, excavated anterointernally and with vague metaconid; talonid with shallow basin, papillated. M1-2 with low, blunt, almost median protoconid and smaller metaconid posterointernal to it. Talonids basined, with blunt hypoconid; entoconid region papillated without clear definition of cusps. Ms reduced, essentially a flattish crown with a central basin surrounded by a vaguely cuspidate rim.

Discussion: The apparently degenerate characters of the highly peculiar cheek teeth of this genus are like those in the Recent *Perodicticus* but may be even more extreme in the fossils. The details are so extremely variable in both that it is conceivable that almost any apparently diagnostic feature of the fossils might recur in the living forms, but in fact I cannot match all the

characters listed above in a fair series of *Perodicticus* jaws and generic distinction seems warranted. Another possible distinction is that all three of the fossil jaws seem to deepen forward in a peculiar way that I have not seen in *Perodicticus* but that is approached in some specimens of its Asiatic vicar *Nycticebus*.

The three specimens on which this genus is based were not known to Clark and Thomas, and they have not heretofore been mentioned in print.

Propotto leakeyi Simpson, 1967

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Holotype: Specimen R of consolidated list.

Hypodigm: Specimens R, S, and T.

Known distribution: Miocene, Songhor, and probably Rusinga.

Diagnosis: Only known species of the genus. Measurements of lower teeth as in Table 4.

Discussion: The generic diagnosis calls attention to the most striking features of these specimens, and the accompanying figures obviate need for further verbal description. These peculiar dentitions are unlike any previously described in fossils and resemble those of the living pottos but are perhaps even more extreme in modification of the more primitive, more insectivore-like cheek teeth of other lorids. The three specimens vary markedly in size, proportions, and structure of the teeth and at first sight seem hardly compatible with reference to a single species. However, *Perodicticus potto* is almost if not quite as variable in the same ways, and valid separation of species among the fossils will require larger samples, if it is called for at all.

SKULLS AND UPPER DENTITIONS

No associated upper and lower dentitions are known. The one Uganda specimen, the facial part of a skull, is the only nomenclatural type including upper teeth. As it cannot be compared with the Kenya types, it is a special case.

The Kenya specimens of skulls and upper teeth are from the same localities or areas as the lower jaws, including the five nomenclatural types, and it is presumable that they may for the most part if not altogether represent the same species. As is almost always true of fossil mammals, more lower than upper teeth and jaws are known.

Lower teeth of *Progalago dora*, *P. songhorensis*, *Komba robustus*, and *K. minor* fall into four corresponding size groups without overlap among the known samples. *Propotto leakeyi* is of about the same size as *Progalago dora*, but it is so distinctive morphologically that its upper teeth could almost surely be recognized if found. (They are not known.) The smallest species known from lower dentitions, *Komba minor*, should be recognizable by its size, or, at least, upper teeth too large to occlude can be excluded from consideration. In fact none of the upper teeth here available could belong to this species.

There remain three species of different sizes based on lower jaws and teeth: *Progalago dora*, *P. songhorensis*, and *Komba robustus*, in sequence of decreasing size. Of the eight specimens with upper teeth, one (AI of consolidated list) is so poorly preserved as not to warrant any attempt at identification. The other seven do fall into three groups as regards size. They are here taken as possibly corresponding with the three named species based on lower jaws. These references are, however, quite uncertain. The skull and upper tooth specimens are not included in specific hypodigms (except *Mioeuoticus bishopi*) or definitely referred to species. They are here described as cf. *Progalago dora*, etc.

Pteropus Brisson, 1762

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15. PTEROPVS.

Dentes Primores IV vtrinque,

Palmae corpore longiores, pollicibus brevissimis, membrana inter digitos expansa et ad crura postica extensa, pro volatu.

Mammae duae pectorales.

Pteropus comorensis Nicoll, 1908

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Pteropus comorensis.

On one occasion we visited a small island situated in the middle of the harbour. This island, which is marked on the charts as "Buzi," was thickly covered with vegetation, and in some places it was impossible to force a way through the prickly clumps of acacia trees. The heat was intense, and I have never felt the sun so much as I did on Buzi.

...

This little island was tenanted by a large colony of fruit bats.‡ They were, however, in the thick trees on the inaccessible side of the island, where it rises straight up from the sea. Every evening numbers of them flew across to the main island,

‡ *Pteropus comorensis*

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and one evening, on our way back to the ship after a day's collecting in the mangrove swamps, we shot one as it flew over the steam launch. Its fur was of a reddish colour, thick and soft. A most unpleasant skunk-like smell clings to one's hands after touching one of these bats, and this is especially noticeable when the animal has just been shot. On several occasions I saw these fruit bats splash into the salt-water of the harbour, but whether they were drinking or washing I do not know.

[The first paragraph mentioned above doesn't actually belong to the description, but was separated from it by a paragraph on swifts (*Cypselus mayottensis*). However, I did include it since it refers to "this little island" in the first sentence of the description, which might indicate that the type locality can be restricted to "Mayotte: Buzi Island". Furthermore, the name *Pteropus comorensis* was mentioned as a footnote, hence the use of the "‡" symbol.]

- Victor Van Cakenberghe, December 2009.

Pteropus dupréanus Schlegel, 1867

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Pteropus dupréanus, n. sp.

Allied to *P. stramineus*

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from Sennaar and Senegambia; but tail longer (7 lines), and the general colour of fur greyish yellow.

Pteropus fuscus E. Geoffroy Saint-Hilaire, 1803

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1. Esp. La roussette brune. *Pteropus fuscus*. (Geoff.).

Catact. Pelage brun, des poils roux sur la face, autour de l'anus, et sur les parties latérales du dos.

Canis volans ternatatus orientalis, Seba, Mus. I, p. 91, t. 57, Fig. 1, 2.

Vespertilio vampyrus, Lin. Gml. p. 45.

Chien volant Daub., Mém. de l'Acad. 1759, p. 384.

Roussette, Buff. Daub. t. 10, p. 55 - 66, pl. 14.

Descript. T. 0,^m22 (8 p. 2 l.). Envergure, 0,^m88 (2 pi.

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8 p. 1 l.). Incisives supérieures séparées entr'elles presque également, les latérales plus courtes; point de queue; corps brun-noirâtre, différentes teintes de roux-jaunâtre s'étendant de chaque côté du dos et des lombes, depuis les épaules jusqu'à la croupe.

Patrie. Les Indes.

N°. LXXXVI. Individu vu en dessous, les ailes étendues, envoyé de l'île Bourbon, par M. de la Nux, conseiller au ci-devant conseil royal de cette île.

N°. LXXXVII. Autre individu vu dans la même position, mais différant un peu du précédent par quelques teintes d'un jaune plus clair sur la tête, envoyé de l'île de Bourbon par M. de la Nux.

N°. LXXXVIII. Autre individu envoyé de Madagascar, par le citoyen Macé, naturaliste.

N°. LXXXIX. Foetus conservés dans l'esprit de vin; ils sont nus et blanchâtres.

Pteropus Gambianus Ogilby, 1835

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Two undescribed *Pteropi*, brought over by Mr. Rendall, present some modifications of dentition which have not been observed in other species, and which appear to indicate a subgenus, probably representing the common Asiatic forms on this coast of Africa. These animals have the incisors and canines of the same form and number as the rest of the genus, but there are only three molars in the upper and five in the lower jaw. The incisors are small and regular, the canines of intermediate size; the first false molar in the lower jaw is small and of the normal form, but the second in this jaw and the first in the upper are of the same form as the canines, and very little inferior to them in size, so that when the mouth is opened there appear to be four canines in each jaw; next follows in either jaw a tooth with a large fang upon the outer edge and a smaller one within, which is of intermediate form between the true and false molars; after which come two normal molars in the lower and one in the upper jaw. All the molars are separated from one another by a vacant space on each side; this gap is particularly large between the real and spurious canines or first false molars in the upper jaw, the corresponding space in the lower having, in its centre, the small false molar already mentioned.

Pteropus Gambianus.

Length from the nose to the centre between the thighs: $6\frac{3}{4}$ in.

Length of the head from the nose to the root of the ear: $1\frac{3}{4}$

Expanse of the wings: 1 f 8 in.

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The fur is of a very soft woolly texture, and of a uniform reddish mouse colour over every part, only rather lighter on the sides of the neck and belly than on the superior surface of the body. The wings are ample, naked except upon the thighs and arms, and of a light brown colour; there is no real interfemoral membrane; but the whole posterior face of the thighs and body is margined with a narrow band of integument about half an inch broad, and covered above with the same description of hair as the back. The ears are small, naked, erect and elliptical, and the eyes placed much nearer to them, and consequently at a greater comparative distance from the muzzle, than in the ordinary *Pteropi*

Pteropus Haldemani Hallowell, 1846

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Description. - General expression ferocious; head resembling that of a dog; ears of moderate size, smooth for the most part, obtuse at the tip, hairy at base externally; there is no tragus; body dark brown above; neck, occiput and vertex same colour, but lighter than upon the back; wings and interfemoral membrane of a sienna brown colour above and below; thorax and upper part of abdomen and sides brown; the rest of the abdomen is white; there are two long and thin hairs upon the muzzle; lips full, nostrils prominent, their margins being surrounded by a fold of the skin; eyes rather large, irides - ; wings long; that portion of the membrane included between the phalanges naked, the remainder more or less hairy above and below; upper surface of the interfemoral membrane hairy, with the exception of a small part at its posterior extremity which is naked; under surface also hairy, but much less so than upper; no tail; tibia and fibula included within the membranes; four slender toes, compressed, of nearly equal length, the outer one being a little shorter than the others; they are sparsely furnished with thin hairs, varying in length; the terminal phalanx of each is provided with a robust, sharp and incurvated nail. The index finger like the thumb is also furnished with a short and incurvated nail.

Measurements. Inches.

Total length: $3\frac{1}{2}$

Length of head: $1\frac{3}{4}$

Distance between anterior margin of nostril and anterior canthus of eye: $\frac{5}{8}\frac{1}{10}$

Distance between angle of mouth and anterior canthus of eye: $\frac{3}{8}$

Length of neck, body and tail: 3

Length of forearm; 3

Length of tibia: $1\frac{1}{4}$

Spread: $14\frac{1}{4}$

Length of thumb: $\frac{3}{4}$

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Incisors. $2\cdot 2/2\cdot 2$ Canines. $1\cdot 1/1\cdot 1$ False Molars. $1\cdot 1/2\cdot 2$ Molars. $2\cdot 2/3\cdot 3$

This species I have named after my esteemed friend, S. S. Haldeman, Esq., author of the N. American Limniades, who obtained it with other African animals from Dr. Goheen, Physician to the American Colonization Society.

Pteropus hottentottus Temminck, 1832

p. 3

Primum hoc animal idem visum est, ac P. AMPLEX-CAUDATUS GEOFFR.,

p. 4

qui in insulis Timor, Amboina, ac Sumatra occurrit; e description tamen aliquanto fusiore atque accuratior contrarium mox elucebit.

Jam magnitudine utrumque animal insigniter differt: etenim PT. HOTTENTOTTUS quinquies major est, oculi magis superiora versus collocata, longius a naso, quam ab auribus distant: membrana ejusdem conformationis, atque illa AMPLEXI-CAUDATI; in parte tamen introrsum versa frequentibus obsita est pilis.

Basis membranae interfemoralis ad latus internum pilosa. Ad basin caudae perbrevis cernitur incisura, figuram inversae V prae se ferens. Juxta hanc cauda, quae plane est libera, exoritur. Cauda cum corpore comparata, habet tantummodo longitudinem dimidiam intervalli, quod est inde ab anteriore oculi margine, usque ad nasi apicem. Illa autem AMPLEXI-CAUDATI est longior; quippe quae totâ ejus intervalli longitudine gaudeat. Dentes incisores sunt perparvi, uniformiter seriati, sibique approximati; dentes autem molares aequali intervallo a se invicem distant. Aures breves et rotundatae. - Rostrum longum ac compressum.

Corpus haud crassum, vestitur pilis brevibus, mollibus ac laevibus, qui per superiorem ejus partem duplicem offerunt colorem; ad basin nitide griseum, ad apicem vero fuscum; qui etiam in speciminibus masculis praevalet: in inferioribus partibus uterque sexus coloris est subgrisei.

Denique haec species praeterea notas gravioris momenti nonnullas offert; quales sunt forma membranae interfemoralis ad Coxygem; cauda libera ac brevis, et quidquid praeterea a nobis jam indicatum est: quae omnia sane sufficiunt, ut primo adspectu PT. HOTTENTOTTUS TEMM. ab AMPLEXI-CAUDATO distinguatur.

Mensurae,

Universa longitudo in speciminibus adultis 0,13.

Extensio alarum (1) 0,53.

Longitudo antibrachii 0,09.

Caudae 0,02.

Excisura membranae interfemoralis 0,25.

(1) Occurrunt specimina quaedam, in quibus alarura extensio quoque major est.

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Habitat haec species circa Urbem Capensem; reperitur quoque in plagis longius remotis. Complura ejus specimina, quae tum pellibus, tum sceletis constant, conservantur in splendidissima collectione, quam Museum Lugduno-Batavum continet.

Pteropus labiatus Temminck, 1837

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PTEROPUS LABIATUS.

ROUSSETTE LABIAIRE.

Portrait du mâle Pl. 39 fig. 1, 2 la femelle 3.

Cette belle roussette, dont le mâle est remarquable par la longueur extrême des pinceaux onctueux et par le prolongement labial au deux mâchoires, est de la taille de notre *vespertilion murin* d'Europe. Son museau est long; ses dents incisives sont fines et entassées; un appendice membraneux de peu de largeur sert d'interfémorale, et quoique entourant le coccyx par un rudiment elle n'est point visible sur cette partie, étant cachée et totalement couverte en dessus comme en dessous par les poils qui revêtent une grande portion de cette membrane adhérente au tibia: membranes des ailes naissant immédiatement des flancs. Oreilles longues et pointues.

Pelage cotonneux sur toutes les parties du corps, particulièrement sur le dos, moins, ou plus lisse aux parties inférieures.

Le mâle, porte tout le long du bord des deux mâchoires un prolongement de la partie labiale qui tombe de plusieurs lignes au-delà du bord inférieur et cache totalement l'ouverture ou la fente latérale de la bouche, à-peu-près de la même manière comme dans le chien dogue de forte race; ce singulier appareil, propre aux deux mâchoires, donne à la tête du mâle de cette espèce une physionomie toute particulière. Le pelage cotonneux des parties supérieures revêt aussi toute la région humérale et le bord de la membrane alaire adhérente aux flancs; ce pelage ainsi que celui de la tête qui est court et ras, porte une teinte isabelle roussâtre, mais d'un ton plus roux vers le dos et surtout à la croupe; insertion des oreilles et bord postérieur de celles-ci couvertes de poils blancs; de chaque côté du cou qui est d'un brun roussâtre, naît une ample touffe de longs poils d'un blanc pur; ces larges pinceaux qui recouvrent probablement un appareil onctueux forment deux panaches dont les poils sont divergent d'un centre commun. La poitrine, la région humérale, les flancs et la région du coccyx sont

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roux clair; le milieu du ventre où les poils sont assez courts et lisses est d'un blanc terne. Toutes les membranes ont une teinte couleur feuille-morte.

La femelle dont nous donnons la tête vue de profil, pl. 39 fig. 3, ne diffère point remarquablement par la coloration du pelage, mais elle manque de tout vestige d'appendice labial; et n'a point de pinceaux ou de poils divergens aux côtés du cou.

Longueur totale 4 pouces 1 ou 2 lignes; envergure 15 pouces; antibrachium 2 pouces 4 lignes.

Patrie: M. Botta a déposé au Musée de Paris deux individus de cette belle et nouvelle espèce, découverte dans son voyage en Abyssinie. Les sujets du musée des Pays-Bas ont été acquis, l'un à Londres sans indication de patrie, l'autre à Paris, provenant des collections de Mr. Botta.

Pteropus Leachii A. Smith, 1829

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Pteropus Leachii, mihi.

P. suprà fusco-cinereus, infrà sordido-cinereus, caudâ liberâ.

Colour above a sort of brownish gray, beneath a dirty pale smoke gray; incisors short, strong, regular, and rounded at tips; head long; ears of moderate length and rounded at the apices; membranes blackish, interfemoral one only edging the inner side of each hinder extremity; tail free. Length from forehead to root of tail, four inches; length of tail, three-quarters of an inch; expanse of wings, thirteen inches. Found abundantly in the gardens about Cape Town during the fruit season, and often proves very destructive to vineyards in the night. A larger species of the same genus inhabits the interior, but I have not yet been able to obtain a perfect specimen. Named after the celebrated naturalist Dr. Leach, F.R.S.

Pteropus macrocephalus Ogilby, 1835

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The whole length of this species is barely 6 inches, the length of the head 2 inches, and the expanse of the wings about 1 foot 3 inches. The colour, form and appearance are much the same as in the last species, but the *Pter. macrocephalus* is at once distinguished by the great size of the head, as well as by the colour of the flying membranes which are very dark brown, nearly approaching to black. The canine teeth also, as well as the head, are of much larger size, and the interfemoral margin is narrower. Dr. Horsfield, from the great length of the head, thinks that this species may approximate to the *Macroglossus* of M. F. Cuvier, the *Pter. rostratus* of his own 'Zoological Recsearches in Java.' It is to be observed, however, that it differs in denritation from that animal, as well as from all other *Pteropi* hitherto described; and, with the *Pter. Gambianus*, may furnish the type of a new geuns to those who regard such modifications as amounting to generic characters. Mr. Rendall's collection contains numerous specimens of both the species here described

Pteropus mascarinus Mason, 1907

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Pteropus mascarinus, sp. n.

Skull. - With moderately long but heavy muzzle, flattened and scarcely concave frontal region. Sagittal crest weakly developed, almost obsolete.

Measurements. - A few of the measurements can only be given approximately owing to the damaged condition of the specimen: -

Upper length (approximate) 49 mm.; condylo-basal length (approximate) 46; basilar length (approximate) 44; median palate length 28; breadth between outer sides of canines 9.8, inside canines 4.3 ; outside p.4 13.8, inside p. 4.8; outside

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m.2 13, inside m.2 9.2; zygomatic breadth (approximate) 27; least interorbital breadth 7.8; the dimensions of the structure behind postorbital process cannot be taken; greatest breadth of brain-case 15; occipital depth 11; mandible 40.6; maxillary tooth-row exclusive of incisors (alveoli) 20.1; mandibular tooth-row exclusive of incisors (alveoli) 22.3.

Teeth. - With the longitudinal grooving characteristic of the genus *Pteropus*, very strongly developed, somewhat large and heavy for the size of the animal; canines long and sharp, both above and below, with unusually acute and prominent postero-internal basal ledges, those of the maxillary in particular; first upper premolars very minute, but would have been still persistent in the type and standing in the tooth-row.

Measurements. - The sizes of the upper incisors and second upper molars are omitted, as these teeth are wanting in the specimen. Vertical length of upper canine 7.5 mm.; horizontal length of upper canine 3.5; horizontal length of p.3 3.8, width 2.3; horizontal length of p.4 3.8, width 3; horizontal length of m.1 4.1, width 2.1. Lower teeth - combined breadth of incisors - ?; height of canine (from basal ledge behind) 5 ; horizontal length of anterior premolar 2; of p.3 3.8, width 2.1; horizontal length of p.4 3.5 ; of m.1 4.3, width 2; horizontal length of penultimate molar 3, width 1.8; horizontal length of m.3 1.5, width 1.4.

Locality. - Round Island, North-east Mauritius.

As compared with the now existing *Pteropi* of the Mascarenes, this species occupies a place intermediate between *Pteropus vampyrus* and *Pt. rodricensis*, in size only, the dentition being typical of *Pteropus*, whereas the two above species fall into the subgenus *Spectrum*. The only other fruit-bat occurring in the Mascarenes is *Pt. rubricollis*, and this has been placed by Matschie in his subgenus *Sericonycteris*. A great analogy, in fact, exists between the dentition of *Pt. mascarinus* and those species of *Pteropus* (*tonganus*, *Gouldii*, and *conspicillatus*) inhabiting Australia and the islands of the Pacific Ocean.

The limb-bones call for no special description, the most perfect specimen being the right femur, which measures 40 mm.

Since the above was written a few additional bones referable to this new species, and representing two younger individuals, have been received. Their fragmentary condition has, however, failed to add any additional particulars to the above description of this interesting and lost species.

Pteropus mollipilosus H. Allen, 1862

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Head small; ears large. Envergure great. Interfemoral membrane scanty. Tail very small, free. General hue olive brown: brighter upon the back of the neck, where a faint dusky-brown line is seen traversing the thick fur of this region. Antero-posteriorly a patch of the same hue lies upon each shoulder; that upon the back is thinner and darker. The interfemoral membrane is well furnished with soft olive brown hair, which becomes more scanty and shorter as it runs down the legs and the back of the feet to the claws. Back of the humerus and ulna thinly covered with the hair of the prevailing hue; anterior part of neck russet; belly of the same prevailing color as the back, with an inclination to dusky red upon the thighs.

* This disposition of the superior incisors is peculiar. We know that these teeth frequently vary; as, for instance, in the genus *Hypoderma*, where the young animal has $i \frac{4}{4}$ and later in life $\frac{2}{2}$; the elderly animal possessing but $\frac{2}{0}$. Thus, the difference in the number of the incisors is not of the importance which might at first sight be supposed. It is interesting, however, to know that this is the only instance of incisorial variation as yet seen among the species of *Epomophorus*.

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Fine straggling hairs clothe the inferior surface of the ulna and humerus, and stretch out upon the membrane down along the sides of the body and upwards upon the interbrachial membrane. Everywhere the hair is very soft.

The skull is of medium size, not crested. The postorbital processes are long and incline outward and downward to within two lines of the zygomas. The intermaxillary bone is not thrown forward and downward as in other *Pteropi*, but is on the same level as the roof of the mouth. The first premolar above is larger than the incisors. The same tooth on the lower jaw is also of good size, but less tubercular than in other species.

The dimensions are as follows:

Length from nose to base of tail: 8.0
 " " " commencement of membrane: 3.0
 " of head: 2.3
 " from nose to eye: 1.0
 " " eye to ears: 0.6
 Between eyes: 0.9
 " ears: 1.3
 Height of ears: 0.10
 Length of tail: 0.3
 " superior extremity: 6.6
 " thumb: $1.7\frac{1}{2}$
 " first joint: $0.4\frac{1}{2}$
 " second joint: 1.3
 " index finger: 3.0
 " third " : 8.0
 " fourth " : 6.5
 " fifth " : 5.0
 Envergure: 26.0

It will be observed that the length of the inferior extremity and the expanse of wing membrane are greater than usual, when the other proportions are taken into consideration.

Hab. Western Africa; discovered by M. DuChaillu.

Pteropus rodricensis Dobson, 1878

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Ears slightly longer than the muzzle, but projecting by their upper third only from the long fur surrounding them; ear-conch moderately broad in lower three fourths, very abruptly narrowed above by flattening of the upper fourth of the inner margin and a deep concavity of the corresponding part of the outer margin, terminating in a narrow acute tip; more than two thirds of the concave surface of the conch is well clothed with moderately long hairs, the upper fourth and the back of the ear naked or with a few short hairs only (Plate III. Fig. 1).

Fur long and dense, in quality intermediate between that of *Pt. Edwardsii* and *Pt. Vulgaris*; that covering the back of the head, neck, and shoulders very long, on the back shorter but not appressed, the hairs directed backwards, extending thickly upon the wings almost as far outwards as a line drawn from the proximal third of the

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humerus to the knee, upon the femur, knee, and adjoining wing-membrane along the proximal third of the tibiae, about two thirds of which are thickly covered with straight hairs directed backwards, their distal thirds being nearly naked as well as the feet; and, although the fur of the back extends upon the interfemoral membrane, it does not conceal its posterior margin, and its lower third, supported by the calcanea, is naked. Beneath, the antebrachial membrane is covered with long thinly-spread hairs, and similar hairs clothe the wing-membrane between the humerus and femur, and extend outwards in a broad band behind the forearm. Face reddish brown, with a few greyish or shining hairs: chin and throat darker brown; top of the head and nape reddish or yellowish brown, passing into a band of bright yellow, which extends across the back of the neck from shoulder to shoulder, and downwards on the sides of the neck and thorax, limited by a longitudinal band of dark fur passing backwards from the chin along the thorax to the abdomen, which is clothed with dark brown fur, of which some hairs have greyish or shining extremities; fur of the back behind the shoulders dark brown, the extremities of the hairs more or less yellowish. In the male the extremities of the hairs on the head and back are much brighter-coloured than in the female.

First upper premolar deciduous; the second upper premolar and the second and third lower, also the first and second upper molars and the first lower, have each a small but distinct posterior basal cusp; even the small first lower premolar has an indication of a posterior basal cusp; last upper molar very small and circular, not as large as the first lower premolar; last lower molar slightly larger than the first lower premolar.

Length (of an adult M preserved in alcohol), head and body 7", head 2".2, ear 0".9, ear from tip of nostril 2", eye from tip of nostril 0".8, forearm 4".9; thumb-metacarp. 0".35, ph. And claw 1".6; third finger-metacarp. 3".3, 1st ph. 2".4, 2nd ph. 3".55; fourth finger-metacarp. 3".25, 1st ph. 2", 2nd ph. 1".85; fifth finger-metacarp. 3".35, 1st ph. 1".5, 2nd ph. 1".45; tibia 2".25, calcaneum 0".6, foot 1".15.

Hab. Island of Rodriguez.

A-d. M & F juv. Et ad., al. Rodriguez. Royal Society.

E. F ad. Rodriguez. Royal Society.

F. Ad. Sk. Rodriguez. Royal Society.

G. Skeleton. Rodriguez. Royal Society.

Pteropus ruber E. Geoffroy Saint-Hilaire, 1803

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3. Esp. La roussette à cou rouge. *Pteropus ruber*. (Geoff.)

Caract. Pelage gris, dessus du cou rouge.

Pteropus fuscus, Briss.

Roussette, Daub. Mém. de l'Acad. 1759, p. 385.

Rougette, Buff. t. 10, p. 55, pl. 17.

Descript. T. 0,^m17 (6 p. 4 l.). Envergure 0,^m66 (2 pi. 1 p.), incisives rapprochées, les inférieures par paire; point de membrane interfémorale; pelage entièrement de couleur centrée mêlée de brun; dessus du cou garni de poils roux-vifs, nués d'orangers; poils longs et un peu crépus.

Patrie. Les Indes Orientales.

N°. XCI. Individu envoyé de Madagascar par le citoyen Macé.

Pteropus rufus E. Geoffroy Saint-Hilaire, 1803

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2. Esp. La roussette marron. *Pteropus rufus*. (Geoff.)

Caract. Pelage roux, le dos, depuis les épaules, de couleur marron-foncée.

The great bat from Madagascar. Edwards, Aves, t. 4, pl. 180.

Descript. T. 0,^m29 (10 p. 7 l.). Envergure 0,^m86 (2 pi. 6 p.), incisives supérieures intermédiaires plus grandes et plus rapprochées entr'elles que les latérales, point de queue; dessus du corps roux, beaucoup plus clair sur la tête; les épaules et le cou bruns en dessous; museau noir.

Patrie. Madagascar.

N°. XC. Individu envoyé par le citoyen Macé, naturaliste.

Pteropus rufus princeps K. Andersen, 1908

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Pteropus rufus princeps, subsp. n.

Similar to *Pt. rufus rufus* (*Pt. edwardsi* auct.), but skull and external dimensions conspicuously larger. Total length of skull 77 mm. (69 - 73.8 in the typical form of the species); mandible 62 (54.5 - 58.2); forearm 170.5 (158.5 - 165.5).

Type. M ad. Alc. And skull, Fort Dauphin, S.E. Madagascar; collected by M. Cloisel ; B.M. 91.11.30.10.

Remarks. - The typical smaller form of the species is apparently confined to the northern and central part of Madagascar.

Pteropus Schoënsis Rüppell, 1842

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Pteropus schoënsis (Rüppell).

Diagn. *Pteropus ecaudatus*, auriculis brevibus, halluce elongato, corporis colore cervino, regione epigastrica canescente; macula albicante ante et post auriculas, ad latera faciei a naribus per oculos fascia umbrina; unguibus nigris.

Ich hatte Zweifel, ob dieses Thier nicht etwa das Jugendalter von *Pteropus Whitei* Bennet, (*Pt. labiatus* Botta) sey. Die wenig abgenutzten Doppelspitzen der Schneidezähne des einzigen von mir untersuchten Thieres deuten auf ein junges Individuum hin. Aber bei genauer Vergleichung ergab es sich, dass bei der hier zu beschreibenden Art das Daumenglied bei weitem länger, und die Ohren vergleichlich viel kürzer sind als bei dem Bennet'schen *Pteropus*. Ich bedauere, dass die Untersuchung der Schädel beider Thiere nicht thunlich ist, um jegliche Ungewissheit über die Artenverschiedenheit zu lösen. Die Körperausmessung gab folgendes Resultat:

Zoll. Lin.

Ganze Körperlänge von der Nasenspitze bis zum Mittelrand der sehr kurzen Interfemural-Membran: 8 9

Flugweite: 18 -

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Vorderarmknochen: 1 11

Daumenglied, ohne Nagel: - 6 $\frac{1}{2}$ Verticalhöhe des Ohrs: - 4 $\frac{1}{2}$

Die Grundfarbe der ganzen Behaarung ist rehfarbig, welches auf dem Bauch und der Kopfmitte ins Hellgraue, und auf dem Rücken mehr ins Röthliche ober geht; von den Nasenlöchern aus verläuft durch die Augen nach dem Ohre zu ein kaffeebrauner Streifen; oberhalb und unter der Basis des Ohrs ist ein Büschel grauweißer Haare. Auf der innern Seite der Flughaut ist die Behaarung längs des Körpers und an dem Vorderarm hellgrau, und auf der Rückenseite der Flughaut rothbraun. Die robusten, etwas comprimierten Nägel sämtlicher Zehen sind schwarzbraun.

Das hier beschriebene Exemplar scheint weiblichen Geschlechts zu seyn; es ward mir aus dem Reiche Schoa, südlich von Abyssinien, zugeschickt.

[*Pteropus stramineus*](#) E. Geoffroy Saint-Hilaire, 1803

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4. Esp. La roussette jaune. *Pteropus stramineus*. (Geoffr.)

Caract. Pelage jaunâtre, une petit queue.

Seba. Thes. I, p. 91, tab. 57, fig. 1, 2.

Pennant, p. 362, pl. 31, fig. 91.

Descript. T. 0,^m15 (5 p. 6 l.). Envergure 0,^m64 (1 pi. 11 p.), les incisives intermédiaires très-écartées, les inférieures contiguës, égales; la membrane interfémorale cessant au point d'insertion de la queue qui est longue de 0,^m010 (6 l.).

Patrie. Inconnue.

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N°. XCII et XCIII. Deux individus donnés au cabinet par le professeur Fourcroy.

N°. XCIV. Cet autre individu vient de la collection du Stathouder; c'est sans doute l'original qui a servi à la description et à la mauvaise figure qu'en a donnée Albert Seba.

[*Pteropus Wahlbergi*](#) Sundevall, 1846

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molliter rufescenti-villosus, auriculis oblongis, ad basim, utrimque, maculâ albo-villosa. - Patagium fuscum, ad corpus late denseque villosum: caudale totum cum pedibus posticis, et brachia supra subtusque Villosissima. Cauda parva, *apice sub patagio libera*, prominula. Pili laterales colli, in MF, radiantes; M præterea fasciculo albopiloso ante humeros ornatus. Prope Port-Natar et in Caffraria interiore occisus: M +/- 50 millim. Caput 60. Cubitus 85.

[*Pteropus Whitei*](#) Bennett, 1836

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Pter. pallidè brunneus, posticè pallidior; ventre albido; scopâ humerali albâ magnâ.

Long. tot. 6 ³/₄ poll.; *capitis*, 2 ¹/₄; expansio alarum, 12.

Hab. in regione Gambiensi, *D. Rendall*.

OBS. *Scopa humeralis* forsan maribus propria.

In naming this remarkable *Bat* in commemoration of an individual who had no share in its discovery and by whom it was never seen, I may seem, strictly speaking, to have erred; but it is time that technical zoology should record the name of one who was by

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no means unversed in its mysteries, and who has long maintained the highest rank as a popular zoologist. Gilbert White, of Selborne, was the first Englishman who ascertained the existence of indigenous *Bats* other than the two known as European to Linnaeus: to that good man and excellent observer be this singular species dedicated, - a species belonging to one of the few groups of *Bats* that are of direct utility to the human race, and which, preying not on other animals, rest contented with the simple fruits of their native woods.

[*Pterygistes azureum*](#) Thomas, 1901

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Very similar to the other species, which scarcely differ among themselves except in size, but smaller than any of them.

General characters, so far as can be made out on the single specimen, as in *P. Leisleri*, to which this form was assigned by Drouet and Peters. But the size is markedly less, the general build is lighter, the ears appear to be rather broader and more rounded, while the tragus is slightly narrower and more pointed.

Colour, in a spirit-specimen, apparently of the usual dark brown characteristic of *Pterygistes*. Hairs of back about 6 millim. in length.

Skull, as compared with that of *P. Leisleri*, smaller and more delicate throughout, smooth, unridged, no sagittal and scarcely any lambdoid crests perceptible in an old male. Teeth practically identical throughout with those of *P. Leisleri*.

Dimensions of the type (an old male, in spirit): -

Forearm 37 millim.

Head and body 54; tail 42; ear 12; tragus, length on inner edge 3, breadth 2; third finger, metacarpus 36.5, first phalanx 15, second phalanx 10.5; fifth finger, metacarpal 31,

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first phalanx 8, second phalanx 4.7; lower leg 17; hind foot (s. u.) 7.7.

Skull: greatest length 14.2; basipalatal length 11; zygomatic breadth 9.4; posterior breadth 9; breadth between orbits 6.4; constriction 4.4; length of palate 5; breadth between outer borders of m^2 6.2; front of canine to back of m^3 5.1.

Hab. St. Michael's, Azores.

Type. Male. B.M. no. 65.10.2.1. Collected and presented by F. DuCane Godman, Esq. One specimen only.

The single specimen has all its claws, both of feet and thumb, worn down quite blunt. Whether this is accidental or due to the nature of its habitat remains to be seen.

From the gigantic *P. maximus* and *lasiopterus*, through *P. noctula*, *stenopterus*, and *Leisleri*, there is a regular gradation in size down to this small Azorean representative of the group, all of them differing from each other in little else but size and the correlated development of the cranial ridges.

I accept with some hesitation the generic separation of the *noctula* group from *Pipistrellus*, advocated by Gray, Jerdon, H. Allen, Miller, and Mehely, on the ground that not only is the general build of the body different, but that the wings even are different in shape, owing to the reduction in length of the fifth digit. At the same time it must be admitted that some of the Indian species of *Pipistrellus*, such as *P. ceylonicus*, *chrysothrix*, *mordax*, &c., approximate to a certain extent to *Pterygistes*, and render the distinction less marked than it appears to naturalists who have only compared with each other such extreme forms as *Pterygistes noctula* and *Pipistrellus pipistrellus*.

[*Pterygistes madeiræ*](#) Barrett-Hamilton, 1906

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Pterygistes madeiræ, sp. n.

This bat is similar to *P. Leisleri*, Kuhl, of Britain, but perhaps smaller and with quite different skull. The colour is

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unascertainable in the old spirit-specimens, which alone are available for examination.

The *skull*, as compared with that of *P. Leisleri*, is slightly smaller, less massive, and relatively longer and narrower, characters which, although quite apparent to the eye, do not stand out so clearly when expressed in words. It is not at all like the much smaller *P. azureum* Thomas, with its somewhat inflated cranium and depressed nasal region.

The *dimensions* in millimetres of a female (in spirit) are as follows: -

Head and body 55; ear 12; tragus 4; greatest expanse of wing 240; thumb and claw 6; longest digit 74; basal joint of fifth digit 32; basal joint of second digit 40; forearm 43; tibia 16; hind foot (without claws) 7.5.

Skull (of the type): greatest length 15; basipalatal length 11.5; zygomatic breadth 9.5; posterior breadth 9; breadth between orbits 8, at constriction 4.5; length of palate 6.5; breadth between outer borders of m^2 6.75; length from anterior border of canine to posterior border of m^3 5.25.

Hab. Madeira.

This species is evidently the representative of *P. Leisleri* in Madeira, and it is remarkable how different it is from its neighbour *P. azureum* of the Azores. It is described from specimens in the R. F. Tomes collection procured by Mason. According to the custom of his day, Tomes identified his specimens with *P. Leisleri* from which species, however, it is clearly differentiated by its characteristic skull.

Thetype is no. 1 of the Tomes collection in the British Museum (a female).

Rhinopoma cystops macinnesi Hayman, 1937

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A dwarfed form of the well-known species of Egypt and the Sudan, having a shorter forearm and a much lighter and smaller skull.

Type. - Brit. Mus. no. 36.11.4.45. Adult male in alcohol, collected on Bat Island, near Central Island, Lake Rudolf, April 24th, 1934. Another male and a female examined taken at the same time and place.

Description of Type. - Externally similar to true *cystops*, but having a forearm measurement of only 46 mm. (47 mm. in the other two specimens). In adult *cystops* the forearm averages 52, sometimes reaching 55. The colour of the fur is similar to that of the typical form, a pale plumbeous-fawn, paler at the base.

The skull is notably smaller and weaker than that of *cystops*; the sagittal crest is very weak, being barely indicated, while in *cystops* it is strongly developed in both sexes. The rostrum is narrower, the bullæ smaller, and the reduction in the size of the teeth is illustrated by the breadth of m^2 in *macinnesi* being 1.8, while in the type and another specimen of adult *cystops* the figures are 2.2 and 2.3 respectively. Compared with the skulls of *Rhinopoma pusillum* Thos., from Persia, and *muscatellum* Thos., from Muscat, two species of approximately similar external dimensions, the skull of this new form is quite distinct, being smaller and narrower, with smaller teeth. In the following table the measurements of three specimens of *macinnesi* are compared with those of the types of *cystops*, *pusillum*, and *muscatellum*: -

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[table removed - eds.]

This discovery extends the known range of the genus into Uganda, and I have pleasure in associating with it the name of Mr. D.G. MacInnes, to whom much of the success of the mammal-collecting was due.

Rhinolophus abæ J.A. Allen, 1917

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Type, No. 49113, F ad. (skin and skull), Aba, Uele district, Belgian Congo, Dec. 15, 1911; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No. 1756.

General coloration drab-brown, a little darker above than below, the middle of the back slightly darker (blackish brown) than the rest of the upperparts, sides of the shoulders a little lighter. Fur almost uniformly colored from base to tip, the median zone being only slightly lighter than the base and tips.

Ears long and pointed, the anterior border slightly convex (increasingly so apically), the posterior nearly straight (slightly concave toward the apex), the antitragus large and high (vertical depth anteriorly 5-6 mm.). Breadth of horseshoe in skins (when not unduly shrunken), 8.5-9.5 mm. Nose-leaf broad at base (about 6.5 mm.), acutely pointed, and about 6 mm. in length. Sella about 3 mm. broad at base and 4 in height (as nearly as can be judged in softened skins), slightly concave on anterior face, with a posterior extension of about 3 mm., highest in front with a slight median axial depression.

Type, collectors' measurements: Total length, 85; head and body, 63; tail, 24; foot, 12; ear, 24. Forearm (from skin), 52.3; tibia, 20; foot, 11.

Skull (type), total length, 22.9; zygomatic breadth, 11.9; breadth at canines, 5.8; maxillary breadth, 8.1; upper toothrow (front of $c-m^3$), 7.8; lower jaw (incisors to condyle), 15.1; lower toothrow ($c-m^3$), 8.8.

Represented by ten specimens, skins and skulls (2 males, 8 females) all collected at Aba, December 15-17, 1911. Collectors' measurements (1 male, 7 females, all adult):

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Total length: 85 (80 - 89); Head and body: 61 (57 - 64); Tail: 24 (22 - 27); Foot: 12 (11 - 13); Ear: 22.9 (22 - 24)

Forearm, same specimens, 53 (50-54). Skull (6 specimens), total length, 23 (21.9-23.6); zygomatic breadth, 11.7 (11.3-12.2); breadth at base of canines (9 specimens), 5.9 (5.6-0.2); maxillar breadth (across front corner of m^3), 8.3 (7.8-8.6).

P^2 is present on both sides in 3 skulls, on one side in 2 skulls, and absent on both sides in 4 skulls.

The series is very uniform in coloration; an immature male differs from the others in being lighter and grayer, and is somewhat smaller.

This form, appears to be a northern representative of the *Rhinolophus auger* group, of which several forms have been described from the Zambesi region and southward by K. Andersen.

Rhinolophus acrotis Heuglin, 1861

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Rh. medius, digitis valde elongatis, alis ad apicem caudae usque porrectis; auriculis mediis, acutis, margine externo S-forme, lobulo lato accessario obtuse marginato instructis: - ferro equino duplice margine superioris libero, medio antico-inciso, margine (ferri equini) inferioris labiis connato; glandulis violaceis circumdato; latere sellae anteriore medium versus angustato, basi concava, plicis duabus ad nares usque ductis; apice vix rectangulare; prosthema latiore, lanceolato, acuminato, parte anteriore plicis duabus transversalibus erectis ornato; phalangis exterioribus quarti et quinti digiti apice paullo bifurcatis, digito secundo phalangis duabus, tertio, quarto et quinto phalangis tribus instructis. Vellere pallide murino, subtus pallidior. Long. a naso ad basin caudae 2" - extensio alarum 13" - caud. 1" 3" - auriculis 9 $\frac{1}{4}$ " - antibrach. 1" 9".

Gleicht in Rücksicht auf Construction des Nasenbesatzes, Form der Ohren

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u. s. w. sehr der *Rh. minimus*, Heugl., ist aber viel grösser, die Ohren verhältnissmässig kleiner und spitziger, Nasenblatt doppelt, jedoch nur der Rand des oberen frei, die *Sella* mit breiter Vorderhand, die Endglieder des vierten und fünften Fingers an der Spitze gegabelt. Zeigfinger zweigliederig. Im Ohr 6 deutlichere und 1 - 2 minder hervortretende Querfalten. Zahnsystem - so weit dies ohne Dissection beobachtet werden kann - regelmässig; untere Schneidezähne je 2 auf jeder Kieferseite, nicht sehr klein, am schaufelförmigen Rande der Schneide dreizackig.

Nasenblatt, wie gesagt, doppelt, das untere mit einer regelmässigen Reihe von violetten Drüsen umgeben. Der Körper der *Sella* seitlich zusammengedrückt, die höchste Spitze ziemlich erhaben über die Spitze der vorderen Wand. *Prosthema* kurz, lanzettförmig mit 2 nach vorne aufgeschlagenen Querfalten, an der Basis jederseits mittelst eines kleinen Lappchens mit dem Hufeisen verbunden. Im Gaumen drei grosse und dahinter drei schmalere Querfalten, welche letztere ihrer Länge nach nochmals getheilt sind. Flughaut und Finger sehr lang, das erste Daumenglied beiderseits vom *Patagium* eingeschlossen; ebenso der Schwanz, in dessen Spitze das *Patagium interfemorale* spitz verläuft. Das letztere, sowie die Partie zwischen Armen und Füssen innen fein behaart.

Die Farbe des Pelzes hell-mausgrau, Unterseite noch etwas lichter als der Rücken, Flughäute, Ohren und Nasenbesatz fleischfarb-grau. Nägel hellfleischfarb mit weisslicher Spitze und rostbraunem Fleck in der Mitte. Zehenrücken mit weisslichen feinen borstenartigen Haaren besetzt.

Rhinolophus acrotis brachygnatus K. Andersen, 1905

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Diagnosis. - Like *Rh. acrotis Andersoni*, but upper and lower tooth-rows, as well as mandible, distinctly shorter.

Skull and dentition (two skulls examined). - Although being apparently, on an average, larger than *Rh. a. Andersoni*, the present form has a smaller skull, smaller teeth, shorter tooth-rows and mandible (conf. measurements).

Colour. - The general colour of the single skin at my disposal (F imm.) is much darker than in the type of *Rh. a. Andersoni* (M imm.): greyish "drab" on the upperside, tinged with "mouse-grey" on the hinder back; "ecru-drab" on the underside; base of hairs on the back of the same colour as the underside.

Type. - M ad. (in alcohol). Ghizeh, Lower Egypt, Dec. 16th, 1891; collected and presented by Dr. J. Anderson. B. M. no. 92.9.9.7. There is a second specimen (F imm., skin) in the Museum from the same locality.

Distribution. - Known only from Ghizeh.

Remarks. - The absence, even in immature individuals, of the lower p_3 and upper p^2 distinguishes this form sufficiently from the only species with which it could possibly be confounded, viz. *Rh. euryale* and *Rh. clivosus*. Up to the present time three forms of *Rh. acrotis* have been described: *Rh. acrotis* (typicus) from Keren, Erytrea; *Rh. a. Andersoni* from the Eastern Egyptian Desert, about 22° N., 35° E.; and *Rh. a. brachygnathus* from the environs of Cairo.

Measurements of *Rh. acrotis brachygnathus* and *Andersoni*.

[table removed - eds.]

Rhinolophus acrotis schwarzi Heim de Balsac, 1934

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Un spécimen F de Tamanrasset, n° 114, 5-11-27.

L'histoire de ce spécimen est singulière: il a été déjà mentionné par MONOD sous le nom erroné de *Rhinolophus fumigatus* RÜPPEL (= *Rh. antinorii* DOBSON). La fausse détermination n'est pas imputable à MONOD, mais bien à l'illustre O. THOMAS, à qui le spécimen avait été soumis. L'erreur est explicable par le fait que O. THOMAS avait négligé d'examiner le crâne et n'avait pas dépouillé l'animal. Toutefois, la petite taille du spécimen a elle seule aurait pu le frapper.

Rh. acrotis était connu jusqu'ici de l'Erythrée et de l'Égypte (remonte jusqu'au Caire). Sa présence à Tamanrasset ainsi qu'à Djanet d'où nous avons reçu trois autres spécimens grâce à l'obligeance du Dr FOLEY montre que ce Chiroptère est bien établi dans les massifs centraux du Sahara et qu'il ne s'agit pas d'individus erratiques ou migrants. Son aire de répartition se trouve ainsi étendue considérablement vers l'Ouest (d'une distance de l'ordre d'au moins 2.500 kilomètres). Le fait a une importance biogéographique certaine et il se reproduit d'ailleurs pour d'autres Mammifères sahariens.

Au surplus les spécimens des massifs centraux constituent une race géographique particulière qui mérite d'être nommée.

La race nominale, *Rh. acrotis acrotis* v. HEUGLIN, a été décrite de l'Erythrée (Keren). Postérieurement THOMAS décrit une race

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plus petite, *Rh. acrotis andersoni*, d'après un spécimen immature, provenant de l'Est de l'Égypte (22° N - 35° E). Cette race ne fut pas admise par ANDERSEN, l'infériorité de taille d'un spécimen immature ne pouvant être prise en considération. Ultérieurement le même ANDERSEN décrit une race de petite taille, *Rh. acrotis brachygnathus* en provenance de la Basse-Égypte (Caire, Giseh). Mais nos spécimens du centre du Sahara sont encore plus petit que *brachygnathus*. E. SCHWARZ a bien voulu comparer le crâne de Tamanrasset à ceux d'*acrotis* (= *andersoni*) et de *brachygnathus* du Br. Muséum. Il nous a confirmé par écrit que ce spécimen était de façon marquée, plus petit que les animaux du Musée britannique, et qu'il méritait d'être décrit. D'autant plus qu'il s'agit ici d'une forme géographiquement très éloignée des autres, probablement isolée dans les massifs sahariens centraux, et en tout cas représentant un extrême.

Nous proposons de la dédier au Dr E. SCHWARZ sous le nom: *Rhinolophus acrotis schwarzi* nov. subsp.

Type: F adulte en alcool, Djanet, Tassili des Azdgers (nous préférons ne pas prendre pour type le spécimen de Tamanrasset dont les avant-bras sont brisés et qui ne peuvent être exactement mesurés).

Diagnose: Forme plus petite que *Rhinolophus acrotis brachygnathus* (avant-bras 45-46 mm., long, condylo-incisive 17 mm.); de teinte gris cendré clair (4 spécimens examinés).

[table removed - eds.]

Rhinolophus adami Aellen and Brosset, 1968

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TYPE. - F adulte, en alcool. Grotte de Kimanika, Kouilou, République du Congo, 9.1.1967; coll. J.-P. Adam, no orig. 12/8G. Muséum de Paris 1968-408.

PARATYPES. - M adulte. Grotte de Meya-Nzouari, Kouilou, République du Congo, 26.7.1963; coll. J.-P. Adam, Muséum de Genève 1129.84.

2 FF subadultes. Mêmes lieu, date et collecteur que le type, no orig. 3/8G et 8/8G. Muséum de Paris 1968-409 et 1968-410.

DIAGNOSE. - *Rhinolophus* de taille moyenne (avant-bras 47 à 49 mm; long crâne 20,3 à 20,6 mm). Feuille nasale grande (long. 15 à 16 mm, larg. 8,5 à 9 mm) à connectif arrondi dépassant quelque peu le sommet de la selle; lancette non acuminée, à bords droits ou un peu convexes; selle à dossier (procès vertical) large et à bords concaves. Troisième métacarpe non réduit, supérieur à 90 % du cinquième métacarpe. P² dans la rangée dentaire, séparant nettement C de P⁴. P₃ implanté au bord externe, mais bien développé et séparant P₂ de P₄. Crâne étroit: largeur zygomatique (qui est inférieure à la largeur mastoïde) mesurant moins de la moitié de la longueur totale (au bord antérieur de C). Pont palatal long; son bord médian antérieur arrive au niveau du bord antérieur de M¹.

DESCRIPTION. - Les dimensions externes (sauf la feuille nasale) entrent pour la plupart dans les limites de variation de *R. capensis*. Les oreilles sont très grandes, elles mesurent plus de la moitié de la longueur de l'avant-bras. Le bord interne est assez régulièrement convexe de la base au sommet; celui-ci est bien marqué et forme un angle droit, ou même un peu aigu, toutefois fortement arrondi à la pointe; le bord externe est nettement concave vers le sommet, puis très légèrement convexe jusqu'à l'antitragus. Celui-ci est régulièrement convexe et séparé du pavillon par une encoche à angle très obtus; il mesure environ 13 mm de l'insertion de base à l'encoche.

La feuille nasale est très caractéristique de cette espèce; elle est grande, surtout longue. Le fer à cheval recouvre presque tout le museau et présente une large échancrure au milieu de son bord antérieur. La selle est très large (3.2 mm) et son procès vertical (dossier) est environ aussi large que haut; ce procès est un peu plus large à la base qu'au sommet et ses bords latéraux sont légèrement concaves; il en résulte un étranglement au milieu de la hauteur (voir tableau de mensurations). Le connectif a une pointe régulièrement arrondie qui dépasse quelque peu le sommet du dossier de la selle. La lancette, triangulaire, possède des bords latéraux légèrement convexes et une pointe peu aiguë, donc absolument pas acuminée comme c'est le cas chez les espèces comparables, *simulator*, *bembanicus* et *capensis*.

La lèvre inférieure présente trois sillons délimitant deux lobes bien développés, comme chez *simulator* et *capensis*.

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Le patagium s'insère à la partie distale du tibia ou à la cheville.

Le troisième métacarpe n'est pas réduit; il mesure toujours plus des 90 % du cinquième. La queue est plus longue que la moitié de l'avant-bras.

La coloration de cette nouvelle espèce ne présente rien de particulier. Le type (F) est brun assez clair sur le dos; la face ventrale est gris-brunâtre. Les paratypes sont plus foncés dessus, mais le ventre est aussi gris-brunâtre; l'un (8/8G) est même blanchâtre au bas-ventre. La feuille nasale est très claire, couleur chair. Les oreilles sont brunes, un peu plus foncées au sommet. Le patagium est brun foncé.

Le crâne présente des proportions semblables à celles de *simulator-bembanicus*. Il est très étroit: sa plus grande largeur n'atteint pas la moitié de la longueur totale (mesurée au bord antérieur de C). Le renflement nasal est fortement marqué; le profil général du crâne rappelle beaucoup celui de *bembanicus* figuré par SENNA (1914, p. 2). La largeur zygomatique est égale ou inférieure à la largeur mastoïde. Le rapport largeur zygomatique sur longueur C-M³ est nettement inférieur (1.26) à celui mesuré sur *capensis* (1.38 à 1.41), et *simulator-bembanicus*

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[table removed - eds.]

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(1.30 à 1.40); ce rapport illustre particulièrement bien cette étroitesse du crâne, le pont palatal, mesuré au milieu, est remarquablement long et comparable à celui de *simulator-bembanicus*: il constitue les 38 à 39.5 % de la longueur C-M³, alors que chez *capensis* il ne représente que les 34 à 35 % (*simulator*: 38.5 %; *bembanicus* 42.5 %). Le milieu du bord antérieur du pont palatal arrive au niveau du bord antérieur de M¹; le point médian du bord postérieur se situe entre M² et M³. Un long pont palatal est un caractère archaïque chez les *Rhinolophus*. Dans notre nouvelle espèce, cette ancienneté se manifeste aussi dans la denture.

Au maxillaire supérieur, P² est situé dans la rangée dentaire et sépare nettement C de P⁴; les pointes de C et P⁴ sont séparées de 2 mm. La hauteur de P² atteint ou dépasse légèrement le cingulum de C.

Au maxillaire inférieur, P₃ est inséré sur le bord externe de la rangée dentaire et sépare P₂ de P₄. P₂ est remarquablement petit; cette dent n'atteint pas la moitié de la hauteur de P₄ et rappelle, par ce caractère, *R. alticolus*.

REMARQUES. - Il est curieux de constater la découverte dans une région géographique restreinte - l'ancienne AEF - de deux espèces nouvelles de *Rhinolophus* en l'espace d'une dizaine d'années. *R. silvestris*, décrit en 1959, est une espèce bien individualisée ayant son plus proche parent dans une espèce orientale, *deckenii*. La nouvelle espèce *R. adami* est proche, elle, de deux formes d'Afrique du Sud-Est, deux formes que plusieurs auteurs pensent devoir être synonymes: *simulator* et *bembanicus*. *R. bembanicus* n'est connu que par le type, provenant de Zambie. *R. simulator* a été signalé en Tanzanie, au Malawi, en Zambie, au Katanga (?), en Rhodésie, au Mozambique, au Botswana et en République d'Afrique du Sud (Transvaal et Natal).

Afin de pouvoir comparer facilement notre nouvelle espèce à ses plus proches parents, nous avons divisé les espèces africaines du groupe *ferrumequinum* en deux sous-groupes ainsi définis:

Groupe *ferrumequinum*: connectif à sommet arrondi et bas, ne dépassant pas ou seulement de peu le procès vertical de la selle. Largeur du fer à cheval inférieure à 9 mm (atteignant exceptionnellement 9 mm).

Sous-groupe *ferrumequinum*: P² externe ou absent. C et P⁴ en contact ou presque: espèces: *ferrumequinum*, *augur*, *darlingi*, *clivus*.

Sous-groupe *capensis*: P² dans la rangée dentaire, séparant nettement C de P⁴: espèces: *capensis*, *alticolus*, *denti*, *swinyi*, *simulator*, *bembanicus*, *adami*.

Rhinolophus aethiops diversus Sanborn, 1939

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Type from Bakel, Senegal, French West Africa. No. 19.7.7.2774 British Museum (Natural History). Adult female in alcohol. Collected September 27, 1887. From the Lataste collection.

Characters. - Averages smaller than either *a. aethiops* or *a. eloquens*, with a narrower horseshoe and sella and a shorter lancet. The type, when dried, is darker than *aethiops*, being in general a faded brown. The hairs above are of uniform color to the base; below they are slightly darker at the base. The type, however, has been in alcohol for many years and the only skin is more than one hundred years old.

The type skull is like that of *aethiops* but much smaller. Upper premolar two is present on one side only and is so minute that it is hard to see even with a lens. Lower premolar three is present on both sides and is also very minute.

Measurements. - Type (maximum and minimum of topotypes in parentheses): Forearm 53.2 (52.8 - 55.6); third finger, metacarpal 40.6 (39 - 40.7), first phalanx 16 (16 - 17.7), second phalanx 30.3 (30 - 31.5); fourth finger, metacarpal 41.9 (41 - 42.7), first phalanx 9.6 (9.8 - 10.3), second phalanx 18.7 (17.9 - 18.7); first finger, metacarpal 41.9 (42 - 44), first phalanx 12.6

(12.5 - 13.2), second phalanx 13.9 (14.1 - 14.6). Ear 27 (27 - 28); tail 28.6 (28.8 - 32.7); tibia 22.6 (22.8 - 23.5). Skull of type: greatest length 23.1; condyle-basal

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length 19.6; palatal length 2.9; width across nasal swellings 6.1; interorbital width 2.6; zygomatic width 11.9; mastoid width 10.5; width of braincase 9.5; upper toothrow 8.8; width across canines 6.6; maxillary width 8.7; supraorbital length 6.4.

Specimens examined. - Total 7. Bakel, Senegal, 2 males (alc.), 2 females (alc.); "River Gambia," 1, no sex. Nerokoro, near Tambikunda, Sierra Leone, 1 male (alc.), 1 female (alc.). All in British Museum.

Remarks. - All the forms of *R. aethiops* are very much alike except *a. hildebrandti*, which is the largest. *R. diversus* does not agree very well either with typical *aethiops* from Angola or with *a. eloquens* from the Sudan, being smaller and darker. The two specimens from Sierra Leone are not included in the measurements as they are both subadult and are only referred to *R. diversus*.

Rhinolophus alcyone Temminck, 1853

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RHINOLOPHUS ALCYONE.
RHINOLOPHE ALCYONE.

C'est la seule espèce de cette section qui ne soit parvenue de la partie littorale de l'Afrique, dont nous nous occupons. Elle est munie d'une follicule en fer-de-lance, large à sa base, mais peu haute et munie de deux rangées de petites carités: un socle repose sur le bourrelet et est entouré du large fer-à-cheval bordé d'un petit plis du derme. Les oreilles sont grandes et hautes, terminées par un oreillon mobile.

Nous n'avons pu observer qu'une femelle; il est dès-lors incertain s'il faut attribuer l'existence d'un syphon, dans le mâle.

Le pelage de la femelle est pour toutes les parties su-

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périeures; d'un rouge-vif couleur de briques cuites; en dessous d'une teinte plus claire, couleur de rouille. Les oreilles n'ont que la base poilue; leur pointe est recourbée en dehors; celles-ci; de même que toutes les autres parties du derme sont noires.

Longueur jusqu'au bout de la queue 3 pouces; la queue seule 9 lignes; envergure 11 pouces; antibrachium 1 pouce 9 lignes.

Patrie. La rivière Boutry à la Guinée.

Rhinolophus alcyone alticolus Sanborn, 1936

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Type from cave at 5,800 feet on Mount Cameroon, Cameroon Mandate. No. 42596 Field Museum of Natural History. Adult female. Collected July 2, 1934, by R. and L. Boulton.

Diagnosis. - A member of the *landeri* group with reduced second phalanx of fourth finger. Like *alcyone* but much smaller. With the exception of the ears, which are larger, about the size of *lobatus*. Skull with characters of *alcyone*; size of *lobatus*, but narrower.

Color. - General color of upper parts dark brown, between Raw Umber and Mummy Brown (Ridgway, 1912). Base of hairs much lighter. Below brownish gray.

Skull. - As in *alcyone* in that p_3 is partly in tooth-row and separates p_2 and p_4 , but much smaller. Length as in *lobatus*, but with shorter tooth-rows and narrower throughout.

Measurements. - Type and three topotypes: forearm 46.2 (46.2 - 47.1); third digit metacarpal 31.8, first phalanx 14.3, second phalanx 25.5; fourth digit metacarpal 34.6, first phalanx 7.7, second phalanx 15.7; fifth finger metacarpal 35, first phalanx 10.1, second phalanx 17.7. Ear, from meatus 22, from crown (dry) 15, width (dry) 14.1. Tibia 17.3; foot 9; calcar 11.9. Skull: total length 19.2 (19.2 - 19.6); total length to canine 18.5 (18.4 - 18.9); basilar length to canine 14.9 (14.7 - 15); interorbital width 2.4 (2.4 - 2.5); zygomatic width 9 (9 - 9.1); mastoid width 9.1 (9 - 9.1); width of brain case 8 (7.9 - 8); width across canines 4.4 (4.3 - 4.5); width across m^3 - m^3 , 6.4 (6.3 - 6.4); width of nasal swelling 4.8 (4.8 - 4.9); upper tooth-row c- m^3 , 6.6 (6.6 - 6.7); lower tooth-row c- m^3 , 6.9 (6.7 - 6.9); mandible 11.9 (11.8 - 12.1).

Specimens examined. - Type and three topotypes.

Remarks. - This race appears to be a highland form most closely related to *alcyone* and a western representative of *lobatus* from East Africa. Thomas (Ann. Mag. Nat. Hist., (8), 7, p. 458, 1911) listed a *Rhinolophus* from Panyon (4,000 feet), Nigeria,

as "near *R. alcyone*" which may be the same. Andersen (Ann. Mus. Civ. Stor. Nat., Genova, (3), 2, p. 190, 1905-06) has published an excellent series of measurements of this group.

Rhinolophus Andersoni Thomas, 1904

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Rhinolophus Antinorii, Anderson and de Winton, Mamm. Egypt, p. 96, pl. xvi. fig. 2 (1902) (nec Dobs.).

Allied to *R. clivosus*, Rüpp., but with no small anterior premolars and different colour.

General characters as in *R. clivosus*, the nose-leaf and ears being apparently very much as in that species; perhaps the front face of the median vertical process is slightly more narrowed upwards, but the difference is very slight; horseshoe small, not covering the muzzle laterally. Wings from the ankles.

Colour drab-grey, the hairs above slightly darkened terminally, those of the belly drab-grey throughout. In *R. clivosus* the belly is white. Membranes transparent greyish, rather darker along the centres of the digital interspaces.

Skull with a broader heavier muzzle and nasal region than in *R. Dobsoni* (the bat considered by Dobson as *R. clivosus*) or than in *R. euryale*.

Small premolars, both upper and lower; absent in every specimen, the large upper premolar pressed close against, even overlapping, the canine. In *R. clivosus* the small premolars are present both above and below.

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Dimensions of the type (measured in the flesh): -

Forearm 46 mm.

Head and body 53; tail 28; ear 22.

Skull: front of canine to occiput 19; length in middle line 16; front of bony palate to basion 12.5; breadth of palate across m^2 7.5; tip of upper canine to tip of large premolar 1.6; front of lower canine to back of m_3 8.3.

Type. Male. Original number 26. Captured 3rd August, 1903. Four specimens.

"Arabic name 'Wat-wat'; Bisharin name 'Humush-Kilite.'" - A. M. M.

In the British Museum collection there is already a spirit-specimen of this bat, obtained by Dr. J. Anderson at Gizeh. This is the *R. Antinorii* of that naturalist's 'Mammals of Egypt,' a note being appended by Mr. de Winton drawing attention to its possible identity with *R. clivosus*. It is certainly not *R. Antinorii*, which is larger in all ways and has especially a much larger nose-leaf, the horseshoe of which nearly covers the muzzle.

With *R. clivosus* it has no doubt a nearer affinity, but seems to differ sufficiently by its different colour and the constant absence of the minute premolars to be recognized as a distinct species. I have also ventured to distinguish the Kordofan bat described by Dobson under the name of *R. clivosus*, as it has the small premolars quite separating the canines from the large premolars.

Whether the queer greenish colour of the two specimens of *R. Dobsoni* ("sulphur-brown above, beneath canary-colour," Dobson) is natural or the result of defective preservation can only be determined when further examples are obtained.

Rhinolophus augur K. Andersen, 1904

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Short diagnosis. - Apparently allied to *Rh. ferrum-equinum*, but readily distinguished by its much smaller ears and shorter tail. Forearm 54-57 mm.

Nose-leaves. - Horseshoe not covering muzzle laterally. Sella pandurate, as in *Rh. ferrum-equinum*, but decidedly narrower. Posterior connecting-process triangular, obtusely pointed, its upper margin straight, shorter than the vertical height of sella. Lateral margins of lancet deeply emarginate.

Ears. - Not reaching tip of the muzzle, when laid forward. Tip attenuated and acutely pointed. Outer margin strongly concave below the tip of the ear. Greatest width of ear equal to its length from notch on outer margin to tip.

Wings. - General characters as in *Rh. ferrum-equinum*, but forearm on the average shorter, its maximum length (57.2 mm. in a series of 15 specimens) being almost exactly like mean measurements in *Rh. ferrum-equinum* (57 mm., 31 specimens from Europe). 5th metacarpal slightly longer than 4th. 2nd joint of 3rd finger almost twice the length of 2nd joint of 5th finger. In the folded wing, tip of 1st joint of 3rd and 5th fingers opposite posterior point of elbow. Plagiopatagium inserted at the tarsal joint.

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Tail. - Much shorter than in *Rh. ferrum-equinum*, its length (31.3 mm, on average) being almost exactly the same as the 2nd joint of 3rd finger (30.7 mm.); in *Rh. ferrum-equinum* much longer (36.7 against 29.5 mm.). Hinder border of interfemoral, between tip of calcars, slightly triangular.

Colour (16 skins). - General colour above "drab," with a tinge of fawn. Individual hairs "ecru-drab" at base, drab at tip. On the middle of the back an ill-defined (in some individuals scarcely perceptible) horseshoe-shaped patch, approaching "wood-brown"; branches of this patch on the shoulder-region, convexity backwards. Length of hairs on middle of back about 7.5 mm. Underside bright ecru-drab, more or less tinged with vinaceous buff on the breast and sides of the body. Ears (dried) transparent brown, membranes dark brown. Adult individuals vary but very slightly in colour, some being more greyish, others more fawn in hue. Young specimens (5 skins) are much duller coloured; fur above greyish brown; horseshoe-patch on back indicated, but rather indistinct; below bright grey, with a slight rosy tinge.

Dentition. - Essentially as in *Rh. ferrum-equinum*. Upper c and p^4 closely approximated, their cingula actually touching each other or separated only by an almost hair-fine interspace. Upper p^2 minute, blunt, placed in the outer angle formed by c and p^4 , hardly rising to the level of the cingulum of the canine; this small tooth is present in all of the fourteen skulls examined, but will no doubt be found wanting in very old individuals with much-worn teeth (cf. *Rh. augur zambesiensis*). Upper p^4 as high as two thirds of the canine. Lower p_2 one third of p_4 . Lower p_3 , when present, extremely small, placed on the external side of the tooth-row, blunt, as high as the cingulum of p_4 ; in nine out of fourteen skulls examined this tooth is wanting, and every trace of the alveole has disappeared; in a tenth specimen (adult) the tooth is wanting on both sides, but the left alveolus is still to be traced: in the remaining four skulls p_3 is present on both sides; three of these latter are skulls of young bats, the fourth of an apparently adult individual, though still with almost unworn teeth. As proved by these facts, the presence or absence of p_3 in this species varies individually, without any bearing on differences in geographical distribution: of six specimens from the same cave, taken by the same collector and in the same season of the year, two have lost both the tooth and every trace of the alveolus, while in four specimens it is present on both sides; evidently it is most often to be found

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in *young* individuals, more rarely in full-grown, perhaps never in very old. Lower p_4 a little higher than antero-exterior cusp of m_1 .

Measurements. - Below under *Rh. augur zambesiensis*.

Type. - M ad. (skin). Kuruman, Bechuanaland, 4000 feet, collected by Mr. R. B. Woosnam, April 19th, 1904. B.M. no. 4.10.1.1. Original no. 26. "Caught in an old mine; of fifteen taken there was only one female."

Distribution. - Specimens have been examined from the following localities: - Zuurbron, Wakkerstrom, S. Transvaal (1 specimen in alcohol); De Kaap, S. Transvaal (2, alcohol); near Krügersdorp, S. Transvaal, 4900 feet (6 skins); "Transvaal" (1, alcohol); Vredefort Road, N. Orange River Colony (1, alcohol; 2 skins); Kuruman, Bechuanaland, 4000 feet (8 skins). It will probably be found generally distributed over the whole of that part of South Africa which is irrigated by the Orange River and its confluent ("Orange River Area," in the zoogeographical sense of that term).

Remarks. - The present species has been confused with *Rh. ferrum-equinum* and *Rh. capensis* to such extent that it will be difficult to disentangle its synonymy without examination of the specimens recorded in literature. *Rh. capensis* differs mainly in the following points: - It is much smaller: forearm 49.5 mm. (average of eleven specimens), against 56 in *Rh. augur*; the ears much longer (20.5 mm. from base of inner border) and broader (16.7 mm.); tip of the ear obtusely pointed; sella only very slightly narrowed in the middle, its lateral margins subparallel in their upper half; posterior connecting-process shorter, its upper margin concave; all metacarpals and finger-joints much shorter, therefore the wing very much narrower in antero-posterior direction; even a badly prepared skin of *Rh. capensis*, in which the shape of the ears and nose-leaves is unrecognizable and the length of the tail not to be relied upon, can always be at once distinguished from *Rh. augur* by taking the measurement of the 2nd joint of the 3rd finger (22-26 mm., against 28.7-32 in *Rh. augur*); the tail remarkably short (20.8 mm.), only about the same length as the tibia, far shorter than the 2nd joint of 3rd finger, &c. The skull of *Rh. capensis* is slightly smaller; the nasals more swollen; the auditory bullae larger; the basioccipital therefore still narrower; the tympanic ring larger. As far as the available material goes, *Rh. capensis* seems to be restricted to the *western part of the Cape Colony*, as far north as the mountains on which the coast-rivers spring, eastwards to Winter-Bergen.

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Rh. augur bears, in fact, much more resemblance to *Rh. ferrum-equinum*. The differences between the two species have already been pointed out above. Whether, however, this resemblance is indicative of true relationship, or rather of convergence owing to similar habits and similar effects of natural surroundings, is open to question. It is in this connexion worth while mentioning that no doubt all the previous records of the occurrence of *Rh. ferrum-equinum* in Africa south of the Mediterranean subregion rest on confusion with other species; that, consequently, the two species, at the present time at least, are separated from each other by almost the whole of the tropical region of Africa.

Rhinolophus augur zambesiensis K. Andersen, 1904

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Short diagnosis. - Very dark-coloured. In size somewhat intermediate between *Rh. augur* and *Rh. a. zuluensis*. Forearm 53.3-54.8 mm.

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Colour (1 skin). - In the two forms just described the more or less distinct dark zone on the back takes the form of a horseshoe-patch. In the present race this patch extends over the whole of the back, giving the upperside of the animal an almost uniform brown colour, approaching "mummy-brown." Underside light "drab," somewhat darker in shade than in *Rh. a. zuluensis*. Ears and membranes (dried) black; in spirit-specimens the ears are "raw umber," the wings blackish brown.

Dentition. - The only skull examined is of a very old individual with much-worn teeth; both upper p^2 and lower p_3 are wanting, and no trace of their alveoli is to be seen. In younger individuals the dentition will no doubt be found to agree with that of the other races.

Type. - M ad. (skin). Fort Hill, N. Nyasa, about 9° 40' S., 33° 20' E., collected by Mr. A. Whyte, July 1896, presented by Sir Harry Johnston. B.M. no. 97.10.1.18. Original no. 136.

Distribution. - Specimens examined: - Mazoe, Mashonaland, 17° 30' S., 23° 30' E. (3, in alcohol); Zomba, Nyasa, 15° 30' S., 35° 30' E. (1, alcohol); and the type. - From these localities it may be supposed that the present form is generally distributed over the whole of the area which is irrigated by the Zambesi and its confluent.

Remarks. - As will be seen from the above, the three forms of *Rh. augur* just described are characteristic of three distinct zoogeographical districts of S. Africa: - *Rh. augur* of the Orange River District (in wider sense); *Rh. a. zuluensis* and *zambesiensis* of the areas from which I have derived their names*.

Rhinolophus augur zuluensis K. Andersen, 1904

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Short diagnosis. - A small and darker-coloured race of *Rh. augur*. Forearm 51.2-54.5 mm.

Colour (8 skins). - Fur above approaching "wood-brown." Horseshoe-patch on back nearest to "russet." Beneath light "drab," tinged with pinkish buff on the sides of the body.

Measurements. - See table under *Rh. augur zambesiensis*.

Type. - F ad. (skin). Jususic Valley, 20 miles N.W. of Eshowe, Zululand, 1000 feet, collected by Mr. C. H. B. Grant, Nov. 17th, 1903, presented by O. D. Rudd, Esq. B.M. no. 4.5.1.8. Original no. 602. "Caught in old gold-driving."

Distribution. - Specimens examined: - King William's Town (1, in alcohol); Port Natal (1, alcohol); typical locality (8 skins). This race will probably be found generally distributed over the whole of the Natalese Province, southwards to Winter-Bergen, northwards to about the latitude of Delagoa Bay, westwards to the mountain-range in which the coast-rivers rise.

Remarks. - The obvious difference in size and colour, combined with the distinct geographical distribution, entitle this form to subspecific rank. It cannot be confounded with young individuals of the typical form: young *Rh. augur* are greyish brown above, bright grey with a slight rosy tinge beneath; *Rh. a. zuluensis*, owing to a stronger development of the darker-coloured tips of the hairs, next to wood-brown above, light drab beneath.

Rhinolophus axillaris Allen, 1917

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Plate LI, Fig. 2.

Type, No. 49175, M ad. (skin and skull), Aba, Uele district, Belgian Congo, Dec. 17, 1911; Herbert Lang and James P. Chapin. American Museum Congo Expedition. Orig. No: 1807.

Apparently a member of the *R. landeri* group. Has two phases, a dark brown and a rufous phase.

Dark phase. - Type, above dark hair-brown with a faint rufescent tone in strong light, the fur dull gray-brown basally, with long hair-brown tips (in some specimens forming a dark broad V-shaped area on the lower back); below much lighter, the fur pale brownish plumbeous basally, the tips darker, giving a pale brownish wash. At the base of each arm is an axillar patch of short, stiff, dark rust-colored hairs, about 4 by 6.5 mm. in area, in strong contrast with the surrounding fur, both in color and texture. Ears and feet pale brown, membranes darker.

Total length (collectors' measurements), 77 mm.; head and body, 52; tail, 23; foot, 11; ear, 18 1.

Rufous phase. - Above rich cinnamon-brown, the fur basally pale gray, with long dark cinnamon-brown tips; below pale buffy white, with a grayish tone.

The red phase is represented by a single specimen, in which the glandular axillary area is pale yellow. The dark phase is represented by 4 specimens, in one of which an axillar glandular area is not evident, and in a second is less strongly marked than in the other two. All were taken at the same locality and practically on the same date (three on Dec. 17, the other two respectively on Dec. 13 and 15).

Collectors' measurements (3 males, 2 females): Total length, 79 (75-84); head and body, 54 (52-58); tail, 26 (23-29); foot, 11 (10-12); ear, 19 (18-22).

The skull of the type is badly crushed. The measurable skulls give the following: Total length, 3 skulls, 20 (all 20); zygomatic breadth, 2 skulls, 10.2 and 9.6; breadth at base of canines, 4 skulls, 5.4; breadth at m^3 , 6.9 (6.7-7.2).

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P^2 is greatly reduced but stands in the toothrow, closely crowded between the canine and p^3 .

A peculiar and strongly marked feature of this species is the axillar gland, rendered conspicuous by a short tuft of stiff rust-colored hairs.

Rhinolophus brockmani Thomas, 1910

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Rhinolophus brockmani, sp. n.

A very pale-coloured species, allied to *R. lobatus* and *dobsoni*.

General characters as in *R. dobsoni*, the nose-leaf apparently quite as in that species. Colour remarkably pale, the hairs of the upper surface white with a brown tip about a millimetre in length, and of the under surface wholly white from base to tip.

Skull as compared with that of *R. dobsoni* rather longer and narrower, with a more elongate brain-case. Opening of posterior nares rather broader.

Cheek-teeth markedly smaller, the molars (particularly m^3) narrower, and the inner lobe of m^1 and m^2 smaller, so as to have a larger space between each tooth. Small upper premolar large, in the centre of the unusually extended space between the canine and p^4 , a space double as broad as in *R. dobsoni*. P^4 with its antero-external corner projecting forward in front of the anterior edge of the narrow inner lobe, the latter being in *R. dobsoni* practically the most anterior part of the tooth, no angular external projection being present; inner lobe of the same tooth much narrower than in *R. dobsoni*.

Dimensions of the type (the starred measurements taken in the flesh): -

Forearm 44 mm.

Head and body* 45; tail* 26; ear* 18. Third finger, metacarpus 29; 1st phalanx 14, 2nd phalanx 25; lower leg and foot (c. u.) 27.

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Skull: greatest length 18; front of canine to back of m^3 6.6.

Hab. Upper Sheikh, Somaliland. Alt. 4500'.

Type. Adult female. B.M. no. 9.12.17.4. Original number 237. Collected 13th November, 1909, and presented by Dr. R. E. Drake-Brockman. Two specimens.

The most striking character of this bat is its peculiar whitish colour, other members of the group being generally some shade of grey. Unfortunately the colour of the original specimens of its nearest ally *R. dobsoni*, from Kordofan, cannot be determined, but the difference in the skulls and teeth is quite sufficient to separate the two.

A number of *R. acrotis*, Heugl., were obtained at the same time and place.

Rhinolophus caffer Sundevall, 1846

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plicis postnasalibus duabus integerrimis, rotundatis (ut in div. *Rh. nobilis*); fuscescens, pilis dorsi albidis, basi apiceque fuscus; gastræi griseis, basis fuscis; auriculis orbicu-

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laribus apiculo brevissimo, acuto, lateraliter, supra incisuram levem, prominulo. - Antitragus ("lobulus") auris parvus, sed altus, semicircularis, cum membrana conchæ connatus, vix ullâ incisurâ distinctus. Parvus: cubitus 48 mm. - Circa Port-Natal inventus.

Rhinolophus clivosus Cretzschmar, 1828

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Rhinolophus clivosus.
(Mus. Francof.)

Diagnos. *Rhinolophus* apparatus olfactorio externo clivis gradatim elatis non dissimili. Fossae nasali ferro equino membranaceo circumdatae interpositus scyphus parvulus - sequitur membrana transversalis concavata, antrorsum eminens, culmine obtusa tunc membrana recta, conjungens posteriorem transversarie positam, hastatam. Corporis colore ex fusco cinerascens.

Ausmessungen

-; Fuss; Zoll; Lin.

Länge des Körpers mit dem Kopfe; -; 2; 6

Länge des Kopfes; -; -; 9

Länge des Ohren; -; -; 8

Länge des Schwanzes; -; 1; -

Breite der ausgespannten Flügel; -; 10; -

Beschreibung

Die Farbe der zarten wolligten Haare der hügeligten Hufeisennase ist ein liches graubraun, das auf dem Rücken dunkler, auf dem Unterleibe heller scheint. Die äusseren Riechwerkzeuge sind von sehr zusammengesetzter Art. Die hufeisenförmige wulstige Membran hat in ihrem Mittelpunkt eine Vertiefung, in welcher der Nasenknorpel sich zu einem

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kleinen Trichter gestaltet; hinter demselben erhebt sich ein quer aufgerechtester Knorpel, der ziemlich stark an seiner Spitze, flach zugerundet in seiner Mitte, länglich ausgehöhlt und über den Trichter etwas herübergebogen ist. Von diesem läuft eine gerade nach hinten gerichtete dünne und gezackte Membran zu der hintersten quer stehenden lanzettförmigen, welche sie mit der vorderen verbindet. Die hinterste Membran ist dünn, schliesst sich nach vorne an die hufeisenförmige Membran an, nach hinten geht sie in die allgemeine Haut über, und ist deswegen an ihrer hinteren Fläche mehr behaart, als an der vorderen. Rings um diese Riechwerkzeuge stehen einige Borstenhaare. Von vorne gesehen nehmen sich diese sämtlichen Membranen aus wie übereinander aufgeschichtete Hügel, von denen die hinterste die Spitze bildet. Die Ohren sind am äusseren Rand etwas ausgeschnitten, und wie die Flügel und Schwanzhäute, braun.

Vaterland. Die hügelichte Hufeisennase wurde bei Mohila gefunden.

Aufenthalt. Mauern und Felsen.

Rhinolophus clivosus hillorum Koopman, 1989

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HOLOTYPE: AMNH 257044, a female obtained by Andrew S. Voros (5924) in early October at the John Hegbe Farm near Zozoma, ca. 2 mi SW Voinjama in Lofa County, extreme northwestern Liberia at ca. 500 m elevation. This would place it at ca. 8°25'N, 9°35'W. The holotype consists of an entire

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specimen preserved in alcohol with the skull extracted and cleaned.

DIAGNOSIS: Larger than other subspecies of *R. clivosus*. Much larger than any of the northern subspecies (*bocharicus*, which many regard as a separate species, *clivosus*, *brachygnathus*, *schwarzi*, *acrotis*) and somewhat larger than the eastern and southern *keniensis*, *zuluensis*, and *augur*. Selected measurements are as follows (range of *hillorum* followed by combined range of *keniensis*, *zuluensis*, and *augur*): forearm length (52 - 57, 50 - 56); condylocanine length (20.5 - 21.7, 18.7 - 20.5); zygomatic breadth (12.7 - 13.3, 11.1 - 12.2); mastoid width (10.8 - 11.5, 9.7 - 11.1); width across last upper molars (9.0 - 9.4, 7.6 - 8.6); maxillary toothrow length (8.9 - 9.4, 7.6 - 8.8). The size difference is particularly evident in the breadth of the palatal area and the volume available for the temporalis musculature as shown by the width across the molars and the zygomatic breadth. Going along with this are the larger teeth of *hillorum* and the absence in all three known specimens of the anterior upper premolar. This tooth is almost always present, albeit greatly reduced in *keniensis*, *zuluensis*, and *augur* (only 1 out of 40 skulls examined for this character). It seems evident that even if the very similar *keniensis*, *zuluensis*, and *augur* were combined into one subspecies, *hillorum* would still remain distinct.

ETYMOLOGY: I name this subspecies after the two (often confused) eminent mammalogists named John E. Hill: J. Eric (1907 - 1947), and J. Edwards (1928 -). I have known both of them and feel that each, in different ways, has been important in my understanding of African bats, to which both made outstanding contributions.

LOCALITY RECORDS: LIBERIA: Voinjama: AMNH 257044; Tokadeh, Nimba, 600 m: BM 79.459. CAMEROON: Lake Manenguba, Manenguba Mountains, 1800 m: ZFMK 69.485. All three specimens are preserved in alcohol with the skulls extracted and cleaned.

[*Rhinolophus Commersonii*](#) E. Geoffroy Saint-Hilaire, 1813

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J'ai trouvé cette espèce parmi les dessins, et manuscrits de Commerson, avec la désignation et les caractères suivans:

Chauve-souris du fort Dauphin, île de Madagascar. *Vespertilio, obscure caudatus, auribus simplicibus, amplis, acuminatis, erectis, patulis; naso duabus valvis transversis, late secedentibus hiulco.*

On ne pourroit confondre cette chauve-souris qu'avec le

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diadème: mais outre qu'elle est un peu plus petite que celui-ci, sa feuille est d'un tiers moins large, de même que sa queue est aussi du tiers plus courte. De plus, la membrane interfémorale qui se termine dans le diadème par un bord à angle saillant, est à angle rentrant dans cette espèce.

Il n'y auroit que l'éloignement des lieux, *Madagascar* et *Timor*, où se trouvent ces deux rhinolophes, pour autoriser la conjecture qu'ils forment deux espèces très-différentes.

J'ai annoncé plus haut que les rhinolophes se ressemblent par les couleurs et la disposition des poils: dans tous, en effet, la toison est épaisse, bien fournie et moelleuse: le poil est fauve en dessus, et blanc jaunâtre en dessous.

J'ai trouvé le trident dans les plus profondes excavations des montagnes, en Egypte et notamment dans les parties les plus reculées des tombeaux des rois et du temple de Denderah.

Ainsi dans, les pays chauds comme dans les pays froids, les rhinolophes recherchent également les lieux écartés; l'état de la température ne leur en fait donc pas une nécessité: en aurois-je trouvé la vraie raison en l'attribuant au défaut d'oreille interne?

Les rhinolophes de France ne rentrent pas tous les soirs en été dans les cavernes qu'ils habitent l'hiver. Songeroient-ils à s'épargner des allées et venues, et les fatigues du trajet dans leurs demeures souterraines? On ignore où ils se retirent pour passer le jour.

[*Rhinolophus Darlingi*](#) K. Andersen, 1905

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Diagnosis. - Differs from all other South-African *Rhinolophi* by the combination of these two characters: cingula of the upper canine and p^4 in immediate contact with each other; forearm 46.7-47.7 mm.

Nose-leaves. - Extreme length 14-14.5 mm. Horseshoe covering almost the whole of the muzzle; no tooth-like projection on either side of the median notch; greatest width 8.1-8.5 mm. Sella naked, slightly constricted below the middle, broadly rounded off at summit; vertical height 4 mm.; breadth at base, at constriction, and at summit

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2.1, 1.5, and 1.7 mm. Posterior connecting-process very short, rounded, in side view very nearly semicircular, scarcely projecting beyond the summit of the sella (this kept in vertical position). Lancet very long, scarcely constricted, its lateral margins evenly converging towards the tip; distance from the posterior transverse bridge to the tip of the lancet 4.2 mm.

Ears. - Reaching tip of muzzle, when laid forward: not attenuated below the tip; this latter blunt. Width of ear about equal to its length from notch on outer margin to tip.

Wings. - 5th metacarpal equal to (or only a mere trifle longer than) 4th. 1st phalanx of 4th finger about $\frac{2}{3}$ the length of 2nd phalanx. Plagiopatagium inserted on tarsus.

Tail. - Much longer than the tibia, about the same length as the 3rd metacarpal. Hinder margin of interfemoral triangular.

Colour. - General colour of adult individuals (2 skins) rather dark brownish drab above, "drab-grey" beneath. Base of hairs on back of the same colour as the underside. Length of fur on middle of back about 10 mm. - Immature specimens (2 skins) have a more greyish tinge on the back, but the difference from adult examples is very slight (but may perhaps be more pronounced in still younger individuals).

Skull. - Hinder border of palatal bridge opposite (or very slightly behind) the middle of m^3 .

Dentition. - Cingula of the upper canine and p^4 so closely approximated as to completely touch each other. Upper p^2 quite on the external side of the tooth-row; in one skull (F ad.) with unworn teeth it is as high as the cingula of the adjacent teeth; in two other skulls with slightly worn teeth it is hair-fine, hardly perceptible without a lens (therefore presumably lost in still older individuals). Lower p_3 , when present, external to the tooth-row, not reaching the upper margin of the cingula of p_2 and p_4 ; in a little more advanced age p_3 is lost.

Type. - Ad. (skin). Mazoe, Mashonaland, 4000 ft., June 13th, 1895; collected by J. ff. Darling, Esq. B. M. no. 95.8.27.1. Original no. 35. "Taken in mining adit." Five paratypes (three skins, two in alcohol) are in the British Museum.

Distribution. - A young *Rhinolophus* from Angola (B. M. no. 64.8.16.3*) is indistinguishable from the species here

* *Rh. æthiops*, specimen "e" in Dobson's Cat. Chir. Brit. Mus. (1878) p. 122.

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described, in external characters as well as in skull and dentition. Its geographical distribution therefore, probably, is from Angola across South Africa, eastwards at least to Mazoe.

Remarks. - The short diagnosis given above will, in every case, prevent confusion of the present species with any other hitherto known South-African *Rhinolophus*. *Rh. Landeri*, *lobatus*, *simulator*, and *Denti* differ, altogether, in having the upper p^2 placed in the tooth-row, separating the canine and p^4 by a distinctly perceptible interspace. *Rh. Landeri* is much smaller (forearm about 43 mm.), with much shorter ears, much narrower sella, higher connecting-process, much shorter tail (18-21 mm.), and with the 1st phalanx of the 4th finger peculiarly shortened (less than, or equal to, half the length of the 2nd phalanx). *Rh. lobatus*, which is of about the same size as *Rh. Darlingi*, may at once be separated by having the posterior connecting-process long, pointed, projecting like a small "horn" far beyond the summit of the sella. *Rh. simulator* is smaller (forearm 43.5-45 mm.); the front face of the sella is "rough" owing to the presence of very short hairs (most easily to be observed under a lens). *Rh. Denti* is much smaller (forearm 41-43 mm.); front face of sella as in *Rh. simulator*.

The species is named in honour of Mr. J. ffolliott Darling, to whom the British Museum is indebted for so many valuable accessions to its collection of South-African mammals.

[table removed - eds.]

Rhinolophus darlingi barbertonensis Roberts, 1924

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Rhinolophus darlingi barbertonensis subsp. nov.

Similar in structural characters to *R. darlingi* Andersen (Ann. and Mag. N.H. (7), xv, p. 20), which occurs in the highveld from Nyasaland to Transvaal, but differing therefrom in its consistently smaller size, especially in the skull. The following dimensions will serve to show this difference, those in brackets being of the typical form (from a specimen taken at Pretoria) and the others from the type taken in the district of Barberton; altogether ten specimens of the smaller form, from Barberton, Louws Creek, Gravelot Mine (near Leydsdorp) and Mokeetsi, all below the Drakensberg Mountains, and twelve of the larger, from Pretoria, have been compared, and the figures given represent the normal, hardly any overlapping being perceptible in the external dimensions, and less in the cranial. Length of head and body 45 (55), tail 33 (32), hind foot with claws 8 (8), ears 20 (21), all taken in the flesh. Length of forearm 45 (49); third finger, metacarpal 34 (36), first joint 4.5 (16.8), second joint 24.5 (26); fourth finger, metacarpal 33.2 (34.5), first joint 8.4 (9.8), second joint 15.2 (18); fifth finger, metacarpal 34 (36), first joint 11.2

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(11.8), second joint 4 (16.5); tibia and foot with claws 26 (28), Skull: length from front of canines to occiput 18.8 (19.5); width of brain case 8.8 (9.2), width across canines 5 (5.5); length of upper tooth row, from front of canines to back of hindmost molar 6.8 (7.2).

Type: Adult female, T.M. No. 2476, taken at Louws Creek, Barberton, on March 4th, 1920.

Rhinolophus darlingi damarensis Roberts, 1946

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A paler form than those of the eastern parts of South Africa, paler drab grey above and whiter below, the wings also averaging longer, though the skull is only slightly larger on the average.

Measurements of type from Oserikari, and minima and maxima of seven with skulls from Kochina, Great Karas Mountains, of a skin without a skull from Neudamm, and for comparison, in brackets, six with skulls from Transvaal of *R. d. darlingi* K. Andersen: Head and body 60, 53 - 56 (56 - 60); tail 30, 28 - 31 (27 - 33); hind foot (c.u.) 11 - 13 (8 - 10); ear 20, 18 - 21 (19 - 21); wing span 344, 300 - 315 (2), (290 - 315 in six). Forearm 50, 48 - 50.5 (Neudamm 51), (47.5 - 49); 3rd finger metacarpal 33.5, 32.5 - 35 (Neudamm 36), (31.2 - 32); 1st phalanx 18.5, 14.5 - 17.5 (15.8 - 16.8); 2nd phalanx 26, 25 - 29 (25 - 27.5); 4th finger metacarpal 37.5, 34 - 38.5 (34 - 36); 5th finger metacarpal 37.5, 36 - 39.5 (35 - 37). Skull, greatest length from canines 20.5, 19.5 - 20.5 (19 - 20); dorsal median length 17.7, 16.5-17.7 (16.5 - 17.2); zygomatic width 10.7, 9.9 - 10.5 (9.8 - 10.5); width of brain

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case at mastoid 8.4, 8 - 9 (9.1 - 9.2); width across m³ 7.9, 7.4 - 7.5 (7.4 - 7.7); width across canines 5.7, 5.1 - 5.7 (5.2 - 5.5); c-m³ series 7.7, 7.1 - 7.6 (7 - 7.4).

Type: T.M. No. 9474, adult F, Oserikari, Okahandja district, South-west Africa. Also seven with skulls from Kochina, Great Karas Mountains, Great Namaqualand, and one without skull from Neudamm, near Windhoek, Southwest Africa.

Rhinolophus Deckenii Peters, 1868

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Rh. fumigatus Ptrs., non Rüppell, Monatsber. 1866. p. 885).

Außenrand des Ohres mit einer stumpfwinkeligen Einbucht; die hintere Spitze des Sattels abgerundet, viel kürzer als seine vordere Oberfläche, welche nur sehr wenig in der Mitte verschmälert erscheint und an der Spitze abgerundet ist; Hufeisen jederseits am mittleren Einschnitt mit einem spitzen Zacken, am Rande flach wellenförmig. Der erste obere kleine Prämolargraben niedrig und stumpf, nach außen gedrängt. Flughaut geht etwas über den Hacken herab. Die Behaarung ist lang und fein, oben dunkler, unten heller rötlichbraun.

Masse eines ausgewachsenen Weibchens.

Meter.
 Totallänge: 0.097
 Kopf: 0.0265
 Ohrlänge: 0.023
 Ohrbreite: 0.019
 Länge des ganzen Nasenbesatzes: 0.0172
 Breite des Hufeisens: 0.011
 Schwanz: 0.029
 Oberarm: 0.035
 Vorderarm: 0.055
 L.1.F. Mh. 0.0055; 1 Gl. 0.003; 2 Gl. 0.0027: 0.010
 L.2.F. - 0.041: 0.041
 L.3.F. - 0.037; - 0.019; - 0.029; Kpl. 0.005
 L.4.F. - 0.042; - 0.011; - 0.0175; - 0.0023
 L.5.F. - 0.043; - 0.013; - 0.0165; - 0.0023
 Oberschenkel: 0.023

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Unterschenkel: 0.025
 Fufs: 0.015
 Sporn: 0.014

Das einzige Exemplar stammt aus der Sammlung des Baron C. von der Decken, von der Zanzibarküste. Erst neuerdings erhielt ich ein Exemplar von Rüppell's *Rhinolophus fumigatus* in Weingeist zur Untersuchung, woraus ich ersah, daß es nicht, wie ich glaubte, mit dieser Art zu vereinigen sei.

Rhinolophus Denti Thomas, 1904

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Allied to the European *R. euryale*, but smaller.

Size very small, among the smallest species of the genus. Leading characters (in the order used in Dobson's synopsis): posterior upper premolar separated from the canine, though not very widely, the small anterior premolar in the tooth-row, towards its outer side; horizontal portion of the sella not widely expanded, though (allowing for shrinkage in the dried skin) it would appear to be more so than is usual in the allied species; upper margin of the posterior connecting process forming a marked projection, rounded terminally, rising considerably above the summit of the front of the sella; sides of the vertical process of the sella parallel, summit broadly rounded off; antitragal notch shallow.

Horseshoe large, covering most of the muzzle, circular, its anterior edge sharply notched in the centre; lancet short, conical, its sides evenly convergent upwards, thickly covered with fine fur, similar in colour and quality to that of the head. Ears of medium size, their inner margin evenly convex, tip sharply pointed, upper half of outer margin slightly concave; antitragal notch not deep and the lobe itself comparatively little convex. Hind limbs slender and delicate. Wings from the lower third of the tibiae. Interfemoral membrane finely fringed posteriorly.

Fur close and fine, about 7 mm. long on the back. General colour above pale grey, the individual hairs dull whitish, with dark brown tips. Under surface nearly white. Membranes brown, the plagiopatagium and interfemoral inconspicuously edged with white.

Skull with the nasal convexity more developed than in *R. euryale*, less than is figured in Peters's *R. lobatus*. Palate ending opposite the posterior edge of the internal lobe of m^2 .

Dimensions of the type (those in inverted commas taken by the collector in the flesh): -

Forearm 42 mm.

"Head and body 41"; "tail 21"; "ear 20"; nose-leaf (dry) 9.2 X 6.3; lower leg and foot (c.u.) 25.5.

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Skull: greatest length 17; basal length to front of canines 13.2; breadth of brain-case 7.6; palatal bridge 1.9; front of upper canine to back of m^3 5.9; front of lower canine to back of m_3 6.6.

Hab. Kuruman, Bechuanaland. Alt. 1300 m.

Type. Male. B.M. no. 4.4.8.2. Original number 7. Collected 24th January, 1904, by R. E. Dent. Two specimens.

"Caught in a house."

This species, the smallest of South-African *Rhinolophi*, seems to represent *R. euryale*, but may be readily distinguished from that, as from all others, by its proportions, its pale colour, the high attachment of its wing-membranes, and its unusually hairy lancet.

Rhinolophus denti knorri Eisentraut, 1960

p. 3

Material: 5 Exemplare, 1 M (Tgb. Nr. 90), 15.XI.1956, 1 M (Tgb. Nr. 165), 27.XI.1956, bei Nyembaro am Salung-Plateau, 12 km westlich Kolente. - 1 M, 2 FF (Tgb. Nr. 127-129), 19.XI.1956, bei Nerebili, 400 m Meereshöhe.

Die 5 vorliegenden Exemplare von *denti* sind der erste Nachweis der bisher nur in südlichen Gebieten Afrikas gefundenen Art für Westafrika. Während die Nominatform nach den bisherigen Funden nordwärts kaum weit über den 20. südlichen Breitengrad hinausreicht, liegen die Fundorte in Guinea etwa auf dem 10. nördlichen Breitengrad. In der Luftlinie gemessen sind die nächsten Fundorte in den beiden Verbreitungsgebieten über 4000 km voneinander getrennt. Bei dieser großen Entfernung könnte man recht erhebliche morphologische Unterschiede der beiden Formen erwarten. Dies ist jedoch keineswegs der Fall. Immerhin berechtigen die vorhandenen Abweichungen zu einer subspezifischen Abtrennung.

Die Rasse *knorri* ist etwas kleiner als die Nominatform. Besonders auffallend ist die geringe Länge der Ohren und Hinterfüße. Dagegen ist das Gebiß nicht nur relativ, sondern auch absolut kräftiger. Die Felfärbung ist dunkler und zeigt einen stärkeren bräunlichen Ton gegenüber einem mehr blaßgrauen Ton bei der Nominatform.

Typus: M (Tgb. Nr. 165 A), 27.XI.1956, Höhle bei Nyembaro am Salung-Plateau, 12 km westlich Kolente, KNORR leg., Staatliches Museum für Naturkunde Stuttgart, Katalog-Nr. 6111. Kopf-Rumpf-Länge: 40 mm, Schwanz: 17 mm, Ohr: 17 mm, Hinterfuß: 6.5 mm, Unterarm: 37.5 mm, Tibia: 15.5 mm, gesamte Schädelänge: 17.0 mm, Jochbogen: 8.0 mm, maxillare Zahnreihe: 6.0 mm, Mandibellänge: 10.5 mm, mandibulare Zahnreihe: 6.0 mm.

Knorri ist in der Größe nur wenig von der typischen Form *denti* unterschieden. Sehr deutlich zeigt sich dies beim Vergleich der immer am korrektesten festzustellenden Schädelmaße (Tabelle 2). Die Durchschnittswerte für die gesamte Schädelänge und die Länge der Mandibeln sind bei *knorri* nur um ein geringes kleiner, und die Variationsbreiten greifen weit ineinander oder überdecken sich. Das gleiche gilt auch für andere Schädelmerkmale, und deren Durchschnittswerte ergeben meist nur geringe Differenzen.

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[table removed - eds.]

p. 6

[table removed - eds.]

Auffallend sind die durchweg etwas geringeren Körpermaße von *knorri*, wobei sich aber auch wieder in den meisten Fällen die Variationsbreiten überschneiden. Hervorzuheben sind die deutlich kleinere Ohrlänge und die geringere Länge der Hinterfüße. Sofern diese erheblichen Differenzen nicht auf verschiedener Meßmethode der verschiedenen Sammler beruhen, dürften die geringeren Maße für *knorri* typisch sein, zumal sich auch die Variationsbreiten nicht überschneiden.

Sehr charakteristisch für *knorri* erscheinen die Unterschiede in der Ausbildung des Gebisses. Zwar sind die Zahnproportionen selbst etwa die gleichen, ganz allgemein aber besitzt die Guinea-Form trotz ihrer geringeren Körper- und Schädeldimensionen

ein kräftigeres Gebiß. Beim Vergleich (Abb. 2) fällt besonders die größere Breite der Molaren auf. Diese Unterschiede zeigt auch folgende Gegenüberstellung:

Durchschnittliche Breite der oberen Molaren (M¹; M²; M³)

knorri (5 Exemplare): 1.5; 1.52; 1.43

denti (2 Exemplare): 1.3; 1.4; 1.3

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Die Fellfärbung ist bei der neuen Form dunkler als bei der Nominatform. Die Tönung der Oberseite ist bei vier Exemplaren der vorliegenden kleinen Serie mittelgrau Braun und als "Brownish Olive" zu bezeichnen. Bei F Nr. 127 ist sie etwas heller und geht mehr nach "Dresden Brown" über. Die Unterseite ist grauweißlich und bei dem letztgenannten Tier schwach cremefarben. Bei allen ist die Bauchmitte heller, die Seitenpartien etwas dunkler. In der Gesamtfärbung heben sich die 5 Stücke von *knorri* deutlich gegen die von mir durchgesehenen Exemplaren von *denti* ab. Letztere sind auf der Oberseite bedeutend heller (Buffy Brown) und mehr grau; nur einige zeigen einen etwas bräunlichen Ton. Ebenso ist bei allen die Unterseite heller, und zwar schmutzigweiß. Die Flughäute sind bei *denti* mehr bräunlich, bei *knorri* mehr grauschwärzlich.

Die vorliegenden Stücke wurden ebenso wie die der vorhergehenden Art, zum Teil mit diesen am gleichen Platz, in Höhlen gefangen, die bei Nyembaro und Nerebili im Hinterland von Guinea, etwa 150 km von Conakry entfernt, liegen.

Rhinolophus Dobsoni Thomas, 1904

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Rhinolophus clivosus, Dobs. Cat. p. 120 (nec Rüppell).

Type. Specimen *b*. F. B.M. no. 47.5.7.49. (Forearm 44 mm.)

Hab. Kordofan.

As Peters has shown, the true *R. clivosus* of Rüppell, from Mohila, Arabia, is one of the group with the large upper premolar pressed close against the canine, the small premolar being in the outer angle. In Dobson's bat, on the other hand, although he puts it in the same group, the anterior premolar separates the second premolar from the canine. This difference cannot be due, as he supposed, to immaturity, one of the specimens at least being fully adult.

Rhinolophus empusa K. Andersen, 1904

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Short diagnosis. - Anterior upper and middle lower premolars placed in the tooth-row. Sella wedge-shaped (deltoid). Horseshoe as broad as muzzle. A conspicuous dark patch under each eye. Forearm 48 mm.

Nose-leaves. - Horseshoe covering the muzzle laterally; a small tooth-like projection on each side of median notch; underlying leaf easily visible all round. Sella wedge-shaped; breadth at base about three fourths of vertical height; lateral margins plainly converging (with a scarcely perceptible constriction in the middle) towards the summit, which is subacutely pointed. Posterior connecting - process long, pointed, slightly curved forwards, as an erect horn, about five sixths the vertical height of the sella, thus projecting far beyond the summit of the sella. Lateral margins of lancet concave in the middle, tip behind constriction about 2.3 mm.

Ears. - Rather short, scarcely reaching tip of muzzle, when pressed forwards along the sides of the head. Tip obtusely pointed. Outer margin concave below the tip. Notch on outer margin shallow, almost rectangular. Extreme breadth of ear somewhat greater than the distance from the notch to the tip.

Wings. - 5th metacarpal longer than 4th. 1st joint of 4th finger equal to three fifths the length of 1st joint of 3rd finger. On the folded wing the tip of the 1st joint of the 5th finger reaches as far back as the posterior point of the elbow; tip of 1st joint of the 3rd finger falling only a trifle short of the same point. Plagiopatagium inserted at base of metatarsus.

Tail. - Much longer than tibia, about the same length as the combined 1st and 2nd joints of the 4th finger. Tip of tail projecting 2 mm. beyond the membrane (a character

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which, however, no doubt will prove to be subject to a certain amount of individual variation). Hinder border of interfemoral, between tips of calcars, almost square.

Colour (of spirit-specimen). - Light brown above, greyish beneath. Sides of face yellowish white, with a strongly marked dark brown patch under each eye. Membranes dark brown; free margin of plagiopatagium very narrowly bordered with whitish. Forearm, metacarpals, and fingerjoints yellowish.

Skull. - Essential characters as in *Rh. Blasii*, but parietal region of skull still more globular; frontal region considerably less constricted. Extreme width of brain-case slightly greater than zygomatic width. Median anterior point of palatal bridge

opposite front of m^1 ; median posterior point drawn out into a tooth-like projection. Upper margin of mandible from condyle to base of coronoid process almost straight. Coronoid process very small, projecting only a trifle beyond the highest cusp of m^1 .

Dentition. - Small anterior upper premolar (p^2) placed in the tooth-row. Upper canine and posterior premolar (p^4) widely separated. Cusp of p^4 three fourths the height of the canine. Middle lower premolar (p^3) extremely small, but *placed entirely in the tooth-row*, separating p^2 and p^4 from each other; p^2 one third the height of the canine. Cusp of lower p^4 not quite as high as the antero-exterior cusp of m^1 .

Type. - F ad. Zomba, Nyasa, collected by Mr. A. Whyte, Jan. 1893, presented by Sir Harry Johnston. B.M. no. 93.7.9.33.

Distribution. - Only one specimen, the type, examined.

Remarks. - This fine species stands quite alone among the African *Rhinolophi*, being the only hitherto known species with a wedge-formed sella. Its nearest ally is the South European *Rh. Blasii*. In dentition *Rh. empusa* occupies a slightly more primitive position than *Rh. Blasii*: in the former the lower p^3 is still entirely in the tooth-row, in the latter it is just on the point of being driven out on the exterior side of the tooth-row.

Measurements of the type.

Nose-leaves: mm.

Front of horseshoe to posterior point of lancet: 13.8

Extreme width of horseshoe: 9

Vertical height of sella: 3.1

Width of sella at base: 2.3

Ears:

Base of lower margin to tip: 15.2

Notch on outer margin to tip: 11.8

Extreme width: 13.2

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Wings: mm.

Forearm: 48

3rd finger, metacarpal: 30.8

" 1st joint: 15.7

" 2nd joint: 24

4th finger, metacarpal: 33.7

" 1st joint: 9

" 2nd joint: 15

5th finger, metacarpal: 35

" 1st joint: 11

" 2nd joint: 12

Tail, from anus: 25.3

Tibia: 20.2

Hind foot (c. u.): 9.8

Skull:

Extreme length: 19.8

Basal length: 15.2

Extreme width of brain-case: 9.3

Zygomatic width: 9

Maxillar width, across outer margin of m^3 : 6.1

Orbital constriction: 2.8

Palatal bridge: 2.6

Med. post. point of palate to front of premaxilla: 6.2

" " " for. magnum: 9

Mandible; condyle to front of incisors: 13.1

Teeth:

Front of upper canine to back of m^3 : 6.5

Front of lower canine to back of m^3 : 6.8

[*Rhinolophus euryale tuneti*](#) Deleuil and Labbé, 1955

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Nous avons pris un certain nombre de Rhinolophes euryales (*Rhinolophus euryale* BLASIUS 1853) dans la grotte d'El Haouaria (Cap Bon), et un unique exemplaire dans une grotte très profonde située entre Testour et El Aroussa. En mai, puis en septembre ces Rhinolophes étaient fréquents; en novembre, ils devinrent rares; le 12 décembre, nous n'en trouvâmes qu'un seul, comme d'ailleurs à Tsstour le 25 décembre. Migre-t-il en hiver?

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TROUESSART ne fait que supposer *Rhinolophus euryale* BLASIUS, en Tunisie. Au Maroc, on a trouvé la sous-espèce *Rhinolophus euryale barbarus* (K. ANDERSEN et MATSCHIC 1904) à Rabat (CABRERA 1932).

Le tableau 3 (p. 52) donne ses mensurations en millimètres.

DIAGNOSE. - Les côtés de la selle, vue en avant, sont parallèles. Le connectif, rétréci après le milieu et à bords très concaves, présente, vu de profil (cf pl. XXI) une pointe inférieure aiguë et très en avant, et une pointe supérieure obtuse qui tend à la surplomber sans toutefois la dépasser.

La lancette reste conique, vue de face, comme dans le Grand Rhinolophe et le Petit Rhinolophe.

Le patagium, extérieurement, s'insère au talon, tandis qu'intérieurement l'uropatagium ne se déploie qu'à 3 ou 4 mm du talon. La membrane interfémorale reste plus ou moins triangulaire. Les membranes alaires sont de couleur sombre

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bistre, comme l'oreille; celle-ci, de même couleur, présente une pointe aiguë tournée dehors et non échancrée et un antitragus arrondi dont la largeur est le double de la hauteur. La face dorsale est beige cendré sombre, tandis que la face ventrale est d'un blanc légèrement beige. La feuille nasale ne couvre pas tout le museau.

DISCUSSION. - *Rhinolophus euryale* nov. subsp. de Tunisie ne peut se comparer à *Rhinolophus hipposideros vespa* Laurent 1937, car il est beaucoup plus grand, et son connectif, de profil

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ne ressemble pas du tout au connectif obtus et très postérieur du *Rhinolophus hipposideros*.

On peut penser au Rhinolophe de Méhély, espèce orientale, mais le nôtre est nettement plus petit. Sa lancette est conique et non à bords très concaves, devenant presque parallèles et la pointe postérieure du connectif n'est jamais aiguë, mais obtuse et tend à surplomber la pointe aiguë inférieure (cf. fig. 2). Enfin, les dimensions du crâne de *Rhinolophus mehelyi carpetanus* CABRERA sont supérieures à celles du crâne du Rhinolophe tunisien. Il se rapprocherait de *Rhinolophus blasii* PETERS, de la région méditerranéenne orientale, mais en diffère par son connectif, sa fourrure moins foncée, ses oreilles bistres, etc.

En comparant notre sous-espèce tunisienne à *Rhinolophus euryale barbarus* (ANDERSEN et MATSCHIC), type Tanger, elle en diffère par ses dimensions plus petites (avant-bras, doigts), par son antitragus deux fois au moins plus large que haut, par son connectif moins aigu, plus obtus, plus postérieur; ses ailes et oreilles sont plus foncées, presque comme celles de *Rhinolophus Mehelyi*.

Enfin, en comparant les dimensions de notre Rhinolophe à celles des Rhinolophes euryales de l'Institut Scientifique Chérifien, d'origine non précisée, données par M. PANOUSE, on constate que les premières sont, dans l'ensemble, supérieures aux secondes. Nous pensons donc que le *Rhinolophe euryale* type *El Haouaria* (Cap Ron) pourrait être une sous-espèce originale tunisienne que nous nommerons *Rhinolophus euryale tuneti* DELEUIL et LABBE.

[The name *tuneti* first appears on p. 53 - eds.]

Rhinolophus foxi Thomas, 1913

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Closely allied to *R. deckeni*, Peters, of East Africa, but greyer in colour and with smaller teeth.

Size rather less than in *R. deckeni*. General colour above "drab-grey," below more whitish grey. Nose-leaf and ears apparently as in *deckeni*.

Skull as in *R. deckeni*, but rather smaller; the nasal

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swelling just about as in *deckeni*, larger than in *ferrumequinum* and *augur*. Bony palate extending slightly further forward, to the level of the junction between p^4 and m^1 . Teethall slightly smaller than in *deckeni*, the combined length of p^4 - m^2 , on outer edge, being 5.3 mm. as against 5.8 in that species. Minute premolar still further reduced, absent on one side in each of the specimens, and on the other quite minute, smaller than an incisor, and placed in the narrow outer angle between the closely jammed canine and large premolar.

Dimensions of the type (the italicised measurements taken in the flesh): -

Forearm 50 mm. (F 51).

Head and body 57; tail 29; ear 25; third finger, metacarpal 36.5, first phalanx 16; lower leg and foot (c. u.) 30.

Skull: greatest length to front of canines 22.2; naso-occipital length 18.7; palatal length 3; front of canine to back of m^3 8.5

Hab. Kabwir, Bauchi Plateau, Northern Nigeria. Alt, 2500 feet.

Type. Adult male. B.M. no. 13.2.5.1. Original number 45. Collected 14th November, 1912, and presented by Mr. J.C. Fox, of the Cambridge University Mission. Two specimens, male and female.

This is evidently a western representative of *R. deckeni*, but the difference in the general size of the teeth and the reduction of the small premolar prevent my treating it as a subspecies of that bat, from which it is most readily distinguished by its much paler colour.

The British Museum possesses a good example of *R. deckeni* from Tanganiko, near Mombasa, collected and presented by Mr. A. Blayney Percival.

Rhinolophus fumigatus Rüppell, 1842

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Rhinolophus fumigatus (Rüppell).

Diagn. *Rhinolophus* alis octava parte majoribus *Rhinolophus* clivoso, (cui posthemale auriculisque persimilis) vellere villosissimo, colore cinerascens fuscus.

Der fleischige Nasenaufwuchs ist bei dieser Art ganz wie bei *Rhinolophus clivosus*, nämlich zwei über einander gestellte, nach oben einfach zugespitzte Hautlappchen, unten durch einen hufeisenförmigen Fleischausschwuch begrenz; der seitliche Rand der Ohren ist bei beiden gleich ausgeschweift, und der untere Rand derselben halb elliptisch aufwärts umgebogen; auch sind die Verhältnisse der einzelnen Körpertheile so ziemlich die nämlichen, nur ist bei der Schoënsen Art der Schwanz kürzer, dagegen der Vorderarm und die ganze Flugweite merklich grösser, wie aus nachstehender Zusammenstellung ersichtlich ist;

Rhinoloph. clivosus. / *Rhinoloph. fumigatus.*

Zoll. Lin. / Zoll. lin

Ganze Körperlänge von der Nasenspitze bis zum Schwanzende: 3 3 / 3 4

Schwanz besonders gemessen: 1 - / - 10

Verticalhöhe des Ohres: - 8 / - 9

Länge des Vorderarms: 1 8 / 1 11

Grösste Flugweite: 9 9 / 11 -

Daumenglied (ohne Nagel): - 2 / - 2

Tibia: - 8 / - 10

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Der ganze Körper ist dick mit einem ziemlich langen, dunkelrauchgrau gefärbten Pelz bedeckt, wodurch sich der Gesamthabitus dieses Thiers gleichfalls von *Rhinolophus clivosus* unterscheidet.

Vaterland: das Reich Schoa, südlich von Abyssinien.

Rhinolophus fumigatus exsul K. Andersen, 1905

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Diagnosis. - Apparently similar in all essential characters to *Rh. fumigatus* (= *Antinorii*); but decidedly smaller. Forearm 49.5 mm.

External characters. - To judge from a dried skin, the shape of the nose-leaves and ears is the same as in *Rh. fumigatus*. The lateral margins of the sella bear the dense clothing of long hairs characteristic of the *fumigatus*-group of species; the hairing on the front face of the sella seems to be sparser and shorter than in the typical form. General colour above dark greyish "drab" darkest on the hinder part of the back, brighter drab on the underside of the body; base of hairs, throughout, dark smoke-grey.

Measurements. - Under the next form.

Type. - Ad. (skin). Kitui, British East Africa, 3500 ft., Jan. 3rd, 1901; collected and presented by Mrs. H. Hinde. B. M. no. 1.5.6.3. Original no. 68. The type is the only specimen known to me.

Distribution. - Probably the whole of the Ukamba district.

Remarks. - This is evidently a southern offshoot of *Rh. fumigatus*, cut off from the (Abyssinian) main stem of the species, and developed into a distinct race. From *Rh. Deckeni*, an inhabitant of nearly the same regions, it is easily distinguished by the longer and differently shaped nose-leaves, by having the margins of the sella clothed with long straight hairs, by the much smaller size (forearm in *Rh. Deckeni* 54.7 mm.), by having the 4th metacarpal only about 2 mm. longer than the 3rd (in *Rh. Deckeni* 5 mm. longer), by the comparatively very short 2nd phalanx of the 5th finger, &c. I have not seen a skull of this form; it will, no doubt, be found to have lost both the lower p_3 and the upper p^2 , agreeing in this respect with *Rh. fumigatus* and differing from *Rh. Deckeni*.

Rhinolophus Geoffroyii A. Smith, 1829

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Rhinolophus Geoffroyii, mihi.

R. suprà ligneo-fuscus ; subtùs sublignicolor, membranis nigris, interfemorali transversè venosâ; caudâ vix præter apicem porrectâ; superiore membranæ nasalis lobo suprà acuminato.

Colour above wood brown, beneath lighter; membranes black, interfemoral one veined longitudinally, and the tip of the tail just extending beyond its posterior margin; upper lobe of nasal membrane pointed above; eyes small and nearly in front of the inner edges of auricles; ears large and patulous, nearly as long as the head, pointed, and each with an inferior lobe towards the base of the outer margin, which is separated from the principal by a deep intervening notch: interfemoral membrane semicircular behind. Length from the tip of nose to com-

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mencement of interfemoral membrane, three inches; length of latter, one inch; expanse of wings, thirteen inches. Inhabits the whole of South Africa.

This species decidedly differs from both the *Rhinolophus affinis*, and *Rhinolophus minor*, of Horsfield, not only in size and colour, but also in the depth of the notch in the outer edge of the auricle, as well as in the form of the nasal appendages, &c.

Rhinolophus Hildebrandti eloquens K. Andersen, 1905

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Diagnosis. - A small race of *Rh. Hildebrandti*. Forearm 57.7-60.5 mm.

Wings. - *Rh. Hildebrandti* is among the largest species of

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its genus; *Rh. H. eloquens* barely surpasses *Rh. Ferrum-equinum*. The reduction in size affects mainly the forearm and metacarpals, far less the phalanges. In the length of the forearm the largest specimens of *Rh. H. eloquens* fall short of the smallest individuals of the typical form; this character, therefore, evidently affords a most reliable mark of distinction between the two races. As to the metacarpals, maximum measurements of *Rh. H. eloquens* approach, very closely, minimum measurements of *Rh. H. typicus*.

Colour. - *Rh. Hildebrandti* (4 skins) is dark greyish "drab" on the upperside, bright drab beneath. *Rh. H. eloquens* (3 skins) is tinged with "wood-brown" above, next to "broccoli-brown" beneath. Base of the hairs, on the upper and under sides, dark smoke-grey. Immature individuals (1 skin) are slightly duller in colour than adults.

Skull and dentition. - All the measurements markedly smaller than in typical *Rh. Hildebrandti*. The length of the upper and lower tooth-rows, as being almost quite independent of the individual variation, is the best character by which to distinguish between the skulls of the two forms (conf. measurements below). In *Rh. Hildebrandti* (6 skulls) the upper p^2 is nearly always present and easily perceptible; the lower p_3 very minute, but generally not completely wanting. In *Rh. H. eloquens* p^2 is exceedingly small, hair-fine, even under a strong lens; p_3 (as it appears from examination of four skulls) almost always wanting, even in immature specimens.

Type. - Ad. (sk.), unsexed. Entebbi, Uganda; collected and presented by F. J. Jackson, Esq. B. M. no. 99.8.4.4. Three other specimens, from the same locality, are preserved in the Museum.

Distribution. - Probably the whole of the district surrounding Albert, Albert Edward, and Victoria Nyanza.

Remarks. - The present form is of great interest from a phylogenetic no less than from a zoogeographical point of view. *Rh. Hildebrandti* and *Rh. Fumigatus* (= *Antinorii*)* form a well-defined group of species, agreeing in the general form of the ears, in the size and shape of the horseshoe, the sella, and the posterior connecting-process, in having the margins and front face of the sella clothed with long hairs, in the general form of the wings, the comparative shortness of the tail, and the more essential characters of the skull

* Together with *Rh. Aethiops*; but this species, although extremely interesting as the hitherto only known West-African representative of the group, has a more isolate position, differing much more from the two other species than these latter inter se.

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and dentition. Until now, however, the very large *Rh. Hildebrandti* and its small counterpart, *Rh. Fumigatus*, have been without any intermediate link, widely separated in size as well as in geographical distribution. *Rh. H. eloquens* greatly reduces the gap between the two species; in all the points in which *Rh. H. eloquens* differs from *Rh. Hildebrandti typicus*, it approaches *Rh. Fumigatus* - in size, in colour, in dentition. *Rh. Hildebrandti* is decidedly the most primitive form: the upper p^2 is distinctly perceptible, the lower p_3 only occasionally wanting; in the Uganda race p_2 is markedly reduced in size, p_3 almost always

completely lost; in the northern *Rh. Fumigatus* p^2 is but very rarely present, p_3 never. Uganda, the typical locality of *Rh. H. eloquens*, is, so to speak, "on the way" between the areas of *Rh. Hildebrandti* (from Mazoe to Mombasa) and *Rh. Fumigatus* (S. Somali and Abyssinia). It is not improbable that, when the region between Uganda and Abyssinia is better explored, we shall have to regard *Rh. Hildebrandti* and *Rh. Fumigatus* no longer as distinct "species," but rather as *peripheral forms* of one species, bound together by races intermediate in characters and in geographical distribution.

The subjoined table of measurements gives the difference in size, from the smallest form of *Rh. Fumigatus* to the largest form of *Rh. Hildebrandti*. If I had been arranging the forms from a phylogenetic point of view, the order would have been just the reverse (the names of the races read from right to left).

[table removed - eds.]

Rhinolophus hildebrandti perauritus de Beaux, 1922

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Somalia mer., territ. dei Rahanuin, 10.XI.11. *Int. in alc.* C. E. 12238; 3 FF; *Cotipi*. Le dimensioni somatiche superano di poco i maxima segnati da Andersen (Ann. Mag. Nat. Hist. XV, 1905, p. 76) per il *Rh. fumigatus*. Le orecchie e le membrane fogliacee nasali sono però più grandi. Le dimens. dei denti sono quelle del *Rh. hildebrandti eloquens*, And. (l.c.). P_2 ben percettibile. Lungh. della testa in carne mm. 25; test + tronco 57; coda 31; avambraccio 54-55; 3° metacarpo 39; 4° mtc. 41-41.5; 5° mtc. 41-42. Padiglione: lungh. mssm. 20; largh. in posiz. natur. 18; lungh. membrana fogliacea nas. 20; lungh. del ferro di cavallo 9-10; sua largh. 11-12; lungh.

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$C-M_3$ 8.5-9; $C-M_3$ 9.5-10. - *Colore*: Parti super. grige chiare (light drab, Ridgw. XLVI); singoli peli grigi chiarissimi nei $\frac{5}{6}$ basali (pale drab gray, ibid.); drab nel $\frac{1}{6}$ apic.. Parti infer. grige biancastre (pale olive buff, Ridgw. XL). Membrana alare bruna fuliginosa.

Annotaz.. Fui lungamente in dubbio se attribuire questo *Rh.* alla forma *fumigatus fumigatus*, And.; ma dopo accurati confronti con *fumig.* tipici determ. dall'a. della spec., e con *hildebrandti*, Pet. dovetti decidermi a farne una nuova sottospec., la quale costituisce daltronde un ulteriore appoggio alla supposiz. già espressa dallo stesso And. nel 1905, *che hildebrandti e fumigatus costituiscano una specie unica*.

Rhinolophus Hildebrandtii Peters, 1878

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(Taf. 1. Fig. 1.)

Diese Art schliesst sich durch die Form des Nasenbesatzes am nächsten an *Rh. fumigatus* und *aethiops* an, ist aber grösser, die grösste unter den bisher aus Africa bekannt gewordenen Arten. Das Hufeisen ist a am Rande einfach abgerundet, ungekerbt, vorn eingeschnitten, neben dem Einschnitte wulstig, aber ohne zahnartigen Vorsprung. Jederseits neben dem Hufeisen befindet sich, wie bei *Rh. aethiops*, nur vorn eine rudimentäre Falte. Die vordere Seite des Sattels ist etwas biscuitförmig eingebuchtet und überragt etwas den vorderen Theil des oberen bogenförmigen Sattelrandes; der ganze Sattel ist höher und Länger als bei *Rh. aethiops*. Der ganze Nasenbesatz ist mehr oder weniger dicht behaart. Die Ohren sind gross und breit, am Ende zugespitzt, aussen mit 10 Querfalten. Die Behaarung ist weich und reichlich, einfarbig, oben dunkelblond, unten graubraun.

Mafse nach getrockneten aufgeweichten Bälgen:

Meter

Totallänge ungefähr: 0.095

Kopflänge: 0.029

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Ohrhöhe: 0.029

Ohrbreite: 0.022

Länge des Nasenbesatzes: 0.022

Breite des Nasenbesatzes: 0.012

Höhe des Sattels: 0.0057

Länge des Sattels: 0.001

Vorderarm: 0.062

L.1.F. Mh. 0.0015; 1.GL. 0.0035; 2.GL. 0.003; 0.011

L.2.F. - 0.0415;) 0.005; 0.045

L.3.F. - 0.013; - 0.019; - 0.035; Kpl. 0.0035

L.4.F. - 0.015; - 0.011; - 0.022; - 0.002

L.5.F. - 0.047;) 0.0135; - 0.019; - 0.002

Schwanz: 0.034

Unterschenkel: 0.0275

Sporn: 0.021

Fuss: 0.0175

Aus Ndi (Taita).

Rhinolophus hilli Aellen, 1973

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Type. - F adulte, en alcool. Uwinka, Préf. Cyangugu, Rwanda, alt. 2300 m, 25.8.1964; coll. U. Goepel. Zoologisches Muséum der Universität Zürich, n° 481.

Derivatio nominis: en l'honneur de John Edwards Hill, du British Muséum, dont les remarquables travaux ont beaucoup contribué à la connaissance de la systématique des chiroptères.

Diagnose. - Appartient au groupe *philippinensis* et est voisin de *ruwenzorii* Hill. Diffère de ce dernier par: a) une taille générale plus petite, b) le connectif de la feuille nasale bien développé, c) le crâne à boîte crânienne et le rostre plus larges.

Description. - Les dimensions externes et crâniennes sont un peu plus faibles que celles de *ruwenzorii*: inférieures en moyenne de 5 à 10 %. L'avant-bras mesure 53 mm, alors que chez *ruwenzorii*, il est de 55.5 mm (56 et 57) pour la F, et un peu plus faible pour le M.

Les oreilles, remarquablement grandes, sont très semblables à celles de *ruwenzorii*. Comme chez ce dernier, le bord antérieur de l'antitragus n'est pas concave et ne porte pas de sec à sa base (caractères de *maclaudi*).

La feuille nasale, qui ressemble en général à celle de *ruwenzorii*, en diffère surtout par le connectif. Dans notre nouvelle espèce, la selle (partie basale) est en forme de coupe (calice), moins évasée que chez *maclaudi*. Chez

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ruwenzorii, les bords latéraux de la partie basale de la selle seraient plus étalés (cf. fig. 2, in Hayman, 1960); mais chez le spécimen de Butembo que j'ai examiné, la selle ressemble beaucoup à celle de *hilli*. Ce caractère semble donc variable; il est d'ailleurs fort possible que sur l'animal vivant on puisse observer les deux positions cup-like et wing-like, expansions en rapport avec les émissions d'ultrasons: étalées, les "ailes" recouvrent les orifices des narines, repliées en calice, elles les dégagent un peu. Le procès vertical de la selle est semblable chez les deux espèces, de même que le fer-à-cheval et la lancette. Par contre, le connectif est nettement différent. Obsolète chez *ruwenzorii*, il est parfaitement développé chez *hilli*, entre le procès vertical de la selle (dont il n'atteint pas le sommet) et la lancette. Ce caractère me paraît important et justifie la séparation en deux espèces. Sur les côtés du museau, en avant, il y a une foliole secondaire (secondary leaflet, cf. Rosevear, 1965, fig. 49) de chaque côté, aux 3/4 cachée par le fer-à-cheval, et non rattachée directement au reste de la feuille nasale. Cette foliole fait défaut chez *maclaudi*, mais est présente chez *ruwenzorii*, bien que non encore signalée.

La lèvre inférieure présente un sillon médian net et deux échancrures latérales.

Le patagium s'insère au milieu du tarse.

Les longueurs respectives des métacarpes et des phalanges des 3e, 4e et 5e doigts correspondent exactement au schéma donné par Andersen pour *philippinensis* (1905: 247), c'est-à-dire que l'aile est de structure très primitive. Le 3e métacarpe est très peu raccourci (environ 90 % du 5e), le 4e métacarpe est à peu près égal au 5e. Ces deux caractères correspondent à ceux de la section *philippinensis*. Par contre, un certain allongement des 2es phalanges, et surtout de celle du 3e doigt, est un caractère propre aux sections *trifolius* et *sedulus* (du groupe *philippinensis*). La structure de l'aile de *ruwenzorii* est tout à fait semblable à celle de *hilli*.

Les plis palataux sont du même type que celui décrit chez *ruwenzorii*, mais ils sont tous, sauf peut-être le 6e, séparés au milieu. La fourrure est douce et bien fournie, comme chez *ruwenzorii*. La coloration est quelconque et très uniforme: brun foncé dessus et dessous, les oreilles et le patagium un peu plus clair.

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Le crâne présente une forme et des proportions semblables à celles du *ruwenzorii*. Toutefois, il est relativement plus large au niveau du rostre: le rapport largeur M³-M³ sur longueur C-M³ (alv.) est plus grand que 1 (1.05) chez *hilli*, alors qu'il est inférieur ou égal à 1 chez *ruwenzorii* (0.92 à 1.0), moyenne de 3 spécim. 0.95). La boîte crânienne est aussi plus large: le rapport largeur mastoïde sur longueur basale, utilisé par Hill (7), est de 0.6 chez *hilli* et seulement de 0.57 à 0.58 (2 spécim.) chez *ruwenzorii*. Enfin, la largeur zygomatique est pratiquement égale à la largeur mastoïde chez *hilli*, alors qu'elle est plus faible chez *ruwenzorii*.

mesures en mm: *R. hilli* type F; *R. ruwenzorii* type F, M Butembo

tête et corps: 60; 67, 66

avant-bras: 53; 57, 56

3e doigt, métacarpe: 37.5; 40, 40

- 1ère phalange: 17; 18.5, 17.5

- 2e phalange: 29.5; 32, 30

4e doigt, métacarpe: 41.5; 44, 43

- 1ère phalange: 11; 12, 11.5

- 2e phalange: 19; 20, 19.5
 5e doigt, métacarpe: 42; 46.8, 43.5
 - 1ère phalange: 12.5; -, 13
 - 2e phalange: 18; -, 18
 queue: 30; 31, -
 tibia: 23; 23.5, 23
 pied, sans griffe: 12.5; 12, 12
 oreille, longueur: 29; 32.5, 32
 - largeur: 22; -, 26
 feuille nasale, long; 22; -, (18)
 fer-à-cheval, larg; 12; -, (13)
 crâne:
 longueur totale: 23.8; 25.9, -
 long, condylobasale: 21.1; 23.3, -
 longueur basaie (C): 18.0; 20.7, -
 pont palatal: 3.4; 3.6, 3.4
 largeur zygomatique: 10.6; 11.2, -
 largeur mastoïde: 10.8; 12.0, -
 larg. interorbitaire: 2.7; 2.6, 2.7
 largeur M³-M³: 8.0; 8.2, 7.5
 largeur C-C: 5.7; 5.9, 5.4
 longueur C-M³ (alv.): 7.6; 8.2, 7.9
 longueur C-M³ (cour.): 8.1; 8.9, 8.3
 longueur P⁴-M³: 6.0; 6.3, 6.2
 long, mandibule: 15.1; -, 15.9
 longueur C-M₃: 8.3; -, 8.7

Le pont palatal comme chez *ruwenzorii* est remarquablement long; il représente environ 45 % de la longueur de la rangée dentaire C-M³. C'est un caractère propre au groupe *philippinensis*.

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Au maxillaire supérieur I¹ a une couronne plate, usée; il est fort possible que cette dent était bifide à l'état jeune, comme chez *ruwenzorii*. P² est situé au bord externe de la rangée dentaire et sépare C de P⁴. Très petite, cette dent n'arrive pas à la hauteur du cingulum de C. Chez le type de *ruwenzorii*, P² est plus développé, mais c'est un caractère quelque peu variable, car chez *ruwenzorii* de Butembo, la dent est moins développée. P⁴, M¹, M², et M³ sont semblables chez *hilli* et *ruwenzorii*.

Au maxillaire inférieur, I₁ est bicuspidé, mais probablement tricuspidé à l'origine (disparition par usure de la cuspidé centrale), plus petit que I₂. I₂ est bicuspidé à droite et tricuspidé à gauche. P₃ est minuscule et déporté en dehors de la rangée dentaire, de sorte que P₂ est en contact avec P₄; chez *ruwenzorii*, P₃ sépare encore un peu P₂ de P₄. P₂ est plus petit que la 1/2 de P₄.

Il est évident que ces constatations, basées sur un très petit nombre de spécimens, ne seront peut-être pas confirmées lorsqu'on disposera de plus de matériel.

Remarques. - Dans sa revision des *Rhinolophus* du groupe *philippinensis*, Andersen n'avait connaissance pour l'Afrique que de l'espèce *maclaudi* Pousargues, qu'il incluait sans hésiter dans ce groupe d'espèces orientales. Laurent, lui, concluait que *maclaudi* constituait un type africain archaïque isolé, n'entrant pas dans l'un des groupes d'Andersen, n'ayant donc pas d'affinités avec des espèces d'Asie. On connaît maintenant 9 spécimens de *maclaudi*, provenant tous de la Guinée, aux environs de Conakry.

Il me semble que les conclusions d'Andersen restent valables, surtout depuis que l'on connaît l'existence d'une deuxième espèce de ce groupe en Afrique, soit *Rhinolophus ruwenzorii*, décrit par Hill en 1942 et retrouvé à plusieurs reprises dans la région du Mt Ruwenzori et de l'Ituri. Il y a actuellement huit spécimens de *ruwenzorii*: le type F de la Butahu Valley, 1 F de R. Mahoma, 3 MM du Mt Hoyo, 3 spécimens de Butembo, du Mt Hoyo et de la cave de Matata. Tous ces spécimens proviennent du Zaïre (ancienne Province du Kivu), sauf celui de la R. Mahoma qui est de l'W de l'Ouganda.

L'étude morphologique indique des affinités certaines avec les espèces orientales du groupe *philippinensis*. La nouvelle espèce décrite ci-dessus vient encore confirmer ces affinités. A mon avis, il s'agit de types archaïques reliques, maintenant isolés, mais ayant une origine commune avec les espèces orientales du groupe *philippinensis*. Andersen a subdivisé son groupe en trois sections: aucune des espèces africaines ne cadre exactement dans l'une de ces sections, mais elles répondent parfaitement à la définition même du groupe.

Rhinolophus hilli est nettement plus proche de *ruwenzorii* que de *maclaudi*. Il est possible, qu'à la lumière de nouvelles observations basées sur un matériel plus abondant, on doive considérer le *hilli* seulement comme une sous-espèce de *ruwenzorii*. Cela n'a d'ailleurs pas de grande importance. Par contre, il me semble impossible d'entrer dans les vues de Hayman et Hill, qui pensent que *maclaudi* et *ruwenzorii* pourraient être conspécifiques. La structure et la forme différentes des feuilles nasales et des oreilles, pour ne parler que de ces formations externes, ne peuvent pas être des caractères subspécifiques.

Biologie. - Les rares spécimens de *maclaudi* et de *ruwenzorii* dont on connaît l'origine précise, proviennent de grottes. Le type de *hilli* a été capturé au filet dans une forêt de montagnes. Il s'agit d'une F adulte portant un embryon de 15 mm.

Rhinolophus keniensis Hollister, 1916

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Type from west side of Mount Kenia, British East Africa, at 7,000 feet altitude. United States National Museum No. 166352, adult male in alcohol with skull removed. Collected August 27, 1909, by Edmund Heller. Orig. No. 1154.

Description. - A small member of the *Rhinolophus augur* group, differing from *R. a. zambesiensis* in the small size of the nose-leaf (greatest width of horse-shoe, 6.8; compared with 8.0-8.3 in *zambesiensis*), shorter forearm, and shorter tibia. Skull somewhat larger, with slightly more robust teeth, than in *zambesiensis*.

Measurements of type. - Forearm, 52 mm.; tibia, 21; greatest length of ear from anterior margin, 18.9; greatest width of ear, 12.2; third finger metacarpal, 34.8; first phalanx, 17.4; fourth metacarpal, 39.5; first phalanx, 11.3; fifth metacarpal, 40.3; first phalanx, 12.8. Skull: Greatest length, 22.8; condylobasal length, 20.4; zygomatic breadth, 12; postorbital constriction, 3; mastoid breadth, 10.7; mandible, 15. Teeth: Canine to m^3 , 8.6; breadth across upper canines, 6.6; greatest breadth across upper last molars, 8.4; lower canine to m_3 , 9.

Remarks. - This new bat is readily separable from all the other species of *Rhinolophus* known from British East Africa by the combination of narrow horse-shoe, hairless sella, and position of small upper premolar entirely without the tooth row. It is doubtless a northern representative of *R. augur*, and perhaps it intergrades directly into *R. a. zambesiensis*. Doctor Lönnberg and Mr. Oldfield Thomas have recorded *R. a. zambesiensis* from Kilimanjaro, but I am not aware that a member of the group has up to now been noted in British East Africa.

Specimen examined. - One, the type.

Rhinolophus landeri guineensis Eisentraut, 1960

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Material: 10 Exemplare, 1 M, 3 FF (Tgb. Nr. 27-30), 5.X.1956, nahe Tahiré, am Fuße des Kelesi-Plateaus. - 1 M, 1 F (Tgb. Nr. 121, 126), 19.XI.1956, nahe Nyembaro am Salung-Plateau, 12 km westlich Kolente. - 2 MM, 2 FF (Tgb. Nr. 133, 136-138), 23.XI.1956, bei Darsalam am Kankuja-Plateau, 8 km westlich Kolente.

Diese neue Form stimmt in den wesentlichen Merkmalen mit von mir am Kamerunberg gesammelten Stücken von *Rh. landeri landeri* Martin überein, zeichnet sich aber durch größere Körper- und Schädelmaße aus und zeigt darin gewisse Übereinstimmung mit der im Kongogebiet und in Ost- und Südafrika lebenden Subspezies *lobatus* Peters. Ferner sind die bei den MM vorkommenden Drüsenhaarbüschel unter der Achsel nicht bräunlich getönt, sondern rein weiß. Es scheint daher berechtigt, diese westlichste Form als besondere Subspezies abzutrennen. Ich schlage für sie den Namen *guineensis* vor. Soweit ich die Literatur überblicke, sind die von KNORR gesammelten Stücke der Erstnachweis der Art für Guinea.

Typus: M ad. (Tgb. Nr. 28), 5.X.1956, Tahire am Fuße des Kelesi-Plateaus, etwa 500 m Meereshöhe, KNORR leg., Staatliches Museum für Naturkunde Stuttgart, Katalog-Nr. 6103. Kopf-Rumpf-Länge: 50 mm, Schwanz: 26 mm, Ohr: 17 mm, Hinterfuß: 8.5 mm, Unterarm: 44.3 mm, gesamte Schädellänge: 20.6 mm, Jochbogenbreite: 10 mm, maxillare Zahnreihe: 7.3 mm, Mandibellänge: 13.3 mm, mandibulare Zahnreihe: 8 mm.

In Tabelle I sind die Körper- und Schädelmaße für *guineensis* angegeben und den entsprechenden Maßen der Nominatrasse und der Rasse *lobatus* Peters (= *axillaris* J. A. Allen, vgl. HAYMAN in SANDERSON) gegenübergestellt. Fast sämtliche Durch-

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schnittswerte von *guineensis* liegen höher als die von *landeri*, dagegen berühren oder überschneiden sich die Maximum- und Minimumwerte sehr häufig. Besonders auffallend ist die größere Länge der Tibia und ferner die höheren Werte für die gesamte Schädellänge, die Mastoidbreite, die Mandibellänge und die Länge der oberen und unteren Zahnreihe. In Abb. 1 kommen die unterschiedlichen Schädelmaße zum Ausdruck. Bei der Gegenüberstellung der Guinea-Rasse mit der Rasse *lobatus*, für die in der Literatur nur verhältnismäßig wenige Angaben vorliegen, ist die weitgehende Übereinstimmung der Unterarm- und Tibiallänge festzustellen.

In Übereinstimmung mit der bedeutenderen Körpergröße sind auch die an den frishtoten Tieren festgestellten Körpergewichte bei *guineensis* mit 9.1 (8 - 11) g wesentlich höher als bei *landeri* (von Kamerun) mit 6.2 (5 - 7) g.

Entsprechend der größeren Schädelmaße bei *guineensis* gegenüber *landeri* ist auch das Gebiß kräftiger entwickelt. Hervorzuheben ist die relative Kleinheit von P_1 , der ähnlich wie bei *alcyone* höchstens die halbe Länge von P_3 erreicht, während er bei meinem Material von *landeri* aus Kamerun (EISENTRAUT 1956, vgl. auch die Angaben von AELLEN bis zu $\frac{2}{3}$ der Höhe von P_3 ansteigt. Im übrigen aber entsprechen die Zahnproportionen denen der Nominatrasse.

Weitgehende Übereinstimmung zwischen *guineensis* und *landeri* besteht hinsichtlich der Ausbildung des Nasenaufsatzes, lediglich übertreffen bei erstgenannter Form die absoluten Maße - entsprechend den allgemein größeren Körpermaßen - die von *landeri*. Während z. B. die Breite des Hufeisens bei einem Alkoholstück von *guineensis* 9 mm beträgt, erreicht sie bei *landeri* nur etwa 7 mm.

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Die Färbung der mir vorliegenden Guinea-Stücke läßt oberseits eine Variation von dunklerer zu hellerer graubräunlicher Tönung erkennen (nach RIDGWAY von Clove Brown über Olive Brown zu Tawny Olive). Ein Stück (Nr. 136) zeigt eine blaßrotbraune Felfärbung, jedoch ist auf der hinteren Rückenmitte eine sich stark abhebende, dunkelgraue Haarpartie vorhanden (vielleicht Jugendmauser?), bei einem weiteren Stück (Nr. 126) ist diese Färbungstendenz in ganz schwach angedeuteter Form ebenfalls zu erkennen. Die Unterseite zeigt die übliche fahle Aufhellung.

Bei den bisher bekannten Formen der Art wurde eine charakteristische rotbraune Färbung der Achseldrüsenhaare (soweit vorhanden) bei den MM hervorgehoben. Im Gegensatz dazu zeigen drei MM von *guineensis* zwar ebenfalls die Ausbildung eines solchen Haarschopfes, dieser ist aber reinweiß und hebt sich von den umliegenden grauen Haarpartien scharf ab. Einem M (Nr. 39) fehlt dieses Merkmal ganz (vielleicht jugendliches Exemplar).

Das F Nr. 133 (Alkohol) besitzt deutlich ausgeprägte, gleichlange Afterzitzen; sein Uterus ist in völligem Ruhezustand, die Uterushörner jedoch sind gut entwickelt, und zwar das rechte etwas stärker als das linke.

Die von Herrn KNORR gesammelten Stücke wurden während der Tagesruhe in Felshöhlen erbeutet, wo sie zusammen mit anderen Fledermausarten (*Rhinolophus denti*, *maclaudi*, *foxi*, *Hipposideros caffer*, *jonesi*, *Nycteris aethiopica* und *gambiensis*) vorkommen. Die Fundplätze befinden sich nahe den Orten Tahiré, Nyembaro und Darsalam in einem etwa 150 km von der Küste (Conakry) entfernt liegenden Gebiet von Südkindia.

Rhinolophus lobatus Peters, 1852

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Tafel IX. Tafel XIII. Fig. 16, 17.

Rh. colore opace umbrino, subtus pallidior; alis nigrofuscis; auriculis elongatis, ellipticis, acuminatis, parum excisis, lobulo alto; plica transversali prosthematici superiore distincte lobata; deutibus molaribus superioribus utrinque quinis, inferioribus senis; metacarpo digiti quarti longiore quam digiti quinti; alis usque ad tarsum porrectis.

Longitudo tota 0,085; caudae 0,026; antibrachii 0,045; volatus 0,290.

Habitatio: Africa orientalis, Sena, Tette, 17° Lat. Aust.

Die Ohren sind elliptisch, zugespitzt, um ein Fünftel kürzer als der Kopf; der Einschnitt zwischen der Ohrmuschel und dem Ohrklappen ist gering; der Ohrklappen selbst ist hoch, an beiden Ecken gleichmäßig abgerundet, an seiner äußeren Fläche so wie die Ohrmuschel über zwei Drittel behaart. Das Hufeisen zeigt drei concentrische Falten; die vordere Querfläche des Sattels ist etwas unter der Mitte zusammengedrückt, biscuitförmig; die vor der Lanzette sich erhebende Spitze ist dreieckig, so lang wie die Querfläche; die Lanzette ist länger als breit, nach unten gleichmäßig, ohne vorspringende Seitenlappen, erweitert; die Scheidewand zwischen dem mittleren und oberen Zellenpaar ist zu beiden Seiten lappenartig abgesetzt. Auf den vorspringenden Falten des ganzen Nasenbesatzes finden sich zerstreute borstenförmige Haare. Das Maul ist abgerundet, bis unter den hintern Rand des Hufeisens gespalten; die Lippen haben in der Mitte einen nackten wulstigen Rand; die innere Fläche der Oberlippe ist jederseits durch einen großen warzigen Vorsprung ausgezeichnet. Die kleinen Augen stehen doppelt so weit von der Schnauzenspitze wie von den Ohren entfernt. Am Gaumen sieht man sieben quere Schleimhautfalten, drei vordere größere, entfernter von einander stehende, und vier hintere, weniger hervorragende und näher an einander gedrängte; die letzte derselben ist grade und ungeliebt, die übrigen bilden doppelte in der Mitte zusammenstoßende Bögen. Die Behaarung des Körpers ist an der Rückenseite länger als an der Bauchseite. Die Pha-

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langen des Daumens sind ganz frei. Das Mittelhandglied des vierten Fingers ist das längste von allen, wie bei *Rh. hippocreps*, Hermann, und das des dritten Fingers das kürzeste, wie bei allen bis jetzt bekannten Arten der Gattung *Rhinolophus* im engeren Sinne. Die erste Phalanx des vierten Fingers ist sehr kurz, weniger als halb so lang wie die erste Phalanx des dritten Fingers. Die zweite Phalanx des fünften Fingers ist um die Hälfte länger als die erste desselben. Alle vier unbenagelten Finger tragen ein kurzes knorpeliges Endglied. Die Flughäute gehen bis zur Fußwurzel herab; sie sind oben nackt, an der Bauchseite dagegen zwischen Oberschenkel und Oberarm mit zerstreuten langen Härchen bewimpert. Der Schwanz ist um ein Viertel länger als der Unterschenkel, und ragt mit seinem kurzen knorpeligen Endgliede aus der Schenkelflughaut hervor; das vierte und fünfte Endglied sind die längsten und von gleicher Größe. Die Schenkelflughaut ist oben und unten bis zum vierten Endglied mit zerstreuten Haaren bewimpert. Die Sporen sind nur sehr schwach. Die Nägel der Zehen sind ein wenig länger und gekrümmter als die Daumenkrallen.

Die Farbe des Rückens und der Halsseiten ist braun, wie die eines dunklen Umbers und die Bauchseite zeigt ganz denselben Farbenton, aber viel blasser und verwaschen. Die Haare sind sämtlich einfarbig, nur werden die Rückenhaare nach dem Grunde zu alle mählig immer blasser. Die kleinen Haarbüschel auf der Unterseite der Flughäute, welche selbst von schwarzbrauner Farbe sind, erscheinen weißlich. Die Ohren sind von der Farbe der Flughäute. Das Gesicht ist um das Hufeisen herum dunkler. Die Nägel sind gelblich.

Der Schädel (Tafel XIII. Fig. 16, 17), obgleich beträchtlich größer, nähert sich in seine Gestalt am meisten dem von *Rhinolophus hippocreps* Herm.; auch der Fortsatz des Unterkieferwinkels hat dieselbe horizontale Lage und fast rechtwinklig abgehende Richtung wie bei dieser kleinen europäischen Hufeisennase. Der kleine obere erste Lückenzahn ist aber nicht lang und spitzig, sondern ganz niedrig und ragt mit seiner stumpfen Spitze nicht über den Rand der benachbarten Zähne hervor, und im Unterkiefer findet sich zwischen den beiden falschen Backzähnen noch ein ganz kleiner nach außen gedrängter Zahn, so daß die Zahl der untern Backzähne sich im Ganzen auf sechs beläuft. Die Formel des Gebisses ist daher bei diesem Thiere $3\frac{2}{3}1\frac{1}{2}1\frac{1}{4}1\frac{2}{3}2\frac{3}{2}3 = 32$. Die Wirbelsäule besteht aus 7 Halswirbeln, 11 Rückenwirbeln, 7

Lendenwirbeln, 3 Kreuzbeinwirbeln und 11 Schwanzwirbeln. Die beiden ersten Rippen sind mit dem letzten Halswirbel und mit den beiden ersten Rückenwirbeln verwachsen, die vier folgenden verbinden sich durch normale Zwischenstücke mit dem Brustbein, und die siebente und letzte wahre Rippe steht durch ein sehr breites plattes Knochenstück mit demselben in Verbindung. Der Magen ist kugelförmig und geht zunächst in einen weiten Darm über, der sich gegen die Mitte seiner Länge plötzlich um die Hälfte verengert, und erst nahe vor seinem Ende sich wieder etwas erweitert. Die Länge des Darmcanals ist 145 Millim. Die Leber ist dreilappig, und trägt unter dem mittleren Lappen eine gestreckte hirnförmige Gallenblase. Die Milz ist $3\frac{1}{2}$ Mm. breit, und etwa dreimal so lang wie breit. Die Nieren sind ungelappt, bohnenförmig, an ihrem oberen Ende etwas dicker als um unteren. Die Ruthe des Männchens

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enthält einen 3 Mm, langen Knochen. Die Luftröhre bildet unter dem Kehlkopf eine kleeblattförmige blasige Erweiterung. Die Lungen bestehen jederseits nur aus einem einzigen unregelmäßig quadratischen Lappen, dessen Ecken sehr spitz ausgezogen sind.

Von dieser Art besitze ich vier Exemplare, Männchen und Weibchen, von denen eins aus Tette, drei aus Sena herkommen. Die aus Sena wurden unter dem vorstehenden Dache eines alten Gebäudes gefunden. Nach dem Inhalte des Magens und Darmcanals zu urtheilen, besteht ihre Nahrung aus Insecten.

Maße in Millimetern.

Fem.; Fem.; Mas.

Länge von der Schnauze bis zur Schwanzspitze: 80; 80; 85

Flugweite: 290; 295; 290

Länge des Kopfes: 20; 20; 20

Länge oder Höhe des ganzen Ohres: 16; 16; 16

Breite des Ohres: 12; 13; 12

Länge des ganzen Nasenbesatzes: 11; 11; 12

Breite des Nasenbesatzes: $7\frac{1}{4}$; 7; 7

Länge des Oberarms: $27\frac{1}{2}$; 27; 27

Länge des Vorderarms: $44\frac{1}{2}$; 46; 45

Länge d.1.Fg. (Mittelh. 4, 4, 4. 1.Gl. 3, 3, $3\frac{1}{2}$. 2.Gl. $1\frac{1}{2}$, $1\frac{1}{2}$, 2): $8\frac{1}{2}$; $8\frac{1}{2}$; $9\frac{1}{2}$

Länge d.2.Fg. (Mittelh. $30\frac{1}{2}$, 35, $33\frac{1}{2}$. 1.Gl. $\frac{1}{2}$, $\frac{1}{2}$, $\frac{1}{2}$): 34; $35\frac{1}{2}$; 34

Länge d.3.Fg. (Mittelh. 29, 32, $30\frac{1}{2}$. 1.Gl. 13, $14\frac{1}{2}$, $14\frac{1}{2}$. 2.Gl. 23, 25, 24. 3.Gl. $1\frac{1}{2}$, $1\frac{3}{4}$, 2): $66\frac{1}{2}$; $73\frac{1}{4}$; 71

Länge d.4.Fg. (Mittelh. 33, 36, 34. 1.Gl. $5\frac{1}{2}$, $6\frac{1}{2}$, 7. 2.Gl. $14\frac{1}{2}$, 16, 16. 3.Gl. 1, 1, 1): 54, $59\frac{1}{2}$; 58

Länge d.5.Fg. (Mittelh. 32, $34\frac{1}{2}$, $32\frac{1}{2}$. 1.Gl. 9, $9\frac{1}{2}$, $9\frac{1}{4}$. 2.Gl. 13, $14\frac{1}{4}$, $13\frac{1}{4}$. 3.Gl. 1, $1\frac{1}{4}$, $1\frac{1}{4}$): 55, $59\frac{1}{2}$; 57

Länge des Oberschenkels: 18; 19; 19

Länge des Unterschenkels: 18; 19; 19

Länge des ganzen Fusses mit den Krallen: $9\frac{1}{2}$; 10 10

Länge des Schwanzes: 24; 25; 26

Länge des Sporns: 10; 10; 10

Die in der Diagnose angegebenen Kennzeichen characterisiren diese Art hinreichende, um sie von den verwandten africanischen Formen zu unterscheiden. *Rh. capensis*, Lichtenstein, hat nur vier obere Backzähne, die Flughäute lassen einen Theil des Schienbeins frei und das Mittelhandglied des fünften Fingers wird nicht vom vierten überragt. *Rh. clivosus*, Rüppell, hat zwar dieselbe Form der Backzähne, aber die Farbe ist röthlich, die Form des Nasenbesatzes und das Verhältniß der Gliedmaßen verschieden. *Rh. fumigatus*, Rüppell, welche in der Größe mit unserer Art übereinstimmt, weicht durch Färbung, durch die noch größere Verlängerung des Vorderarms, und, da sie mit *Rh. clivosus* sonst übereinstimmen soll, auch in dem Verhältniß der Mittelhandglieder und in dem Bau der Lanzette von derselben ab.

[*Rhinolophus Macclaudi*](#) Pousargues, 1898

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Parmi les Mammifères nombreux et intéressants recueillis par M. le Docteur Macclaud dans l'île de Conakry, sur les côtes de la Guinée française, se trouvait un Cheiroptère du genre *Rhinolophus* présentant un ensemble de caractères qui le distinguent nettement de toutes les autres formes congénériques.

Cette nouvelle espèce est de grande taille, plus grande que toutes celles signalées jusqu'à présent sur le continent africain, car elle dépasse en dimensions le *R. Hildebrandti* (Pet.) de l'Afrique orientale et ne le cède à cet égard qu'au géant du genre, le Rhinolophe deuil, *R. luctus* (Tem.), des hautes terres de l'Inde continentale et archipélagique. Sa taille n'est pas le seul caractère qui la rapproche des grandes espèces indiennes pour lesquelles Gray avait créé les sous-genres *Aquias* et *Phyllotis*; elle leur ressemble

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aussi par le mode de conformation de la feuille nasale et en particulier de la selle ou partie centrale. Cette dernière peut être comparée, dans son ensemble, au pétale labellé de certaines fleurs: sa portion basilaire est cordiforme; chacune de ses deux ailes symétriques s'élève d'abord verticalement au-dessus de la cloison nasale, puis s'étale et s'épanouit latéralement et enfin s'incurve en haut vers son bord supérieur, de manière à ménager entre elles une cavité assez profonde en forme de coupe évasée, analogue à celle que l'on remarque chez les *R. mitratus* (Blyth) et *R. philippinensis* (Wat). La partie supérieure de la selle, bien développée en longueur comme en largeur, dessine une sorte de languette dirigée en haut et en avant. Un autre caractère spécial à cette nouvelle espèce consiste dans le mode de structure de la partie antérieure de la feuille nasale ou fer à cheval, qui, sur son bord interne longeant extérieurement chacun des orifices nasaux, se redresse verticalement en une

lame membraneuse assez élevée, qui vient doubler de chaque côté, comme un calice, l'espèce de corolle formée par la portion basilaire de la selle. Enfin je signalerai tout particulièrement la longueur démesurée des conques auditives, dont les dimensions relatives sont énormes et qui, au point de vue absolu, dépassent même en longueur, celles du *R. luctus*, le mieux pourvu sous ce rapport.

La diagnose de cette espèce remarquable peut être résumée de la façon suivante:

La partie terminale ou supérieure de la feuille nasale, bien développée,

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triangulaire, remonte presque verticalement jusqu'entre les oreilles et présente latéralement les trois replis et les vacuoles ordinaires. La partie antérieure horizontale de la feuille ou fer à cheval est large, légèrement échancrée au milieu de son bord antérieur et surplombe toute la lèvre supérieure; le long de son bord interne attenant aux narines elle se relève de chaque côté en une lame verticale très développée dont les amorces descendent en s'atténuant graduellement vers son bord antérieur.

La portion basilaire de la selle est cordiforme, creusée en coupe, beaucoup plus large que sa portion supérieure verticale (section b' du *Synopsis de Dobson*), chacune de ses ailes s'étalant latéralement et se relevant ensuite verticalement. La partie supérieure de la selle a la forme d'une languette dressée, assez longue, et conserve la même largeur jusqu'à son sommet, arrondi et recourbé en avant. La lame membraneuse verticale (*posterior connecting process*) qui rattache cette languette à la partie supérieure de la feuille est concave, peu élevée et n'atteint même pas son sommet. La lèvre inférieure ne présente qu'un seul sillon médian peu profond. Les oreilles sont énormes, d'un quart plus longues que la tête et hors de proportion avec la taille de l'animal; leur bord interne, garni près de la base d'un duvet rare et court, est régulièrement convexe jusqu'au sommet assez peu aigu, légèrement arrondi et dirigé en dehors; leur bord externe, un peu échancré et concave dans le quart supérieur, devient ensuite régulièrement convexe. A l'intérieur se voient douze replis parallèles très accentués. L'antitragus, très élevé, est séparé du bord externe par une encoche profonde et anguleuse; sa surface bosselée présente en haut une partie bombée et convexe, plus bas une dépression circulaire profonde formant cuvette; son bord supérieur, arrondi et convexe, se replie en dehors en s'enroulant en ourlet; son bord antérieur, profondément concave, vient se terminer comme un soc au-dessous et au niveau de l'œil. La membrane alaire, très développée, d'un brun sombre, prend ses attaches aux malléoles. La queue est relativement courte, et sa pointe extrême dépasse un peu la membrane interfémorale. Celle-ci est de grandeur médiocre, triangulaire en arrière et relativement moins développée que chez le *R. luctus*.

Le pelage est doux, bien fourni, mais assez court, d'un blond châtain uniforme, un peu plus clair sur la face inférieure du corps.

La première prémolaire supérieure est petite, peu élevée, assez distante de la canine, au contraire très rapprochée de la deuxième prémolaire,

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contre laquelle elle est accolée. La deuxième prémolaire inférieure est très réduite, enchâssée dans l'angle qui sépare les première et troisième prémolaires et pincée par conséquent un peu en dehors de la rangée dentaire.

Le seul spécimen recueilli par M. le Docteur Maclaud est une femelle adulte, prise pendant la période d'allaitement, comme l'indiquent ses mamelles extrêmement turgides. Sur la région pubienne se voient les deux appendices tétiniformes ordinaires.

Ses principales dimensions sont les suivantes:

Longueur de la tête et du corps: 96^{mm}0
 - de la queue: 41 0
 - de la tête: 33 0
 Dimensions de la feuille nasale: 26 X 16 0
 Longueur totale de la selle: 14 0
 Largeur maximum de sa partie basale: 9 5
 Largeur de la partie supérieure ou languette: 5 0
 Longueur de l'oreille, de la base de son bord interne au sommet: 44 0
 Longueur de l'antitragus, de la pointe de son bord externe au sommet: 17 0
 Longueur de l'avant-bras: 68 0
 - du métacarpien du 3^e doigt: 47 0
 - de la 1^{re} phalange du 3^e doigt: 23 0
 - de la 2^e phalange du 3^e doigt: 36 0
 - du métacarpien du 5^e doigt: 50 0
 - de la 1^{re} phalange du 5^e doigt: 17 0
 - de la 2^e phalange du 5^e doigt: 22 0
 - du tibia: 31 0

Cette espèce, que je dénommerai *Rhinolophus Maclaudi* en l'honneur de M. le Docteur Maclaud, a été prise sur l'île même de Conakry, mais elle doit se rencontrer également un peu plus au large, sur les îles de Los, et d'autre part, sur les côtes et dans l'hinterland de nos possessions de la Guinée française; elle vient grossir le nombre, assez restreint du reste, des espèces du même genre, dans cette partie de l'Afrique où l'on n'a encore signalé que le *R. alcyone* (Tem.), le *R. fumigatus* (Rüpp.) et le *R. clivus* (Rüpp.).

Rhinolophus maendeleo Kock, Csorba and Howell, 2000

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(Figs. 1, 4, 7)

1994 *Rhinolophus* sp. nov., BURGESS & MUIR, Frontier-Tanzania Coastal Forests Workshop. Coastal forests E. Afr.: Biodiv. & conserv.: 30; Mkulumuzi (Amboni).

1995 *Rhinolophus* n. sp., CLARK & STUBBLEFIELD, Frontier-Tanzania Techn. Rep., 16: 13; Amboni Caves (KMH 7673) and E-Usambara Mts.

1998 *Rhinolophus* sp. 1 (cf. *swinyi*), COCKLE, KOCK, STUBBLEFIELD, HOWELL & BURGESS, Mammalia, 62 (1): 59, tab. 2; Mkulumuzi Forest.

Holotype male ad. (alc., skull, os penis), SMF 79643: Amboni Cave Forest, 05°05'S - 39°02'E, 0 - 80 m, Mkulumuzi River Gorge, 2.5 km W of Tanga, Tanga Distr., NE-Tanzania, 25.II.1992, L. STUBBLEFIELD leg. (KMH 7673); mistnetted 19:10 - 19:30h in opening in forest on path in front of stream.

Paratype female ad. (alc., skull): Mazumbai Forest Reserve, 04°25'S - 38°15'E, east ridge of West Usambara Mts., Lushoto Distr., NE-Tanzania, 29.VIII.1985, SMF 66960, H. GROSSMANN leg. (KMH 3216); mistnetted in garden.

Etymology: From Swahili "maendeleo" for progress; a noun in apposition. Named in allusion to the increasing knowledge of the Tanzanian bat fauna.

Diagnosis: Medium sized bat, forearm length 48 - 49. Horseshoe and sella of the noseleaf wide, the lancet nearly straight-sided (Fig. 1). Lower lip with three well-defined mental grooves. Skull slender, mastoid width equals or slightly exceeding zygomatic width; rostral inflations of nasal sinus high and wide; palatal bridge more than one-third of upper toothrow length (Fig. 4). These features are combined with the absence of a bony bar over the foramen infraorbitale. Distal part of baculum (os penis) strongly flattened dorsoventrally; dorsal part of the basal cone projecting proximally and deeply incised (Fig. 7).

Measurements (of holotype followed by paratype; in mm, if not stated otherwise). - External: Head and body 46, 49; tail from last vertebra to anus 23, 25.8; hindfoot s.u. 8.0, 8.2; ear from outer base 24.5, 24; horseshoe width 8.4, 8.2; tibia 19.4, 18.9; forearm including carpalia 48.2, 49.1; forearm without carpalia 47.2, 48.3; metacarpal III (between proximal and distal end of bone, excluding wrist) 32.1, 34.6; 1st phalanx digit III 14.6, 14.7; 2nd phalanx digit III 24.3, 24.2; metacarpal IV 36.1, 38.1; 1st phalanx digit IV 8.3, 8.15; 2nd phalanx digit IV 14.4, 14.2; metacarpal V 35.1, 36.2; 1st phalanx digit V 10.3, 10.9; 2nd phalanx digit V 12.15, 12.9; weight 6 g, -.

Skull: Greatest length of skull 20.75, 20.64; occipito-canine (crown) length of skull 20.07, 20.03; occipito-canine (alveolus) length of skull 19.57, 19.60; condylobasal length of skull 17.74, 17.72; condyle-canine (crown) length of skull 17.72, 17.75; condyle-canine (alveolus) length of skull 17.57, 17.59; mastoid width 9.39, 9.42; braincase width 8.5, 8.8; zygomatic width 9.37, 9.35; length of palatal bridge between anterior and posterior emarginations 2.70, 2.78; width across C1-/C1/ crowns 4.74, 4.79; width across M3-/M3/ (crowns) 6.58, 6.55; C-M3/ crown length 7.25, 7.20; C-M3/ alveolar length 6.83, 6.82; C-P4/ crown length 3.30, 3.28; greatest width of anterior rostral inflations 5.01, 4.96; least interorbital width 2.53, 2.49; mandible length from outer point of proc. articularis to III alveolus -, 13.07; mandible length from outer point of proc. condylaris to III alveolus -, 13.14; C-M3/ crown length 7.62, 7.43; C-P4/ crown length 2.83, 2.70.

Description: Pelage dorsally brownish, ventrally beige turning to whitish on lower abdomen and loins; a darker brown collar around lower neck and upper chest. Ears large, reaching beyond the tip of nose when laid forward, tips bluntly rounded. The noseleaf covering almost all of the muzzle; horseshoe wide, no supplementary noseleaf. Sella naked, wide at its base, rising from a rather well-developed cup and constricted in the middle;

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connecting process hairy, forming a continuous arch and its basal part almost parallel to the sella; the tip of lancet nearly straight-sided (Fig. 1). Three mental grooves in lower lip. Fourth and fifth metacarpals subequal in length, the third metacarpal shorter. First phalanx of digit IV more than one half length of 2nd phalanx. The last caudal vertebra is free from uropatagium.

Skull (Fig. 4): Slender, with long rostral part; anterior median rostral inflations enlarged, bulbous, with the posterior rostral inflations of nasal sinus reduced which results in a markedly concave rostral profile with moderately deep rostral depression flanked by strong postorbital ridges. Weak sagittal and lambdoid crest in the male holotype and almost absent in the female paratype. In both specimens the foramen infraorbitale is open by natural absence of a bony bar. Palatal bridge more than one-third of the C-M3/ length. Interpterygoid groove deep, with lateral ridges posteriorly.

Teeth: C1/ with small posterior cingulum cusp in unworn stage, considerably exceeding PM4/ in height; upper PM2/ medium sized, in toothrow, separating C1/ from PM4/; I/1 two-third the bulk of I/2; the small PM3 partly extruded from toothrow, posteriorly covered by cingulum of PM4/; PM2 and PM4 not in contact.

Baculum: 2.66 long, its distal half dorsoventrally compressed; the dorsal part of the basal cone enlarged and deeply emarginated forming two long wings; the ventral incision less deep (Fig. 7).

Comparison

Rh. maendeleo n. sp. externally can be distinguished from the closely related *Rh. adami* by the shape of the lancet (longer in *Rh. adami* with convex sides; Fig. 2). Cranially, *Rh. adami* is larger in the following measure

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ments (n = 6): palatal bridge length 2.99 - 3.23; greatest width of anterior rostral inflation 5.08 - 5.46; interorbital width 2.64 - 2.81; mastoid width 9.47 - 9.88; width across M-M3/ 6.64 - 6.87. The bony bar across the foramen infraorbitale is present. The interpterygoid groove is deep in both species but its walls run nearly parallel in *maendeleo* n. sp. while they are constricted close to the proximal end in the case of *Rh. adami* (Fig. 5). The baculum of *Rh. adami* (Fig. 8 - 9) has a much shorter basal cone with shallow dorsal and ventral incisions.

In the area where *Rh. maendeleo* n. sp. has been found, the only other taxon with similar external appearance is *Rh. s. simulator* ANDERSEN 1904. This latter, however, is smaller in forearm length (42.0 - 47.2) and has a more pronounced, wider based connecting process and hastate lancet (Fig. 3). Cranially the two taxa are distinguishable by the less inflated narial swellings (Fig. 6), presence of bony bar across the foramen infraorbitale, and smaller measurements of *Rh. s. simulator* (n = 42: occipito-canine length of skull 17.27 - 19.25; C-M3/ crown length 6.31 - 7.08; mastoid width 8.62 - 9.33; articular length of mandible 11.60 - 12.54; C- M3 crown length 6.66 - 7.35).

Besides in both specimens of *Rh. maendeleo* n. sp., the bony bar across the foramen infraorbitale is not developed on one side of one specimen of *Rh. cognatus* ANDERSEN 1906 (Bay of Bengal, S. Andaman, Port Blair, BMNH 6.12.1.12) and absent from both sides of one

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specimen of *Rh. fumigatus* RÜPPELL 1842 (Murka, Tavo West N.P., Kenya, SMF 41816).

Another similar horseshoe bat is *Rh. simulator alticolus* SANBORN 1936, a West African forest endemic. The proper taxonomic status of this taxon (originally assigned as subspecies to *Rh. alcyone*) is controversial (HILL & MORRIS 1971, KOOPMAN 1975, 1993), sometimes being regarded as separate species (EISENTRAUT 1956, ROSEVEAR 1965, HAYMAN & HILL 1971) ALTHOUGH ONLY ON THE BASE OF A COMPARISON WITH *RH. ALCYONE* AND *RH. LANDERI*. *RH. S. ALTICOLUS* IS SHOWING SOME AFFINITIES TO *RH. MAENDELEO* N. SP. AND *RH. ADAMI* BY ITS WIDE HORSESHOE, LARGER SELLA AND THE SIDES OF THE LANCET MORE OR LESS STRAIGHT, BUT CRANIALY AND BY MEASUREMENTS IT IS VERY SIMILAR TO *RH. S. SIMULATOR*.

SYSTEMATICS

AMONG THE AFROTROPICAL *RHINOLOPHUS* SP., THE FOLLOWING SET OF CHARACTERS APPLY TO *RH. MACLAUDI* POUSARGUES 1897 (CURRENTLY INCLUDING *RUWENZORII* HILL 1942 AND *HILLI* AELLEN 1973), *RH. ADAMI* AELLEN & BROSSET 1968 AND *RH. MAENDELEO* N. SP. ONLY: VERY LARGE EARS AND SELLA, TIP OF LANCET STRAIGHT-SIDED OR ROUNDED (NOT HASTATE), THREE MENTAL GROOVES, NARROW SKULL (MASTOID WIDTH SUB EQUAL TO OR WIDER THAN ZYGOMATIC WIDTH), BULBOUS NARIAL INFLATIONS, PM2/ IN TOOTHROW AND LONG PALATE. MANY OF THESE CHARACTERS ARE REGARDED AS BASAL FEATURES (ANDERSEN 1905) AND IN THE INDOMALAYAN REGION ARE TYPICAL FOR THE SPECIES OF THE PHILIPPINENSIS-GROUP OF BOGDANOWICZ (1992). OF THIS GROUP, *RH. MACLAUDI* IS CHARACTERIZED BY A VERY ELABORATE NOSELEAF AND SPECIALLY FORMED NASAL INFLATIONS, AND, ACCORDING TO BOGDANOWICZ (1992), CLUSTERS FAR FROM THE PHILIPPINENSIS-GROUP; ITS PHYLOGENETIC POSITION IS UNCERTAIN. *RH. ADAMI* (NOT INVESTIGATED BY BOGDANOWICZ) WAS PLACED ORIGINALLY NEAR THE AFRICAN MEMBERS OF THE *FERRUMEQUINUM*-GROUP (AELLEN & BROSSET 1968), but based on the above mentioned features, along with *Rh. maendeleo* n. sp. represents a different clade within African rhinolophids which is here formally established as the *adami*-group.

Habitat, status and faunal assignment

The Amboni Cave Forest is a coastal lowland forest of 350 ha at an altitude from sea level to 80 m, consisting of degraded forest and evergreen thicket in a limestone gorge, under the Protection of Monuments (Amboni Caves) Order, 1937, originally protected during the German administration (CLARKE & STUBBLEFIELD 1995). - The Mazumbai Forest is a primeval lower montane rain forest of 450 ha, at an altitude of 1400 - 1900 m (REDHEAD 1981, LOVETT & NORTON 1989), protected by its status as University Natural Forest Reserve of the Sokoine University of Agriculture, Morogoro.

The known range of the new species is wholly within protected areas. The habitat characterises *Rh. maendeleo* n. sp. as a faunal element of the Zanzibar-Inhambane phytogeographic zone (see SHEIL 1992), respectively the Swahili regional centre of endemism of BURGESS et al. (1998), whose forests extend inland to the foothills of some of the Eastern Arc mountains. Although Mazumbai Forest is even further inland and at a higher altitude of the West Usambara Mts., this recently defined Swahili regional centre is still not definitely delimited.

Local congeners of *Rhinolophus* sp.: Until present, no *Rhinolophus* sp. is reported for Mazumbai Forest (REDHEAD 1981), and *Rh. maendeleo* n. sp. represents the first record of the genus.

The bat fauna of the Amboni Caves (= Mkulumuzi Caves, = Siga Caves) is comparatively well known having been investigated under several biological aspects (SJÖSTEDT 1910, ALLUAUD & JEANNEL 1914, AIELLO ET AL. 1960, COCKLE et al. 1998). However, there are rather few species of *Rhinolophus*. The cave and surrounding forest are inhabited by *Rh. hildebrandtii* PETERS 1878 (KULZER 1959, COCKLE et al. 1998, SMF 79633), *Rh. eloquens* (SMF 81201: Amboni Caves), while *Rh. d. swinnyi* GOUGH 1908 listed by COCKLE et al. (1998) is the present new species.

Biogeography

The East African forests of the coastal belt from Somalia to Mozambique have a unique diversity of flora

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(SHEIL 1992) and mammal fauna (KINGDON & HOWELL 1993) with a high number of endemics. These forests are relics of a once continuous forest belt from West to East Africa, broken up several times during drier climatic periods (HAMILTON 1982).

Among bats, the members of the newly established *adami*-group provide an interesting addition to this list. Endemic Chiroptera of the Zanzibar-Inhambane phytogeographic zone are: *Pteropus voeltzkowi* MATSCHIE 1909, *Myonycteris relicta* BERGMANS 1980, *Taphozous hildegardeae* THOMAS 1909, *Rh. deckenii* PETERS 1867, *Glauconycteris kenyacola* PETERSON 1982, *Kerivoula africana* DOBSON 1878, and *Tadarida (Mops) brachyptera* (PETERS 1852). *Rh. maendeleo* n. sp. emphasises the biogeographic identity of a separate SE-African faunal region (BURGESS et al. 1993).

The relationship of the bats of this Zanzibar-Inhambane zone with species of the Guinea-Congolian lowland forest on the supraspecific level is documented by *Rh. adami* of the Congo (B) lowlands as the closest relative of *Rh. maendeleo* n. sp., by *Rh. silvestris* AELLEN 1959 as nearest relative of *Rh. deckenii*, and by *Tadarida (Mops) leonis* (THOMAS 1908), the closest relative of *T. (M.) brachyptera*. At species level, W-African lowland forest taxa in common with the E-African coastal lowland forests are: *Hipposideros gigas* (WAGNER 1845), *H. cyclops* (TEMMINCK 1853), and possibly *H. camerunensis* EISENTRAUT 1956, which reaches east to W-Kenya only. At subspecies level *Miniopterus m. minor* PETERS 1866 in the East and *M. minor newtonii* BOCAGE 1889, and *M. m. occidentalis* JUSTE & IBAÑEZ 1992, in the West are mutual representatives.

Rhinolophus Martini Fraser, 1843

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Mr. Fraser exhibited and described a new species of Bat, belonging to the genus *Rhinolophus*, and four new species of Birds from Western Africa.

Rhinolophus Martini. *Rhin. auribus magnis apud frontem inter se spatio angusto sejunctis; rostro fossá oblongá supernè, anticè quatuor appendiculis carneis, vix elevatis (duobus utrinque), tectd:*

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prosthema nasali longitudinalitèr diviso; fossá frontali posticè culmine semicirculari dense vellere induto, collimetatá; caudá longá et cartilagine bifurcá terminatá; patagio ad pedes basim solummodó ducto: colore cinereo-fusco, subtùs canescente.

unc. lin.

Long. tot.: 3 5

Volatús amplitudo: 10 0

---- latitudo maxima: 2 4

Antebrachium: 1 6

Auris: 0 9

Cauda: 1 7

Hab. Fernando Po.

This species of *Rhinolophus* is remarkable for having the complicated fleshy appendages of the muzzle divided in the longitudinal direction. Each half of this apparatus is composed of two leaflets, the margins of which are free, though but little elevated; the foremost of these is shaped somewhat like the human ear, and terminates in front in a small prominent lobe, which is situated over the opening of the nostril; the second or hindermost leaflet on each side approaches to a circular form. Upon separating these four leaflets a large pit is observable on the upper surface of the muzzle, and the hinder margin of this pit terminates in a nearly semicircular and slightly elevated fleshy ridge, which is densely clothed with fur. The ears are large, rounded at the extremity, but inclining to a pointed form, and separated from each other on the top of the head by a space of about two and a half lines in width; on the inner side, and towards the base, is a narrow oblique ridge: the tragus is about two and a half lines in length, narrow, rounded at the extremity, and somewhat dilated near the base on the outer margin. The wing and interfemoral membranes join the foot at the base; the latter extends to the extremity of the tail, which terminates in a bifurcated cartilage; numerous minute papillæ are observable on the margin of the interfemoral membrane. The general colour of the animal, in spirit, is gray-brown, but with an ashy tint on the under parts of the body, and is darker than that of the *Rhinolophus Hipposideros* of authors.

Rhinolophus micaceus de Winton, 1897

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Rhinolophus micaceus, sp. n.

Colour above and below dark soft rich purplish brown, flecked with silver-grey. The fur is so fine that it is difficult to see if the hairs are truly annulated; but as the silvery speckling comes and goes as the animal is turned in the light, it appears to arise wholly from the glint of the light on the brown fur. It is not possible to give particulars of noseleaf, the specimen being dried. Length of fur on the back 11.5 millim.

Measurements taken in flesh by collector: -

Head and body 90 millim.; tail 23; hind foot 17; ear 26; forearm 69.

Skull: greatest length 28 millim.; greatest breadth 15.3; breadth of brain-case 11.5; intertemporal constriction 3.9; basal length 22.9; between tips of canines 6.5; length of entire tooth-row 10.5.

Type (F): Como River, 75 miles from Gaboon, 6th July, 1897.

Collected by Mr. G. L. Bates.

Rhinolophus ruwenzorii J. Eric Hill, 1942

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TYPE. - Amer. Mus. Nat. Hist. 82394, F adult, alcoholic with skull removed. Brought in by natives with a *Rousettus lanosus* from a cave, December 24, 1926, south side of Butahu Valley, western slope of Mount Ruwenzori, altitude 7,500 feet.

DIAGNOSIS. - Like *Rhinolophus macclaudi* Pousargues (1897, Bull. Mus. d'Hist. Nat., Paris, III, pp. 358 - 361, 2 figs.) from French Guinea, but smaller, with lateral margins of nostrils less developed and with less concavity in the anterior margin of the antitragus. Skull with relatively broader braincase, shorter palatal bridge, more abrupt nasal swellings and narrower zygomatic spread.

Rhinolophus ruwenzorii is larger in body than *R. aethiops* or *R. hildebrandti*, but the measurement of the forearm is approximately as in these species.

The noseleaf is quite unlike that of any African species except *R. macclaudi*, and it differs from the noseleaf of that form only in minor details: lateral margins of the nares form a low ridge, rather than "coming to double, as a chalice, the sort of corolla formed by the basal part of the sella" (Pousargues, describing *R. macclaudi*); horseshoe more widely divided anteriorly and without the shallow emarginations near the medial side of the anterior margin.

The ears are much like those of *R. macclaudi* but differ in details: notch separating the antitragus less acute; anterior concavity of the antitragus less marked; terminal plowshare-like process ("soc," described and figured by Pousargues) obsolete in the new form.

Pelage abundant and soft, about 13 mm. long on the shoulders. Coloration (possibly faded after 15 years in alcohol): upperparts darker than Bister; underparts near Benzo Brown; wings and ears near Fuscous-Black. (Names of colors in capitals from Ridgway, 1912, "Color Standards and Color Nomenclature.") *R. macclaudi* was described as light chestnut ("blond chatain"); it appeared darker than this in 1937, more like our specimen.

SKULL. - Similar to that of *R. macclaudi* (examined and photographed in 1937, while studying material in European museums under a grant from the Carnegie Corporation of New York), but smaller (13 to 20 per cent in most measure-

Rhinolophus ruwenzorii, *Rhinolophus macclaudi*

n. sp., Pous.

Type, Type,

AMNH 82394, Paris 1897-981

F ad., F ad.

Butahu Valley, Conakry Isl. Fr. Guinea

EXTERNAL MEASUREMENTS:

Head and Body: 67.0, 96.0

Tail: 31.0, 41.0

Hind foot (s.u.): 12.0, -

Tibia: 23.5, 31.0

Ear (from notch): 32.5, 44.0

Forearm: 57.0, 68.0

Digit II, Metacarpal: 43.4, 50.3

Digit III, Metacarpal: 40.0, 47.0

1st phalanx: 18.5, 23.0

2nd phalanx: 32.0, 36.0

Digit IV, Metacarpal: 45.0, 50.8

Digit V, Metacarpal: 46.8, 50.0

SKULL:

Greatest length: 25.9, 30.9

Basal length: 20.7, 25.3

Palatal bridge: 3.6, 4.5

Zygomatic breadth: 11.2, 14.0

Lacrimar breadth: 7.1, -

Temporal constriction: 2.6, 3.1

Mastoid breadth: 12.0, 13.8

Front of orbit to prosthion: 9.3, -

Height of occiput: 5.9, 7.2

Outside breadth across tooth row M³-M³: 8.2, 10.0

Maxillary alveoli: 8.2, 10.2

Crowns P⁴-M³: 6.3, 8.1

ments) with relatively broader braincase (mastoid-basal index 58 rather than 54.5) and less spreading zygomatic arches (their spread less than, rather than greater than, mastoid breadth). Nasal swelling more pronounced and abrupt; intertemporal "neck" shorter; maxillary root of

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zygoma weaker than in *R. macclaudi*; palatal bridge shorter. A well-developed pit between the posterior roots of the pterygoid plates in both species.

DENTITION. - Cheek teeth smaller; upper incisor bifid; P^2 in toothrow, but minute, about as high as cingulum of C^1 ; M^1 with poorly developed hypocone, this is little more than a cingulum in M^2 (the molars are similar in *R. macclaudi*); lower incisors tricuspidate, the medial pair smaller, overlapping lateral ones; P_3 minute, "squeezed" laterally between P_2 and P_4 ; the former large, squarish in outline from above, about threefourths as high as P_4 .

The palatal ridges are most like Seabra's schema for *R. hipposideros* (Jorn. Sci. Math. Phys. Nat., Lisbon, [21 V, pp. 248-252], but the second ridge runs between the minute first premolars, and the last four ridges are incomplete medially.

Rhinolophus silvestris Aellen, 1959

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Type. - M adulte, en alcool. Grotte de N'Dumbu, Lastoursville, Gabon, 4.8.1957; coll. P. Strinati et V. Aellen. Muséum d'Histoire naturelle de Genève, n° 965.40.

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Cette nouvelle espèce a été signalée par SANBORN (1953) sous le nom de *Rhinolophus f. fumigatus* Rüppell. L'auteur américain a examiné un spécimen M provenant de la grotte de Bongolo, près de Lébamba, Gabon; il l'a comparé à neuf *R. fumigatus* typiques du British Muséum et l'a trouvé identique, à part une rangée dentaire plus longue et une largeur maxillaire plus grande. Il est curieux que le biotope de forêt équatoriale, opposé à celui de savane ou de steppe de *R. fumigatus*, n'ait pas attiré l'attention de SANBORN. Les mensurations publiées par ce dernier montrent à l'évidence que son spécimen appartient à ma nouvelle espèce. En fait, les différences existant entre le Rhinolophe du Gabon et *R. fumigatus* ne permettent même pas d'attribuer le premier, en qualité de sous-espèce, au second.

Il est probable que *R. silvestris* a été confondu parfois sous le nom de *Rhinolophus ferrumequinum* (Schreber). Ainsi, un nouvel examen du spécimen "u" provenant du Gabon, que DOBSON (1878) classe sous ce nom, montrera très probablement qu'il s'agit de la nouvelle espèce.

Diagnose. - *Rhinolophus* de taille plutôt grande (avant-bras 50 mm, long, du crâne 23,4 mm). Feuille nasale à connectif à sommet arrondi et très bas, ne dépassant pas le processus vertical (dossier) de la selle; fer à cheval large (10 mm) recouvrant tout le museau. Troisième métacarpe réduit, inférieur à

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90% du cinquième métacarpe. P^2 très réduit et externe, de sorte que C et P^4 sont presque en contact. P_3 tout à fait rudimentaire, non fonctionnel; P_2 et P_4 se touchent.

Description. - Les oreilles atteignent le bout du museau, lorsqu'elles sont rabattues en avant; elles sont peu acuminées; l'antitragus est séparé du pavillon par une encoche à angle droit ou un peu obtus; le bord externe est sinueux, un peu concave avant le sommet; le bord interne est convexe dans la partie basale, droit au sommet. La feuille nasale est large et recouvre tout le museau; le fer à cheval est fortement échancré au milieu et un peu festonné sur les bords; l'échancrure n'est pas accompagnée, de chaque côté, d'une petite dent, comme PETERS (1867) l'indique pour *R. deckeni*. La selle est large (3,2 mm) et son processus vertical (dossier) étranglé au milieu, il mesure 1,9 mm de large au sommet. La pointe du connectif est très basse et largement arrondie. La lancette est en forme de triangle équilatéral; ses côtés sont droits, à peine concaves vers le sommet. Il y a un seul sillon au milieu de la lèvre inférieure. Le patagium s'insère à la cheville. Le troisième métacarpe est réduit, c'est-à-dire qu'il mesure moins des 90 % du cinquième métacarpe, exactement dans les mêmes proportions que chez *R. deckeni*. Les deuxième phalanges du troisième et du quatrième doigt sont relativement courtes: elles mesurent respectivement moins du double des premières phalanges. La queue est assez longue et mesure plus de la moitié de la longueur de l'avant-bras.

La coloration est tout à fait semblable à celle de *R. ferrumequinum* (Schreber).

Le crâne présente un renflement nasal bien prononcé. Les palatins s'arrêtent, en avant, au niveau de la moitié antérieure de M^1 . La première prémolaire supérieure (P^2) est très réduite et externe; la canine et la seconde prémolaire (P^4) sont presque en contact. La réduction de P^2 est toutefois moins accentuée que chez *R. fumigatus* et semble du même ordre que chez *R. deckeni*. A la mandibule, la première prémolaire (P_2) est très basse, bien que de section appréciable; elle n'atteint pas la moitié de la hauteur de la troisième prémolaire (P_4). La seconde

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prémolaire (P_3) est tout à fait rudimentaire, non fonctionnelle et externe; P_2 et P_4 sont en contact.

Remarques. - Il n'est pas étonnant que cette espèce ait été confondue par SANBORN avec *R. fumigatus*. Elle lui ressemble beaucoup, du moins superficiellement: la feuille nasale et les caractères dentaires sont très semblables. Cependant, on peut relever les différences essentielles suivantes:

R. silvestris, *R. fumigatus*

Longueur de l'avant-bras: environ 50 mm, 51 à 56 mm
 Longueur du 3e doigt: 76,5 à 80 mm, 82,2 à 96 mm
 Longueur du 3e métacarpe par rapport au 5e: inférieur à 90 %, supérieur à 90 %
 Longueur de la queue: super, à la moitié de l'avant-bras, infér. À la moitié de l'avant-bras
 3e doigt, 1^{re} phalange: super, à la moitié de la 2e phal., infér. À la moitié de la 2e phal.
 4e doigt, 1^{re} phalange: super, à la moitié de la 2e phal., infér. À la moitié de la 2e phal.

D'autre part, les proportions du crâne ne sont pas les mêmes. Le crâne de *R. silvestris*, qui est plus court que celui de *R. fumigatus*, présente une série dentaire C-M³ plus longue et une largeur maxillaire M³-M³ plus large. P² est moins rudimentaire, semble-t-il, chez *silvestris* que chez *fumigatus*.

La répartition géographique actuellement connue de *Rhinolophus f. fumigatus* Rüppell comprend l'Erythrée, l'Éthiopie, le Soudan¹, les Somalies anglaise et italienne, le Kenya, le Tanganyika², la Rhodésie du Nord. D'iverses formes ont été rapportées à *R. fumigatus*, en tant que sous-espèces ou synonymes:

aethiops Peters, 1869. Loc. Typ.: SW Africain.
 = *R. f. aethiops*, sec. ELLERMAN, MORRISON-SCOTT et HAYMAN (1953).

macrocephalus Heuglin, 1877. Loc. Typ.: Éthiopie.
 = *R. f. fumigatus*, sec. ANDERSEN (Ann. Mag. Nat. Hist. (7) 14: 453, 1904).

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antinorii Dobson, 1885. Loc. Typ.: Éthiopie.
 = *R. f. fumigatus*, sec. ANDERSEN (Ann. Mag. Nat. Hist. (7) 14: 453, 1904).

exsul Andersen, 1905. Loc. Typ.: Kenya.
 = *R. f. exsul*, sec. ANDERSEN (Ann. Mag. Nat. Hist. (7) 15: 74, 1905).

eloquens Andersen, 1905. Loc. Typ.: Ouganda.
 = *R. aethiops eloquens*, sec. SANBORN (Zool. Ser. Field Mus. Chicago 24: 42, 1939).
 = *R. f. eloquens*, sec. ELLERMAN, MORRISON-SCOTT et HAYMAN (1953).

Il y aura lieu, certainement, d'assimiler encore les formes suivantes à *Rhinolophus fumigatus*:

foxi Thomas, 1913. Loc. Typ.: N Nigeria.
abae J. A. Allen, 1917. Loc. Typ.: Uélé, Congo belge.
perauritus de Beaux, 1922. Loc. Typ.: Somalie.
diversus Sanborn, 1939. Loc. Typ.: Sénégal.

Rhinolophus silvestris a, comme plus proche parent, non pas *R. fumigatus*, mais *R. deckeni* Peters (1867). Ce dernier n'est malheureusement pas bien connu. La localité typique est la côte NE du Tanganyika, face à l'île de Zanzibar; la description de PETERS est assez sommaire et seules les mensurations externes sont données. DOBSON (1878) rapporte *R. deckeni* à *R. ferrumequinum* (Schreber). THOMAS, en 1910, cite *R. deckeni* à Rombo (Tanganyika), mais quelques années plus tard, il précise que la détermination est erronée et qu'il s'agit de *R. augur zambesiensis*. Par contre, dans le même travail, THOMAS (1913) dit que le British Muséum possède un bon exemplaire de *R. deckeni*, provenant du Kenya (près de Mombasa); il en donne même une mesure crânienne; il le compare à sa nouvelle espèce, *R. foxi*, en précisant que ce dernier est "evidently a western representative of *R. deckeni*". On a vu, plus haut, que je rattache *R. foxi* à *fumigatus*. Ceci montre bien l'étroite ressemblance de ces formes. Enfin, *R. deckeni* a été récemment signalé à Kisolo, en Ouganda³. En résumé, on ne connaît de

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la répartition géographique de *R. deckeni* que trois localités de l'Afrique orientale: côte du Tanganyika opposée à Zanzibar, près de Mombasa (Kenya) et Kisolo (Ouganda).

D'après les mesures publiées, *R. deckeni* est une espèce nettement plus grande que *R. silvestris* (cf. table des mensura-

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tions), mais en présentant tous les caractères essentiels, aussi bien externes que dentaires, pour autant que permettent d'en juger la description originale et les précisions de THOMAS (1913). Nous pouvons reprendre l'expression de ce dernier, à propos de *R. foxi*, en la modifiant ainsi: *R. silvestris* est très probablement le représentant forestier de *R. deckeni*, auquel il sera peut-être possible de le rattacher comme sous-espèce, lorsque l'on connaîtra mieux ce dernier.

Rhinolophus silvestris semble localisé au Gabon. Les régions forestières, assez bien prospectées du S du Cameroun et du bassin du Congo, n'ont fourni aucun *Rhinolophe* référent à la nouvelle espèce.

Dans le tableau de mesures ci-dessous, j'ai indiqué celles données par SANBORN (1953) pour son "*R. f. fumigatus*" du Gabon; on peut constater qu'elles sont presque identiques à celles du type de *silvestris*. A titre de comparaison, j'ai mis en parallèle les mesures de *R. deckeni*, selon PETERS (1867) et THOMAS (1913) et celles de *R. f. fumigatus* trouvées dans la littérature.

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Rhinolophus silvestris: M type, M SANBORN; *Rhinolophus deckeni* F type; *Rhinolophus f. fumigatus* (min - max)

Avant-bras: 49, 50.3; 55; (51 - 56)
 3e doigt, métacarpe: 33, 35.2; 37; (35.6 - 42)
 " 1^{re} phalange: 16, 16.7; 19; (15.8 - 18)
 " 2^e phalange: 27.5, 28; 34; (29.5 - 36)
 4e doigt, métacarpe: 37, 39.2; 42; (37 - 44)
 " 1^{re} phalange: 9.5, 9.8; 11; (9 - 11)
 " 2^e phalange: 16.5, 16.7; 19.8; (18 - 24)
 5e doigt, métacarpe: 38, 40.2; 43; (38.6 - 45)
 " 1^{re} phalange: 11, 11.6; 13; (12.1 - 14)
 " 2^e phalange: 14.5, 15.4; 18.8; (13.5 - 17)
 Tibia: 19.5, 21.4; 25; (20.3 - 23)
 Pied (avec griffes): 11, -; 15; (10.5 - 11.5)
 Queue (de l'anus): 28, 32; 29; (21 - 27)
 Oreille (longueur): 22, 21.9; 23; (20.3 - 26)
 Feuille nasale (long.): 15, -; 17.2; (17 - 18)
 Fer à cheval (larg.): 10, -; 11; (9.5 - 10.2)
 Crâne:
 Long. tôt. (prémaxill.): 23.4, -; -; (-)
 " " (à la canine): 22.8, 22.4; -; (24)
 " condylobasale (C): 20, 19.6; -; (-)
 " naso-occipitale: 19, -; -; (-)
 " palatale: 2.8, 2.8; -; (-)
 Larg. renflement nasal: 6, 6; -; (6)
 " interorbitaire: 2.4, 2.5; -; (3)
 " zygomatique: 11.6, 11.8; -; (12)
 " mastoïde: 10.7, 10.6; -; (11)
 " C-C (externe): 6.8, 6.4; -; (-)
 " M³-M³ (externe): 8.5, 8.7; -; (8 - 8.4)
 Long, rangée dentaire C-M³: 8.7, 8.6; -; (7.7 - 8.3)
 " " " P⁴-M²: 5.1, -; 5.8; (-)
 " mandibule: 15.6, -; -; (16.3)
 " rangée dentaire I-M₃: 9.9, -; -; (-)
 " " " C-M₃: 9.2, -; -; (9.2)

footnotes:

¹ Voir THEODOR, O., *Parasitology* 47: 478, 1957.

² Voir SWYNNERTON, G. H., *Annual Report of the Game Department 1953, Dar es Salam*: 55, 1954.

³ Voir JOBLING, B., *Rev. Zool. Bot. afr.* 50: 101, 1954.

Rhinolophus simulator K. Andersen, 1904

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Short diagnosis. - Small upper premolar separating the canine and posterior premolar. In outer aspect somewhat like a peculiarly small and long-tailed *Rh. capensis*. Forearm 43.5-45 mm.

Nose-leaves. - Horseshoe as in *Rh. capensis*; underlying leaf distinct, under the margin of the horseshoe. Sella longer (higher) and slightly broader than in *Rh. capensis*; constriction below the middle very shallow; lateral margins subparallel in their upper half; summit broadly rounded off; width at summit equal to half the vertical height of sella.

* I am very glad to see these facts in perfect accordance with the zoogeographical subdivision of this part of S. Africa as proposed, five years ago, by Professor Matschie in his admirable book 'Die Megachiropteren des Berliner Museum für Naturkunde' (Berlin, 1899, p. 40).

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[table removed - eds.]

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Posterior connecting-process very short, rounded off above, scarcely projecting beyond the summit of sella (this kept in vertical position). Margins of lancet abruptly narrowed; tip of lancet behind constriction with slightly converging (nearly parallel) margins, about 2 mm. in length.

Ears. - Large; reaching far beyond the tip of the muzzle, when placed along the sides of the head. Tip of ears obtusely pointed, as in *Rh. capensis*. Outer margin strongly concave below tip. Notch on outer margin shallow, obtuse. Extreme width of ear somewhat greater than the distance from notch to tip.

Wings. - General form as in *Rh. capensis*, but all dimensions markedly smaller. 4th metacarpal slightly longer than 5th. In the folded wing the tip of the 1st joint of 4th finger reaches very nearly the posterior point of the elbow; tip of 1st joints of 3rd and 5th fingers slightly beyond. Plagiopatagium inserted at basal [corrected on paper in tarsal - eds.] joint.

Tail. - Considerably longer than in *Rh. capensis*, uropatagium therefore much broader. Hinder margin of uropatagium, between tips of calcars, square (concave in *Rh. capensis*) on account of the shorter tail). Tip of tail projecting about 1 mm.

Colour (1 skin). - Exactly as in *Rh. augur*. Length of hairs on middle of back about 8 mm. Young individuals (1 skin) considerably duller than adult ones; details exactly as in *Rh. augur*, juv.

Skull. - Very different from that of *Rh. capensis*: - Much smaller. Nasal swellings unusually prominent; nasal region deeply hollowed out behind swellings; for both these reasons the facial portion of the skull, from the swellings to crista sagittalis, more concave than in *Rh. capensis*. Palatal bridge much broader: median anterior point opposite front of m^1 , in *Rh. capensis* opposite middle of the same molar.

Dentition. - Widely different from that of *Rh. capensis*: - Distance between tips of upper canines 3.8 mm., in *Rh. capensis* 5 mm. Upper canine and p^4 distinctly separated from each other, also internally; distance between tips of these teeth 2.1 mm.; in *Rh. capensis*, although the skull is much larger, 1.9 mm. Upper p^2 placed in the tooth-row, with a minute cusp distinctly projecting above the cingulum of canine. Highest cusp of p^4 somewhat more than half the height of canine. Inner lobes of molars less developed than in *Rh. capensis*: m^2 as long as broad, in *Rh. capensis* decidedly broader than long. Right p_3 extremely minute, in the outer angle formed by the cingula of p_2 and p_4 ;

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cingula of these latter in immediate contact internally; left p_3 wanting, without any trace of the alveolus.

Type. - M ad. (in alcohol). Mazoe, Mashonaland, collected by J. ff. Darling, Esq. B.M. no. 2.2.7.10.

Distribution. - Besides the type specimen I have seen only two skins, from the same locality and the same collector.

Remarks. - Owing to our still very defective knowledge of the Ethiopian *Rhinolophi*, the true affinities of this interesting species are somewhat obscure. I have compared it with *Rh. capensis*, only because it, by quite superficial inspection, could be taken for a curiously small and long-tailed form of that species. The totally different skull and dentition prove that it, in fact, has nothing to do with *Rh. capensis*, but, most probably, is to be regarded as an aberrant member of the *Rh. Landeri* group. It cannot be confounded with any other species; *Rh. Landeri*, *lobatus*, and *Denti* may, apart from many other differences, at once be distinguished by their very narrow sella.

Measurements.

M ad, type; M ad, typical locality

Nose-leaves: mm. mm.

Front of horseshoe to posterior point of lancet: 13.7; -

Extreme width of horseshoe: 8.3; -

Width of sella at base: 2.4; -

" " summit: 2.2; -

Ears:

Base of inner margin to tip: 20; -

Notch on outer margin to tip: 13.2; -

Extreme width: 14.5; -

Wings:

Forearm: 43.5; 45

3rd finger, metacarpal: 30; 30.8

" 1st joint: 13.6; 14.5

" 2nd joint: 22.9; 22.8

4th finger, metacarpal: 33.2; 34

" 1st joint: 8.1; 8

" 2nd joint: 14.8; 13

5th finger, metacarpal: 32.2; 33.3

" 1st joint: 10.7; 10.5

" 2nd joint: 12.5; 12

Tail, from anus: 25.7; -

Tibia: 19.2; -

Skull:

Basal length: -; 15.1

Extreme width of brain-case: -; 9.1

Zygomatic width: -; 9

Maxillar width across outer margins of m^3 : -; 6.2

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Skull (con.):

Orbital constriction: -; 2.4

Palatal bridge: -; 2.7

Med. post. point of palate to front of premaxilla: -; 6

Med. post. point of palate to for. magnum: -; 9.1

Mandible: condyle to front of incisors: -; 12.3

Teeth:

Front of upper canine to back of m^3 : -; 7

Front of lower canine to back of m^3 : -; 7.5

Rhinolophus swinnyi Gough, 1908

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Two skins of an apparently new species of *Rhinolophus* were found in a small collection of skins prepared by Mr. H. H. Swinny, Ngqeleni, District W. Pondoland, which I propose to describe under the name.

Rhinolophus swinnyi, n. sp.

Size very small, not as large as the smallest yet known South African species of the genus.

Posterior upper praemolar separated from the canine, which is very large, anterior praemolar very small, in the tooth row, towards its outer side.

[Fig 1 & 2 - removed]

Horizontal portion of sella narrow, upper margin of the posterior connecting process rising above the sella, forming a marked projection, rounded terminally; sides of the vertical process of the sella parallel above, summit truncated with rounded off sides. Antitragal notch very shallow.

Horse shoe large, circular, its anterior margin notched in the centre; lancet moderate, its sides strongly concave, covered posteriorly with fur similar to that on the rest of the head, anteriorly somewhat less dense. Ears of medium size, the under edge convex in the distal half, then concave, convex again proximally, the concave portion very short. Tip sharply pointed, upper half of the outer margin concave,

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antitragal notch very shallow, lobe strongly convex. Hind limbs slender. Wings from the ankle. Interfemoral membrane finely fringed posteriorly.

Fur close and fine, about 7 mm. long on the back. General colour mouse grey, lighter below, individual hairs ash grey with brown tips. Membranes dark brown or black, interfemoral membrane with a fringe of grayish hairs.

Dimensions of Type

Forearm, 40 mm.; "head and body, 42 mm.;" "tail, 18 mm.;" "ear, 18 mm.;" noseleaf, 10.5 by 6.5; lower leg and foot, 25 mm.; "foot, 8 mm."

The measurements in inverted commas were taken in the flesh by the collector.

Skull : Greatest length, 17 mm.; basal length to front of canines, 14.2 mm.; breadth of braincase, 6 mm.; front of upper canine to back of m^3 , 6.4 mm.; front of lower canine to back of m^3 , 6.8 mm.; from palatal notch to front of incisors, 5.8 mm.; width from the outside of m^3 , 6.5. Length of free portion of canine, 2.5 mm.

Type, adult male (dried skin), in the Transvaal Museum.

The co-type, also a male, is slightly larger; forearm, 40.7; "head and body, 42 mm.;" "tail, 19 mm.;" "hindfoot, 8 mm.;" "ear, 18 mm."

Rhinolophus swinnyi rhodesiae Roberts, 1946

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Structurally similar to *R. swinnyi swinnyi* Gough, from Port St Johns, Pondoland, in respect of the flying integument starting from near the ankles and in its small size; but the tail a little longer, ear shorter and in colour bright ochraceous orange, the upper parts of body darker than the lower on account of the presence of dark-tipped hairs; the skull is slightly smaller than in the typical subspecies.

It has been previously recorded as *R. denti* (*Ann. Transv. Mus.* IV, 180; 1914).

Measurements of the type, and, in brackets for comparison, of eight specimens of *R. swinnyi swinnyi*: Head and body 52 (42 - 45); tail 22 (18 - 19); ear 15 (16 - 18). Forearm 40 (40 - 42); 3rd finger metacarpal 28.3 (27.9 - 29); 1st phalanx 12.5 (13 - 13.5); 2nd phalanx 22 (23 - 24); 4th finger metacarpal 31.5 (30.5 - 32.3). Tibia and foot (c.u.) 24 (25 - 25.5). Skull, greatest length from canines 17 (17.2 - 17.6); dorsal median length 14.7 (14.5 - 15); zygomatic width 8.3 (8.6 - 8.9); width of brain case at mastoid 8.5 (8.5 - 8.9); width across m^3 6.3 (6.2 - 6.4), across canines 4.2 (4.1 - 4.4)•

Type: T.M. No. 1325, adult F, Bezwe River, tributary of Wanetsi River, Southern Rhodesia.

Rhinolophus tridens E. Geoffroy Saint-Hilaire, 1813

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J'ai donné le nom de *trident* à ce rhinolophe, en l'empruntant de sa feuille qui est terminée par trois pointes bien distinctes; le fer à cheval; ou cette membrane située au-devant des narines, existe dans cette espèce, et dans les autres rhinolophes, tout comme dans les précédents: mais la feuille offre bien moins de complication: ce n'est qu'une lame formant bourrelet à sa base par l'effet d'un repli, et dont la partie moyenne, en-virtu de la même cause, fait corps avancé sur les bords latéraux placés plus en arrière.

Les oreilles sont plus larges que dans les précédents rhinolophes et en même temps moins fermées sur le devant: ce qui tient à une bride tégumentaire qui les attache en partie au chanfrein.

La queue est fort courte et, de plus, remarquable en ce qu'elle est dans un tiers de sa longueur, libre au-delà de la membrane interfémorale; celle-ci est coupée carrément et supplée à ce qui lui manque en longueur par plus de largeur

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Les principales dimensions du rhinolophe trident sont les suivantes.

Longueur du corps 0^m,055; - de la queue 0^m,024; - de l'envergure 0,240; - de la membrane interfémorale 0,006; - largeur de celle membrane 0,060.

J'ai trouvé ce rhinolophe en Egypte, et je l'ai figuré dans le grand Ouvrage sur l'Egypte; art. des Mammifères, pl. 2, n°. I.

Rhinopoma cystops Thomas, 1903

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Size comparatively small (forearm averaging about 52 millim., and rarely attaining 55). Nose-leaf more developed than in *R. microphyllum*. Ears proportionally large, the frontal band joining them particularly high. Tail very long and slender, longer than the forearm.

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Skull small, narrow and delicate (greatest length 16 - 17 millim., as compared with 19 - 20 in *R. microphyllum*); muzzle with two inflated projections, one on each side of and above the nasal opening, with a longitudinal groove between them. In *R. microphyllum* this region is almost flat, and its angles, although thickened, are not conspicuously inflated. Sagittal crest but little developed, not connected anteriorly with the nasal projections.

Dimensions of the type (an adult female, measured in spirit before skinning): -

Forearm 53 millim.

Head and body 53; tail 59; hind foot (s. u.) 11.5; ear 17.

Skull: greatest length 16.7; basal length 14; occipito-nasal length 14.5; zygomatic breadth 10; breadth of brain-case 7.2; front of canine to back of m^3 5.6.

Hab. (of type). Luxor, Lower Egypt. Other specimens from many Egyptian localities, southwards to Gebel Auli, Soudan (*H. F. Witherby*). Examples from Palestine (*Tristram*), Midian (*Burton*), and Aden (*Percival & Dodson*) are also assignable to *R. cystops*.

Type. Adult female. B.M. no. 2.1.17.2. Collected and presented by the Hon. N. Charles Rothschild.

Not only, however, are these two Egyptian species distinct from each other, but the members of the genus elsewhere appear to fall into two groups corresponding to them both in external proportions and in the characters of the skull.

To the first or *microphyllum* group, besides the type species, whose range extends from Egypt and Palestine (*Tristram*) to Persia (*Witherby*), there belongs the large form described below as *R. sumatræ*.

To the second, besides *cystops* itself, the species recognizable are *Hardwickei* from India and *muscatellum* from Muscat.

Apart from the differences in size and length of tail that separate the two groups, the species are very like each other externally; but the skull-characters are in all cases most obvious, and leave no room for doubt as to the distinctness of the various forms.

Rhinopoma hardwickei sennaariense Kock, 1969

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... Nach freundlicher Mitteilung von Dr. K. F. KOOPMAN existiert der Typus zu *sennaariense* FITZINGER (nec emendatio HEUGLIN 1877) im Mus. Wien und ist nach seiner Untersuchung eindeutig *hardwickei* zugehörig: dieser Name wird damit zum ältesten für eine Population verfügbaren. Da der geographische Begriff Sennar sehr verschieden Landschaft oder Provinz verstanden und gebraucht wurde, deren Grenzen Geschichte wechselten, beschränke ich den Fundort des Typus ausnahmsweise zweite Herkunftsangabe FITZINGER'S, Unter Fazogli wurde zwar ebenfalls Distrikt verstanden, der jedoch bedeutend geringere Ausdehnung als „Sennar“ Ortschaft Fazogli (heute Fazughi) liegt am linken Ufer des oberen Blauen Nil am Fuß des gleichnamigen Berges.

Rhinopoma Lepsianum Peters, 1859

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Rhinopoma Lepsianum n. sp.; supra dilute cinnamomeum, subtus flavidum; cranium regione interorbitali cordiformi, concava, intumescentiis anteorbitalibus nullis.

Long, a rostri apice ad caudae basin 0,076; caudae 0,066; cap. 0,026; antibr. 0,070; crur. 0,026; exp. alar, 0,360.

Diese Art, welche von Hrn. Lepsius am blauen Nil entdeckt wurde, ist nicht allein durch viel beträchtlichere Grösse und andere Färbung, sondern auch durch eine andere Gestalt des Schädels von *Rh. Microphyllum* verschieden. Bei letzterer ist der Schädel jederseits vor und über den Augenhöhlen durch eine bereits von Geoffroy beschriebene blasige Auftreibung aus-

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gezeichnet, welche bei dieser neuen Art ganz fehlt. Auch ist das Foramen infraorbitale bei dieser letzteren viel grösser und von länglicher Gestalt.

Rhinopoma microphyllum tropicalis Kock, 1969

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Typus: Jebel Talao, 2 km NE von Kadugli (Kordofan), 550m, 12.11.1963; M jung ad. (Balg, Schädel und Skelett): SMF 33265.

Paratype: Fundort wie Typus, 13.X.1962, 12.11.1963, 6.1.1965; 4 M, 1 F, alle ad. (2 Bälge und Schädel mit Skeletten, 3 in alc., Schädel herauspräpariert): SMF 33198-33200, 33214-33215.

Körpermaße des Typus: KR 79, Schw 73.2, Ohr 22.9, Hf 15.8, Tb 30.0, Ua 71.5, 2.F 82.5, 5.F 67.3, Spw 409.

Schädelmaße des Typus: Crn 21.0, Cbl 19.4, Mast 10.5, Hk 9.6, Zyg 12.7, c-c¹ 5.4; M-M³ 9.6, C-M³ 7.2, lor 2.6, Ukf 15.4, C-M₃ 8.1.

Körpermaße der Typeserie: KR 79 - 86 (M 81.2), Schw 64.9 - 74.5 (M 72.3), Ohr 21.1 - 22.9 (M 22.1), Hf 15.5 - 16.4 (M 15.9), Tb 20 - 30 (M 28.9), Ua 70.2 - 74.0 (M 72.5), 3.F 76.° - 84.8 (M 80.9), 5.F 66.4 - 71.3 (M 68.7), Spw 404 - 422 (M 412).

Schädelmaße der Typenserie in Tabelle 9.

Diagnose: Die neue Unterart *tropicalis* unterscheidet sich von *R. m. microphyllum* aus Unter-Ägypten durch bedeutendere Größe und allen Körper-

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und Schädel dimensionen. Der Schwanz ist relativ länger, in den meisten Fällen länger als der Unterarm. Die Größe der Hinterfüße ist übereinstimmend. Die Mesopterygoid-Gruben sind breit u-förmig. - Trotz des Geschlechtsdimorphismus ist das einzige Weibchen größer als männliche Stücke anderer Populationen. Die Färbung (SETZER 1952, HARRISON 1964a) ist nicht grau, sondern bräunlich, Unterseiten nicht weißlich, sondern heller braun.

Die Unterart *sumatrae* kommt *tropicalis* in der Größe am nächsten, ist jedoch im Mittel kleiner mit relativ kürzerem Schwanz und dunkler brauner Färbung; die Mesopterygoid-Grube ist spitz v-förmig.

Die Unterart *kinneari* ist in der Färbung am ähnlichsten, doch durch alle übrigen genannten Merkmale unterschieden, also ebenfalls durchschnittlich kleiner mit einer mehr v-förmigen Mesopterygoid-Grube.

tropicalis ist von allen bisher gefundenen Unterarten von *Rhinopoma microphyllum* geographisch getrennt. Das von ROSEVEAR (1965) aus Nigeria beschriebene Exemplar rechne ich ebenfalls zu *tropicalis*.

THOMAS (1903b) scheint keine *Rhinopoma microphyllum* aus dem Sudan untersucht zu haben, da sich bei ihm nur die "Egyptian Pyramids" als einziger afrikanischer Fundort finden, was umso überraschender ist, da gerade im Brit. Mus. ein *microphyllum*-Exemplar vom Jebel Aulia (leg. WITHERBY 1900) aufbewahrt wird (HARRISON in litt.). - Meine Nachforschungen

ergaben weitere Stücke von *microphyllum* aus Kordofan (im BM, HARRISON in litt.), worauf sich wahrscheinlich schon DOBSON (1878) bezieht, vom Jebel Azraq bei Omdurman (Coll. HARRISON) und aus dem Gebiet 10 miles N von Khartoum (HAPPOLD in litt., 1967). Dieses bisher bekannte Material ist insgesamt zur Unterart *tropicalis* zu rechnen.

Aus Sennar meldet AELLEN (1957) ebenfalls *microphyllum* (leg. 1846) (Anm. Verf.: ? evtl. von KOTSCHY). Die Identität dieses Einzelstückes bedurfte einer Überprüfung. Dr. AELLEN (Genf) teilte mir freundlicherweise einige Maße mit (Ua gebrochen, jedoch über 63, 3.F 83.5, 4.F 70, 5.F 69, Schw 60) und machte mir die Angabe, daß er dieses Stück nach WROUGHTON (1912a) determiniert habe. Auf Grund der bereits geschilderten Variationsbreite und der teilweisen Unsicherheit von WROUGHTON's Auffassungen war es bisher nur vermutlich als *microphyllum* anzusehen. Die angegebenen Längen der Finger sind nahezu identisch mit den Mittelwerten des vorliegenden Sudan-Materials und bestätigen die Zuordnung.

Rhinopterus Miller, 1906

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Rhinopterus gen. nov. (Vespertilionidæ).

Type. *Glauconycteris floweri* de Winton.

Characters. Externally like a small *Vespertilio*, but upper surface of forearm, tail, and tibia thickly sprinkled with pointed, horny excrescences resembling those on edge of ear in some *Molossidæ*, but larger. Skull differing from that of *Vespertilio* in the much greater relative breadth of anterior portion of braincase, shorter, lower rostrum, and in the form of the upper tooththrows, which are more concave on inner side and more convergent anteriorly.

Species. *Rhinopterus floweri* (de Winton).

Rhinopterus lowei Thomas, 1915

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Larger than *R. floweri*; white instead of buffy below. General characters as in *R. floweri*, but size, as gauged by

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skull, distinctly larger. Colour apparently, judged by spirit-specimens, very much as in *R. floweri* above, but the whole under surface is entirely white, while in the type of *R. floweri* it is buffy, "light buff" posteriorly, and "warm buff" on the throat and chest.

Skull much larger than in *R. floweri*, the muzzle less shortened in proportion to the brain-case.

Dimensions of the type (measured on the spirit-specimen): -

Forearm 40 mm.

Head and body 45; tail 36; ear 12.5; third finger, metacarpal 36.5, first phalanx 11.5; lower leg and hind foot (c.u.) 12.

Skull: greatest length 13.5; condyle to front of canine 12.9; zygomatic breadth 9; interorbital breadth 4.6; intertemporal breadth 3.2; palato-sinual length 5.3; front of canine to back of m^3 5.

Hab. Sudan. Type from the Blue Nile, 20 miles above Sennar.

Type. Adult male. B.M. no. 15.3.6.70. Collected 13th December, 1913, by Willoughby P. Lowe.

Mr. Willoughby Lowe obtained five specimens of *Rhinopterus* at different localities on the Blue and White Nile. On examination of the skulls these prove all to belong to a larger form of the genus than *R. floweri*, living side by side with the latter in the same area - a phenomenon not unusual with the smaller bats. The *Rhinopterus* obtained by Dr. A. F. Wollaston at Shendi, and referred to in Anderson and de Winton's 'Mammals of Egypt,' also belongs to this larger form. Whether the difference in the colour of the under surface is constant remains to be seen.

Rousettus aegyptiacus thomensis Feiler, Haft and Widmann, 1993

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Holotypus: MTD, B 16753, M, São Tome, bei Generosa, ca. 800 m ü. NN, 26.3.1991, Balg und Schädel.

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Paratypen: M, São Tome, bei Generosa, ca. 800 m ü. NN, MTD, B 16750, 27.3.1991, B 16751, 28.3.1991, Balg und Schädel; Staatl. Museum für Naturkunde Stuttgart, 8 1803, 1989, in Alkohol. F, Sao Tome, bei Generosa, ca. 800 m ü. NN, MTD, B 16752, 20.4.1991, B 16754, 19.4.1991, Balg und Schädel, B 16755, 26.3.1991, in Alkohol.

Diagnose: Die Tiere von São Tome besitzen eine extrem dunkle Oberkopffärbung, die sie von den Tieren des nahen westafrikanischen Festlandes unterscheiden. Morphometrisch gleichen sich beide Formen nahezu, wie ein Vergleich mit EISENTRAUT (1963) und seinem Material ergab. - Bemerkenswert ist das Vorkommen einer eigenen Fledermausfliegenart (*Eucampsipoda iterfacta* KOCK, 1993).

Das Gaumenfaltenmuster für die Exemplare von São Tome stimmt mit jenen des Festlandes überein (EISENTRAUT, 1976). Von uns untersuchte 4 MM und 4 FF hatten vom je 4 durchgehende Falten, gefolgt von je 4 unterbrochenen Falten mit Ausnahme eines M, das nur 3 unterbrochene Falten besaß. Die letzte, das Muster abschließende Falte war immer durchgehend. Die Formel lautet folglich: 4 - 4/3 - 1. Interessanterweise hatte EISENTRAUT (1963), wie wir auch, bei annähernd gleich vielen Exemplaren eine Farbmutante dabei: ein extrem helles und rotbraunes Tier. - Da Gewichtsangaben bei anderen Autoren meistens fehlen, teilen wir die folgenden für unser Material mit: 8 MM, 130 - 170 g, Mittelwert 153 g; 3 FF, 130 - 145 g, Mittelwert 135 g.

Nach einem Nachweis der Art (FEILER, 1984) schlossen sich weitere 1989 durch J. HAFT und J. FAHR und später 1991 durch alle Teilnehmer unserer São Tome-Reise an. Sie ist von der Küste bis in die Berge hinein verbreitet, nur auf der Spitze des Pico de São Tome haben wir sie nicht beobachtet. In der Bucht von Angra Toldo befindet sich ein *Rousettus*-Quartier. Die Höhle ist von See aus gut zu sehen und nur schwer zugänglich. Es ist eine Meeresgrotte, die am Wasserspiegel etwa 1.5 m breit ist und bis in eine Höhe von 7 bis 8 m reicht, wo sie spitz ausläuft.

R. aegyptiacus thomensis war des Nachts sowohl durch seine klickenden Ultraschall-Laute als auch durch kreischende Soziallaute auszumachen. Große Mengen von Höhlenflughunden fanden sich allabendlich in fruchtenden Brotfruchtbäumen nahe der Hauptstadt ein. In Gefangenschaft bevorzugten die Tiere Nektarersatz als Nahrung. Die bevorzugten Früchte sind hier Mango und noch leicht grüne Bananen. In Mägen freilebender Tiere fanden wir Samen, die wahrscheinlich von *Musanga spec.* stammten. Die Flughunde nahmen auf São Tome auch im Lager herumliegende Papayas und Bananen auf.

Nach HAFT fingen sich die 1991 gefangenen Tiere (32 Ex.) an zwei verschiedenen Fangorten und bei annähernd gleichem Wetter zu unterschiedlicher Zeit: in São Nicolau ausschließlich vor 0 Uhr, meist zwischen 20 und 22 Uhr, in Cascata verstreut über die Nacht, viele jedoch erst gegen Morgen.

Die angesaugten Zitzen mehrerer gefangener Weibchen (26. u. 28.3.) lassen auf Jungtiere schließen. Ein flugfähiges Jungtier wurde im Juni 1983 auf São Tome in einem knapp über dem Boden hängenden Netz gefangen. FAHR hatte in Makokou (Gabun) am 2. und 3. Oktober relativ kleine Jungtiere vorgefunden (Unterarmlänge etwa 80 mm). EISENTRAUT (1963) stellte in Kamerun Ende November ein neugeborenes Jungtier fest, einige Weibchen waren zu dieser Zeit trächtig. Trächtige Weibchen fand er ebenfalls im Januar. Unsere Beobachtungen bestätigen einmal mehr, daß die Art keine eng begrenzte Fortpflanzungszeit hat.

Rousettus angolensis ruwenzorii Eisentraut, 1965

p. 3

Typus: M ad, Ruwenzori East, 5000 ft., März 1906, R. E. Dent leg. (British Museum Natural History, London, Nr. 6.12.4.1.). Kopf-Rumpf = 117, Schwanz = 13.5, Hinterfuß = 17.5, Ohr = 24.5, Unterarm = 76, Tibia = 31; größte Schädellänge = 43.1, Hirnkapselbreite = 16.1, Jochbogenbreite = 25.8, Interorbitalbreite = 6.9, Intertemporalbreite = 8.0, maxillare Zahnreihe = 16.7, Breite über oberen C = 9.0, Breite über oberen M = 13.3, Mandibellänge = 33.5, mandibulare Zahnreihe = 19.0.

Die Ruwenzori-Rasse, von der ich im Britischen Museum außer dem zum Typus ausgewählten Stück 6 weitere Exemplare (2 MM ad, 3 FF ad, 1 F sad), vergleichen konnte, stellt also die größten Vertreter des Rassenkreises. Außerdem zeichnet sie sich durch ihre besondere Langhaarigkeit gegenüber der Nominatrasse aus, wohingegen die mir zur Verfügung

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stehenden Stücke der Rasse *smithi* von Guinea besonders kurzhaarig sind. Im Durchschnitt beträgt die Länge der Rückenhaare bei *smithi* von Guinea 8 mm, bei *angolensis* von Kamerun 8.5 - 9 mm, bei *angolensis* von Fernando Poo 9 - 10 mm und bei *ruwenzorii* vom Ruwenzori East 11 - 12 mm.

In Abb. I kommt die unterschiedliche Schädelgröße der 3 Rassen noch einmal deutlich zum Ausdruck. Ferner ist in Abb. 2 die an dem vorhandenen Material festgestellte Variationsbreite der Korrelation von größter Schädellänge: Jochbogenbreite graphisch wiedergegeben. Diese Darstellung veranschaulicht noch einmal die fortlaufende Größenentwicklung der einzelnen Populationen von West nach Ost.

Entsprechend der Zunahme der Schädelmaße finden wir auch eine Zunahme der Zahngröße. Die durchschnittswerte der Zahnmaße sind in Tabelle 2 zusammengestellt.

[table removed - eds.]

Bezüglich der Färbung des Rückenfalls sind die Guinea- und Ruwenzori-Stücke jeweils relativ einheitlich. Erstere sind die hellsten von allen. Ihre Tönung ist mittelbraun und variiert ein wenig von brussels brown bis sudan brown. Die Stücke vom Ruwenzori haben einen tiefen kastanienbraunen Ton, der als brownish olive bis saccardo's umber zu bezeichnen ist. Innerhalb der beiden mir zur Verfügung stehenden sehr großen Serien vom Kamerunberg und von Fernando Poo ist die

Variation der Färbung relativ groß und reicht etwa von sudan brown über saccardo's umber bis mummy brown. Ganz allgemein möchte ich jedoch der Färbung keine allzu große Bedeutung für die Rassencharakterisierung bei *R. angolensis* beimessen.

Leider liegen über das Vorkommen des Angola-Flughundes in einigen Teilgebieten innerhalb des Areals bisher offenbar keine oder nur sehr allgemeine Angaben vor, so z. B. für Nigeria, Ghana, Südkamerun und Gabun. Im Kongogebiet scheint die Art nur sehr sporadisch vorzukommen (vgl.

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u.a. Schouteden 1948; Frechkop 1954). Hier wäre noch zu untersuchen, ob die entsprechenden Stücke zu *angolensis* zu rechnen sind, oder bereits Annäherung an die Ruwenzori-Rasse erkennen lassen. Es ist durchaus möglich, daß ein spärliches Vorkommen oder vollständiges Fehlen in manchen Gegenden mit dem Mangel an geeigneten Biotopen zusammenhängt. Ich selbst kenne die Art aus dem Gebiet des Kamerungebirges und auf Fernando Poo als einen sehr häufigen Vertreter der Megachiropteren und als typischen Felshöhlenbewohner. Demgegenüber finde ich nur einmal, und zwar bei Hayman (1954), die Angabe, daß die von ihm untersuchten Togo-Stücke in hohlen Bäumen gefangen wurden. Da in den von mir besuchten Gebieten Felshöhlenquartiere relativ häufig sind, erklärt sich daraus wohl das beobachtete Massenaufreten. Vielleicht beruht das Fehlen der Art in anderen Gegenden auf dem Mangel an solchen geeigneten Tagesquartieren.

Rousettus kemp Thomas, 1909

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Allied to, but with heavier teeth than, *R. lanosus*.

External characters as in the Ruwenzori Mountain Fruit-bat, *R. lanosus*, except that the long hairs of the back, rump, and interfemoral region tend to be greyer. Palate-ridges 4 - 3 - 1.

Skull as in *lanosus*, the brain-case equally strongly deflected.

Teeth larger, broader, and heavier throughout, more as in ordinary Rousets, not so highly specialized as in *R. lanosus*. Incisors, canines, and anterior premolars as in *lanosus*. P^4 3 mm. in horizontal length as against 2.4 in *lanosus*; m^1 3.0 x 1.5 (*lanosus* 2.5 x 1.2); m^3 2.8 x 1.5; p_4 2.4; m_1 2.9 X 1.4; m_2 1.9 X 1.4.

Forearm of the type 86 mm.

Head and body (measured in flesh) 125 mm.; tail 15; ear 23.

Hab. Kirui, Mt. Elgon, British East Africa. Alt. 6000'.

Type. Subadult male. Rudd Collection. Original number 269. Collected 16th September, 1909.

This is evidently the Elgon representative of the Ruwenzori *A. lanosus*; the Shoa specimen mentioned by Dr. Andersen would appear, as is natural, to be the same as the Elgon rather than the Ruwenzori species.

Rousettus lanosus Thomas, 1906

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Rousettus lanosus, sp. n.

A member of the *R. collaris* group; the limbs thickly hairy.

Size and general characters about as in *R. collaris*. Fur loose and shaggy, very abundant, its limits not sharply defined as in *R. collaris*; that on the head about 8 mm. in length, mixed with a large number of much longer hairs, attaining over 20 mm. On the back the fur is thick and woolly, passes out on to the proximal half of the upperside of the forearms, and is continued thickly down the hind limbs and the wing-membrane external to them to the ankles, the feet being also thinly clothed above. Interfemoral thickly furry above, except just along its posterior margin. Below, the wings are thinly hairy to a point level with the middle of the forearm, the hind limbs and interfemoral membrane being also less thickly haired than on the upper surface. Ears narrow, rather longer than the muzzle. Palatal ridges as in *R. collaris*.

General colour above bistre-brown, becoming warmer posteriorly; head darker. Under surface near broccoli-brown, but with a yellowish suffusion; some of the longer hairs quite yellow.

Skull rather more delicately built than in *R. collaris*, the bones thinner and lighter, muzzle rather longer and narrower; postorbital processes very thin; anterior palatine opening unusually broad.

Teeth conspicuously smaller throughout, each molar and premolar slightly shorter and very much narrower than in the allied species.

Dimensions of the type (measured on the spirit-specimen): -

Forearm 88 mm.

Head and body 134; tail 16, tail free from membrane 9; head 46; ear 23; third finger, metacarpus 60, first phalanx 41, second phalanx 57; lower leg and foot (c. u.) 62.

Skull: length to tip of nasals 42; basal length 37.5; zygomatic breadth 25; breadth of brain-case 17.4; palate length from anterior palatine foramina 19; breadth of palatine foramina 3.4; length of upper tooth-row from front of canine 14.5; the same below, 16.2; first upper molar 2.3 x 1.2.

Hab. Ruwenzori East, at 13,000'.

Type. Adult male (in spirit). B.M. no. 6.7.1.2. Collected by K. B. Woosnam. Three specimens obtained.

This species is allied to *R. collaris*, but is at once distinguishable from that and every other species by the thick

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woolly covering of its hind limbs and its remarkably narrow molars.

The occurrence of a fruit-bat at such an altitude as 13,000' is very noteworthy.

Rousettus sjöstedti Lönnberg, 1908

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Usambara: 1 specimen from the Mkulumusi caves near Tanga, ⁴/₅, 1905.

Resembling *R. angolensis* (Bocage). Fur woolly, greyish brown above, rather pale brownish grey beneath, a little lighter on the neck and this light colour extends up on the sides of the neck so as to form a collar which is narrowed and almost interrupted on the back. Head coloured like back, darker on snout and on both eyelids. Wing-membrane blackish speckled with light spots. Not quite half the forearm is hairy. Tibia naked. On the lower side the forearm is woolly in a similar degree as above but the wing-membrane is sparsely beset with woolly hairs between humerus and forearm and along the outer side of the latter. Tibia almost wholly naked below. Interfemoral membrane above and below with some very few scattered hairs. Wing-membrane rising from back of second toe but at the end curving to the interspace between first and second.

Palatal elevations six in number, three anterior complete, and three posterior mesially interrupted (in addition to these there is a rudiment of a fourth interrupted fold on one side between the last and next last). Behind these there is an angular serrated fold on the back of the palate.

Dimensions of *R. sjöstedti* (in spirit, F); *R. angolensis* (according to BOCAGE) M, F (in mm).

Length of head and body: 125; 120, 140

" " tail: 17; 13, 13

" " head: 43; 45, 48

Distance from eye to nostril: 16; 15, 15

Length of ear: 25; 23, 22

Forearm: 86; 80, 79

Thumb: 34.5; 32, 33

Metacarpus of third finger: 59; 56, 60

First phalanx of third finger: 38 (39); 40, 40

Second " " " " : 53; 50, 53

Metacarpus of fourth " " : 57; 53, 56

First phalanx of " " : 32; 26, 26

Second " " " " : 35; 28, 29

Tibia: 35; 32, 33

Hindfoot: 25; 20, 21

Metacarpus of fifth finger: 56; -, -

First phalanx of fifth finger: 28.5; -, -

Second " " " " : 26; -, -

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Length of skull of *R. sjöstedti* about 40 mm. Frontal region of skull between postorbital processes somewhat convex. Molars not very narrow transversal diameter of first molar 2 mm. and last premolar still thicker.

This new bat is most nearly related to *Rousettus angolensis* (BOCAGE) and differs like that one from *R. leachi* (A. SMITH) = *R. collaris* auct., which lived in the same caves, most conspicuously with regard to the structure of the palate. Unlike *R. leachi*, the new species has only 3 complete palatal elevations and behind those 3 mesially interrupted folds. The presence of only 3 complete palatal elevations was pointed out by BOCAGE as a characteristic for the by him 1898 described species *angolensis*. MATSCHIE used the same characteristic as well for the definition of his new genus *Myonycteris*, which comprised the species *torquata* and *angolensis*. The latter has, however, proved to a true *Rousettus* by cranial characteristics. The same is also the case with the present new species. As a specific characteristic the number of complete palatal elevations serves very well,

and by among other characteristics the new species is easily separated from another East African bat *R. lanosus*. From *R. angolensis* with the same number of complete palatal elevations, *R. sjöstedti* differs in the structure of the palate having only three (instead of four) mesially interrupted posterior palatal folds. This characteristic may, however, be variable as in the type specimen there are rudiments on one side of a fourth divided fold between the second and third. In addition to this there are many differences in dimensions and other exterior characteristics as is proved by the description above. *R. angolensis* is more hairy, for instance, on the tibia and interfemoral membrane. The former is said in *R. angolensis* to be covered "en dessous de poils longs et serrés" --- "jusqu'à l'articulation du pied", --- "en dessous jusqu'au premier tiers de la jambe" and the latter "presque entièrement couverte en dessous de poils aussi long at aussi fournis que ceux du dos", while in *R. sjöstedti* both these parts are practically naked.

The tibia of *R. sjöstedti* is longer than that of *R. angolensis* but shorter than that of *R. lanosus*. The latter is easily recognized by its small molars.

Rousettus smithii Thomas, 1908

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Most nearly allied to *R. angolensis*, with which it forms a distinct section of the genus, but differing in the following characters: - Size much smaller, the skull also narrower and with less widely expanded zygomatica. Fur shorter and more resembling that of ordinary Rousets (that of *R. angolensis* being unusually long and silky), and not extending so far down the hind limbs, the proximal half only of the tibiae being clothed. Ears narrower. Colour dull brown without rufous suffusion; neck more greyish.

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Skull more lightly built than in *R. angolensis*, but agreeing with it in all essential respects, such as the very slight deflection of the brain-case, the co-ossification of the premaxillae, and the swollen supraorbital margins. Teeth of the same squarish form, but smaller throughout, and similar in relative proportions, with the exception that the last molar, both above and below, is very much smaller, about one-third instead of one-half the size of the tooth immediately preceding it.

Dimensions of the type (not fully adult): -

Forearm 70 mm.

Head and body (c.) 112; tail 11; pollex (c. u.) 28.5; third finger, metacarpal 49.5, first phalanx 32.5, second phalanx 41; lower leg and hind foot (c. u.) 46.

Skull: greatest length 38.5; zygomatic breadth 20.5; supraorbital foramina to tip of nasals 18; breadth of braincase 15; front of canine to back of m^3 14.8; p^4 2.3 X 1.8; m^2 1.4 x 1.2; p_4 2.7 x 1.7; m_3 1.3 x 1.1.

Hab. Sierra Leone.

Type. Nearly adult female. B.M. 8.9.11.1. Collected and presented by Canon F.C. Smith.

The many important characters by which *Rousettus angolensis* differs from all other members of the genus have recently been brought out in Dr. K. Andersen's admirable notes on the group, so that no comparison of *R. smithii* with other species is required. From *R. angolensis* it is at once distinguishable by its smaller size (allowing, of course, for the slight immaturity of the type), smaller teeth, and, especially, by its much smaller posterior molars.

I have much pleasure in naming this Rouset after its discoverer, to whom the National Museum is indebted for various acceptable specimens.

Scabrifer notius G.M. Allen, 1908

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Type. - Adult male, alcoholic, No. 4555, Museum of Comparative Zoölogy; collected at Cape Town, Africa, and received from E. L. Layard.

General Characters. - Size as in *S. floweri* (De Winton), but the fur very dark basally, with light tips. The curious wart-like papillae characteristic of this genus

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are sparsely scattered on both surfaces of the humerus, the forearm, finger bones, tail, and tibiae.

Description of the Type. - Color of the fur in alcohol dorsally and ventrally nearly mummy brown, lighter, approaching hair brown at the tips. As the type has been immersed in alcohol for many years, it is possible that the fur is still darker in the living animal. The ears and muzzle are pale.

Dorsally the fur covers the body thickly, but is practically absent from the membranes. The bases only of the ears are clothed, and the somewhat swollen muzzle is thinly clad with short minute hairs and sparser longer ones. Ventrally the membranes are likewise naked, save for the proximal half of the tail and interfemoral, which are thinly covered with short hairs.

The ear, laid forward, reaches nearly to the nostril; it is rounded at the inner basal angle, then nearly straight in outline to the broadly rounded tip. Externally there is a slight concavity below the tip, and the lower half is again nearly straight to the basal notch that separates off the low rounded terminal lobe. The tragus is narrow, bluntly pointed, and rather short.

The wing membrane arises from the base of the toes. The calcar is strongly keeled, but without a terminal lobe. The tip of the tail is free for about 2.5 mm. The curious wart-like papillae characteristic of this genus seem much fewer than in *S. floweri* from the Soudan and are practically absent from the membranes, whereas in the latter species the upper side of the forearms, legs, and tail, and all the proximal portion of the wings and interfemoral membrane are thickly studded with them. In the new species there are dorsally some half-dozen papillae on the humerus, a few at the proximal end of the forearm, but a thicker cluster of from 15 to 20 at the distal end of the forearm; a few are also scattered along the metacarpals and phalanges, and on the dorsal surface of the femora, tibiae, and metatarsals, as well as at the region of the joints on the upper side of the tail. Ventrally the distribution of these warts is about the same, but they are more numerous on the humerus and almost wholly lacking on the fingers, tibiae, and tail.

Skull and Teeth. - As stated by Miller, the skull resembles that of a small *Eptesicus*, and is somewhat flattened, with an almost straight dorsal profile. The inner upper incisors are long and slender with a faint indication of a cusp near the tip, laterally. The outer incisor is not quite half the height of the inner, and like that tooth has a strongly developed cingulum that forms a slight cingulum cusp. The outer cusps of the mandibular molars are long and rather slender.

Measurements: - The external measurements of the type are as follows: head and body, 49; tail, 37; ear, 13.5; tragus, 5; forearm, 35; thumb, 5.3; 2d digit, metacarpal, 30.5; 3d digit, metacarpal, 32.3; 1st phalanx, 12.7; 2d phalanx, 9; 3d phalanx and tip, 7; 4th digit, metacarpal, 32.5; 1st phalanx, 12; 2d phalanx and tip, 8; 5th digit, metacarpal, 32.5; 1st phalanx, 9.2; 2d phalanx and tip, 5.6; tibia, 12.8; foot, 6.

Skull; greatest length, 14.3; palatal length, 6.6; interorbital constriction, 3.6; mastoid breadth, 8; palatal breadth outside second molar, 5.6; mandible, 10.3; maxillary tooth row (exclusive of incisors), 5; mandibular tooth row (exclusive of incisors), 5.5.

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Remarks. - *Scabrifer floweri* from the White Nile and Soudan seems to be a pallid species, while the new species here described is dark-furred, with a much less development of the dermal papillae that give an encrusted appearance to those parts where they occur. The two appear not to differ in size.

Scoteinus schlieffeni australis Thomas and Wroughton, 1908

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M. 1967, 1975, 1993, 2000, 2005, 2007. F. 1995, 2001.

On laying out all the specimens of this species in connection with the identification of the present series, it became evident that there are several well-marked geographical races separable on colour characters.

Typical *S. schlieffeni* was based by Peters on a specimen from Cairo. He described it as "supra rufescens, subtus ex albo rufescens." A second species, *S. minimus*, based on a F from Tanganyika was described by Noack as "oben olivengelbbraun unten weissgelb. Seiten hell umbra" Unfortunately we have no undoubted specimen of either of these for comparison, but we consider that we are justified in describing three forms as certainly distinct from either typical *S. schlieffeni* or *S. minimus* (which latter is at most a local race of the former). These are: (1) a pale desert form from the Aden Hinterland; (2) a white-bellied desert form from Upper Egypt; and (3) the present series from S. Africa.

[possibly incomplete - eds.]

Scoteinus schlieffenii fitzsimonsi Roberts, 1932

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Differing in colour from its nearest geographical allies *S. s. australis* Thomas and Wroughton, of the eastern littoral and *S. s. minimus* Noack, of Central Africa in being clay colour above and cinnamon buff to pinkish buff below.

Type : T.M. No. 6550, adult M, Tsotsoroga Pan, Northern Bechuanaland, 19th June, 1930, No. 833. Vernay-Lang Kalahari Expedition. Length of head and body 48, tail 32, hind foot (c.u.) 6, ear 11, forearm 30. Skull : length, canines to occiput, 12.3, zygomatic width 8, width of brain case 6.2, width of palate across outside of m² 5.5, length of upper tooth-row (c-m³ 4.3 mm.

Scotæcus Thomas, 1901

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Like *Scotophilus*, but with the following special characters: - The two lower premolars are subequal, instead of the first being far smaller than the second; the upper canines are flattened and grooved anteriorly; the last upper molars have much larger posterior lobes, so that they are triangular instead of equally narrow externally and internally; the talon of the last lower molar is bi- or tricuspidate and practically as large in cross-section as the main part of the tooth; the brain-

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case is decidedly more flattened and less dome-shaped; and both nasal and anterior palatal notches are much deeper, the latter reaching nearly to the line of the front of m^1 .

Type "*Scotophilus*" *albofuscus*, Thos.

In my first description of it *S. albofuscus* was only placed provisionally and with great doubt in *Scotophilus*. Mr. de Winton, in describing *S. hirundo*, expressed his opinion that neither were properly congeneric with the previously described species, and, now that a third member of the group has turned up, I think it advisable to form a special genus for all three. In the characters above noted the three precisely agree, and differ from any of the members of the true *Scotophilus*.

In some respects, perhaps, *Scotæcus* is even more nearly allied to the American *Nycticeius*, which shares with it the triangular m^3 and more complicated m^3 , but still has, like *Scotophilus*, the marked disproportion between the sizes of the two lower premolars, the canines rounded in front, and the comparatively shallow nasal and prepalatal notches

Scotæcus albigula Thomas, 1909

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Closely allied to *S. hindei*, Thos. (locality Kitui, Ukamba), but the whole under surface nearly uniform whitish, the bases of the hairs only faintly greyer (brownier on the lower abdomen), the whitish underside of neck in particular contrasting markedly with the brown upperside. Fur a little softer and more woolly, as is natural at the higher elevation. A band of fine whitish hairs along the membrane just external to the forearm, this part being quite naked in *S. hindei*.

Skull slightly longer than that of *hindei* and its palate rather broader. A minute p^1 present on each side in, or close behind, a deep notch in the cingulum of the canine. No such tooth is present in the type of *S. hindei*, but it occurs in the specimen from Nyasaland mentioned by Mr. Wroughton in 1907.

Dimensions of type (the starred measurements taken in the flesh): -

Forearm 38 mm.

Head and body * 58; tail * 35; ear * 13; third finger, metacarpal 36.5, 1st phalanx 12.5; lower leg and hind foot (c. u.) 20.

Skull: condylo-basal length 14.5; basi-squial length 11; front of canine to back of m^3 6; breadth across palate outside molars 7.8.

Hab. Kirui, Mt. Elgon, Brit. E. Africa. Alt. 6000'.

Type Adult male. Rudd Collection. Original number 200. Collected 5th September, 1909, by R. Kemp.

Scotoecus artinii de Beaux, 1923

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(N. 1443). Tipo 1 F, a secco, cranio estratto, danneggiato dopo la misurazione. Paratipi 1 M, 1 F, in alc., crani non estraibili.

Archers Post, 26 12.1919. L. FRANCHETTI et L. TONKER.

Dedico la presente specie al chiarissimo Prof. E. Artini, Direttore del Museo civico di Storia Naturale di Milano.

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Il rivestimento peloso ha brillantezza sericea. La colorazione d'insieme delle parti dorsali varia secondo i soggetti dal bruno cuoio nel Tipo (snuff brown, Ridgw. XXIX) al bruno oliva nel % (olive brown, XL), ed i singoli peli mostrano l'apice decisamente più chiaro. Le parti ventrali si avvicinano alla colorazione dello *S. albigula*, Thos. (Ann. M. Nat. Hist. IV, 1919, p. 544): la gola è bianca, il torace è superficialmente quasi peli brunastra (wood brown, XL); sull'addome il colore bruno aumenta quantitativamente. Il muso è bruno o bruno nerastro secondo i soggetti. Il mento è nerastro o porta almeno una larga macchia nerastra, diffusa, marginale mediana.

Il patagio è bruno nerastro, sottilmente marginato di chiaro e mostra sulla superficie ventral lungo l'avambraccio dei minutissimi peluzzi chiari scarsissimamente disseminati su di una striscia larga 5 mm.

Il palato è molto largo. Un piccolo P_1 è presente in tutti egli esempl.

Misure del Tipo (e paratipi: F - M): Avambraccio 32 (34 - 34). Testa e tronco ca. 53 (54 - 51). Coda 26 (29 - 26). Dito terzo, metacarpo 31 (31 - 31), prima falange 11.5 (12.5 - 11.5). Tibia e piede con unghia 10.5 (19.5 - 19). (Lunghezza del pene nel prepuzio 8.5). Il tipo fu misurato *prima* di essere messo in pelle.

Cranio del Tipo. Lungh. Occipite - C 14.2. Lungh. condilo-basale 13. Lungh. basi-sinuale 10. Larghezza orbitale 6.5. Largh. intertemporale 5. Larghezza della cassa cranica 8. Largh. mastoidea 8.1. Larg. estern massima sulle file dentali molari 7.5, Fila $C-M_3$ 5.8; P_4-M_2 3.8; $C-M_3$ 6.1.

Sc. artinii si distingue facilmente dalle specie geograficamente più vicine. Dal *hindei*, Thos. (Ann. M.N. Hist., VII, 1901, p. 264), Kitui, Br. E. Afr., pel colore delle parti dorsali che in quest'ultimo è molto più scuro (chocolate, XXVIII) e delle parti inferiori che in quest'ultimo sono brune. Dall'*albigula*, Thos. (o.c. IV, 1909, p. 544), Mt. Elgon, che, come cortesemente mi confermano con lettera del 1.II.1923 i signori Kershaw e Hinton del Museo Britannico, ha le parti dorsali bruno cioccolato, pel colore delle medesime e per le parti in-

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feriori decisamente meno chiare nell'*artini*. Dal *woodi* Thos. (o.c. XIX, 1917, p. 280), Nyasaland, per il colore delle parti superiori, che in quest'ultimo sono molto più scure (mummy brown XV), della membrana alare che in quest'ultimo è bianca e delle parti inferiori che nel medesimo sono brune. Dal *cinnamomeus*, Wettst. (Anz. Ak. Wissensch. Wien. 53, 1916, p. 191), Cordofan, per il colore delle parti superiori, che, come mi conferma l'autore, corrisponde in quest'ultimo al *cinnamon* di Ridgw. XXIX e per il colore del patagio, che nel medesimo non ha solo il margine ma anche intere porzioni marginali bianche.

Spiccate differenze di colorazione lo distinguono pure dalle specie geograficamente più lontane: *albofuscus*, Thos. (Ann. M. C. Genova, XXIX, 1890, p. 86), Gambia, che è più scuro ed ha le ali biancastre; *hirundo*, Winton (Ann. M. N. Hist. IV, 1899, p. 355), Costa d'Oro, che è molto più chiaro (grigio topo); *falabae*, Thos. (o.c. 1919, XVI, p. 447), Kabwir, Nigeria, sett. che ha i singoli peli più chiari nella porzione basale che nella apicale.

Debbo infine aggiungere, che a mezzo di un accurato controllo della colorazione di *Pipistrellus nanus*, Pet., *Scoteinus schlieffenii africanus*, Allen, e *Taphozous mauritanus*, Geoffr., conservati in identiche condizioni mi sono assicurato che l'alterazione di colore subita dai soggetti nell'alcool è assolutamente minima.

Scotoecus cinnamomeus Wettstein, 1916

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Am ähnlichsten *S. albofuscus* (Thom.), aber Rücken "cinnamon", Unterseite weißlich, Tragus länger als bei allen gleichgroßen Formen. Flughautrand zwischen fünftem Finger und Tibia weiß gesäumt, mit weißer Verbreiterung in der Mitte dieser Strecke. Schwanzspitze über 1 mm frei aus der Flughaut herausstehend. Außenrand des Ohres gerade. Innenrand des Tragus gerade, 3 mm lang, Außenrand etwas unter der Mitte des Innenrandes stark konvex, dann gerade, am Ende schmal abgerundet.

Maße des Typus: Kopf und Körper 47, Schwanz 33, Unterarm 32, Ohrhöhe 12, Außenrand d. Tragus 4.8, Tragusbr. 2 mm.

Typus: 1 F Nubbaka, Kordofan, 17.III.1914

Cotypen: 1 F Nubbaka, 17.III

1 F Dilling, Kordofan, 20.III.1914.

Scotoecus falabæ Thomas, 1915

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Scotoecus falabæ, sp. n.

A medium-sized brown species, rather larger than *S. hirundo*.

General characters very much as in *S. hirundo*, to which the new form is most nearly allied. Size rather larger.

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Fur 3.5 mm. in length on the shoulders. General colour sepia-brown, but the light bases of the hairs showing through, especially across the shoulders, where the hairs are prominently buffy whitish for the greater part of their length, the tips only brown. Under surface dull buffy whitish, the throat clearer white, but nowhere so pure a white as in *S. hirundo*. Membranes blackish brown throughout. Tragus short, broad, rounded, slightly incurved, the inner margin concave. Wings to the end of the metatarsus. A distinct postcalcarial lobule present.

Skull, as compared with that of *S. hirundo*, larger, more rounded, higher, and less flattened than in *S. hirundo*, and still more so in comparison with the other species. The interorbital broadening less noticeable than in any known species.

Dentition as in *S. hirundo*, except that minute anterior premolars are present on each side, standing in the notch in the posterior base of the canines.

Dimensions of the type (the italicized measurements taken in the flesh): -

Forearm 36 mm.

Head and body 51; *tail* 36; *ear* 12; third finger, metacarpus 34, first phalanx 11.2; lower leg and hind foot (c.u.) 20.

Skull: occiput to base of canine 13.7; condylo-basal length 13.3; interorbital breadth 6.8; intertemporal breadth 4.8; breadth of brain-case 8.5; front of canine to back of m^3 5.6; front of p^4 to back of m^2 3.7.

Hab. Kabwir, Northern Nigeria. Alt. 2500'.

Type. Young adult female. B.M. no. 15.10.8.1. Original number 70. Collected July 28, 1915, and presented to the National Museum by Dr. J. C. Fox.

This species differs from *S. hirundo* by its larger size, higher skull, and more obviously bicolor fur. The type of *S. hirundo* is a very old female with worn teeth.

The inconvenient variability in the presence or absence of the small upper premolars is a character of *Scotoecus* unique in the Vespertilionidæ. How far these teeth will prove to be generally constant within each species remains to be seen, but in *S. hindei*, at least, they are absolutely inconstant.

The specific name is given in commemoration of Dr. Fox's escape from the peaceful liner 'Falaba,' barbarously sunk on its way out to Africa.

Scotoecus Hindei Thomas, 1901

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Size largest of the genus. Fur close and soft, hairs about 3 mm. in length on the back; not extending either above or below on limbs or membranes, except slightly on and around the base of the tail. General colour above chocolate-brown, the bases of the hairs darker; below the colour is paler, the hairs being brown basally and dull buffy terminally. Muzzle and chin blackish brown. Ears of medium length, their tips rounded, with a slight and even concavity below them. Tragus short, of equal breadth above and below, an indistinct lobe at its external base. Wings to the metatarsus. Postcalcaneal lobe well-defined. Penis very long, apparently provided with a bone.

Skull very similar to that of *S. hirundo*, but larger and heavier throughout; nasal notch very deep, its depth, measured from the base of i^1 , going only about three times in the distance from it to the occiput; prepalatal notch reaching to the level of the front of m^1 ; sagittal crest fairly developed.

Upper incisors short, stout, strongly convergent. Lower incisors transverse, slightly overlapping. Anterior lower premolar three fourths the height of the posterior one.

Dimensions of the type: -

Forearm 35 millim.; height of ear 12, of tragus (on inner edge) 2.4.

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Skull: greatest length 15; basal length 12.9; orbital breadth 8; intertemporal breadth 4.9; breadth of brain-case 8.2; nasal notch 3.3×2.5 ; palate from bottom of prepalatal notch 5; front of upper canine to back of m^3 6; front of lower canine to back of m^3 6.5.

Hab. Kitui, British East Africa; altitude 3500 feet.

Type. Male. B.M. no. 1.2.5.1. Original number 63. Collected 26th November, 1900, by S. L. Hinde.

Its larger size and different colour will readily distinguish this species from its nearest ally *S. hirundo*.

Scotoecus woodi Thomas, 1917

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Near *S. albofuscus* of the Gambia, but smaller.

Size about the smallest of the genus. General colour above dark brown (near mummy-brown), the tips of the hairs paler brown; under surface little paler, near Prout's brown. Wings coloured as in *S. albofuscus*, the forearms, digits, hind limbs, and tail blackish, the membranes internal to a line from elbow to knee, and the interfemoral dark brown, those external to forearms dull whitish, rather darker; terminally. Ears short, with large external basal lobe; tragus short and broad, its inner margin slightly concave.

Skull short and stumpy, of the characteristic broad shape usual in the genus, the lacrymal breadth even greater than in *S. albofuscus*. Nasal notch very deep. Median part of zygoma absent in type.

Incisors slender, their bases not touching the canines. Canines broadened transversely, their basal area broader than long, and flattened behind, close and parallel to the front edge of the large premolar; no small premolar or place for it present.

Dimensions of the type (the italicized measurements taken in the flesh): -

Forearm 28.5 mm.

Head and body 56 mm.; tail 21; ear-12.

Third finger, metacarpus 28, first phalanx 10, second phalanx 8; lower leg and hind foot (c. u.) 17.5.

Skull: greatest length 13.2; median upper length 11; basi-sinual length 9.8; greatest breadth 10.3; lacrymal breadth 6.7; mastoid breadth 9.1; palato-sinual length 4.5; front of canine to back of m^3 4.9.

Hab. Southern Nyasaland. Type from Chiromo; alt. 200'

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Type. Adult male. B.M. no. 17.2.1.1. Original number 173. Collected 2nd October, 1916, and presented by Rodney C. Wood, Esq.

This species may be distinguished from its only close, though geographically very distant, ally *S. albofuscus* by its smaller size, proportionally even broader skull, and the different shape of the base of its canines. The other members of the genus all have uniformly brown wing-membranes.

I may note that of twelve, skulls of *Scotæcus*, including examples of all the described species, only two have complete zygomata, although all have been prepared by that most skilful skull-cleaner Mr. W. Sherrin. Imperfection or, at least, excessive tenuity of the zygoma would therefore appear to be an additional character of the genus *Scotæcus*. Of forty skulls of *Scoteinus* similarly prepared by Mr. Sherrin, nearly all have perfect, although very slender, zygomata.

Scotonycteris bedfordi Thomas, 1904

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[Proc. zool. Soc. Lond., 1904, vol. i. - 2 August 1904]

Scotonycteris bedfordi, Thomas, Abstr. P.Z.S. 1904, No. 2, p. 14, March 8.

Footnote: The complete account of the new species described in these communications appears here; but as the names and preliminary diagnoses were published in the 'Abstract,' such species are distinguished here by being underlined. - EDITOR

Near *S. zenkeri*, but ears much smaller.

Size rather less than in *S. zenkeri*. Fur close, fine and woolly, the hairs on the back about 6 mm. in length. Fleishy part of forearms, wing-membrane bordering flanks and hind limbs, upper side of legs to ankles, and interfemoral membrane (except terminal part near calcar) well clothed with fur; the centre of the interfemoral quite buried in thick fur. Below, the fur was less extended, the proximal part of the wing-membrane and the centre of the interfemoral.

General colour above mixed whitish and ochraceous buff, the hairs dark brown at base, then silvery white, their tips pale brownish buffy or "clay-colour," the peripheral hairs of membranes wholly of the latter colour. Head rather browner than back. Central white muzzle-patch present, as in the type species; eyes with dark rims, interrupted behind by small white postocular spots, only about 2 mm. in diameter. Owing to the general light colour, these patches are but little prominent. Ears very small and narrow, conspicuously smaller than in *S. zenkeri*. Throat and centre of chest creamy whitish, passing on shoulders, sides of body, and inguinal region into brown.

Dimensions of the type: -

Forearm 48 mm.

Head and body (original measure in the flesh) 71; ear (in flesh) 11; thumb (c. u.) 23; first finger - metacarpal 34, 1st phalanx 24, 2nd phalanx 30; fifth finger 63; tibia 20; calcar 6.

Hab. Fish Town, Fernando Po.

Type. Female. Original number 31. Collected 2 January, 1904, by E. Seimund.

This interesting Bat, which had been named in honour of its joint donor, the President of the Society, was the first representative of the the genus *Scotonycteris* received by the National Museum, and was therefore a most welcome accession. It differed from *S. zenkeri*, of the Cameroons, by its conspicuously smaller ears, which were only 11 mm. in length, while those of *S. zenkeri* were stated by Dr. Matschie to be 17 mm. long, a measurement that quite agreed with their appearance in his plate. *S. bedfordi* also seemed to have smaller postocular spots and a more hairy interfemoral membrane than its mainland ally.

Scotonycteris ophiodon Pohle, 1943

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Vor fast 50 Jahren, in der Sitzung vom 16.10.1894, legte MATSCHIE der Gesellschaft naturforschender Freunde einen Flughund vor, der eine neue Art einer neuen Gattung darstellte: *Scotonycteris zenkeri* Mtsch. Heute kann ich der Gesellschaft die zweite Art dieser damals geschaffenen Gattung vorlegen: *Scotonycteris ophiodon* sp. n.

Typus: Nr. 50051 des Berl. Mus.; F juv.-ad. von Bipindi, Bez. Kribi, Kamerun, Mai 1899, GEORG A. ZENKER S.V.. Die Zähne (auch die Eckzähne) sind voll in Stellung, die Basalnaht ist noch offen. Das Tier liegt in Alkohol; der leider auf der

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rechten Seite unter Verlust der Hirnkapselwand verletzte Schädel ist gereinigt.

Diagnose: Ein *Scotonycteris* von erheblicher Größe. Das vorliegende Weibchen ist ebenso groß wie ein gleich altes von *Epomops franqueti* Tomes. Das Tier ist also weit größer als *Sc. zenkeri* Mtsch. Unterarm über 75, größte Schädelhöhe über 36 mm. Ohren länger (über 20 mm) als bei *zenkeri* und zugespitzt. Gaumenfalten ähnlich denen von *Casinycteris argynnis*. Schädel weniger breit (Gr. Länge: Jochbogenbreite = 1.71 gegen 1.51 bei *zenkeri*). Gaumen sehr stark gewölbt. Backenzähne stärker als die von *zenkeri*, aber auch fast ebenso lang wie breit und geradezu ungeheuer hoch und stark gekrümmt, so daß die Reihe C-M¹ in Profilansicht an ein Schlangengebiss erinnert.

Außere Merkmale: Schnauze verhältnismäßig noch breiter als die von *zenkeri* und daher etwa wie eine Miniaturausgabe einer Bernhardiner-Schnauze wirkend. Ohren relativ länger (20.5 mm) bei relativ gleicher Breite (13 mm), zugespitzt und mit 8 deutlich erkennbaren queren Verstärkungsleisten. Schwanz als Höcker gut fühlbar. Flughäute denen von *zenkeri* entsprechend mit 20 - 22 Vertikalfalten auf dem Mesopatagium. Sie setzen sich an die erste Zehe (und zwar an deren erste Phalange) an. Schwanzflughäute wohlentwickelt, in ihrer ganzen Länge mindestens 7 mm breit. Ueber die Fingerlängen sagt umseitige Tabelle aus, daß sie zwar absolut erheblich größer, relativ aber etwa die gleichen sind wie bei *zenkeri*. Nur der Metacarpus des 2. Fingers ist auch relativ länger. Die übrigen Unterschiede liegen innerhalb von Fehlergrenze und Variationsbreite.

Verteilung der Behaarung wie bei *zenkeri*. Pelz lang, weich und dicht; Länge der Haare auf dem Rücken um 10 mm, der Leithaare um 14 mm; auf dem Bauch sind die Haare gröber und bis 7 mm lang. Die Rückenseiten der proximalen Hälfte des Unterarmes und der beiden proximalen Drittel des Unterschenkels sind (in distaler Richtung abnehmend, an den Gelenken aber dicht) behaart. Farbverteilung auch etwa wie bei *zenkeri*, doch sind die Töne erheblich heller und enthalten mehr Gelb. Da der Typus 44 Jahre in Alkohol gelegen hat, ist aber die exakte Beschreibung der einzelnen Farbtöne sinnlos. Auf dem Nasenrücken ein scharf begrenzter weißer Fleck vom Umriß eines Apfelkernes. Seine Spitze liegt etwa in der Mitte des behaarten Nasenrückens, sein abgerundetes Ende wenige Millimeter hinter den inneren Augen-

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[table removed - eds.]

winkeln. Hinter den Augen und an der Ohrbasis kein weißer Fleck. Dagegen sind Maul und Nasenballen weiß eingefärbt (etwa 3 - 4 mm breit).

Gaumenfalten: Bei *zenkeri*: 5 glattrandige Gaumenfalten zwischen den Zähnen. 1 gleich dahinter; dann 1 mm breite faltenfreie Fläche, und schließlich 6 eng gedrängte, gezähnte, postdentale Falten. Von der ersten, kräftigsten, direkt hinter den Eckzähnen liegenden Falte an nehmen sie an Stärke ab, an Krümmung zu. Die 12. besteht nur aus einzelnen Höckern. Zweite Falte zwischen den P³, die dritte verbindet die P⁴, die 4. die Zwischenräume zwischen den P⁴ und M¹ und die 5. die M¹. Von der vierten an sind sie in der Mitte geteilt, die 6. besteht aus zwei Halbbogen; vor der 5. ein unpaarer mittlerer Höcker, vor der 6. jederseits einer. Von den sechs postdentalen ist die erste auch geteilt, die vier nächsten aber einheitlich.

Bei *ophiodon* haben wir auch die Andeutung der Teilung in eine vordere und eine hintere Faltengruppe, doch ist die Trennung unscharf, da ein faltenfreier Streifen fehlt. Auf 6 ungeteilte inter-

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dentale Falten mit glatten Rändern folgen etwa 12 teils in der Mitte geteilte, teils vollständige, teils unvollständige, postdentale, deren Teile oft durch Schaltstücke verbunden sind. Diese 12 hinteren Falten haben gezähnelte Ränder. Die erste aller Falten steht noch vor den C, dicht hinter den I, die zweite gleich hinter den C. Die dritte verbindet die P³. Sie ist schon stark vorgewölbt, wird aber hierin von der 4. bis 6. erheblich übertroffen. Die vierte beginnt hinter der Mitte der P⁴, ihre weiteste Vorwölbung trifft die Verbindungslinie der Hinterränder der P³. Die 5. beginnt vor der Mitte der M¹ und wölbt sich bis über die Hinterränder der P⁴, die 6. beginnt hinter der Mitte der M¹ und wölbt sich bis zur Mitte der P⁴. Die ersten 5. sind dicke Falten mit hohem Kamm und breiter Basis. Die 6. dagegen hat den Charakter der postdentalen: sie ist flach und hat gezähnelte

Ränder. Die Lage der postdentalen siehe Abbildung 1. Das Gaumensegel trägt keine Falten, sondern nur unregelmäßig angeordnete Höcker (in der Abb. fortgelassen). Insgesamt entspricht diese Anordnung der Gaumenfalten der von *Casinonycteris argynnis* und nicht der von *Scotonycteris zenkeri*.

Schädel: In der Aufsicht und im Profil dem von *Nannonycteris* ähnlicher als dem von *Scotonycteris zenkeri*. Rostrum sehr kurz, relativ kürzer als bei *zenkeri*; seine Länge (Linie Nasalaspitze bis Orbitalrand) beträgt hier $26\frac{1}{2}\%$ der größten Schädelhöhe, bei *zenkeri* 29 % (bei dem vorliegenden gleich alten F Schädel Nr. 10029 von *Epomops fr. franqueti* von Yaunde 31 %). Damit ist die Länge des Rostrums geringer als der Abstand der Außenseiten der P⁴ von einander; bei *zenkeri* ist sie ebenso groß, bei *Epomops*, *Nannonycteris* und *Micropteropus* aber größer. Natürlich ist sie auch etwa gleich der Lacrymalbreite, aber dieser Vergleich trennt die kurzschnauzigen Gattungen der Epomophori nur schlecht von den F von Epomops, da auch bei diesen die Lacrymalbreite etwa gleich der Länge des Rostrums ist. Höhe des Rostrums über dem Diastema C-P³ relativ etwas

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niedriger als bei *zenkeri*, fast genau gleich der Hälfte der Länge des Rostrums. Höhe hinter den M¹ größer als Länge des Rostrums. Der Winkel, den die obere Profillinie des Rostrums mit der Alveolarlinie bildet, ist ein größerer als bei *zenkeri*, somit wird der Abfall des Schädels nach vorn plötzlicher. Postcanine Einschnürung des Rostrums sehr schwach, etwa $\frac{1}{10}$ mm betragend (bei *zenkeri* über $\frac{2}{10}$). Aufsteigende Aeste der Intermaxillaria bis oben hinauf etwa gleich breit, nicht zugespitzt; alveoläre Aeste verhältnismäßig hoch, nicht miteinander verwachsen, Vorderrand im Profil nicht senkrecht, sondern oberes Ende vorragend. Der Gaumen ist von rechts nach links sehr stark gewölbt, in der Hauptsache dadurch, daß der Alveolarfortsatz des Maxillare von vorn nach hinten immer mehr aus der Gaumenfläche heraustritt, Zahnreihen divergieren nach hinten so stark, daß der Abstand der buccalen Flächen der M¹ von einander (12.5 mm) fast doppelt so groß ist wie die kleinste Gaumenbreite (6.7 mm). Der hinter dem Jochbogenansatz liegende Teil des Gaumens ist etwa ebenso lang (6.4 mm) wie durchschnittlich breit (9.5 mm hinter M¹, 6.2 mm an der Choane). Er verjüngt sich also stark nach hinten wie auch bei *zenkeri*. Durch diese Verjüngung steigt der fast geradlinig verlaufende Rand dieses Gaumenteles allmählich fast bis zur Höhe der Gaumenwölbung. Das hat zur Folge, daß in der Profilansicht dieser Rand nicht die geradlinige Fortsetzung der Alveolarlinie ist (wie bei *zenkeri*), sondern mit ihr einen nach oben offenen stumpfen Winkel bildet.

Auch die basicraniale Achse bildet mit der basifacialen einen stumpfen Winkel; der Hirnschädel ist also gegen den Gesichtsschädel abgelenkt (wie bei *Nannonycteris*). Die Verlängerung der Alveolarlinie läuft aber trotzdem unter den Condylen hindurch. Es liegt dies daran, daß durch die Gaumenwölbung der Gaumenrand (die Alveolarlinie) von vorn nach hinten immer stärker hervortritt, also von vorn nach hinten fällt. Im Bereich des postdentalen Gaumens steigt dann die Profillinie wieder an und dann kommt die Abknickung der Schädelbasis, die nun etwa parallel der Alveolarlinie verläuft. Würde man die Mittellinie des Gaumens verlängern, so würde sie das obere Ende der Condylen treffen, bei *zenkeri* dagegen das untere. - Postorbitalfortsatz kurz und dick (Jugendmerkmal?). Temporalleisten weit getrennt, nur bis zur Kranznaht zu verfolgen (Jugendmerkmal?). In der Basalansicht treten die Hinterränder der Proc. zyg. max. nicht

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(wie bei *zenkeri* und *argynnis*) fast senkrecht zur Mittelebene des Schädels aus diesem heraus, sondern bilden damit einen spitzen Winkel (wie bei *Nannonycteris* und *Micropteropus*). Jochbogen dünn, wenig aufwärts gebogen und schwächer nach außen gewölbt als bei *zenkeri*. - Horizontaler Ast des Unterkiefers kräftig, am dicksten unter P₄ und M₁, dahinter dünner werdend; wie bei *zenkeri* und *argynnis* nach außen konvex, nicht fast gerade wie bei *Nannonycteris* und *Micropteropus*. Coronoidfortsatz relativ niedriger als bei *zenkeri* (Jugendmerkmal?), aber ebenso stark, Proc. ang. und Condylus wie bei *zenkeri*.

Gebiß: $\frac{I_1}{I_2} \frac{I_2}{I_2} \frac{C}{C} \frac{P_1}{P_1} \frac{P_3}{P_3} \frac{P_4}{P_4} \frac{M_1}{M_1} \frac{M_2}{M_2} \times 2 = 28$, also wie bei *zenkeri* und fast allen Epomophori. Schneidezähne wie bei *zenkeri*, im Oberkiefer jedoch höher, stärker gekrümmt und weniger rückwärts geneigt; I¹ deutlich höher als I². Im Unterkiefer bei *zenkeri* I₂ höher und kräftiger als I₁; auch hier ist der I₂ viel kräftiger, in der Höhe aber ist ein Unterschied wohl vorhanden aber nicht auffällig. Alle übrigen Zähne erheblich höher (auch relativ), kürzer und stärker rückwärts gekrümmt als bei *zenkeri*. Eckzähne oben und unten etwa ebenso breit wie lang (siehe Maßtabelle), am stärksten gekrümmt von allen Zähnen. Am Oberkiefer-Eckzahn laufen von der Spitze jederseits ein Cingulum in der Krümmung des Zahnes zur Basis; hier biegen beide nach hinten um und vereinigen sich unter Bildung des Basalwulstes. Zwischen jedem Cingulum und dem Zahn befindet sich eine vertikale Furche, die von der Zahnschmelz bis zur Basis zieht. Durch die Cinguli wird die Zahnkrone (außer im basalen Teil) breiter als lang (in halber Höhe: Breite 2.0, Länge 1.5 mm). Das innere Cingulum bildet in etwa $\frac{2}{3}$ der Zahnhöhe durch plötzliche Verbreiterung einen Höckeransatz. Von hier nach unten läuft dann auch auf der Zahnvorderseite eine Furche bis zur Basis. Die übrigen oberen Backenzähne ähneln denen von *argynnis*, doch sind sie höher und stärker rückwärts gekrümmt. Die Innenhöcker sind am P³ wohl niedriger als dort, aber scharf abgesetzt vom Hauptzacken, am P⁴ und M¹ wie bei *argynnis*. - Auch die Unterkiefer-Caninen tragen die beiden vertikalen Furchen auf der caudalen Fläche und damit den von der Spitze außen abwärts laufenden, an der Basis nach hinten umbiegenden, als Basalwulst über die rückseitige Basis laufenden, nach oben umbiegenden und wieder zur Spitze laufenden Wulst, der

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wieder auf der Innenseite in etwa $\frac{3}{4}$ der Zahnhöhe infolge plötzlicher Verbreiterung eine Nebenzacke andeutet. Von hier aus läuft auch auf der Vorderseite des Zahnes eine kurze Vertikalfurche. Der P₁ ist deutlich stärker als ein I₂. Der P₃ ist fast ebenso hoch wie der Eckzahn. Sein innerer Höcker ist scharf abgesetzt, rostral durch eine Schmelzkerbe, caudal durch eine vertikale tiefe Grube. Auch bei ihm ist durch eine vertikale Furche auf der caudalen Fläche ein von der Spitze zur Basis laufendes Cingulum abgesetzt, das dann umbiegend einen Basalwulst bildet, der dann an der Nebenspitze wieder emporsteigt. P₄ und M₁ entsprechen ungefähr den gleichen Zähnen von *argynnis*, doch sind sie und die Innenspitze etwas höher.

Verbreitung: Bisher nur bekannt von Bipindihof, wahrscheinlich also ähnlich verbreitet wie *zenkeri* und viele andere Säuger (z. B. *Colobus satanas* Waterhouse), die von Fernando Po, dem südlichen Kamerun und dem nordwestlichen Gabun bekannt geworden sind.

Maße: Aeußere Maße: Kopfrumpflänge 105 mm, Schwanzlänge unter 1 mm, Hinterfußlänge 14 mm, Ohrlänge 20.5 mm.

Schädelmaße: Größte Länge 36 + mm ; Palation bis Foramina incisiva 17.6; Palation bis Basion 12.2; Länge des postdentalen Gaumens 6.4; Länge Orbiturand bis Nasaliaspitze 9.2; Breite der Hirnkapsel über dem Jochbogenansatz 16; Jochbogenbreite 21; Breite über den Kronen der M¹, außen 12.6; Lacrymalbreite 9.4; Breite über den Caninen, außen 6.7; Intertemporalbreite 8.9; Spitzenabstand 11.2; Interorbitalbreite 6.4; Choanenbreite 5.0; Mittlere Breite des postdentalen Gaumens 7.5; Innenabstand der M¹ von einander 7.9; desgl. der P⁴ 6.2; desgl. der Basen der C¹ 2.5; Durchmesser der Orbita 9; Unterkieferlänge (vom Condylus aus) 16.5; Höhe des Coronoidfortsatzes 11.2 mm.

[table removed - eds.]

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Verwandschaft: *ophiodon* zeigt eine eigenartige Mischung von Charakteren der Arten *Scotonycteris zenkeri* und *Casinycteris argynnis* mit Merkmalen, die ihm eigentümlich sind. Zur ersten Gruppe nenne ich das Fehlen des weißen Fleckes vor den Ohren, die Form des Intermaxillaria. Zur zweiten Gruppe gehören die längeren Ohren, die Anordnung der Gaumenfalten, die Höckeranordnung an den Backenzähnen. *ophiodon* eigentümlich sind die erheblichere Größe, die weiße Einfassung von Maul und Nasenballen, der Umriß des Schädels in der Aufsicht, die starke Gaumenwölbung, die Abnickung des Hirnschädels u. a. Den größten Teil der Merkmale aber hat er mit den beiden Arten gemeinsam. Seine systematische Stellung ist also nicht zweifelhaft, er gehört zwischen die beiden.

In welche Gattung muß er aber gestellt werden? Mancher Säugetierspezialist würde die Frage einfach beantworten: In eine neue. Nach dieser Methode erhalten wir aber einen Wust neuer Gattungsnamen, die nichts weiter sagen, als daß die Arten eben verschieden sind. Wir haben in der Säugetierkunde schon viel zu viele Gattungsnamen, denn ganze große Familien enthalten nur monotypische Gattungen. Wir müssen uns daran gewöhnen, Gattungen nur dort neu zu schaffen, wo wirklich prinzipielle Unterschiede vorliegen. Das ist aber in unserem Fall nicht zu behaupten. *ophiodon* gehört sicher in eine der beiden schon beschriebenen. Wir müssen also auf unsere Frage zurückkommen. Umm sie entscheiden zu können, wollen wir zunächst ANDERSON's Ausführungen über *Casinycteris* (pg. 671/2) zitieren, wobei wir seine Einschaltungen über Aehnlichkeiten mit *Pteropus*-Arten als unwesentlich für unsere Frage fortlassen: "If the single species of this genus were known only from its external appearance, it would unhesitatingly be considered a *Scotonycteris*, differing from *S. zenkeri* in nothing but its larger size, relatively larger ears, and small white ear-patches; so extraordinary is its resemblance in all external features to *S. zenkeri*, even in the general colour of the fur and the very peculiar white markings of the head. In the characters of the incisors and canines, the peculiarly short-and-broad, subcircular outline of the premolars and molars, and the heightening of their outer cusps the dentition of this genus is very similar to that of *Scotonycteris*, and the details in which it differs (chiefly the stronger development of the inner cusp of p³ and p₃, the divergence of the outer and inner cusps of the large premolars and molars, and the less

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pronounced reduction of the posterior molars) would in themselves be far from justifying its generic separation from *Scotonycteris*. With this close external and dental resemblance to *Scotonycteris*, the present genus combines, however, a modification of the bony palate absolutely unique in Megachiroptera and more closely approaching to the type of palate predominant in Microchiroptera." Die Gattung *Casinycteris* ist also einzig und allein begründet auf die tiefe Spalte, die sich bei *argynnis* von der Choane bis zu den letzten Oberkieferzähnen zieht oder richtiger, die in diesem Bereich an die Stelle des harten Gaumens getreten ist. Dieses Merkmal scheint unzweifelhaft um so mehr Gewicht zu haben, als es sonst in der ganzen Unterordnung der Megachiroptera nicht wieder vorkommt. Man müßte also ANDERSON und THOMAS ohne weiteres Recht geben in der Aufrechterhaltung dieser Gattung *Casinycteris*, wenn nicht von beiden eine Kleinigkeit übersehen worden wäre, die diesem Merkmal ein ganz anderes Gesicht gibt. Es ist dies die Tatsache, daß dieses Merkmal bei vielen Säugern als Mißbildung beobachtet worden ist und wahrscheinlich bei allen vorkommen kann als sogenannter Wolfsrachen oder Gaumenspalte. Wir können von *argynnis* nur einen einzigen Schädel, eben den des Typus der Art; wir können also nicht einmal behaupten, daß die Gaumenbildung bei allen Tieren der Art die vom Typus beschriebene ist. Es ist schon möglich, daß wir von dieser seltenen Art ausgerechnet nur ein teratologisches Stück in die Finger bekommen haben. Aber selbst wenn sich das Vorhandensein der Gaumenspalte bei allen Tieren der Art erweisen sollte, dann ist dieses Merkmal noch nicht ausreichend, um damit die Unterbringung der Art in einer anderen Gattung zu begründen. Merkmale, die innerhalb der ganzen Säugetierreihe als Anomalie erblich auftreten können, müssen durch eine verhältnismäßig einfache Genänderung bedingt sein und sind deshalb zu klein, um als Gattungsmerkmale zu gelten. Wenn nicht daneben noch andere Unterschiede vorhanden wären, so würden sie ja nicht einmal zur Begründung einer Art ausreichen. Die Gattung *Casinycteris* ist also zu streichen; *argynnis* ist als Art zur Gattung *Scotonycteris* zu stellen und in diese selbe Gattung kommt als dritte Art unser oben beschriebener *ophiodon*.

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Bemerkungen: Es ist nicht mehr als Pflicht darauf hinzuweisen, daß wir auch die Kenntnis dieser Art dem ausgezeichneten Sammler GEORG A. ZENKER verdanken, von dem ja das Berliner Museum so viele andere neu zu beschreibende Säugetierarten und Unterarten erhielt, von denen hier nur *Scotonycteris zenkeri* Mtsch., *Idiurus zenkeri* Mtsch., *Zenkerella insignis* Mtsch., *Cercocebus albigena zenkeri* Schwarz genannt seien. G. ZENKER ist am 12.2.1922 nach einem an Abenteuer reichen Leben in seiner Wahlheimat Kamerun (Bipindihof) im Alter von 67 Jahren verstorben. Er ist dem Berliner Museum treu geblieben wie wenige Sammler und hat sein Material hierher gegeben, obwohl wir ihm nur wenig dafür bezahlen konnten, und obwohl er wußte, daß er von London ein Mehrfaches der von uns bezahlten Preise hätte erhalten können.

Material: Nur der Typus.

Scotonycteris ophiodon cansdalei Hayman, 1946

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Type. - F skin and skull, B.M. No. 46.229, collectors' number 420. Collected at Oda, Oda District, Central Province, Gold Coast, December 24, 1945, by G. S. Cansdale. A very young specimen, No. 46.230, preserved in alcohol, was taken clinging to the type., "Suspended in tree in forest; solitary." (Note by collector.)

Diagnosis. - Closely similar to *Scotonycteris ophiodon* Pohle (1943) in size and most external, cranial and dental characters, but differing externally in having a conspicuous white patch at the posterior angle of each eye, an inconspicuous white spot at the anterior base of the ears (both spots lacking in *ophiodon*), and in the white border of the upper lips only reaching two-thirds of the way forward towards the nostrils (in *ophiodon* the white lip border is said to include the nostrils); and cranially in the long and thin postorbital processes (short and thick in the unique type of *ophiodon*), in the horizontal basicranial axis (deflected in *ophiodon*), and in having a distinct, though low, sagittal crest; the latter may well be an age character. (For measurements see Table I.)

Colour. - As the type of *Scotonycteris ophiodon* had been in spirit for forty-four years when Pohle described it, he was not in a position to give much account of its colour, beyond stating that it appeared to have the same general pattern as in *S. zenkeri* Matschie (1894), but somewhat lighter and yellower, a difference which might well be due, in my opinion, to the bleaching effect of long immersion in alcohol. The following general account of the colour

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of the Gold Coast form, *cansdalei*, based partly on Cansdale's notes from the fresh specimen, partly on the dry skin, shows that it has a striking similarity to *zenkeri* and also to *Casinycteris argynnis* Thomas (1910).

The dorsal hair above has three colour zones, dark brown basally, dirty white medially, fawn distally, the general effect varying according to the length of hair and length of the whitish zone. On the neck and shoulders the hairs are longer than on the hinder portion of the back and have a much wider whitish zone, which shows through the fawn tips. Further back the whitish zone is very narrow and the hair is shorter, so that the general effect is much darker, and on the rump the pale zone is absent and the hairs are almost uniformly brown. Below, the hair on throat and lower neck is very sparse, is almost white, and the pale flesh skin is exposed. From the lower neck a pale abdominal patch, a dirty whitish heavily washed with dull buff, continues for some way, and is set off by the contrasting dark brown of the sides of the belly, the flanks and the anal region. On the head the white patch on muzzle, the white border of the lips (not reaching the nostrils), and the white patch behind the eyes, are conspicuous, but the white spot at the anterior base of the ears mentioned by Cansdale *in litt.* is so small and indistinct that in the dry skin it could easily be overlooked, and it seems likely that the type of *ophiodon*, after forty-four years in alcohol, would probably be too bleached to show such an inconspicuous feature. According to the collector, the wing membranes are yellowish-brown, and the bare skin of mouth, eyelids and ears dark yellow (in the dry skin the three latter are dark brown). The wing membranes are prominently reticulated, and the joints of phalanges and metacarpals are pale yellow, much as in *Casinycteris*.

Remarks. - In 1943 Pohle described *S. ophiodon* from Bipindi, Cameroons, as the second species of the genus, previously known only by the genotype, *S. zenkeri*, whose range included the Cameroons and Fernando Po. *S. ophiodon* was characterised by its much greater size (forearm 75, in *zenkeri* 48 - 54), and by striking dental peculiarities, of which the most important are the secondary cusps on the inner edges of upper and lower canines, and the heightening of the canines and cheek

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[table removed - eds.]

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teeth, the latter being provided with prominent inner cusps. *S. ophiodon* also lacks the white spot behind the eye found in *zenkeri*, although agreeing with the latter in most other respects in facial pattern and body colour.

Pohle considered that some of the characters of *ophiodon*, particularly those of the cheek teeth, showed affinity to *Casinycteris argynnis*, whose close external similarity to *Scotonycteris* has been commented on by Andersen (1912); in fact Pohle expressed the view that the extraordinary shortening of the palate in *Casinycteris*, the chief diagnostic feature of the genus, is merely an abnormality, akin to the "cleft palate" sometimes found in mammals, and as such to be discounted. Moreover, he also took the view that even if it were not an abnormality in itself, such a feature, being found nowhere else in the Megachiroptera, should not on the evidence of one skull be regarded as a character of sufficient stability to justify the founding of a genus. On these grounds he proposed to relegate *Casinycteris* to the synonymy of *Scotonycteris*, while retaining the species *argynnis*. Unfortunately for this theory, however, it must be pointed out that Pohle's declaration that only the type skull is known of *argynnis* indicates that he overlooked the fact that at the time at least two other skulls of this rare bat had been reported on. One was recorded by J. A. Allen (1917) from the Eastern Congo (with comments on its peculiar skull and palate by Lang and Chapin (1917a); another was recorded from the Uele District of the Eastern Congo by Schwarz (1920), who gave measurements showing a similar extreme shortening of the palate as in the type. In view of these facts it seems clear that Pohle's dismissal of the chief generic character of *Casinycteris* as an abnormality must be disregarded, and that the genus must stand.

Although the heightening of the inner cusps of the cheek teeth in *ophiodon* and *cansdalei* is a feature more nearly approaching the dentition of *Casinycteris argynnis* rather than *Scotonycteris zenkeri*, the normal palate makes inclusion in the former genus out of the question; the prominent secondary cusps on all canines form the most marked feature in which both *ophiodon* and *cansdalei* differ from both *S. zenkeri* and *Casinycteris*, but I am not prepared to add a generic or subgeneric name to the

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literature on that account. I have used a subspecific name for this Gold Coast *Scotonycteris* chiefly for convenience in expressing its relationship to *ophiodon*, but am quite prepared to find eventually that *cansdalei* will be raised to specific rank; the cranial differences, particularly the absence of a basicranial deflection, combined with the external difference in the head markings, may later be regarded as of specific value.

The discovery in the Gold Coast of this most interesting fruit bat by Mr. Cansdale, whose name I am glad to link with it, shows that special search for Chiroptera in the forest zones of Africa is likely to produce yet more surprises, both in new forms and in furthering our knowledge of the distribution of species already known elsewhere.

Scotonycteris zenkeri occidentalis Hayman, 1947

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Type. - British Museum number 46.898, adult female skin and skull (originally an alcoholic specimen). Collected at Oda, Oda Province, Gold Coast, 1946, by G. S. Cansdale. "Found sleeping in plantain leaves. G. S. Cansdale."

Diagnosis. - In size, form, cranial and dental characters as well as face markings, similar to *Scotonycteris zenkeri* Matschie (1894), whose distribution is limited, so far as is known, to the Cameroons and Fernando Po, but general colour considerably darker, the overall hue above being sepia rather than russet or rust-brown.

As in *S. zenkeri*, the dorsal hairs have three distinct colour-zones, a very dark brown basal third, a pale grey median zone, and a terminal zone of sepia instead of the russet or rust-brown of *zenkeri*, giving the animal its distinctive dark appearance. On the head the white markings are well denned, the white patch on the rostrum extending back to the level of the anterior angle of the eye, and the white spot behind each eye, being similar in size and position to those found in *zenkeri*. Below, the rather sparse hairs on throat and sides of the neck are whitish, and a narrow line of white hairs runs forward from the angle of the mouth for a short way along the upper lip. The centre of the chest and abdomen is greyish white, rather sharply differentiated from the brown flanks. As compared directly with two specimens of *zenkeri* (including the type of *S. bedfordi* Thomas (1904) = *zenkeri*) it is noticeable that the general darkening extends to the wing-membranes, which are dark chocolate instead of light brown. The collector's notes on colour of the fresh specimen tally with the description given above, so that there is no question of the colour having changed as a result of a brief immersion in alcohol.

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Measurements. - Head and body 70 mm., hind foot 13, ear 13, forearm 52, pollex 19, fifth metacarpal 32. Skull: total length 24.6, palatal length 13, post-dental palate 4, rostrum (front of orbit to tip of nasals) 6.7, breadth of brain-case at zygomatic roots 11.7, zygomatic breadth 15.9, breadth across crowns m^1-m^1 7.6, breadth across crowns of upper canines 5.6, interorbital breadth 5, upper toothrow $c-m^1$ 7.9.

Palate ridges. - Andersen's description (1912) of the palate ridges in *S. zenkeri*, as shown also in the figure given by Matschie (1899), applies also to those of *occidentalis*, in that there are six main ridges separated by a narrow space from five weaker and more crowded posterior ridges. There is also a weak ridge between the incisors and canines, not mentioned by Andersen, but indicated in Matschie's figure.

Remarks. - The discovery in the Gold Coast of a local race of *S. zenkeri*, itself a rarity in collections and restricted to the Cameroons and Fernando Po, comes soon after Mr. Cansdale's capture of a Gold Coast race of *S. ophiodon* Pohle (1943), which, like *zenkeri*, was previously known only from the Cameroons. The collection in 1946 of *S. ophiodon cansdalei* Hayman (1946) and of *S. zenkeri occidentalis* demonstrates that our knowledge of the distribution of African bats is still far from complete.

Scotophilus Leach, 1821

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Gen. 3. *Scotophilus*

Dentes incisores $^4I_6$: superiores inæquales: laterales brevioribus bifidis.
inferiores subtrifidi.

--- *molaes* $^8I_8$, processibus acuminatis armati.

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Gen. 3. *Scotophilus*

Dentes incisores $^4I_6$: superiores inæquales, acuminati: intermedii longiores, simplices; laterales æqualiter bifidi.
inferiores obsoletè trifidi.

--- *lanarii* $^2I_2$: superiores longiores, ad latus posticè processu armati.

interiores ad latus anticè processu armati.

--- *molares* $\frac{8}{8}$, processibus acuminatis armati.

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Pedes antici indice 1-articulato: digito medio 3-articulato; digitis quatro et quinto 3-articulatis.

--- *postici* digitis mediocribus, subæqualibus: *Ungues* compressi, arcuati: *Membrana* ad caudæ apicem producta, hinc posticè acuminata: *Suspensorium* flexuosum, in membrana inclusum.

Aures distantes: *Auriculæ* parvæ.

Cauda ad membranæ apicem producta: articulis quinque osseis exsertis.

Scotophilus albofuscus Thomas, 1890

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Size small; body: stout and heavy compared to the size of the wings. Head very broad and flat; muzzle considerably swollen, the glandular prominences much developed. Ears rather thick and fleshy, especially along their outer edge; laid forward they reach to just about halfway between the eye and the tip of the nose; their internal basal lobe scarcely developed at all; inner margin nearly straight, tip narrowly rounded off; outer margin evenly convex in its upper third, slightly concave in its middle third and terminating in a long fleshy lobule running forwards towards the angle of the mouth. Tragus unusually short (see measurements below), its inner and outer edges straight and nearly parallel, tip broadly rounded off, external basal projection fairly well developed. Wings to the metatarsus. Post calcaneal lobe broad and prominent. Extreme tip of tail projecting.

Fur short and close, scarcely encroaching at all upon the membranes or limbs. Colour of body above and below dark umber brown; naked skin, of all those parts which are visible when the wings are folded dark brown or black, i.e. muzzle, ears, lines of arms, forearms and fingers, wing membranes internal to a line drawn from the elbow to the knee, and whole of hind-limb, interfemoral membrane and tail; on the other hand

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some of the more aberrant species of *Vesperugo*, and especially from those placed by M.^r Dobson in his subgenus *Scotozous*, to which perhaps it may be thought the new species should have been referred. *Scotozous* was founded on the Indian V. (*S.*) *dormeri*, Dobs., a species with all the *facies* of a *Vesperugo*, with two upper premolars, and with a long and pointed anterior lower premolar, these characters being strictly those of *Vesperugo*, from which in fact it only differs by having but one upper incisor on each side, as in *Scotophilus*. On the other hand the second species referred to the subgenus by M.^r Dobson, "*Nycticejus schlieffeni*" Peters, ought, it appears to me, to be rather called a *Scotophilus*. Before entering into this point however, some reference to its specific history is unfortunately necessary, as there has been considerable confusion in regard to it.

Its first description was given by D.^r Peters in 1859 unless, *Vespertilio marginatus*, Cretzschm. be the same species, but the type of, the latter form was distinctly stated by Blasius to be referable to *Vesperugo kuhli*, Natt., and if he was right the species will stand under Peters's name. In 1878 M.^r Dobson referred the bat, with one Museum specimen, to his subgenus *Scotozous*, of the genus *Vesperugo*, but at the same time, misled by an erroneous locality, he placed a second specimen of it under *Scotophilus pallidus*, a species described by him two years previously in India. That this specimen was wrongly named has since been proved by the receipt of genuine examples of *S. pallidus* from India, but in the mean time a comparison with it had caused the Marquis G. Doria and myself to refer to *S. pallidus* a bat from Massowa collected by Signor Gustavo Frasca; this extension of the range of the latter, commented upon by us at the time, must now be cancelled, the bat being, like the

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[there seems to be something wrong with the sequence here. Some text seems to be missing. - eds.]

the wing-membrane external to the same line is pure white "above and below, contrasting very markedly with the dark skin of the limbs and digits.

Number of teeth strictly as in *Scotophilus*. Upper incisors long, slender, unicuspid, close to the canines; without posterior ledges, but each with a very minute postero-external basal cusp. Lower incisors tricuspid, overlapping, placed at right angles to the direction of the jaws. Anterior lower premolar not as in *Scotophilus*, crushed in between the posterior one and the canine, but more as in *Vesperugo*, long, as long as the posterior one, and quite as large in section basally, its long simple cusp directed somewhat outwards, out of the general line of the teeth.

Measurements of the type, an adult male, preserved in spirit: Head and body 50 mm.; tail 31; head, length 17, breadth of muzzle across eyes 10.8; ear, length above crown 8.8, from base of outer margin 14.5; tragus, length of inner margin 2.0, of outer margin 3.7; forearm 31 (= 1.22 in.); lower leg 12; hindfoot without claws 7.1; calcaneum 13.

Hab. Bathurst, River Gambia. Collected by D.^r Percy Rendall.

This species may be readily distinguished from all its allies by its white wings, which contrast very markedly with the dark coloured body and limbs, all the other known members of the genus having the body and wings more or less uniformly

coloured. In this respect the bats of the Gambia present rather a remarkable instance of geographical isomorphism, a considerable proportion of the smaller forms, belonging to several different genera, being dark with white wings, a style of coloration comparatively rare elsewhere.

D.^f Rendall is to be congratulated on his discovery of this striking species, which forms a worthy companion to the *Vesperugo* (*Vesperus*) *rendalli* described by me last year.

The characters of *Scotophilus albofuscus* add still further to the difficulty of properly distinguishing the genus *Scotophilus* from p. 87

specimen with which it was compared, an example of *S. schlieffeni*. Finally in 1887 D.^f H. Noack, when determining bat from Marungu, Central Africa, thinking (and as I believe rightly) that he had before him a true *Scotophilus*, described it as new under the name of *S. minimus*, but his detailed description leaves no doubt as to its identity, with *S. schlieffeni*, with which he naturally did not compare it, owing to the latter form being referred to *Vesperugo* in D.^f Dobson's classical work.

Now this animal, round whose history so much confusion has gathered, has distinctly the general facies of *Scotophilus*; it has ordinarily no minute anterior upper premolar, and the corresponding tooth in the lower jaw is as small and almost as much crushed in between its neighbours as is the case in several of what are admittedly *Scotophili*. It is true that it has occasionally an anterior upper premolar, but this only occurs in one of the many specimens known to science, and may be simply an individual variation, perhaps due to atavism. In fact M.^f Dobson's own reference of a specimen of it to "*Scotophilus*" *pallidus*, and D.^f Noack's description of it as "*Scotophilus*" *minimus* as already referred to, both strongly support this view.

But even if *S. schlieffeni* is a *Scotophilus*, it by no means follows that *S. albofuscus* is, for the latter is still further away from the typical members of the genus, owing to its long uncrowded anterior lower premolars, which agree more with those of the true *Vesperugo* or of *Scotozous dormeri*. In fact it tands in regard to the latter form just where *Vesperus* does to *Vesperugo*, *Glauconycteris* to *Chalinolobus*, and *Dasypterus* to *Atalapha*, differing, so far as dentition is concerned, only by the absence of the minute upper premolars. But whatever may be

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the ultimate arrangement of these various annectent species, it seems better for the present to call the new form a *Scotophilus* on account of its dental formula, irrespective of the shape of the teeth, rather than to consider it a member of the large genus *Vesperugo*, in which the number of its teeth would necessitate the foundation of a new subgenus for its reception.

There is another bat whose claim to generic distinction is considerably weakened, if not, as I believe, altogether removed, by the discovery of *Scotophilus albofuscus*, namely the American species called by D.^f H. Allen and others "*Nycticejus crepuscularis*". This bat's sole, or at least chief, generic characteristic lies in the uncrushed state of its anterior lower premolars, a peculiarity which as already stated, *S. albofuscus* shares with it, while geographical considerations would prevent the two species from being regarded as specially closely related. It may also be noted that since Rafinesque's *Nycticejus humeralis* is, as D.^f Allen has suggested, no doubt identical with Le Conte's "*N. crepuscularis*" and is earlier in date, the species ought to bear the former instead of the latter specific name. Its proper designation would therefore appear to be *Scotophilus humeralis*, Raf.

Scotophilus alvenslebeni Dalquest, 1965

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Type. - Adult male, skin and skull, no. 18767 Walter W. Dalquest, Midwestern University Collection, taken on the south bank of the Save River, 212 km SSW Beira, Mozambique.

Diagnosis. - A very large, dark *Scotophilus*, exceeded in size only by *Scotophilus gigas* among bats of this genus known from the African mainland. Sagittal crest enormously developed, extending posteriorly over the occiput in an exaggerated crescent. Forearm, 77 mm; condylobasal length of skull, 26 mm.

Comparison. - This species needs comparison only with *Scotophilus gigas* Dobson, for no other *Scotophilus* from the African mainland approaches it in size. I have not seen specimens of *S. gigas*, but the forearm of that species is said to be more than 85 mm in length (Ellerman, Morrison-Scott and Hayman, p. 85), which is considerably greater than that of *alvenslebeni*. The fur of *alvenslebeni* is dark sooty or dusky brown. The ears and membranes are black. *S. gigas* is yellow in color. *S. alvenslebeni* has not been compared with *S. robustus* of Madagascar, but it is highly improbable that the insular form would occur on the African mainland.

Measurements. - External measurements of the type and only known specimen are: total length, 175; length of tail, 77; length of hind foot, 14; height of ear from notch, 20; height of tragus, 9. Cranial measurements are: length of skull from canines, 31.0; condylobasal length, 26.2; zygomatic breadth, 20.0; braincase breadth, 14.8; width across upper M3's, 12.4; width across upper canines, 9.6; upper canine-M3 length, 9.9; mandible length plus incisor, 21.4; lower canine-M3 length, 12.1; forearm, 77.

Remarks. - The type was shot as it flew over the camp clearing at Zinave in the late evening. The species is named for Baron Werner von Alvensleben, in partial recognition of his labors in behalf of our work in Mozambique.

It seems unlikely that this form will be found to intergrade with *S. gigas*. *S. gigas* is known from Nigeria, Nyasaland and Southern Rhodesia. The specimen from Southern Rhodesia is in the British Museum, and has a forearm in excess of 85mm.

The forearm of the "type", from Nigeria, has a forearm 3.4 inches (85 mm). The Southern Rhodesian bat thus does not approach *S. alvenslebeni* in size.

Scotophilus angusticeps Shortridge and Carter, 1938

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Scotophilus angusticeps sp. n.

Two specimens ♀♀ from near Citrusdal.

General colour above drabby umber-brown (rather as in *Eptesicus capensis*), without gloss or sheen; below light drabby brown, centre of abdominal and anal regions strongly washed with rufous, throat tinged with sienna-brown; base of fur, above and below, dark seal-brown; ears and flying membranes dusky black.

Type (in the Kaffrarian Museum). - Adult female, original number 1937, collected 28th December 1937 at Hex River Estate, 10 miles north of Citrusdal, N.W. Cape Province.

Co-type. - Adult female, original number 3308, collected 3rd January 1938.

Dimensions of Type and Co-type. - H. and b., 77 (74); tl., 58 (57); hf., 12 (11.5); ear, 18 (17.5); forearm, 53 mm.

Skulls. - Total length, 20; basal length, 17(17.2); zygomatic width, 13.5; width of brain case, 10 (9.5); interorbital constriction, 4.5; length of palate, 9; upper dental series (from front of canine), 7 (7.5); lower dental series (from front of canine), 7.5 (8); mandible, 16 (15.5); height of brain case, 7.5 mm.

Skull flatter, less massive and with a markedly narrower zygomatic width than in *S. nigrita dingaani* or *S. n. herero*; cranially *S. n. herero* is in some respects intermediate between *S. n. dingaani* and *S. angusticeps*, although much more closely resembling the former.

Scotophilus damarensis Thomas, 1906

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Quite like *S. nigrita herero*, but markedly smaller. Colour above similarly "wood-brown." Below it is a pale whitish brown, without the buffy tone generally present in *herero*.

Dimensions of the type: -

Forearm 48 mm. (other specimens up to 51).

Skull: greatest length to occipital crest 17.3; zygomatic breadth 12.6; interorbital breadth 7.2; palate length in middle line 6.7; front of upper canine to back of m^3 6.5; greatest breadth of palate between outer corners of m^2 8.1; front of lower canine to back of m_3 7.

Hab. Elephant's Vley, Northern Damaraland.

Type. Male. Original number 276. Collected 10th August, 1859, by C. J. Andersson and received in the Tomes Collection. Four specimens.

Mixed up with the series of *S. nigrita herero* there occur four specimens of this smaller bat, otherwise indistinguishable from its larger ally, to which it would bear the same relationship that in Nigeria *S. nigritellus*, de Wint., does to the true *S. nigrita*. But as it is not quite so small as *nigritellus*, and specimens from the intermediate area are not yet known, I give it a binomial name, though I think it will very probably prove to be a subspecies of that form, from which it may be readily distinguished by its pale brown colour.

Scotophilus darwini Tomes, 1859

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The next species which I have to describe has been presented to me by Mr. Darwin, with the information that it had been received from the Canary Isles.

In a collection of Bats from Madeira, given to me also by Mr. Darwin, I could only enumerate two species, both European, viz. *S. leisleri* and *S. marginatus*; and I was somewhat surprised to find in the present species one which I had not before met with. None of the descriptions of African species in the works of Temminck, Wagner, Peters, Smith, and others, apply to this species; and I therefore regard it as new, and describe it as follows: -

It is one of the same group as the species just described, and as the *S. kuhlii* and *S. pipistrellus* of Europe. It is characterized by a somewhat more robust make than these species, and has rather broader ears and tragi.

The head is rather broad and flat, the crown being but little raised above the facial line; the glands of the lips are considerably developed, and bulge sufficiently to occasion the nostrils to open nearly straight forward, although the interruption in the outer margins of the latter sufficiently indicates that with a more pointed

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muzzle they would open sublaterally; were the specimen taken from the spirit in which it is preserved and dried, it is probable that this would be the case. In the middle of the face is a kind of hollow, occasioned by the labial glands on each side being developed in an upward direction, thus leaving a depression between them*. Between the nostrils is a space of moderate extent, and but very faintly emarginate. The ears are rather large, triangularly oval, as broad at the base as they are long, and have their tips brought to a rounded point; about the middle of their outer margin they have a distinct but shallow notch, below which is a lobular portion, as in many other species of this group, but differing from all others which I have seen in having a small but very well-defined notch about its middle. These organs altogether are more like those of *S. kuhlii* than of any other species, but are larger, besides having the double emargination just noticed**. The tragus is rather short and broad, curved inwards, and with the end very much rounded; on its outer margin, near the base, is a projecting angular point, without any accompanying notch.

The wing-membranes extend to the base of the toes, and the latter are half the length of the foot. The thumb is moderate, with the basal phalange much the shortest. The terminal vertebra of the tail is free.

The fur of the head extends forwards to between the eyes, and thence in a narrow strip towards the nose. Over each eye is a wart bearing a bundle of stiff hairs; and a similar tuft springs from the top of the labial glands; the upper lips are also slightly fringed with similar hairs, most conspicuous about the corners of the mouth. The remainder of the face, the ears, and the tragus are naked. The fur of the back spreads on the upper surface of the interfemoral membrane, sparingly, for nearly half its length, as in *S. kuhlii*, and similarly to a small extent on the membranes near the sides of the body. Beneath, the membrane immediately around the pubes is dusted with very short hairs, more abundant on the vertebræ of the tail than elsewhere. On the membrane contiguous to the sides of the body, fur of a much longer kind extends, to a much greater degree than in *S. kuhlii*.

On both surfaces of the body the fur is bicoloured: above, very dark brown at the base, tipped with lighter and more rufous brown, that on the membranes wholly of the latter colour; beneath, it is dark at the base, tipped with paler brown, with less of the rufous tinge than that of the upper parts. On the under surface of the membranes the fur is uniformly of the same colour as the tips of the hairs on the belly, but on the pubes it is paler. Membranes dark brown.

Such appear to be the colours of the fur, so far as can be gathered from the examination of a specimen in spirit; but it is necessary to

* In the *Romicia calcarata* of Dr. Gray the lip-glands are so much developed as to leave a deep pit between them. It belongs to the present group.

** I am here comparing a specimen in spirit with others in skin, - a plan not always attended with perfectly satisfactory results.

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consult others in skin before this point can be determined with accuracy.

Although in its external appearance *S. darwini* bears considerable resemblance to *S. kuhlii*, it differs, besides having a somewhat differently shaped ear and broader tragus, in the form and arrangement of the fore teeth. In *S. kuhlii* the upper incisors are rather long and slender; the inner ones are deeply forked at their apices, and longer than the outer ones, which are slender and pointed, somewhat like, small canines; and there is a visible interval between the points of the inner and outer ones. In *S. darwini*, on the contrary, they are short and obtuse, of nearly equal length, the inner ones faintly cleft at their points, and the outer ones so closely packed to them as to leave no space even between their points. Again in *S. kuhlii* there is a space between the canine and the "carnassier" or sectorial tooth; in which is placed a small and conical premolar, within the line of the teeth, but distinctly visible from the outside; whereas in *S. darwini* the canine and the "carnassier" are contiguous, and there is a very small anomalous premolar placed in the inner angle formed at their bases, visible only from inside.

These differences in the dentition are alone sufficient to distinguish the species from *S. kuhlii*. From *S. marginatus*, *S. ursula*, and *S. nathusii* it may be also recognized by the form of the upper incisors; and these are the only European species with which it could be confounded.

" "

Length of the head and body: 2 1

----- of the tail: 1 5

----- of the head: 0 8

----- of the ears: 0 4 1/2

----- of the tragus: 0 2

Breadth of the tragus: 0 1 1/2

Length of the fore-arm: 1 5

----- of the longest finger: 2 6

----- of the fourth finger: 1 8

----- of the thumb: 0 2

----- of the tibia: 0 6 1/4

----- of the foot and claws: 0 3

----- of the *os calcis*: 0 5

Expanse of wings: 9 9

Hab. Palma, Canary Isles.

Obs. The Madeiran species being European ones, and one of them African also (i. e. *S. marginatus*), renders it not unlikely that the species inhabiting the Canaries may also occur in Africa, and perhaps in Europe. With a view to the chance of this, I have compared this species with what now remains of the types of *Vespertilio aristippe*, *V. leucippe*, *A. alcythoe*, *V. vispistrellus*, and *V. savii*, but find nothing which leads me to regard it as referable to any of them; and I have therefore given such a detailed description as will be amply sufficient to distinguish it from all recorded European species.

Scotophilus gigas Dobson, 1875

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Ear-conch and tragus like those of *S. borbonicus*, Geoffr.; but the internal basal lobe of the ear is more rounded and its inferior horizontal margin is straight, not in the least degree concave; the upper third of the outer margin of the conch is slightly but distinctly concave; and the tragus has the narrow ridge, proceeding from the base of its inner margin across its front margin, as well developed as in *S. Temminckii*.

Wings to the metatarsus near the base of the toes; last two caudal vertebræ and half the third last vertebra free.

Fur above deep chestnut, beneath yellowish white. The fur on the upper surface is short and does not extend anywhere upon the membranes, terminating by a well-defined line, and not extending posteriorly as far as the root of the tail; beneath, the wing-membrane is thinly covered as far as a line drawn from the elbow to the knee-joint, and a band of fur passes outwards, posterior to the forearm, to the carpus, as in *Vesperugo noctula*. The face in front of the eyes is nearly naked.

Upper incisors with a posterior horizontal expansion of the cingulum, as in *S. borbonicus*; lower incisors crowded. The other teeth as in *S. Temminckii*.

Length (of an adult female preserved in alcohol), head and body 4.6 inches, tail 3.6, tail free from membrane 0.35,

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head 1.45, ear 0.9, tragus 0.45 X 0.1, forearm 3.4, thumb 0.7, second finger 5.75, fourth finger 4.15, tibia 1.4, foot and claws 0.75.

This is by far the largest species of the family Vespertilionidæ yet described, its forearm exceeding that of *Vesperugo molossus* (hitherto known as the largest species) by half an inch, and exceeding by quite one inch the forearm of the largest species of *Scotophilus*.

Hab. Lagos, west coast of Africa. Type in the collection of the British Museum.

Scotophilus hirundo de Winton, 1899

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Colour of the body above mouse-grey, beneath silvery white; wings and interfemoral membrane wholly dark, with perhaps the exception of the postcalcaneal lobe, which in the dry skin appears pale grey. The tragus is short and rounded, spatulate in form as in *S. albofuscus*, with which species the dentition also closely agrees.

Type no. 99.6.15.8 in British Museum.

Sex not known. Gambaga, 1300 feet, 30th Nov., 1898.

Measurements (taken from the dry skin): -

Head and body c. 43 millim.; forearm 31.5; thumb 5.3.

Measurements of skull: -

Greatest length 14.5; across brain-case 9; between

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orbits 6.9; tip to tip of canines 4.1; tip to tip of incisors 1.9; front of canine to back of molar series 5.2; front of palate to foramen magnum 10; mandible, greatest length 10.2, height at coronoid 4.2.

The canines have a deep broad groove in front, the tips diverging; the incisors are strongly convergent. The lower incisors are not crowded, but grow in the direction of the jaws. The first premolar in the lower jaw is large and longer than the second premolar.

In all important characters this new species closely resembles *S. albofuscus*, the colour of the fur and wing membranes alone distinguishing the two species. There can be no doubt that these two species should be placed in a distinct genus from *S. nigrita* and company, the form of the canine teeth alone being a sufficient reason; but provisionally I follow Mr. Thomas (Ann. Mus. Genov., Feb. 1890, p. 3) in placing them in this genus until the whole of this group of bats shall have been worked out.

Scotophilus minimus Noack, 1887

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(Vergl. Taf. X, Fig. 36 - 37.)

LEACH in: Transact. Linn. Soc. XIII, S. 71; PETERS, Säugethiere S 65 - 67; PETERS in: M. B. Berl. Acad. 1866, S. 679; DOBSON in: Proc. L. Z. Soc. 1875, S. 369; DOBSON, Catalogue of the Chiropt. S 256 - 266.

Nachdem LEACH die Familie *Scotophilus* 1820 aufgestellt hatte, bestimmte PETERS den Namen *Scotophilus* für die östlichen, *Nycticejus* für die amerikanischen Arten, denen nahe verwandt ist die Gattung *Atalapha*.

Diagnose von *Scotophilus* nach DOBSON:

Körper klein, mit starken kurzen Beinen und starker Flughaut. Kurze konisch abgestumpfte nackte Schnauze, Nasenlöcher ziemlich genähert, vorn nach der Seite geöffnet, mit vorgestrecktem innerem Rande, Ohr länger als breit, kurzer als der Kopf, oben gerundet, der untere Rand hinter dem Mundwinkel nach vorn und in einer convexen Grube in die Höhe gezogen, Tragus zugespitzt, Schwanz kürzer als Kopf und Leib, nur die Spitze zwischen der Flughaut frei, Sporn schwach. Flughaut zwischen den Schenkeln dick und lederartig. Schädel dick, Hinterhaupt concav; die Gaumenplatte endet hinter M 3 M in der Mitte der Jochbogen, Raum zwischen den Bullae aud. breit. Färbung olivellbraun oder kastanienbraun, unten gelblich weiss.

Zähne $I^{1+1/6} C^{1/1} P^{1+1/2} M^{3/3}$. Pullus mit einem kleinen 2. Schnedezahn aussen. Schneidezähne mit Basalwulst, stark nach hinten und innen gerichtet mit einer Furche. Der Untere erste Praem. klein.

Scotophilus minimus, F mit Schädel, Qua Mpala, Marungu. August. Der ohne Schwanz nur 34 mm lange *Scotophilus* unterscheidet sich schon durch seine Zwerggestalt von allen bisher bekannten afrikanischen Arten und hat nur in dem australischen *Scotophilus greyii* einen eben so kleinen Genossen. Im Habitus und der Färbung hat er Aehnlichkeit mit dem fast doppelt so grossen *Scotophilus borbonicus*

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(vergl. v. d. DECKEN, Reise, S. 7; DOBSON in: Proc. L. Z. S. 1875, S. 376), ebenfalls mit dem noch grösseren *Scotophilus temminckii*. Nase und Lippen sehr wenig behaart, schwärzlich, Nasenkuppe nackt, gespalten, braun, die Nasenlöcher mit wulstigem Rande, 1 mm von einander entfernt, Lippenränder warzig, Mundspalte reicht hinter das Auge, letzteres nahe am Ohr, 1 mm entfernt, von der Nase 5 mm, Wangen dünn behaart, Ohr nackt, gelbbraun, elliptisch zugespitzt, hinterer Ohrrand unten nach vorn umgelegt und mit einem Zacken fast bis zur Mundspalte reichend, Tragus über $1/2$ Ohrlänge, schmal elliptisch, in der Mitte mit einer Grube. Ohren vorn 8 mm von einander entfernt, Flughaut stark, braun, zwischen den Schenkeln gelbbraun, an den Seitell mit starken Papillen, zwischen den Schenkeln mit 8 hellen Querstreifen ähnlich wie bei *Plecotus auritus*, zwischen denen Längenpapillen stehen.

Daumen frei, an den Seiten reicht die Flughaut bis zu den Zehen, hinten bis zum Fersengelenk, 5 mm vor der Schwanzspitze mit starker Ausbuchtung, Beine kurz, kräftig, Füsse kurz, breit, Nägel kurz, gelbbraun. Schwanz stark, nach der Spitze verjüngt, 1 mm frei. Färbung oben hell olivengelbbraun, unten an der Kehle mehr gelb, Seiten hell umbra, Haar lang, fein und dicht. Flughaut unbehaart, nur die vordere Hälfte des Humerus behaart, Körper 34, Schwanz 23, Ohr 8 lang, 7 breit, Tragus 5 lang, 1.5 breit. Humerus 17, Radius 29, Daumen a 1.5, b mit Nagel 4.5; I 20; II a 30, b 12, c 12; III a 30, b 12, c 7; IV a 29, b 8, c 5. Femur -, Tibia 11, Fuss 5.5.

Schädel. Schnauze breit, Orbitalrand vorn wulstig, kleine Scheitelleiste, Schädelkapsel gewölbt, blasig nach hinten aufsteigend, Hinterhaupt schräg nach vorn abfallend, nach oben spitz zulaufend, keine Hinterhauptleiste über dem For. occipit., Bullae aud. bestehen aus 2 hinter einander liegenden Kapseln, deren vordere flache die grosse Ohröffnung enthält. Hinterhauptloch 3 mm breit, 2 hoch, Jochbogen zart, sehr dünn, in der Mitte hochgezogen, hinter M 3 beginnend und vor der Ohröffnung endend, Einschnürung zwischen den Augen mässig, Gaumen sehr gewölbt, hinten 1 mm hinter M 3 gradlinig endend, hinten nach unten gezogen, wie bei *Epomophorus*, Zahnfläche concav, vorn nach oben gezogen, Zwischenkiefer knorpelig, Nasenöffnung klein, Scheitellänge 13.5, Scheitelbreite 6, zwischen den Augen 4, Breite zwischen den Jochbogen 9, Gaumenlänge 6, Breite zwischen M 3.5, zwischen C 3, Zahnreihe 7, Jochbogen: Länge von M 3 an 3.5.

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Unterkiefer. Horizontaler Ast gerade, unten wenig gebogen, aufsteigender Ast nach oben gezogen, Rahenfortsatz gross, dreieckig, hinter M 3 steil etwas nach vorn gerichtet aufsteigend, die Spitze nach aussen gebogen, Condylus liegt dicht über dem kleinen mit der Spitze nach unten und aussen gebogenen Eckfortsatz. Länge bis zum Condylus 9.5, Höhe des horizontalen Astes 1.5, Zahnreihe 5.5; Breite bei M III 4, Rabenfortsatz 3 mm hoch.

Zähne wie oben angegeben, gelblich transparent, die beiden kleinen I oben mit Basalwulst, Spitzen nach innen, hinten ausgekehlt; C 1.5 lang, hinten aussen und breiter innen ausgekehlt mit scharfer Kante dazwischen, hinten und vorn kleiner Basalhöcker, $P = 2/3$ C, aussen C-artig, ausgekehlt, innen hinten mit tiefer M-artiger Furche, deren vordere Leiste gebogen, kleiner scharfer innerer Seitenhöcker. M I und II identisch, dreizackiger, in der Kaufläche W förmiger, innen durch eine tiefe

Furche getheilter Aussenhöcker, Aussenzacken 1 und 3 nach hinten und vorn gerichtet, scharfer Innenhöcker über $\frac{1}{2}$ M hoch. Basalwülste aussen unbedeutend, M III verkleinert, hinten nach oben gezogen, 1 scharfer hinten ausgekehlter Aussenzacken, 2 niedrigere ausgekehlte mittlere und ein niedriger mehr nach innen gerückter Innenzacken.

Unten die 6 kleinen I dreilappig auf hohen schmalen stielartigen Wurzeln, C 1 mm hoch, hinten ausgekehlt mit Basalwulst. Alle Basalwülste unten stärker als oben. P I sehr klein, P II grösser, C-artig, zweiwurzlig, Höhe $\frac{2}{3}$ C, hinten stark ausgekehlt, innen vorn und hinten mit kleinem Basalhöcker, M identisch, mit 3 hohen innen ausgekehlten Aussenzacken und 2 kleineren Innenzacken, der hintere AUssen- und Innenzacken bei M III niedriger. Die Details der Zähne nur bei starker Vergrösserung zu erkennen, Höhe der sehr spitzen Aussenzacken 0.75 mm.

Von *Scotophilus* leben ausser *minimus* in Afrika: *borbonicus* quer durch Afrika, *gigas* in West-Afrika; *temminckii* in Südost-Asien, *heathii*, *pallidus*, *ornatus*, *coromandelicus* und *emarginatus* in Indien, *rueppellii* and *greyii* in Australien

Scotophilus nigrita colias Thomas, 1904

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A richly yellow (almost orange) bellied race of *S. nigrita*.

General characters as in this species, which is Dobson's "*S. borbonicus*." Fur long, rather shaggy; hairs of back 8 - 9 mm. in length. General colour above (of the tips of the hairs) olivaceous, but the bases of the hairs are a dull sulphur-yellow, which shows through on the upper surface. Below,

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the central line is a rich chrome-yellow, deepening laterally on the sides of the belly to a golden yellow, which is especially bright on the broad band of fur extending on the wing-membrane between the elbows and knees.

Dimensions of the type: -

Forearm 55 mm. (57 in a second specimen).

Skull: greatest length 20.5; zygomatic breadth 14.5; cheek-tooth series 5.7.

Hab. Fort Hall, Kenya District, British East Africa.

Type. Male. B.M. no. 2.7.6.11. Original number 107. Collected 25th Jan., 1902, and presented by Mrs. Hinde.

The bats referable to *S. nigrita* seem divisible by colour into several geographical subspecies, of which *S. n. Dingani*, Smith, would be the Cape one, and *S. n. leucogaster*, Cretzschm., the Abyssinian. Specimens representing the true Senegalese *S. nigrita* and the Mozambique forms described by Peters are still wanting to the Museum Collection.

From any member of the group as yet described *S. n. colias* seems readily distinguishable by its brilliantly yellow under surface.

Scotophilus nigrita herero Thomas, 1906

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Hairs of back about 5.5 mm. in length.

General colour above "wood-brown," the hairs rather paler though not yellowish, towards their bases. Below dull "cream-buff."

Dimension of the type: -

Forearm 59 mm.

Greatest length of skull 20.1. Front of canine to back of m^3 7.1.

Hab. Northern Damaraland. Type from Elephant's Vley, about 18° S., 17° 30' E. Other specimens from Otjoro.

Type. Female. Original number 260. Collected 8th August, 1859, by Mr. C.J. Andersson, and received in the Tomes Collection. Twenty specimens examined.

This is the bat referred by Mr. Tomes to *Nycticejus planirostris*, Peters, but it is far paler, both above and below, than either that or any other described race of the widely distributed *S. nigrita*. The variation in colour among the large series collected by Mr. Andersson is much less than one would gather from Mr. Tomes's note. I can find no specimens which are anything like "as dark in colour as the

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figure given by Dr. Peters." Nor, in fact, in view of the faunal characteristics of the two regions concerned, would one expect to find any that were so.

[*Scotophilus nigrita nux*](#) Thomas, 1904

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A chestnut-brown subspecies of *S. nigrita*.

General characters of the smaller forms of the widely distributed *S. nigrita*. Fur short, close and fine; hairs of back about 5 mm. in length. Colour above uniform chestnut-brown, or "burnt umber" (Ridgway), the bases of the hairs slightly paler than the tips; very different therefore from the other pale brown or olivaceous representatives of the species. Under surface a rather lighter brown, approaching "russet" (Ridgway), the other forms being all yellowish or whitish below. Fur of body scarcely extending on the wing-membranes below.

Dimensions of the type (measured in spirit before skinning): -

Forearm 55 mm.

Head and body 70; tail 47; ear 15.

Skull: greatest length 20.5; zygomatic breadth 14.7; upper cheek-tooth series 5.8.

Hab. Efulen, Cameroons.

Type. Adult male. B.M. no. 3.2.4.5. Collected by Mr. G. L. Bates.

Although conspicuously different in colour from any of the known forms of *S. nigrita*, this bat so clearly represents that species in the West-African forest country that for the present I prefer to give it only subspecific rank.

[*Scotophilus nigrita pondoensis*](#) Roberts, 1946

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Differs from *S. n. dinganii* (A. Smith, *S. Afr. Quart. J.* II, 59; 1834: 'Between Port Natal and Delagoa Bay') in its much darker shade of colour, like the

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smaller species *S. viridis* (Peters), above 'antique brown', below near 'cadmium yellow'.

Type: In the South African Museum, Cape Town, S.A.M. No. 6858, collected by Captain G. C. Shortridge at Port St Johns, Pondoland, 22 August 1902. Named with the kind permission of Dr K. Barnard, Acting Director of the South African Museum.

Measurements: Head and body 89; tail 50; hind foot 12.6; ear 16. Skull, greatest length from canines 22; zygomatic width 14.8; width of brain case 10.1; mastoid width 12.5; width across molars 9.8, across canines 7.7; c-m³ series 7.2.

[*Scotophilus nigritellus*](#) de Winton, 1899

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Size smaller than *S. nigrita* (*S. borbonicus*), but apparently agreeing with that species in general form. The colour above is dark olive, the bases of the fur lemon-yellow. This colour is somewhat nearly matched in occasional specimens of both *S. nigrita* and the Indian species *S. kuhli*; but size alone is sufficient to distinguish it from either of these forms.

The ears and tragus, also the teeth and skull, closely resemble those of *S. nigrita*, but are very much smaller in size.

Type no. 99.6.15.9 in British Museum.

M, Gambaga, 1300 feet, 9th Jan., 1899.

Measurements (taken in the flesh): -

Head and body 70 millim.; tail 37; hind foot 8; ear 15; forearm 44.5; thumb with claw 5.5.

Measurements of skull: -

Greatest length 18 millim.; greatest breadth 12.9; across brain-case 9; between orbits 6.5; tip to tip of canines 5.5; tip to tip of incisors 3.5; front of canine to back of molar series 6.1; front of palate to foramen magnum, middle line, 12; mandible, greatest length 12.5, height at coronoid 5.5.

Tragus long and pointed as that of *S. nigrita*.

Canines rounded in front. Incisors very slightly convergent at their tips. First premolar in lower jaw small, barely half the length of second.

Scotophilus robustus A. Milne-Edwards, 1881

p. 1034

Le Muséum d'Histoire naturelle vient de recevoir de Madagascar une collection fort importante et riche en objets rares ou inconnus; elle lui a été offerte par M. Humblot, qui a surtout exploré, sur la côte est, la région comprise entre Fou. apointe te le lac l'Aincatre.

p. 1035

Une très grosse Chauve-Souris, du genre *Scotophilus* (*Sc. robustus*), se distingue de toutes les espèces déjà décrites; elle est de près d'un quart plus grande que le *Sc. borbonicus*; sa tête est relativement plus grosse, l'oreillon est long et effilé, le poil est partout d'un brun fuligineux.

Scotophilus rusticus Tomes, 1861

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The present diminutive species resembles in the shape of its head, ears, and incisor teeth the *S. kuhlii* and the *S. minutus*. Top of the head nearly on a level with the face, nostrils somewhat crescent-shaped, and the snout grooved as in *S. minutus*; ears small and triangular, as in *S. minutus*; tragus of uniform breadth, rounded at the end and curving inwards, the notch in its outer margin, at the base, small, and the lobe below it small and pointed. As in *S. minutus* the outer margin of the ear does not advance nearer the angle of the mouth than the base of the tragus.

Thumb and feet rather large in relation to the size of the animal, being equal in size to those of *S. minutus*. They are proportioned just as in that species.

The fur is confined to the body, both above and beneath. It is everywhere thick, soft, of moderate length, and without gloss. On the upper surface it is dark brown at the roots, with the terminal half light cinnamon-brown; on the under parts of the body, dusky at the roots, with the terminal half brownish white, excepting on the pubes, where it is almost of a uniform dirty white. The nose, ears, and wing-membranes are dark brown, the latter very narrowly edged with white, which is most distinct on each side of the feet.

The cranium resembles that of *S. kuhlii*, both in general conformation and in the number and arrangement of its teeth, being possessed of one more premolar than *S. minutus*.

Dentition: M. $2 \cdot 2 / 6$; Can. $1 \cdot 1 / 1 \cdot 1$; Premol. $2 \cdot 2 / 2 \cdot 2$; Mol. $3 \cdot 3 / 3 \cdot 3 = 18 / 18$.

The upper incisors are unequal in size, just as in *S. minutus*; the canines are relatively stouter than in that species, which is due in some measure to a slight fullness, or rounded prominence, in the middle of the thin hinder edge of the tooth; the first premolar is very small, being a mere tubercle, and is placed between the contiguous canine and premolar, and is visible only from within; the

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second resembles in shape the first and only one in *S. minutus*, which it represents. The true molars do not need mention; and the lower jaw with its teeth is so much like that of *S. minutus*, that it may in like manner be dismissed.

Scotophilus variegatus Tomes, 1861

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This handsome species, is about the size of the *S. leisleri* of Europe.

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The head is very short; the muzzle short and obtuse, but not very broad or depressed, being about as deep as it is wide. The forehead is elevated in only a trifling degree above the facial line; the nostrils are somewhat prominent, rather small, and open sublaterally, almost laterally; they are nearly round, with a narrow posterior elongation or slit not exceeding in length the

diameter of the nostril; the lips are simple, thick, and rather overhanging. The ears are small, of an irregular roundish ovoid form, their inner margin having a kind of lobe, the lower outline of which projects downwards over the forehead, and is there distinct; but blends off in an upward and outward direction to the tip of the ear, which is rounded, and has a somewhat outward direction; the outer margin approaches in some degree to a straight line, and a very narrow piece of membrane passes forward along the side of the head to near the corner of the mouth, where it terminates in a small fleshy lobulus of a rounded form. The ear-conch is rather thin, but not papillated, and has a few distinct sulci near its outer margin, and a longitudinal seam near the inner one. The tragus is broad and short, and shaped somewhat like that of the *S. borealis* of Europe. Outwardly at its base is a descending point, and above this a broad notch, which varies in depth, but is usually rather shallow; and above this it is of nearly equal breadth, the outer margin being rounded, and forming with the inner margin, which is nearly straight, a blunt angular tip, which is directed inwards.

The penultimate phalange of the thumb is a little longer than the basal one, the claw short and not very hooked. Toes rather longer than the remaining part of the foot; the claws small and black. Wing-membranes extending barely to the roots of the toes. Calcaneum long, with a considerable cartilaginous lobe near the middle of its lower edge.

The fur of the forehead extends almost to the end of the nose, in some specimens quite as far forward as the nostrils; the sides of the face sprinkled rather thickly with fine, short, whitish hairs; over the eye and between it and the nostrils are some longish black ones, which are straight and stiff. Inside the ears are a few fine adpressed hairs; and their outer surface is thickly clothed with fine fur for halfway from their base. The fur of the back extends on to the membranes on all parts, but to a greater extent on the interfemoral membrane than anywhere else. Beneath, it spreads on to the membranes, chiefly on and under the humerus, occupying the space (but not thickly) between the elbow and the knee. On their upper surfaces the toes are clothed with fine adpressed hairs; but the other parts of the foot are perfectly naked.

On all parts of the body the fur is long, soft, and unicoloured, or very nearly so; that of the upper parts cream-coloured, palest on the head and neck, and becoming considerably darker on the hinder part of the back and on the interfemoral membrane; beneath, it is of a dirty-white colour. All that of the body, but not of the membranes, has a slight ashy tinge at its roots.

The membranes are so peculiarly marked as to deserve especial

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notice. They are rather ample, thin, and remarkably marked with veins, but are not papillated. The ground-colour is pale yellowish-brown, everywhere strongly marked with veins of a deep brown colour, which are faintest near the body and on the interbrachial membrane: on the interfemoral membrane they are very distinct, not very numerous, and have a transverse disposition; beneath the humerus they have a tessellated arrangement; but beneath the forearm they are more branched; whilst between the fingers and at the ends of the wings they are run together, and a dark-brown mottled appearance is produced. The ears and muzzle pale brown; legs, tail, and wing-bones darkish brown.

The skull is remarkable for its shortness and great depth in relation to its breadth, and for the almost total absence of ridges or crests. Although the facial line of the cranium makes a near approximation to a straight line, yet the occipital region is so high as to form a facial angle of 48 degrees*. This is due in some degree to the shape of the maxillary bones, the lower or alveolar margins of which curve upwards from the root of the zygoma to the intermaxillary bones. Some approach is made to this form of cranium by that of some of the species of *Lasiurus*, as the *L. noveboracensis*; but in that species the facial angle does not exceed 35 degrees*.

The upper incisors are four in number, very unequal in size, the outer ones being almost rudimentary. The inner ones are of moderate size, acute, and with a moderate cingulum, the outer ones very short and conical, with an evenly developed and broad cingulum. Canines rather slender and pointed. First and only premolar well developed and of the usual *carnassial* form; the three true molars of ordinary form, excepting the hinder one, which is very narrow from front to back, but has the usual number of cusps.

The lower jaw has a less prominent posterior angle than in other species of the genus, but does not otherwise differ materially. The incisors have the usual external trilobed appearance; but they are thicker from front to back than is usual, and the hinder part is produced into a kind of basal lobe, which has sufficient prominence to have the appearance of a fourth cusp. The canines are rather acute, and have the cingulum produced anteriorly into an acute point, a little above the level of the incisors. The first premolar is small, conical, and with a regular cingulum; the second somewhat similar, but considerably longer, and it is succeeded by the three true molars, of usual size and proportions, excepting the posterior one, which is rather smaller than usual.

Dentition: M. $2 \cdot 2/6$; Can. $1 \cdot 1/1-1$; Premol. $1 \cdot 1/2-2$; Mol. $3 \cdot 3/3-3 = 14/18$.

"", ""

Length of the head and body: 2 4; 2 3

----- of the head: 0 8; 0 9

----- of the ears: 0 4; 0 4

Breadth of the ears: 0 4; 0 4

Length of the tragus: 0 $2\frac{1}{2}$; 0 $2\frac{1}{2}$

* Taking the lower margin of the maxillary bone as the horizontal or base-line.

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Breadth of the tragus: 0 1; 0 1

Length of the fore-arm: 1 9; 1 8

----- of the thumb: 0 4; 0 $3\frac{3}{4}$

----- of the first finger: 1 10; 1 9

----- of the second finger: 3 5; 3 2
 ----- of the third finger: 2 7; 2 4
 ----- of the fourth finger: 2 2; 2 0
 ----- of the tibia: 0 9; 0 8¹/₂
 ----- of the foot and claws: 0 4²/₃; 0 4¹/₂
 ----- of the tail: 2 0; 1 10
 ----- of the *os calcis*: 0 9; 0 9
 Expanse of wings, about: 13 6; 12 9
 Length of skull from front of intermaxillary bone to the occipital suture: 0 6; 0 6
 Breadth behind the zygomatic arches: 0 4; 0 4
 Length from the condyloid fossa to the front of the intermaxillary bone: 0 4¹/₄; 0 4¹/₄
 Length of the dental series of the upper jaw, exclusive of the incisors: 0 2¹/₂; 0 2¹/₃
 Length of the bony palate: 0 2; 0 2¹/₄
 Breadth between the points of the upper canines: 0 2; 0 2
 Breadth between the two posterior molars: 0 2; 0 2
 Depth from the occipital suture to the bottom of the auditory bullæ: 0 4¹/₂; 0 4
 Greatest length of lower jaw: 0 5¹/₄; 0 5
 Breadth of lower jaw vertically from the coronoid process: 0 1³/₄; 0 1²/₃
 Length of the dental series, exclusive of the incisors: 0 2³/₄; 0 2¹/₂

Hab. "Otjoro, December 1st, 1859."

Obs. I have chosen to give an ample description of this species because it differs in some respects from any other species of the genus which I have seen, the differences appearing to me to be such as may (if found in any other species) be sufficient to point out another minor group, parallel with those which have been partially indicated by Blasius, and more fully by Kolenati. Broadly, these differences may be stated thus: - *Cranial portion of the skull much raised, instead of being depressed; muzzle not depressed; cutaneous system thin and elaborately veined, instead of being thick and leathery; fur long, soft, and unicoloured, and covering nearly the whole of the face.*

Scotophilus welwitschii Gray, 1866

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Among an interesting series of Bats from Angola, collected by Dr. Welwitsch, and most kindly presented to the British Museum, is a very interesting and ornamental species of *Scotophilus*, with the wings coloured like *Vespertilio pictus* of Pallas.

This coloration of the wings seems common to several Bats belonging to different genera; but I have not before observed it in a species of *Scotophilus*.

SCOTOPHILUS WELWITSCHII (Pl. XXIV.)

Brown. paler beneath; hair of the back black, with brown tips, which are longer and paler on the hairs of the under surface. The ears rather elongate, longer than head. tip rather acute; tragus elongate, lanceolate, acute, nearly half as long as the ear. The wings blackish brown. yellow-dotted, and yellow (or red brown perhaps when alive) near the body, and on and near the arms and fingers, and between the shoulders and arm-bone; interfemoral membrane yellow, black-dotted, and with a dark hinder edge, the upper surface near the base of the tail hairy; heel-bone elongate, as long as the shin. Feet pale yellow; toes black at the end; .wings to the base of the toes. Thumb - upper joint black, much longer than the lower, which is yellow. The face hairy to the end of the nose; just above the nostrils. Upper cutting-teeth 1,1 ? . large, blunt; premolars ²/₁ ²/₁, the front upper large, triangular; the hinder small. rudimentary. Forearm-bone 2 inches 1 line long.

Hab. Angola (Dr. Welwitsch; B.M.).

Selysius Bonaparte, 1841

Selysius

Dei *Vespertilionini* non dati per ora, ma forse da aggiungersi alla Fauna italiana, nomineremo il *V. mystacinus*, Leisler, che facciamo tipo di nuovissimo genere ben caratterizzato dalla forma del cranio, dalla dentatura, dal folto pelo, dall'opacità e intaccatura delle orecchie, e soprattutto dall'inceppamento dei piedi: il nome di *Selysius* che gli apponiamo fa commendevole ricordo di stima al giovane naturalista di Liegi, autore della *Fauna belgica* e di altri applauditissimi scritti. Nè taceremo del vero *V. emarginatus*, Geoffr. suo congenere; avvertendo qui essere quello da noi figurato sotto tal nome il *V. nattereri*, Kuhl, anello di congiunzione tra i due generi *Selysius* e *Vespertilio*. E qui giova dire che per timore di confusione usiamo il nome *Vespertilio* a denotare quel gruppo cui per abbaglio lo restringemmo, ma che tal nome giustamente dovrebbe trasferirsi a quella suddivisione contenente il vero *Vespertilio murinus* di Linneo. Avendoci finalmente accertati il colore del vello, la frequenza in Svezia, ed altre ragioni inoppugnabili, che il *V. murinus*, L. corrisponde al *V. discolor*, Leisl. deve a questo ripristinarsi almeno il nome specifico, comunque siasene abusato, applicandosi nella Svezia stessa dal Nilsson, ora ricredutosi, al *V. daubentoni*, Leisl., dagl' Inglesi tenacemente al *V. pipistrellus*, Daub., e da noi stessi travolti nella piena corrente al *V. myotis*, Bechst.; cui preghiamo non chiamisi altramente d'ora innanzi, il nome *Myotis* potendo anzi divenir generico per chi non tollerasse eccezione di regola.

Spectrum vulgare Gray, 1870

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Ears small; fur soft, abundant, spreading, blackish chestnut; head pale yellow ; the upper arm, sides of the shoulders, back, and

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the rump bright golden yellow; underside of thighs and pubis yellowish.

La Roussette, *Brisson*. La Rossette, *Buffon*., H. N. x. t. 14. *Vespertilio vampyrus*, *Schreb*. Not *Linn*. *Pteropus vulgaris*, *Geoff*. *Ann. Mus.* XV. P. 92 ; *Temm. Mon.* i. p. 132 ; ii. P. 74, t. 38 (young).

Hab. Isle of France; Bourbon; Madagascar ?

Sphyrocephalus A. Murray, 1862

p. 8

Dental formulary :-

Incisors. 4/4 Canines. 1/1 Molars: Premolars? 1/2 True molars? 2/3

Head very large and oblong; the lips largely developed and expanded. Ears rather large, without tragus. Thumb and index-finger of hand unguiculate, the other fingers without claws. Tail wanting.

Sphyrocephalus labrosus A. Murray, 1862

p. 8

Brown, with a few whitish hairs at the base of the ears. The head very large, massive, half as long as the whole body, oblong, and as broad at the muzzle as at the top of the head, with some resemblance to a hammer, whence the name hammer-headed (*Sphyrocephalus*), rather more than twice as long as deep; ears rather large, destitute of tragus; eyes rather large; eyelids provided with eyelashes; nostrils large and tubular; lips extraordinarily developed; both upper and lower lips of a very smooth semi-muco-membranous texture, corrugated and tuberculated at the margins; the upper lip with a tuberculated prominence in the line of the two outer incisors, and a more elongated tuberculated ridge further up, in the line of the two inner incisors; the external lateral margin expanded into a sort of stiff semi-cupshaped flap with a tuberculated edge, rising to meet the nostril and then descending, following its lower edge, and terminating in a curved scroll-like coil in the nostril.

The same semi-muco-membranous lip is spread over the front of the lower jaw, forming a sort of chin. The skin in the neighbourhood of the lips, and extending upwards and backwards along the nasal bones, is covered with the same sort of fine velvety down which surrounds a horse's nostrils; the hair on the rest of the head is flocky; on the downy portion there are on the sides of the upper

p. 9

lip three rows of papillæ, each with a long whisker-hair springing from it. In the specimen before us these papillæ are arranged four in the two first rows and three in the last; similar papillæ and hairs run up the downy space covering the long nasal bones, in three rows, past the eyes and quite to the forehead, numbering each nine or ten papillæ, the middle row being shorter than the two others. The gape of the mouth is large, extending back fully a third of the head; the lip does not encroach on the outside of the face along the gape; it is only directly in front that it is so much developed; the upper lip is connected with the gum by a broad thick ridge uniting them together in the line of the symphysis of the intermaxillary bones.

The disposition of the teeth is as follows:-

They are all well separated from each other, none touching each other except, perhaps, the last molars; the incisors of the upper jaw are minute rounded points; in the lower jaw they are equally minute, but transversely oblong and bilobed. The canine teeth in both are well developed and of the usual form; beyond the canine there is a minute tooth (a mere point) in the lower jaw which is wanting in the upper jaw; the next tooth beyond it is almost exactly of the form of the canine, and is probably a pre-molar; the remaining teeth, two in the upper and three in the lower jaw (probably true molars), have their crown divided longitudinally; in the upper jaw each ridge slopes backwards, in the under jaw the external ridge is bilobed. The palate has strong, elevated, transverse ridges running across from interspace to interspace between each tooth. The tongue is rather large, and covered with a sort of tessellated pavement of large flat papillæ; it is free very far back. Under it and lying in the hollow of the mouth, occupying the whole breadth for a short space in front between the rami of the lower jaw, is a very curious membrane fringed with slips or plaits - a sort of second tongue, calling to recollection a somewhat similar organ or structure under the tongue of the Loris and Lemur. In these it assumes the form of an aponeurotic lamina, which is divided at its anterior thinner end into filaments or slips. "This arrangement (a development of the frenum of the tongue)," says Van der Hoeven, "has been described incorrectly, in my judgment, as though the tongue were double, or even as if a bird's tongue were under the mammalian tongue." It may be a development of the frenum, because two things connected together,

although at opposite ends, may always be said to be parts of the same thing however distant they may be. But I would only observe that in the present instance the frenum of the tongue is situated very far back, and it seems to me that it may just as possibly be a development of the floor of the mouth as of the frenum of the tongue. I do not suppose that Dr. Van der Hoeven would think it necessary to look elsewhere than to the walls of the œsophagus to find the structure from which the elongated papillæ lining the œsophagus, in the hawk-billed turtle for instance, had been developed. There is a tendency to similar structure in other parts than the tongue in many animals - on the palate and back of the mouth, for example; and I should not be disposed to seek further than the place from

p. 10

which it springs for the source of this development. It does not extend much further back than the lower canine teeth; but there is a slight plait or perceptible line running on each side all the way to the back of the mouth, giving the appearance of a second thin flat tongue lying in the hollow of the mouth, tied down like the tongue of a crocodile; but the separation here is a mere trace, and it is only the fringed margin in front which is free.

The neck is distinct, and the skin has the appearance of having some cellular space between it and the muscles.

The body is oblong and compact; the ribs descend far, and the lower ribs are very large; the stomach is moderate in size, furnished internally with several large transverse folds; the small intestines are not very long; there is no cæcum.

The arms have the thumb and index unguiculate, the rest of the fingers are without claws; the thumb has two phalanges, the rest have three phalanges. The thumb is united to the wing by a membrane stretching on both sides over the whole of the proximal and half of the distal phalanx; the proximal phalanx of the thumb is shorter than the distal. The winged membrane is not extended across the back, but is very ample; the winged space between the third and fourth fingers, and between the fourth and the body, has in its middle numerous longitudinal bundles of muscular fibre interwoven without attachments, and one or two similar transverse bundles; these are probably for the purpose of assisting in the folding of the wing. Along these bundles of muscular fibre the membranous wing is closely wrinkled; and there is little doubt that they will also strengthen the membrane where they occur.

The hind feet are uniform and all unguiculate; they are united to the body by an interfemoral membrane, which has a single large bundle of muscular fibres stretching obliquely across from the foot to the coccyx.

The testicles are situated under the skin on each side of the male organ, and are round.

There is no tail.

The length of the whole body, in the specimen from which the above description is taken, is nearly 7 inches; the length of the head 3 1/4 inches, its depth about 1 1/4 inch. The stretch of wings is 28 inches across.

The most remarkable features in this animal are its large hammer-shaped head, and the great external development of its lips. Its whole structure is essentially that of a Pteropine Bat, with some modifications showing a tendency towards the *Rhinolophi*. No species having any of the nasal appendages peculiar to that section of the Bats has yet been found among the Pteropine Bats. They are strictly frugivorous, and have the nose like that of a fox or dog. The present species, although it has not any nasal appendages, has labral expansions which may possibly be analogous to them, and the animal may possibly have peculiar habits to which the structure of these organs is especially adapted.

Unfortunately, in the only specimen yet received, the stomach and

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intestines were wholly empty; so that we cannot speak of its food with positive certainty. The teeth are Pteropine in character, but not so absolutely so as to preclude the possibility of this creature being at least partially insectivorous, the molars showing a tendency to mammillation on the external side of the longitudinal ridges into which they are separated. The large folds in the interior of the stomach seem to point to a vegetable diet.

The sublingual fringed membrane is also an interesting peculiarity, not only on account of its rarity, but because one of the few other instances where it has been noticed is in an animal having no one thing in common with the present, except that of living in the same country. We sometimes see this happen; an abnormal structure or peculiarity occurring in an animal restricted to one country will be found repeated in some other animal of that country no way connected with or allied to it.

This Bat was sent to me by my excellent friend, the Rev. Wm. C. Thomson, one of the missionaries of the United Presbyterian Church of Scotland, stationed at Old Calabar - a true Christian, an excellent naturalist, and one whose devotion to the cause he has undertaken, viz. the amelioration of the African negro, has been proved by the greatest sacrifices from his youth upwards.

footnote p. 8:

* Since this paper was in print, the last number of the 'Proceedings of the Academy of Natural Sciences of Philadelphia' has been received in this country (the first copies arrived on 19th February, 1862); and in it I find a description of a new Bat, which probably belongs to this species, by Dr. Harrison Allen (Proc. Acad. Nat. Sc. Phil. July 1861, p. 156). It is said to be taken from a specimen collected by M. Du Chailu, and is named by Dr. Allen *Hypsignathus monstrosus*. If it is the same species, of course Dr. Allen's name must take precedence. His description does not quite correspond with mine, but, judging from the description of the nose, may, perhaps, have been taken from a dried skin, whereas mine is

from a fine example in spirits. M. Du Chaillu has exhibited no specimen of this Bat in England.

Tadarida (Chaerephon) cyclotis Brosset, 1966

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Type M ad. - Bélinga. - Janvier 1963.

Description

Molosse d'assez grande taille (avant-bras 51.5 mm), séparable à première vue des espèces africaines de dimensions voisines par des oreilles larges et arrondies, des formes moins massives et un pelage relativement long et épais. La morphologie crânienne, la dentition, le tragus présentent des particularités distinctes.

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Dimensions (en mm):

Longueur tête et corps: 70

Queue

- partie incluse: 20

- partie libre: 18

Oreilles

- largeur maximale: 20

- longueur maximale: 20

Tragus

- longueur: 1.8

- largeur: 1.1

Anti tragus

- hauteur: 5

- largeur: 6

Avant-bras: 51.5

3^e doigt

métac.: 49

1^{er} phal.: 21

2^e phal.: 20

3^e phal. incomplètement ossifiée: 9

4^e doigt

métac.: 47

1^{er} phal.: 17

2^e phal.: 11

3^e phal. incomplètement ossifiée: 3

5^e doigt

métac.: 29

1^{er} phal.: 15

2^e phal.: 6

Tibia: 19

Pied (sans les griffes): 12

Pouce (sans l'ongle): 8.5

Crâne:

Longueur totale: 21.4

Longueur condylo basale: 18

Largeur zygomatique: 12.3

Largeur boîte crânienne: 11.3

Rétrécissement interorbitaire: 4.4

Rangée dentaire supérieure (de l'incisive à la pointe postérieure de M 3): 9,1

Ecartement à la pointe des canines: 4.8

Longueur de la mandibule: 15

Longueur de la rangée dentaire inférieure: 9.7

Pelage et membranes:

Pons de couleur uniforme sur toute leur longueur. Le dos est brun rougeâtre. La partie ventrale présente une teinte plus claire, rougeâtre-orangée d'une tonalité chaude et brillante .. base de

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membrane réunissant les oreilles en avant existe l'amorce d'une "crête" basse, formée de poils mous et serrés, de couleur brun noirâtre. Le pelage se prolonge sur le patagium, de chaque côté du corps sur une bande d'environ 1 cm de largeur. L'uropatagium est couvert d'une pilosité décroissante, de la naissance de la queue jusqu'à la partie libre de celle-ci. Cette dernière est complètement nue. Sur la face dorsale, plantés au niveau du fémur, on note quelques longs poils raides. Les pattes, les doigts externes surtout, montrent des rangées de poils raides, clairsemés et courts, de couleur jaune doré. Des

poils semblables, mais plus courts existent également sur un "bouton" sous mentonnier, aux commissures des lèvres, au point de jonction des deux oreilles, à l'extrémité supérieure de la membrane réunissant celle-ci.

La peau des avant-bras est brun-clair dessus, dépigmentée dessous, la face interne des avant-bras étant de couleur chair. La partie interne de l'uropatagium est également très claire. La lèvre supérieure est brune, l'inférieure est blanchâtre.

Tête et oreilles:

La lèvre supérieure est profondément plissée. La partie nasale est fortement proéminente, comme chez la plupart des Molossidés. Les oreilles sont grandes et larges. Le bord externe est arrondi. L'antitragus est grand, aplati, et bien détaché de l'ensemble de l'oreille. Le tragus est petit et caché par l'antitragus (fig. 9).

Une membrane sagittale, épaisse et haute de 6 mm réunit les deux oreilles en avant.

Corps:

Le corps ne présente pas les formes lourdes habituelles chez les *Tadarida*. Les membres inférieurs, du type molossidé, sont relativement grêles, tandis que la queue est également assez courte.

Le cinquième doigt est très court par rapport au troisième, ce qui réalise une aile longue et étroite, de faible surface portante. Ce rapport est caractéristique des formes à vol direct, élevé et rapide. Les ailes de *T. cyclotis* sont très semblables à celles de *T. condylura*.

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Le grand développement du pouce et des callosités du pouce désignent *T. cyclotis* comme un grimpeur apte à se glisser dans des fissures étroites.

Crâne:

Sa structure d'ensemble est légère par rapport à celle des autres *Tadarida*. La boîte crânienne relativement allongée est marquée sur le dessus d'une dépression accusée au niveau de la grande suture transversale. La région occipitale est fortement convexe. La crête sagittale est nulle et les crêtes lambdoïdes peu développées. Les bulbes auditifs sont par contre grands, en liaison avec l'oreille externe qui est elle-même de forte taille (fig. 10).

La partie nasale est relativement étroite. Le rétrécissement interorbitaire est prononcé. Le palais est allongé, fortement concave et trapézoïdal. L'échancrure du prémaxillaire, en arrière des incisives, qui est un caractère du genre *Tadarida*, n'apparaît plus clairement. Mais le séjour du type de *T. cyclotis* dans le formol a durci les cartilages du crâne, qui ont pu oblitérer l'échancrure, si elle existe.

La dentition est peu spécialisée. Elle est plutôt faible, quoique la canine soit longue et mince. PM 1 est bien développée et située dans la rangée dentaire. Les incisives inférieures, au nombre de quatre, sont plantées en demi-cercle. Elles sont fortement bifides.

Mieux que toute description, le dessin ci-contre donne une excellente idée du crâne de *T. cyclotis*.

Comparaison entre *T. cyclotis* et les espèces africaines les plus voisines *Tadarida condylura* à l'avant-bras plus court; les formes sont plus lourdes, les oreilles beaucoup moins larges. L'antitragus est moins grand, le crâne est moins long, mais plus massif. Les crêtes lambdoïdes et sagittales sont beaucoup plus développées. La dentition enfin présente de notables différences. Le Muséum de Paris possède des séries de *T. condylura* qui nous ont facilité ces comparaisons.

Reste *T. trevori*, espèce plus rare, et que nous n'avons pu examiner. D'après les descriptions d'ALLEN et de VERSCHUREN, *T. trevori* aurait des dimensions et un aspect voisins de *T. cyclotis*. Mais le rapport du 3^e doigt et du 5^e est différent, réalisant chez *trevori* une aile moins étroite. L'oreille est beaucoup moins large, et le tragus n'est pas carré chez *cyclotis* comme il l'est chez *trevori*. Enfin le crâne de cette dernière espèce est plus grand dans toutes ses dimensions, et le rapport de sa longueur à sa largeur, ainsi que l'écartement des maxillaires, doivent lui donner un aspect massif et puissant, différent de celui de *cyclotis*.

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L'aspect de la tête et du crâne de *Tadarida cyclotis* rappelle un peu celui des Molossidés du genre *Otomops*. On notera qu'il s'agit là probablement de formes forestières, alors que les *Tadarida* sont en général liés aux zones découvertes: savanes et semi-déserts. L'évolution vers une structure plus légère et un grand développement de l'appareil auditif est peut-être caractéristique des Molossidés forestiers, circulant à la cime des arbres; ces espèces auraient développé une morphologie tendant à augmenter la maniabilité du vol, par un allègement des formes et une hypertrophie de l'appareil écholocateur.

Biologie

Ce beau Molosse se fit prendre au crépuscule dans un filet japonais, tendu sur une levée de terre qui traverse le marigot bordant le terrain d'aviation de Bélinga. L'animal venait de toute évidence boire à la surface de la nappe d'eau. Par la suite, nous avons passé une dizaine de soirées à cet endroit à faire le guet, pour essayer de voir et de tirer d'autres individus. Malheureusement, aucun Chiroptère de ce type ne fut observé. La biologie de *Tadarida cyclotis* reste donc inconnue. Tout au plus peut-on dire, compte tenu de la localisation de la capture, qu'il s'agit probablement, d'une forme forestière.

Tadarida (Chaerephon) faini Hayman, 1951

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Type (in the Congo Museum, Tervuren): M skin and skull, Congo Mus. n° 20509, original number 1, collected in June 1951 by Dr. A. FAIN at Wago Forest, between Blukwa and Lake Albert, Belgian Congo. (Blukwa lies at about 1° 45' N., 30° 37' E. at about 2.000 metres above sea-level). The paratype, a F skin and skull, original, number 2, Brit. Mus. n° 51.629, from the same locality and collector, has been presented to the British Museum (Natural History).

Diagnosis: The species is distinguished primarily by its light ochraceous-tawny body colour above and below, its semi-translucent wings and its medium size (forearm 38 mm. in the type).

Description: The head and body of the type, both dorsally and ventrally, are clothed with short hair of an almost uniform ochraceous-tawny colour. The underside is slightly lighter in tone than the back, and in the median line of the abdomen there is a slightly yellowish wash. The hairs everywhere are slightly paler basally, but the paler bases do not show through to affect the general colour. On the underside of the wing membrane a narrow band of yellow hair runs from the base of the humerus to the base of the femur, flanking the side of the body. The crown of the head is nearly naked, and the lappet of skin joining the bases of the ears is clothed with dense but short hairs of the same colour as those on the head and body.

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The ear membranes are dark, but the wing and interfemoral membranes are semi-translucent. The wing membranes are attached to the middle of the tibia.

The female skin, apart from being slightly paler above than the type, shows no differences.

The skull and teeth are characteristic of the subgenus.

Measurements: (in the absence of original measurements, those now given, taken from the dry skins, must be regarded as approximate only, except in the case of the forearm measurements). Type and paratype: (all measurements in millimetres), head and body 59, 60; tail 33, 29; hind foot 10, 10; ear 15, 16; forearm 38, 39. Skulls of type and paratype: greatest length 15.7, 15.8; condylobasal length 14.8, 14.5; zygomatic breadth 10, 10.1; mastoid breadth 9.8, 9.4; maxillary breadth 6.8, 6.9; interorbital width 4, 4.1; breadth at base of canines 4.1, 4.1; upper tooth row, including canine, 5.7, 5.7; length of mandible 11.1, 10.9; lower tooth row, including canine, 5.7, 5.9.

Remarks. No other African member of the group combines the distinct colouration and size of the present species.

Nyctinomus ochraceus J. A. ALLEN (1917) from Medje, Ituri Forest, is one of the more brightly coloured forms of about the same overall size, but according to the description is dark chestnutbrown above, with only the bases of the hairs ochraceous buff- and is orange below. It has also a considerably longer skull.

SIMPSON (1945) has regarded *Chaerephon*, as well as *Mops*, as no more than subgenerically distinct from *Tadarida*, which antedates *Nyctinomus*. TATE (1941) has pointed out that certain skull characters used in the past to distinguish *Chaerephon* are not constant, and it therefore seems best to treat *Chaerephon* as a subgenus of *Tadarida*.

Tadarida (Chaerephon) gallagheri Harrison, 1975

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Holotype.

Harrison Zoological Museum (Reg. No. 1.7797), now deposited in the British Museum (Natural History). Adult male, obtained on 14th Nov. 1974 at Scierie Forest, 30 km SW of Kindu, Zaire (3° 10' S, 25° 49' E), by Major M. D. Gallagher.

Diagnosis.

A small bat, forearm 37.5 mm; greatest length of skull 16.0 mm. General features are those of subgenus *Chaerephon*, the conjoined ears with a frontal crest, the premaxillae co-ossified and m³ complete. This species is, however, unique amongst Molossid bats in the possession of large paired nasal inflations highly reminiscent of *Rhinopoma*. It is also at once distinguishable from all other African Molossids by the presence of a deep, well-defined inter-aural pocket, containing the frontal crest. The anterior extension of this pocket projects into the inter-aural membrane as a bulbous anterior protrusion, which in profile overhangs and projects in front of the muzzle.

Description.

The holotype is the only known specimen. The ears are large and extend well in front of the muzzle when laid forwards; their anterior

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borders are squarish; there is a well-developed subrectangular anti-tragal lobe, at the base of their convex, external margin; the tragus is very small, its upper surface with two small indentations distinct minute basal lobule posteriorly. The ears are

united across the forehead by an inter-aural membrane, with a flat-topped convex superior margin, resembling a truncated triangle; a prominent globular swelling projects from the middle of its anterior surface, projecting above and well in front of the muzzle in profile. This swelling, which has a faintly marked median rib dorsally, is in fact the anterior extremity of a remarkable inter-aural pocket, situated on the crown of the head, behind the inter-aural membrane. The pouch is deep, lined by two vertical hairless folds of skin 3.2 mm high, which extend from the posterior aspect of the inter-aural membrane on each side of the globular swelling and unite with each other on the crown of the head. The floor of the pocket is smooth, but it contains a frontal crest of hairs, which are uniformly dark brown; in colour and some 9 mm in length. The muzzle is blunt, with a transverse ridge above the nostrils, and the upper lip is marked by vertical furrows. The wings are long and narrow; the wing membranes, tail membrane and ears uniformly greyish black in colour. The long tail projects for three-quarters of its length beyond the short tail membrane. The pelage is uniform in colour, umber brown, so far as can be ascertained

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in the single alcoholic holotype, and scarcely lighter on the ventral surface, where there is no pattern. It extends as a narrow band on to the ventral wing membrane from axilla to groin, but not significantly elsewhere on to the membranes. The toes are fringed with short whitish hairs.

The skull is extremely distinctive, its general form elongated and narrow. The rostrum is very long and terminated anteriorly by pronounced paired nasal inflations, surrounding the broadly flask-shaped nasal aperture and imparting an extremely unusual truncated aspect to the front of the rostrum. The inflations extend laterally so as to be visible from the ventral aspect of the skull. Pronounced lachrymal projections are present at the anterior orbital margins and the infraorbital foramina open on the sides of the rostrum between these and the back of the nasal inflations, at the level of the anterior border of the second upper premolar. The interorbital region is long, with slightly swollen sides; suprabital ridges are almost absent. The zygomatic arches are slender and not widely flared, rather straight-sided when viewed from above. The braincase is moderately inflated, lacking any sagittal crest and the lambdoid crests are weak. The dorsal profile is rather low, with the braincase only slightly elevated above the rostrum. The premaxillae are completely co-ossified and there is no palatal emargination; the palate is long, with a median concavity and slightly excavated on each side postero-laterally behind the level of m^3 . The mesopterygoid space is broadly rounded anteriorly; the basal pits are deep and rounded, well defined posteriorly; the tympanic bullae are moderately large. The mandible is long and slender, with a blunt angular process and low coronoid.

The dentition is not heavy; the upper incisors are relatively large; well, separated from one another, flattened pyriform in shape, with their points directed inwards slightly and each with a small outer basal cingular cusp. The upper canines are slender, with small posterior basal cingular cusps. The small upper premolar is situated in the toothrow just behind and lateral to the cingular cusp of the canine; the second upper premolar is well developed, attaining fully three-quarters of the height of the canine; m^1 and m^2 are subequal, each with well developed hypocones; the metacone dominant in each and the styles well formed; m^3 has a well developed third commissure. Four well spaced lower incisors are present, the inner pair larger and faintly bifid. The lower canines are slender with anterior and posterior basal cingular cusps. The second lower premolar clearly exceeds the first in crown area and height, while m_1 is clearly larger than m_2 , especially in width of the talonid; m_3 is distinctly smaller; in all the lower molars the protoconid is dominant.

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Measurements of the Holotype.

Total length 77 mm; forearm 37.5 mm; hind foot (s. u.) 7.8 mm; tail 28 mm; ear 19 mm. Cranial measurements: greatest length 16.0 mm; condylobasal length 14.9 mm; zygomatic width 8.9 mm; breadth of the braincase 7.8 mm; postorbital constriction 4.2 mm; width of the rostrum across nasal inflations 5.3 mm; median palatal length 7.0 mm; mandible 10.6 mm.

Remarks.

This remarkable little bat is at once distinguishable from all other known Molossids by its unique nasal inflations, highly reminiscent of *Rhinopoma*, a feature so distinctive as to demand consideration of generic status for it. It is clear, however, that in most features it represents a highly specialised *Chaerephon*, a swift-flying species which has perhaps developed some special modification of the nasal region for echolocation. Amongst the African Molossids, which have been

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recently listed and keyed by Hayman and Hill (1971), it finds resemblances with *Tadarida* (*Chaerephon*) *major* (Trouessart, 1897), which however lacks a clearly defined inter-aural pocket, as well as having several cranial differences in addition to the lack of nasal inflations. These are an upturned occipital region, wide flattened braincase and shallow, diffuse basal pits. It is also larger, with a forearm length of 42 - 44 mm and greatest cranial length 18.5 - 19 mm (Rosevear, 1965). In respect of the remarkable inter-aural pocket *T. gallagheri* most closely approaches an Oriental species, *Tadarida* (*Chaerephon*) *johorensis* (Dobson, 1873), which has a rather well defined pocket, differing however, in some details of structure from this species. The structure of the pocket in *johorensis* has been well described by Hill (1974). It differs particularly by being partially divided by a median septum, and the anterior face of the membrane does not project forwards in such a striking manner. *T. johorensis* also differs in some cranial features, as well as lacking nasal inflations, thus the narial emargination is rounded posteriorly, instead of being flash-shaped, with a pointed posterior extension as in *T. gallagheri*. It is also a larger species (forearm 46 - 50 mm). The discovery of a female *T. gallagheri* will be awaited with interest as its aural structure particularly may differ from the male.

Tadarida (Chaerephon) tomensis Juste and Ibáñez, 1993

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Holotype. - Adult female; skin, skull, and partial skeleton; Estacion Biologica de Donana (EBD) 18256, field number B7085; from Praia das Conchas, 3 km NW Guadalupe, Sao Tome Island, Republic of Sao Tome and Principe (0°24'N, 6°37'E); obtained on 22 April 1988 by Javier Juste and Antonio Ayong.

Paratypes. - Two adult females; in alcohol, skulls removed; EBD 17371 and 17734. EBD 17371 is from Agua Izé, São Tomé Island, and was collected by Javier Juste and Carlos Ibáñez on 11 April 1988. EBD

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17734 is from Praia das Conchas, São Tomé Island, and was collected by Javier Juste and Antonio Ayong on 22 April 1988.

Distribution. - Known only from São Tomé Island where it has been collected at two localities.

Diagnosis. - Interaural membrane bearing a characteristic globular swelling on the anterior margin, similar to that found only in *T. (Chaerephon) gallagheri* among all African species. In lateral view, the interaural swelling extends slightly forward of the muzzle (Fig. 1). The anterior rostrum is inflated on each side of the anterior nares from (and including) the premaxillae to the lacrimals. The nasal opening is elongated when viewed from above (the nasals appear emarginate) with the posterior margin extending posterior to the lacrimal processes and the maxillary root of the zygomatic arches (Fig. 2). Also in this dorsal view the nasal orifice is shaped as an elongated bell; not flaskshaped as in *T. gallagheri*.

Description. - A small free-tailed bat (length of forearm of holotype, 38.5 mm; other measurements in Table 1) with round-tipped ears wider than long that extend forward slightly ahead of the muzzle. The well-developed antitragal lobe is subrectangular. The tragus is minute, and bears three lobes on its upper margin and a lower basal lobule.

A striking feature is the appearance of the interaural membrane (Fig. 1). The anterior margin of this membrane has the form of a truncated triangle when displayed that is slightly elevated centrally and has a small depression. The anterior margin of the in-

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teraural membrane also bears a central globular mass that extends a little beyond the muzzle. This mass is divided longitudinally by a medial groove and surrounds a 3-mm-deep pouch connected to the interaural membrane by a small median ridge. The base of the pouch is lined with a dense layer of brown hair that contrasts with the essentially naked interaural membrane.

The wing membrane is narrow, whitish, and nearly transparent in the middle and at the tips of the wings, and brownish black proximally along the sides of the body and the uropatagium. The body is dark brown dorsally, brown on the ventral sides and flanks, but whitish to pale-brown on the venter medially. The body pelage is dense, relatively long, and extends as a narrow strip along the flanks and uropatagium, where it is most conspicuous on the venter. The dorsal pelage is longest (ca. 7.0 mm) and has a whitish to pale-brown base; that of the venter is shorter (4.0 mm) and the individual hairs are tricolored-whitish basally, brown medially, and with pale-brown tips.

The rostrum is inflated laterally from the lacrimals to and including the premaxillae (Fig. 2a) on each side of the comparatively long and broad narial opening. The lacrimal processes are well developed, but not as long as in *T. gallagheri*. The braincase is rounded and only slightly elevated above the rostrum. In the specimens examined (females), supraorbital ridges and a sagittal crest are lacking; lambdoidal crests are weakly developed. The anterior insertion of zygomatic arches shows a conspicuous flattened space anterior to the orbital fossa. The zygomatic arches are slender and relatively straight when viewed dorsally. The premaxillae are fused; the degree of anterior palatal emargination resembles level 2 of Freeman (1981). The palate is broad, concave medially, and achieves its anterior maximum width at the level of the last premolar. The mesopterygoid fossa becomes wider posteriorly. The basisphenoidal pits are well defined (diameter, 1.2-1.6 mm) and deep (level 4 of Freeman, 1981), separated by a 0.2-mm septum (Fig. 2c). A line connecting the glenoid fossae passes through the anterior one-half of the pits. The auditory bullae are of moderate size. The mandible is slender, has a low coronoid process and a somewhat more developed angular process.

The dental formula is $i\ 1/2$, $c\ 1/1$, $p\ 2/2$, $m\ 3/3$, total 30. The upper incisors are large and separated by a space equal to the width of the cingulum of either incisor. The canines have a small posterior cusp on the posterolingual cingulum. The first upper premolar is small, aligned with the toothrow, and in contact with the canine and second upper premolar. The last upper premolar is well developed with a high crown and > 50 % the length of the canine. The first two upper molars are similar in size; labially the metacone is higher than the paracone, and lingually the protocone is much better developed than the hypocone. The third commissure of M3 is well developed and about the same length as the second (stage 2 of Freeman, 1981, scale).

Each of the lower incisors is small and separated from each other, the inner pair is larger and weakly bilobed. The lower premolars are caniniform, the second almost twice the size of the first. The first two lower molars are subequal in size; the third is smaller. The protoconid is the dominant cusp, followed by the hypoconid.

Comparisons. - Among African members of the subgenus *Chaerephon* with interaural membranes, only *T. major* shows some similarity to *T. tomensis* and *T. gallagheri* in the interaural structure, but *T. major* lacks a well-defined interaural pocket and shows striking differences in the skull, such as its flattened braincase and its shallow basisphenoidal pits (Freeman, 1981; Harrison, 1975).

Based on morphology of the interaural membrane and longer rostral inflations in the skull, the closest relative to *T. tomensis* among all African molossids is *T. gallagheri*, known only from the holotype, a male, from Kindu (Kivu region, East Zaire) and

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[table removed - eds.]

described by Harrison (1975). Body measurements are similar, but the two female specimens from São Tomé Island are slightly larger; the remaining female from São Tomé Island is noticeably smaller (Table 1). The interaural membrane in *T. tomensis* differs from that of *T. gallagheri* because the globular swelling is less projected over the muzzle. The fold that borders the pouch in *T. gallagheri* (Harrison, 1975) is absent in *T. tomensis*.

The most conspicuous differences between the two species occur in the skull (Figs. 2a and 2b). Specimens from São Tomé Island have a narrow postorbital constriction; yet a broader zygomatic width, especially as shown by the females from São Tomé Island, when compared with the holotype of *T. gallagheri* (Table 1). The shapes of the anterior insertions of the zygomatic arches are wider and flared in *T. tomensis* (Fig. 2a). While the nasal inflations are much less conspicuous in *T. tomensis* than in *T. gallagheri*, the nasal aperture is longer in *T. tomensis*, and the posterior border lies behind a line across lacrimals. The form of the nasal opening clearly is different for the continental and insular species (Fig. 2a). The palate differs by being narrower posteriorly in *T. tomensis*. Basisphenoidal pits are larger

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and deeper, and the septum is narrower in *T. gallagheri* (Fig. 2c). Finally, the occipital is blunter in *T. tomensis* than in *T. gallagheri* when seen laterally, being more similar to that of *T. pumila*. Although the teeth are similar, they all average significantly larger in *T. tomensis*, especially the canines (Fig. 2b) and upper first premolars (minuscule in *T. gallagheri*). Because the usual pattern of sexual dimorphism in *Tadarida* is one in which males typically exhibit greater body size, larger skulls, and more powerful teeth (Peterson, 1969, 1970, 1972), we argue that *T. gallagheri* and *T. tomensis* are not conspecific. *T. tomensis* clearly differs from the other similarly sized molossid on the island, *T. pumila*, because it has a different pattern of color in body pelage and because it has conspicuous differences in the interaural membrane and skull morphology.

Remarks. - The specimens from São Tomé Island show the unique interaural membrane that Dobson (1878) used to define the subgenus *Chaerephon*. They also share other features typical of the group such as an unreduced upper last molar, well-developed antitragal lobule, a relationship between the second phalanx and the total length of the fourth digit around 14 - 17 % (17.3 % in the holotype, 16.4 and 15.4 % in the paratypes of *T. tomensis*); a range assigned to *Chaerephon* by Freeman (1981), and finally, all dental attributes established by Legendre (1984). Indeed, on a supraspecific level, the specimens of São Tomé Island belong to *Tadarida* (*Chaerephon*). Nevertheless, at this level, and supported by the appearance of *T. tomensis*, we could restrict the definition of the subgenus *Chaerephon* to the original character state defined by Dobson (1878). This would reduce the taxon to include only the species *johorensis*, *gallagheri*, and *tomensis*. However, the presence of a variably complex interaural membrane in other related species, as in *T. major*, makes it difficult to establish clear boundaries and therefore the restriction of *Tadarida* (*Chaerephon*) requires more detailed studies. The similarities of the structure of the interaural membrane clearly indicate a close phylogenetic relationship among *johorensis*, *gallagheri*, and *tomensis*. Such a relationship, however, poses an interesting biogeographic dilemma: how does one reconcile the geographic gap of > 2,000 km between the two African species within the same forest block, and between these and *T. johorensis* from Sumatra and the Malayan Peninsula (Hill, 1961, 1974)? This great second gap could be explained by an early/middle Miocene forest corridor connecting these two areas (K. F. Koopman, pers. comm.). Among African species, the simpler structure of the interaural membrane and the less marked inflations in the cranial rostrum suggest the presence of less derived features in the insular *T. tomensis*, in contrast to *T. gallagheri* (D. L. Harrison, pers. comm.).

We expect that males of *T. tomensis* will be larger in size, and will show the same interaural structure described here for females. The ear formation is similar in both sexes of *T. johorensis* (Hill, 1974).

The discovery of this molossid increases to five the number of families of bats known for São Tomé Island; the emballonurid *Taphozous mauritanus* also was captured for the first time on the island. The known bat fauna of São Tomé Island now includes *Eidolon helvum*, *Rousettus aegyptiacus*, *Myonycteris brachycephala*, *Taphozous mauritanus*, *Hipposideros commersoni*, *Hipposideros ruber*, *Miniopterus minor*, *Tadarida pumila*, and *Tadarida tomensis*.

Tadarida [(Xiphonycteris)] petersoni El Rayah, 1981

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Holotype

Adult male in alcohol with skull extracted (ROM 55813) from 15 km S Kumba, Cameroun (04°39'N 09°26'E), collected 27 January 1970, by R. L. Peterson (Field No. 0364).

Specimens Examined (Holotype and 36 Paratypes)

CAMEROUN: Holotype and 18 topotypes (ROM MM 55748, 55752, 55753, 55812, 55813, 55814; FF 55780, 55746, 55747, 55749, 55754, 55755, 55756, 55815, 55816, 55817, 55818, 55819); Mawutu Village, 30 km N Buea Station (03°10'N 12°20'E) 6 (ROM FF 69096, 69097, 69098, 69100, 69102, 69104); 6 km SE Eseka (03°35'N 10°44'E) 3 (AMNH M 236332, F 236333), (CM F 42293); 30 km E Nanga Emboko (04°38'N 12°21'E) 2 (AMNH M 241076, F 241078); near Okola (04°00'N 11°33'E) 1 (AMNH M 241075). GHANA: 12 km NE Kade (06°05'N 00°50'W) 6 (ROM MM 36563, 36564, 36600, 39427, FF 36601, 39380).

Distribution

Known only from the above localities in Cameroun and Ghana.

Diagnosis

A small member of the subgenus *Xiphonycteris* as defined by Koopman (1975) with open anterior palate and reduced third commissure, similar in skull size to *Tadarida nanula* and *T. spurrelli* but with a short distinct third commissure and deeper arched cranium. Wing bones longer than either *T. nanula* or *T. spurrelli* but shorter than *T. brachyptera* with which it is allied in overall shape characteristics.

Measurements of the Holotype

Total length, 93 mm; tail length, 27 mm; hind foot, 9 mm; ear, 19 mm; tibia, 13.3 mm (for other measurements see Tables 1 and 2).

Description

Muzzle short with vertically wrinkled thick upper lips (Fig. 1). Ears medium in size and joined on the muzzle by a deeply cleft band; tragus small, rectangular (about 1 mm wide and 1.8 mm high); antitragus somewhat rectangular (about 4.3 mm long and 3.3 mm high).

Feet small (about 9.0 mm), sparsely covered with hair to tip of toes. Calcar short (about 6.5 mm). Wing membrane insertion about 3.5 mm above the ankle at the base of the metatarsus. Dorsal pelage uniform Prout's Brown and extending to the sides of the neck immediately ventral to the antitragus. Base of dorsal hairs pale brown, darker distally. Ventrally, fur colour varies from Pale Ochraceous-Buff on chin and throat to light Pinkish Cinnamon on chest and abdomen. Pelage Cinnamon-Drab ventrolaterally along the insertion of the endopatagium to the base of the humerus and femur. Ventral pelage with white basal band, darker distally. Dorsally hair short (about 2 mm), ventrally slightly longer only on throat (about 4 mm). Hair extends to the endopatagium above and below. Posterolateral edge of uropatagium sparsely fringed with long bristles (about 6.5 mm).

Skull (Figs. 2 and 3) with first upper premolar small, in line with toothrow and filling the space between canine and second premolar; upper incisors vertical, parallel, separated from canines by pre-canine diastema (about 0.6 mm). Anterior palatal emargination extends posterior to upper incisors (covered by the soft palate in the holotype). Soft palate transversed by five palatal ridges. Third commissure on the last upper molar (M^3) reduced, much shorter than second (Fig. 3). Lower incisors small, bifid, and filling the space between canines. First lower premolar as high as second but wider at the base. Basisphenoid pits similar in shape to those of *T. brachyptera*, well defined, and oval in shape with a dividing septum, that is wide posteriorly and gradually narrowing anteriorly.

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Comparisons

Differs from *Tadarida nanula* in having longer and wider wings and skull; septum separating basisphenoid pits small and more triangular but pits larger; smaller canines; narrower postorbital constriction and larger braincase (Table 1). Differs from *T. spurrelli* (forearm 26.6-30.0) in having longer and wider wings and skull; upper and lower canines without enlarged cingula; higher braincase and longer mandible. Differs from *T. brachyptera* (=leoni) in having smaller wings and skull; shorter upper and lower canines; shorter mandible and the absence of well-developed sagittal or lambdoidal crests (Fig. 3).

Taphozous Dobsoni Jentink, 1879

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First phalanx of middle finger folded (in repose) on the upper surface of the metacarpal bone. Radio-metacarpal pouch well developed. Gular sac large in the male; as we have received no female, we cannot state if she too possesses a similar organ.

Ears shorter than the head; inner margin not papillate. Lower lip with a distinct groove.

Wings from the ankles. Interfemoral membrane greatly developed.

Under surface of the base of the thumbs with small fleshy pads. There is also a little rounded cushion near the base of the under surface of the footsoles.

The face in front of the eyes is covered with short blackish brown hairs.

General color as in *Taphozous mauritanus*: fur above buff brown near the base, then brown with grey extremities. Chest and abdomen pure white. Hairs round gular sac colored as the sides of throat which is embellished with a chestnut collar.

On the upper surface the wing-membrane as far outwards as a line drawn from the ankle and the thumb to the elbow and also the hairy antehumeral and interfemoral membranes are brown, the hairs being colored as on the back: the wing membranes for the rest white, except a brown patch inside the first phalanx of the longest finger: beneath all the hairs and membranes are white, except the brown-colored interfemoral membrane.

Lower incisors distinctly trifold.

Length of the only specimen, an adult male, preserved in alcohol.

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Head and body: 97

Tail: 22

Free end of tail: 12

Ear: 13

Tragus: 6

Forearm: 62

Thumb: 7

Third finger-metacarp: 62

" " 1st phalanx: 22

" " 2nd phalanx: 23

Fourth finger-metacarp: 47

" " 1st phalanx: 12.5

" " 2nd phalanx: 8

Fifth finger-metacarp: 36.5

" " 1st phalanx: 12

" " 2nd phalanx: 10

Tibia: 26

Foot and claws: 10

Calcaneum: 17.5

Hab : Madagascar, Mahambo (J. Audebert).

This species is the largest among the other hitherto known African species of the genus *Taphozous*, which are provided with a distinct radio-metacarpal pouch.

[*Taphozous hamiltoni*](#) Thomas, 1920

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A fairly large species of the group with a naked gular patch in the female - a pouch therefore probably present in the male.

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Size rather smaller than in *hildegardæ*, decidedly larger than in *sudani*. General external appearance as to colour and distribution of fur much about as in *perforatus* and its allies. Fur covering, but restricted to, the body, short; hairs of back barely 3 mm. in length. Colour above dark sepia-brown, the extreme tips of the hairs lighter, their bases white; below similar, but paler, the light tips being longer. Throat with a sharply defined naked patch, no doubt indicating that the male has a gular pouch.

Skull broad and stout, much more heavily built than that of *T. sudani*, and approaching that of the large *T. nudiventris*, though its muzzle is conspicuously shorter than in that animal and is without the great projection forward of the incisors. Forehead broad and flat, little hollowed out, the rise of the brain-case behind it not nearly so great as in processes well developed, short. Brain-case broad, more parallel-sided, less oval, than in *sudani*. Mesopterygoid fossa penetrating the palate to the level of the hinder edge of m^2 . Basial pits broadly triangular, not very deep.

Teeth as usual, rather stout and heavy throughout, breadth across canines greater than in other species of the same size.

Dimensions of the type: -

Forearm (c.) 66 mm.

Head and body 80; tail 35; third metacarpal 60.

Skull: condyl to front of canines 22; zygomatic breadth 15; interorbital breadth 7.3; intertemporal breadth 5; breadth of brain-case 11.2; mastoid breadth 13; palato-sinual length 6.2; postpalatal length 11.2; basial pits, length 3, combined breadth 5. Teeth: front of canine to back of m^3 9.7; front of p^4 to back of m^2 6.5.

Hab. Mongalla, Sudan.

Type. Adult female. B.M. no. 19.12.18.1. No. 118 of Stevenson Hamilton collection. Collected 13th June, 1918. Presented to the National Museum by Wellcome Research Laboratories.

It is difficult to say to which of the older-known species this *Taphozous* is most nearly allied. Its skull is much stouter than that of *perforatus*, *sudani*, and their allies, while, of course, the widely different fur-distribution of *nudiventris* and the peculiar colour of *mauritanus* at once separate those forms from it. *T. hildegardæ* has a much narrower and

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more slender skull, and no naked gular patch in the female. It is to be hoped that male specimens will shortly be obtained, so that the pouch-structure in that sex can be observed.

Major Stevenson Hamilton states that the specimen was captured in the verandah of his house.

Taphozous hildegardeæ Thomas, 1909

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Taphozous hildegardeæ, sp. n.

An African representative of the Asiatic black-bearded *T. melanopogon*.

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Radio-metacarpal pouch distinct. Lower lip scarcely grooved. No gular sac in male or female; a well-marked blackish beard in the male, covering the whole under surface of the throat. Upper surface pale brown, the bases of the hairs white, their tips chocolate-brown. Lips and chin anterior to the beard pale brown. Belly white, a few hairs tipped with brown. Limbs pale brown. Upperside of antibrachial, interfemoral, and wing-membranes near the body also brown, the more distal part of wing-membrane whiter; underside of all membranes white, as is the fur on the membrane on each side of the body. Ears, feet, and tail about as in *T. melanopogon*.

Skull as in *T. melanopogon*, but rather larger, and the brain-case broader.

Dimensions of the type (measured on the spirit-specimen): -

Forearm 70 mm. (another M 67.5, a F 65.5).

Head and body 83; tail 22; ear 20; third finger, metacarpal 61, first phalanx 21.5, second phalanx 24; lower leg and foot (c. u.) 37.5.

Skull: greatest length 21.7; front of canine to back of m³ 9.2.

Hab. Mombasa district. Type from Rabai, 700'; others from Shimoni, sea-level.

Type. Adult male. B. M. no. 9.6.12.7. Original number 613. Collected and presented by Dr. and Mrs. Hinde. Four specimens.

This is a most interesting species, as it represents in Africa the Asiatic black-bearded *T. melanopogon*, not known west of India proper. From this it is distinguished by its larger size, the greater extension over the throat of the black beard (more as in *T. theobaldi*), and its broader skull. No known African species are at all like it.

I have named the species in honour of Mrs. Hinde, who has so ably assisted in her husband's East-African collecting-work, and who has taken a special interest in bats.

Taphozous leucopterus Temminck, 1835

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TAPHOZOUS LEUCOPTERUS.

TAPHIEN LEUCOPTERE.

Planche 60. fig. 7.

Taille intermédiaire entre le *Taphien à ventre nu* et le *Taphien perforé*. Museau glabre; base des oreilles poilues; partie de l'antibrachium, membrane inter-brachiale et moitié de la partie supérieure de l'interfémorale très poilues; le reste couvert de poils ras et rares; osselets du calcaneum très longs; queue perforant la membrane vers le milieu de sa longueur; une grande partie de la membrane des ailes d'un blanc pur. Oreilles à peu-près rondes, tragus en forme de large fer-de-hache. Dents incisives supérieures nulles, 4 très petites et trilobées à la mâchoire inférieure: canines à talon pointu dirigé en avant; molaires 5 en haut, dont une, large et très courte anormale; l'arrière molaire de moitié moins large que sa voisine: en bas 5 molaires dont 4 parfaites et une petite anormale. La denture du jeune-âge n'a pas été examinée.

Pelage court, bien fourni, tricolore aux parties supérieures et sur les côtés du cou, unicolore en dessous. Tout le dessus du corps et les côtés du cou ont une teinte cendrée fourrée de petit-gris; les poils sont d'un brun clair à leur base, noirâtres au milieu et terminés de gris-clair; tout le dessous du corps, du cou et les rubans de poils le long des flancs et de l'antibrachium sont d'un blanc pur. La membrane alaire de l'antibrachium est en grande partie blanche, et celle entre les doigts de l'aile d'un gris-clair; l'interfémorale et la membrane qui s'étend le long des flancs sont noirâtres.

Longueur totale depuis la pointe du museau au bout de l'interfémorale 3 pouces 6 ou 7 lignes; osselet du calcaneum 8 lignes; envergure 12 pouces 4 ou 6 lignes; antibrachium 2 pouces ou 1 ligne de plus.

Cette espèce est nouvelle; elle a fait partie d'un envoi d'objets recueillis dans l'intérieur de l'Afrique méridionale; les mœurs n'ont pas été observées; elle fait partie du Musée des Pays-Bas.

Taphozous nudiventris Cretzschmar, 1830-1831

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Nackbäuchiger Grabflatterer

Diagnos. *Taphozous* facie usque ad regionem ophthalmicam, sumine, inguinibus, prymna et uropygio, nudis; corporis colore supra ex fusciscente griseo, infra albedo.

Ausmessungen.

Fuß. Zoll. Lin.

Länge von dem Scheitel bis zu der Schwanzspitze: - 4 3

Flügelweite: 1 4 6

Länge des Kopfes: - 1 -

Länge der Ohren: - - 9

Beschreibung.

Gesicht spitz zulaufend und von der Nase bis zu der Augengegend nackt. Lippen wulstig mit einzelnen Härchen besetzt. Unter dem Kinn ein kleiner Blindsack der durch eine Hautfalte entsteht. Der innere Rand des Ohres etwas ausgezackt, der Ohrdeckel kurz, zugerundet. Die Nasenlöcher klein, nach vornen gerichtet; die Augen mittelmäßig groß, schwarz von Farbe. Die Farbe auf dem oberen Theil des Körpers aus dem Lichtbraunen in das Weisgraue spielend, auf dem Unterkörper noch heller, ins Weißliche übergehend. Die Flug- und Interkrural-Membranen hellbraun; die letztere nach vornen aufgeschürzt, von dem Schwanzende durchbohrt. Der Steiß, die Schwanzgegend, der Unterbauch und die Schamgegend sind nackt und fleischfarbig. An dem Nagelglied jeder Zehe ein Büschel von steifen, gekrümmten Härchen.

Der hier beschriebene *Taph. nudiventris* unterscheidet sich von dem von Geoffroy in Egypten aufgefundenen *Taph. perforatus* nicht nur durch die Dimensionen seines Körpers *), indem er um ein Drittheil größer ist als der letztere, sonderit durch die Nacktheit aller Körpertheile, welche der hinteren und vorderen Seite der Beckengegend angehören und bei allen, sowohl alten als jungen Thieren bemerkt wird. Bei *Taph. perf.* Geoffr. sind dagegen alle diese Theile vollkommen behaart. Eben so ist die Färbung der Haare bei dieser Art durch eine oben dunklere unten hellere graue Farbe verschieden. Auch die Schädel beider Thiere sind sehr verschieden und wir können um so mehr eine zuverlässige Bestimmung darüber erlassen, da uns Rüppell aus verschiedenen Gegenden Egyptens viele alte und junge Individuen beider

*) Die Abbildung ist um 1/3 kleiner als die naturliche Größe.

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Arten eingesendet hat. Auf der Tafel 27 haben wir zur besseren Verdeutlichung unter No. 1.2.3. die obere und untere Fläche des Schädels und den Unterkiefer von *Taph. nudiventris*, so wie unter 4.5.6. dieselben Theile von *Taph. perforatus* (Geoffr.) abbilden lassen. An dem Oberkiefer des letzteren sind die Backenzähne so sehr abgenutzt, daß ihre Schmelzleisten ganz geebnet und nicht mehr zu erkennen sind, woraus wir auf das hohe Alter dieses Individuums zu schließen uns berechtigt glauben. Somit sind denn beide Arten durch Größe des Körpers, Farbe und Vertheilung der Behaarung, so wie durch den Schädelbau vollkommen verschieden und können unbezweifelt als Species gelten.

Vaterland. Egypten, Nubien.

Aufenthalt. Häufig in den Pyramiden von Gize.

Taphozous peli Temminck, 1853

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TAPHOZOUS PELI.

TAPHIEN DE PEL.

Pelage peu abondant, très-court et lustré; couvrant seulement le corps et la tête, et laissant complètement

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à nu toutes les parties postérieures, de même que tout le système cutané.

Le pelage du corps, du cou et de la tête est en dessus d'un marron vif et lustré, en dessous d'une teinte un peu plus claire et terne. Sous le menton qui est à nu, s'ouvre une poche, dans les replis de laquelle s'opère la sécrétion de la matière onctueuse. La face n'est couverte que de quelques poils rares et noirs. Les oreilles sont grandes; à leur base se forme un prolongement du derme, dirigé sur les joues; le tragus est grand, en fer-de-hache. La queue assez longue; perce la

membrane interfémorale vers le milieu de son étendue: toutes les autres parties du derme sont nues et noires. Les dents sont en tout semblables et en même nombre que dans les autres Taphiens; les quatre incisives inférieures sont trilobées.

Longueur de la pointe du museau au bout de l'interfémorale 6 pouces; de la queue 1 pouce 4 lignes, dont 6 lignes de libre; envergure 19 à 20 pouces; antibrachium 3 pouces 1 ou 2 lignes.

Patrie. Cette espèce nouvelle a été tuée sur la rivière de Boutry, côte de Guinée.

Taphozous Sudani Thomas, 1915

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T. perforatus group. No gular sac. Feet comparatively large.

Size slightly greater than in *T. perforatus*. Distribution of fur about as in that species, except that the longer hairs are less extended on the thighs and interfemoral membrane. General colour above and below sepia-brown, the hairs white for their basal two-thirds. Wing-membranes whitish or white beyond a line connecting the elbow with the knee; rest of the membranes brown, more whitish below. No gular sac in male or female, nor any trace of it on the skin. Feet decidedly larger than in *T. perforatus*, about 13 mm. in length or upwards, as compared with about 11 mm. in the allied species. Wing-membrane inserted on the end of the tibia, instead of the ankle.

Skull in general like that of *T. perforatus*, but slightly

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larger and the brain-case more swollen. Male skull decidedly larger than that of the female.

Dimensions of the type (measured on the spirit-specimen): -

Forearm 64 mm.

Head and body 71; tail 23; ear 17; third finger, metacarpal 58; first phalanx 21.5; lower leg and hind foot (c.u.) 39; hind foot only 13.5

Skull: greatest length to base of canine 20.2; condyle to front of canine 19.7; interorbital breadth 6.1; breadth of brain-case 10; palato-sinual length 6.5; maxillary tooththrow 8.8.

Hab. Upper Nile. Type from Mongalla, just north of Lado; other specimens from Khartoum (*Major H. N. Dunn*) and Lake No (*Willoughby Lowe*).

Type. Adult male. B.M. no. 2.7.4.2. Collected and presented by W. L. S. Loat, Esq.

This species is distinguishable from *T. perforatus* by its larger feet, more swollen brain-case, and whitened wing-membranes; from *T. hildegardeæ* by its smaller size, the absence of any gular beard, and generally darker colour.

Taphozous sudani australis Harrison, 1962

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As type specimen of this new form an adult male, no. 9932, is nominated. It was obtained on the 5 May 1960 at the junction of the Limpopo and Shashi Rivers, Southern Rhodesia. The specimen is now in the National Museum of Southern Rhodesia, Bulawayo.

DESCRIPTION OF THE TYPE

The specimen agrees in all essential characters with the typical form of the species. The gular sac is absent, nor is there any gular beard. The radio-metacarpal pouch is present. Hind feet large, 13 mm. in the dried skin, without claws. Wing membranes strongly whitish beyond a line connecting the elbow with the upper tibia. Skull decidedly lighter and

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more delicate than in the typical form, not exceeding that of *Taphozous perforatus* in size, although with the same relatively swollen braincase in comparison with the rostrum as in *T. s. sudani*. Hence in comparison with typical *T. s. sudani* the rostrum and interorbital region are very distinctly lighter and narrower and in relation to this the dentition also is more delicate, this being particularly noticeable in respect of the upper and lower canines. The external and cranial measurements are given in the tables below and it will be seen that the cranial measurements are distinctly smaller, the external measurements differing but little from the typical form.

[table removed - eds.]

The brown tips of the hairs on the dorsal surface are slightly darker than the specimens of the typical form examined (nearly matching Hudson Seal A9, Pl. 8 of Maerz & Paul, 1950) while the pale creamy buff hair bases where they are exposed over the shoulders contrast more sharply with the dark brown hair tips than in the typical form. On the ventral surface the pelage extends uniformly over the whole abdomen to the base of the tail, the distal part of the belly therefore not naked. On the throat and neck the fur is brown and little lighter than the dorsal surface, while over the chest and abdomen it becomes lighter and tinted with ashy grey (closely matching Slate - Gy + Kasha - Beige Oriental Pearl -, A2, Pl. 14 of Maerz & Paul). Some whitish hairs are found in the inguinal region. A band of ashy grey hairs extends along the proximal

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part of membrane from the axilla to the upper femur, while a narrow band of whitish hairs extends along the lateral border of the radius as far as the base of the fifth metacarpal. The ears, proximal wing membranes and whole interfemoral membrane are dusky, blackish, the latter rather lighter ventrally. The distal wing membranes are whitish beyond a line joining the elbows to the upper tibia, but the tips of the wings are again slightly dusky although translucent. The form of the ears and tragus does not differ significantly from the typical form. The upper canines are tall and slender, much more delicate than typical *T. s. sudani* with the antero-medial and postero-medial cusps both less prominent. The lower canines are similarly lighter but no other significant dental characters are observed in comparison either with *T. s. sudani* or with *T. p. perforatus*.

[table removed - eds.]

The paratypical series is in general closely similar to the type, although in one example (9928) the tips of the hairs on the dorsal surface are decidedly more rufous brown than the others, especially on the head, while the whole distal wing membrane in this specimen is strongly whitish. It has been previously noted (Harrison, 1958) that in the West African *T. p. swirae* Harrison the distal wing membrane is slightly paler in some specimens than is usual in *T. perforatus*, but in all the specimens of this series the wing membranes are far more strongly white than in any example of *T. perforatus* yet seen. The cranial measurement of the series average smaller than those of *T. s. sudani* but the lachrymal

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breadth and tooth-rows are constantly smaller, thus confirming the cranial characters noted in the type (see Table 6). The discovery of this bat so far to the south of its previously known range is of great interest and the constantly large hind feet and whitish wing membranes confirm the existence of two species of the *T. perforatus* group in Africa, both of which are now known to be widely but differently distributed. The small-footed and dusky winged *T. perforatus* extending from Egypt southwards to Kenya and Sudan, eastwards to southern Arabia, Cutch and Kathiawar in India and westwards at least to northern Nigeria, while the large-footed *T. sudani* with whitish wings is known from Sudan, Belgian Congo, Togoland (Hayman, 1954) (British Museum specimen) (*vide* Booth, 1956) and now by the newly described form from Southern Rhodesia.

Triænops aurita G. Grandidier, 1912

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Cette Chauve-Souris, qui appartient au genre *Triænops*, a été capturée par M. le D^r Mazières aux environs de Diego-Suarez, dans l'extrême Nord de Madagascar; des trois espèces de ce genre connues jusqu'à ce jour dans la grande île, deux vivent dans la forêt tropicale de la côte orientale, d'où M. Humblot a rapporté les premiers exemplaires, tandis que j'ai recueilli les individus types de la troisième dans la grotte de Sarondrano, à quelques kilomètres de Tulear, c'est-à-dire dans le Sud-Ouest de notre colonie. On voit donc que non seulement ce nouveau *Triænops*, comme nous allons le montrer, se distingue de ses congénères par des caractères anatomiques importants, mais aussi par son habitat.

Le *Triænops aurita* n'est représenté dans nos collections que par un unique exemplaire mâle adulte qui malheureusement a été plongé dans le formol, puis desséché accidentellement au cours du voyage, de sorte que, quoique son état de conservation extérieure soit très bon, il a été impossible de dégager le crâne de la peau. Néanmoins les caractères externes sont si nets qu'il n'y a aucun doute qu'il faille rapporter ce nouvel animal au genre *Triænops*. Ses appendices nasaux cutanés se composent d'un fer à cheval formé d'une seule feuille repliée en volute dans sa partie antérieure; au fond du fer à cheval, en arrière, se trouvent trois petites éminences que dominent trois lobes en fer de lance de longueur à peu près égale; sur

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l'éminence centrale se dresse une petite languette. Les fossettes sont situées, l'une grande et bien ouverte en avant et au pied des trois éminences, et les autres, latérales, à la base du trident.

Les oreilles sont proportionnellement très grandes pour une Chauve-Souris de ce genre; pointues à leur extrémité supérieure, très rapprochées sur le sommet du crâne, elles dominent toute la tête; elles sont plus longues que larges et à peine échancrées. A première vue, seules elles suffiraient à différencier le *Triænops aurita* des autres espèces du genre.

La queue est courte, ne dépasse pas la membrane qui forme un angle assez ouvert et non pas aigu comme chez le *Tr. Rufus* ou comme chez *Tr. Humbloti*. Son extrémité n'atteint pas les pieds, tandis que chez les autres *Triænops*, elle les dépasse beaucoup.

Le pelage est gris noir sur le dos (la base des poils est presque blanche, tandis que la pointe est fréquemment noire, surtout sur le dos et les reins); le ventre est gris sale. Les poils ne dépassent pas le corps proprement dit et n'empiètent ni sur la membrane claire ni sur l'interfémorale.

Le *Triaenops aurita* semble être jusqu'à présent la plus petite espèce du genre. Ses dimensions sont, en millimètres: tête et corps, 35; queue, 15; tête, 16; avant-bras, 45; pouce (ongle compris), 5; deuxième doigt, 38; troisième doigt, 60; oreilles: longueur, 16; largeur, 11; tibia, 20; pieds (ongles compris), 7.

La silhouette que nous donnons ci-dessus et qui représente la tête du *Triaenops aurita* au double de la grandeur naturelle n'a pour but que de montrer les proportions relatives de la face et des oreilles, et la disposition de ces dernières.

Triaenops persicus majusculus Aellen and Brosset, 1968

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TYPE. - M adulte, en alcool. Grotte de Doumboula, Loudima, République du Congo, 19.6.1964; coll. J.-P. Adam. Muséum de Paris 1968-412.

PARATYPES. - 5 MM ad., 5 FF ad. Mêmes lieu, date et coll. Que le type. Muséum de Paris.

4 MM ad., 2 FF ad. Mêmes lieu et date que le type, coll. Taufflieb (3403-1, 3403-4). Muséum de Genève 1074.41 à 46.

2 FF ad. Grotte de Meya-Nzouari, 29.11.1966, coll. Adam (503 A, 504 A). Muséum de Paris.

DIAGNOSE. - Semblable à *Triaenops persicus afer* Peters, mais plus grand: avant-bras 53.4 à 60.1 mm (moyenne 55.0 mm), crâne, longueur totale 18.9 à 21.2 mm (moyenne 20.4 mm).

DESCRIPTION. - Les spécimens de notre série ont été comparés à des *T. persicus afer* du Muséum de Paris, provenant de Zanzibar. Ils sont semblables, sauf pour les dimensions, qui sont plus grandes dans la nouvelle sous-espèce (cf. tableau ci-dessous).

Le caractère invoqué par DORST (1948: 18) pour séparer *afer* de *persicus*, soit le prolongement lancéolé de la selle bifide chez le premier, unicuspidé chez le second, ne nous paraît pas très valable, bien que nous n'ayons pas eu à notre disposition des *persicus* typiques. Dans notre série du Congo et sur les exemplaires de Zanzibar, cette lancette présente un bord supérieur très oblique, droit ou légèrement concave, ce qui détermine parfois un semblant de deux pointes. La variabilité de ce caractère n'avait d'ailleurs pas échappé à DOBSON (1879: 717), qui ne trouve aucune différence importante entre ces deux formes.

Il nous paraît superflu de reprendre ici une description détaillée de notre nouvelle forme, car nous ne ferions que répéter ce qu'ont dit plusieurs auteurs sur *persicus* sensu lato. Le lecteur pourra se référer aux descriptions originales et à celles de TATE (1941), de DORST (1948), de HARRISON (1955, 1963, 1964).

Actuellement, le statut taxonomique et la répartition géographique des formes du genre *Triaenops* sont les suivants:

Triaenops Dobson, 1871
furcula Trouessart, 1906: Madagascar
rufus Milne-Edwards, 1881: Madagascar
humbloti Milne-Edwards, 1881: Madagascar
persicus Dobson, 1871

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persicus Dobson, 1871: Perse
macdonaldi Harrison, 1955: Oman
afer Peters, 1877: Aden, ex-Somaliland, Kenya, Tanzanie, Zanzibar, Mozambique
majusculus, subsp. Nov.: Congo (Brazzaville)

Dans sa révision, DORST (1948) considère encore *afer* comme spécifiquement distinct de *persicus*. Mais, HARRISON (1964), qui est certainement l'auteur qui a examiné le plus de matériel de *persicus* s. l., admet que *afer* n'est qu'une sous-espèce de *persicus*, suivant en cela la notion ancienne de DOBSON (1879).

D'après les descriptions et mensurations publiées, les quatre sous-espèces aujourd'hui reconnues de *Triaenops persicus* se distinguent avant tout par leur taille:

[table removed - eds.]

Précisons encore que dans notre nouvelle sous-espèce la première prémolaire supérieure (P^2) est située dans la rangée dentaire, sur le bord externe, et sépare nettement la canine © de la deuxième prémolaire (P^4). Cela correspond à la figure de *T. afer* donnée par DORST (1948: fig. 2 C).

Il nous paraît intéressant, d'autre part, d'étudier l'os pénien de notre nouvelle sous-espèce, puisque celui de la forme la plus voisine, soit *T. persicus afer*, est déjà connu et peut servir de comparaison (MATTHEWS, 1941).

La forme générale est assez semblable, mais l'extrémité proximale est sagittée et non lancéolée. Cependant, la différence la plus remarquable est la taille: chez *p. afer*, le baculum mesure 2.8 mm, selon la figure 15 D de MATTHEWS; il n'atteint pas 2 mm chez *p. majusculus* (1.88 et 1.57 mm sur deux spécimens examinés).

REMARQUES. - La présence d'un *Triænops* au sud de la République du Congo est une donnée inattendue. En effet, le genre passait pour typiquement oriental. Cette notion classique est à revoir. En fait, ces chiroptères ont dû pousser vers l'ouest, à partir du Zanzibar et de la Tanzanie, et à travers le couloir écologique constitué par les lisières méridionales du Massif congolais, des populations don't

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[table removed - eds.]

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l'extrême pointe de répartition se situe près de l'Atlantique. On peut penser qu'une exploration plus complète des régions intermédiaires conduirait à la découverte de nouveaux jalons illustrant cette progression des *Triænops* vers l'occident. C'est du reste la forme qui peuple l'Afrique orientale qui est la plus proche de notre nouvelle sous-espèce.

[*Triænops rufus*](#) A. Milne-Edwards, 1881

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Le Muséum d'Histoire naturelle vient de recevoir de Madagascar une collection fort importante et riche en objets rares ou inconnus; elle lui a été offerte par M. Humblot, qui a surtout exploré, sur la côte est, la région comprise entre Fou.apointe te le lac l'Aincatre.

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Plusieurs Chauve-souris n'étaient pas connues; j'indiquerai d'abord deux espèces remarquable par la complication des appendices cutanés de la face et appartenant au genre *Triænops*; elles se distinguent du seul *Triænops* propre à l'Afrique (*Triænops afer* Peters) par la forme lancéolée et simple de la feuille centrale du nez et du *Triænops* de Perse (*Triænops persicus* Dob.) par la disposition des conques auditives, don't le bord interne est plus profondément échancré. L'un de ces *Triænops*, que j'ai désigné sous le nom de *T. rufus*, se reconnaît facilement à la couleur rousse de son pelage; l'autre (*Triænops Humbloti*) est un peu plus grand, et son poil est d'un gris qui rappelle à la teinte de nos *Rhinolophes*.

[*V\[espertilio\] acetabulosus*](#) Hermann, 1804

V[espertilio]. acetabulosus. Caudatus , fusco-niger , pilis ad extremos plantarum digitos longissimis, membrana acetabulosa nuda ad sternum. Commers. mscr. n.° 51.

La petite Chauve-souris du Port Louis. Magnitudo europaeorum. Volatus amplitudo pollicum decem.

Dentes in maxilla superiore duo parvi anteriores, disjuncti. Duo maximi etiam remoti et uncinati, seu introflexi. Molares quinque vel sex contigui.

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In maxilla inferiore dentes incisores quatuor, minimi, vix conspicui. Canini duo maximi inter maximos et minores maxillae superioris suscipiendi. Molares quinque vel sex.

Aures magnae, acutae, nudaе, basi tantum pilosae, apice nonnihil deflexo quasi falcatae.

Odor mellis recentis.

[*V\[espertilio\] mauritanus*](#) Hermann, 1804

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Commerson in *Fauna Borbonica* manuscripta sequentes vespertiliones novos describit:

V[espertilio]. mauritanus. Ecaudatus , capite dorsique lateribus rufis, collo, dorso et corpore subtus saturate ferrugineis, aoribus simplicissimis , palmis biunguiculatis. Commers. mscr. n.° 42.

La grande Chauve-souris de Maurice. Magnitudo eadem quae Vespertilionis insulae Bonro. (cujus descriptio autem non occurrit.) Dentes iidem.

Habitat in sylvis. Carnis sapor exquisitus, leporinae accedens.

Vespertilio Rüppelii J.B. Fischer, 1829

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V. Rüppelii; corpore supra ex cinereo fuscato, infra albo. *V. Temminckii* (Mus. Francof.) Cretzschmar in Rüpp. zool. Atl. p. 17. t. 6. ?.

Longit. corporis (cum capite) 1 poll. 10 lin., capitis 6 lin., auriculatum $3\frac{1}{2}$ lin., hallucis 2 lin., caudae 1 poll. 2 lin.; alarum extensio 7 poll. Caput valde pilosum, fasciculus pilorum ultra rhinarium fuscescens 1 lin. longum et 1 lin. latum prominet; pinnae nasi validae, torosae, nares sub-laterales; auriculae rotundae postice nudaе, antice pilo tenero vestitae; tragus foliiformis, apice latior et rotundatus; alae fuscae utrobique nudaе; patagium anale postice pilosiusculum. ? et pulli nullo modo diversi. Cretzschm. l.c. In sylvestribus et, fruticosis Dongolæ. Rüppel.

Vespertilio tricolor Temminck, 1832

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ADDITAMENTUM AD Genus VESPERTILIONUM,
quod memorabatur supra pag. 7 seq.
Species V. TRICOLOR. (Nova Species.)

Magnitudo novae hujus speciei tertiâ parte minor est, quam ea VESPERTILIONIS SEROTINE. Auriculae longae sunt, obtusis ac reflexis acuminibus, in anticâ parte emarginatae.

Tragus longus est, falcatus et introrsum flexus. Rostrum breve ac tumidum. Dentes $2\frac{1}{2}$ acuminati sunt in maxillâ superiore; in inferiore sex, tribus lobis instructi, adsunt.

Pili sunt laeves, in superioribus partibus minus longi ac triplicis coloris; scilicet brunneo-nigri ad basin, luteo-albi in media parte, atque ad apicem rubri.

Inferior corporis pars duplicem speciem offert; nimirum fuscâ ad basin: at in reliquâ portione exstat color luteo-albus; latera colli ac pectus colore lucide rufo tinguntur.

Membranae sunt pallide fuscae.

Universa longitudo est trium pollicum ac decem linearum.

Alarum vero extensio aequat duodecim pollices.

Antibrachium pollicem unum cum decem lineis longum est.

Vansonia Roberts, 1946

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Vansonia n.g., genotype *Pipistrellus vernayi* Roberts

Ann. Transv. Mus. XV, 16; 1932: Tamalakane River, Ngamiland.

This genus differs from *Pipistrellus* and related genera, in which the cranium is slightly raised above the level of the muzzle and with the same number of teeth, in having the inner of the two upper incisors deeply bifurcate, while the outer upper incisor is much reduced in size; the anterior upper premolar, though small, lies within the tooth row and separates the canine and large premolar. Presumably this name displaces *Alobus* Peters, genotype *Vespertilio temminckii* (1867) (which I have not examined), which is preoccupied by *Alobus* Leconte (1856).

Vespertilio Bocagii Peters, 1870

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Vespertilio Bocagii. Nov. sp.

V. rufus, subtus pallescens, pilis basi nigris; auriculis externe emarginatis; trago elongato, antice recto, postice convexo; patagio ad digitorum basin extenso; cauda âpice libera; patagio nudo; incisivi inferiores ad directionem marginis mandibulae positi; âpice incisivi superioris primi postice dente secundo paullo brevior.

Long. tota 0^m,0900
" capitis 0^m,0170
" auris 0^m,0130
" " marg. ant. 0^m,0105

Lat. auris 0^m,0075
 Long. tragi 0^m,0055
 " caudae 0^m,0400
 " humeris 0^m,0230
 ". antibrachii 0^m,0330

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" digit 1. (mtc. 0,0020; ph. 1. 0,0035; ph. 2. 0,0020) 0^m,0080
 " " 2. (" 0,0340; " 0,0027) 0^m,0370
 " " 3. (" 0,0340; " 0,0150; ph. 2. 0,0110; cart. 0,0065) ... 0^m,0665
 " " 4. (" 0,0320; " 0,0105; " 0,0080; " 0,0020) ... 0^m,0525
 " " 5. (" 0,0330; " 0,0100; " 0,0060; " 0,0030) ... 0^m,0520
 " fem 0^m,0140
 " tibiae 0^m,0175
 " plantae 0^m,0100
 " cale 0^m,0160

Duque de Bragança. (Anchieta)

[*Vespertilio borbonicus*](#) E. Geoffroy Saint-Hilaire, 1803

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7. Esp. Le vespert. de l'île de Bourbon. *Vespertilio borbonicus*. (Geoff.)

Caract. Orecchie corte, ovali; orecchio in semi-coro; pelo marrone in sopra, biancastro in sotto.

Especie inedita.

Descript. T. 0,008 (2 p. 11 l.). Questa specie si appropinqua molto della Noctule, dalla quale differisce però per l'orecchio più lungo e in forma di semi-coro, e per i colori del ventre: i peli sono d'un tinte uniforme in tutta la loro lunghezza.

Patria. L'île de Bourbon.

N°. CXXII. Individuo n'ayant que deux incisives à la mâchoire supérieure.

N°. CXXIII. Cet autre individu n'a que trois incisives, ce qui prouve que les deux autres manquent de même dans l'individu précédent: ces deux individus nous ont été envoyé par Macé, naturaliste.

[*Vespertilio Capaccinii*](#) Bonaparte, 1837

Vespertilio capaccinii

VESPERTILIONE DEL CAPACCINI

VESPERTILIO auriculis laterialibus, capite tertio brevioribus, ovato-lanceolatis, margine integris; trago brevi, gracillimo: pedibus robustis a patagio anali angusto villosissimo valde excedentibus: vellere griseo-rufescenti, subtus cinereo-flavido.

Si astenero dalle orgie di Bacco le tre Miniadi sorelle, e sdegnarone perfino di credere che concepito da Semele compiesse quel favoloso Nume i suoi nove mesi dentro un femore del padre Giove. Nemiche di tai crapule e di tai ciance, quel tempo che le altre femmine spendevano in feste ed in sì poco ragionevoli riti, consumavano esse nelle faccende domestiche e nelle arti di Minerva:

Solae Minaeides intus
 Intempestiva turbantes festa Minerva,
 Aut ducunt lanas, aut stamina pollice versant,
 Aut haerent telae, famulasque laboribus urgent.

Ma siccome tutto in que'giorni era Bacco, e della sua onnipotenza non si dovea dubitare, dicesi che ammonite invano da' prodigi raccontati da Eliano nella sua storia varia, fossero finalmente colpite dallo sdegno divino, e in brutti Vespertilioni tramutate, traendo con una morale soverchiamente naturale, che le odiatrici di tanta luce giustamente si condannavano ad assumere le mostruose forme descritteci dal Sulmonese:

Lucemque perosae
 Nocte volant, seroque trahunt a vespere nomen.

I diversi tempi però, i diversi costumi, le utilità conosciute delle arti, che ancor diciam di Minerva, massime a rimpetto della dannosa licenza de'Bacchanali, assai diversa ci additano l'applicazione della favola; ed invece di scorgere in questo animale gli effetti della miscredenza e della empietà, siamo piuttosto condotti da miglior senno a deplorare la poetica finzione che a lai volatili apparentemente brutti, messaggieri e compagni della tristezza notturna, assegnasse una origine da evitarsi, ch'è quanto

dire si spendessero ad accreditare il pazzo culto di Bacco. Ad oggetto pertanto di rivendicar la memoria delle buone e studiose Miniadi, più che all'uomo di stato vogliam raccomandarla al filosofo che in mezzo a gravissimi affari coltiva le già familiari scienze naturali, cui tutto modestamente amerebbe in pace di dedicarsi. Ond' è che intitoliamo a Monsignor Francesco Capaccini, il quale degna del suo favore la nostra Iconografia, questa nuova specie di Vespertilione a' suoi meriti non del tutto disacconcia, sì per lo amore sviscerato che alberga nel cuore per le arti di Minerva già da lui grandemente protette, sì per la ritiratezza, assiduità, vigilanza, cui fin da giovinetto consecratosi ad onore del sacerdozio, sempre vieppiù si dedica coll'adempiere scrupolosamente gli officii dell' altissimo ministero senza curarne lo abbagliante splendore.

Pervenutoci dalla poco esplorata Sicilia ha questo Vespertilione il trago assai più gracile e più sottile dello stesso *V. emarginatus*. La grandezza del pollice alare, la esimia robustezza de' piedi pochissimo inceppati dalla membrana interfemorale piena, tenace, assai pelosa, e scarsamente estesa, son poi caratteri tutti suoi proprj, mercè de' quali ad altri gruppi si approssima, che non sono di questa parte di mondo.

Il suo muso è molto erto e grosso, sporgente al di là delle orecchie quanta proprio è la distanza che corre tra l'una e l'altra: il naso ottusissimo offre larga ma superficiale depressione tra le narici: gli occhi grandicelli e molto infossati son quasi tre volte più vicini all' orecchio che alla punta del muso, e tanto distanti tra loro quanto dalla sudetta punta: la bocca è pochissimo fessa, non giungendo all'angolo anteriore dell' occhio; i denti sono piccoli e sottili: le orecchiette collocale molto in basso sui lati del capo sotto la linea degli occhi, son lunghe una volta e mezza appena di quanto son larghe, e due terzi al più della lunghezza del capo; la forma loro è lanceolato-ovale, e non mostrano intaccatura, ma un semplice dolcissimo seno sotto la metà del lembo esterno; il trago che non arriva a mezz' orecchia è retto, ed oltre modo sottile: le estremità del muso e le labbra sono scarsamente coperte di peli, che però folleggiano moltissimo sulla fronte, e nello spazio interoculare: mostrasi di sotto il mento una grossa glandula. Le bucce, alte. circa la metà di lor lunghezza, se chiudonsi addosso al corpo oltrepassano di poco l'estremità del muso. La membrana interfemorale assai tarpata obliquamente di qua e di là, riesce a forma di triangolo acuto che lascia molto liberi i piedi, orlando strettamente la coda, della quale lascia sporgere una puntarella sottilissima; spargesi di molti peli dalla base fino alla metà, non meno sopra che sotto, alcuni dei quali le fanno ciglio io prossimità dei piedi: pelosissima è parimente la membrana delle ali lungo i fianchi, particolarmente al di sotto. La coda uguale all' antibraccio è poco più breve del corpo. Le cinque dita dei piedi ugualissime fra loro portano lunghi peluzzi bianchi, e son terminate da unghiette bianche, robuste, molto uncinato, acutissime. Il pelame è molle e folto, di color cannellino chiaro che verge alquanto in rossastro, inferiormente cinereo-giallastro, essendo i peli di sopra cinerei dalla base fino alla metà, e di un bel castagno cupo fino ai due terzi nelle parti inferiori. Le bucce, la membrana interfemorale e le stesse orecchie rosseggiano alquanto invece di tendere più o meno al filigine come in altre specie. Il cranio è similissimo per le forme e le proporzioni a quello del *V. emarginatus*, essendo lungo quasi due volte quanto la propria altezza, con l'encefalo assai rigonfio e senza cresta occipitale.

poll. lin.

Lunghezza totale dalla punta del muso all'apice della coda: 3 2

" del capo: " 8

" delle orecchie: " 5 1/2

" della coda: 1 6

Estensione delle bucce: 10 "

Altezza delle medesime: 2 "

Lunghezza del braccio: " 11

" dell'antibraccio: 1 6

" del pollice: " 3

" della coscia: " 6

" della gamba: " 8

" del piede: "

Vespertilio Capensis A. Smith, 1829

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Vespertilio Capensis, mihi.

V. supra flavo-fuscus, subtus flavO-albus; membranis nigro-fuscis, interfemorali longè ultra pedes extensâ et posticè acuminatâ, ubi caudæ est terminatio; auriculis indentatis externè, et apicibus subacutis, trago lineari, subfalciformi.

Above yellowish brown, beneath yellowish white; membranes blackish brown, the interfemoral one extending considerably beyond the feet and pointed behind, where the tail terminates; auricles indented on their outer edges and with their tips sub-acute; tragus linear and falciform; head short, its sides and the muzzle bare and black, lips margined towards the angles of the mouth with strong brownish hair. Length of body, an inch and three-quarters; length of tail, three-quarters of an inch; expanse of wings, nine inches.

This species has a strong affinity to the *Vespertilio Temmickii* of Dr. Horsfield, and perhaps when the two are accurately compared together will be found to be the same, and the trifling apparent differences to be dependent upon changes which take place during the preparation of specimens. In the Cape example the posterior part of the interfemoral membrane is inclined to a semicircular form, whilst in that of Java it will be found to be more acuminate. Other slight differences are also observable, independent of those of colour and size.

Vespertilio d'asythrix Temminck, 1840

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VESPRTLIO D'ASYTHRIX.
VESPRTLION À POIL TOUFFU.

Ressemble plus ou moins au *vespertilion blépote* de l'archipel des Indes; mais il est moins grand, et ses oreilles quoiqu'aussi amples et presque totalement arrondies ont une conque plus développée et leur forme est en angle très ouvert.

Museau très obtus et large, garni d'un pelage abondant jusqu'à la pointe de muffle, où deux bouquets de poils raides sont disposés en moustache. Oreilles plus larges que hautes, orbiculaires, mais terminées en un angle très ouvert; à moitié poilues en dehors, un repli s'avance vers la commissure des lèvres; tragus court, en feuille arrondie. L'articulation du pied avec les métatarses totalement libre. Queue très longue, à membrane développée. Museau court; boîte cérébrale bombée. Dents incisives supérieures 4 par paire et d'égale grandeur; inférieures 6 petites et trilobées; molaires 5 en haut dont une fausse, et 6 en bas dont deux fausses.

Pelage de moyenne longueur, touffu, cotonneux et mat; face très poilue; base de l'interfémorale poilue seulement en dessus; membranes des flancs poilues à claire-voie en dessous seulement; pieds totalement libres; les membranes très amples, ne prennent attache qu'au dessus de l'articulation des métatarses. Couleurs des parties supérieures, de la tête, et des touffes mystacales d'un noir mat; celles des parties inférieures d'un noir enfumé, également mat; ruban de poils à claire voie, disposé le long des flancs, d'un noir grisâtre terne. Membranes d'un noir clair.

Longueur totale 3 pouces 9 lignes, dont la queue prend 1 pouce

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6 lignes; tendon fémoral du calcaneum 8 lignes; envergure 10 pouces; antibrachium 1 pouce 8 lignes.

Cette nouvelle espèce, remarquable par ses membranes développées, son large museau obtus, ses larges oreilles et son pelage laineux, a été rapportée de l'intérieur de la Caffrérie par Mr. Verreaux de Paris. Le sujet sur lequel cette description est prise, fait partie du musée des Pays-Bas.

On peut classer cette nouvelle espèce d'Afrique après le *vespertilion leucomèle* de cette monographie pag. 204.

Vespertilio Dinganii A. Smith, 1833

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Vespertilio Dinganii, Smith. (Dingan's Bat.) Fur long and silky; above, the surface colour is dull olive-green, beneath a pale greenish yellow; head short and broad; nostrils prominent; ears blackish brown, and slightly pointed; tragus long, falciform, and tapering towards the tip, which is rounded; auricular appendage semicircular, very distinct, and in a measure separated from the anterior and outer margin of the auricle, inside of the latter with a thin sprinkling of short olive-green hair; wing membranes black; interfemoral membrane pointed, with part of the last joint of the tail protruding beyond it; claws pale greenish yellow; lower incisors placed obliquely across the jaw. Length from nose to base of tail three inches and a half; length of tail two inches; expanse of wings about twelve inches. - Above the hairs are yellowish green, towards their bases below they are of one colour throughout.

Inhabits South Africa, - between Natal and Delagoa Bay.

Vespertilio goudotii A. Smith, 1834

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Vespertilio Goudotii, Smith. (Goudot's Bat.) Hair of moderate length, longer on the face, head, and upper parts of

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body, than on the lower parts. Head rather small; point of nose bare and black; nostrils round, and opening laterally; upper lip on each side behind nostrils thickly covered with long hair; ears as long as the head, slightly rounded at points, and each with a large deep notch on its outer edge; on the outer surface towards base covered with chesnut hair, on the inner surface toward inner edges with a thin sprinkling of short tawny hair; tragus long and tapering to a point. The surface colour of the upper parts of head and body light maroon or dull chesnut; of the under parts dull tawny or pale brown; the bases of the hairs above dusky, of those beneath dull black; membranes brown with a purplish tint; tail of the same length as the interfemoral membrane. Length from nose to base of interfemoral membrane two inches and a half; length of interfemoral membrane one inch and three quarters; expanse of wings nine inches.

Inhabits Madagascar.

Vespertilio hesperida Temminck, 1840

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VESPERTILIO HESPERIDA.
VESPERTILION HESPÉRIDE

Taille d'un tiers moins forte que la *pipistrelle* d'Europe; système cutané peu développé en raison du volume du corps; museau court, obtus; oreilles courtes, aussi larges que hautes, à tragus en feuille courbée et à pointe arrondie; membrane interfémorale veinée en losange, un peu poilue à sa base; les quatre membres nus. Dents incisives 4 par paire en haut et 6 en bas, petites et bien rangées: les molaires n'ont pas été observées.

Pelage court, lisse et bien fourni, de deux couleurs partout, en dessus noirâtre à la base et brun roussâtre à la pointe; en dessous noir à la base et cendré roussâtre à la pointe; la pointe du museau très poilue et noire; les membranes brunes veinées de brun plus clair.

Longueur totale du mâle adulte 2 pouces 8 lignes dont la queue prend seulement 1 pouce; antibrachium 1 pouce. La femelle a des dimensions moins fortes.

Synonymie. Pas indiquée dans l'atlas du voyage de Mr. Rüppell en Egypte.

Habite. Les bords de la Mer rouge vers les côtés d'Abyssinie; des sujets font partie du Musée de Francfort et des Pays-Bas.

Vespertilio hispidus Schreber, 1774

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Tab. LVI.

Campagnol volant. DAUB., *mém.* 1795, p. 388.
Autre chauvefouris. BUFF. *hist. nat.* 10. p.88. tab.20. f. 1.2.
Bearded bat. PENN. *fyn.* p. 367. n. 282.

Die Schnauze ist lang, unter der Stirne stark eingedrückt. Die Nase ist von einer besondern Struktur. Die Nasenlöcher, sagt Herr Daubenton, sind nicht, wie an andern Thieren, durch eine bis vor gehende Scheidewand von einander abgesondert; sondern jedes Nasenloch befindet sich an der Spitze einer kleinen Rinne, welche von einem Ende zum andern oben offen ist. Der innere Rand der Rinne ist sehr klein, der äussere stärker, und dehnt an seinem hintern Ende in eine Wust aus. Die äussern Ränder der beiden Rinnen vereinigen sich über der Oberlippe, und formiren durch diese Vereinigung den Anfang einer starken Furche, welche sich von da bis and die Stirne erstreckt, wo sich eine breite und kahle Vertiefung bedinget, deren Rand mit langem Haar besetzt ist. Die Ohren sind land und schmal. Das Haar auf dem Rücken von dem Scheitel an ist rothbraun; das übrige weißlich mit einem gelbbräunlichen,

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an andern auch mit einem aschgrauen Schatren schwach überlansen. Die Ohren und die Flughaut rötlich und schwartzlich braun. Der Schwanz liegt ganz in der Flughaut, und ist fast so lang as der Leib, desser Länge anderhalb Zoll beträgt.

Das Vaterland dieser Fledermaus is Senegal. Herr Adanson hat sie entdeckt.

Vespertilio Hottentota A. Smith, 1833

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Vespertilio Hottentota, Smith. (Hottentot Bat.) Above a dull indistinct dark brown, sometimes nearly black; beneath the surface colour is a sort of pale tawny brown or tawny white, lighter towards the pubis; he middle and basilar portions of the hair brownish black. Ears about half the length of the head, black and acutely rounded at their tips, the inner surface bare, the outer hairy towards the base; tragus linear tapering, and with a rounded point; muzzle covered with dense fur till near its point; wing membranes brownish black; interfemoral membrane with a sprinkling of fine short brownish fur for half its length above; claws pale flesh-coloured. Length from nose to base of tail two inches and a half, of tail two inches; expanse of wings twelve inches.

Inhabits South Africa, - Uitenhage and Albany.

Vespertilio isabellinus Temminck, 1835-1841

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VESPERTILIO ISABELLINUS.
VESPERTILION ISABELLE.

Planche 52 fig. 1 et 2.

Taille du *vespertilion discolor* d'Europe, mais les formes semblables à celles du *V. murin*. Museau obtus; oreilles larges ovoïdes, munies vers leur bord antérieur d'un repli longitudinal et saillant, qui forme comme la duplication du derme; un lobe distinct termine le bord postérieur et s'étend vers la commissure des lèvres. Tragus court en feuille; bout de la queue libre. Toutes les membranes nues, fortement veinées; le tendon du calcaneum portant un lobe externe. Dents incisives 4 par paire en haut, l'interne longue et forte, bilobée dans le jeune, pointue chez l'adulte; l'interne vers la canine très petite; 6 incisives en bas, trilobées dans le jeune âge, tranchantes chez l'adulte; molaires 4 en haut sans fausse; 5 en bas dont une petite fausse molaire pointue.

Pelage de moyenne longueur, fin, soyeux, peu fourni. Les parties supérieures dans l'adulte, d'une belle teinte isabelle; la pointe du museau et les lèvres noires; toutes les parties inférieures d'une teinte isabelle très claire; la base externe des oreilles couverte de poils. Les membranes d'un brun noirâtre et abondamment veinées de lignes jaunâtres. Une grande portion du bout de la queue libre.

Les jeunes ont le pelage court, peu fourni, d'un isabelle cendré, en dessous d'un blanchâtre légèrement teint de couleur isabelle.

L'adulte porte en longueur totale 4 pouces 2 ou 3 lignes, dont la queue prend 1 pouce 7 lignes; envergure 11 pouces 7 ou 8 li-

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gnes; antibrachium 1 pouce 8 lignes. - Le jeune de l'année porte 3 pouces 6 ou 7 lignes en longueur totale, 9 pouces 6 lignes d'envergure, antibrachium 1 pouce 6 lignes. - Le jeune moins avancé, n'a que 3 pouces en longueur sur 7 pouces d'envergure.

Cette espèce que je crois nouvelle, a été envoyée au musée des Pays-Bas, en un grand nombre d'exemplaires, par les soins de Mr. le consul Clifford van Breugel.

Habite l'Afrique septentrionale, vit en grand nombre dans les environs de Tripoli.

[*Vespertilio leucomelas*](#) Cretzschmar, 1826

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Zweifarbige Fledermaus.

Diagnes. *Vespertilio corporis colore supra nigro; infra ex nigro et albo variegato. Rhinarium prominens incisura ad frontem porrecta utrinque labro sejunctum. Auriculae medio frontis contiguae.*

Ausmessungen.

Fuß. Zoll. Lin.

Länge von dem Scheitel bis zu dem Schwanz: - 1 9

Länge de Schwanzes: - 1 3

Höhe der Ohren: - - 7

Länge des Kopfes: - - 6

Beschreibung.

Oberkörper, Kopf, Gesicht, Nase, Ohren, Hals und der obere Theil der Brust schwarz. Auf der Brust haben die Haare weiße Spitzen und werden am Unterleib ganz weiß, so daß an diesen Theilen eine aus Schwarz und Weiß vermischte Färbung entsteht. Die Nase erhebt sich wulstig über die Oberlippe, ist auf dem Rücken breit, und schmal an dem Lippenrand; so daß sie wie ein mit der Spitze nach unten gekehrtes Dreieck aussieht. Dieses entsteht durch zwei Einschnitte, welche den Nasenkörper gleichsam von der Oberlippe auf beiden Seiten trennen und nach der Stirne hin verlaufen. Die Nasenlöcher oben, auf dem breiten Theile der Nase. Hinter der Nase eine Vertiefung. Die Ohren sind am Rande dieser Vertiefung zusammengewachsen und zur Seite dieser Vertiefung liegen die Talgdrüsen. Der Ohrdeckel breit an der Wurzel, an der Spitze schmal auslaufend. Die Flug- und Intercrural-Membranen braun. Die Flughaut an ihrer Verbindung mit dem Körper mit feinen weißen Härchen besetzt.

Der ganzen Gestalt nach würde dieser Cheiropter mit unserem *Vespert. auritus* namentlich in der Ohren- und Nasenbildung übereinstimmen, und mit diesem eine Abtheilung der Gattung *Vespertilio* bilden. Die Zahnbildung ist aber verschieden, und verdient hier angeführt zu werden. Von den vier oberen Schneidezähnen sind die zwei mittleren oder inneren am längsten und an der Spitze zweizackig. Backenzähne haben wir oben nur vier bemerken können, selbst der erste steht ganz nahe an dem Eckzahn, weswegen wir Anstand nehmen den fünften als ausgefallen zu betrachten. Ist diese Zahnbildung constant, so würde sich dieser Cheiropter besser an die Gattung *Noctilio* anreihen.

Vaterland. Das petraeische Arabien

Aufenthalt. Alte verödete Wohnungen

Vespertilio marginatus Cretzschmar, 1830

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Saumrandige Fledermaus.

Diagnos. Vespertilio corporis colore supra rubiginoso, infra ex rubido cinereo. Patagium diilale ET intercrurale limbo flavescenti marginatum.

Ausmessungen.

Fuß. Zoll. Lin.

Länge von dem Scheitel his zu dern Schwanze: - 1 4

Länge des Schwanzes: - 1 1

Flügelweite: - 6 10

Länge des Kopfes: - - 6

Länge des Daumens: - - 2

Buschreibung.

Die saumrandige Fledermaus ist die kleinste von den Fledermäusen, welche Rüppell in dem nordöstlichen Afrika aufgefunden hat. - Die Stirne, der Scheitel, Nacken, Seiten des Halses und ganzer Oberkörper sind rothbraun von Farbe und alle diese Theile mit einem wolligen dichten Haar besetzt. Der ganze Unterkörper ist mit einem mehr länglichen glatten Haare bedeckt, welches von der Wurzel an schwärzlich grau ist und mit graubrauner Spitze endiget, wodurch dieser Theil eine etwas gemischte graubraune Farbe erhält. Die Nase ist in der Mitte gefurcht und die Nasenlöcher seitlich, beinahe trichterförmig hervorgetrieben. Die Augen klein, und über denselben so wie auf den Lippen einzelne schwarze Borstenhaare. Die Flug- und Intercrural-Membranen schwarzbraun und an dem unteren Rande mit einem gelblichen wulstigen Saum eingefaßt. Der Sporn-Knorpel stark, mit seiner Spitze nach außen gerichtet, welches der Intercrural-Membran eine winkelige Gestalt gibt; an seinem äußeren Rande mit einer blätterigen, fast knorpeligen Membran versehen, welche in der Nähe des Fußes breiter ist und schmal endiget. Der Daumen mit einem sehr kurzen stark gekrümmten Nagel versehen.

Vaterland. Nubien und das petraeische Arabien.

Aufenthalt. Wurde überall einzeln und selten gefunden, aber immer in der Nähe von Wäldern oder Gebüsch, welche sie zu bewohnen scheint.

Vespertilio megalurus Temminck, 1835-1841

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VESPERTILIO MEGALURUS

VESPERTILION MÉGALURE

Le corps grêle, avec une tête déprimée, le museau pointu et les narines rapprochées, mais les membranes du vol et celles de la queue très développées. Oreilles médiocres, distantes et pointues vers le bout; tragus très long, en forme de feuille de saule. Queue très ample par l'extension de sa membrane soutenue par des cartilages métatarsiens ou calcanés très longs, et par une queue très longue à grand bout libre. Chanfrein et botte déprimés. Dents incisives 4 par paire; celles internes longues et convergentes, les externes près de la canine à peine visibles; molaires 4 en haut sans dent anormale, en bas 5 molaires dont une très petite anormale.

Pelage long, lisse, soyeux, bicolore partout: en dessus noirâtre à la base et d'un brun olivâtre jusqu'à la pointe: en dessous brun à la base; les poils du devant du cou et du ventre d'un brun cendré jusqu'à la pointe, ceux des flancs d'une teinte isabelle, et la région pubinale toute blanche depuis la base des poils jusqu'à leur pointe. Flancs en dessous poilus; seulement un ruban poilu le long des flancs et autour du coccyx aux parties supérieures.

Longueur totale 4 pouces 3 lignes dont la queue prend à peu-près 2 pouces; envergure de 11 pouces à 11 pouces 6 lignes; antibrachium 1 pouce 10 lignes; dimensions de l'adulte; celles des jeunes ne sont pas connues.

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C'est probablement à cette espèce caractérisée par la large membrane interfémorale qu'il faudra rapporter le *Vespertilio capensis* très succinctement indiqué par Smith. Zool. Jour. part. 16; pag.435. Le sujet dont il est question a été comparé au *Vespertilio Temminckii* de Horsfield; ce dernier est un *Nycticee*. Je présume que le sujet de Mr. Smith est la même espèce que notre *mégalure*, mais dans un développement moins parfait et plus jeune que les nôtres.

Quelques sujets ont fait partie d'un envoi d'objets rassemblés, très récemment, dans l'intérieur de l'Afrique méridionale. On ne sait rien du genre de vie, ni sur les lieux qu'elle habite de préférence: elle fait partie du Musée des Pays-Bas.

Vespertilio minuta Temminck, 1840

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VESPERTILIO MINUTA.
VESPERTILION MINUTA.

Je n'ai eu connaissance de cette très petite espèce africaine, que longtemps après la publication de la dissertation zoologique sur les mammifères du Cap de Bonne Espérance, publiée par Mr. Smuts, ce qui est la cause que je n'ai pu faire placer l'indication dans l'opuscule mentionnée.

Elle est plus petite que notre *pipistrelle* d'Europe, dont elle a les formes totales. Les oreilles sont ovoïdes, aussi larges que hautes; le tragus en feuille, à pointe arrondie et moitié longueur de l'oreille. Le museau est nu, garni de quelques poils clair-semés. Le pelage est court, bicolore partout; en dessus noir à la base et la pointe des poils d'un brun foncé couleur de terre d'ombre; en dessous à base noire avec la pointe d'un brun-clair. Les membranes sont noires.

Dents 4 par paire en haut; l'incisive accolée aux canines, extrêmement petite, l'autre longue bifurquée; 6 en bas, entassées; molaires

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4 en haut et 5 en bas; les premières sans fausse molaire et les inférieures avec une fausse, très petite et pointue.

Longueur totale 2 pouces 6 ou 7 lignes; envergure 7 pouces; antibrachium 1 pouce 1 ligne.

Elle vit comme notre pipistrelle et habite les lieux boisés près des eaux dans plusieurs parties de la pointe méridionale de l'Afrique.

Le Musée des Pays-Bas a reçu quelques individus par les soins de Mr. Horstok, médecin au Cap de Bonne Espérance.

Vespertilio minutus somalicus Thomas, 1901

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Closely allied to the typical *V. minutus*, Temm., but distinguished by smaller size, at least of head, and much paler coloration.

General characters as in *V. minutus*, the description of that animal in Dobson's Catalogue agreeing in all details except those referring to colour, though it may be also noticed that the inner margin of the ear, instead of being "slightly convex to the tip," is flat or even faintly concave for its middle third.

General colour of fur above dull buffy, the hairs blackish slate for two thirds of their length, with their tips buffy; beneath the bases are equally dark, the tips being dull whitish. Hairs of back rather more than 8 millim. long. Wing-membranes smoky brown, interfemoral membrane paler brown, both prominently edged with whitish, recalling the white edging of *Pipistrellus Kuhlii* on the same parts.

Skull very small and delicate, smaller than South-African

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specimens; although the forearm measurement does not appear to be less. The head is therefore apparently rather smaller in proportion.

Dimensions of the type (measured in flesh): -

Forearm 31 millim.

Head and body 44; tail 85; ear 10.

Skull: greatest length 12.5; basipalatal length 9.6; zygomatic breadth 8; interorbital breadth 3.1; breadth of braincase 6.7.

Dimensions of a spirit-specimen (F): -

Forearm 30.

Head and body 42; tail 35; ear 11; tragus on inner edge 3.3; lower leg 13.

Hab. Somaliland. Type from Hargaisa, 3500 feet; other specimens from Berbera.

Type. B.M. no. 98.6.9.1. Collected 14th November, 1897, and presented by R. McD. Hawker, Esq. Other specimens presented by E. Lort Phillips, Esq.

This subspecies may be readily distinguished from the typical form by its far paler colour and prominently white-edged wing-membranes.

Vespertilio nanus Peters, 1852

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Tafel XVI. Fig. 2.

V. magnitudine pipistrelli; colore supra umbrinofusco, subtus pallidior; pilis in basi piceis; alis nigris; auriculis retro excisis; lobulo semicirculari, trago securiformi; rostro tumido, prominente; dentibus primoribus inferioribus secundum mendibulae directionem positis.

Longitudo tota 0,080; caudae 0,040; antibrachii 0,030; volatus 0,220.

Habitatio: Africa orientalis, Inhambane, 24° Lat. Aust.

Von der Gröfse des *Vespertilio pipistrellus*, Daubenton. Der hintere Rand des Ohres ist ausgeschnitten und endet in der Höhe der Mundspalte, $2\frac{1}{2}$ Mm. hinter dem Mundwinkel; das Ohr ist weniger breit als die Höhe des vorderen Randes; die Entfernung der vorderen Ohrränder von einander ist gröfser als die Entfernung derselben von der Schnauzenspitze; in der hintern Hälfte des Ohrs befinden sich fünf fächerförmig auseinander strahlende Querfalten; der Tragus ist am Ende breit, beilförmig, mit seiner Spitze nach innen gerichtet; der Antitragus halbkreisförmig, vorn eben so hoch wie hinten. Die Schnauze ist breit, stumpf und in der Mitte niedergedrückt; die Lippen sind wulstig, die Oberlippe an jeder Seite inwendig mit einem warzigen Vorsprunge versehen. Die Gaumenfalten haben dieselbe Zahl und Lage wie bei *V. pipistrellus*; es sind sieben, von denen die beiden ersten ungetheilt sind. Der fünfte Finger ragt über die erste Phalanx des dritten Fingers hervor, und die erste Phalanx des fünften Fingers reicht bis zum letzten Drittheil der ersten Phalanx des vierten Fingers. Die Flughäute sind bis zur Zehenwurzel angewachsen und längs dem Unterarm unbehaart. Die Schenkelflughaut ist dagegen auf den Querfalten der Bauchseite mit Haaren bewimpert.

Die Farbe des Kopfes und der Rückenseite ist umbrabraun, an der Bauchseite blasser. Sämmtliche Haare sind an den unteren zwei Drittheilen pechschwarz. Flughäute und Ohren sind schwarz, die Nägel schmutzig weifs.

Gebifs $3\frac{2}{3} 2\frac{1}{2} 1\frac{2}{6} 1\frac{2}{6} 1\frac{2}{6} 2\frac{3}{2} 3$. Die beiden oberen Vorderzähne stehen entfernt vom Eckzahn, sind gleich lang, und an der Basis von gleicher Dicke; der innere ist am Ende in zwei parallele, nicht divergirende, Spitzen getheilt; der äufsere zeigt hinten und oben einen spitzen Absatz. Die unteren Vorderzähne sind dreilappig und nach der Richtung des Kiefferrandes befestigt. Der erste kleine obere Lückenzahn ist ganz nach innen gedrängt. Die Wirbelsäule besteht aus 7 Halswirbeln, 12 Rückenwirbeln, 5 Lendenwirbeln, 4 Kreuzheirwirbeln und 9 Schwanzwirbeln. In dem Bau der Eingeweide zeigt sich die gröfste Uehereinsummung mit *V. pipistrellus*.

Ich erhielt diese Art in gröfser Anzahl aus unbenutzten Schornsteinen in Inhambane, im 24° S. Br.

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Mafse in Millimetern.

Fem.; Mas.

Ganze Länge von der Schnauze bis zur Schwanzspitze: 80; 80

Flugweite: 210; 220

Länge des Kopfes: 14; 14

Länge des ganzen Ohres: $11\frac{1}{2}$; 12

Breite desselben: 7; 7

Länge des vordern Ohrrandes: 9; $9\frac{1}{2}$

Entfernung der vorderen Ohrränder von einander: 7; 7

Entfernung der Schnauzenspitze vom vordern Ohrrande: 6; 6

Länge des Tragus am vordern Rande: $3\frac{1}{2}$; $3\frac{1}{2}$

Gröfste Breite des Tragus: 2; 2

Länge des Oberarms: 19; 20

Länge des Vorderarms: 30; $31\frac{1}{2}$

Länge des Daumens (Mittelh. 2, 2. 1.Gl. $2\frac{1}{4}$, $2\frac{1}{4}$. 2.Gl. $1\frac{1}{4}$, $1\frac{1}{4}$): $5\frac{1}{2}$; $5\frac{1}{2}$

Länge d. 2ten Fingers (Mittelh. 27, 28. 1.Gl. 2, 2): 29 30

Länge d. 3ten Fingers (Mittelh. 28, 29. 1.Gl. 11, $11\frac{1}{2}$. 2.Gl. $9\frac{1}{2}$, $9\frac{1}{2}$. 3.Gl. 6, 7): $54\frac{1}{2}$; 57

Länge d. 4ten Fingers (Mittelh. $27\frac{3}{4}$, $28\frac{3}{4}$. 1.Gl. $9\frac{1}{2}$, 10. 2.Gl. $6\frac{1}{2}$, 7. 3.Gl. $1\frac{1}{2}$, 2): $45\frac{1}{4}$; $49\frac{3}{4}$

Länge d. 5ten Fingers (Mittelh. 27, $28\frac{1}{2}$. 1.Gl. $7\frac{1}{2}$, 8. 2.Gl. 6, $6\frac{1}{2}$. 3.Gl. $2\frac{1}{4}$, 3): $43\frac{3}{4}$; $45\frac{1}{2}$ Länge des Oberschenkels: $12\frac{1}{2}$; 13

Länge des Unterschenkels: $12\frac{1}{2}$; 12

Länge des Fulses mit den Krallen: 7; 7

Länge des Schwanzes: 39; 40

Länge des Sporns: 14; 16

Länge des Schädels: 12; 12

Vespertilio minutus, Temminck, unterscheidet sich von dieser Art durch die Ohren, welche ebenso breit wie hoch sind, durch den schmalen, um hintern Rande convexen Tragus, durch die Kleinheit der nahe an die Eckzähne angedrängten obern äufsern Vorderzähne und durch die Kürze des fünften Fingers, welcher nur bis zur Mitte der ersten Phalanx des vierten Fingers reicht. *Vespertilio Temminckii*, Rüppell, ist schon wegen der Farbe der Bauchseite, deren Haare bis zur Basis weifs

sind, nicht mit ihr zu verwechseln. *Vespertilio marginatus*, Rüppell, läßt ebenso wenig eine Verwechselung mit derselben zu, da sowohl die Form der Ohren als die Färbung eine ganz andere ist. Bei *Vespertilio platycephalus*, Temminck, betrügt die Länge des Schwanzes nur ein Drittel der Totallänge, und der hintere Ohrrand bildet an seinem Ende hinter dem Mundwinkel einen breiten häutigen Anhang. *Vesperugo subtilis*, Sundevall, mit oblongen, ganzrandigen Ohren, kurzem, halbovalen Tragus und schmaler Schnauze ist ebenfalls eine durch ihre Größe verwandte Art.

[*Vespertilio Natalensis*](#) A. Smith, 1833

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Vespertilio Natalensis, Smith. (Natal Bat.) Fur soft and of moderate length, the surface colour above deep clear brown; beneath light brown, inclining to tawny brown; the baselar

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half of all the hairs clear black. Ears naked, broad towards base, and slightly emarginate on the outer side, the tips rounded; tragus narrow, straight, and rounded at the point; muzzle and upper lip black, with a thin sprinkling of short black hair, nostrils opening directly in front; no appearance of a cleft in the fore part of the upper lip; wing and interfemoral membranes black. Length from nose to base of tail two inches, of tail an inch and a quarter; expanse of wings eight inches and a half.

Inhabits South Africa, - towards Natal.

[*Vespertilio Nigrita*](#) Schreber, 1774

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Die spizöhrige Fledermaus. *Vespertilio Nigrita*

Tab. LVIII.

Marmotte volante. DAUBENTON mém. 1759. p. 385.
Chauve-souris étrangère. BUFF. hist. nat. 10. p.82. tab. 18.
Senegal bat. PENN. syn. p. 3669. n. 281.

Die Schnauze ist von mittelmässiger Länge, vorn dick, die Ohren dick und spizig, die Ohrendeckel schmal, fast halb so lang als die Ohren, und zugespizt. Der Schwanz um ein Viertel kürzer als der Leib, mit der Schwanzhaut umgeben, bis auf die zwei letzten Wirbelbeine, welche über den Rand derselben hinausragen. Das Haar hat auf dem Kopfe, Halse und Rücken eine gelblich braune mit etwas aschgrau melirte, auf der Brust und dem Bauchseite ganz bleich gelblich braune ins graue spielende Farbe. Die Schwanzhaut ist schwärzlich. Die Länge vier Zoll.

Sie ist in Senegal von Herrn Adason entdeckt worden.

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Die spizöhrige Fledermaus. *Vespertilio Nigrita*
Tab. LVIII.

Marmotte volante. DAUBENTON mém. 1759. p. 385.
Chauve-souris étrangère. BUFF. hist. nat. 10. p.82. tab. 18.
Senegal bat. PENN. syn. p. 3669. n. 281.

The snout is of average length, in front thick, the ears are thick and pointed, the tragus is narrow, almost half as long as the ears, and pointed. The tail is about a quarter shorter as the body, enclosed in the tailmembrane, to the two last vertebrae, which project beyond the edge. The hairs on the head, throat and back are yellowish brown mixed with ashgrey; on the breast and belly entirely pale yellowish brown to grey colour. The tail membrane is blackish. The length is four inches.

This is in Senegal discovered by Mr. Adanson.

[*Vespertilio Pipistrellus varietas africana*](#) Rüppell, 1842

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footnote: Ganz gleiche Grösse und Verhältnisse wie bei den europäischen *V. Pipistrellus*, nur der Bauch etwas mehr gelbbraun gefärbt.

Vespertilio platyops Thomas, 1901

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A large pale-coloured species, with the bicuspid incisors of *V. serotinus*, but with a broad blunt tragus.

Size rather less than in the Serotine. Head very flat. Ears of medium length, their inner margin strongly convex in its basal third, evenly slightly convex above to the rounded tip; outer margin slightly concave above, then convex to the notch, and ending in a low rounded lobe. Tragus broad, expanded above its broadest point at about the middle of its inner margin; inner margin straight, joining above at a rounded angle the strongly convex outer margin; a small triangular external basal lobule present.

Wings from the base of the toes. A small postcalcareal lobule. Tail projecting from the membrane by the last vertebra.

Colour above (in spirit) uniform dull fawn or sandy; below dull whitish. Membranes brown, the plagiopatagium and interfemoral inconspicuously edged with whitish behind.

Skull very broad and flat; crests meeting posteriorly, but

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not drawn out into a marked projecting angle, as in the Serotine.

Teeth very much as in the Serotine. Inner upper incisors bicuspid, outer minute. Lower incisors tricuspid, overlapping, their line more directly transverse and less angular than in the Serotine. Anterior lower molar slightly smaller in proportion to the second than in the allied species.

Dimensions of the type (in spirit): -

Forearm 46 millim.

Head and body 62; tail 39; hind foot (s. u.) 8.5; ear 16; tragus, length on inner edge 4, breadth 3; third finger, metacarpal 43, first phalanx 16, second phalanx 14; lower leg 20.5.

Skull: greatest length 19; basipalatal length 15; zygomatic breadth 14; breadth between orbits 8; constriction 4.3; palate, length 7.5; breadth across outer corners of $m.^2$ 8.5.

Hab. Lagos.

Type. Female in spirit. B.M. no. 88.5.17.3. Presented by the late Dr. G. E. Dobson.

The characters of this species are strongly confirmatory of the view that "*Eptesicus*" should not be separated from *Vespertilio*, for with the large size, heavy build, and more or less the crested skull of the former it has the broad tragus of the typical *Vespertilio*.

V. megalurus, the only other African species as large as *V. platyops*, has unicuspidate middle upper incisors.

Vespertilio smithii Wagner, 1855

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A. Smith's *V. minutus* (zool. of South Afric. I. Tab 51) = dessen *V. capensis* kann nicht zur Temminck'schen *V. minutus* gehören, da die von Smith angegebenen Maasse eher auf *V. megalurus* passen. Körper 2 ½", Schwanz 2", Vorderarm 1" 8". Ohren so breit als hoch, Klappe halb so lang als das Ohrn Schwanzende nicht vorragend. Backenzähne jeder seits 4/6. Oberseite tief röthlichbraun, die Haare am Grunde leberbraun; Unterseite ebenso, an den Haarspitzen blass gelblichbraun. Flughäute zwischen bräunlichroth und leberbraun. Man könnte diese Art, die von *V. minutus* Temm. Entschieden abweicht und doch auch nicht recht zu *V. megalurus* passen ???, als *V. Smithii* bezeichnen.

Vespertilio sylvicola A. Grandidier, 1870

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Vespertilio sylvicola

d'un roux marron; abdomen gris foncé. Queue entièrement enveloppée dans la membrane interfémorale.

Corps et queue, 72 mill.; queue seule, 34 mill.; envergure, 21 cent. Hab. forêts d'Ambaravarantou.

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Vespertilio sylvicola

Of a reddish-brown; abdomen grey. Tail entirely included in the interfemoral membrane.

Head and body, 72 mm; tail alone, 34 mm, wingspan, 21 cm. Hab. Ambaravarantou forest.

Vespertilio temminckii Cretzschmar, 1826

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Taf. 6

Vespertilio Temminckii (mas.),
(Mus. Francof.)

Temmincks Fledermaus.

Diagnos. Vespertilio, corpore supra ex cinereo fuscato, infra albo.

Ausmessungen.

-; Fuss; Zoll; Lin.

Länge des Körpers mit dem Kopfe; -; 1; 10

Länge des Kopfes; -; -; 6

Länge des Ohren; -; -; 3 1/2

Länge des Schwanzes; -; 1 ; 2

Breidte des ausgespannten Flügel; -; 7; -

Länge des Daumens; -; -; 2

Beschreibung

Der Kopf und Rücken dieser kleinen Fledermaus ist dunkel mäusegrau mit bräunlichem Ueberzug, der ganze untere Körper glänzend schneeweiss. Der Kopf ist stark behaart und ein Büschel von Haaren ragt über die eine Linie lange und eine Linie breite Nasenkuppe (Rhinarium) hervor, welche ganz nackt und bräunlich ist. Die Nasenflügel sind stark, wulstig, und die Nasenlöcher beinahe ganz zur Seite geöffnet. Die runden Ohren sind hinten ganz nackt, vorn mit feinen grauen Härchen bekleidet. Das Ohrfläppchen ist blattförmig, an der Spitze breiter und zugerundet. Die braunen Flügelhäute sind an der vordern und hinteren Fläche ganz nackt, die hintere Fläche der Schwanz - Flughaut etwas behaart.

Die männliche Thiere sind von den weiblichen eben so wenig im Äusseren verschieden als die jungen von den alten.

Vaterland. Nubien. Rüppel hat 7 Exemplare in der Umgegend von Dongola gesammelt.

Aufenthalt. Fliegt in der Abenddämmerung und wurde an Waldungen und buschigten Gegenden gefunden.

Die mit der Redaction dieses Werkes Beauftragten haben, durch die Benennung der hier beschriebenen Fledermaus, die Huchachtung zu erkennen geben wollen, welche sie für die grossen Verdienste hegen, die

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sich Herr Director Dr. Temminck um die Erweiterung und Berichtigung der Naturwissenschaften durch seine gehaltvollen Arbeiten erworben hat. Sie ergreifen zugleich diese Gelegenheit, um ihrem verehrten Freunde im Namen der Senkenbergischen naturforschenden Gesellschaft öffentlichen Dank abzustatten, für die derselben so oft erwiesene Theilnahme und Mitwirkung an ihrem Gedeihen.

Der von Horsfield beschriebene und mit Temminck's Namen bezeichnete Cheiropter*) gehört zu der von Rafinesque aufgestellten Gattung *Nycticejus*. Dieses Geschlecht wird von Herrn Temminck in seinem Monographieen der Säugethiere nächstens bearbeitet und die *vespertili Temminckii* (Horsfield) als *Nycticejus* aufgeführt erscheinen, worauf wir einstweilen hinweisen wollten.

*) Die von Horsfield in seinen *Zoological researches of Java* angegebene Beschreibung desselben ist aufgefasst wie folgt: *Vespertilio Temminckii*, capite cuneato supra lateribusque planis; auriculis capite brevioribus, oblongis, rotundatis, margine exteriore parum excisis, trago elongato falcato; vellere sericato, pilis brevissimis supra fuscis, subtus sordide flavis, lateribus capitis corporique nitore dilute rufescente.

Dentes primores $1-1/6$, Ianiarii $1-1/1-1$ molares $4-4/5-5$, Körperlänge 4" 6"', Kopflänge 1", Länge der ausgebreiteten Flügel 12'.

Vesperugo (Vesperus) grandidieri Dobson, 1876

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Head, ears, and tragus very similar to those of *Vesperugo abramus*, Temm. Ears short, rounded off above; outer margin of the ear-conch straight or faintly concave in upper two thirds, emarginate opposite the base of the tragus, terminating in a small rounded lobe midway between the base of the tragus and the angle of the mouth: crown of the head scarcely elevated above the face-line; nasal apertures separated widely, opening forwards, their margins not projecting.

Wing-membrane from the base of the toes; postcalcaneal lobe very shallow; half the last caudal vertebra free.

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Fur short on both surfaces, scarcely extending upon the membranes; reddish brown above, paler beneath.

Upper inner incisors bicuspidate, the shorter outer cusp nearly equalled in vertical extent by the unicuspidate outer incisor; lower incisors trifid, not crowded; the single upper premolar close to the canine.

The other species of this subgenus known from Southern Africa are *V. minutus*, Temm., and *V. capensis*, Smith. From both *V. grandidieri* is easily distinguished by the large size of the outer upper incisor.

Length: - head and body 1.75 inch; tail 1.4; ear 0.5; tragus 0.22 x 0.08; forearm 1.25; thumb 0.28; second finger - metacarp. 1.2, first phalanx 0.45, second phalanx 0.55; fourth finger - metacarp. 1.1, first phalanx 0.28, second phalanx 0.2; tibia 0.5; foot and claws 0.3.

The above measurements are taken from the only specimen of this species yet obtained, an adult female with *fœtus in utero*, preserved in alcohol in the Paris Museum, which, by the kindness of H. Alph. Milne-Edwards, I have been enabled to examine and describe. It was brought from Zanzibar by M. Grandidier (who has added so much to our knowledge of the fauna of South-eastern Africa and Madagascar), with whose name I have much pleasure in associating the species.

Vesperugo (Vesperus) Innesi Lataste, 1887

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Vesperugo (Vesperus) Innesi, n. sp.

Formule dentaire (un seul coté de chaque mâchoire étant considéré):

2 / 3 + 1 / 1 + 1 + 3 / 2 + 3

L'incisive inférieure externe a disparu par caducité sur le sujet dont j'ai préparé le crâne.

La pointe des incisives supérieures est trop usée, sur mes deux sujets, pour que je puisse dire si ces dents étaient fourchues ou non; pour tout le reste, le système dentaire de l'espèce nouvelle est très-semblable à celui de l'espèce *V. serotinus* Schreber. L'incisive supérieure externe, notamment, est, de même, très distante de la canine et accolée à l'incisive interne (¹), les centres de figure des quatre incisives supérieures se trouvant

(¹) Les incisives supérieures de *Miniopterus Schreibersi* Natterer sont bien différentes de celles de *Vesperugo serotinus* Schreber, mais elles ne me paraissent pas plus écartées que ces dernières soit de la canine, soit entr'elles sur la ligne mediane. C'est donc à tort que G. E. Dobson voit, dans cet écartement, un caractère propre au groupe des *Miniopteri*, dans la famille de *Vespertilionidae* (*loc. cit.*, p. 170, B).

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situés sur une même ligne droite; l'incisive externe, est en outre très-petite et presque entièrement cachée dans la gencive.

Les crânes des deux espèces ont aussi la même forme générale; mais celui de l'espèce nouvelle est beaucoup plus petit. Le crâne du sujet que j'ai préparé (M, donne au Musée Civique de Gênes) mesure 16^{mm},3 de longueur maximum, tandis que celui d'un *V. serotinus* subsp. *isabellinus* Temminck (F, n° 2408) mesure 19^{mm},3. Les bulles osseuses de l'espèce nouvelle sont aussi très-semblables à celles de *V. serotinus* et *isabellinus*; elles présentent le même développement considerable de l'os tympanique, mais elles sont relativement beaucoup plus grosses.

Dimensions (en millimetres) des deux sujets (conservés dans l'alcool):

F, M
Corps (tête et tronc), jusqu'à l'origine de la queue: 58, ?
Queue: 40, 35
Tête: 20.3, ?
Oreille (depuis l'origine de son bord externe): 15, 15
Oreillon (depuis l'origine de son bord externe): 6.5, 6.5
Largeur maximum de l'oreillon: 2, 2
Avant bras: 42, 40
Pouce (ongles compris): 6, 6.5
Troisième doigt: 70, 65
Cinquième doigt: 50, 47
Jambe: 20, 19.5
Calcaneum: 18, 18
Pied (ongles compris): 9, 9

Description:

La tête de l'espèce nouvelle est très-semblable à celle de *V. serotinus*; son front est plus convexe; son museau est plus

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plat, par suite du développement plus considérable des bourrelets glanduleux latéraux; il est aussi un peu moins velu. Les oreilles sont plus courtes que la tête et construites sur le même

type que celles de *V. serotinus*; le lobe antéro-inférieur de leur bord interne est proportionnellement plus développé en avant et plus arrondi; le lobe antéro-inférieur de leur bord externe est relativement plus saillant et il est isolé de ce bord par une encoche moins élargie et plus profonde. L'oreillon est assez effilé, en pointe obtuse, et il présente, à la base de son bord externe, un tout petit lobule triangulaire; au-dessus de ce lobule, le bord externe est d'abord assez fortement convexe, puis légèrement concave, presque rectiligne; le bord interne est droit. Quand on rabat l'oreille sur les côtés du museau, elle couvre l'oeil et son sommet arrive à peu près au niveau du bout de la mâchoire inférieure.

Callosités du pouce et du tarse, comme chez *V. serotinus*. Insertion de l'aile, au même point que chez ce dernier. Lobe postcalcanéen, très étroit. Queue, à huit vertèbres, dont la dernière, sans compter le cartilage terminal, est libre, la longueur de cette partie libre sur mes deux sujets variant presque du simple au double. Les orteils, même sans les ongles, sont plus longs que la moitié du pied; le premier est sensiblement plus court que les autres, ce qui n'a pas lieu chez *V. serotinus*.

En dessus, le poil s'étend moins loin sur les ailes et, surtout, sur la membrane interfémorale que chez *V. serotinus*; sur. Cette

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dernière membrane, il dépasse à peine le corps, et, sur les premières, il ne s'avance qu'à sept ou huit millimètres de lui. Le lobe antéro-inférieur du bord externe de l'oreille est nu, tandis qu'il est velu chez *V. serotinus*.

Couleur, isabelle très-pâle en dessus, plus pâle, presque blanche, en dessous. Les membranes sont plus foncées. L'aile est bordée par un fin liseré blanc, comme celle de *V. Kuhli* Natterer ⁽¹⁾.

Habitat:

L'espèce nouvelle habite les maisons de la ville du Caire, en Égypte. Je suis heureux de la dédier à M. Walter Innes, conservateur du Musée de l'Ecole de Médecine de cette ville; car c'est à lui que je dois les deux sujets, M et F, conservés dans l'alcool, qui ont servi à cette description, ainsi que beaucoup d'autres Mammifères et Reptiles égyptiens que j'ai utilisés ou que j'utiliserai de mon mieux au profit de la science.

Le sujet dont le crâne a été préparé, le male, a été offert par moi au Musée civique de Gênes; la femelle est conservée dans ma collection.

Affinités et caractères distinctifs.

Par son oreillon, relativement long, atténué au sommet, incliné en dehors, atteignant sa plus grande largeur au-dessous du milieu de son bord interne, à bord externe convexe, l'espèce nouvelle se rapproche du groupe de *Vesperus* sud-américains qui comprend les espèces *V. velatus* Is. Geoffroy, *V. macrotus* Poeppig, *V. montanus* Philippi et *V. magellanicus* Philippi; il se distingue, d'ailleurs, des trois premières, par ses oreilles absolument libres et nullement réunies par un pli cutané, et, des quatre, par ses oreilles plus courtes que la tête; les oreilles sont un peu, chez *V. magellanicus*, et beaucoup, chez les trois autres, plus longues que la tête.

(¹) Plis du palais:

Entre les incisives, une éminence, prolongée, à droite et à gauche, derrière chaque incisive, par une crête courte; entre les canines, un pli transversal, ses extrémités repliées en arrière; un autre pli semblable entre les prémolaires. Cinq paires de plis, arqués et à concavité postérieure, entre les molaires, les derniers moins saillants et moins arqués.

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Parmi les autres espèces du sous-genre *Vesperus*, nous pouvons de suite éliminer les *V. tenuipinnis* Peters, *V. pumilus* Gray, *V. Grandidieri* Dobson, *V. propinquus* Peters, *V. borealis* Nilsson, *V. discolor* Natterer, *V. atratus* Blyth, *V. pachyotus* Dobson et *V. albigularis* Peters, dont l'oreillon a une forme bien différente, atteignant sa plus grande largeur plus ou moins haut et jamais au-dessous du milieu de la hauteur de son bord interne.

Restent les espèces *V. serotinus* Schreber avec sa sous-espèce *V. isabellinus* Temminck, *V. Andersoni* Dobson, *V. Hilairei* Is. Geoffroy, *V. platyrrhinus* Dobson, *V. minutus* Temminck, *V. capensis* Smith, *V. megalurus* Temminck et *V. nasutus* Dobson, qu'il est nécessaire de comparer une à une à l'espèce nouvelle.

Je ne possède, pour cette comparaison, que les *V. serotinus* et *isabellinus*; je m'en rapporterai, pour les six autres espèces, aux descriptions qui en ont été données, notamment par Dobson (Catalogue Chiroptera, 1878).

L'espèce nouvelle est certainement voisine de *V. serotinus* et, surtout, de sa sous-espèce *isabellinus*; mais elle s'en distingue par sa plus petite taille, par son oreille moins largement arrondie au sommet et plus profondément échancrée vers le tiers supérieur de son bord externe, par son oreillon plus long et plus atténué au sommet, par l'angle postérieur plus aigu de sa membrane interfémorale, par sa couleur beaucoup plus pâle, par le fin liseré blanc qui borde sa membrane alaire, etc.

Les autres espèces du même groupe ci-dessus mentionnées me paraissent différer à leur tour de celle que je viens de décrire:

V. Andersoni, par sa taille beaucoup plus grande, par le petit développement du lobe situé à la base du bord interne de son oreille, et par la forme différente de son oreillon; le bord externe de celui-ci présente un petit lobe arrondi et non triangulaire, et, à partir de l'échancrure qui isole ce lobe, il demeure convexe jusqu'au sommet;

V. Hilairei, par ses oreilles plus longues, atteignant presque le bout du museau, et échancrées, dans le tiers supérieur de

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leur bord externe, plus profondément même que les oreilles de *V. serotinus*; en outre, par son habitat sud-américain;

V. platyrrhinus ⁽¹⁾, par la forme de son oreille, nullement échancrée vers le haut de son bord externe et coupée anguleusement à la base de son bord interne, par la forme de son oreillon, plus élargi au dessus de sa base et plus courtement aigü au sommet, par le développement plus considérable et la forme différente des bourrelets glanduleux du museau;

V. minutus, par sa taille considérablement plus petite;

V. capensis, par l'insertion de la membrane alaire à la base de l'ongle du cinquième orteil et non à la base du cinquième orteil;

V. megalurus, par sa taille plus grande, égale à celle de *V. serotinus*, par son oreillon plus court même que celui de *V. serotinus*, par son incisive supérieure externe, dont la base est large et située exactement entre l'incisive interne et la canine, au lieu d'être étroite et située sur le même niveau transversal que l'incisive interne;

V. nasutus, enfin, par la forme de son museau, conique et projeté en avant comme dans le genre *Nyctinomus*, par l'absence de lobe arrondi à la base du bord interne de l'oreille, etc.

Paris, 8 mai 1887.

(¹) Voir Dobson, *loc. Cit.*, pl. XII, fig. 1.

Vesperugo (Vesperus) Moloneyi Thomas, 1891

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Vesperugo (Vesperus) Moloneyi sp. n.

Vesperus with the tragus extraordinarily short, with the outer upper incisors nearly as long as the inner, and with the anterior lower premolar minute.

Size of body medium, but the extremities so short that the forearm-length makes the species appear to be among the smallest of the genus. Head very broad and flat, much as in *Nycticejus** or in *V. pachypus*: facial glands swollen, but not raised vertically above the level of the centre of the muzzle. Ears very short, their edges evenly continuous, not emarginate externally either above or opposite the base of the tragus; the small keel usually present just behind the base of their inner margin nearly or quite obsolete. Tragus (see figure) extraordinarily small, quite unlike that of any other member of the group, its height, measured along its inner edge, less than half its breadth, its upper and its outer margins rounded; its outer base without any projection. Anterior extremities much reduced throughout, especially distally; the forearm but little more than half of the combined lengths of the head and body, the thumb very short, the two phalanges of the middle finger together only about half the length of the short forearm, and those of the fifth finger less than a fifth of it; finally the usual distal cartilaginous extension of the third finger is nearly obsolete. Hind legs unusually thick and muscular. Calcar feeble, post-calcareal lobule absent. Extreme tip of tail only free from membrane.

Fur short, uniformly very dark brown or black above and beneath.

Upper inner incisors long, their tips bicuspid; outer ones

* When showing (Ann. Mus. Genov. (2) ix. p. 88, 1890) that the American *Nycticejus humeralis* could not be separated generically from the Old-World *Scotophilus*, I did not notice that the name *Nycticejus* (1819) was anterior in date to *Scotophilus* (1822). Mr. Blanford has since pointed this out to me; and it is evident that the former name must be used for all the species hitherto called *Scotophilus*.

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cylindrical, unicuspid, not reaching quite to the level of the outer cusps of the inner incisors. Upper premolars very close to and but little shorter than the canines. Lower incisors tricuspid, overlapping. Anterior lower premolar very small, scarcely exceeding in height the cingulum of the large posterior one.

Dimensions of the type, a male preserved in spirit: -

Head and body 50 millim.; head, length 16, breadth across muzzle 11; ear, length from base of inner edge 9, length from base of outer 11.5; tragus, length of inner margin 1.0, length of outer margin 3.8, breadth above 1.9, height of base 3.2. Forearm 29 (= 1.15 inch); thumb, including claw, 5.5; third finger, metacarpal 29, first phalanx 8.5, second phalanx 7.8; lower leg 11; hind foot, including claws, 8.2; tail 30.

Hab. Lagos, West Africa. One specimen, collected and presented by Sir Alfred Moloney.

This most remarkable species is distinguished by its proportionally large body and head and the reduction in length of all its extremities, including in this term the ears, tragus, wings, legs, and tail. This reduction, combined with the markedly more muscular condition of the legs, no doubt indicates a less exclusively aerial manner of life; and we may be prepared to find when its habits are known that it seeks for its prey creeping about either the trunks and branches of trees or the rocks of cliffs and caves, rather than flying about in the open.

Vesperugo (Vesperus) Rendalli Thomas, 1889

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Crown of the head flat, scarcely raised above the level of the muzzle; muzzle broad, the glandular prominences much swollen. Ears short; laid forward they reach barely more

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than one half the distance between the eye and the tip of the nose; their inner margin straight, tip narrowly rounded off, upper third of outer margin straight, middle third slightly convex, basal third with a marked lobe separated by a notch opposite the base of the tragus.

Tragus of about the same breadth from base to tip, its inner margin evenly but slightly concave, its tip rounded, its outer margin evenly convex, its base with a triangular lobule; when laid forward its tip reaches to between the posterior canthus and the middle of the eye.

Pads at the bases of the thumbs scarcely more marked than usual, but the soles of the feet broad, swollen, and wrinkled, projecting posteriorly behind the level of the origin of the calcaneum. Wings from the metacarpus close to the base of the toes. Calcaneum reaching about halfway from the heel to the end of the tail, its tip forming a small projecting point like that found in *V. Daubentoni*; postcalcaneal lobule distinct, but rather narrow. Tail involved in the interfemoral membrane to its extreme tip. Penis with a distinct bone.

Fur scarcely extending on the membranes except on the interfemoral in the centre below.

Colour of body, above and below, of the skin or the forearm, digits; and legs greyish brown, of the membranes everywhere white, the reticulations well marked.

Teeth. - Upper incisors (figs. b-d), or at least the outer

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pair, quite different from those of any other member of the genus. The inner one with a large secondary cusp about halfway up its posterior edge, and the hinder edge of the main cusp, above the secondary one, markedly sinuous, as though the tooth were occasionally trifold (fig. b). Outer incisor well developed, triangular in section at the base (see fig. c), its angles each with a cusp, the antero-internal bearing the long main cusp, three fourths the height of the inner incisor; cusp on the postero-internal angle about half the height of the secondary cusp of the inner incisor; cusp on the external angle minute, but quite distinct (fig. d); postero-external side of the tooth close to the base of the canine (fig. c). No trace of a minute anterior premolar; large premolar pressed closely against the back of the canine and leaving no room for another tooth between.

Lower incisors trifold, overlapping, placed at right angles to the direction of the jaws. Second lower premolar two thirds the height of the canine, and first premolar two thirds its height.

Dimensions of the type (an adult male in spirit): - Head and body 50 millim.; tail 39; forearm 36 (= 1.4 inch); head 16.5; muzzle to eye 7.0; ear, above crown 9.0, from base of external margin 13.0; thumb, including claw, 6.2; lower leg 13.6.

The type specimen of this most interesting bat was obtained by my friend Dr. Percy Rendall at Bathurst, on the River Gambia, and I have much pleasure in connecting his name with it.

Vesperugo Rendalli presents a combination of characters which prevents it falling into anyone of the subgenera of *Vesperugo* as recognized and defined by Dr. Dobson (Cat. Chir. B.M. p. 184). But I am far from regarding it as representing a new subgenus, and only consider it as showing that *Hesperoptenus*, Peters, in which a penis-bone is present and the outer incisors are minute, should be amalgamated with *Vesperus*, in which there is no penis-bone and the outer incisors are well developed. The swollen and probably adhesive hinder pads of this species give it a second point of resemblance to *V. ("Hesperoptenus") Blanfordi*, Dobs.; but, as Dr. Dobson has shown, this character is not one on which generic or subgeneric division can be based.

Vesperugo [(Vesperugo)] maderensis Dobson, 1878

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Ears and tragus generally like those of *V. kuhlii*, but the outer margin of the ear-conch is much more deeply emarginate above, and the middle third consequently more abruptly convex (Plate XII, fig. 5, ear, double natural size).

Postcalcaral lobe distinct, placed on the calcaneum about the length of the foot from the ankle; extremity of the tail free; wings from the base of the toes; feet small.

Fur, above, dark brown with light brown extremities to the hairs; beneath, the hairs are dull black with yellowish-brown tips. No trace of a white margin to the membranes.

On the upper surface the fur extends upon the wings almost as far as a line drawn from the proximal third of the humerus to the knee, and upon the base of the interfemoral membrane; beneath, the wing-membrane is clothed as far as a line drawn from the

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elbow to the knee; fur longer on the ventral than on the dorsal surface.

Inner upper incisor on each side very slender, long, and unicuspidate, the outer incisor very small and close to its base; lower incisors at right angles to the direction of the jaw; second upper premolar separated by a narrow space from the canine, through which the summit of the minute first premolar can in some specimens be seen.

Length (of an adult M), head and body 1".6, tail 1".5, head 0".55, ear 0".5, tragus 0".22, forearm 1".3, thumb 0".22, second finger 2".25, fourth finger 1".6, tibia 0".45, foot 0".25.

(Three specimens (from Palma and Madeira) have been examined and measured by me and found to agree in all respects.)

Hab. Madeira and Canary Isles.

This species is closely allied to *V. kuhlii*, but may be at once distinguished by the much shorter body and longer tail, by the very deep emargination on the outer side of the ear, and by the complete absence of the white margin to the membranes.

a. F ad., al. (type). Madeira. W. S. MacGillivray [C.].

b. M ad., al. Palma.

Vesperugo Anchieta Seabra, 1900

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Treze fêmeas de Cahata representam esta nova espécie angolense que caracterizamos assim: cabeça notavelmente deprimida; o focinho curto; narinas proeminentes e bem distanciadas; as orelhas grandes, largas na sua base; o chanfro da orla externa muito escavado, a ponta arredondada; o tragus bem desenvolvido e ligeiramente serrilhado na orla posterior; o anti-tragus bem visível e triangular. A pelagem abundantíssima tanto na cabeça como sobre o dorso e ventre; de um pardo-escuro um pouco avermelhado sobre o dorso, mais claro no ventre; membrana branchial nua; a inter-femural coberta de pelos tanto pela parte superior como pela parte inferior, até meio do comprimento da cauda. Estes pelos, como em geral succede, são muito claros e curtos na parte inferior d'esta membrana. Na região compreendida entre o humero e o femuro, pela parte inferior (região ventral), o pelo estende-se com bastante abundancia e é longo. Pela parte dorsal estende-se apenas ligeiramente sobre o femuro. O paladar apresenta os caracteres genericos; os lóbulos anteriores são deprimidos e as pregas encontram-se, como procuramos representar no schema 22, quasi igualmente distanciadas entre si.

Dentição: incisivos superiores simples; o interno bem desenvolvido, o externo attingindo apenas metade do comprimento d'este; o canino separado apenas por um pequeno espaço do segundo premolar e deixando entre si e este ultimo dente o primeiro premolar pequenissimo e difficil do reconhecer. Os molares perfeitamente conformados. Os incisivos inferiores trilobados.

Dimensões: cabeça e corpo 40 mili.; cabeça 15; orelha (da base do tragus á penta) 11; cauda 32; ante-braço 35; pollex 4; 3.º dedo: 3.º dedo: metacarpô 31; 1.ª phalange 12; 2.ª phalange 16; 4.º dedo: metacarpô 30, 1.ª phalange 10, 2.ª phalange 8; 5.º dedo: metacarpô 20, 1.ª phalange 9, 2.ª phalange 5; tibia 43; pé 6. Nos outros indivíduos as dimensões correspondem exactamente a estas.

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Thirteen females from Cahata represent this new Angolan species that are characterised as follows: notably depressed head; short snout; prominent nostrils and well-spaced; big ears, large at the base; the bevel of the external edge is concave, with a rounded tip; the tragus is well developed and slightly serrated on the back edge; the anti-tragus is very visible and triangular. The coat of hair is equally abundant on the head, back and belly; from a dark brown it becomes a little red on the back, and clearer on the belly; the branchial membrane has no hair; the inter-femur is covered by hairs on the top and the bottom, halfway of the length of the tail. These hairs, as generally, are very light and short on the bottom part of the membrane. In the region between the humerus and femur, on the bottom part (belly region), the hair extends abundantly and is long. On the dorsal part it extends over the femur. The palate presents general characteristics; the anterior lobes are depressed and the folds are, as we sought to represent scheme 22, they are almost equally spaced.

Dentition: simple superior incisors; the interior is under developed, the external reaches about half the length of this one; the canine separated by a small space between the second premolar and in between these two is a small first premolar which is difficult to identify. The molars are perfectly conformed. The inferior incisors are tri-lobed.

Measurements: head and body 40 mill.; head 15; ear (from the base of the tragus to the tip) 11; tail 32; forearm 35; thumb 4; third finger: metacarpal 31; 1.^a phalange 12; 2.^a phalange 16; fourth finger: metacarpal 30, 1.^a phalange 10, 2.^a phalange 8; fifth finger: metacarpal 20, 1.^a phalange 9, 2.^a phalange 5; tibia 43; foot 6. The measurements of other individuals correspond exactly with these.

Vesperugo hypoleucus Heuglin and Fitzinger, 1866

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In Sennaar, zwischen Kereri, Halfäye und Surerat am Nil in der Nähe von Chartum gesammelt. Ausgezeichnet durch die rein weiße Färbung des Bauches.

Vesperugo pagenstecheri Noack, 1889

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Taf. II, Fig. 3, Taf. V, Fig. 64 - 65.

Zwei Spiritus-Exemplare F. Netonna. Im Wipfel einer Kokuspaluie gefangen. Coll. Hesse.

Diagnose. Kleiner *Vesperugo*, der *Vesperugo nanus* PETERS nahe steht, aber erheblich kleiner und viel heller gefärbt ist. Ohr lang und schmal zugespitzt, hinten etwas ausgeschnitten, der äussere Ohrrand mässig gegen den Mundwinkel vorgezogen, der kleine und schmale Tragus innen gerade, aussen convex. Färbung oben hell gelblich olivenfarben, unten noch heller weisslichgelb mit olivenfarbenem Anfluge.

Beschreibung. *Vesperugo pagenstecheri* in zwei identischen Exemplaren unterscheidet sich von *V. nanus* durch die geringere Grösse, die längeren und schmalen Ohren, den anders geformten Tragus, den längeren zweiten Finger und die viel hellere Färbung.

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Der Kopf ist klein, die Nase, deren Seiten mit borstigen, nach vorn gerichteten Wimpern besetzt sind, stumpf, die seitwärts geöffneten Nasenlöcher mit wulstigem Baude durch eine seichte Furche getrennt, die Unterlippe ohne Warzen, fein behaart. Die langen, niedergedrückt die Nasenspitze überragenden Ohren sind sehr schlank zugespitzt, aussen nur wenig eingebuchtet, der untere Dach vorn vorgezogene, etwas verdickte Rand endet in einen hochstehenden Lappen von 0.75 mm Länge. Der schmale Tragus ohne Furche steht ziemlich vertical und ist nicht beilförmig wie bei *V. nanus*, sondern oben einfach abgerundet, die innere Kante gerade, die äussere etwas gebogen. Der Daumen ist frei. Die unbehaarte Flughaut am Ende des zweiten Fingers scharf zugespitzt, der letzte Wirbel des Schwanzes ist frei. Die im Basaltheile dichter, unten spärlicher mit gelblichen, oben mit bräunlichen Härchen besetzte Schwanzflughaut ist schlank zugespitzt und entbehrt der Ausbuchtung, die Schenkelflughaut reicht bis über die erste Phalange des Fusses. Bein und Fuss sehr zart, Nägel weisslichgelbbraun. Flughaut schwarzbraun, Schenkelflughaut heller, gelbbraun, Humerus, Unterarm, zweiter Finger, Unterschenkel weisslichgelb. Wimpern und Nagenseite schwärzlichbraun. Haarbasis hell graubraun, bei *nanus* schwarzbraun, Haar lang, fein, flockig, hell gelblichumbra mit Stich ins Olivenfarbene, unten heller mit vielfach durchschimmernder graubrauner Haarbasis. Bei *V. nanus* ist die Färbung viel dunkler braun. Vergl. PETERS, Säugethiere Mosamb., p. 63, Taf. 16, 3; DOBSON, Cat. Chiropt., p. 237.

Maasse. *V. pagenst.*, *V. nanus*.

Körper: 40, Körper und Schwanz: 28, Schwanz: 80

Ohr lang: 9.5, 11.5 - 12

" breit: 6, 7

Tragus: 3, 3.5

Humerus: 16, 19 - 20

Unterarm: 30, 31.5

Daumen: 5, 5.5

II: 35, 29 - 30

III: 52, 54 - 57

IV: 44, 45 - 49

V: 37, -

Femur: 12; 12.5 - 13

Unterschenkel: 11, 12

Fuss mit Krallen: 6, 7

Sporn: 5, -

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Der Schädel (Tf. V, Fig. 64) ist ziemlich schmal, Scheitel und Hinterhaupt abgerundet, Scheitel- und Lambdanaht fehlen, das sehr grosse Hinterhauptloch wird erheblich von der Squama occipit. überragt, die Stirn ist gewölbt, die Nasenbasis eingesenkt, die rundlichen Nasenbeine durch eine mediane Furche getrennt. Der Orbitalrand ist vorn wulstig, die sehr feinen Jochbogen wenig gebogen, der kurze Oberkiefer nach oben gebogen, der Gaumen ist hinten breit, vorn zugespitzt. Von den 7 Gaumenfalten gehen die beiden ersten quer durch, die folgenden sind gebrochen und die inneren Bogen stark nach hinten

gezogen. Am Unterkiefer ist der aufsteigende Ast niedrig und sehr breit, der Proc. coron. mit scharfer Spitze, etwas nach vorn gerichtet, die Massetergrube tief mit scharfen Leistenrändern.

Maasse. Scheitellänge 11 (bei *Vesperugo nanus* 12), grösste Scheitelbreite 6.25, Einschnörung 3.5, Schädelkapsel 7, Nasenbreite 3.5, Hinterhauptloch 2.5 hoch und breit, Höhe der Squama occipit. über dem For. occipit. fast 2, hintere Gaumenbreite 3.5. Unterkiefer bis zum Condylus 7, aufsteigender Ast 2, Höhe des horizontalen Astes 1.

Gebiss (Taf. V, Fig. 65) von *Vesperugo*: $I^{2+2}/6 C^{1+1}/1+1 P^{2+2}/2+2 M^{3+3}/3+3$.

I innen mit der Spitze nach innen und vorn gerichtet, zweispitzig, der hintere Zacken stark, nicht ganz so lang wie der vordere. I aussen vorn mit kräftigem Basalwulst. kürzer und schmaler als I innen. C mit breitem Basaltheil, aber schlank zugespitzt und nach vorn gerichtet, vorn, aussen und hinten mit scharfer Kante, innen stark cannelirt. Der Basalwulst springt nach hinten höckrig vor und ist röthlich gefärbt, wie auch P und M einen hellröthlichen Anflug haben. P I ein sehr kleiner, rudimentärer, nach innen gerichteter Zacken. P II = $3/4$ C, länger als M. Basalwulst vorn höckrig, hinten ausgezogen, der Innenzacken vertieft. M I und II identisch, die beiden Aussenzacken gleich hoch, der Innenhöcker vertieft mit nach vorn gerichtetem Innenzacken. M III kleiner, der hintere Theil des W unbedeutend, der Innenzacken nach vorn gerichtet. C ist 1 mm hoch, die übrigen Zacken entsprechend niedriger.

Unten I nach vorn gerichtet, klein, gestielt, stark dreilappig, C schmal und gerade, der Basalwulst innen mit kleinem Nebenzacken. P und M mit Basalwulst, P I und II einspitzig, I etwas niedriger. Bei M der vordere Aussenzacken etwas höher als der hintere, innen 3 Zacken, der vordere nach vorn, der hintere nach hinten gerichtet. M III etwas kleiner

Vesperugo pulcher Dobson, 1875

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Crown of the head slightly elevated above the face-line; muzzle shaped more like that of some species of *Vespertilio* (*V. emarginatus*, e. g.), and the glandular prominences between the nostrils and eyes less developed than in most species of *Vesperugo*. Ears about four fifths the length of the head, the ascending inner margin straight or faintly convex, the upper half of the outer margin slightly concave, terminating in a line directly below the eye, but on a lower level than the angle of the mouth; tragus nearly half the length of the ear, shaped like that of *V. serotinus*.

Posterior margin of the interfemoral membrane festooned with distinct papillæ arranged like the teeth of a comb.

Fur above sienna-brown, with shining tips, beneath wholly pure white; the dark-coloured fur of the back does not pass beyond a line connecting the angle of the mouth (when the head is extended) and the under surface of the humerus. Interfemoral, antebrachial, and wing-membranes between the humerus and femur white; the remaining parts of the wing-membrane pale brown.

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Inner upper incisor long, with a second cusp on its outer side near the extremity. Outer incisor short, lying close to the base of the inner incisor. Second premolar in the upper jaw separated from the canine by a short interval, through which the small first premolar is distinctly visible from without. Lower incisors not crowded.

Length (of an adult male preserved in alcohol), head and body 2".0; tail 1".6; head 0".7; ear 0".6; tragus 0".25; forearm 1".35; thumb 0".28; second finger 2".6; fourth finger 1".9; tibia 0".52; foot and claws 0".3.

Hab. Zanzibar.

Type in the collection of the British Museum.

This species resembles *V. (Alobus) temminckii*, Rüpp., but may be at once distinguished by the form of the tragus, which is narrowed towards the tip and subacutely pointed, by the deeply fimbriated margin of the interfemoral membrane, by its much greater size, &c.

Vesperugo pusillulus Peters, 1870

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Vesperugo pusillulus. Nov. sp.

V. supra ferrugineus, subtus pallidior, pilis omnibus basi nigris; auriculis triangularibus rotundatis, externe vix emarginatis, plicis 5 transversis; trago versus apicem dilatato, ut in V. nochila; antitrigo rotundato-triangulari; patagio ad digitorum basin affixo; cauda usque ad apicem patagio involuta; patagio femorali supra basi pilosa, subtus lineis transversis ciliatis; apice

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dentis incisivi superioris primi postico ciente incisivo secundo vix altiore; praemolari primo superiore a latere visibili.

Long. tota 0^m,0600

" capitis 0^m,0125
 " auris 0^m,0085
 " " marg. ant 0^m,0060
 Lat. auris 0^m,0070
 Long. tragi 0^m,0035
 " caudae 0^m,0240
 " humeri 0^m,0165
 " antibrachii 0^m,0250
 " digit 1 0^m,0050
 " 2. (mtc. 0,0235; ph. 1. 0,0011) 0^m,0246
 " " 3. (" 0,0255; " 0,0097; ph. 2. 0,0073; cart. 0,0045) ... 0^m,0470
 " " 4. (" 0,0247; " 0,0078; " 0,0050; " 0,0015) ... 0^m,0390
 " " 5. (" 0,0244; " 0,0063; " 0,0027; " 0,0015) ... 0^m,0349
 Long. fem 0^m,0100
 " tibiae : 0^m,0090
 " plantae 0^m,0045
 " calcar 0^m,0090

Exempla duo ex Loango. (Anchieta)

Vesperugo senarensis Heuglin, 1877

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Vesperugo senarensis, Heugl. - Fitz. u. Heugl. Säugeth. p. 10. - In der Umgegend von Chartum.

Vesperugo sennaariensis Heuglin and Fitzinger, 1866

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In Sennaar, in der Umgegend von Chartum.

Vesperugo stampflii Jentink, 1888

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An adult male, Farmington River (Stampfli).

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The smallest *Vesperugo*-species hitherto recorded. Ears rounded off, inner margin convex, outer margin straight for its greatest part, then convex, terminating in a small rounded lobe at a short distance behind the angle of the mouth. Inner margin of tragus slightly concave, outer margin convex, tip rounded off.

Wings from base of toes, calcaneum long with projecting tip; extreme tip of tail free. Interfemoral and wing-membranes margined with white.

Fur above black with brownish red tips, beneath black with light brown tips. Ears and membranes black.

Inner upper incisors bifid, outer incisors unicuspidate; lower incisors trifid. First upper premolar very small, hardly visible from without; first lower premolar three fourth of the size of the second premolar.

Measurements of the adult male, in alcohol (in mm):

Length of head and body: 38

" " tail: 24
 " " ear: 9
 " " tragus: 4
 " " forearm: 33
 " " thumb: 5
 " " third finger: 45
 " " fifth: 36
 " " tibia: 10
 " " foot: 5

I call it after its discoverer, Mr. Stampfli.

Vesperus bicolor Bocage, 1889

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Museau obtus, large et renflé. Oreilles courtes, mesurant à peine $\frac{2}{3}$ de la longueur de la tête, avec le bord interne fortement convexe à partir de la base et l'externe émarginé dans sa moitié supérieure; l'extrémité arrondie. Tragus allongé, cultriforme, à bords parallèles et avec un petit lobe triangulaire à la base du bord externe. Ailes par-

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tant de la base du 1^{er} orteil; un lobe calcanéen distinct et convexe. Queue presque entièrement comprise dans la membrane interfémorale.

Poils des parties supérieures brun-marron avec les extrémités d'un roux ferrugineux; ceux des parties inférieures d'un marron plus foncé, presque noir, avec les extrémités blanches; les poils du museau roux-ferrugineux. Les membranes alaires transparentes, d'un brun-roussâtre pâle; les membres de cette même couleur.

Incisives supérieures internes indistinctement bilobées, larges et courtes, les externes rudimentaires; pas de trace de la première prémolaire à la mâchoire supérieure; les incisives inférieures trilobées, disposées transversalement par rapport au bord de la mâchoire; la première prémolaire inférieure petite, ayant à peine la moitié de la hauteur de la deuxième.

Deux individus femelles de *Caconda* par M. d'Anchieta.

De la taille à peu-près du *V. minutus*, Temm., et du *V. tenuipinnis*, Peters, et se rapprochant du second par ses couleurs. Malheureusement ces deux espèces manquent aux collections du Muséum, de Lisbonne, de sorte que nous avons à nous prononcer d'après les descriptions de ces deux espèces: il doit ressembler au *V. minutus* par la forme du tragus, mais s'en écarter par la forme et les dimensions relatives de ses incisives supérieures et aussi par ses couleurs; ces derniers caractères semblent s'accorder mieux avec ceux du *V. tenuipinnis*, mais le tragus de cette espèce doit avoir une forme toute différente, ayant été comparé par Peters au tragus du *V. noctula*.

Voici maintenant les dimensions des parties les plus intéressantes à comparer chez les trois espèces:

V. bicolor, nova sp.; *V. minutus* (Dobs.), *V. tenuipinnis*, (Peters)

Tête et corps: 43 mm.; 45 mm; 46 mm.

Queue: 26; 27; 30

Oreille: 12; 11; 11

Tragus: 6; 5.7; 4.5

Avant-bras: 30; 31; 29

Pollex: 6; 5.5; 5

3^e doigt: 53; 50; 53

Jambe: 11; 10; 11

Vesperus damarensis Noack, 1889

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Taf. V, Fig. 59.

Ueber *Vesperus* s. DOBSON, Cat. Chiropt., p. 184 ff.

Drei Exemplare F, zwei in Spiritus. Damaraland, Omburo und Golabu, Coll. SCHINZ.

Diagnose. Mittलगrosser *Vesperus* mit langen, oval zugespitzten Ohren, deren äusserer Rand aber etwas eingebuchtet ist, und geradem, aussen halbmondförmig abgerundetem Tragus, mit dunkelbrauner, zwischen Schenkeln und Schwanz hell gelbbrauner, von weissen parallelen Adern durchzogener Flughaut, oben von hell sepiagelber mit Graubraun gemischter, unten von weisegrauer mit Schwarzbraun gemischter Farbe der ziemlich langen und flockigen Haare.

Beschreibung. Die spärlich behaarte Nase ist kurz und breit, schwarz gefärbt, die nach aussen geöffneten Nasenlöcher mit wulstigem Rande sind weit vorgestreckt und durch eine breite Furche getrennt, die Oberlippe verdickt, die Stirn stark behaart. Die 12 mm lange Zunge schmal, hinten nur wenig verdickt. Das oval zugespitzte Ohr reicht niedergedrückt bis zur Nasenspitze, nur der Basaltheil des vorderen Randes und die hintere Basis sind behaart. Der äussere Rand ist wenig ausgebuchtet und unten verdickt, der untere Lappen mässig, bei weitem nicht bis gegen den Mundwinkel vorgezogen. Der Tragus ist etwas nach vorn geneigt, der innere Rand desselben gerade, der äussere halbmondförmig convex und unten umgebogen. Daumen und Zehen dünn behaart, der kleine Daumen mit basaler Schwiele und weissem oder weisslichbraunem Nagel frei, der dritte Finger stark

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verlängert, daher die Flughaut zwischen zweitem und drittem Finger stark zugespitzt; die distale Hälfte des Oberarmes nackt, die Flughaut zwischen Arm und Schenkeln mit starken Papillen, von weissen parallelen Adern durchzogen, oben am proximalen Rande und unten etwas weiter hin dünn behaart. Die Schwanzflughaut schliesst den Schwanz ganz ein, ist hinter dem Caleaneus stark ausgebuchtet und oben und unten am proximalen Theile dünn weiselich behaart; auch hier verlaufen weisse parallele Adern schräg nach dem Schwanz.

Das lange weiche Haar ist von ungleicher Länge, erscheint daher flockig, die Basis dunkel schwarzbraun, die Spitze weisslich-gelb mit Sepiabeimischung und Silberglanz. Am Oberarm und der Unterseite sind die Haarspitzen weisslich. Stirn, Scheitel und Ohr sepiagelbbraun, Kinn und Gesichtsseiten etwas heller. Die Färbung ist der von *Vespertilio lanosus* SMITH ähnlich; letztere, der SMITH ungeheuerlicher Weise einen halbmondförmigen an seinem Exemplar offenbar zufällig entstandenen Ausschnitt des äusseren Ohrandes zuschreibt, ist falsch bestimmt und muss nach DOBSON *Kerivoula lanosa* heissen, da sie gar nicht das Gebiss von *Vespertilio* hat.

Maasse: Körper 43 - 50 mm, Schwanz 26 - 30, Ohr 9 - 10 lang, 7 - 8 breit, Tragus 4, Humerus 19, Unterarm 32 - 33, Daumen 4.5 - 5, II 57.5 - 60, III 52 - 54, IV 45 - 47, Oberschenkel 12, Unterschenkel 11 - 12, Sporn 10 - 10.5.

Schädel. Die obere Profillinie ist ziemlich gerade, die Nase sehr wenig eingebogen, die Stirn sehr wenig convex, die Scheitelnah schwach, die Stirnbeine nach hinten rundlich pfeilförmig (im maurischen Kielbogen) verlaufend, der Schädel zwischen den Stirnbeinen eingeschnürt. Die Nasenbeine und Zwischenkiefer sind durch die Wurzeln von I und C verdickt. Der Jochbogen sehr fein, in der Mitte etwas nach oben gezogen. Hinterhaupt rundlichdreieckig, etwas gewölbt und unten nach vorn abfallend, das Foramen occipit. sehr gross. Das Tympanum an der Aussenseite flach mit etwas verdicktem Rande, der knöcherne Gaumen hinten mit einem Zacken. Das rundliche Foramen Infraorbit. sitzt hoch, dicht am Orbitalrande.

Der horizontale Ast des Unterkiefers ist mässig gebogen, nach hinten verjüngt, der Proc. coron. wie gewöhnlich dreieckig, etwas nach vorn gerichtet, die Massetergrube tief, vorn mit dreieckigem scharfem Rande.

Maasse: Scheitellänge 19.5 - 20, Basilarlänge 18 - 18.5, Höhe des Hinterhauptes und Scheitelhöhe 5, Breite des Hinterhauptes zwischen

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dem Tympanum 8.5, hinter den Jochbogen 8, Weite zwischen den Jochbogen 10, Scheitelbeine fast 7, Nasen- und Stirnbeine 5, knöcherner Gaumen 6.5, Breite vor den Jochbogen 5, Länge des Jochbogens 5, For. occipit. 2.5 hoch, 3 breit. Unterkiefer bis zum Proc. cor. 8, bis zum Condylus 10.25; Höhe des horizontalen Astes fast 1, unter dem Proc. coron. 2.75.

Gebiss von *Vesperus*: $I^{2+2}/6 C^{1+1}/1+1 P^{1+1}/2+2 M^{3+3}/3+3$

Oben I (Taf. V, Fig. 59) mit Basalwulst, I innen klein, mässig gebogen mit sehr kleinem hinterem Zacken, der bei *Vesperus* im Alter verschwindet und nur bei einem Exemplar noch deutlich sichtbar ist. I aussen sehr klein, eng an I innen gedrängt, etwas nach vorn gerichtet. C mässig gebogen, senkrecht zur Kieferachse gestellt, hinten und innen ausgekehlt mit scharfer äusserer Kante, $P = 1/2 C$, mit starkem hinterem Basalzacken, vorderem Basalwulst und starkem innerem Zacken mit vertiefter Kaufläche, Spitze nach innen gerichtet. M I und II W-förmig, die Aussenzacken doppelt, die innere Reihe höher, vor dem ersten Aussenzacken noch ein kleiner Basalzacken. Der innere tief liegende Kauhöcker innen gefurcht mit scharfem mittlerem Zacken. M III nach hinten gerichtet = $1/2 M III$, aussen ein, daneben, nach innen zwei Zacken, hinten zwei Zacken, der innere höher.

Unten I mit gestielter Wurzel, die Krone scharf dreilappig, die I stehen nicht senkrecht, sondern schräg nach vorn zur Achse des Kiefers C senkrecht zur Kieferachse, schlank, mässig gebogen, mit Basalwulst, der nach hinten und vorn in einen Zacken endet, wie auch bei P II und M. Die hintere Fläche von C ist mässig ausgekehlt. P I klein, eng zwischen C und P II eingekeilt, kleiner als $1/2 C$, etwas höher als $1/2 P II$; $P II = 3/4 C$. M I aussen und innen mit je 2 Zacken, I aussen höher mit kleinem vorderen Nebenzacken, M II aussen 2, innen 3 Zacken, ebenso M III, der erste viel höher als der zweite, innen der letzte mit kleinem Hinterzacken.

Von den Afrika, resp. die südafrikanische Subregion bewohnenden *Vesperus*-Arten bind zu vergleichen *V. serotinus* und *capensis*, da, wie aus der Beschreibung erhellt, *V. minutus*, *andersoni*, welcher viel grösser ist, *tenuipennis* und *grandieri* nicht in Frage kommen, vollends nicht *V. nasutus* und *platyrhinus*. Es ist an und für sich nicht wahrscheinlich, dass *V. serotinus* sich bis in die südafrikanische Subregion erstreckt, und da bei diesem das nach vorn gebogene Ohr nur über die Mitte zwischen Auge und Muffel, bei *damarensis* bis zur Nasenspitze reicht, ist eine Identifizierung mit *V. serotinus* nicht mög-

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lich. Das Ohr von *V. damarensis* hat die Proportion wie bei dem kürzlich von F. LATASTE (in: Ann. Mus. Civ. Stor. Nat, Genova (Ser. 2), Vol. 4, 1887) aus Kairo beschriebenen *Vesperus innesi*, nur ist der Tragus ganz anders. Auch das Ohr von *V. capensis* unterscheidet sich erheblich, hier liegt der verstärkte Aussenrand des Ohrs viel höher als bei *V. damarensis*, der Tragus ist bei *damarensis* unten an der Aussenseite umgebogen, die äussere Schwanzspitze ist frei, die Ausbuchtung der Schenkelflughaut ist viel kleiner, die Färbung ist oben rötlichbraun mit hellerer Haarspitze, die inneren I oben stehen bei *damarensis* weiter von den äusseren ab als bei *capensis*, die unteren I stehen bei *V. capensis* senkrecht zur Kieferachse, der hintere Ansatz von P oben ist breiter

Vesperus guineensis Bocage, 1889

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Museau obtus, portant de chaque côté entre la narine et l'œil un renflement glandulaire arrondi. Oreilles plus courtes que la tête, triangulaires, arrondies au bout, avec le bord externe distinctement émarginé dans sa moitié supérieure. Tragus plus court que la moitié de la hauteur de l'oreille, cultriforme, à bords droits et à pointe arrondie, atteignant sa plus grande largeur vers le milieu du bord externe, qui porte à la base un petit lobe triangulaire. Ailes partant de la base du premier orteil; le lobe postcalcanéen convexe à son bord libre. Queue

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comprise jusqu'à près de l'extrémité dans la membrane interfémorale.

Les poils en dessus et en dessous d'un beau marron uniforme, à l'exception de ceux qui couvrent la région pubienne, la membrane alaire des deux côtés de l'abdomen et la membrane interfémorale, lesquels sont blancs teints légèrement de fauve. Les membres et la membrane alaire marron foncé, comme le corps.

Incisives supérieures internes bilobées, les externes pointues et arrivant presque à l'extrémité de la pointe extérieure de la 1^e incisive. La prémolaire supérieure accolée à la canine et à la première molaire, triangulaire et assez forte. A la mâchoire inférieure la première prémolaire de la même forme que la seconde et ayant deux tiers de la hauteur de celle-ci. Les incisives inférieures trilobées et implantées en travers des bords de la mâchoire.

Plus petit que le *V. tenuipinnis*, Peters; inférieur même en dimensions aux *Vesperugo pipistrellus* et *V. Kuhli*, desquels il se rapproche par son système de coloration. L'absence de la première prémolaire supérieure ne permet pas de le confondre avec ces espèces. La couleur des régions inférieures et la forme du tragus, cultriforme et non pas securiforme, doivent servir à le distinguer du *V. tenuipinnis*, qui vient également de la Guinée.

Dimensions: Tête et corps 35 mm.; queue 25; oreille 9; tragus 5; avant-bras 26; pollex 4; 3^e doigt 43; jambe 10; pied 5.

Deux femelles envoyées de Bissau en 1879 par M. Rodrigo da Costa.

Vesperus Humbloti A. Milne-Edwards, 1881

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Le Muséum d'Histoire naturelle vient de recevoir de Madagascar une collection fort importante et riche en objets rares ou inconnus; elle lui a été offerte par M. Humblot, qui a surtout exploré, sur la côte est, la région comprise entre Fou.apointe te le lac l'Aincatre.

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Un petit *Vesperus* (*V. Humbloti*, nov. sp.) se rapproche beaucoup du *V. minutus*, mais ses dimensions sont plus fortes et ses dernières vertèbres caudales sont complètement incluses dans la membrane interfémorale.

Vesperus pusillus Noack, 1889

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Taf. II, Fig. 2, Taf. V, Fig. 60 u. 61.

Spiritus-Exemplar M. Boma, März, Coll. HESSE.

Diagnose: Winzig kleiner *Vesperus* mit langem, schmalem, wenig eingebuchtetem Ohr, nach oben breit dreieckig verbreitertem, mit der Spitze nach innen gebogenem Tragus, kurzer stumpfer Nase, dreizackigen äusseren Incisiven, geringer Ausbuchtung der Schenkelflughaut, freiem letzten Schwanzwirbel, von tief russschwarzer, unten wenig hellerer Farbe.

Beschreibung: Nase kurz und stumpf mit runder Muffel, die Nasenlöcher durch eine sehr seichte Furche getrennt. Die wulstige Oberlippe ist durch eine tiefe horizontale Furche abgesetzt. Die Oberlippe an der Seite dick wulstig, unter dem Kinn (nicht an der Oberlippe) eine starke runde Warze. Von den Gaumenfalten sind die beiden ersten ungebrochen, die folgenden durch einen breiteren Zwischenraum getrennt und gebrochen, die Halbbogen auf der Innenseite nach hinten gezogen, die letzte, stark wulstige Doppelfalte innen sehr weit nach hinten reichend. Das lange schmale Ohr mit oval abgerundeter Spitze oben am äusseren Rande etwas eingebuchtet, unten der Lappen fast bis zum Mundwinkel hin vorgezogen. Tragus an der Basis schmal, nach oben verbreitert, innere Kante concav, äussere fast gerade, die Spitze bildet ein rundliches Dreieck, dessen Spitze nach innen gerichtet ist. Die vordere Fläche des Tragus ist ausgehöhlt. Der Daumen ist frei, der Humerus oben in der proximalen Hälfte behaart, unten nur zunächst dem Körper, die Flughaut auf 2 - 3 mm vom Körper dünn behaart, die Schwanzflughaut, aus welcher

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der letzte Wirbel herausragt, ist stark zugespitzt, oben nackt, die Ausbuchtung unbedeutend. Der Hinterleib ist auffallend schwächig, der Penis lang herabhängend. Scrotum ziemlich gross. Färbung des Körpers und der Flughaut tief schwarzbraun, das Ohr tief schwarz, auf der Unterseite sind die Haarspitzen gelblichbraun, doch ist die Gesamtfärbung nur wenig heller als oben.

Maasse: Körper 31 mm, Schwanz 22, Ohr 7, mittlere Breite 4, Tragus 4, Humerus 17, Unterarm 26, Daumen 3; II 20.5; III 35; IV 31; V 28. Femur 10, Unterschenkel 10.5, Fuss 4.5.

An dem sehr zarten Schädel (Taf V, Fig.60) mit etwas blasig aufgetriebenen Stirnbeinen ist die Schädelkapsel doppelt so lang wie die Nase, die Kapsel ziemlich flach und breit, vor den Augen mässig eingeschnürt, die obere Contour mässig gebogen, die

Stirn etwas gewölbt und die Nasenbasis mässig deprimirt. Der Oberkiefer ist vorn nur wenig in die Höhe gezogen, das Hinterhaupt mässig gerundet und unten nach vorn gezogen, besonders am unteren Rande des grossen For. occipit., die ziemlich grossen runden Bullae aud. nicht vom Tympanum getrennt, mit langem seitlichen Halbringe. Die Basis cranii liegt auffallend hoch, so dass die Scheitelhöhe eine geringe ist.

Der Unterkiefer ist nach vorn schmal zugespitzt, die Symphyse kräftig, unten mit rundlichem Zacken, der horizontale Ast ziemlich stark gebogen, so dass der Winkelfortsatz bis in die Höhe der Zahnbasis emporgezogen ist. Der Proc. cor. ist etwas nach vorn, der Winkelfortsatz nach aussen, die hintere Kante des aufsteigenden Astes schwach S-förmig gebogen.

Maasse: Scheitellänge 10.5, Basilarlänge 9, grösste Scheitelbreite in der Mitte 7, am Hinterhaupt 6.5, Einschnürung 3.25; Breite des Oberkiefers 4, Gaumenlänge 4, das runde Hinterhauptloch 3.

Unterkiefer bis zum Condylus 7, Höhe des horizontalen Astes in der Mitte 0.75, aufsteigender Ast unter dem Proc. coron. 2, Breite desselben 2.

Gebiss. Oben I (Taf. V, Fig. 61) innen einspitzig, hinten mit kleinen Basalzacken, hakig gekrümmt, die Spitzen stark convergirend. I aussen sehr klein, dreispitzig, indem der vordere kleinere Zacken eine Doppelspitze hat. Der Zahn steht fast senkrecht, ein wenig nach aussen. C mit unbedeutendem Basalwulst, der hinten etwas ausgezogen ist, schlank, wenig gebogen, nach hinten gerichtet, innen mässig cannelirt. $P = \frac{3}{4} C$, fast gerade, senkrecht, conisch zugespitzt,

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durch eine Lücke von C getrennt, dicht an M I gerückt. M I und II identisch, mit niedrigen Zacken, von den beiden Aussenzacken der erste nach vorn gerichtet, W-förmige Kaufläche, die beiden inneren Nebenzacken höher als die äusseren, der vordere etwas höher, der vertiefte innere Nebenhöcker klein, der innere Rand vorn mit Zacken. M III schmal, der hintere Theil des W von der Mitte des ersten Haarstrichs an fehlt, hinter dem inneren Nebenzacken noch ein ganz kleiner.

Unten I klein, dreilappig. nach vorn gerichtet. C, P und M mit Basalwulst. C schlank, ziemlich gerade, wenig nach vorn, aber nicht nach aussen gerichtet. Zwischen C und P I eine kleine Lücke, die übrigen Zähne in geschlossener Reihe. $P I = \frac{2}{3} C$, mässig gebogen, etwas nach hinten gerichtet, innen cannelirt, der Basaltheil nach hinten ausgezogen. P II etwas höher, aber viel schlanker, fast senkrecht stehend. M I und II identisch, der vordere Aussenzacken höher, fast so hoch wie C, etwas nach vorn gerichtet, die beiden inneren Zacken niedriger, etwas nach hinten gerichtet. Bei M III der hintere Aussenzacken und die beiden Innenzacken niedriger.

Von *Vesperus brunneus* THOMAS (in: Ann. Mag. Nat. Hist. 1880, p. 165, DOBSON, Geogr. distrib. Chiropt., p. 17) unterschieden durch geringere Grösse, längere, schmalere Ohren, die freie Schwanzspitze, die dunklere Färbung. *Vespertilio pusillus* LE CONTE vom Gabun (in: Proc. Ac. Nat. Hist. Philadelphia 1857, p. 10 u. 11) sieht ähnlich aus hat aber ein viel kürzeres Ohr und lanzettförmigen Tragus, ist übrigens, da LE CONTE nichts über die P sagt, so oberflächlich beschrieben, dass die Gattung, geschweige die Species, fraglich ist.

Die genannten drei tief dunklen Chiroptera sind meines Wissens ausser *Vesperugo stampflii* Jent. (in: Notes 1888, p.55), der schwarz mit rothbraunen Flecken ist, die einzigen bisher in Afrika gefundenen, während sich diese tief schwarzbraune Färbung bei südamerikanischen Arten öfter findet.

Vesperus tenuipinnis Peters, 1872

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Eine durch ihre Färbung sehr auffallende Art: oben dunkel rostbraun mit einfarbigen Haaren, Ohren, Gliedmassen und die ganze Bauchseite schneeweiss, nur die Basis der Haare braun; Flughäute farblos durchsichtig.

Der erste obere Schneidezahn ist breit, an seiner Vorderseite gefurcht, fast zweispitzig. Der zweite ist sehr kurz und liegt vor dem äussern Theile des ersten und durch einen Zwischenraum von dem Eckzahn getrennt. Die unteren dreilappigen Schneidezähne stehen dem Kiefferrande fast parallel. Der erste untere Prämolazahn ist etwa $\frac{1}{3}$ niedriger, aber im Querschnitt wenig kleiner als der zweite; er ist ungefähr halb so hoch wie der untere Eckzahn.

Die Ohren sind dreieckig abgerundet, an der Basis des Aussenrandes mit einem abgerundeten, deutlich abgesetzten Läppchen

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versehen und im Allgemeinen ebenso wie die breite beilförmige Ohrklappe denen von *V. noctula* ähnlich.

Die Körperbehaarung dehnt sich seitlich aus über das erste Drittel des Oberarms, die Hälfte des Oberschenkels und die Basis des Schwanzes.

Die Flughäute sind sehr dünn und durchsichtig, so dass man jede Schrift durch sie unbehindert lesen kann und gehen bis an die Zehenbasis. Der Schwanz ist bis auf die äusserste Spitze von der Schenkelflughaut eingeschlossen.

Meter
 Totallänge: 0.076
 Kopf: 0.014
 Ohrhöhe: 0.0115

Vord. Ohrrand: 0.008
 Ohrberite: 0.0085
 Tragus: 0.0045
 Oberarm: 0.016
 Vorderarm: 0.029
 L.1.F. Mh. 0.001; 1 Gl. 0.0023; 2 Gl. 0.0013: 0.005
 L.2.F. - 0.027; - 0.0015; - 0.0285
 L.3.F. - 0.0285; - 0.0103; - 0.0083; Kpl. 0.006
 L.4.F. - 0.0272; - 0.009; - 0.006; - 0.002
 L.5.F. - 0.0267; - 0.006; - 0.0037; - 0.002
 Schwanz: 0.030
 Oberschenkel: 0.011
 Tibia: 0.011
 Fufs: 0.006
 Sporn: 0.013

Ein weibliches scheinbar ausgewachsenes Exemplar aus Guinea.

Xiphonycteris Dollman, 1911

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Size similar to that of the smaller species of *Nyctinomus*. Wings small, membranous area much reduced. Ears, feet, and tail much as in *Nyctinomus*.

Skull with median anterior palatal emargination like that of *Nyctinomus*, but emargination quite small and inconspicuous, though extending behind roots of incisors. Teeth according to the following formula: -

i. $1\frac{1}{1-1}$, c. $1\frac{1}{1-1}$, p. $2\frac{2}{2-2}$, m. $3\frac{3}{3-3}$ = 28.

Upper incisors quite small and closely in contact with canines; widely separated from each other, wider apart at base than at tip. Upper canine rather similar to that of *Nyctinomus*, but with cingulum very prominent and shaft of tooth longer, much flatter anteriorly, and pointing slightly outwards. Base of canines much enlarged, the anterior edges of the cingula on a level with the anterior limit of the upper incisors; when looked at from above the cingula appear as two prominent crescentic ridges with the minute incisors between them, the incisors being so placed as to resemble two anterointernal cusps on the cingula of the canines. Behind the shaft the cingulum is expanded to form a large flat area, its posterior edge closely in contact with the small anterior premolars. Upper premolars and molars as in *Nyctinomus*, the molars with well-developed hypocones. Lower incisors reduced to a single pair of very minute functionless teeth, bridged over by the junction of the lower canines; cutting-edges deeply bifid. Lower canines with well-developed cingula, forming anteriorly prominent secondary cusps, which meet together in the mid-line as a bridge over the minute incisors; posterior portion of cingulum large and well developed, much more so than in the allied genera. Lower molars and premolars as in *Nyctinomus*.

Type, *Xiphonycteris spurrelli*.

This genus would appear to be most nearly related to the

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genus *Nyctinomus*, which it resembles in general external characters and in the presence of the small upper premolar; but the absence of i_2 , the peculiar position of the single pair of minute lower incisors, and the unusual shape and arrangement of the upper and lower canines are more than sufficient reasons for considering the genera as quite distinct. As regards the genus *Chærephon* the same differences hold good, with the additional character of the median anterior palatal emargination present in *Xiphonycteris*.

In the possession of only a single pair of lower incisors, and in having the lower canines in contact with one another, this genus in some ways resembles the South-American genus *Molossops*; but the two genera, on account of numerous important differences, cannot be looked upon as close allies.

Xiphonycteris spurrelli Dollman, 1911

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General appearance and size much as in the smaller species of *Nyctinomus*. Ears roughly triangular in shape, with rounded corners; tragus and antitragus apparently normal. Feet comparatively small; toes hairy, the hairs extending along the margin of the interfemoral membrane for about 10 mm. Wings small, forearm only 27 mm. long; wing-membrane very narrow. Hair on back short, a few longer hairs near base of tail. General colour of back rusty red, hairs rather lighter at base than at tips. Under surface of body buffy white, the hairs becoming brownish on the flanks and sides of neck. Tail, as described above, with terminal half free. Interfemoral membrane naked on upper surface, below thinly clad with very minute white hairs.

Skull with rather narrow brain-case, markedly so in the occipital region. Supraoccipital crest well developed, extending back some distance beyond the exoccipital region. Sagittal crest but little developed. Zygoma fairly strong, expanded posteriorly.

Anteprbital foramen large. Lachrymal ridge faintly indicated. Anterior palatal emargination about as large as base of upper incisor, extending well behind the roots of incisors. Palate slightly arched laterally, nearly flat antero-posteriorly; pterygoids almost parallel, slightly divergent behind; basisphenoid pits moderately developed. Auditory bullæ small and rather flat.

Teeth as described above.

Dimensions of the type (measured in flesh): -

Head and body 56 mm.; tail 24; hind foot 8; ear 13; forearm 27.

Skull: greatest length 17; basilar length 12.2; condylo-basilar length 14; greatest zygomatic breadth 10; inter-

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orbital breadth 3.5; breadth of brain-case (across squamosal region) 8.6; breadth of rostrum (across lachrymal region) 6; palatilar length 5.8; width of palate inside m^2 3.3; width across palate outside m^2 7; postpalatal length 6.5; length of upper tooth-row from front of canine to back of m^3 6.4; length of upper tooth-row from front of first premolar to back of m^3 4.9; height of canine 3.2.

Hab. Bibianaha, 60 miles W. of Kumasi, Gold Coast. Altitude 700 feet.

Type. Adult male. B.M. no. 11.1.11.1. Original number 34. Collected on December 8th, 1910, by the donor, Dr. H. G. F. Spurrell.

It gives me great pleasure to associate this new and interesting bat with the name of Dr. Spurrell, to whom the National Collection is already indebted for many rare and unique West-African mammals.